Computational Biology Lecture 12: HMM and beyond

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Hidden Markov Machines - Recap

 Viterbi algorithm finds most probable path given probabilities and states

$$q^* = \operatorname*{argmax}_p P(oldsymbol{x}, q_p)$$

► Forward and Backwards algorithms finding the posterior probability of a state..

$$P(q_t = i|\mathbf{x}) = \frac{P(x, q_t = i)}{P(\mathbf{x})} = \frac{f_i(t)b_i(t)}{P(\mathbf{x})}$$

▶ are simplifications. Actually need the parameters:

$$q^* = \operatorname*{argmax}_p P(\mathbf{x}, q_p | \mathbf{\theta}) \ P(q_t = i | \mathbf{x} | \mathbf{\theta}) = \frac{f_i(t)b_i(t)}{P(\mathbf{x} | \mathbf{\theta})}$$

.. where θ are the parameters of the model.

We can sometimes infer these parameters if we know alot about the system generating the sequence.. however usually we don't

Training HMMs - Introduction

Given a sequence and a state path, we can count

- ▶ A_{ij} count of $i \rightarrow j$ transitions
- \triangleright $E_i(I)$ count of letter I when in state i

To maximise likelihood of the sequence:

$$e_i(I) = \frac{E_i(I)}{\sum_{l \in \mathcal{A}} E_i(I)} \quad a_{ij} = \frac{A_{ij}}{\sum_{k \in \mathcal{Q}} A_{ik}}$$
(1)

Training HMMs - Introduction

Need to know:

- probabilities to define the state sequence
- state sequence to work out the probabilities

How do we get round it?

Training HMMs - Introduction

Need to know:

- probabilities to define the state sequence
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How do we get round it?

Expectation Maximisation

Training HMMs - Baum Welch

- make initial guess at transition and emission probabilities
- use this to estimate posterior probabilities Expectation
- calculate the new transition and emission probabilities given the state posterior probabilities calculated - Maximisation

The second and third steps can be repeated until and good enough estimation is reached

Training HMMs - Baum Welch

One more point of detail is Pseudocounts

$$e_i(I) = \frac{E_i(I)}{\sum_{I \in \mathcal{A}} E_i(I)} \quad a_{ij} = \frac{A_{ij}}{\sum_{k \in \mathcal{Q}} A_{ik}}$$
 (2)

Problem:

- ► HMM may want to allow transitions that we don't see much in the training data
- Can solve by adding pseudo counts

Training HMMs - Baum Welch

One more point of detail is Pseudocounts

$$e_i(I) = \frac{E_i(I)}{\sum_{I \in \mathcal{A}} E_i(I)} \quad a_{ij} = \frac{A_{ij}}{\sum_{k \in \mathcal{Q}} A_{ik}}$$
 (2)

Problem:

- ► HMM may want to allow transitions that we don't see much in the training data
- Can solve by adding pseudo counts

$$e_{i}(I) = \frac{E_{i}(I) + r_{i}(I)}{\sum_{I \in \mathcal{A}} E_{i}(I) + r_{i}(I)} \quad a_{ij} = \frac{A_{ij} + r_{i}(I)}{\sum_{k \in \mathcal{Q}} A_{ik} + r_{i}(I)}$$
(3)

There is more detail but I will not cover it here. For more see Durbin *et al*, Biological Sequence Analysis

The structure of an HMM is usually designed by an expert. Need to decide on:

- number of states
- what transitions are allowed
- what order

Number of states:

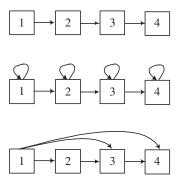
Too few states:

- ► fail to model significant differences
- miss detail
- underfit

Too many states:

- model noise
- poor generalisation
- overfit

In Biology HMMs are usually built from blocks
These represent common motifs in biological sequences



Given a set of motifs, a genetic algorithm can also generate good structures

For more complex problems, they are better than experts

Transitions between states:

- should represent real physical structure
- ▶ fully connected gives best likelihood but can overfit

Order

The Markov property of HMM imposes too little short range structure

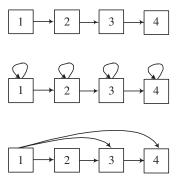
They can be more generalised to second order:

$$P(x_i|x_{i-1},x_{1-2},...,x_1) = P(x_i|x_{i-1},x_{i-2})$$

and to nth order

$$P(x_i|x_{i-1},x_{1-2},...,x_1) = P(x_i|x_{i-1},...,x_{i-n})$$

In Biology HMMs are usually built from blocks
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Given a set of motifs, a genetic algorithm can also generate good structures

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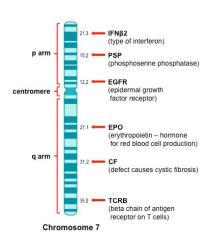
HMM and Beyond - Finding Genes

- ► There are about 20,000 genes in the genome
- ▶ 25% of the genome is genes

Genes contain:

- promoters
- enhancers
- silencers
- insulators
- ▶ coding 1%

Finding Genes on the genome



HMM and Beyond - CpG islands

The 'CG' base dinucleotide sequence in DNA is less common than expected

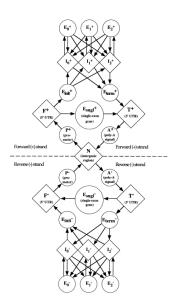
- methylation of the Cytosine base is common
- more likely to mutate in to a Thymine base

Methylation is supressed in promoter regions of the genome

HMM and Beyond - HMM Example

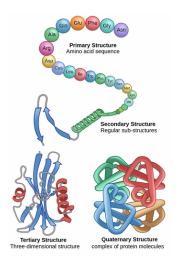
GENESCAN

- found genes
- ▶ fifth order Markov model
- ▶ Burge and Karlin 1997
- looks for areas of high 'CpG' content

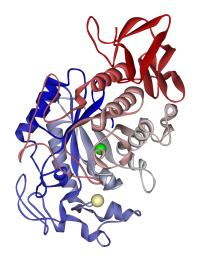


Proteins structure:

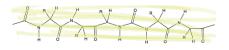
- Primary amino acid sequence
- Secondary α helices, β sheets or coil
- Tertiary 3D structure of polypeptide
- Quaternary protein complex



Amylase enzyme protein

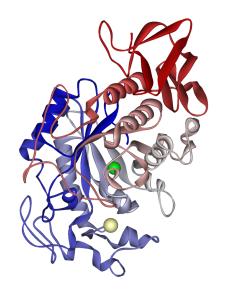


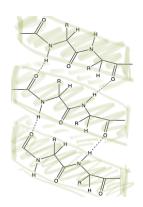
This is the amylase in your saliva, breaks down starch in to sugars. If you chew bread or plain pasta for a while, it goes sweet because of this enzyme.



Each strand is made of a sequence of amino acids

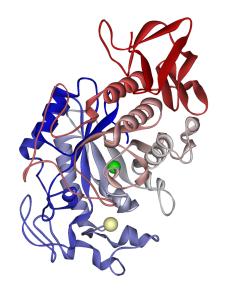
Amylase enzyme protein

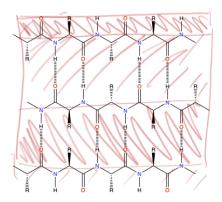




The spiral shapes are called 'alpha helix', and are the amino acid chains stacked up together, held together with hydrogen bonds

Amylase enzyme protein





The flat ribbons are called 'beta sheets', and have the amino acid chain forming a flat structure, held together with hydrogen bonds

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Can the amino acid sequence be used to generate the *secondary structure?*

Difficulties:

- Non local
- Different lengths of input
- Representation

However:

- There is now good training data
- Can use windows to get a fixed input

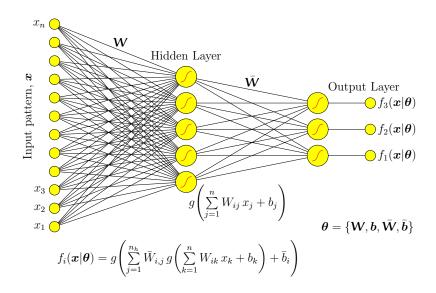
Yes.

HMM and Beyond - Secondary Structure Prediction

Qian and Sejnowski 1988:

Adapted their 'NETtalk' system from a word sequence to an amino acid sequence

- mean squared error loss
- one hot encoding representation
- windowed input



Loss function - mean squared error

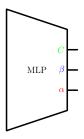
$$L(\theta, \mathcal{D}) = \frac{1}{m} \sum_{i=1}^{m} ||y_i - f(x_i|\theta)||^2$$

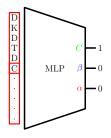
Follow negative gradient wrt. parameters $\boldsymbol{\theta}$ Make a step in the direction of negative gradient

$$\theta(t+1) = \theta(t) - r\nabla L(\theta, \mathcal{D})$$

r is learning rate

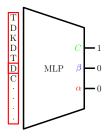
How to make a variable length sequence a fixed length input?

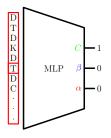


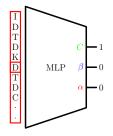


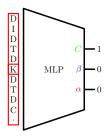
```
CCCCCααααααααααCCCββββββββCCCCC

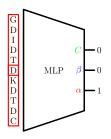
· · · · CDTDKDTD I DG I Y DWCMK E N L KWA NMA P N I · · · · · ·
```

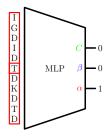




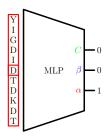


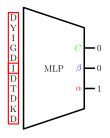




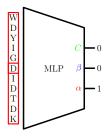


```
CCCCCαα<mark>αααααααα</mark>αCCCβββββββCCCCC
····CDTDKDTDIDGIY</mark>DWCMKENLKWANMAPNI·····
```

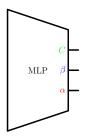




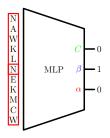
```
CCCCCαααααααααCCCβββββββCCCCC
. . . . . CDTDKDTDIDGIYDWCMKENLKWANMAPNI . . . . .
```

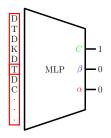


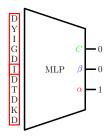
Need to shuffle:

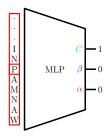


```
CCCCCααααααααααCCCββββββββCCCCC
....CDTDKDTDIDGIYDWCMKENLKWANMAPNI.....
```









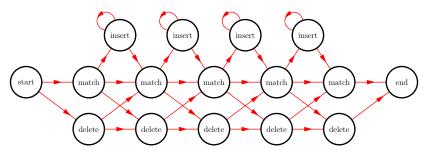
Qian Sejnowski - 1988 - 64.3% 'Coil' more common, 47%, easier for network to recognise. Improved Rost and Sander - 1993

- profiling
- balanced training
- structural context
- jury of networks

Profiling Homologous proteins

- same tertiary structure
- approximately same secondary structure

Add each aligned protein coding up Using sequence alignments of a family of proteins - HMM



Allows relationships between different proteins to be expressed in the coding

Increases accuracy by about 5%

Coding Coding amino acids is problematic

▶ Use a number 1-20

Coding

Coding amino acids is problematic

► Use a number 1-20 Gives a false representation 19 is not similar to 20

Coding

Coding amino acids is problematic

- ► Use a number 1-20 Gives a false representation 19 is not similar to 20
- Use chemical features

Coding

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- ► Use a number 1-20 Gives a false representation 19 is not similar to 20
- Use chemical features Should work better than it does.

Coding

Coding amino acids is problematic

- ► Use a number 1-20 Gives a false representation 19 is not similar to 20
- Use chemical features Should work better than it does.
- One hot encoding works very well

One hot encoding:

For words,

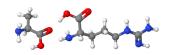
- $a \rightarrow [1, 0, 0, 0, 0, 0, 0, 0, \dots, 0]$
- $\begin{tabular}{ll} \bullet & \mbox{aa} \to \\ & [0,1,0,0,0,0,0,0,\dots,0] \end{tabular}$
- ► aardvark \rightarrow [0, 0, 1, 0, 0, 0, 0, 0, 0, ..., 0]
- ► aardwolf \rightarrow [0,0,0,1,0,0,0,0,...,0]





Similarly for amino acids, however only 20 amino acids makes this easier.

- Ala \rightarrow [1,0,0,0,0,0,0,0,...,0]
- $\begin{array}{c} \blacktriangle \text{ Arg} \to \\ [0,1,0,0,0,0,0,0,\dots,0] \end{array}$
- Asn \rightarrow [0,0,1,0,0,0,0,0,...,0]
- Asp \rightarrow [0,0,0,1,0,0,0,0,...,0]



Profiling - using one hot encodings and alignments of similar proteins

Protein	G	Υ	ı	Υ	D	Р	Α	V	G	D	
Alignments	G	Υ	I	Υ	D	Р	Ε	V	G	D	
	G	Υ	1	Υ	D	Р	Α	V	G	D	
	G	Υ	Ε	Υ	D	Р	Α	Ε	G	D	
	G	Υ	Ε	Υ	D	Р	Α	Ε	G	D	
G	5	0	0	0	0	0	0	0	5	0	
Α	0	0	0	0	0	0	4	0	0	0	
Р	0	0	0	0	0	5	0	0	0	0	
D	0	0	0	0	5	0	0	0	0	5	
Е	0	0	2	0	0	0	1	2	0	0	
V	0	0	0	0	0	0	0	3	0	0	
I	0	0	3	0	0	0	0	0	0	0	
Υ	0	5	0	5	0	0	0	0	0	0	
:	:	:	:	:	:	:	:	:	:	:	

Balanced Training

The distribution of secondary structure types is unbalanced:

- \triangleright 32% α helix
- \triangleright 21% β sheet
- ▶ 47% coil

These are sampled in equal quantities so in the training data:

- \triangleright 33.3% α helix
- \triangleright 33.3% β sheet
- ▶ 33.3% coil

This reduces performance over all slightly, but does increase accuracy on the α helix and coil structures.

Structural context:

- ho helices are typically 10 amino acids long
- \triangleright β sheets are typically 6 amino acids long

The prediction: $\alpha \alpha \alpha \beta \beta \alpha \alpha$ $\alpha \alpha$ makes no sense.

Should be $\alpha \alpha \alpha \alpha \alpha \alpha \alpha \alpha \alpha$

A structure to structure MLP is attached at the end to propagate this training signal back.

This is known as stacking. Gives 2 - 3% improvement

For example, β sheet

Secondary Structure prediction:

- Qian Sejnowski 1988 64.3%
- ► Rost and Sander 1993 69.7%

and..?

HMM and Beyond - PSI PRED

PSI BLAST:

- Position Specific Interative Basic Local Alignment Search Tool - PSI BLAST
- Derives a Position Specific Scoring Matrix PSSM
- ► This matrix is used to search for new matches
- PSSM is then updated
- repeated to give both near and distant relationships between proteins

PSI PRED:

- Predicts secondary structure based on PSI BLAST profiles
- uses MLP

Gets 77% accuracy

Secondary Structure prediction:

- Qian Sejnowski 1988 64.3%
- Rost and Sander 1993 69.7%
- ► PSI PRED 1999 77%

Little progress until 2010...

Next Lecture: Deep Learning

Multi class classification is better with cross entropy loss and softmax. Deals better with minority classes.

Why? - We want to maximise the likelihood of the correct classification given the data

Posterior probability for class k is:

$$P(C_k|\mathbf{x}) = \frac{P(\mathbf{x}|C_k)P(C_k)}{\sum_j P(\mathbf{x}|C_j)P(C_j)}$$

Given:

$$a_k = \ln P(\mathbf{x}|C_k)P(C_k) \approx f_{C_i}(\mathbf{x}_i|\theta)$$

$$P(C_k|\mathbf{x}) = \frac{\exp(a_k)}{\sum_j \exp(a_j)}$$

Softmax: Allows output to be interpreted as the probability of the classification

Cross Entropy Error

Maximising the relative entropy is maximising the log-likelihood of the data (over N patterns)

$$P(\mathbf{x}|\boldsymbol{\theta}) = \prod_{i=1}^{N} P(C_i|\mathbf{x}_i,\boldsymbol{\theta})$$

Take logs:

$$\log P(\mathbf{x}|\boldsymbol{\theta}) = \sum_{i=1}^{N} \log P(C_i|\mathbf{x}_i, \boldsymbol{\theta})$$

We approximate that probability using the output

$$\log P(\boldsymbol{x}|\boldsymbol{\theta}) = \sum_{i=1}^{N} \log f_{C_i}(\boldsymbol{x}_i|\boldsymbol{\theta})$$

Maximising the cross entropy we maximise the log probability. Gives more and better information to the machine than squared error loss.

HMM and Beyond - Summary

Machine learning tools used on language have been adapted for biological sequence analysis

- ► HMM still used today
- ► MLP need to tune correctly

tools to use:

- Cross entropy loss
- Soft max
- windowing
- profiling

Now deep learning predominates - next lecture