

# What drove the Stony Coral Tissue Loss Disease outbreak in Florida ?

**Thomas Dobbelaere**<sup>1,\*</sup>, **Erinn Muller**<sup>2</sup>, **Lewis Gramer**<sup>3,4</sup>, **Dan Holstein**<sup>5</sup> and **Emmanuel Hanert**<sup>1,6</sup>

<sup>1</sup> *Earth and Life Institute (ELI), UCLouvain, Louvain-la-Neuve, Belgium*

<sup>2</sup> *Coral Health and Disease Program, Mote Marine Laboratory, Sarasota, FL, USA*

<sup>3</sup> *Cooperative Institute for Marine and Atmospheric Studies (CIMAS), University of Miami, Miami, FL, USA*

<sup>4</sup> *Atlantic Oceanographic and Meteorological Laboratory (AOML), NOAA, Miami, FL, USA*

<sup>5</sup> *Department of Oceanography and Coastal Sciences, College of the Coast and Environment, Louisiana State University, Baton Rouge, LA, USA*

<sup>6</sup> *Institute of Mechanics, Materials and Civil Engineering (IMMC), UCLouvain, Louvain-la-Neuve, Belgium*

Correspondence\*:

Earth and Life Institute (ELI), UCLouvain, Croix du Sud 2 box L7.05.16, B-1348

Louvain-la-Neuve, Belgium

thomas.dobbelaere@uclouvain.be

## 2 ABSTRACT

For about six years, the Florida Reef Tract (FRT) has been experiencing an outbreak of the Stony Coral Tissue Loss Disease (SCTLD). First reported off the coast of Miami-Dade County in 2014, the SCTLD has since spread throughout the entire FRT with the exception of the Dry Tortugas. However, the causative agent for this outbreak is currently unknown. Here we show how a high-resolution bio-physical model with a modified patch SIR epidemic model can inform on the potential characteristics of the causative agent of the disease and its vector. In this study, the agent is assumed to be transported within composite material (such as coral mucus, dying tissues and/or resuspended sediments) driven by currents and potentially persisting in the water column for extended periods of time. In this framework, our simulations suggest that the SCTLD is likely to be propagated within neutrally buoyant materials driven by mean barotropic currents. Calibration of our model parameters with field data show that corals are then infected within a mean transmission time of 6.45 days. Furthermore, the propagation speed of the disease through the FRT is shown to be very sensitive to the value of an infection threshold, defined the proportion of infectious corals that causes an exponential growth of the disease on the colony. Our results present a new connectivity-based approach to understand the spread of the SCTLD through the FRT. Such method can provide a valuable complement to field observations and lab experiments to support the management of the epidemic as well as the identification of its causative agent.

**Keywords:** stony-coral-tissue-loss disease, biophysical modeling, Florida reef tract, spatial epidemiology, connectivity

## 1 INTRODUCTION

Coral diseases are a major threat to coral reef ecosystems and have led to significant declines in coral cover especially within the Caribbean region (Richardson et al., 1998; Sutherland et al., 2004; Aronson and

23 Precht, 2001; Harvell et al., 2007; Miller et al., 2009; Brandt and McManus, 2009). Indeed, the Florida  
24 reef tract, which was dominated by *Acropora palmata* and *Acropora cervicornis*, and often had 30% coral  
25 cover until the 1970s/80s (Dustan and Halas, 1987; Porter and Meier, 1992), is now dominated by bare  
26 substrate, octocorals, and macroalgae with only approximately 5% stony coral cover remaining (Ruzicka  
27 et al., 2013). The loss of the branching Acroporid species was attributed primarily to a disease outbreak,  
28 termed white band disease (Aronson and Precht, 2001), but several other threats such as habitat reduction,  
29 eutrophication, overfishing, and bleaching likely all contributed to these species decline (Team, 2005).  
30 Subsequent losses of coral cover within the region were often linked to additional disease incidences and  
31 repeated regional coral bleaching events as a result of global climate change (Kuta and Richardson, 1996;  
32 Richardson et al., 1998; Sutherland et al., 2004; Gardner et al., 2003; Aronson and Precht, 2006; Kuffner  
33 et al., 2015; Manzello, 2015). A novel coral disease outbreak, termed Stony Coral Tissue Loss Disease  
34 (SCTLD), is now threatening the last vestiges of coral throughout the Florida reef tract.

35 SCTLD was first documented off the coast of Miami-Dade County in the summer of 2014 by Precht et al.  
36 (2016) and has since spread throughout the entire Florida reef tract with the exception of the Dry Tortugas.  
37 To date, SCTLD has been observed affecting over 20 different stony corals species. A case definition  
38 of SCTLD has been compiled to describe the visual appearance and ecology of SCTLD (NOAA, 2018).  
39 Briefly, the gross morphology of SCTLD is described as focal or multifocal, with locally extensive to  
40 diffuse areas of acute to subacute tissue loss distributed basally, peripherally, or both. In some cases, tissues  
41 bordering areas of chronic tissue loss show indistinct bands (1–5 cm) of pallor, progressing to normal  
42 pigmentation away from the denuded skeleton. There is also a range in coral susceptibility to SCTLD, with  
43 species categorized as highly susceptible (e.g., *Dendrogyra cylindrus*, *Dichocoenia stokesii*, *Meandrina*  
44 *meandrites*), moderately susceptible (e.g., *Orbicella* spp., *Montastraea cavernosa*, *Siderastrea siderea*,  
45 *Stephanocoenia intersepta*), or tolerant (e.g., *Porites* spp., *Acropora* spp.). Unfortunately, SCTLD has  
46 not remained isolated in the Florida reef tract and has now been recorded in Mexico (Alvarez-Filip et al.,  
47 2019), the USVI (Blondeau et al., 2020) and several other location around the Caribbean. The continued  
48 persistence of the outbreak, the high number of species affected, and the large geographical range of reports  
49 consistent with the case definition suggests that SCTLD is the largest coral disease outbreak ever recorded.

50 Large-scale spatial epidemiologic analyses showed that the reefs in Florida with SCTLD are clustered,  
51 supporting a contagious mode of transmission (Muller et al., 2020). Similarly, aquaria-based experiments  
52 indicate SCTLD can be transmitted through direct contact or indirectly through the water column (Aeby  
53 et al., 2019) suggesting water can be a SCTLD vector, at least within a controlled setting. The initial  
54 exponential increase in spread among reefs from the disease epicenter (Precht et al., 2016) and the persistent  
55 subsequent linear rate of spread of SCTLD (Muller et al., 2020), north along South Florida reefs and south  
56 into the Florida Keys, indicates that water currents may play a role in disease transmission. Furthermore,  
57 the rate of spread, estimated at 100 m per day, suggests surface currents are likely too fast to have spread  
58 SCTLD within the region. These results imply that the bottom boundary layer, which is significantly slower  
59 than surface currents, may be the vertical layer in which transmission occurs (Aeby et al., 2019). However,  
60 to date, there have been no efforts linking local hydrodynamic modeling efforts with the spatio-temporal  
61 dynamics of SCTLD in Florida.

62 Estimating the transport of infectious material from reef to reef by currents cannot be performed  
63 empirically. However, experimentally-calibrated numerical models that simulate currents can provide a  
64 realistic picture of the dispersal of infected matters. Nonetheless, accurately modeling water circulation at  
65 the spatial scales that affect this dispersal remains a key challenge, as small-scale flow features such as  
66 recirculation eddies around reefs and islands strongly impact exchanges between reefs (Wolanski, 1994;

Burgess et al., 2007; Figueiredo et al., 2013). In this context, models that can explicitly simulate flow features down to the reef scale are needed. This represents a spatial resolution of the order of 100-1,000 m in dense reef systems. As of today, most regional ocean models using traditional numerical methods cannot achieve such resolution because of the computational resources it requires. To our knowledge, the best resolution currently available among these models in the FRT is  $\sim 900$  m with the FKEYS-HYCOM model that has been developed for the Florida Keys region (Kourafalou and Kang, 2012; Sponaugle et al., 2012; Vaz et al., 2016). Unstructured-mesh ocean models offer a potential solution to this resolution issue by locally increasing the model resolution close to reefs and islands (Lambrechts et al., 2008; Thomas et al., 2014, 2015), in order to focus the computational resources where they are most needed. High resolution bio-physical dispersal models can be used to build the potential connectivity between reefs and therefore approximate exchanges between colonies in the complex topography of the coral reef systems (Frys et al., 2020).

Marine diseases differ significantly from better studied terrestrial diseases, namely due to the potential for long environmental residence times, during which pathogens may survive and disperse through the water (Harvell et al., 2007; Sokolow, 2009). Several recent studies have attempted to adapt traditional epidemic models (Susceptible-Infectious-Recovered, or SIR models) to coral reef systems (Sokolow, 2009; Bidegain et al., 2016a,b). Novel approaches have included developing pathogen pools (Bidegain et al., 2016a,b), and to model at the metapopulation scale, rather than at the scale of coral holobionts (Sokolow, 2009). Both these approaches are attempting to address the same issue: Infection occurs between patches of entirely sessile animals, through the dispersal of pathogen(s). Thus, there are internal within-patch disease dynamics and metapopulation-scale between-patch dynamics occurring simultaneously. The epidemic model developed in this study utilizes the same basic architecture of previous coral reef SIR models, but rather than assume pathogen pools (e.g. Bidegain et al. (2016a,b)) or ignore internal patch dynamics (e.g. Sokolow (2009)), we have modeled both within-patch disease dynamics and the dispersal of pathogen explicitly using potential connectivity networks.

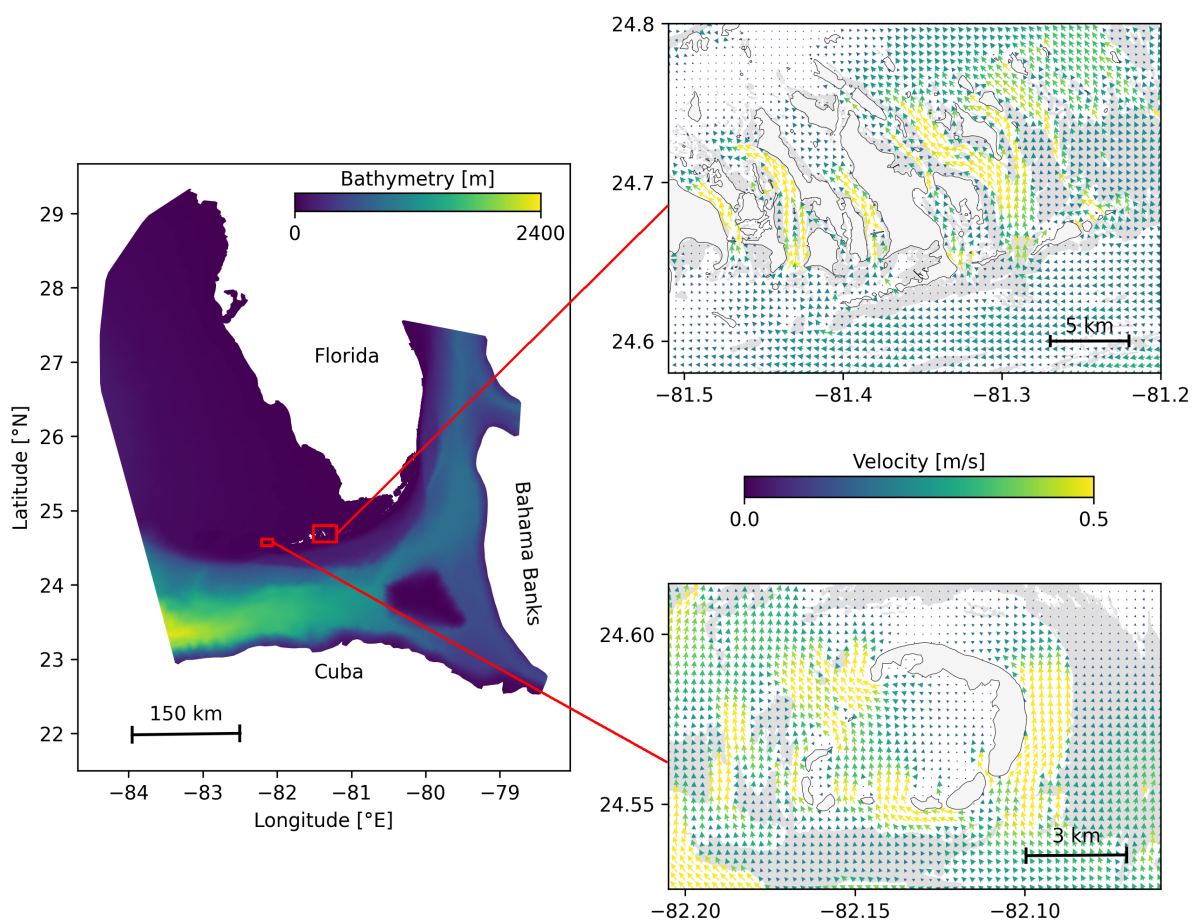
The objective of this study is to deduce the probable propagation mechanism of the SCTL throughout the FRT by developing an experimentally-calibrated epidemio-hydrodynamic model. With a resolution of about 100 m, this model can capture potential exchanges of infectious matter between reefs that would be ignored by coarser models. By reproducing the observed spread of disease between 1st May 2018 and 1st April 2014, we provide insight on the characteristics of the disease agent and its vector. Ultimately, our model, coupled with lab and field works, would support the management of the epidemic and the identification of its causative agent.

## 2 METHODS

### 2.1 Modeling reef connectivity

In this study, we focus on the exchanges of infectious material between coral reefs driven by ocean currents, which therefore have to be accurately simulated. An ocean model should provide a realistic large-scale circulation while also resolving small-scale flow features down to the scale of individual reefs. In this study, we use the unstructured-mesh depth-integrated coastal ocean model SLIM<sup>1</sup> to simulate ocean currents over an area that includes the FRT but also the Florida Strait and part of the Gulf of Mexico (Fig. 1). By using an unstructured mesh, we can increase the model resolution only over the FRT and hence concentrate computational resources where they are most needed. SLIM, being a depth-averaged model, is

<sup>1</sup> <https://www.slim-ocean.be>



**Figure 1.** Model computational domain with the bathymetry (left) and close-up views of the mesh with snapshots of the currents on May, 25 2018 at 00:00, for the Marquesas Keys (bottom) and the Lower Keys (top). This illustrates the benefits of unstructured meshes to represent the fine-scale details of the topography and hence simulate currents down to the scale of individual reefs (shown in darker grey) and islands (shown in lighter grey with black contours).

107 well suited to shallow-water flows. Details of the model formulation and validation are provided in Frys  
 108 et al. (2020).

109 The mesh resolution depends only on the distance to the coast but we distinguish between the coastlines  
 110 along the FRT where we impose a maximum resolution of 100 m and the other coastlines along which  
 111 the maximum resolution is 2500 m. The mesh has been generated with the open-source mesh generator  
 112 GMSH (Geuzaine and Remacle, 2009) and has about  $7 \times 10^5$  elements. The coarsest elements, far away  
 113 from the FRT, have a size of about 10 km. An illustration of ocean currents simulated on that mesh are  
 114 shown in Fig. 1. It shows how a 100-m spatial resolution allows us to simulate fine-scale details of the flow,  
 115 such as recirculation eddies and currents within the dense reef system in the Lower Keys that consist of  
 116 many individual reefs with narrow passages in between.

117 The simulated currents can then be used to model dispersal of infectious material throughout the FRT.  
 118 In this study, 3 types of potential vector carrying the disease causative agent were considered: positively  
 119 buoyant (e.g. mucus and surfactant), neutrally buoyant (e.g. fines, pelagic organisms) and negatively  
 120 buoyant (e.g. sediments, composites, demersal organisms). As SLIM is a depth-averaged model, the  
 121 mean currents it generates are well suited to model the dispersal of neutrally buoyant material remaining



within the water column. However, these currents must be modified to correctly represent the dynamics of materials evolving in the surface and bottom boundary layers. Therefore, surface current response to winds is estimated by adding 1.5% of the wind speed to SLIM currents with a windage angle of 45° to the right for positively buoyant particles. Such parametrization is shown to be an accurate approximation of wave-induced Stokes drift and quasi-Eulerian surface currents by (Ardhuin et al., 2009). For negatively buoyant materials, on the other hand, bottom currents are obtained by taking 60% of SLIM currents velocity with a veering angle of 15° to the left. [\[Some references about the parametrization for bottom currents\]](#)

Using these three velocity fields, virtual particles are then released on all the reefs composing the FRT to model the dispersal of infected materials carrying the disease causative agent. The locations of the reefs of Florida are extracted from the "coral reefs and hardbottom" layer of the Unified Florida Reef Tract Map (FWC, 2017). The polygon of this reef map are then further divided into 500 m squares in order to track the propagation of the disease with a finer geographical resolution, generating a total of 16823 polygons. At the beginning of each simulated months and for each type of currents, a total of about  $1.5 \times 10^6$  particles are released over all the reef polygons. These particles have a state composed of their polygon of origin as well as their mass, initialized to 1, that they loose at a constant rate  $\gamma$  as they are moved by surface, mean or bottom currents. In this study, the value of  $\gamma$  is chosen so that particles have a half life of 30 days. When the particles are brought over reef polygons by currents, the amount of infected mass that lands on the polygon is recorded in monthly potential connectivity matrices whose entries are denoted  $C_{ij}$ . The matrix rows correspond to the source reefs and the columns correspond to the destination reefs. Hence  $C_{ij}$  represents the mass of infected material originating from sub-reef  $i$  that has settled on sub-reef  $j$ . This matrix is then normalized by dividing each of its rows  $i$  by the the total mass of particles released on polygon  $i$  in order to obtained the normalized potential connectivity matrix  $\tilde{C}$ , whose entry  $\tilde{C}_{ij}$  gives the probability that infectious material produced on sub-reef  $i$  settles on sub-reef  $j$ . Connectivity matrices are computed for each type of current and for each month of the simulated period.

These connectivity matrices can be more easily handled by interpreting them as large graphs whose vertices are reefs. They can then be analyzed using graph theory tools. In this study, four potential connectivity measures are used to interpret the monthly computed graphs. These indicators are described in Table 1. The first indicator is the weighted connectivity length (WCL), that gives the average dispersal distance from origin to destination for material produced on a given reef. The weighted connectivity of reef polygon  $i$  writes:

$$WCL_i = \frac{\sum_j \tilde{C}_{ij} L_{ij}}{\sum_j \tilde{C}_{ij}} \quad (1)$$

where  $L_{ij}$  is the distance between origin reef  $i$  and destination reef  $j$ . Another measure of the spreading potential of reef  $j$  is its out-degree, *i.e.* the product of the number of connections originating from reef  $j$  by the quantity of infectious matter its sends to the network. This indicator is obtained by computing the number non-zero entries of row  $i$  of the potential connectivity matrix  $C$  and multiplying it with  $\sum_j C_{ij}$ . The information given by the out-degree is complemented by the proportion of infectious elements produced on reef  $i$  that successfully settles on a reef, called the fraction exchanged of reef  $i$ . This indicator is given by  $\sum_j \tilde{C}_{ij}$ . Finally, the isolation of reef  $i$  in the network is given by the self recruitment, *i.e.* the proportion of infectious matter settling on reef  $i$  that originates from reef  $i$ , computed by  $C_{ii} / \sum_j C_{ji}$ . A large self-recruitment value indicates that few infectious matter produced elsewhere settles on the reef and thus that it is isolated from the rest of the network.

Indicators	Description	What it shows
Weighted connectivity length (WCL)	Average dispersal distance from origin to destination reef for all infectious elements released over a reef	Average distance at which a reef can send infectious matter
Out-degree	Number of out-going connections originating from a given reef multiplied by the total mass exchanged	Potential for a reef to spread the disease
Fraction exchanged	Fraction of infectious material produced on a given reef that settles on another reef	Success rate of potential disease spread
Self recruitment	Fraction of infectious material settling on a given reef that has been released on the same reef	Potential for disease to settle on a given reef

**Table 1.** Indicators used to analyze the modeled exchanges of infected material for each considered type of currents and for each simulated month

## 2.2 Epidemiological modeling

### 2.2.1 Model equations

The spread of the SCTL throughout the FRT is simulated using a connectivity-based Kermack-McKendrick SIR model (Brauer, 2008). SIR models are among the most standard epidemiological models. They divide individuals into three compartments: susceptible (S), infectious (I) and removed (R). When affected by the disease, susceptible individuals become infectious and infect other susceptible individuals until they are removed, either by recovery or death. Such models usually rely on the hypothesis of an homogeneous, well-mixed population. To account for the spatial heterogeneity of the FRT, the basic SIR formulation is here modified by considering the proportions of susceptible ( $S_j$ ), infectious ( $I_j$ ) and removed ( $R_j$ ) corals of each polygon reef  $j$ . In this epidemiological model, individual reefs interact through the exchange of infectious material as represented by the connectivity matrix. For each sub-reef  $j$  and at any time, the following relations hold:  $0 \leq S_j, I_j, R_j \leq 1$  and  $S_j + I_j + R_j = 1$ . The evolution of these proportions through time is governed by the following equations:

$$\begin{aligned}
 \frac{dS_j}{dt} &= -\beta \sum_i \frac{A_i}{A_j} I_i \tilde{C}_{ij} S_j - \beta'(I_j) S_j I_j \\
 \frac{dI_j}{dt} &= \beta \sum_i \frac{A_i}{A_j} I_i \tilde{C}_{ij} S_j + \beta'(I_j) S_j I_j - \sigma I_j \\
 \frac{dR_j}{dt} &= \sigma I_j
 \end{aligned} \tag{2}$$

where  $\tilde{C}_{ij}$  is the entry of reef  $(i, j)$  of the normalized potential connectivity matrix [-],  $A_i$  is the area of reef polygon  $i$  [ $\text{km}^2$ ],  $\sigma$  is the removal rate [ $\text{day}^{-1}$ ], and  $\beta$  and  $\beta'(I_j)$  are the inter- and intra-reef disease transmission rates [ $\text{day}^{-1}$ ], respectively. In this model, infectious corals of reef  $i$  can infect corals of reef  $j$  if there is non-zero probability of infectious material exchange from reef  $i$  to reef  $j$ , given by  $\tilde{C}_{ij}$ . Moreover, to account for coral resistance to the disease, the intra-reef transmission function  $\beta'(I_j)$  has the shape of a

smooth step function of the proportion of infectious corals  $I_j$  and writes:

$$\beta'(I_j) = \frac{\beta'_0}{2}(1 + \tanh[(I_j - I_0)/\tau]), \quad (3)$$

where  $I_0$  is a threshold on the infection population above which intra-reef transmission becomes significant, and  $\tau$  is a measure of the interval over which the transition for low to high transmission occurs. As long as the proportion of infectious corals on reef  $j$  is below  $I_0$ , the only infection mechanism taking place is connectivity-driven transmission at rate  $\beta$ . Once the threshold is exceeded ( $I_j \geq I_0$ ), intra-reef transmission with rate  $\beta'_0$  is activated. A larger value of threshold  $I_0$  corresponds to a greater resistance to disease for corals and therefore a slower spread of the disease within reef  $j$ . Coral birth and natural (*i.e.* non SCTL-related) death rates are not taken into account in this model, which amounts to assume that they balance each other. For this study the same values were used for  $\beta$  and  $\beta'_0$ .

## 2.2.2 Calibration

Transmission and removal parameters of  $\sigma$  and  $\beta'_0$  are fitted to disease prevalence observations averaged over all colonies from all monitored sites in the FRT in order to accurately simulate the temporal evolution of  $S_j, I_j, R_j$  on each infected reef polygon. **[Could you provide some details on these data prevalence observations ?]** To relate our model framework to the compiled data, Eqs. 2 are simplified to a single-reef SIR model:

$$\begin{aligned} \frac{dS}{dt} &= -\beta'_0 SI \\ \frac{dI}{dt} &= \beta'_0 SI - \sigma I \\ \frac{dR}{dt} &= \sigma I \end{aligned} \quad (4)$$

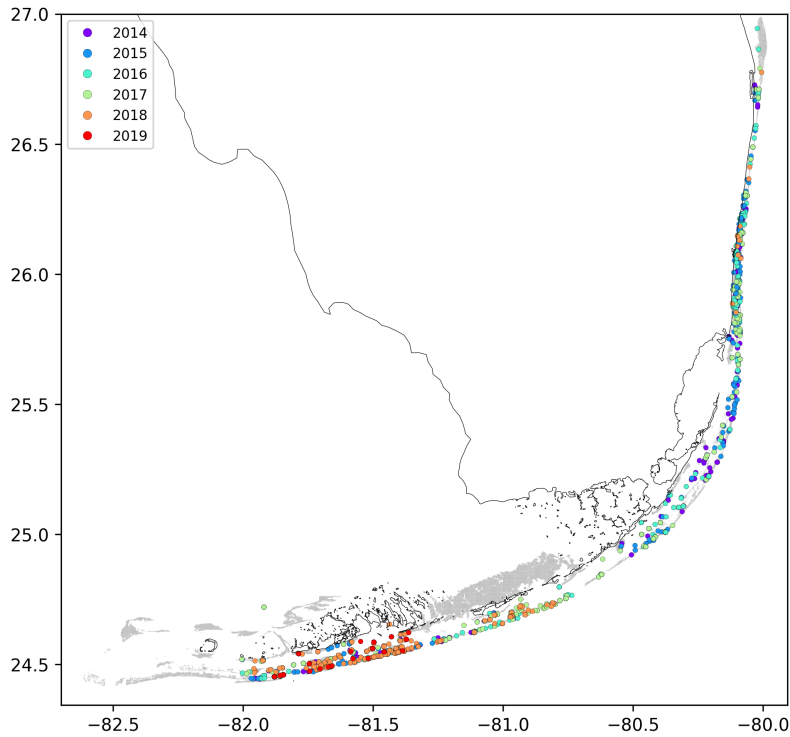
Due to the low values of the entries in the normalized connectivity matrix  $\tilde{C}_{ij}$ , intra-reef transmission, when activated, is the dominant infection mechanism of Eqs. 2. Consequently, Eqs. 4 give a reasonable approximation of the evolution of the disease on sub-reefs for which  $I_j > I_0$ . Using this approximation, the ratio  $\beta'_0/\sigma$  is imposed by matching the modeled proportion of susceptible corals remaining after the disease has vanished ( $S_\infty$ ) with observations. A standard property of a SIR model solution is that

$$S_\infty - \frac{\sigma}{\beta'_0} \log(S_\infty/S_0) = 1 \quad (5)$$

where the initial proportion of susceptible corals ( $S_0$ ) is taken equal to  $1 - I_0$  (see for instance Murray (2007)). In the framework of Eqs. 4, the ratio  $\beta'_0/\sigma$  gives the value of the basic reproduction number  $R_0$ , defined as the average number of secondary cases produced by one infected individual introduced into a population of susceptible individuals (Keeling and Rohani, 2007). This number is used in epidemiological models to determine whether an emerging infectious disease can spread in a population ( $R_0 > 1$ ) or not ( $R_0 < 1$ ). The obtained basic reproduction number is then used to express  $\sigma$  in terms of  $\beta'_0$  and calibrate its value in order to reproduce as well as possible the temporal evolution of the colonies-averaged susceptible population shown in Fig. 5.

## 2.2.3 Initialization

In order to solve Eqs. 2, initial conditions are needed, *i.e.* proportions of susceptible, infectious and recovered corals at the beginning of the simulated period. This information is constructed based on different



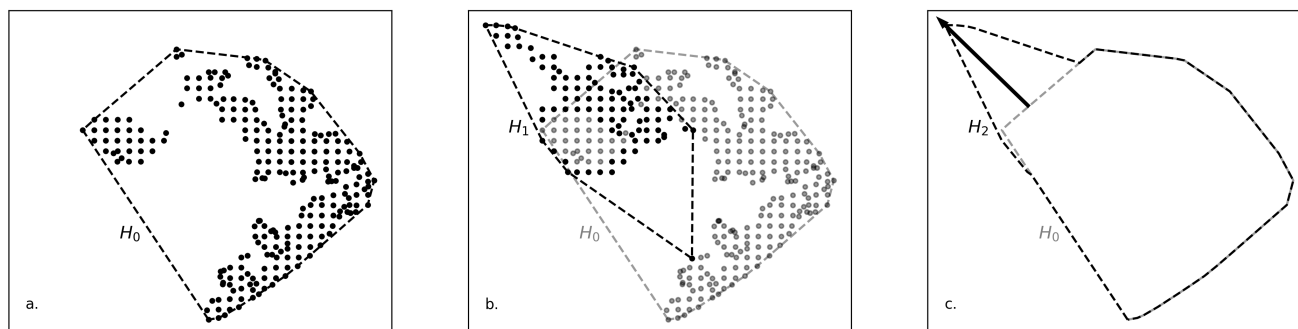
**Figure 2.** Locations of the disease observations between 2014 and 2019 recorded in the data sets used in this study

field-collected datasets: (i) Coral Reef Evaluation and Monitoring Project (CREMP; 2014–2017), (ii) CREMP Presence/Absence Data (CREMP P\_A; 2016–2017), (iii) Southeast Florida Coral Reef Evaluation and Monitoring Project (SECREMP; 2014–2017), (iv) Florida Reef Resilience Program Disturbance Response Monitoring (FRRP; 2014–2017), (v) Hurricane Irma Rapid Reef Assessment (IRMA; 2017, Viehman et al. (2018)), (vi) the Southeast Florida Action Network citizen science program (SEAFAN; 2014–2017), and (vii) the Southern Coral Disease Margin field effort (2017; Neely (2018)). These datasets give the locations and dates at which the SCTLD has been observed throughout the FRT. The locations of these observations are shown in Fig. 2. Using this information, we first delineate an infected zone by constructing the concave hull of the points where the disease was observed before May 2018. The reefs infected prior to the beginning of our simulated period are then defined as the reefs located inside the constructed zone. The time of observed infection is then spatially interpolated on each reef of the infected zone by kriging with a Gaussian semivariogram using Python `pyKrig` module. Assuming an initial state  $(S, I, R) = (1 - I_0, I_0, 0)$  when the disease was observed, the proportions of susceptible, infectious and removed corals on each reef of the infected zone on the 1st May 2018 is finally approximated using the simplified equations 4. Reefs outside of the infected zone are initialized with a population of 100% of susceptible corals.

#### 2.2.4 Computation of front speed

Muller et al. (2020) estimated the speed of the spreading STCLD epidemics at around 92 m/day in the southern section of the FRT. In order to assess our simulation results in regard to this value, we developed a methodology to compute the displacement of the disease front during a given time interval within our simulated period. First, the concave hull of the infected polygons at the beginning of the time interval  $H_0$  is computed. Then the concave hull of the polygons infected during the time interval  $H_1$  is computed while





**Figure 3.** Method used to compute the disease front displacement during a simulated time interval. **a.** Concave hull of the infected polygons at the beginning of the simulated period  $H_0$ . **b.** Concave hull of the polygons infected during the simulated period  $H_1$ . **c.** Arrow showing the computed front displacement during simulated time interval between  $H_0$  and  $H_2$ , the union of  $H_0$  and  $H_1$ .

the concave hull  $H_2$  is defined as the union of  $H_0$  and  $H_1$ . This methodology is illustrated in Fig. 3. The distance traveled by the disease front is then obtained by computing the maximum distance between a pair of points of  $H_0$  and  $H_2$ . The epidemics front speed is finally computed by dividing the resulting distance by the number of days in the simulated time interval.

## 2.3 Transmission experiments

In parallel to this modeling study, laboratory-based transmission experiments of SCTLD were conducted by several independent groups for various end points including transmission dynamics and samples for molecular and histological analysis. Requests for transmission data were sent to members of the ‘Transmission’ sub group of the Florida Disease Advisory Committee’s ‘Research’ working group as well as any other additional researchers that may have been conducting transmission studies on stony coral tissue loss disease. Data that was requested and subsequently provided included the location, dates, and duration of the experiment, the species used as the diseased colony (donor of disease agents) and apparently healthy colony (exposed to diseased agents), the number of successful transmissions as well as incubation period following a contact with disease agents prior to disease signs. Additional information included the size of the colonies used in the experiment, the percent tissue loss of the diseased (donor) colony at beginning of the experiment, and whether the apparently healthy (exposed) fragment was touching the diseased colony or not.

The average probability of successful disease transmission was determined by taking the mean of the number of colonies exposed to the disease in each study divided by the total number of coral colonies exposed to diseased colonies. The ‘incubation’ period was identified as the average number of days after an apparently healthy coral colony was exposed to a diseased colony before visual disease signs occurred (i.e., active tissue loss). Only corals that eventually showed disease signs were integrated within the incubation period calculation.

Data was provided from 8 different research groups representing 15 institutions and 19 total collaborators providing a total of 109 data points. After amalgamating the contributed data, the mean probability of transmission of SCTLD to an apparently healthy coral had a likelihood of approximately  $44.8 \pm 3.6\%$ . The probability of transmission ranged from 0 to 100% depending on the experiment. Additionally, the time between exposure of an apparently healthy coral to a diseased coral and subsequently showing initial signs of tissue loss (i.e., incubation period) was  $9.7 \pm 1$  days.

### 3 RESULTS

#### 3.1 Exchanges of infected materials

Among the three modes of transport, bottom currents exhibit the lowest range as they generate the networks with the smallest weighted connectivity length (Fig. 4). However, infectious matters transported by bottom currents have the largest settlement success rate as these currents generate the graphs with the largest fraction exchanged. Therefore, bottom currents tend to transport more infectious material on closer reefs compared to the two other modes of transport.

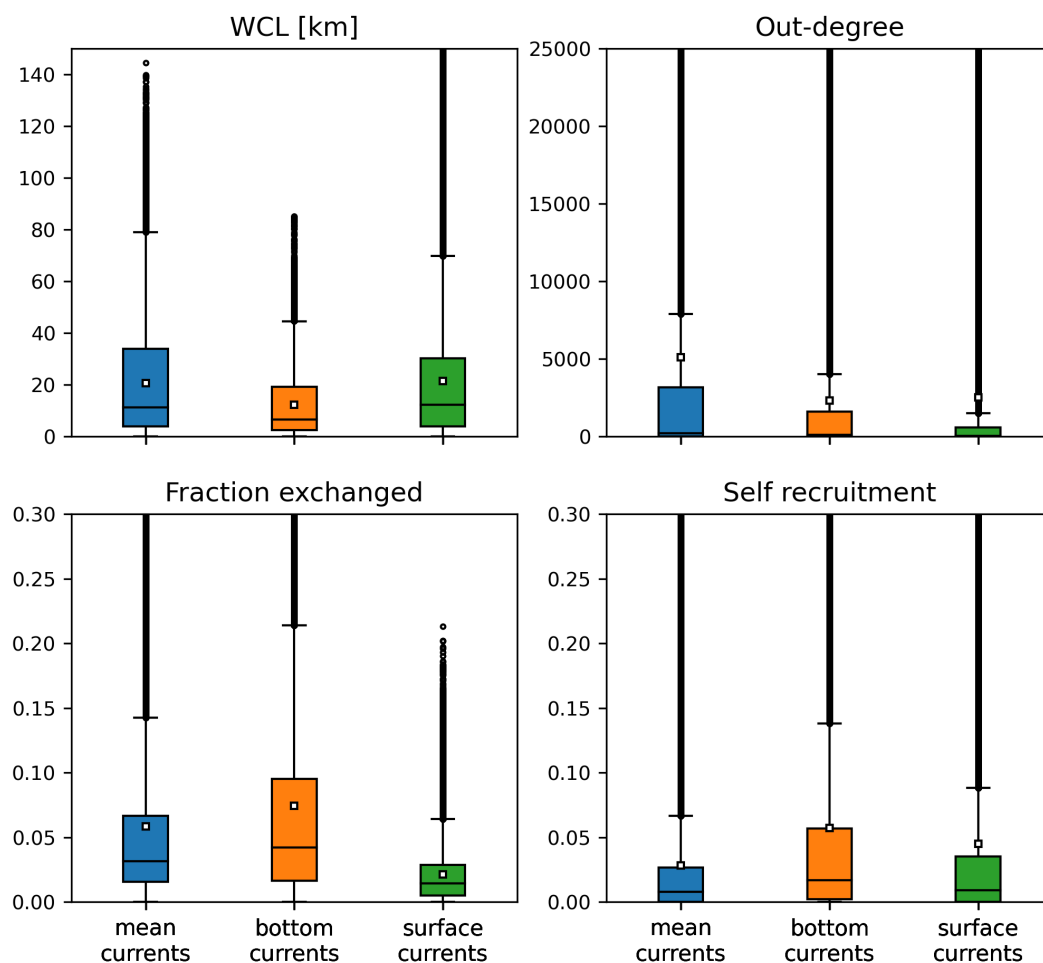
Mean and surface currents, on the other hand show similar spreading ranges with mean WCL of 20.63 km and 21.39 km respectively. However, the infectious matters that surface currents transport have the weakest probability to successfully settle on reefs. Consequently, surface currents and bottom currents produce networks with similar mean out-degree, although surface currents have the potential to transport infectious matter on more reefs. Nonetheless, networks have larger median out-degree with bottom currents than with surface currents, which suggests that surface currents have a lower spreading potential than bottom currents. As a result of their large WCL and fraction exchanged, barotropic currents on the other hand exhibit the largest mean out-degree, which suggest that they have a strong dispersal potential.

Self recruitment gives the proportion of infectious material settling on a reef that was produced on the same reef. The greater its values, the more the reef is isolated from the rest of the network. Since infectious material is less likely to settle on isolated reefs, self recruitment measures the probability for the disease to settle on a given reef, whereas all three other indicators inform on the reef spreading potential. Fig. 4 shows that the disease is more likely to settle on the reefs of networks generated by mean currents. This result is consistent with the values of the other connectivity measures, as reefs tend to be more strongly connected with mean currents. On the other hand, reefs are more isolated with bottom currents, as they produce the graphs with lowest WCL and out-degree. Finally, surface currents generate larger self recruitment values than mean currents as they exhibit the lowest fraction exchanged.

#### 3.2 Epidemiological model results

Best fit to averaged disease prevalence observations is obtained with transmission rate  $\beta_0^{-1} = 6.45$  days and removal rate  $\sigma^{-1} = 6.99$  days. Comparison of the evolution of the state described by Eqs 4 results with observations is shown in figure 5. Our model results accurately reproduce the observed fraction of susceptible individuals on colonies through time. However, the modeled fraction of removed individuals overestimates observations by about 5%. These discrepancies might be explained by the presence of "Unknown" values in our data sets as well as the simplifying assumptions of SIR models. Transmission and removal rates are found to have fairly close values with basic  $\beta_0/\sigma$  being equal to 1.0345. This proximity in rate values is imposed by Eq. 5, as aggregated observations show a proportion of susceptible individuals of about 85% at the end of the outbreak. Since infection and removal occur at very close rates, the instantaneous proportion of infectious individuals on the reefs remains pretty low through the outbreak, with a maximum value of about 0.4%.

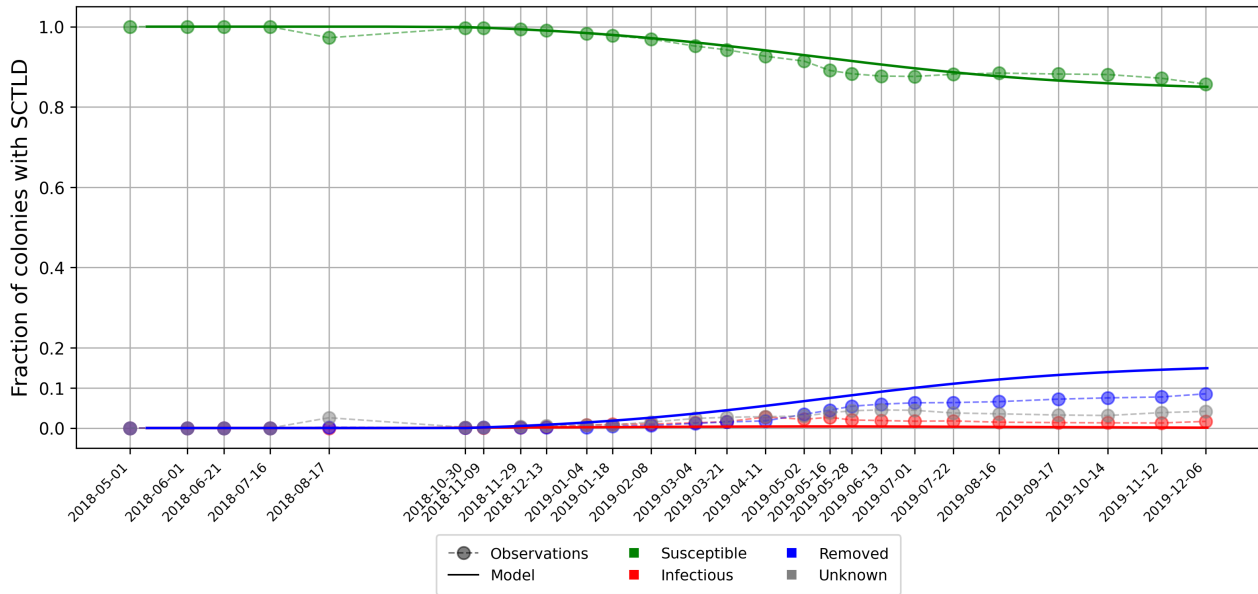
Using the above calibrated epidemiological parameters, epidemiological model simulations were performed from 1st May 2018 to 1st April 2019 for each type of currents and different values of the infection threshold  $I_0$ . A summary of the simulation results are shown in Fig. 6. Two metrics are used to assess the accuracy of the model. First, the modeled front speed is compared to the reference rate of 92 m/day derived by Muller et al. (2020). Furthermore, we computed the mean of the distances between each point where the SCTL has been observed during our simulated period and the centroid of the closest reef



**Figure 4.** Distribution of the indicators derived from the monthly connectivity matrices computed for each type of current during our simulated period. Mean values are indicated by white squares

304 polygon predicted to be infected by our model during the same period. Bottom currents produce the slowest  
 305 modeled disease propagation with a maximum front speed of  $\sim 20$  m/day, while simulations performed  
 306 with surface currents spread the disease at a maximum speed of of about 60 m/day. However, surface  
 307 currents tend to propagate the disease to the north, rather than westward, along the Florida Keys. This  
 308 explains why bottom currents predict infection closer to observations despite exhibiting slower front speed.  
 309 Finally, Mean barotropic currents outperform other types of current regarding both criteria with a front  
 310 speed of 107 m/day and a mean geographical accuracy of  $\sim 1.2$  km. This suggest that the causative agent  
 311 of the disease might be transported within neutrally buoyant material driven from reef to reef inside the  
 312 water column by mean currents.

313 Moreover, Fig. 6 shows a strong dependence of model results to infection threshold  $I_0$ , that gives  
 314 the proportion of infectious individual that colonies can withstand before exponential disease growth is  
 315 triggered on reef. Front speeds of both mean and bottom currents reach a plateau for values of infection  
 316 threshold between  $I_0 = 0.05\%$  and  $I_0 = 0.1\%$ , while the minimal prediction error is attained around  
 317  $I_0 \approx 0.078\%$  with mean currents. For  $I_0 > 0.1\%$ , intra-reef infection is strongly impeded and populations  
 318 of infectious individuals on infected reefs are not able to become sufficiently large to infect other colonies  
 319 on reefs they are connected to. For values of  $I_0$  lower than  $0.05\%$  on the other hand, intra-reef infection  
 320 dominates and coral population on infected reefs is removed too fast to efficiently spread the disease



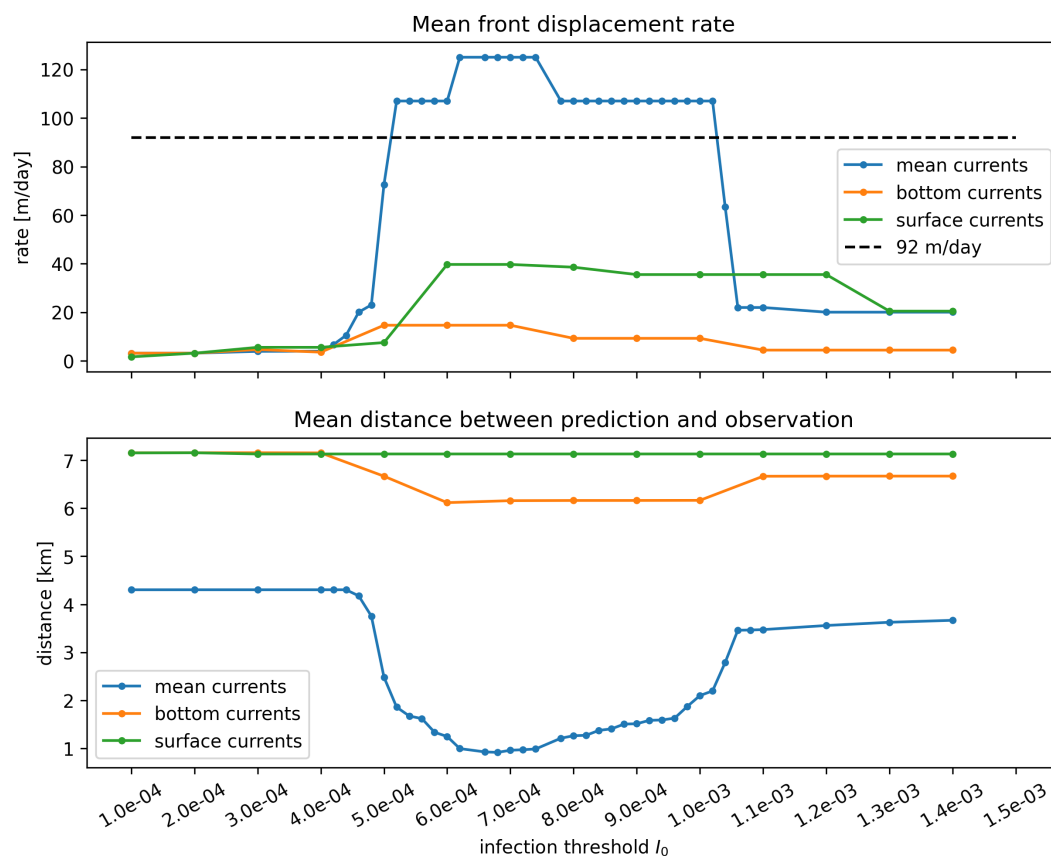
**Figure 5.** Disease prevalence averaged over all monitored sites over time as modeled by Eqs. 4 using calibrated transmission and removal parameters  $\beta_0^{-1} = 6.45$  days and  $\sigma^{-1} = 6.99$  days.

through the network. Since disease propagation throughout the FRT only occurs for fairly small values of  $I_0$  in our model, corals are expected to have low resistance to the causative agent of the SCTL D.

The results shown in Fig. 6 were obtained by removing the large reef located North to Vaca key, denoted Vaca reef in Frys et al. (2020), from our reef polygons. Preliminary simulations showed that this reef has close to no impact on the modeled spread of the disease to the rest of the FRT, as it sends very little infectious material to southerly and easterly neighboring reefs. Moreover, Vaca reef has a low coral coverage (0 – 10%), which strongly impedes disease spread on the reef. However, as coral coverage is not taken into account in our epidemiological model, propagation of the disease on the reef was overestimated. This led to unrealistically strong modeled front speed variations due to the large size of the reef. Consequently, and in the absence of SCTL D observations on Vaca reef, it has been removed from our reef polygons in order to avoid overestimating the front speed.

## 4 DISCUSSION AND CONCLUSIONS

We have developed an epidemio-hydrodynamic model to simulate the spread of the SCTL D through the entire FRT. Calibrating our model with colonies-averaged prevalence observations, we estimate the species-averaged reproduction number  $R_0$  to be barely larger than one. Our model simulations suggest that only the barotropic currents are able to reproduce the observed spread of the disease. Bottom current do not spread infectious material far enough while surface currents do not allow infectious material to spend enough time over the reefs to strongly infect them. The causative agent of the SCTL D is therefore expected to be transported within neutrally buoyant particles inside the water column. With this mode of transport, the propagation of the disease from reef to reef only occurs for a well-defined range of values of the infection threshold  $I_0$ . This threshold is defined as the proportion of colonies that have to be infected to trigger a rapid spread of the disease over the entire reef. Our result suggest that this occurs as soon as 0.05 – 0.1% of colonies are infected. On average, corals are thus expected to have low resistance to the SCTL D.



**Figure 6.** Summary of epidemiological model simulations with calibrated transmission parameters. **Top:** Modeled disease front speed for each type of current with respect to intra-reef infection threshold  $I_0$ . **Bottom:** Mean distance between predicted infected reefs and observed disease points. These results show that mean barotropic currents outperform other modes of transport at reproducing the observed spread of the disease. The appearance of a plateau suggests that model predictions are very sensitive to the value of the infection threshold  $I_0$ .

After calibration, we estimated the species-averaged basic reproduction number  $\beta'_0/\sigma$  to be equal to 1.0835. This value being close to 1, modeled infectious individuals are removed from the system almost as fast as susceptible individuals get infected. This causes the proportion of infectious corals on the reefs to remain pretty low (*i.e.*  $\leq 0.4\%$ ) through time. This suggests that only a small fraction of the colony causes the disease to spread on the reef during the outbreak. The observation-based species-averaged transmission period of 6.45 days used in this model seems to be a reasonable estimation of the disease transmission dynamics as it is of the same order of magnitude as the experimentally-derived mean incubation period of 9.7 days. The difference between the two values can be explained by field measurement uncertainties as well as the inability to perfectly mimic field conditions in laboratory. In this study, the same values were used for inter- and intra-reef rates  $\beta$  and  $\beta'_0$ . This implies that the infectiousness of the causative agent is not reduced during its journey from reef to reef. However, to assess the impact of this assumption, epidemiological model simulations were performed with  $\beta = \beta'_0/2$ . The resulting disease front speeds did not exceed 20 m/day. This strong decrease can be explained by the interplay between inter- and intra-reef infection. Reducing inter-reef transmission rates decreases the proportion of infectious corals on reefs attained by infectious materials, which in turn reduces the amount of infectious matter sent to the rest of the network. This suggests that, to reproduce the observed spread, inter- and intra-reef transmission rates



360 must have similar magnitude, *i.e.* that the causative agent is almost not degraded while traveling from reef  
361 to reef. **Is this consistent with Rhodobacterales and Rhizobiales ?**

362 The fact that mean barotropic currents outperform the other modes of transport can be explained by  
363 considering the trajectories of the particles used to model the transport of the vector of the disease causative  
364 agent. Due to the impact of winds on positively buoyant materials, particles driven by surface currents  
365 are likely to be blown away from the reefs. Moreover, even when winds are pushing particles along the  
366 reef line, these particles spend less time over the same region than particles driven by mean and bottom  
367 currents. Smaller amounts of particle mass will therefore settle on reef polygons, leading to lower entries  
368 of the potential connectivity matrix, *i.e.* lower exchange of infectious material between reefs. Hence,  
369 despite being able to transport the disease over greater distances, surface currents are less likely to drive  
370 the propagation of the disease. Particles driven by bottom currents, on the other hand, remain longer over  
371 the same region, producing larger entries of the potential connectivity matrix. Due to these large exchange  
372 probabilities between reefs, bottom currents are better at propagating the disease (Fig. 6). Nevertheless,  
373 bottom currents being relatively slower, exchanges of infected materials occur on a limited geographical  
374 range. Mean barotropic currents, that carry particles on greater distances while allowing for sufficiently  
375 large amounts of infected mass to settle on reef polygons, are thus best suited to propagate the disease (Fig.  
376 6).

377 Since mean currents are the only mode of transport that successfully reproduces the observed propagation  
378 speed of the disease in our model, the disease causative agent is expected to be transported within neutrally  
379 buoyant material inside the water column. Current-driven propagation seems reasonable as water-borne  
380 transmission is implicated as an important spreading mechanism for multiple coral diseases, including  
381 white band disease, white plague disease, white pox disease, white syndrome disease, *Porites* ulcerative  
382 white spots diseases, skeletal eroding band disease (Shore and Caldwell, 2019). **[examples of neutrally**  
383 **buoyant vectors ? mucus ?]**. The causative agent might also be transported within fine sediments such  
384 as silt, as suggested by Rosales et al. (2020). Such sediments are easily eroded in shallow areas around  
385 coral reefs and would therefore be mostly transported inside the water column by mean barotropic currents.  
386 This hypothesis might be tested by adapting the deposition rate  $\gamma$  used in our experiments to be consistent  
387 with the sedimentation rate of silt. However, such modification of  $\gamma$  would alter the entries of our potential  
388 connectivity matrices. Nonetheless, the sensitivity of the connectivity matrices to the value of  $\gamma$  has been  
389 briefly assessed by generating new matrices using particles with a half-life of 15 days ( $\gamma$  increased by a  
390 factor two). Although these matrices exhibited stronger short-range connectivity, the impact on connectivity  
391 indicator values remained limited ( $< 10\%$ ). This suggests that the main results of this study would remain  
392 valid for fine sediments.

393 Coral resistance to the SCTL outbreak is represented by parameter  $I_0$ , defined as the maximum proportion  
394 of the colony that can get infected without causing the disease to spread to the rest of the colony. The  
395 plateau shown in Fig. 6 highlights the impact of this parameter on the modeled propagation of the disease.  
396 On the one hand, when corals are strongly susceptible to the disease, infectious individuals are removed  
397 from the system too fast to become sustainable sources of infectious materials in the network. On the  
398 other hand, if corals are weakly susceptible to the disease, very few corals get infected and the disease  
399 barely propagates. Our simulations suggest that this value must be fairly low (around 0.01%) in order to  
400 successfully spread the disease throughout the FRT. **This seems to imply that susceptible coral species**  
401 **have very weak defense mechanisms against the causative agent of the disease. (?)**.

402

As with any modeling study, it is important to understand the assumptions on which the model is based. Here, we have used a 2D barotropic ocean model coupled with the 3D model HYCOM (Chassignet et al., 2007) in order to indirectly represent baroclinic phenomena. Such model is well suited to simulate the fate of neutrally-buoyant matters in shallow regions. However, as depth-averaged currents do not accurately approximate the motion of particles in the bottom and surface layers, SLIM velocities have therefore been modified to simulate the exchanges of negatively and positively buoyant matters. Surface current response to wind parametrization is based on the results of Ardhuin et al. (2009), consistent with observations. In this study, measured surface currents are shown to be in the order of 1.0% – 1.8% of the wind speed, in a direction 10° – 40° to the right of the wind. [discuss bottom currents parametrization]. Although such estimation of surface and bottom currents is disputable, using a 2D model allows for reef-scale resolution throughout the whole FRT. Such high-resolution allows to capture recirculation eddies around islands and reefs, that significantly impact the weighted connectivity length as well as the local retention on the reefs.

The appearance of an interval of optimal values of threshold  $I_0$  for the propagation of the disease in our results highlights the impact of coral resistance on the spread of SCTLD through the FRT. Therefore, a further step in our modeling approach would be further dividing coral populations of our polygons into highly susceptible (e.g. *Dichocoenia stokesii*, *Meandrina meandrites*), intermediately susceptible (e.g. *Orbicella faveolata*, *Montastrea cavernosa*), and weakly susceptible (e.g. *Acropora Palmata*, *Acropora cervicornis*) sub-populations. The proportions susceptible, infectious and removed individuals within these sub-populations would then be modeled with specific transmission ( $\beta$ ,  $\beta'_0$ ) and removal ( $\sigma$ ) rates as well as specific infection thresholds  $I_0$ . Such approach would however require a fine knowledge of the distribution of the different coral species throughout the FRT. This knowledge about coral coverage could also be used to avoid overestimation of the front propagation, as in the case of Vaca reef.

Despite the limitations of its current formulation, we believe that our model brings unprecedented perspectives on the propagation mechanism of the SCTLD through the FRT. Using a reef-scale spatial resolution, we determined the most probable mode of transport for the vector of the disease agent and deduced its species-averaged reproduction number based on prevalence observations. Besides, our model formulation provides a framework to quantify coral resistance to the disease. As our model results are continuous through time, they can exhibit the variability of the propagation of the SCTLD through time and therefore bring additional insight to observation data. This study therefore provides much-needed complementary insight on the identification of the causative agent of the Stony Coral Tissue Loss disease and the management of the crisis it generates. *Say that our approach could be applied to other areas as well where there is still time to do active management of the disease.*

## CONFLICT OF INTEREST STATEMENT

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## AUTHOR CONTRIBUTIONS

## FUNDING

This paper is a result of research funded by the Florida Department of Environmental Protection under award XXX to Mote Marine Laboratory.

## ACKNOWLEDGMENTS

439 Computational resources were provided by the Consortium des Équipements de Calcul Intensif (CÉCI),  
440 funded by the F.R.S.-FNRS under Grant No. 2.5020.11.

## REFERENCES

- 441 Aeby, G., Ushijima, B., Campbell, J. E., Jones, S., Williams, G., Meyer, J. L., et al. (2019). Pathogenesis of  
442 a tissue loss disease affecting multiple species of corals along the florida reef tract. *Frontiers in Marine*  
443 *Science* 6, 678
- 444 Alvarez-Filip, L., Estrada-Saldívar, N., Pérez-Cervantes, E., Molina-Hernández, A., and González-Barrios,  
445 F. J. (2019). A rapid spread of the stony coral tissue loss disease outbreak in the Mexican Caribbean.  
446 *PeerJ* 7, e8069
- 447 Ardhuin, F., Marié, L., Rasche, N., Forget, P., and Roland, A. (2009). Observation and estimation of  
448 Lagrangian, Stokes, and Eulerian currents induced by wind and waves at the sea surface. *Journal of*  
449 *Physical Oceanography* 39, 2820–2838
- 450 Aronson, R. B. and Precht, W. F. (2001). White-band disease and the changing face of Caribbean coral  
451 reefs. In *The ecology and etiology of newly emerging marine diseases* (Springer). 25–38
- 452 Aronson, R. B. and Precht, W. F. (2006). Conservation, precaution, and Caribbean reefs. *Coral reefs* 25,  
453 441–450
- 454 Bidegain, G., Powell, E., Klinck, J., Ben-Horin, T., and Hofmann, E. (2016a). Microparasitic disease  
455 dynamics in benthic suspension feeders: infective dose, non-focal hosts, and particle diffusion. *Ecological*  
456 *modelling* 328, 44–61
- 457 Bidegain, G., Powell, E. N., Klinck, J. M., Ben-Horin, T., and Hofmann, E. E. (2016b). Marine infectious  
458 disease dynamics and outbreak thresholds: contact transmission, pandemic infection, and the potential  
459 role of filter feeders. *Ecosphere* 7, e01286
- 460 Blondeau, J., Brandt, M., Donovan, C., Eakin, M., Edwards, K., Edwards, K., et al. (2020). Coral reef  
461 condition: A status report for the US Virgin Islands
- 462 Brandt, M. E. and McManus, J. W. (2009). Dynamics and impact of the coral disease white plague: insights  
463 from a simulation model. *Diseases of aquatic organisms* 87, 117–133
- 464 Brauer, F. (2008). Compartmental models in epidemiology. In *Mathematical epidemiology* (Springer).  
465 19–79
- 466 Burgess, S. C., Kingsford, M. J., and Black, K. P. (2007). Influence of tidal eddies and wind on the  
467 distribution of presettlement fishes around One Tree Island, Great Barrier Reef. *Marine Ecology*  
468 *Progress Series* 341, 233–242
- 469 Chassignet, E. P., Hurlburt, H. E., Smedstad, O. M., Halliwell, G. R., Hogan, P. J., Wallcraft, A. J., et al.  
470 (2007). The HYCOM (hybrid coordinate ocean model) data assimilative system. *Journal of Marine*  
471 *Systems* 65, 60–83
- 472 Dustan, P. and Halas, J. C. (1987). Changes in the reef-coral community of Carysfort Reef, Key Largo,  
473 Florida: 1974 to 1982. *Coral Reefs* 6, 91–106
- 474 Figueiredo, J., Baird, A. H., and Connolly, S. R. (2013). Synthesizing larval competence dynamics and  
475 reef-scale retention reveals a high potential for self-recruitment in corals. *Ecology* 94, 650–659
- 476 Frys, F., Saint-Amand, A., Le Hénaff, M., Figueiredo, J., Kuba, A., Walker, B., et al. (2020). Fine-scale  
477 coral connectivity pathways in the Florida Reef Tract: Implications for conservation and restoration.  
478 *Frontiers in Marine Science* in press. doi:10.3389/fmars.2020.00312
- 479 FWC, F. (2017). Unified reef map v2. 0

- Gardner, T. A., Côté, I. M., Gill, J. A., Grant, A., and Watkinson, A. R. (2003). Long-term region-wide declines in Caribbean corals. *science* 301, 958–960
- Geuzaine, C. and Remacle, J.-F. (2009). Gmsh: A 3-d finite element mesh generator with built-in pre-and post-processing facilities. *International journal for numerical methods in engineering* 79, 1309–1331
- Harvell, D., Jordán-Dahlgren, E., Merkel, S., Rosenberg, E., Raymundo, L., Smith, G., et al. (2007). Coral disease, environmental drivers, and the balance between coral and microbial associates. *Oceanography* 20, 172–195
- Keeling, M. and Rohani, P. (2007). Stochastic dynamics. *Modeling Infectious Diseases in Humans and Animals*, 190–230
- Kourafalou, V. H. and Kang, H. (2012). Florida current meandering and evolution of cyclonic eddies along the Florida Keys Reef Tract: Are they interconnected? *Journal of Geophysical Research: Oceans* 117
- Kuffner, I. B., Lidz, B. H., Hudson, J. H., and Anderson, J. S. (2015). A century of ocean warming on florida keys coral reefs: historic in situ observations. *Estuaries and Coasts* 38, 1085–1096
- Kuta, K. and Richardson, L. (1996). Abundance and distribution of black band disease on coral reefs in the northern Florida Keys. *Coral reefs* 15, 219–223
- Lambrechts, J., Hanert, E., Deleersnijder, E., Bernard, P.-E., Legat, V., Remacle, J.-F., et al. (2008). A multi-scale model of the hydrodynamics of the whole Great Barrier Reef. *Estuarine, Coastal and Shelf Science* 79, 143–151
- Manzello, D. P. (2015). Rapid recent warming of coral reefs in the Florida Keys. *Scientific reports* 5, 16762
- Miller, J., Muller, E., Rogers, C., Waara, R., Atkinson, A., Whelan, K., et al. (2009). Coral disease following massive bleaching in 2005 causes 60% decline in coral cover on reefs in the US Virgin Islands. *Coral Reefs* 28, 925
- Muller, E. M., Sartor, C., Alcaraz, N. I., and van Woesik, R. (2020). Spatial epidemiology of the Stony-Coral-Tissue-Loss Disease in Florida. *Frontiers in Marine Science* 7, 163
- Murray, J. D. (2007). *Mathematical biology: I. An introduction*, vol. 17 (Springer Science & Business Media)
- Neely, K. (2018). Surveying the Florida Keys southern coral disease boundary. *Florida DEP. Miami, FL*, 1–15
- NOAA (2018). *Stony Coral Tissue Loss Disease Case Definition*. Available online at: <https://nmsfloridakeys.blob.core.windows.net/floridakeys-prod/media/docs/20181002-stony-coral-tissue-loss-disease-case-definition.pdf> (accessed June 4, 2020)
- Porter, J. W. and Meier, O. W. (1992). Quantification of loss and change in floridian reef coral populations. *American Zoologist* 32, 625–640
- Precht, W. F., Gintert, B. E., Robbart, M. L., Fura, R., and Van Woesik, R. (2016). Unprecedented disease-related coral mortality in southeastern florida. *Scientific Reports* 6, 1–11
- Richardson, L. L., Goldberg, W. M., Kuta, K. G., Aronson, R. B., Smith, G. W., Ritchie, K. B., et al. (1998). Florida's mystery coral-killer identified. *Nature* 392, 557–558
- Rosales, S. M., Clark, A. S., Huebner, L. K., Ruzicka, R. R., and Muller, E. (2020). Rhodobacterales and Rhizobiales are associated with stony coral tissue loss disease and its suspected sources of transmission. *Frontiers in Microbiology* 11, 681
- Ruzicka, R., Colella, M., Porter, J., Morrison, J., Kidney, J., Brinkhuis, V., et al. (2013). Temporal changes in benthic assemblages on florida keys reefs 11 years after the 1997/1998 el niño. *Marine Ecology Progress Series* 489, 125–141

- Shore, A. and Caldwell, J. M. (2019). Modes of coral disease transmission: How do diseases spread between individuals and among populations? *Marine biology* 166, 45
- Sokolow, S. (2009). Effects of a changing climate on the dynamics of coral infectious disease: a review of the evidence. *Diseases of Aquatic Organisms* 87, 5–18
- Sponaugle, S., Paris, C., Walter, K., Kourafalou, V., and Alessandro, E. (2012). Observed and modeled larval settlement of a reef fish to the Florida Keys. *Marine Ecology Progress Series* 453, 201–212
- Sutherland, K. P., Porter, J. W., and Torres, C. (2004). Disease and immunity in Caribbean and Indo-Pacific zooxanthellate corals. *Marine Ecology Progress Series* 266, 273–302
- Team, A. B. R. (2005). Atlantic Acropora status review document. *Report to National Marine Fisheries Service*, 152
- Thomas, C. J., Bridge, T. C., Figueiredo, J., Deleersnijder, E., and Hanert, E. (2015). Connectivity between submerged and near-sea-surface coral reefs: Can submerged reef populations act as refuges? *Diversity and Distributions* 21, 1254–1266
- Thomas, C. J., Lambrechts, J., Wolanski, E., Traag, V. A., Blondel, V. D., Deleersnijder, E., et al. (2014). Numerical modelling and graph theory tools to study ecological connectivity in the Great Barrier Reef. *Ecological Modelling* 272, 160–174
- Vaz, A. C., Paris, C. B., Olascoaga, M. J., Kourafalou, V. H., Kang, H., and Reed, J. K. (2016). The perfect storm: match-mismatch of bio-physical events drives larval reef fish connectivity between Pulley Ridge mesophotic reef and the Florida Keys. *Continental Shelf Research* 125, 136–146
- Viehman, S., Gittings, S., Groves, S., Moore, J., Moore, T., and Stein, J. (2018). NCCOS Assessment: Coral Disturbance Response Monitoring (DRM) Along the Florida Reef Tract Following Hurricane Irma From 2017-10-09 to 2017- 10-18 (NCEI Accession 0179071). *NOAA National Centers for Environmental Information*. Silver Spring, MD: National Centers for Coastal Ocean Science. Available online at: <https://doi.org/10.25921/sscd-6h41>
- Wolanski, E. (1994). *Physical oceanographic processes of the Great Barrier Reef* (CRC Press)

## TABLES AND FIGURES