

Canadian Guidelines for Hereditary Transthyretin Amyloidosis Polyneuropathy Management

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ABSTRACT: Hereditary transthyretin-mediated (hATTR) amyloidosis is a progressive disease caused by mutations in the TTR gene leading to multisystem organ dysfunction. Pathogenic TTR aggregation, misfolding, and fibrillization lead to deposition of amyloid in multiple body organs and frequently involve the peripheral nerve system and the heart. Common neurologic manifestations include: sensorimotor polyneuropathy (PN), autonomic neuropathy, small-fiber PN, and carpal tunnel syndrome. Many patients have significant progression due to diagnostic delays as hATTR PN is not considered within the differential diagnosis. Recently, two effective novel disease-modifying therapies, inotersen and patisiran, were approved by Health Canada for the treatment of hATTR PN. Early diagnosis is crucial for the timely introduction of these disease-modifying treatments that reduce impairments, improve quality of life, and extend survival. In this guideline, we aim to improve awareness and outcomes of hATTR PN by making recommendations directed to the diagnosis, monitoring, and treatment in Canada.

RÉSUMÉ : Lignes directrices sur la prise en charge de l'amylose héréditaire à transthyréline, accompagnée de polyneuropathie, au Canada. L'amylose héréditaire à transthyréline (ATTRh) est une maladie évolutive, causée par des mutations du gène de la transthyréline (TTR), qui entraînent un dysfonctionnement plurisystémique. L'agrégation, le mauvais repliement et la fibrillisation pathogènes de la TTR aboutissent au dépôt de protéines amyloïdes dans plusieurs organes, et affectent souvent le système nerveux périphérique et le cœur. Les troubles neurologiques fréquents comprennent une polyneuropathie sensorimotrice (PN), une neuropathie autonome, une polyneuropathie des petites fibres et le syndrome du canal carpien. Chez bon nombre de patients, la maladie a connu une évolution importante en raison de la pose tardive du diagnostic, la PN-ATTRh ne faisant pas l'objet d'un diagnostic différentiel. Santé Canada a approuvé, depuis peu, deux nouveaux médicaments modificateurs de la PN-ATTRh et efficaces contre l'affection, soit l'inotersen et le patisiran. La pose précoce du diagnostic revêt une importance cruciale dans l'instauration, en temps opportun, de ces tout nouveaux traitements qui atténuent les troubles, améliorent la qualité de vie et prolongent la survie. Les auteurs, par l'élaboration de la nouvelle ligne directrice, espèrent sensibiliser la communauté médicale à la PN-ATTRh, et améliorer les résultats cliniques qui y sont associés, en formulant des recommandations sur le diagnostic et le traitement de la maladie au Canada ainsi que sur la surveillance de son évolution.

Keywords: hATTR, Amyloidosis, Treatment, Diagnosis, Polyneuropathy

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INTRODUCTION

Transthyretin-mediated hereditary amyloidosis (hATTR amyloidosis or ATTR_v/variant amyloidosis) is a progressive disease caused by mutations in the TTR gene, which leads to accumulation of misfolded protein aggregates in multiple organs and tissues.^{1–4} Common disease phenotypes resulting from the deposition of amyloid fibrils include sensorimotor

polyneuropathy (PN), cardiomyopathy (CM), and gastrointestinal symptoms.^{5–8}

Recently, two effective novel disease-modifying therapies for hATTR PN, inotersen and patisiran, have emerged, making diagnosis of this hereditary neuropathy crucial for the early introduction of treatment, improvement in quality of life and survival. In this guideline, we aim to improve awareness

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of hATTR amyloidosis PN and also to make recommendations about the diagnosis, monitoring, and treatment in Canada.

BACKGROUND

Amyloid protein deposition is the common mechanism for the progressive organ damage in human amyloidosis and can result either from hereditary mutations (such as TTR, gelsolin, β_2 microglobulin, apolipoprotein gene mutations, and others) or from the misfolding of wild-type proteins into amyloidogenic fragments in the extracellular space.^{4,9} Both systemic and localized forms have been described, and 36 proteins have already been characterized as amyloidogenic in humans.^{1,9} Currently, the classification of amyloidosis depends on the precursor protein, and the prefix A is added to an abbreviation describing the specific precursor protein type involved (e.g. L/H for immunoglobulin light/heavy chains, A for serum amyloid A, TTR for transthyretin, and so forth).^{1,9}

AL and hATTR amyloidosis account for most cases of systemic amyloidosis, although 14 protein types have been associated with systemic disease.¹⁰ AL amyloidosis results from proliferation of clonal plasma cells, and the amyloid fibrils that originate from the immunoglobulin light chain monoclonal protein can affect the heart, kidney, and peripheral nervous system.^{11,12} In AA amyloidosis, organ damage results from accumulation of serum amyloid A protein, an acute-phase reactant, that is found in chronic inflammatory disorders, infections or hereditary autoinflammatory syndromes,^{12,13} although it almost never affects the peripheral nervous system.

TTR is a homotetrameric (four identical subunits) serum protein responsible for the transport of thyroxine and retinol-binding protein complex, synthesized in the liver, choroid plexus, and in the retinal and ciliary pigment epithelia.⁴ TTR aggregation after tetramer dissociation, misfolding, and fibrillization is the basic mechanism of TTR amyloidosis, and most mutations produce a destabilization of the folded state, which is less stable than the wild-type TTR (wtATTR).^{4,14}

More than 140 mutations in the TTR gene have been identified so far, and most of them are pathogenic.^{15,16} Clinical manifestations are partially determined by the specific TTR mutation, but phenotype–genotype correlations are not always precise, as the inheritance pattern, penetrance, amyloid fibril composition, and geographical distribution can influence presentation.^{17,18} HATTR amyloidosis is considered endemic in some countries, with an estimated global prevalence of 10,186 persons (range: 5526–38,468), and has been reported in more than 30 countries.^{4,19,20} In high-prevalence countries like Portugal, prevalence can range from 1 in 1000 to 1 in 10,000 people, being highly related to specific regions.^{21,22} In Canada, we not only extrapolate figures between 0.5 and 1.5 per 1,000,000 as a non-endemic country but also may find significant regional differences, depending on immigration patterns.^{19,23}

In patients presenting with the classic length-dependent sensorimotor small-fiber neuropathy phenotype and dysautonomia, usually progressing to a large-fiber neuropathy, the most frequent mutation is Val30Met.^{24–26} This mutation is present in early (<50 years old) and late (>50 years old) onset forms of hATTR amyloidosis PN in high-prevalence regions, such as Portugal, Japan, and Brazil.^{4,17,27} In other countries, this mutation is also

commonly identified in isolated families and small clusters, which might also be the case in some areas in Canada.^{4,17,28}

In North America, Val122Ile is the most common TTR mutation, commonly associated with a CM phenotype and affecting older men of African-Caribbean descent, whereas neurologic manifestations are less common.^{8,29} On the other hand, Val122Ile has also been described in Caucasians with small/large-fiber sensory neuropathy, autonomic neuropathy, and carpal tunnel syndrome, and sometimes as isolated features, without CM.^{8,30,31} Late-onset forms of Val30Met have also been associated with cardiac abnormalities, which can be the presenting feature.^{6,27} Significant variability exists in penetrance according to the genotype; therefore, early screening to detect asymptomatic mutation carriers is important not only for genetic counseling but also for monitoring patients who need early intervention.

Distinguishing acquired from hATTR amyloidosis is paramount, as the proper diagnosis will direct further investigations and therapy. Common overlapping phenotypes, such as CM and peripheral neuropathy, can occur in both forms. wtATTR is an age-related disorder most common in older men and predominantly causes CM and heart failure.³² Common neurologic manifestations include carpal tunnel syndrome, which is frequently bilateral and may precede cardiac manifestations by years, along with spinal stenosis. Mild peripheral neuropathy has also been described in wtATTR patients.³³

Regardless of the hATTR amyloidosis subtype, many patients have significant progression before the diagnosis can be confirmed, as clinical manifestations can mimic other common diseases such as diabetic neuropathy, chronic inflammatory neuropathies, and other more prevalent cardiac diseases. Outside endemic regions, there is a substantial challenge in the clinical diagnosis of hATTR amyloidosis, which can result in delayed introduction of disease-modifying therapies.^{7,19}

Clinical Presentation of hATTR Amyloidosis PN

A length-dependent, sensorimotor, axonal PN is the most common neurological manifestation of hATTR amyloidosis. The onset is typically that of a predominantly small diameter PN, with painful and numb feet that progresses to lower extremity weakness, imbalance, and autonomic dysfunction,^{27,34} although there is significant clinical heterogeneity that can delay diagnosis. Amyloid depositions in peripheral nerves may be focal, frequently involve bilateral median nerves, and carpal tunnel syndrome may predate a more diffuse neuropathy by years.^{24,35} Late-onset patients may not develop symptoms until the fifth or sixth decade, and consequently, distinguishing hATTR amyloidosis PN from more common causes of neuropathy may be challenging.

The clinician must be astute in recognizing signs and symptoms accompanying PN, which, even if present in small measure, should prompt consideration of a diagnosis of hATTR amyloidosis. Suspicion for both hereditary and AL amyloidosis should remain high in any patient with neuropathy and neuropathic pain without other clearly identifiable cause and in those with evidence of autonomic dysfunction.¹⁵ It is noteworthy that autonomic failure may be subclinical in the early stages and patients may not be forthcoming about specific symptoms such as erectile dysfunction and gastrointestinal manifestations.

Phenotypes of neuropathy beyond the classic length-dependent description are not infrequent, may be mutation-specific, and

highlight the protean nature of this disease.³⁶ These include a pure autonomic neuropathy, a motor-predominant neuropathy mimicking motor neuron disease, pure small-fiber neuropathy, an upper extremity-predominant PN, and a diffuse small- and large-fiber mixed axonal and demyelinating neuropathy with areflexia mimicking chronic inflammatory demyelinating polyradiculoneuropathy (CIDP).³⁷⁻³⁹

Late-onset phenotypes tend to have a more rapid and fulminant clinical course than those presenting prior to age 50, with an average life span of 4.7 years from time of diagnosis.^{27,40} CNS manifestations such as dementia, ataxia, seizures, and cerebrovascular disease have been described with certain *TTR* mutations. However, these manifestations are uncommon and attributed to end-stage disease.¹⁵ There is often an overlapping, mixed phenotype with co-existing hATTR PN and CM; with some mutations, a CM predominates over neurological manifestations.⁸

When to Consider Screening

Accurate and timely identification of hATTR PN is paramount, as currently approved therapies in Canada aim to prevent the progression of an otherwise fatal disease.¹⁵ A genetic diagnosis also has practical implications regarding cardiac monitoring and genetic counseling.³⁹ Late-onset presentations, absence of reliable family history, co-morbid presence of diabetes or a monoclonal gammopathy, demyelinating features on electrodiagnostic studies, and elevations in CSF protein or serum creatine kinase levels may all result in diagnostic uncertainty.^{37,38}

Non-specific manifestations of fatigue and weight loss may be the presenting symptoms and may mimic acquired causes of PN such as primary systemic amyloidosis or peripheral nerve vasculitis. Non-endemic regions have demonstrated greater genetic heterogeneity, which may also translate to phenotypic heterogeneity and variations in clinical course.²⁷ As Canada would be considered a non-endemic region, it must be on alert for non-classic presentations of PN such as ‘atypical CIDP’. Neuropathic pain, small-fiber sensory loss proximal to the wrists, prominence of axonal features on electrodiagnostic studies, and autonomic dysfunction are not features of classic CIDP and should prompt consideration of an alternate diagnosis. Active perusal of ‘red-flag’ signs and symptoms increases the likelihood of prompt hATTR PN disease recognition (Table 1) which is modified from a recent expert paper by Adams et al.²⁷ Screening for hATTR may be considered in patients with unexplained painful peripheral neuropathy without any of the above red flag symptoms. A summary of recommendations for screening is provided in Table 2.

CLINICAL PRESENTATION OF hATTR CARDIAC AMYLOIDOSIS

Cardiac involvement is recognized as an important clinical manifestation of hATTR amyloidosis and AL amyloidosis, and in addition to neurologic disease represents the other predominant organ system involved for ATTR. Certain hATTR genotypes are associated with a predominantly cardiac phenotype. Examples include the Val122Ile mutation, which is highly prevalent among individuals of African-Caribbean descent (although with variable penetrance) and is the most common cause of hATTR CM in North America^{29,41} and the Thr60Ala mutation found in those of Appalachian and Irish descent.²⁹

Table 1. Red flags in axonal sensorimotor polyneuropathy

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1. Rapid progression
 2. Family history of hATTR
 3. Bilateral Carpal Tunnel Syndrome
 4. Autonomic involvement (orthostatic hypotension, erectile dysfunction, post-prandial nausea/bloating)
 5. Gastrointestinal symptoms (diarrhea, constipation)
 6. Unexplained weight loss (>5 kg)
 7. Cardiac disease (Cardiac hypertrophy or arrhythmia)
 8. Kidney disease (proteinuria, renal failure)
 9. Ocular changes (vitreous opacities, glaucoma)
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Heart failure is the most common cardiac manifestation of cardiac amyloidosis, presenting with both left heart (dyspnea, orthopnea, and paroxysmal nocturnal dyspnea) and right heart (edema, ascites, hepatomegaly, early satiety, abdominal bloating, and severe fatigue) signs and symptoms.⁸ Conduction system disease such as bradycardia and heart block and atrial fibrillation are also common. Hypotension, orthostatic lightheadedness, and syncope may also occur, typically reflecting a combination of both heart failure and autonomic dysfunction.

DIAGNOSTIC WORKUP AND PATIENT ASSESSMENT IN hATTR AMYLOIDOSIS

Paraclinical Tests

Large Fibers

There are no electrodiagnostic hallmarks of hATTR amyloidosis PN *per se*, as the pathology of large-fiber involvement may be predominately axon loss, demyelination, or a mix thereof. Axonal sensorimotor PN is the classical finding on nerve conduction studies (NCS).^{42,43} Demyelination is a less common presentation of hATTR amyloidosis PN compared to an axonal pattern. However, patients with late-onset presentations in particular can have a demyelinating neuropathy⁴⁴⁻⁴⁸ which may meet European Federation of Neurological Sciences/Peripheral Nerve Society criteria for CIDP.^{37,38}

The major limitation of standard NCS and electromyography is that they do not assess the function of small nerve fibers, either somatic or autonomic, which are commonly the first fibers involved.⁴⁹ Another limitation of electrodiagnostic studies is that there are anatomical and technical factors that limit the assessment of proximal nerve segments. Nonetheless, due to the wide availability, standardization, and reproducibility, NCS forms an essential component of the initial workup of patients with suspected hATTR amyloidosis PN, even in patients who are asymptomatic, for baseline studies. Furthermore, as focal neuropathies, including carpal tunnel syndrome, can be a presentation of hATTR amyloidosis, NCS can be performed in symptomatic limbs to confirm these suspected peripheral localizations or other unusual sites of focal involvement.

Emerging evidence suggests that magnetic resonance neurography (MRN) is sensitive to injury in proximal and distal lower extremity nerves in carriers of mutant *TTR* as well as patients with known hATTR amyloidosis PN.^{50,51} Similarly, peripheral nerve ultrasonography may identify changes suggestive of nerve injury

Table 2. Summary of recommendations

Screening	<ul style="list-style-type: none"> We recommend screening for hATTR amyloidosis in patients with idiopathic, peripheral neuropathy with one or more of the following features: (1) rapid progression, (2) weight loss, (3) bilateral carpal tunnel syndrome, (4) autonomic neuropathy, (5) unexplained heart failure and/or cardiac arrhythmias, (8) family history of neuropathy, cardiomyopathy/carpal tunnel syndrome, (8) gastrointestinal symptoms (9) vitreous opacities or (10) any unexplained renal insufficiency or nephrotic syndrome. Screening for hATTR may be considered in patients with unexplained painful peripheral neuropathy without any of the above red flag symptoms. All patients with peripheral neuropathy and an immediate family history of hATTR should be offered <i>TTR</i> gene sequencing.
Diagnostic workup and patient assessment	<ul style="list-style-type: none"> All patients with suspected polyneuropathy should undergo NCS and electromyography to confirm and characterize large-fiber nerve involvement. Skin punch biopsy may be performed to assess small-fiber involvement through intraepidermal nerve fiber density in cases where the clinical presentation is not convincing. Other quantitative tests of small-fiber function may complement the diagnostic workup, but there is insufficient evidence to recommend any one technique as the gold standard, and more than one non-invasive test is required to confirm small-fiber dysfunction. Cardiologist referral is recommended to assess patients for cardiac amyloidosis. Screening tests include 12-lead EKG, chest X-ray, echocardiography and serum troponin and BNP or NTproBNP.
Confirming amyloid transthyretin	<ul style="list-style-type: none"> Genetic testing of the <i>TTR</i> gene is suggested in patients where there is any clinical suspicion, suggestive symptoms and positive family history or where there is a positive family history. Predictive testing in asymptomatic family members of hATTR patients may be offered with careful genetic counseling. Confirmation of amyloid deposition via biopsy is not required when the patient has a compatible clinical syndrome and known pathogenic variant in <i>TTR</i>. Skin punch biopsy may be performed to search for amyloid deposition, especially if assessment of small-fiber involvement is required, as described above. Invasive biopsies (cardiac, kidney) should be avoided due to the procedural risks, sensitivity, and availability of genetic testing. Nerve biopsies are not routinely advised in suspected patients for diagnostic purposes due to morbidity and false negative results.
Follow-up and monitoring	<ul style="list-style-type: none"> Longitudinal follow-up of peripheral neuropathy should be done every 6–12 months in patients with hATTR PN. Monitoring should include screening of sensory and autonomic symptoms, staging of PN, comprehensive neuromuscular examination, nerve conduction studies, and if available, tests of small-fiber and autonomic function. Screening for cardiac and renal involvement should be done routinely in patients with hATTR PN. Monitoring of pre-symptomatic <i>TTR</i> mutation carriers should be done annually for 10 years prior to PADO.
Treatment	<ul style="list-style-type: none"> Inotersen and patisiran are recommended as first-line therapy to stop progression of neuropathy and improve PN in early and late stage hATTR PN. A multidisciplinary approach to patients with mixed phenotypes (PN and cardiac disease) is necessary to optimize therapy and to decide first-line treatments. Given the significant peri-operative complications and requirement for lifelong immunosuppression, orthotopic liver transplants should no longer be used as first-line therapy. Conventional management of CTS, with early release surgery, is recommended in hATTR PN patients.

BNP, B-type natriuretic peptide; CTS, carpal tunnel syndrome; NTproBNP, N-terminal proBNP; PADO, predicted age of onset of symptomatic disease; PN, polyneuropathy.

in patients with hATTR amyloidosis, including increased cross-sectional areas of proximal and distal nerve sites.^{52,53}

Small Fibers

In the case of equivocal clinical evidence of autonomic and/or somatic small-fiber involvement, confirmatory testing is required. Skin punch biopsy has become standardized, and the technique of removing 3 mm punches from 10 cm above the lateral malleolus is supported by multiple neurology associations.⁵⁴ Skin biopsy offers simultaneous assessment of autonomic and somatic neurites. The most frequently reported biomarker of skin denervation is intraepidermal nerve fiber density, for which a level less than the fifth percentile confirms small-fiber neuropathy. Intraepidermal nerve fiber density correlates with the severity of neuropathy in hATTR amyloidosis PN^{55–57} and may also be abnormal in the pre-symptomatic stage.⁵⁸ Other findings that can indicate

dysfunction of the small fibers on skin punch biopsy include axonal swellings, abnormal nerve fiber orientation, very fine caliber axons, and excessive or complex nerve fiber branching.⁵⁹

Non-invasive tests for assessment of the small fibers have also emerged in addition to skin biopsy. Of these, measuring orthostatic blood pressure adds no cost, takes minimal time, and is highly standardized in order to detect the 40%–60% of patients with hATTR amyloidosis with PN who experience orthostatic hypotension.⁶⁰ Other tests include quantitative sudomotor axon reflex testing,⁶¹ axon-reflex-flare testing,⁶² sympathetic skin responses,^{63,64} electrochemical skin conductance,⁶⁵ laser Doppler flare imaging studies,⁶⁴ quantitative sensory thresholds,^{61,64} and in-vivo corneal confocal microscopy.⁶⁶ All have shown promise in small studies in hATTR amyloidosis, except laser Doppler imaging, but it was evaluated only in a small study.⁶⁷ At present, no single test has emerged as superior to any other and availability is limited.

Cardiovascular Assessment

It is recommended that patients with known or suspected ATTR or AL amyloidosis undergo routine screening for cardiac involvement by specialist referral, in particular patients with PN and undiagnosed heart failure.⁸ A multidisciplinary approach is essential in comprehensive evaluation of hATTR patients.

Screening evaluation for cardiac amyloidosis includes 12-lead electrocardiogram, chest X-ray, echocardiography, and serum troponin and B-type natriuretic peptide (BNP) or N-terminal proBNP (NTproBNP). Sonographic features characteristic of cardiac amyloidosis include increased biventricular wall thickness with preserved chamber size, biatrial enlargement, thickened cardiac valves, and small pericardial effusion. Ventricular systolic function is often preserved, however may be reduced in late stages of disease.

Cardiac magnetic resonance imaging may be valuable as a more sensitive form of evaluation than echocardiography for some patients, particularly when assessment includes late gadolinium contrast enhancement imaging, which typically shows a diffuse subendocardial or transmural enhancement pattern.⁶⁸ However, cardiac MRI cannot differentiate amyloidosis subtype.⁶ Nuclear scintigraphy using 99mTechnetium pyrophosphate (99mTc-PYP) scanning has a high sensitivity and specificity for ATTR CM, once AL amyloidosis has been excluded.⁵ Increased myocardial uptake similar to or greater than bone by visual assessment, or a quantitative heart-to-contralateral lung uptake ratio (by region of interest analysis) ≥ 1.5 allows for non-invasive diagnosis when the clinical context is supportive, however cannot differentiate hATTR from wtATTR.⁶⁸

Routine genetic sequencing for patients with confirmed ATTR CM to determine disease subtype is recommended.⁸ Right ventricular endomyocardial biopsy remains the gold standard for diagnostic confirmation and is typically performed only at specialized centers in the setting of equivocal non-invasive evaluation or to provide tissue confirmation to support the diagnosis of AL amyloidosis. Mass spectrometry provides highly accurate disease subtyping of tissue biopsy samples when performed by experienced centers.⁶⁹

Patients with hATTR PN without known cardiac involvement may be at risk for developing CM over time, particularly those with a genotype known to be associated with CM. The optimal approach for monitoring such patients for possible development of ATTR CM is uncertain.⁷⁰ Development of heart failure or other signs and symptoms of ATTR CM should prompt further investigation for progression to cardiac involvement. A summary of recommendations for diagnostic workup and patient assessment is provided in Table 2.

Confirming Amyloid Transthyretin

Genetic Testing

Genetic testing for the *TTR* gene is foundational in the diagnosis of hATTR amyloidosis PN. The *TTR* gene found on chromosome 18q12.1 contains 4 exons, and it encodes a 127 amino acid protein.⁷¹ Sequence analysis of *TTR* identifies 99% of pathogenic variants, all of which are found in exons 2 to 4. Variants of *TTR* and other proteins known to cause familial amyloidosis are contained in an online registry with information

including, but not limited to, codon change and reported phenotype.⁷² Genetic tests are readily available and can be done on saliva or blood. Costs and provincial funding approval are a limitation in certain provinces where *TTR* DNA analysis is not publicly funded, but costs are predicted to continue to decrease over time. Pharmaceutical companies have sponsored next-generation DNA panels for hereditary neuropathies via the hATTR Compass program (Akcea) using Ambry Genetics and the Alnylam Act program with Invitae.^{73,74} These are presently available across Canada free of charge, in an effort to identify potentially eligible patients for new-targeted therapies.

Biopsy

Biopsy has historically been required to confirm amyloid deposition in body tissues because of incomplete penetrance of mutations. Nerve biopsy can detect large-fiber dysfunction by assessing axon loss and demyelination and can result in specific diagnosis of amyloidosis with fibrils showing characteristic birefringence. However, the test is less commonly performed currently in clinical practice, as it is invasive, has undesirable potential sequelae, such as infection, is largely confined to the sural nerve, and cannot be used for serial studies to follow the course of the neuropathy. Moreover, there is significant literature on false negatives with nerve biopsies given the patchy nature of amyloid deposition.^{17,51,75,76} In view of the sensitivity of *TTR* genetic testing for detecting pathogenic mutations, confirmation of amyloid deposition in peripheral nerve is not likely required in the context of a demonstrated pathogenic mutation and compatible clinical syndrome.

Skin punch biopsy allows direct observation of amyloid deposition in addition to assessment of small fibers, as described above. Other minimally invasive options for biopsy include abdominal fat pad aspiration,⁷⁷ which has been evaluated in systemic amyloidosis, but not necessarily in phenotypes with neuropathy, and minor salivary gland biopsy.⁷⁸ Invasive biopsy sites include endomyocardium and kidney that can be diagnostic, but these are not routinely recommended unless suspicion of these organs' involvement is high and non-invasive workup is equivocal, in view of the potential procedural risks to the patient, lack of widespread availability, and high sensitivity/specificity of the genetic testing. 99mTc-labeled PYP scans are a non-invasive method of identifying *TTR* in the heart and are increasingly being used to avoid endomyocardial biopsies. In this regard, the absence of amyloid deposition on biopsy should not rule out a diagnosis of hATTR amyloidosis in the context of a compatible clinical syndrome and pathogenic *TTR* mutation.

In summary, in view of the sensitivity of *TTR* genetic testing for detecting pathogenic mutations, confirmation of amyloid deposition via biopsy is not generally required in the context of a demonstrated pathogenic mutation and compatible clinical syndrome. A summary of recommendations for confirming hATTR amyloidosis is provided in Table 2.

FOLLOW-UP AND MONITORING

With the advent of increasing disease-modifying treatment options, it has become essential to follow hATTR amyloidosis PN patients longitudinally and quantify objectively disease progression, in order to guide treatment choices.

Peripheral neuropathy should be characterized at the time of diagnosis and every 6–12 months, depending on disease genotype, phenotype, and evolution. Both somatic and autonomic components should be assessed. Screening for both positive and negative sensory dysfunction (including pain), quality of life, and activity limitations at each visit can be done by history or through patient-reported outcome questionnaires, some of them already validated in the amyloid population.^{79,80}

Comprehensive neurological examination documenting motor, sensory, and reflex dysfunction is critical. PN staging should also be recorded as a gross measure of disease severity. Specifically, in stage 1, patients can walk without assistance and have mild bilateral neuropathy in the lower limbs; in stage 2, they require assistance to walk (crutches or sticks) as neuropathy progresses proximally and/or to upper limbs; and in stage 3, patients are wheelchair-bound or bedridden.^{81,82} Although the modified NIS + 7 (mNIS + 7) was used as the primary endpoint in recent clinical trials,⁸² this is not feasible for use in routine clinical practice due to the need for specialized autonomic assessment and the length of time for evaluation.

Yearly electrophysiologic evaluation with NCS is recommended. Objective measures of small nerve fiber function should be done if available. These include electrochemical skin conductance through sudoscan, sympathetic skin response, quantitative sudomotor axon-reflex, quantitative sensory testing, laser Doppler flare imaging, confocal microscopy of the corneal nerves, and in selected cases, intraepidermal nerve fiber density on skin biopsy. However, orthostatic blood pressure measurement is easily obtainable in the clinic and can partly assess cardiovascular autonomic function.

Non-neurological manifestations of the disease should also be screened routinely. A baseline cardiac assessment, including ECG/Holter, NTproBNP, troponin, echocardiogram (+/– cardiac MRI), and cardiology consultation, is recommended and should guide further routine follow-up. Yearly renal function measurement and ophthalmologic assessment are also recommended, although this would depend on the exact mutation.^{83,84}

For TTR mutation carriers without clinical manifestations, a targeted follow-up with regular monitoring has been proposed.⁸⁵ By establishing the predicted age of onset of symptomatic disease (PADO), which is based on the particular mutation, its' typical age of onset and the age of symptom onset in family members, a structured approach can be offered to individuals in pre-symptomatic stage.⁸⁵ As a general rule, we suggest annual monitoring for 10 years prior to the PADO, for neurologic, cardiac, and mixed phenotypes.

Deciding on what constitutes failure of treatment to motivate change of therapy has not been well defined. While natural history data exist about expected rate of progression for the various measures above,⁸⁶ these decisions are probably best made by an experienced multidisciplinary team, taking into account the different organs involved. A summary of recommendations for follow-up and monitoring is provided in Table 2.

TREATMENT

Overtime, efforts to develop treatments for hATTR have focused on disruption of different steps of the pathophysiologic cascade. This includes expression of mutated TTR, destabilization of TTR tetramers into amyloidogenic monomers, and deposition of amyloid fibrils (Figure 1).⁸⁷

Liver Transplant

This historical therapy is no longer considered to be the first-line treatment for hATTR PN. Given the predominant hepatic expression of TTR, liver transplant has been used in the past (Figure 1) to replace abnormal hATTR secretion with a donor's wtATTR. Large cohort studies have shown efficacy in both clinical progression⁸⁸ and survival^{89,90} in hATTR PN, although survival is less dramatic in individuals with non-Val30Met mutations.⁹¹ Other poor prognostic factors include advanced age, prolonged disease duration, and poor pretransplant nutritional and cardiac status.⁹² The main causes of death after liver transplant are infection related to long-term immunosuppression and cardiac failure.⁹¹ The latter appears to be due to ongoing evidence of cardiac amyloid deposition following liver transplant,⁹³ suggesting benefit of combined heart-liver transplant in select individuals.⁹⁴ Higher rates of cerebral amyloid angiopathy have also been observed in post-transplant patients, likely secondary to persistent perivascular and leptomeningeal amyloid deposition with evidence of TTR expression in the choroid plexus.^{95,96}

Given significant limitations, and as noted above, liver transplant is no longer considered a first-line therapy for hATTR PN. Some of those limitations include organ availability, high costs, requirement for lifelong immunosuppression, surgical morbidity, and the fact that it usually does not completely prevent amyloid deposition. At this time, no studies exist comparing newer genetic therapies to liver transplant. The role of using genetic therapy or a TTR stabilizer after liver transplant remains unclear, although several case series suggest possible benefit.^{97–99}

TTR Stabilizers

Directly targeting the important step of TTR tetramer destabilization into amyloidogenic monomers has led the way to the development and analysis of TTR tetramer stabilizers (Figure 1). The two best studied TTR stabilizers are Tafamidis¹⁰⁰ and Diflunisal,¹⁰¹ both of which bind to the thyroxine binding site of TTR.

Tafamidis

Tafamidis is an orally bioavailable (20 mg oral once daily) non-steroidal anti-inflammatory drug (NSAID) benzoxazole with high affinity for TTR, resulting in stabilization of the TTR tetramer.¹⁰⁰ The first pivotal randomized control trial comparing Tafamidis to placebo in 128 Val30Met ATTR PN patients failed to show treatment response, although efficacy was obtained in the intention-to-treat (ITT) individuals that completed the study per protocol, as well as in secondary endpoints.¹⁰² In non-Val30Met ATTR PN, an open-label 12-month single treatment-arm study of 21 subjects with eight different mutations showed progression of PN.¹⁰³ Tafamidis is not approved by Health Canada for hATTR PN.^{90,104,105}

Treatment of hATTR Cardiac Amyloidosis

Cardiologist referral for patients with known or suspected cardiac amyloidosis is recommended as part of a multidisciplinary approach to management of this complex systemic disease. At present, the TTR stabilizer tafamidis is approved in Canada for the treatment of ATTR CM as a study showed reduced mortality and cardiovascular hospitalizations among patients who did not have end-stage disease.¹⁰⁶ However, it is not widely available and only prescribed by

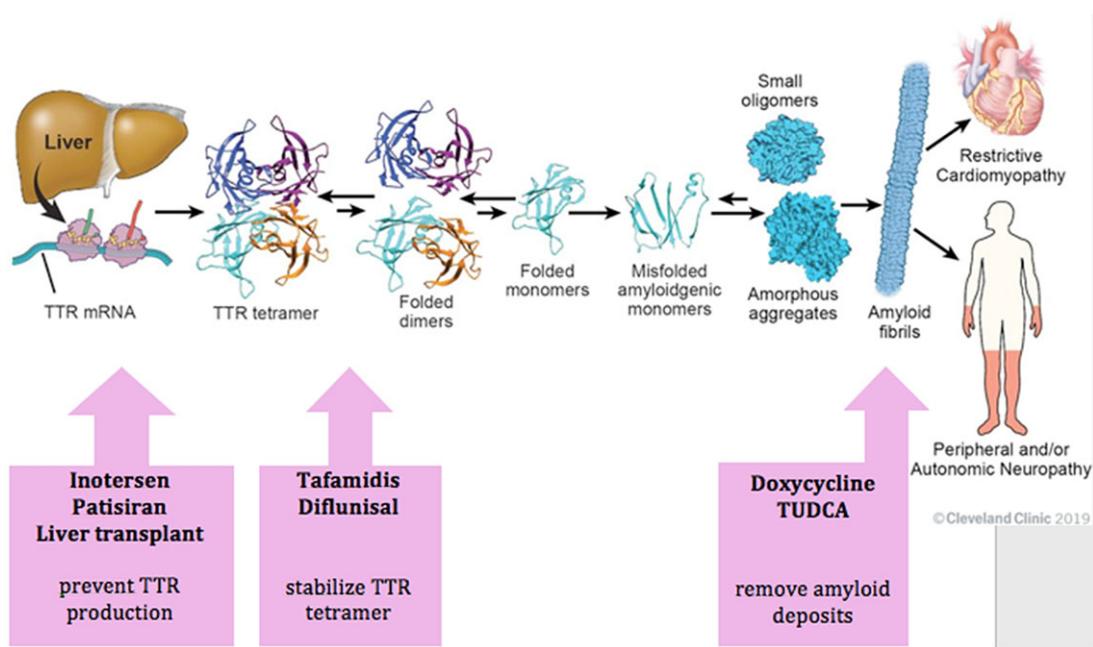


Figure 1. Disease modifying therapies of hATTR according to the pathophysiologic cascade. TTR, transthyretin. TUDCA, taurouodeoxycholic acid. Modified from Ruberg et al. 2019.⁽⁸⁷⁾

specialized centers in Canada. TTR silencer therapies presently approved for treatment of hATTR PN (inotuzumab, patisiran) have not been rigorously studied for hATTR CM; however, a subgroup analysis of the APOLLO trial demonstrated improved cardiac outcomes for patients with suspected cardiac involvement randomized to patisiran compared with placebo.¹⁰⁷

Diflunisal

Diflunisal (at a dose of 250 mg oral twice daily) significantly reduced disease progression in hATTR PN, although studies are limited by high dropout rates which may have biased the outcomes, liver transplantation, and adverse events.^{108–111} Being a NSAID, use of diflunisal is limited by adverse reactions such as gastrointestinal upset, renal dysfunction, and thrombocytopenia.^{109–111} Theoretical concerns regarding worsening of heart failure have also been raised, although recent studies suggest safety¹¹² and potential benefit¹¹³ in ATTR CM.

Diflunisal is relatively inexpensive, but not widely available in Canada and not approved for hATTR PN. Use in TTR PN has been limited due to lack of availability and/or supply in many provinces.

Emerging TTR Stabilizers

Some novel TTR stabilizers, such as Epigallocatechin-3-gallate, CHF5074, and AG10, have been evaluated in few retrospective, pre-clinical, and clinical trials with promising results, although clinical evidence of efficacy in hATTR PN favoring these agents remains to be determined at this time.^{114–118} The catechol-O-methyltransferase inhibitor and antiparkinsonian medication tolcapone have demonstrated TTR stabilization properties, and its availability in the central nervous system may provide benefit in ATTR patients with lepto-meningeal involvement.^{119,120}

Fibril Disruptors/Scavengers

Doxycycline and taurouodeoxycholic acid (TUDCA) have been shown to reduce fibrillar and non-fibrillar TTR tissue deposits in mouse models, respectively (Figure 1).¹²¹ Currently, there are no randomized controlled trials comparing Doxycycline/TUDCA to placebo in TTR PN.

RNA-Targeted Therapies (Current Treatment of Choice)

A broader understanding of the field of RNA processing has permitted the exploitation of various mechanisms of pre-mRNA processing and thus the means by which targeted synthetic oligonucleotides could be developed to inhibit TTR production at the RNA level (Figure 1). In this context, two broad classes of anti-TTR amyloid-specific antisense oligonucleotides (ASOs) have been brought to market that fulfill the related goals of drug delivery, target binding and digestion, and most importantly, patient safety.^{83,84} Two of these novel agents (inotuzumab and patisiran) are currently recommended as the treatments of choice for hATTR PN.

Briefly, ASOs are either single-stranded or double-stranded short (i.e., ‘oligo’) sequences of nucleotides (i.e., either RNA or a combination of RNA and DNA) which bind to specified mature cytoplasmic mRNA known as the sense strand (i.e., antisense). Inotuzumab is a type of single-stranded ASO with specific modifications to confer improved pharmacokinetics, binding, and potency.¹²² Upon binding to nuclear RNase H1, the mRNA–DNA adduct is degraded. By contrast, the double-stranded or duplex oligonucleotide ASOs are hydrophilic because they shield their lipophilic bases and thus require a delivery mechanism. Patisiran is therefore immured in a lipid nanoparticle to facilitate uptake by the hepatocytes. Double-stranded ASOs are also known as small-interfering RNA (siRNA) and utilize the RNA interference pathway to

Table 3. Comparison of recent RNA silencer studies in hATTR PN

	APOLLO (Patisiran)	NEURO-TTR (Inotersen)
Trial duration (weeks/months)	78/18	66/15
Allocation – D:P (n)	2:1 (225)	2:1 (172)
Dose & route of administration	0.3 mg/kg IV q3w	300 mg SC q1w*
Completed – D; P (%)	138/148 (93); 55/77 (71)	87/112 (78); 52/60 (87)
Deaths – D; P (%)	7 (5); 6 (8)	5 (4); 0 (0)
SAE – D; P (%)	54 (36); 31 (40)	36 (32); 13 (22)
Withdrawn – D; P (%)	7 (5); 9 (12)	25 (22); 8 (13)
mNIS + 7† baseline‡	80.9 ± 41.5; 74.6 ± 37.0	79.2 ± 37.0; 74.8 ± 39.0
r mNIS + 7 study exit‡	-6.0 ± 1.7; 28.0 ± 2.6	5.8§; 25.5§
r mNIS + 7 (D – P)	34	19.7
mNIS + 7 total score‡	304	346.3
mNIS + 7% improvement	11.2	5.7

D, drug; P, placebo.

*300 mg was loaded over each of 3 separate days in the first week.

†Lower score translates to improved clinical status and less neuropathy burden.

‡Reported in least-squares mean ± standard error (SE).

neutralize their targets via an RNA endonuclease, Argonaute 2. Therefore, both inotersen, a single-stranded RNA–DNA hybrid, and patisiran, a double-stranded siRNA, are distinct types of ASOs which exploit different pathways targeting the degradation of mRNA. These then represent novel mechanisms to downregulate the production of TTR protein, the root issue in the pathogenesis of hATTR.

Randomized Controlled Clinical Trials of mRNA Knockdown in hATTR

Two landmark clinical trials, APOLLO⁸³ and NEURO-TTR,⁸⁴ reported on the effects of patisiran and inotersen, respectively (Table 3). Briefly, both trials produced strong evidence for disease modification with, drug minus placebo, mNIS + 7 scores. The neurological score at study exit yielding differences of 34 points in APOLLO and 19.7 points in NEURO-TTR. Important differences between the two studies complicate the direct comparison of these primary endpoints given that the total points constituting the mNIS + 7 Alnylam and the mNIS + 7 Ionis were different (304 vs. 368.6, respectively).⁸² Additionally, NEURO-TTR was only 15 months compared to APOLLO, which was 18 months.

The open label extension (OLE) of NEURO-TTR collected data for an additional 104 weeks. The absolute values for mean changes in the mNIS + 7 scores were not reported but on interpolation from Table 3, it appears that the inotersen–inotersen group worsened by ~9–10 points on the mNIS + 7, while the placebo–inotersen arm worsened by ~7–8 points.¹²³ Compared to OLE baseline, 47% of placebo–inotersen patients showed improved mNIS + 7 scores at week 104. By contrast, only 24% of inotersen–inotersen patients showed improved mNIS + 7 scores from NEURO-TTR baseline to OLE week 104, suggesting that its beneficial effect is not as marked over time. Alnylam is continuing to collect data in their global OLE study (ClinicalTrials.gov Identifier: NCT02510261) with completion expected by mid-2022. A preliminary analysis suggests

patisiran–patisiran patients continued to produce a mean mNIS + 7 score at 12 months of less than the APOLLO baseline score which represents preservation of improved clinical status.¹²⁴

Although a direct comparison of the Phase III trials of patisiran and inotersen is limited by different study characteristics, both treatments improve or stabilize PN in long-term follow-up. Patisiran is administered intravenously and has a favorable safety profile. Inotersen is administered subcutaneously and requires close monitoring of platelets and renal function to manage the risk of severe thrombocytopenia and/or glomerulonephritis. Considerations on availability, route of administration and frequency of treatment, according to patient characteristics or formal contraindications influence the exact choice of this therapy.

Both inotersen and patisiran have been approved by Health Canada for use in hATTR PN. Future RNA knockdown therapeutics are currently in the pipeline and under investigation. Alnylam is conducting the HELIOS-A study (ClinicalTrials.gov Identifier: NCT03759379), a phase 3 global open-label study comparing intravenous patisiran to subcutaneous vutrisiran, and Ionis in partnership with Akcea is conducting NEURO-TTRtransform (ClinicalTrials.gov Identifier: NCT04136184), a phase 3 open-label trial comparing their novel ligand-conjugated antisense drug ION-682884/AKCEA-TTR-LRx against inotersen. These drugs are bound to N-acetylgalactosamine which facilitates hepatic uptake by the asialoglycoprotein receptor 1 theoretically permitting smaller dosages and longer intervals between dosing.¹²⁵ If the results of these trials are positive, this next generation of drugs will dramatically simplify the treatment of symptomatic patients with hATTR.

Carpal Tunnel Syndrome (CTS) Management

At this time, no comparison studies of management between hATTR-related CTS and idiopathic CTS have been performed and the risk of recurrent CTS in hATTR patients after decompression is unclear. A recent histopathologic study found amyloid deposition in

the flexor tenosynovium in 9 out of 35 patients with recurrent CTS (recurrence of symptoms 6 months following first carpal tunnel release).¹²⁶ Seven of the nine stained for TTR, although no evidence of systemic amyloidosis was found. This suggests that amyloidosis may be a risk factor for recurrent disease after CTS release, although this finding was not specific to TTR amyloidosis.

In contrast, long-term follow-up of 10 patients with synovial/ligamentous amyloid deposition, six of which stained for TTR (including two with hATTR), found symptom relief following surgery.¹²⁷ So, surgical release is likely still indicated in this patient population, although risk of recurrence may be increased. A summary of recommendations for treatment is provided in Table 2.

Wild-Type TTR

Mainly associated with CM, wtTTR rarely causes PN. Autonomic and sensorimotor nerve damages appear after years of disease if patients benefit from heart transplant.¹²⁸ However, peripheral neurological manifestations may present years prior to CM in the form of carpal tunnel or lumbar spinal stenosis.^{35,128–132}

CONCLUSIONS

Recent significant advances have been made in the diagnosis, follow-up, and treatment of patients with hATTR PN and CM. In countries with a low prevalence such as Canada, the diagnosis can be missed frequently due to the lack of family history and variable penetrance of different mutations. Additionally, the clinical phenotype can reflect other environmental factors unrelated to the TTR mutation.

Early diagnosis is essential for the introduction of disease-modifying therapies, which can potentially reduce impairments and slow down progression in this progressive and fatal disease.

Being a multisystemic disorder, with variable presentation, hATTR amyloidosis requires a combined, multidisciplinary approach. As it can affect many target organs such as the heart, gastrointestinal tract, kidney, liver, and nerves, a specialist team should be involved in patient care as soon as the diagnosis is confirmed. Early management of potential organ damage contributes to improved quality of life and reduced mortality.

This guideline highlights specific recommendations for diagnosis, monitoring, and treatment of patients diagnosed with hATTR PN. Best practices regarding the most effective therapies continue to evolve, including ASOs that have evidence of neuropathy disease modification in blinded and open label trials. Identification of potential candidates for novel therapies is paramount.

CONFLICTS OF INTEREST

Dr. Alcantara has nothing to disclose. Dr. Mezei reports other from Alnylam, other from Akcea, other from Pfizer, other from Alexion, other from CSL Behring, other from Stealth BioTherapeutics, outside the submitted work. Dr. Baker reports other from AKCEA Therapeutics Canada, other from CSL Behring, other from Alnylam Pharmaceuticals, other from Octapharma, grants and other from Grifols, outside the submitted work. Dr. Breiner reports grants from GBS/CIDP Foundation, grants from Grifols Inc, grants from Muscular Dystrophy Canada, personal fees from CSL Behring, personal fees from Allergan, personal fees from Mitsubishi Tanabe, personal fees from Akcea,

personal fees from Alexion, personal fees from Alnylam, outside the submitted work. Dr. Dhawan was co-investigator in two upcoming industry sponsored clinical trials studying treatments in patients with hereditary transthyretin amyloidosis PN (Ionis and Edios trials). Dr. Fiander reports financial relationships with Akcea and Alnylan. Dr. Fine reports grants and personal fees from Pfizer, grants and personal fees from Akcea, grants and personal fees from Alnylam, outside the submitted work. Dr. Hahn reports personal fees from Alnylam, personal fees from Akcea, outside the submitted work. Dr. Katzberg reports personal fees from UCB, personal fees from Octapharma, personal fees from Terumo, grants and personal fees from Takeda, personal fees from Akcea, personal fees from Alnylam, personal fees from CSL Behring, outside the submitted work. Dr. Khayambashi reports grants from Akcea Therapeutics, outside the submitted work. Dr. Rami reports grants and personal fees from Alnylam Pharmaceuticals, grants from Pfizer, outside the submitted work. Dr. Matte reports personal fees from Alnylam Canada ULC, personal fees from Akcea Therapeutics Canada, outside the submitted work. Dr. Putko has nothing to disclose. Dr. Siddiqi has nothing to disclose. Dr. Delgado reports personal fees from Pfizer, Akcea, and Alnylan. Dr. Bril reports personal fees from Alnylam, grants and personal fees from Akcea, grants and personal fees from Ionis, during the conduct of the study.

STATEMENT OF AUTHORSHIP

MA contributed with design, review of the literature, drafting, editing, and reviewing the manuscript; MM contributed with review of the literature, drafting, and reviewing the manuscript; SB contributed with review of the literature, drafting, and reviewing the manuscript; AB contributed with review of the literature, drafting, and reviewing the manuscript; PD contributed with review of the literature, drafting, and reviewing the manuscript; AF contributed with review of the literature, drafting, and reviewing the manuscript; NF contributed with review of the literature, drafting, and reviewing the manuscript; CH contributed with review of the literature, drafting, and reviewing the manuscript; HK contributed with review of the literature, drafting, and reviewing the manuscript; SK contributed with review of the literature, drafting, and reviewing the manuscript; RM contributed with review of the literature, drafting, and reviewing the manuscript; GM contributed with review of the literature, drafting, and reviewing the manuscript; BP contributed with review of the literature, drafting, and reviewing the manuscript; ZS contributed with review of the literature, drafting, and reviewing the manuscript; DD contributed with review of the literature, drafting, and reviewing the manuscript; VB contributed with concept, design, review of the literature, drafting, and critical review of the manuscript, editing, and approval of the final version.

REFERENCES

1. Benson MD, Buxbaum JN, Eisenberg DS, et al. Amyloid nomenclature 2018: recommendations by the International Society of Amyloidosis (ISA) nomenclature committee. *Amyloid*. 2018;25(4):215–9.
2. Sekijima Y. Transthyretin (ATTR) amyloidosis: clinical spectrum, molecular pathogenesis and disease-modifying treatments. *J Neurol Neurosurg Psychiatry*. 2015;86(9):1036–43.

3. Conceição I. Novel RNA-targeted therapies for hereditary ATTR amyloidosis and their impact on the autonomic nervous system. *Clin Auton Res.* 2019;29(Suppl 1):11–7.
4. Adams D, Koike H, Slama M, Coelho T. Hereditary transthyretin amyloidosis: a model of medical progress for a fatal disease. *Nat Rev Neurol.* 2019;15(7):387–404.
5. Yamamoto H, Yokochi T. Transthyretin cardiac amyloidosis: an update on diagnosis and treatment. *ESC Heart Fail.* 2019;6(6):1128–39.
6. Damy T, Maurer MS, Rapezzi C, et al. Clinical, ECG and echocardiographic clues to the diagnosis of TTR-related cardiomyopathy. *Open Heart.* 2016;3(1):e000289.
7. Kapoor M, Rossor AM, Laura M, Reilly MM. Clinical presentation, diagnosis and treatment of TTR amyloidosis. *J Neuromuscul Dis.* 2019;6(2):189–99.
8. Fine NM, Davis MK, Anderson K, et al. Canadian cardiovascular society/Canadian heart failure society joint position statement on the evaluation and management of patients with cardiac amyloidosis. *Can J Cardiol.* 2020;36(3):322–34.
9. Picken MM. The pathology of amyloidosis in classification: a review. *Acta Haematol.* 2020;143(4):322–34.
10. Gertz MA, Dispenzieri A. Systemic amyloidosis: recognition, prognosis, and therapy: a systematic review. *JAMA.* 2020;324(1):79–89.
11. Nuvolone M, Merlini G. Systemic amyloidosis: novel therapies and role of biomarkers. *Nephrol Dial Transplant.* 2017;32(5):770–80.
12. Merlini G, Dispenzieri A, Sanchorawala V, et al. Systemic immunoglobulin light chain amyloidosis. *Nat Rev Dis Primers.* 2018;4(1):38.
13. Brunger AF, Nienhuis HLA, Bijzet J, Hazenberg BPC. Causes of AA amyloidosis: a systematic review. *Amyloid.* 2020;27(1):1–12.
14. Chiti F, Dobson CM. Protein misfolding, amyloid formation, and human disease: a summary of progress over the last decade. *Annu Rev Biochem.* 2017;86:27–68.
15. Gertz MA, Mauerma ML, Grogan M, Coelho T. Advances in the treatment of hereditary transthyretin amyloidosis: a review. *Brain Behav.* 2019;9(9):e01371.
16. Mutations in hereditary amyloidosis; 2020. Available at: <http://amyloidismutations.com/mut-attr.php>; accessed November 19, 2020.
17. Plante-Bordeneuve V, Ferreira A, Lalu T, et al. Diagnostic pitfalls in sporadic transthyretin familial amyloid polyneuropathy (TTR-FAP). *Neurology.* 2007;69(7):693–8.
18. Plante-Bordeneuve V. Transthyretin familial amyloid polyneuropathy: an update. *J Neurol.* 2018;265(4):976–83.
19. Schmidt HH, Waddington-Cruz M, Botteman MF, et al. Estimating the global prevalence of transthyretin familial amyloid polyneuropathy. *Muscle Nerve.* 2018;57(5):829–37.
20. Waddington-Cruz M, Schmidt H, Botteman MF, et al. Epidemiological and clinical characteristics of symptomatic hereditary transthyretin amyloid polyneuropathy: a global case series. *Orphanet J Rare Dis.* 2019;14(1):34.
21. Inês M, Coelho T, Conceição I, Duarte-Ramos F, de Carvalho M, Costa J. Epidemiology of transthyretin familial amyloid polyneuropathy in Portugal: a nationwide study. *Neuroepidemiology.* 2018;51(3–4):177–82.
22. Parman Y, Adams D, Obici L, et al. Sixty years of transthyretin familial amyloid polyneuropathy (TTR-FAP) in Europe: where are we now? A European network approach to defining the epidemiology and management patterns for TTR-FAP. *Curr Opin Neurol.* 2016;29(Suppl 1):S3–13.
23. Namirian D, Chalk C, Massie R. Poor yield of routine transthyretin screening in patients with idiopathic neuropathy. *Can J Neurol Sci.* 2020;47(6):816–9.
24. Conceição I, González-Duarte A, Obici L, et al. “Red-flag” symptom clusters in transthyretin familial amyloid polyneuropathy. *J Peripher Nerv Syst.* 2016;21(1):5–9.
25. Coelho T, Maurer MS, Suhr OB. THAOS - The transthyretin amyloidosis outcomes survey: initial report on clinical manifestations in patients with hereditary and wild-type transthyretin amyloidosis. *Curr Med Res Opin.* 2013;29(1):63–76.
26. Waddington-Cruz M, Ackermann EJ, Polydefkis M, et al. Hereditary transthyretin amyloidosis: baseline characteristics of patients in the NEURO-TTR trial. *Amyloid.* 2018;25(3):180–8.
27. Adams D, Ando Y, Beirão JM, et al. Expert consensus recommendations to improve diagnosis of ATTR amyloidosis with polyneuropathy. *J Neurol.* 2020. doi: [10.1007/s00415-019-09688-0](https://doi.org/10.1007/s00415-019-09688-0).
28. Zhen DB, Swiecicki PL, Zeldenrust SR, Dispenzieri A, Mauerma ML, Gertz MA. Frequencies and geographic distributions of genetic mutations in transthyretin- and non-transthyretin-related familial amyloidosis. *Clin Genet.* 2015;88(4):396–400.
29. Maurer MS, Hanna M, Grogan M, et al. Genotype and phenotype of transthyretin cardiac amyloidosis: THAOS (transthyretin amyloid outcome survey). *J Am Coll Cardiol.* 2016;68(2):161–72.
30. Salhi H, Lefaucher JP, Gorram FG, et al. Phenotypic spectrum and management of 25 patients ATTR Val122Ile. *Neurology.* 2015;84(Suppl 14):P50732015.
31. Stancanelli C, Gentile L, Di Bella G, et al. Phenotypic variability of TTR Val122Ile mutation: a Caucasian patient with axonal neuropathy and normal heart. *Neurol Sci.* 2017;38(3):525–6.
32. Nativi-Nicolau J, Maurer MS. Amyloidosis cardiomyopathy: update in the diagnosis and treatment of the most common types. *Curr Opin Cardiol.* 2018;33(5):571–9.
33. Živković S, Soman P, Lacomis D. Late-onset peripheral neuropathy in patients with wild type transthyretin amyloidosis (wtATTR). *Amyloid.* 2020;27(2):142–3.
34. Carr AS, Pelayo-Negro AL, Evans MR, et al. A study of the neuropathy associated with transthyretin amyloidosis (ATTR) in the UK. *J Neurol Neurosurg Psychiatry.* 2016;87(6):620–7.
35. Nakagawa M, Sekijima Y, Yazaki M, et al. Carpal tunnel syndrome: a common initial symptom of systemic wild-type ATTR (ATTRwt) amyloidosis. *Amyloid.* 2016;23(1):58–63.
36. Luigetti M, Romano A, Di Paolantonio A, Bisogni G, Sabatelli M. Diagnosis and treatment of hereditary transthyretin amyloidosis (hATTR) polyneuropathy: current perspectives on improving patient care. *Ther Clin Risk Manag.* 2020;16:109–23.
37. Lozeron P, Mariani LL, Dodet P, et al. Transthyretin amyloid polyneuropathies mimicking a demyelinating polyneuropathy. *Neurology.* 2018;91(2):e143–52.
38. Ohashi N, Kodaira M, Morita H, Sekijima Y. Electrophysiological demyelinating features in hereditary ATTR amyloidosis. *Amyloid.* 2019;26(1):15–23.
39. Théaudin M, Lozeron P, Algalarrondo V, et al. Upper limb onset of hereditary transthyretin amyloidosis is common in non-endemic areas. *Eur J Neurol.* 2019;26(3):497.
40. Mazzeo A, Russo M, Di Bella G, et al. Transthyretin-related familial amyloid polyneuropathy (TTR-FAP): a single-center experience in Sicily, an Italian endemic area. *J Neuromuscul Dis.* 2015;2(Suppl 2):S39–48.
41. Quarta CC, Buxbaum JN, Shah AM, et al. The amyloidogenic V122I transthyretin variant in elderly black Americans. *N Engl J Med.* 2015;372(1):21–9.
42. Luis ML. Electroneurophysiological studies in familial amyloid polyneuropathy—Portuguese type. *J Neurol Neurosurg Psychiatry.* 1978;41(9):847–50.
43. Kodaira M, Morita H, Shimojima Y, Ikeda S. Electrophysiological features of familial amyloid polyneuropathy in endemic area. *Amyloid.* 2011;18(1):10–8.
44. Misu K, Hattori N, Nagamatsu M, et al. Late-onset familial amyloid polyneuropathy type I (transthyretin Met30-associated familial amyloid polyneuropathy) unrelated to endemic focus in Japan. Clinicopathological and genetic features. *Brain.* 1999;122(Pt 10):1951–62.
45. Briemberg HR, Amato AA. Transthyretin amyloidosis presenting with multifocal demyelinating mononeuropathies. *Muscle Nerve.* 2004;29(2):318–22.
46. Koike H, Kawagashira Y, Iijima M, et al. Electrophysiological features of late-onset transthyretin Met30 familial amyloid polyneuropathy unrelated to endemic foci. *J Neurol.* 2008;255(10):1526–33.
47. Mathis S, Magy L, Diallo L, Boukhris S, Vallat JM. Amyloid neuropathy mimicking chronic inflammatory demyelinating polyneuropathy. *Muscle Nerve.* 2012;45(1):26–31.
48. Mariani LL, Lozeron P, Theaudin M, et al. Genotype-phenotype correlation and course of transthyretin familial amyloid polyneuropathies in France. *Ann Neurol.* 2015;78(6):901–16.

49. Andrade C. A peculiar form of peripheral neuropathy; familiar atypical generalized amyloidosis with special involvement of the peripheral nerves. *Brain*. 1952;75(3):408–27.
50. Kollmer J, Hund E, Hornung B, et al. In vivo detection of nerve injury in familial amyloid polyneuropathy by magnetic resonance neurography. *Brain*. 2015;138(Pt 3):549–62.
51. Kollmer J, Sahm F, Hegenbart U, et al. Sural nerve injury in familial amyloid polyneuropathy: MR neurography vs clinicopathologic tools. *Neurology*. 2017;89(5):475–84.
52. Granata G, Luigetti M, Coraci D, et al. Ultrasound evaluation in transthyretin-related amyloid neuropathy. *Muscle Nerve*. 2014;50(3):372–6.
53. Podnar S, Sarafov S, Tournev I, Omejec G, Zidar J. Peripheral nerve ultrasonography in patients with transthyretin amyloidosis. *Clin Neurophysiol*. 2017;128(4):505–11.
54. Oaklander AL, Nolano M. Scientific advances in and clinical approaches to small-fiber polyneuropathy: a review. *JAMA Neurol*. 2019;76(10):1240–1251.
55. Yang NC, Lee MJ, Chao CC, et al. Clinical presentations and skin denervation in amyloid neuropathy due to transthyretin Ala97Ser. *Neurology*. 2010;75(6):532–8.
56. Ebenezer GJ, Liu Y, Judge DP, et al. Cutaneous nerve biomarkers in transthyretin familial amyloid polyneuropathy. *Ann Neurol*. 2017;82(1):44–56.
57. Chao CC, Hsueh HW, Kan HW, et al. Skin nerve pathology: Biomarkers of premanifest and manifest amyloid neuropathy. *Ann Neurol*. 2019;85(4):560–73.
58. Masuda T, Ueda M, Suenaga G, et al. Early skin denervation in hereditary and iatrogenic transthyretin amyloid neuropathy. *Neurology*. 2017;88(23):2192–7.
59. Freeman R, Chase KP, Risk MR. Quantitative sensory testing cannot differentiate simulated sensory loss from sensory neuropathy. *Neurology*. 2003;60(3):465–70.
60. Palma JA, Gonzalez-Duarte A, Kaufmann H. Orthostatic hypotension in hereditary transthyretin amyloidosis: epidemiology, diagnosis and management. *Clin Auton Res*. 2019;29(Suppl 1):33–44.
61. Kim DH, Zeldenrust SR, Low PA, Dyck PJ. Quantitative sensation and autonomic test abnormalities in transthyretin amyloidosis polyneuropathy. *Muscle Nerve*. 2009;40(3):363–70.
62. Calero-Romero I, Suter MR, Waeber B, Feihl F, Kuntzer T. Axon reflex-mediated vasodilation is reduced in proportion to disease severity in TTR-FAP. *Neurol Genet*. 2018;4(4):e251.
63. Conceicao IM, Castro JF, Scotto M, de Carvalho M. Neurophysiological markers in familial amyloid polyneuropathy patients: early changes. *Clin Neurophysiol*. 2008;119(5):1082–7.
64. Lefaucheur JP, Ng Wing Tin S, Kerschen P, Damy T, Plante-Bordeneuve V. Neurophysiological markers of small fibre neuropathy in TTR-FAP mutation carriers. *J Neurol*. 2013;260(6):1497–503.
65. Castro J, Miranda B, Castro I, de Carvalho M, Conceicao I. The diagnostic accuracy of Sudoscan in transthyretin familial amyloid polyneuropathy. *Clin Neurophysiol*. 2016;127(5):2222–7.
66. Rousseau A, Cauquil C, Dupas B, et al. Potential role of in vivo confocal microscopy for imaging corneal nerves in transthyretin familial amyloid polyneuropathy. *JAMA Ophthalmol*. 2016;134(9):983–9.
67. Zouari HG, Ng Wing Tin S, Wahab A, Damy T, Lefaucheur JP. Assessment of autonomic innervation of the foot in familial amyloid polyneuropathy. *Eur J Neurol*. 2019;26(1):94.
68. Dorbala S, Ando Y, Bokhari S, et al. ASNC/AHA/ASE/EANM/HFSA/ISA/SCMR/SNMMI expert consensus recommendations for multimodality imaging in cardiac amyloidosis: part 1 of 2-evidence base and standardized methods of imaging. *J Card Fail*. 2019;25(11):e1–39.
69. Vrana JA, Gamez JD, Madden BJ, Theis JD, Bergen HR, Dogan A. Classification of amyloidosis by laser microdissection and mass spectrometry-based proteomic analysis in clinical biopsy specimens. *Blood*. 2009;114(24):4957–9.
70. Adams D, Suhr OB, Hund E, et al. First European consensus for diagnosis, management, and treatment of transthyretin familial amyloid polyneuropathy. *Curr Opin Neurol*. 2016;29(Suppl 1):S14–26.
71. Sekijima Y. Hereditary transthyretin amyloidosis. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. *GeneReviews® [Internet]*. Seattle, WA: University of Washington; 1993–2021.
72. Rowczenio DM, Noor I, Gillmore JD, et al. Online registry for mutations in hereditary amyloidosis including nomenclature recommendations. *Hum Mutat*. 2014;35(9):E2403–12.
73. Invitae test requisition form. ALNYLAM ACT (hATTR AMYLOID-OYSIS TRF921-9). Available from https://view.publitas.com/invitae/trf921_invitae_alnylamact_A9Q686httr_order_form/page/1; accessed March 10, 2021.
74. Ambry Genetics test requisition form. hATTR Compass (hATTR – 120110). Available from https://www.ambrygen.com/file/material/view/1290/Akcea_TRF_0420_final%28004%29.pdf; accessed March 10, 2021.
75. Coimbra A, Andrade C. Familial amyloid polyneuropathy: an electron microscope study of the peripheral nerve in five cases. II. Nerve fibre changes. *Brain*. 1971;94(2):207–12.
76. Simmons Z, Blaivas M, Aguilera AJ, Feldman EL, Bromberg MB, Towfighi J. Low diagnostic yield of sural nerve biopsy in patients with peripheral neuropathy and primary amyloidosis. *J Neurol Sci*. 1993;120(1):60–3.
77. van G II, Hazenberg BP, Bijzet J, van Rijswijk MH. Diagnostic accuracy of subcutaneous abdominal fat tissue aspiration for detecting systemic amyloidosis and its utility in clinical practice. *Arthritis Rheum*. 2006;54(6):205–21.
78. de Paula Eduardo F, de Mello Bezinelli L, de Carvalho DL, et al. Minor salivary gland biopsy for the diagnosis of familial amyloid polyneuropathy. *Neurol Sci*. 2017;38(2):311–8.
79. Vinik EJ, Vinik AI, Paulson JF, et al. Norfolk QOL-DN: validation of a patient reported outcome measure in transthyretin familial amyloid polyneuropathy. *J Peripher Nerv Syst*. 2014;19(2):104–14.
80. Pruppers MH, Merkies IS, Faber CG, Da Silva AM, Costa V, Coelho T. The Val30Met familial amyloid polyneuropathy specific Rasch-built overall disability scale (FAP-RODS©). *J Peripher Nerv Syst*. 2015;20(3):319–27.
81. Coutinho P, Martins da Silva A, Lopes Lima J, Resende Barbosa A. Forty years of experience with type I amyloid neuropathy: review of 483 cases. In: Glenner GG, e Costa PP, de Freitas AF, editors. *Amyloid and Amyloidosis*. Amsterdam: Excerpta Medica; 1980, pp. 88–98.
82. Dyck PJB, González-Duarte A, Obici L, et al. Development of measures of polyneuropathy impairment in hATTR amyloidosis: From NIS to mNIS + 7. *J Neurol Sci*. 2019;405:116424.
83. Adams D, Gonzalez-Duarte A, O'Riordan WD, et al. Patisiran, an RNAi therapeutic, for hereditary transthyretin amyloidosis. *N Engl J Med*. 2018;379(1):11–21.
84. Benson MD, Waddington-Cruz M, Berk JL, et al. Inotersen treatment for patients with hereditary transthyretin amyloidosis. *N Engl J Med*. 2018;379(1):22–31.
85. Conceição I, Damy T, Romero M, et al. Early diagnosis of ATTR amyloidosis through targeted follow-up of identified carriers of TTR gene mutations. *Amyloid*. 2019;26(1):3–9.
86. Koike H, Tanaka F, Hashimoto R, et al. Natural history of transthyretin Val30Met familial amyloid polyneuropathy: analysis of late-onset cases from non-endemic areas. *J Neurol Neurosurg Psychiatry*. 2012;83(2):152–8.
87. Ruberg FL, Grogan M, Hanna M, Kelly JW, Maurer MS. Transthyretin amyloid cardiomyopathy: JACC state-of-the-art review. *J Am Coll Cardiol*. 2019;73(22):2872–91.
88. Okumura K, Yamashita T, Masuda T, et al. Long-term outcome of patients with hereditary transthyretin V30M amyloidosis with polyneuropathy after liver transplantation. *Amyloid*. 2016;23(1):39–45.
89. Yamashita T, Ando Y, Okamoto S, et al. Long-term survival after liver transplantation in patients with familial amyloid polyneuropathy. *Neurology*. 2012;78(9):637–43.
90. Coelho T, Inês M, Conceição I, Soares M, de Carvalho M, Costa J. Natural history and survival in stage 1 Val30Met transthyretin familial amyloid polyneuropathy. *Neurology*. 2018;91(21):e1999–2009.

91. Ericzon BG, Wilczek HE, Larsson M, et al. Liver transplantation for hereditary transthyretin amyloidosis: after 20 years still the best therapeutic alternative? *Transplantation*. 2015;99(9):1847–54.
92. Carvalho A, Rocha A, Lobato L. Liver transplantation in transthyretin amyloidosis: issues and challenges. *Liver Transpl*. 2015;21(3):282–92.
93. Yazaki M, Mitsuhashi S, Tokuda T, et al. Progressive wild-type transthyretin deposition after liver transplantation preferentially occurs onto myocardium in FAP patients. *Am J Transplant*. 2007;7(1):235–42.
94. Nelson LM, Penninga L, Sander K, et al. Long-term outcome in patients treated with combined heart and liver transplantation for familial amyloidotic cardiomyopathy. *Clin Transplant*. 2013;27(2):203–9.
95. Sekijima Y, Yazaki M, Oguchi K, et al. Cerebral amyloid angiopathy in posttransplant patients with hereditary ATTR amyloidosis. *Neurology*. 2016;87(8):773–81.
96. Mitsuhashi S, Yazaki M, Tokuda T, et al. Biochemical characteristics of variant transthyretins causing hereditary leptomeningeal amyloidosis. *Amyloid*. 2005;12(4):216–25.
97. Moshe-Lilie O, Dimitrova D, Heitner SB, et al. TTR gene silencing therapy in post liver transplant hereditary ATTR amyloidosis patients. *Amyloid*. 2020;27(4):250–3.
98. Matsushima M, Yabe I, Tsuda M, Sakakibara M, Shimamura T, Sasaki H. Amyloid polyneuropathy and myocardial amyloidosis 10 years after domino liver transplantation from a patient with a transthyretin Ser50Arg mutation. *Intern Med*. 2017;56(23):3231–5.
99. Kon T, Misumi Y, Nishijima H, et al. Effects of liver transplantation and tafamidis in hereditary transthyretin amyloidosis caused by transthyretin Leu55Pro mutation: a case report. *Amyloid*. 2015;22(3):203–4.
100. Coelho T, Merlini G, Bulawa CE, et al. Mechanism of action and clinical application of tafamidis in hereditary transthyretin amyloidosis. *Neurol Ther*. 2016;5(1):1–25.
101. Sekijima Y, Dendale MA, Kelly JW. Orally administered diflunisal stabilizes transthyretin against dissociation required for amyloidogenesis. *Amyloid*. 2006;13(4):236–49.
102. Coelho T, Maia LF, Martins da Silva A, et al. Tafamidis for transthyretin familial amyloid polyneuropathy: a randomized, controlled trial. *Neurology*. 2012;79(8):785–92.
103. Merlini G, Planté-Bordeneuve V, Judge DP, et al. Effects of tafamidis on transthyretin stabilization and clinical outcomes in patients with non-Val30Met transthyretin amyloidosis. *J Cardiovasc Transl Res*. 2013;6(6):1011–20.
104. Gundapaneni BK, Sultan MB, Keohane DJ, Schwartz JH. Tafamidis delays neurological progression comparably across Val30Met and non-Val30Met genotypes in transthyretin familial amyloid polyneuropathy. *Eur J Neurol*. 2018;25(3):464–8.
105. Mundayat R, Stewart M, Alvir J, et al. Positive effectiveness of tafamidis in delaying disease progression in transthyretin familial amyloid polyneuropathy up to 2 years: an analysis from the transthyretin amyloidosis outcomes survey (THAOS). *Neurol Ther*. 2018;7(1):87–101.
106. Mauret MS, Schwartz JH, Gundapaneni B, et al. Tafamidis treatment for patients with transthyretin amyloid cardiomyopathy. *N Engl J Med*. 2018;379(11):1007–16.
107. Solomon SD, Adams D, Kristen A, et al. Effects of Patisiran, an RNA interference therapeutic, on cardiac parameters in patients with hereditary transthyretin-mediated amyloidosis. *Circulation*. 2019;139(4):431–43.
108. Berk JL, Suhr OB, Obici L, et al. Repurposing diflunisal for familial amyloid polyneuropathy: a randomized clinical trial. *JAMA*. 2013;310(24):2658–67.
109. Sekijima Y, Tojo K, Morita H, Koyama J, Ikeda S. Safety and efficacy of long-term diflunisal administration in hereditary transthyretin (ATTR) amyloidosis. *Amyloid*. 2015;22(2):79–83.
110. Azorín SE, Cabib CE, Campistol JM. Diflunisal compassionate use in transthyretin hereditary amyloid polyneuropathy: report of a first Spanish experience. *Amyloid*. 2017;24(Suppl 1):105–6.
111. Wixner J, Westermark P, Ihse E, Pilebro B, Lundgren HE, Anan I. The Swedish open-label diflunisal trial (DFNS01) on hereditary transthyretin amyloidosis and the impact of amyloid fibril composition. *Amyloid*. 2019;26(Suppl 1):39–40.
112. Ikram A, Donnelly JP, Sperry BW, Samaras C, Valent J, Hanna M. Diflunisal tolerability in transthyretin cardiac amyloidosis: a single center's experience. *Amyloid*. 2018;25(3):197–202.
113. Lohrmann G, Pipilas A, Mussinelli R, et al. Stabilization of cardiac function with diflunisal in transthyretin (ATTR) cardiac amyloidosis. *J Card Fail*. 2019;26(9):753–9.
114. Ferreira N, Saraiwa MJ, Almeida MR. Epigallocatechin-3-gallate as a potential therapeutic drug for TTR-related amyloidosis: “in vivo” evidence from FAP mice models. *PLoS One*. 2012;7(1):e29933.
115. Cappelli F, Martone R, Tabacchi G, et al. Epigallocatechin-3-gallate tolerability and impact on survival in a cohort of patients with transthyretin-related cardiac amyloidosis. A single-center retrospective study. *Intern Emerg Med*. 2018;13(6):873–80.
116. Mu Y, Jin S, Shen J, et al. CHF5074 (CSP-1103) stabilizes human transthyretin in mice humanized at the transthyretin and retinol-binding protein loci. *FEBS Lett*. 2015;589(7):849–56.
117. Qiang L, Guan Y, Li X, et al. CSP-1103 (CHF5074) stabilizes human transthyretin in healthy human subjects. *Amyloid*. 2017;24(1):42–51.
118. Penchala SC, Connelly S, Wang Y, et al. AG10 inhibits amyloidogenesis and cellular toxicity of the familial amyloid cardiomyopathy-associated V122I transthyretin. *Proc Natl Acad Sci U S A*. 2013;110(24):9992–7.
119. Gamez J, Salvadó M, Reig N, et al. Transthyretin stabilization activity of the catechol-. *Amyloid*. 2019;26(2):74–84.
120. Pinheiro F, Varejão N, Esperante S, et al. Tolcapone, a potent aggregation inhibitor for the treatment of familial leptomeningeal amyloidosis. *FEBS J*. 2021;288(1):310–24.
121. Cardoso I, Martins D, Ribeiro T, Merlini G, Saraiva MJ. Synergy of combined doxycycline/TUDCA treatment in lowering Transthyretin deposition and associated biomarkers: studies in FAP mouse models. *J Transl Med*. 2010;8:74.
122. Crooke ST, Witztum JL, Bennett CF, Baker BF. RNA-targeted therapeutics. *Cell Metab*. 2018;27(4):714–39.
123. Brannagan TH, Wang AK, Coelho T, et al. Early data on long-term efficacy and safety of inotersen in patients with hereditary transthyretin amyloidosis: a 2-year update from the open-label extension of the NEURO-TTR trial. *Eur J Neurol*. 2020;27(8):1374–81.
124. Gonzalez-Duarte A, Coelho T, Adams D, et al. Long-term use of patisiran, an investigational RNAi therapeutic, in patients with hereditary transthyretin-mediated amyloidosis: 12 month efficacy and safety from global open label extension study. *Lancet Neurol*. 2021;20:49–59.
125. Tanowitz M, Hettrick L, Revenko A, Kinberger GA, Prakash TP, Seth PP. Asialoglycoprotein receptor 1 mediates productive uptake of N-acetylgalactosamine-conjugated and unconjugated phosphorothioate antisense oligonucleotides into liver hepatocytes. *Nucleic Acids Res*. 2017;45(21):12388–400.
126. Scott KL, Conley CR, Renfree KJ. Histopathologic evaluation of flexor tenosynovium in recurrent carpal tunnel syndrome. *Plast Reconstr Surg*. 2019;143(1):169–75.
127. Nakamichi KI, Tachibana S. Amyloid deposition in the synovium and ligament in idiopathic carpal tunnel syndrome. *Muscle Nerve*. 1996;19(10):1349–51.
128. Rosenbaum AN, AbouEzzeddine OF, Grogan M, et al. Outcomes after cardiac transplant for wild type transthyretin amyloidosis. *Transplantation*. 2018;102(11):1909–13.
129. Pinney JH, Whelan CJ, Petrie A, et al. Senile systemic amyloidosis: clinical features at presentation and outcome. *J Am Heart Assoc*. 2013;2(2):e000098.
130. Connors LH, Sam F, Skinner M, et al. Heart failure resulting from age-related cardiac amyloid disease associated with wild-type transthyretin: a prospective, observational cohort study. *Circulation*. 2016;133(3):282–90.
131. Westermark P, Westermark GT, Suhr OB, Berg S. Transthyretin-derived amyloidosis: probably a common cause of lumbar spinal stenosis. *Ups J Med Sci*. 2014;119(3):223–8.
132. Yanagisawa A, Ueda M, Sueyoshi T, et al. Amyloid deposits derived from transthyretin in the ligamentum flavum as related to lumbar spinal canal stenosis. *Mod Pathol*. 2015;28(2):201–7.