

Chemical patterns of colony membership and mother-offspring similarity in Antarctic fur seals are reproducible - R Code

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Used packages

Install with “install.packages”. After installation, packages can be called with ‘library’ oder ‘require’

```
library(GCalignR)
library(vegan)
library(readr)
library(ggplot2)
library(ggbeeswarm)
library(tidyverse)
library(pairwiseAdonis)
```

Alignment procedure

Alignment protocol to align a data frame with GCMS peaks for mother-pup pairs and pure pup colonies.

```
load("RData/objects/seal_raw_dfs.Rdata")
```

```
# index correct MP & pup_colony subset
```

```
index_MP <- c(1:61,72:111)
```

```
index_pupcols <- c(51:160)
```

```
# Aligning mom-pup colonies
```

```
mom_pup_aligned <- align_chromatograms(data = all_dfs[index_MP], # input data
                                       rt_col_name = "RT", # retention time variable name
                                       rt_cutoff_low = 15, # remove peaks below 15 Minutes
                                       rt_cutoff_high = 54.7, # remove peaks exceeding 54.7 Minutes
                                       reference = "P13", # sample with highest overall peak number
                                       max_linear_shift = 0.05, # Premise 1: max. shift for linear co
                                       max_diff_peak2mean = 0.08, # Premise 2: max. distance of a pea
                                       min_diff_peak2peak = 0.03, # Premise 3: min. expected distance
                                       delete_single_peak = T, # delete peaks that are present in jus
                                       write_output = NULL) # add variable names to write aligned dat
```

```
# Aligning six pup colonies
```

```
pup_colonies_aligned <- align_chromatograms(data = all_dfs[index_pupcols], # input data
                                             rt_col_name = "RT", # retention time variable name
                                             rt_cutoff_low = 15, # remove peaks below 15 Minutes
                                             rt_cutoff_high = 54.7, # remove peaks exceeding 54.7 Minutes
                                             reference = "P13", # sample with highest overall peak number
```

```

max_linear_shift = 0.05, # Premise 1: max. shift for linear co
max_diff_peak2mean = 0.08, # Premise 2: max. distance of a pea
min_diff_peak2peak = 0.03, # Premise 3: min. expected distance
delete_single_peak = T, # delete peaks that are present in jus
write_output = NULL) # add variable names to write aligned dat

```

Alignment and preliminary data properties

```

## Load and view GCalignR alignment objects for GCMS scent data
## in two and six breeding beaches
load("RData/objects/mom_pup_alignment_GCalignR.RData")
mom_pup_aligned

## Summary of Peak Alignment running align_chromatograms
## Input: all_dfs[index2]
## Start: 2019-05-21 11:41:08 Finished: 2019-05-21 11:44:23
##
## Call:
## GCalignR::align_chromatograms(data[, data=all_dfs, data=index2, rt_col_name=RT,
## rt_cutoff_low=15, rt_cutoff_high=54.7, reference=P13, max_linear_shift=0.05,
## max_diff_peak2mean=0.08, min_diff_peak2peak=0.03, delete_single_peak=T,
## sep=\t, ...=)
##
## Summary of scored substances:
##      total singular retained
##      157          39        118
##
## In total 157 substances were identified among all samples. 39 substances were
## present in just one single sample and were removed. 118 substances are retained
## after all filtering steps.
##
## Sample overview:
## The following 101 samples were aligned to the reference 'P13':
## M01, M02, M03, M04, M05, M06, M07, M08, M09, M10, M11, M12, M13, M14, M15, M16,
## M17, M18, M19, M20, M21, M22, M23, M24, M25, M26, M27, M28, M29, M30, M31, M32,
## M33, M34, M35, M36, M37, M38, M39, M40, M41, M42, M43, M44, M45, M46, M47, M48,
## M49, M50, P01, P02, P03, P04, P05, P06, P07, P07b, P08, P09, P10, P11, P12, P13,
## P14, P15, P16, P17, P18, P19, P20, P21, P22, P23, P24, P25, P26, P27, P28, P29,
## P30, P31, P32, P33, P34, P35, P36, P37, P38, P39, P40, P41, P42, P43, P44, P45,
## P46, P47, P48, P49, P50
##
## For further details type:
## 'gc_heatmap(x)' to retrieve heatmaps
## 'plot(x)' to retrieve further diagnostic plots
load("RData/objects/pup_colonies_alignment_GCalignR.RData")
pup_colonies_aligned

## Summary of Peak Alignment running align_chromatograms
## Input: all_dfs[index4]
## Start: 2019-05-23 11:36:39 Finished: 2019-05-23 11:40:15
##
## Call:
## GCalignR::align_chromatograms(data[, data=all_dfs, data=index4, rt_col_name=RT,

```

```

##   rt_cutoff_low=15, rt_cutoff_high=54.7, reference=P13, max_linear_shift=0.05,
##   max_diff_peak2mean=0.08, min_diff_peak2peak=0.03, delete_single_peak=T,
##   sep=\t, ...=)
##
## Summary of scored substances:
##   total singular retained
##     143      28      115
##
## In total 143 substances were identified among all samples. 28 substances were
## present in just one single sample and were removed. 115 substances are retained
## after all filtering steps.
##
## Sample overview:
##   The following 110 samples were aligned to the reference 'P13':
##   P01, P02, P03, P04, P05, P06, P07, P07b, P08, P09, P10, P100, P101, P102, P103,
##   P104, P105, P106, P107, P108, P109, P11, P12, P13, P14, P15, P16, P17, P18, P19,
##   P20, P21, P22, P23, P24, P25, P26, P27, P28, P29, P30, P31, P32, P33, P34, P35,
##   P36, P37, P38, P39, P40, P41, P42, P43, P44, P45, P46, P47, P48, P49, P50, P51,
##   P52, P53, P54, P55, P56, P57, P58, P59, P60, P61, P62, P63, P64, P65, P66, P67,
##   P68, P69, P70, P71, P72, P73, P74, P75, P76, P77, P78, P79, P80, P81, P82, P83,
##   P84, P85, P86, P87, P88, P89, P90, P91, P92, P93, P94, P95, P96, P97, P98, P99
##
## For further details type:
##   'gc_heatmap(x)' to retrieve heatmaps
##   'plot(x)' to retrieve further diagnostic plots
## Load raw information for all samples containing
## raw peaks and calculate mean peak number
load("RData/objects/seal_raw_dfs.Rdata")

individual_peak_number <- NULL
for (i in 1:length(seal_dfs.list)) {
  individual_peak_number[i] <- length(seal_dfs.list[[i]]$RT)
}

mean_ind_peaks <- mean(individual_peak_number)
sd_ind_peaks <- sd(individual_peak_number)
cat("\n", "\n", "Mean peaks:", as.character(mean_ind_peaks), "\n", "Peak SD:",
    as.character(sd_ind_peaks))

##
##
## Mean peaks: 34.175
## Peak SD: 10.8445563540296

```

NMDS scaling of mother-pup alignment data

```

load("RData/objects/mom_pup_alignment_GCalignR.RData")

scent_factors_raw <- read_delim("documents/metadata_seal_scent.txt",
                                "\t", escape_double = FALSE, trim_ws = TRUE)
scent_factors_raw <- as.data.frame(scent_factors_raw[-c(194:209),])

# set sample names as row names, ensure there are no duplicates

```

```

scent_factors <- scent_factors_raw[,-1]
rownames(scent_factors) <- scent_factors_raw[,1]

## check for empty samples, i.e. no peaks
x <- apply(mom_pup_aligned$aligned$RT, 2, sum)
x <- which(x == 0)

## normalise area and return a data frame
scent <- norm_peaks(mom_pup_aligned, conc_col_name = "Area", rt_col_name = "RT",
                    out = "data.frame")
## common transformation for abundance data to reduce the extent of mean-variance trends
scent <- log(scent + 1)

## subset scent_factors
scent_factors <- scent_factors[rownames(scent_factors) %in% rownames(scent),]
scent <- scent[rownames(scent) %in% rownames(scent_factors),]

## keep order of rows consistent
scent <- scent[match(rownames(scent_factors), rownames(scent)),]

## get number of compounds for each individual sample after alignment
num_comp <- as.vector(apply(scent, 1, function(x) length(x[x>0])))

## bray-curtis similarity
scent_nmds.obj <- vegan::metaMDS(comm = scent, k = 2, try = 999,
                                trymax = 9999, distance = "bray")

scent_nmds <- as.data.frame(scent_nmds.obj[["points"]])

scent_nmds <- cbind(scent_nmds,
                    age = scent_factors[["age"]],
                    tissue_tag = scent_factors[["tissue_tag"]],
                    colony = scent_factors[["colony"]],
                    family = as.factor(scent_factors[["family"]]),
                    clr = as.factor(scent_factors[["clr"]]),
                    shp = as.factor(scent_factors[["shp"]]),
                    gcms = as.factor(scent_factors[["gcms_run"]]),
                    peak_res = as.factor(scent_factors[["peak_res"]]),
                    sample_qlty = as.factor(scent_factors[["sample_qlty"]]),
                    vialdate = as.factor(scent_factors[["gcms_vialdate"]]),
                    captured = as.factor(scent_factors[["capture_date"]]),
                    sex = scent_factors[["sex"]],
                    num_comp = num_comp)
scent_nmds <- scent_nmds %>% mutate(BeachAge = str_c(colony, age, sep = "_"))
# creates & adds new variable BeachAge and simplifies plotting

```

Colony and family membership in SSB and FWB mom-pup pairs

```

load("RData/objects/mom_pup_nmds_scaling.RData")

## colony membership plot
mp_colony_gg <- ggplot(data = scent_nmds) +
  geom_point(size = 4.5, aes(MDS1, MDS2, color = BeachAge, shape = BeachAge)) +

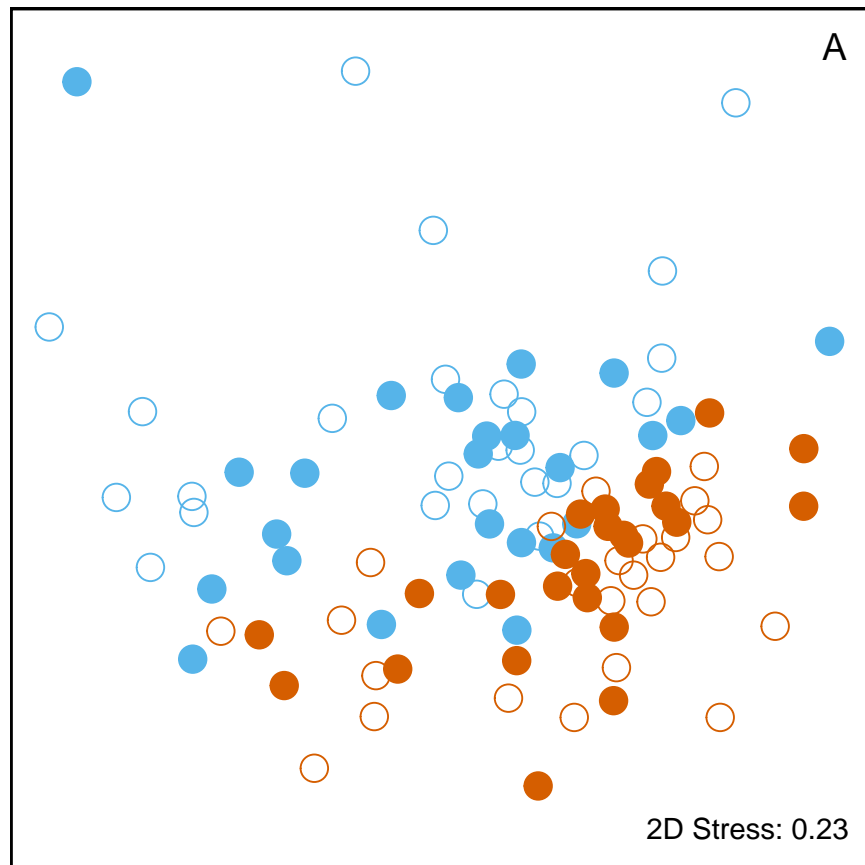
```

```

scale_shape_manual(values = c(19, 1, 19, 1),
  labels = c("FWB mothers ", "FWB pups ",
    "SSB mothers ", "SSB pups ")) +
scale_color_manual(values = c("#D55E00", "#D55E00", "#56B4E9", "#56B4E9"),
  labels = c("FWB mothers ", "FWB pups ",
    "SSB mothers ", "SSB pups ")) +

theme_void() +
ylim(-0.75,1.1) +
annotate("text", x = 0.64, y = 1.1, label = "A", size = 5) +
annotate("text", x = 0.47, y = -0.74, label = "2D Stress: 0.23", size = 4) +
theme(panel.background = element_rect(colour = "black", size = 1, fill = NA),
  aspect.ratio = 1,
  legend.position = "none",
  legend.title = element_blank(),
  legend.background = element_rect(size = 0.3, linetype = "solid", color = "black"))
# call colony membership plot
mp_colony_gg

```



```

## mother-offspring similarity plot
# create color palette for the plot
clr <- c("#D55E00", "red", "#56B4E9", "#009E73", "#000000", "#CC79A7")

# assign pch values for plotting
shp <- c(0,1,2,7,10,5,6,18,16,17,15)

# create unique color-pch pairs

```

```

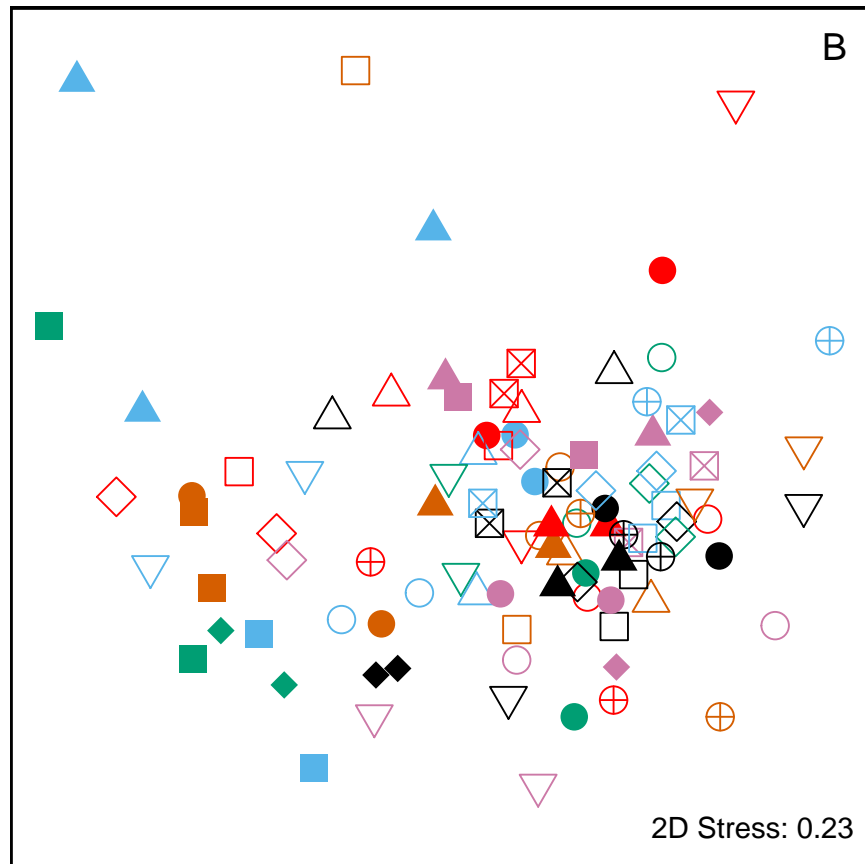
color_shape_pairs <- crossing(clr,shp)

# randomly sample 50 unique pairs (sample without replacement)
set.seed(123) # always get same pairs in a run
color_shape_pairs <- color_shape_pairs[sample(nrow(color_shape_pairs), 50),]

# assign new dataframes to transform scent_nmds$clr & shp with the unique values we created
color_shape_pairs_plot <- rbind(color_shape_pairs[1:25,],color_shape_pairs[1:7,],
                                ,color_shape_pairs[7,], color_shape_pairs[8:25,],
                                color_shape_pairs[26:50,], color_shape_pairs[26:50,])
scent_nmds$clr <- as.factor(color_shape_pairs_plot$clr)
scent_nmds$shp <- as.factor(color_shape_pairs_plot$shp)

# call family plot
mp_family_gg <- ggplot(data = scent_nmds,aes(MDS1,MDS2, color = clr, shape = shp)) +
  geom_point(size = 4.5) +
  scale_shape_manual(values = as.numeric(levels(scent_nmds$shp))) +
  theme_void() +
  ylim(-0.75,1.1) +
  scale_color_manual(values = levels(scent_nmds$clr)) +
  annotate("text", x = 0.64, y = 1.1, label = "B", size = 5) +
  annotate("text", x = 0.48, y = -0.74, label = "2D Stress: 0.23", size = 4) +
  theme(panel.background = element_rect(colour = "black", size = 1,
                                         fill = NA), aspect.ratio = 1,
        legend.position = "none")
mp_family_gg

```



PERMANOVA for mother-pup similarity and colony membership

```
# set seed to reproduce the same outcome (can vary due to different permutations!)
set.seed(123)
```

```
# set counter for while loop
perm_count <- c(99)
```

```
# iterate different significance levels with a while-loop
while (perm_count <= 99999) {# end while-loop after the run for 99999 permutations
  permanova_result_MP <- adonis(scent ~ age+colony+colony:family,
    data = scent_factors,
    method = "bray",
    permutations = perm_count)
  print(permanova_result_MP)
```

```
  perm_count <- (perm_count*10)+9 # ends while-iteration after it increases to 999999
}#while
```

```
##
```

```
## Call:
```

```
## adonis(formula = scent ~ age + colony + colony:family, data = scent_factors,      permutations = perm
```

```
##
```

```
## Permutation: free
```

```
## Number of permutations: 99
```

```
##
```

```

## Terms added sequentially (first to last)
##
##              Df SumsOfSqs MeanSqs F.Model      R2 Pr(>F)
## age              1      0.3217 0.32170  2.6896 0.02253  0.02 *
## colony            1      1.0847 1.08475  9.0692 0.07599  0.01 **
## colony:family     2      1.3870 0.69351  5.7982 0.09716  0.01 **
## Residuals         96     11.4823 0.11961      0.80432
## Total            100     14.2758      1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Call:
## adonis(formula = scent ~ age + colony + colony:family, data = scent_factors,      permutations = perm
##
## Permutation: free
## Number of permutations: 999
##
## Terms added sequentially (first to last)
##
##              Df SumsOfSqs MeanSqs F.Model      R2 Pr(>F)
## age              1      0.3217 0.32170  2.6896 0.02253  0.011 *
## colony            1      1.0847 1.08475  9.0692 0.07599  0.001 ***
## colony:family     2      1.3870 0.69351  5.7982 0.09716  0.001 ***
## Residuals         96     11.4823 0.11961      0.80432
## Total            100     14.2758      1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Call:
## adonis(formula = scent ~ age + colony + colony:family, data = scent_factors,      permutations = perm
##
## Permutation: free
## Number of permutations: 9999
##
## Terms added sequentially (first to last)
##
##              Df SumsOfSqs MeanSqs F.Model      R2 Pr(>F)
## age              1      0.3217 0.32170  2.6896 0.02253 0.0045 **
## colony            1      1.0847 1.08475  9.0692 0.07599 0.0001 ***
## colony:family     2      1.3870 0.69351  5.7982 0.09716 0.0001 ***
## Residuals         96     11.4823 0.11961      0.80432
## Total            100     14.2758      1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Call:
## adonis(formula = scent ~ age + colony + colony:family, data = scent_factors,      permutations = perm
##
## Permutation: free
## Number of permutations: 99999
##
## Terms added sequentially (first to last)
##
##              Df SumsOfSqs MeanSqs F.Model      R2 Pr(>F)

```



```
## age          1      0.3217 0.32170  2.6896 0.02253 0.00403 **
## colony       1      1.0847 1.08475  9.0692 0.07599 1e-05 ***
## colony:family 2      1.3870 0.69351  5.7982 0.09716 1e-05 ***
## Residuals    96     11.4823 0.11961      0.80432
## Total       100     14.2758      1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Post-hoc betadisper and pairwise comparisons for mother-pup pair PERMANOVA results

```
# test for group dispersal
# for different colonies
mod_colony <- betadisper(vegdist(scent), scent_factors$colony, type = "median")
anova(mod_colony)
```

```
## Analysis of Variance Table
##
## Response: Distances
##          Df Sum Sq Mean Sq F value Pr(>F)
## Groups    1 0.0442 0.044201   5.136 0.02561 *
## Residuals 99 0.8520 0.008606
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
# for different ages (mom vs pup)
mod_age <- betadisper(vegdist(scent), scent_factors$age, type = "median")
anova(mod_age)
```

```
## Analysis of Variance Table
##
## Response: Distances
##          Df Sum Sq Mean Sq F value Pr(>F)
## Groups    1 0.01305 0.0130477   1.4726 0.2278
## Residuals 99 0.87718 0.0088604
```

```
# for different combinations of age and colony identity
mod_BeachAge <- betadisper(vegdist(scent), scent_nmds$BeachAge, type = "median")
anova(mod_BeachAge)
```

```
## Analysis of Variance Table
##
## Response: Distances
##          Df Sum Sq Mean Sq F value Pr(>F)
## Groups    3 0.04845 0.0161488   1.9067 0.1336
## Residuals 97 0.82153 0.0084694
```

Pairwise comparison between all possible combinations of mother-pup pairs based on colony identity and age

```
set.seed(123) # for comparability as results do not change when compared to manuscript
pairwiseAdonis::pairwise.adonis(scent, scent_nmds$BeachAge, perm = 99999)
```

```
##          pairs Df SumsOfSqs F.Model      R2 p.value p.adjusted sig
## 1 SSB_1 vs SSB_2 1 0.3515433 2.415725 0.04698416 0.01227 0.07362
## 2 SSB_1 vs FWB_1 1 0.5291662 4.272400 0.08173337 0.00004 0.00024 **
## 3 SSB_1 vs FWB_2 1 0.5542266 4.218847 0.08079165 0.00003 0.00018 **
## 4 SSB_2 vs FWB_1 1 0.8481620 6.618475 0.11899778 0.00001 0.00006 ***
## 5 SSB_2 vs FWB_2 1 0.8397841 6.197227 0.11227424 0.00001 0.00006 ***
## 6 FWB_1 vs FWB_2 1 0.2514355 2.212658 0.04406574 0.01906 0.11436
```

```
# Pairwise group dispersal tests
```

```
# scent, scent_factors and scent_nmds are all data.frames the have the same row setup.  
# Meaning: Each row representing an individual is identical in each data.frame.  
# Thus, we can index pairs based on scent_nmds$BeachAge manually.
```

```
pairwise_betadisper1 <- betadisper(vegdist(scent[scent_nmds$BeachAge == "SSB_1" | scent_nmds$BeachAge ==  
scent_nmds$BeachAge[scent_nmds$BeachAge == "SSB_1" | scent_nmds$BeachAge ==  
type = "median")  
anova(pairwise_betadisper1)
```

```
## Analysis of Variance Table
```

```
##
```

```
## Response: Distances
```

```
##           Df Sum Sq Mean Sq F value Pr(>F)
```

```
## Groups      1 0.00104 0.0010419  0.1024 0.7504
```

```
## Residuals  49 0.49872 0.0101780
```

```
pairwise_betadisper2 <- betadisper(vegdist(scent[scent_nmds$BeachAge == "SSB_1" | scent_nmds$BeachAge ==  
scent_nmds$BeachAge[scent_nmds$BeachAge == "SSB_1" | scent_nmds$BeachAge ==  
type = "median")  
anova(pairwise_betadisper2)
```

```
## Analysis of Variance Table
```

```
##
```

```
## Response: Distances
```

```
##           Df Sum Sq Mean Sq F value Pr(>F)
```

```
## Groups      1 0.02731 0.0273093  3.5184 0.06678 .
```

```
## Residuals  48 0.37256 0.0077618
```

```
## ---
```

```
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
pairwise_betadisper3 <- betadisper(vegdist(scent[scent_nmds$BeachAge == "SSB_1" | scent_nmds$BeachAge ==  
scent_nmds$BeachAge[scent_nmds$BeachAge == "SSB_1" | scent_nmds$BeachAge ==  
type = "median")  
anova(pairwise_betadisper3)
```

```
## Analysis of Variance Table
```

```
##
```

```
## Response: Distances
```

```
##           Df Sum Sq Mean Sq F value Pr(>F)
```

```
## Groups      1 0.00796 0.0079639  0.9519 0.3341
```

```
## Residuals  48 0.40159 0.0083665
```

```
pairwise_betadisper4 <- betadisper(vegdist(scent[scent_nmds$BeachAge == "SSB_2" | scent_nmds$BeachAge ==  
scent_nmds$BeachAge[scent_nmds$BeachAge == "SSB_2" | scent_nmds$BeachAge ==  
type = "median")  
anova(pairwise_betadisper4)
```

```
## Analysis of Variance Table
```

```
##
```

```
## Response: Distances
```

```
##           Df Sum Sq Mean Sq F value Pr(>F)
```

```
## Groups      1 0.03973 0.039734  4.5784 0.03738 *
```

```
## Residuals  49 0.42525 0.008679
```

```
## ---
```

```
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

pairwise_betadisper5 <- betadisper(vegdist(scent[scent_nmds$BeachAge == "SSB_2" | scent_nmds$BeachAge ==
      scent_nmds$BeachAge[scent_nmds$BeachAge == "SSB_2" | scent_nmds$BeachAge ==
      type = "median")
anova(pairwise_betadisper5)

## Analysis of Variance Table
##
## Response: Distances
##           Df Sum Sq Mean Sq F value Pr(>F)
## Groups      1 0.01502 0.015016   1.6302 0.2077
## Residuals  49 0.45134 0.009211

pairwise_betadisper6 <- betadisper(vegdist(scent[scent_nmds$BeachAge == "FWB_1" | scent_nmds$BeachAge ==
      scent_nmds$BeachAge[scent_nmds$BeachAge == "FWB_1" | scent_nmds$BeachAge ==
      type = "median")
anova(pairwise_betadisper6)

## Analysis of Variance Table
##
## Response: Distances
##           Df Sum Sq Mean Sq F value Pr(>F)
## Groups      1 0.00579 0.0057933   0.8476 0.3618
## Residuals  48 0.32807 0.0068347
```

NMDS scaling and colony membership in six pup colonies

```
load("RData/objects/pup_colonies_alignment_GCalignR.RData")
scent_factors_raw <- read_delim("documents/metadata_seal_scent.txt",
      "\t", escape_double = FALSE, trim_ws = TRUE)
scent_factors_raw <- as.data.frame(scent_factors_raw[-c(194:209),])

# set sample names as row names, ensure there are no duplicates
scent_factors <- scent_factors_raw[,-1]
rownames(scent_factors) <- scent_factors_raw[,1]

## check for empty samples, i.e. no peaks
x <- apply(pup_colonies_aligned$aligned$RT, 2, sum)
x <- which(x == 0)

## normalise area and return a data frame
scent <- norm_peaks(pup_colonies_aligned, conc_col_name = "Area", rt_col_name = "RT",
      out = "data.frame")
## common transformation for abundance data to reduce the extent of mean-variance trends
scent <- log(scent + 1)

## subset scent_factors
scent_factors <- scent_factors[rownames(scent_factors) %in% rownames(scent),]
scent <- scent[rownames(scent) %in% rownames(scent_factors),]

## keep order of rows consistent
scent <- scent[match(rownames(scent_factors), rownames(scent)),]

## get number of compounds for each individual sample after alignment
```

```

num_comp <- as.vector(apply(scent, 1, function(x) length(x[x>0])))

## bray-curtis similarity
scent_nmds.obj <- metaMDS(comm = scent, k = 2, try = 999,
                        trymax = 9999, distance = "bray")
## MDS outcome evaluated with PCA for factor colony in metadata table for individuals
scent_nmds <- with(scent_factors, MDSrotate(scent_nmds.obj, colony))

## get x and y coordinates
scent_nmds <- as.data.frame(scent_nmds[["points"]])

## add the colony as a factor to each sample
scent_nmds <- cbind(scent_nmds,
                    age = scent_factors[["age"]],
                    tissue_tag = scent_factors[["tissue_tag"]],
                    colony = scent_factors[["colony"]],
                    family = as.factor(scent_factors[["family"]]),
                    clr = as.factor(scent_factors[["clr"]]),
                    shp = as.factor(scent_factors[["shp"]]),
                    gcms = as.factor(scent_factors[["gcms_run"]]),
                    peak_res = as.factor(scent_factors[["peak_res"]]),
                    sample_qlty = as.factor(scent_factors[["sample_qlty"]]),
                    vialdate = as.factor(scent_factors[["gcms_vialdate"]]),
                    captured = as.factor(scent_factors[["capture_date"]]),
                    sex = scent_factors[["sex"]],
                    num_comp = num_comp)
# creates & adds new variable BeachAge
scent_nmds <- scent_nmds %>% mutate(BeachAge = str_c(colony, age, sep = "_"))

```

Colony membership plot for six pup colonies (Supplementary figure)

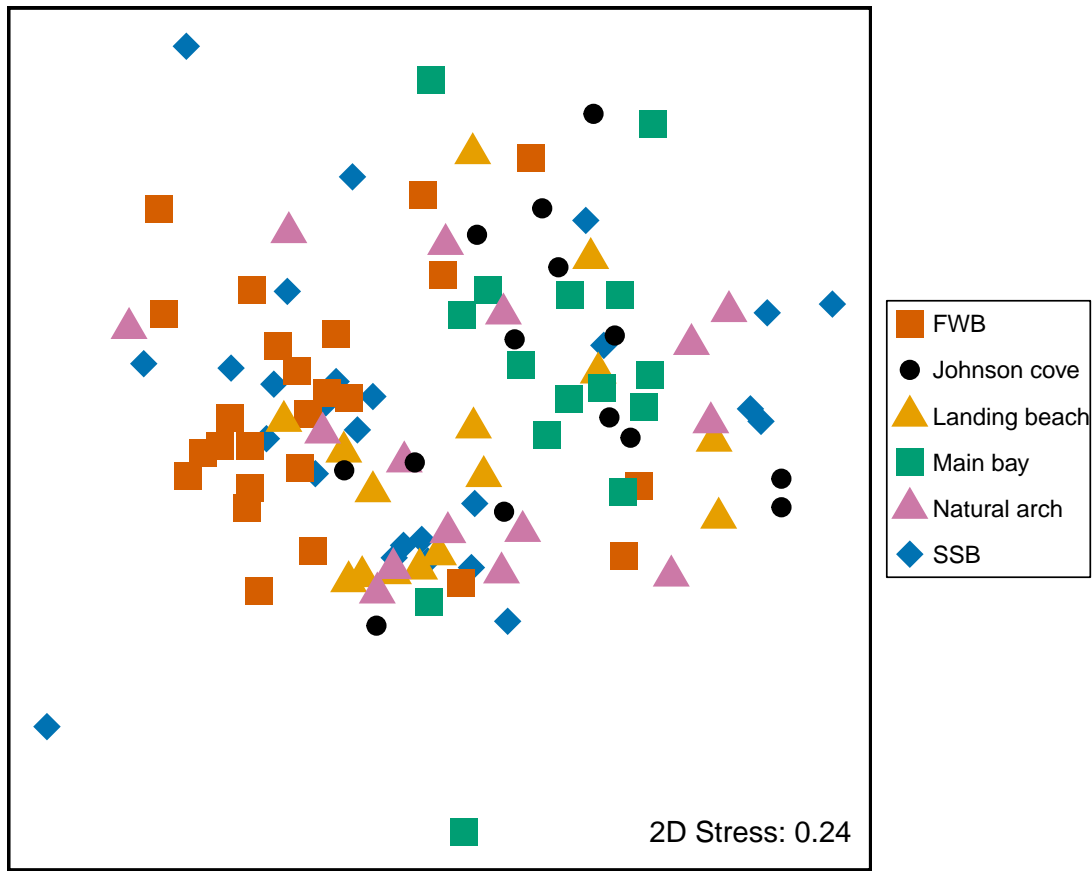
```

load("RData/objects/pup_colonies_nmds_scaling.RData")

pup_colony_gg <- ggplot(data = scent_nmds, aes(MDS1, MDS2, color = colony, shape = colony)) +
  geom_point(size = 4.5) +
  scale_shape_manual(values = c(15,20,17,15,17,18),
                    labels = c("FWB", "Johnson cove", "Landing beach", "Main bay", "Natural arch", "SSI"))
  scale_color_manual(values = c("#D55E00", "#000000", "#E69F00", "#009E73", "#CC79A7", "#0072B2"),
                    labels = c("FWB", "Johnson cove", "Landing beach", "Main bay", "Natural arch", "SSI"))
  theme_void() +
  annotate("text", x = 0.6, y = -0.94, label = "2D Stress: 0.24", size = 4) +
  theme(panel.background = element_rect(colour = "black", size = 1, fill = NA),
        aspect.ratio = 1,
        legend.position = "right", #c(0.1,0.87),
        legend.title = element_blank(),
        # legend.key.size = unit(0.5, "cm"),
        # legend.key.width = unit(0.5, "cm"),
        legend.background = element_rect(size = 0.3, linetype = "solid", color = "black"))

pup_colony_gg

```



PERMANOVA for colony membership in six pup colonies

```
# set seed to reproduce the same outcome (can vary due to different permutations!)
set.seed(123)
```

```
# set counter for while loop
perm_count <- c(99)
```

```
# iterate different significance levels with a while-loop
while (perm_count <= 99999) {# end while-loop after the run for 99999 permutations
  permanova_result_pupcols <- adonis(scent ~ age+colony+colony:family,
    data = scent_factors,
    method = "bray",
    permutations = perm_count)
  print(permanova_result_pupcols)
```

```
  perm_count <- (perm_count*10)+9 # ends while-iteration after it increases to 999999
}#while
```

```
##
```

```
## Call:
```

```
## adonis(formula = scent ~ age + colony + colony:family, data = scent_factors,      permutations = perm
```

```
##
```

```
## Permutation: free
```

```
## Number of permutations: 99
```

```
##
```

```
## Terms added sequentially (first to last)
```

```

##
##              Df SumsOfSqs MeanSqs F.Model      R2 Pr(>F)
## colony              5      3.1874 0.63749  5.1748 0.19128  0.01 **
## colony:family      6      1.4037 0.23395  1.8991 0.08424  0.01 **
## Residuals          98     12.0727 0.12319              0.72449
## Total             109     16.6639              1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Call:
## adonis(formula = scent ~ age + colony + colony:family, data = scent_factors,      permutations = perm
##
## Permutation: free
## Number of permutations: 999
##
## Terms added sequentially (first to last)
##
##              Df SumsOfSqs MeanSqs F.Model      R2 Pr(>F)
## colony              5      3.1874 0.63749  5.1748 0.19128  0.001 ***
## colony:family      6      1.4037 0.23395  1.8991 0.08424  0.001 ***
## Residuals          98     12.0727 0.12319              0.72449
## Total             109     16.6639              1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Call:
## adonis(formula = scent ~ age + colony + colony:family, data = scent_factors,      permutations = perm
##
## Permutation: free
## Number of permutations: 9999
##
## Terms added sequentially (first to last)
##
##              Df SumsOfSqs MeanSqs F.Model      R2 Pr(>F)
## colony              5      3.1874 0.63749  5.1748 0.19128  1e-04 ***
## colony:family      6      1.4037 0.23395  1.8991 0.08424  2e-04 ***
## Residuals          98     12.0727 0.12319              0.72449
## Total             109     16.6639              1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Call:
## adonis(formula = scent ~ age + colony + colony:family, data = scent_factors,      permutations = perm
##
## Permutation: free
## Number of permutations: 99999
##
## Terms added sequentially (first to last)
##
##              Df SumsOfSqs MeanSqs F.Model      R2 Pr(>F)
## colony              5      3.1874 0.63749  5.1748 0.19128  1e-05 ***
## colony:family      6      1.4037 0.23395  1.8991 0.08424  6e-05 ***
## Residuals          98     12.0727 0.12319              0.72449
## Total             109     16.6639              1.00000

```

```
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Post-hoc tests for PERMANOVA results for six pup colonies

# pairwise PERMANOVA
pairwiseAdonis::pairwise.adonis(scent, scent_factors$colony, perm = 99999)

##              pairs Df SumsOfSqs  F.Model      R2 p.value
## 1              SSB vs FWB   1 0.8086332 6.080018 0.11038519 0.00001
## 2      SSB vs landing_beach   1 0.4880064 3.544865 0.08332062 0.00083
## 3      SSB vs main_bay       1 0.8584284 6.181387 0.13681269 0.00001
## 4      SSB vs natural_arch   1 0.5667911 4.172853 0.09665456 0.00010
## 5      SSB vs johnson        1 0.6168108 4.407128 0.10392422 0.00005
## 6      FWB vs landing_beach   1 0.5117674 4.168468 0.09885273 0.00009
## 7      FWB vs main_bay       1 0.9039828 7.289582 0.16095494 0.00001
## 8      FWB vs natural_arch   1 0.9556981 7.905832 0.17221847 0.00001
## 9      FWB vs johnson        1 0.8911846 7.145352 0.16185966 0.00001
## 10 landing_beach vs main_bay   1 0.4229462 3.322412 0.10607138 0.00186
## 11 landing_beach vs natural_arch 1 0.3694950 3.002564 0.09684889 0.00413
## 12 landing_beach vs johnson    1 0.3459312 2.694198 0.09073145 0.01225
## 13 main_bay vs natural_arch    1 0.7381617 5.917530 0.17446818 0.00001
## 14 main_bay vs johnson         1 0.4083747 3.137902 0.10411813 0.00024
## 15 natural_arch vs johnson     1 0.3053377 2.428242 0.08251399 0.01651
##      p.adjusted sig
## 1      0.00015  **
## 2      0.01245   .
## 3      0.00015  **
## 4      0.00150   *
## 5      0.00075  **
## 6      0.00135   *
## 7      0.00015  **
## 8      0.00015  **
## 9      0.00015  **
## 10     0.02790   .
## 11     0.06195
## 12     0.18375
## 13     0.00015  **
## 14     0.00360   *
## 15     0.24765

# test for group dispersal
mod2 <- betadisper(vegdist(scent), scent_factors$colony, type = "median")
anova(mod2)

## Analysis of Variance Table
##
## Response: Distances
##              Df Sum Sq  Mean Sq F value Pr(>F)
## Groups         5 0.02003  0.0040065    0.497 0.7779
## Residuals    104 0.83841  0.0080616
```

Re-evaluation of 2011 field season scent data

Perform non-metric multidimensional scaling

Re-evaluation in PERMANOVA instead of ANOSIM

```
## PERMANOVA
set.seed(123)
adonis(scent ~ age+colony+colony:family,
       data = peak_factors,
       permutations = 99999)

##
## Call:
## adonis(formula = scent ~ age + colony + colony:family, data = peak_factors,      permutations = 99999)
##
## Permutation: free
## Number of permutations: 99999
##
## Terms added sequentially (first to last)
##
##              Df SumsOfSqs MeanSqs F.Model      R2 Pr(>F)
## age           1    0.2014 0.20143   0.9785 0.01013 0.4613
## colony        1    2.5430 2.54300  12.3538 0.12790 1e-05 ***
## colony:family  2    1.2880 0.64400   3.1285 0.06478 1e-05 ***
## Residuals     77   15.8503 0.20585           0.79719
## Total         81   19.8827           1.00000
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

# Test for heterogeneity
anova(betadisper(vegdist(scent), peak_factors$colony))

## Analysis of Variance Table
##
## Response: Distances
##              Df   Sum Sq   Mean Sq F value Pr(>F)
## Groups        1 0.000791 0.0007913   0.222 0.6388
## Residuals     80 0.285197 0.0035650
```

Effect size estimate by PERMANOVA R^2 bootstrap

```
## Load data and assign data to data.frames
load("RData/objects/R2_initial_season_btrap.RData")

old_season_colony <- paov_r2_results[[1]][[2]]
old_season_family <- paov_r2_results[[1]][[3]]

load("RData/objects/R2_replication_season_btrap.RData")
new_season_colony <- paov_r2_results[[1]][[2]]
new_season_family <- paov_r2_results[[1]][[3]]

MP_effectsize <- c(old_season_colony, new_season_colony,
                  old_season_family, new_season_family)

MP_effectsize.groups <- c(rep("Colony S1", 5000),
                        rep("Colony S2", 5000),
                        rep("Family S1", 5000),
                        rep("Family S2", 5000))
```



```
MP_effectsize.df <- data.frame(btrap_combined_results = MP_effectsize,
                              btrap_subset_groups = MP_effectsize.groups)
```

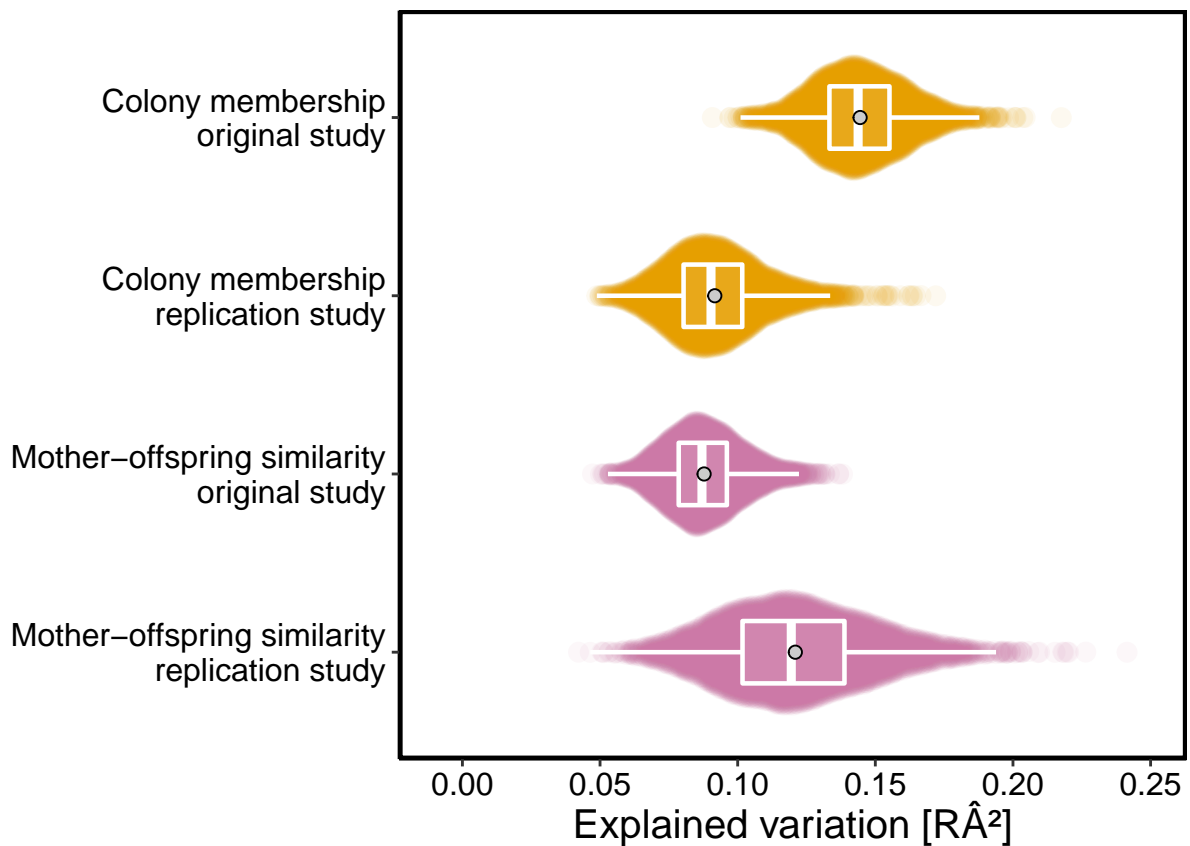
Effect size estimate plot

```
load("RData/objects/effect_size_df.RData")
# point estimates for PERMANOVA on non-bootstrapped (original) data
point_estimate <- c(0.1444734, 0.09168289, 0.08780086, 0.1209394)
# point estimate groups for reasons of comprehensibility
point_estimate_groups <- c("Colony S1", "Colony S2", "Family S1", "Family S2")

# plot commands
MP_effectsize_gg <- ggplot(MP_effectsize.df, aes(y = btrap_combined_results,
                                                  x = btrap_subset_groups,
                                                  color = btrap_subset_groups)) +

  # this arranges the points according to their density
  geom_quasirandom(alpha = 0.06, size = 3, width = 0.3, bandwidth = 1) +
  scale_color_manual(values = c("#E69F00", "#E69F00", "#CC79A7", "#CC79A7")) +
  # makes the boxplots
  geom_boxplot(width = 0.35, outlier.shape = NA, color = "white", alpha = 0.1, lwd=0.8) +
  annotate("point", x = 1, y = point_estimate[4], colour = "#000000",
           fill = "#CCCCCC", size = 2, shape = 21) +
  annotate("point", x = 2, y = point_estimate[3], colour = "#000000",
           fill = "#CCCCCC", size = 2, shape = 21) +
  annotate("point", x = 3, y = point_estimate[2], colour = "#000000",
           fill = "#CCCCCC", size = 2, shape = 21) +
  annotate("point", x = 4, y = point_estimate[1], colour = "#000000",
           fill = "#CCCCCC", size = 2, shape = 21) +
  # this is a possible theme of the plot, there are many others
  theme_classic() +
  # changes the labels on the x axis
  scale_y_continuous(limits = c(-0.01, 0.25),
                    breaks = seq(0, 0.25, 0.05)) +
  scale_x_discrete(labels = c("Family S2" = "Mother-offspring similarity\nreplication study",
                              "Family S1" = "Mother-offspring similarity\noriginal study",
                              "Colony S2" = "Colony membership\nreplication study",
                              "Colony S1" = "Colony membership\noriginal study"),
                  limits = c("Family S2",
                              "Family S1",
                              "Colony S2",
                              "Colony S1")) +
  # geom_hline(yintercept = 0, linetype = "dashed") +
  xlab("") +
  # label for y axis
  ylab("Explained variation [R2]") +
  # flips plot so everything is horizontal
  coord_flip() +
  # adjust theme specifics
  theme(panel.background = element_rect(colour = "black", size = 1.25, fill = NA),
        text = element_text(size = 15),
        axis.text = element_text(colour = "black"),
        legend.position = "none")
```

MP_effectsize_gg



R2 Bootstrap Code

```
## creates function 'scent_btrap_r2_swarm_data' that performs bootstrap

# Bootstrap to track R2 values for randomized subsets. In addition,
# bootstrap cannot only be used to randomize the chemical data frame
# to evaluate R2 distribution as effect size estimates,
# but also to evaluate R2 change for different subsets based on different
# premises. 1) Frequent peaks 2) Strong concentrations 3) Peaks identified by SIMPER

require(vegan)

# path: file path to scent_nmds-mompup2017_ssbfwb.RData",
# objects: scent_nmds, scent_nmds.obj, scent_factors, scent
# df.permutations: number of times the scent.df from loaded data will be permuted
# nmds.permutations: number of permutation in nMDS using Bray-Curtis
# btrap.iterations: number of procedure repeats

scent_btrap_r2_swarm_data <- function(path, df.permutations = 15,
                                       nmds.permutations = 999,
                                       btrap.iterations = 5000){
  # Create a data frame by permuting the data for scent
  # compounds data and also ensure that each population*age occur
```

```

# same amounts of time in the permutation data frame.
#-----

# load data frame with data of aligned fur seal chromatograms
load(path)
scent_factors <- peak_factors
# transfer BeachAge Column from scent_nmds to meta data.frame scent_factors
scent_factors <- cbind(scent_factors,
                      BeachAge = scent_nmds$BeachAge)

# create index column for meta data frame
scent_factors <- cbind(scent_factors,
                      SampleIndex = 1:length(rownames(scent_factors)))

# create data.frame to track PERMANOVA results over repeated tests
nonsubset_results_paov <- data.frame(R2_age = double(), p_colfam = double(),
                                     R2_residual = double(),
                                     F_Het = double(), p_Het = double())
promcomp_results_paov <- data.frame(R2_age = double(), p_colfam = double(),
                                    R2_residual = double(),
                                    F_Het = double(), p_Het = double())
highcomp_results_paov <- data.frame(R2_age = double(), p_colfam = double(),
                                    R2_residual = double(),
                                    F_Het = double(), p_Het = double())
simper_results_paov <- data.frame(R2_age = double(), p_colfam = double(),
                                  R2_residual = double(),
                                  F_Het = double(), p_Het = double())

# create list to store created objects in an iteration
iter_object_container <- list()

for (i in 1:btrap.iterations) {

  # create data.frame subsets (colony subset) by indexing the meta data.frame
  scent.f.ssb.m <- scent_factors[scent_factors$BeachAge == "SSB_1",]
  scent.f.fwb.m <- scent_factors[scent_factors$BeachAge == "FWB_1",]
  scent.f.ssb.p <- scent_factors[scent_factors$BeachAge == "SSB_2",]
  scent.f.fwb.p <- scent_factors[scent_factors$BeachAge == "FWB_2",]

  # int vector of row index number of permuted scent.ssb data.frame
  # row numbers will be used to create a permuted data.frame of
  # evenly distributed draws of individuals
  permute_rows_ssb_m <- sample(scent.f.ssb.m$SampleIndex, df.permutations, replace = T)
  permute_rows_fwb_m <- sample(scent.f.fwb.m$SampleIndex, df.permutations, replace = T)
  permute_rows_ssb_p <- sample(scent.f.ssb.p$SampleIndex, df.permutations, replace = T)
  permute_rows_fwb_p <- sample(scent.f.fwb.p$SampleIndex, df.permutations, replace = T)

  # create overall index number that can be used to
  # index data.frame(scent): index corresponds to correct individual
  perm_index_all <- c(permute_rows_ssb_m,
                     permute_rows_fwb_m,

```

```

        permute_rows_ssb_p,
        permute_rows_fwb_p)

# create new data.frame with indeces found in permutation
# results vector perm_index_all
scent.permute <- scent[perm_index_all,]
scent_factors.permute <- scent_factors[perm_index_all,]
# rownames(scent.permute) == rownames(scent_factors.permute) # TRUE
#-----

# Perform analysis to find 3 subsets based on different premises
# with the permuted data frame.
# Track 15 best performing compounds of an analysis
#-----

## NDMS scale results
## count number of peaks that are not 0 per column
peak_count <- as.vector(apply(scent.permute, 2, function(x) length(x[x>0])))

## add peaks in a column that are not 0 to estimate highest
# concentration peak sum
peak_add <- as.vector(apply(scent.permute, 2, function(x) sum(x)))

## create dataframe with same name properties as scent.RData
compound_subset <- data.frame(name = colnames(scent.permute),
                              peak_count, peak_add)

## sort data frame for most prominent compounds over all samples
most_abundant <- compound_subset %>% arrange(desc(peak_count))

## shorten scent matrix to only the 15 most abundant compounds
scent.promcomp <- scent.permute[colnames(scent.permute) %in%
                                most_abundant$name[1:15]]

## sort data frame for most highly concentrated compounds over all samples
most_concentration <- compound_subset %>% arrange(desc(peak_add))

## shorten scent matrix to only the 15 most abundant compounds
scent.highcomp <- scent.permute[colnames(scent.permute) %in%
                                most_concentration$name[1:15]]

## simper
# simper analysis and results array
sim <- with(scent_factors.permute,
           simper(scent.permute, colony))
best.compounds.simper.btrap <- summary(sim)[[1]]
#filter 15 compounds that contribute most towards dissimilarity of individuals
simper_comps <- as.numeric(rownames(best.compounds.simper.btrap))
best_comps <- simper_comps[1:15]
# subset peak data matrix {scent}
scent.simper.btrap <- scent.permute[,which(colnames(scent.permute) %in%
                                           as.character(best_comps))]
#-----

```

```

# Take 15 identified compounds and limit nMDS of the permuted
# data frame (scent.permute) to only those compounds

#-----

# bray-curtis similarity
scent_nmds_regular.obj <- vegan::metaMDS(comm = scent.permute, k = 2,
                                         try = df.permutations, distance = "bray")
scent_nmds_count.obj <- vegan::metaMDS(comm = scent.promcomp, k = 2,
                                         try = df.permutations, distance = "bray")
scent_nmds_add.obj <- vegan::metaMDS(comm = scent.highcomp, k = 2,
                                      try = df.permutations, distance = "bray")
scent_nmds_simper.obj <- vegan::metaMDS(comm = scent.simper.btrap, k = 2,
                                         try = df.permutations, distance = "bray")

## get x and y coordinates
scent_nmds_regular <- as.data.frame(scent_nmds_regular.obj[["points"]])
scent_nmds_count <- as.data.frame(scent_nmds_count.obj[["points"]])
scent_nmds_add <- as.data.frame(scent_nmds_add.obj[["points"]])
scent_nmds_simper <- as.data.frame(scent_nmds_simper.obj[["points"]])

## add the colony as a factor to each sample
scent_nmds <- data.frame(MDS1r = scent_nmds_regular[["MDS1"]],
                        MDS2r = scent_nmds_regular[["MDS2"]],
                        MDS1c = scent_nmds_count[["MDS1"]],
                        MDS2c = scent_nmds_count[["MDS2"]],
                        MDS1a = scent_nmds_add[["MDS1"]],
                        MDS2a = scent_nmds_add[["MDS2"]],
                        MDS1s = scent_nmds_simper[["MDS1"]],
                        MDS2s = scent_nmds_simper[["MDS2"]],
                        age = scent_factors.permute[["age"]],
                        colony = scent_factors.permute[["colony"]],
                        family = scent_factors.permute[["family"]],
                        BeachAge = scent_factors.permute[["BeachAge"]])
)
#-----

# Perform PERMANOVA on distance matrix based limited scent compounds data
#-----

# not subsetted
nonsubset.df_permanova <- adonis(scent.permute ~ age + colony + colony:family,
                                data = scent_factors.permute,
                                permutations = 9999)
nonsubset.df_hetgeneity <- anova(betadisper(vegdist(scent.permute),
                                                scent_factors.permute$colony))

# track important values of statistical analysis in this run
nonsubset_iter_res_paov <- cbind(R2_age = nonsubset.df_permanova$aov.tab$R2[1],
                                R2_colony = nonsubset.df_permanova$aov.tab$R2[2],
                                R2_famcol = nonsubset.df_permanova$aov.tab$R2[3],
                                R2_residual = nonsubset.df_permanova$aov.tab$R2[4],
                                F_Het = nonsubset.df_hetgeneity$`F value`[1],

```

```

p_Het = nonsubset.df_hetgeneity$`Pr(>F)`[1])

# bind run values to track changes over iterations in the for-loop
nonsubset_results_paov <- rbind(nonsubset_results_paov,
                                nonsubset_iter_res_paov)

#prom comps
promcomp.df_permanova <- adonis(scent.promcomp ~ age + colony + colony:family,
                                data = scent_factors.permute,
                                permutations = 9999)
promcomp.df_hetgeneity <- anova(betadisper(vegdist(scent.promcomp),
                                             scent_factors.permute$colony))

promcomp_iter_res_paov <- cbind(R2_age = promcomp.df_permanova$aov.tab$R2[1],
                                R2_colony = promcomp.df_permanova$aov.tab$R2[2],
                                R2_famcol = promcomp.df_permanova$aov.tab$R2[3],
                                R2_residual = promcomp.df_permanova$aov.tab$R2[4],
                                F_Het = promcomp.df_hetgeneity$`F value`[1],
                                p_Het = promcomp.df_hetgeneity$`Pr(>F)`[1])

promcomp_results_paov <- rbind(promcomp_results_paov,
                                promcomp_iter_res_paov)

# high comps
highcomp.df_permanova <- adonis(scent.highcomp ~ age + colony + colony:family,
                                data = scent_factors.permute,
                                permutations = 9999)
highcomp.df_hetgeneity <- anova(betadisper(vegdist(scent.highcomp), scent_factors.permute$colony))

highcomp_iter_res_paov <- cbind(R2_age = highcomp.df_permanova$aov.tab$R2[1],
                                R2_colony = highcomp.df_permanova$aov.tab$R2[2],
                                R2_famcol = highcomp.df_permanova$aov.tab$R2[3],
                                R2_residual = highcomp.df_permanova$aov.tab$R2[4],
                                F_Het = highcomp.df_hetgeneity$`F value`[1],
                                p_Het = highcomp.df_hetgeneity$`Pr(>F)`[1])

highcomp_results_paov <- rbind(highcomp_results_paov,
                                highcomp_iter_res_paov)

# SIMPER
simper.df_permanova <- adonis(scent.simper.btrap ~ age + colony + colony:family,
                                data = scent_factors.permute,
                                permutations = 9999)
simper.df_hetgeneity <- anova(betadisper(vegdist(scent.simper.btrap), scent_factors.permute$colony))

simper_iter_res_paov <- cbind(R2_age = simper.df_permanova$aov.tab$R2[1],
                                R2_colony = simper.df_permanova$aov.tab$R2[2],
                                R2_famcol = simper.df_permanova$aov.tab$R2[3],
                                R2_residual = simper.df_permanova$aov.tab$R2[4],
                                F_Het = simper.df_hetgeneity$`F value`[1],
                                p_Het = simper.df_hetgeneity$`Pr(>F)`[1])

simper_results_paov <- rbind(simper_results_paov,
                                simper_iter_res_paov)

```

```

#-----

# # pack all this in a list to be later on stored in a list that can be saved again
# create name giving the iteration step
iteration_count <- paste0("iter_", i)

# create list that stores relevant workspace elements for an iteration step
iter_objects <- list(scent.permute = scent.permute,
                    scent_factors.permute = scent_factors.permute,
                    scent.promcomp = scent.promcomp,
                    scent.highcomp = scent.highcomp,
                    sim = sim,
                    scent.simper.btrap = scent.simper.btrap,
                    scent_nmds_regular.obj = scent_nmds_regular.obj,
                    scent_nmds_count.obj = scent_nmds_count.obj,
                    scent_nmds_add.obj = scent_nmds_add.obj,
                    scent_nmds_simper.obj = scent_nmds_simper.obj,
                    scent_nmds_regular = scent_nmds_regular,
                    scent_nmds_count = scent_nmds_count,
                    scent_nmds_add = scent_nmds_add,
                    scent_nmds_simper = scent_nmds_simper,
                    promcomp.df_permanova = promcomp.df_permanova,
                    promcomp.df_hetgeneity = promcomp.df_hetgeneity,
                    highcomp.df_permanova = highcomp.df_permanova,
                    highcomp.df_hetgeneity = highcomp.df_permanova,
                    simper.df_permanova = simper.df_permanova,
                    simper.df_hetgeneity = simper.df_hetgeneity)

# save everything as a list in a container list, that stores
# information/elements of all iteration steps
iter_object_container[[i]] <- iter_objects
names(iter_object_container)[i] <- iteration_count

} # end i

paov_r2_results <- list(regular = nonsubset_results_paov,
                        promcomp = promcomp_results_paov,
                        highcomp = highcomp_results_paov,
                        simper_res = simper_results_paov)
return(list(paov_r2_results = paov_r2_results,
            iter_object_container = iter_object_container))
} # end function

```

Session information

```

## R version 3.5.3 (2019-03-11)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 18363)
##
## Matrix products: default
##
## locale:

```

```

## [1] LC_COLLATE=English_Germany.1252 LC_CTYPE=English_Germany.1252
## [3] LC_MONETARY=English_Germany.1252 LC_NUMERIC=C
## [5] LC_TIME=English_Germany.1252
##
## attached base packages:
## [1] stats      graphics  grDevices  utils      datasets  methods    base
##
## other attached packages:
## [1] pairwiseAdonis_0.0.1 cluster_2.0.7-1      forcats_0.4.0
## [4] stringr_1.4.0      dplyr_0.8.0.1      purrr_0.3.1
## [7] tidyr_0.8.3        tibble_2.0.1       tidyverse_1.2.1
## [10] ggbeeswarm_0.6.0   ggplot2_3.1.1      readr_1.3.1
## [13] vegan_2.5-4        lattice_0.20-38     permute_0.9-5
## [16] GCalignR_1.0.2
##
## loaded via a namespace (and not attached):
## [1] Rcpp_1.0.0      lubridate_1.7.4  assertthat_0.2.0 digest_0.6.18
## [5] R6_2.4.0        cellranger_1.1.0 plyr_1.8.4      backports_1.1.3
## [9] evaluate_0.13   httr_1.4.0       pillar_1.3.1    rlang_0.3.1
## [13] lazyeval_0.2.1  readxl_1.3.0     rstudioapi_0.9.0 Matrix_1.2-15
## [17] rmarkdown_2.1   labeling_0.3     splines_3.5.3   munsell_0.5.0
## [21] broom_0.5.1     compiler_3.5.3   vipor_0.4.5     modelr_0.1.4
## [25] xfun_0.5        pkgconfig_2.0.2  mgcv_1.8-27     htmltools_0.3.6
## [29] tidyselect_0.2.5 crayon_1.3.4     withr_2.1.2     MASS_7.3-51.1
## [33] grid_3.5.3      nlme_3.1-137     jsonlite_1.6     gtable_0.2.0
## [37] magrittr_1.5     scales_1.0.0     cli_1.0.1        stringi_1.3.1
## [41] xml2_1.2.0       generics_0.0.2   tools_3.5.3      glue_1.3.0
## [45] beeswarm_0.2.3  hms_0.4.2        parallel_3.5.3   yaml_2.2.0
## [49] colorspace_1.4-0 rvest_0.3.2      knitr_1.22       haven_2.1.0

```