

My, how you've grown: a practical guide to modeling size transitions for Integral Projection Model (IPM) applications

Tom E.X. Miller^{*a} and Stephen P. Ellner^b

^aDepartment of BioSciences, Rice University, Houston, TX

^bDepartment of Ecology and Evolutionary Biology, Cornell University,
Ithaca, New York

Running header: Better growth modeling for IPMs

*Corresponding author. Department of BioSciences, Rice University, Houston, TX 77005-1827. Email:
tom.miller@rice.edu Phone: 713-348-4218

¹ **Abstract**

² 1.

³ 2.

⁴ 3.

⁵ 4.

⁶ **Keywords**

7 Introduction

8 Structured demographic models – matrix and integral projection models (MPMs and
9 IPMs) – are powerful tools for data-driven modeling of population dynamics and viabil-
10 ity that are widely used in basic and applied settings. In contrast to MPMs for popula-
11 tions with discrete structure (life stage, age class, etc.), IPMs (Easterling et al., 2000) read-
12 ily accommodate populations structured by continuous state variables, most commonly
13 size. A related innovation of the IPM framework is its emphasis on regression-based
14 modeling for parameter estimation, which carries important advantages for making the
15 most of hard-won data (Ellner et al., 2022).

16 A standard workflow allows ecologists to assemble an IPM from data using famili-
17 iar statistical tools to describe growth, survival, reproduction, and other demographic
18 transitions as functions of size (Coulson, 2012; Ellner et al., 2016). The relative ease of
19 the regression-based approach, accommodating multiple covariates (e.g., environmental
20 factors, experimental treatments) and complex variance structures (e.g., random effects,
21 correlated errors), has facilitated a growing body of IPM literature that examines how
22 biotic or abiotic factors affect population dynamics (e.g., Louthan et al., 2022; Ozgul
23 et al., 2010; Schultz et al., 2017) and explores the consequences of demographic hetero-
24 geneity associated with spatial, temporal, and individual variation (e.g., Compagnoni
25 et al., 2016; Crone, 2016; Plard et al., 2018). The vital rate regressions (or “sub-models”)
26 are the bridge between the individual-level data and the population-level model and its
27 predictions; it is important to get them right.

28 Compared to other vital rates, growth is special. The regression sub-models for
29 survival and reproduction provide the expected values of those rates as functions of
30 size (we use “size” as the name for whatever continuous variable defines the population
31 structure, which could instead be immune competence, mother’s weight, etc.). However,
32 for modeling growth, the full probability distribution of subsequent size, conditioned on
33 initial size, must be defined. This distribution defines the growth ‘kernel’ $G(z', z)$ that
34 gives the probability density of any future size z' at time $t + 1$ conditional on current size
35 z at time t . Whenever survival and reproduction are size-dependent, the entire distribu-
36 tion of size transitions can strongly influence IPM predictions because this distribution
37 governs how frequently size changes are much greater or much lower than average.

38 The original template for modeling size transitions in IPMs was provided by East-
39 erling et al. 2000. They first tried simple linear regression, assuming normally dis-
40 tributed size changes with constant variance. Because the residuals from this regression
41 exhibited non-constant variance, they used a two-step approach that estimated the size-

42 dependence in the growth variance (better options soon became available, such as the
43 `lme` function in R). However, even after accounting for non-constant variance, growth
44 data may still deviate from the assumption that size transitions are normally distributed.
45 Size transitions are often skewed such that large decreases are more common than large
46 increases (Peterson et al., 2019; Salguero-Gómez and Casper, 2010), or vice versa (Stub-
47 berud et al., 2019). Size transitions may also exhibit excess kurtosis ('fat tails'), where
48 extreme growth or shrinkage is more common than predicted by the tails of the normal
49 distribution (Hérault et al., 2011).

50 The observation that the normal distribution may poorly describe size transitions
51 in real organisms has been made before, and several studies have emphasized that al-
52 ternative distributions should be explored (Easterling et al., 2000; Peterson et al., 2019;
53 Rees et al., 2014; Williams et al., 2012). Yet, default use of Gaussian growth distribu-
54 tions (often with non-constant variance) remains the standard practice. An ISI Web of
55 Knowledge search on the terms 'integral projection model ecology' (DATE) returned #
56 IPM studies published in the past decade (2010–2020), # of which assumed a Gaussian
57 growth kernel.¹ The general state-of-the-art in the literature appears to remain where it
58 was 20 or so years ago, using the default model without pausing to examine critically
59 whether or not it actually provides a good description of the data. We are guilty of this,
60 ourselves.

61 The persistence of Gaussian growth modeling is understandable. There is a long
62 tradition of statistical modeling built on the assumption of normally distributed residu-
63 als with constant variance. Popular software packages such as `lme4` (Bates et al., 2007)
64 and `MCMCglmm` (Hadfield et al., 2010) make it easy to fit growth models with po-
65 tentially complex fixed- and random-effect structures, but the possible distributions of
66 continuous responses are limited, and default to Gaussian. Abandoning these conve-
67 nient tools for the sake of more flexible growth modeling means, it may seem, sacrificing
68 the flexibility to rigorously model diverse and potentially complex sources of variation
69 in growth, some of which may be the motivation driving the study in the first place.

70 The question we address here is: how can ecologists escape the apparent trade-off
71 between realistically capturing the variance, skew, and kurtosis of size transition data
72 on the one hand, and flexibly including the multiple covariates and random effects that
73 often have substantial impacts on demographic rates. In this article, we offer an answer.

74 Our goal here is to present and illustrate a general 'recipe' that moves growth mod-
75 eling past the standards set over 20 years ago. Like any recipe, users may need to
76 make substitutions or add ingredients to suit their situation. Our approach emphasizes

¹I still intend to do this! But it's a rabbit hole I have not gone down yet.

77 graphical diagnostics for developing and evaluating growth models, rather than a pro-
78 cess centered on statistical model selection. Through a set of empirical case studies we
79 demonstrate how a simple workflow, using tools that were nonexistent or not readily
80 available when IPMs first came into use, makes it straightforward and relatively easy to
81 identify when the default model is a poor fit to the data, and to then choose and fit a
82 substantially better growth model that is no harder to use in practice. We illustrate our
83 approach by revisiting four of our own, mostly published IPM analyses that assumed
84 Gaussian growth.² In each case, the Gaussian assumption does not stand up to close
85 scrutiny. We illustrate how we could have done better, and the consequences of “doing
86 better” for our ecological inferences. All of our analyses may be reproduced from code
87 and data that are publicly available (see Data accessibility statement).

88 A general workflow for better growth modeling

89 The modeling workflow that we suggest runs as follows (Fig. 1):

- 90 1. *Fit a “pilot” model or models assuming a Gaussian distribution but allowing for non-*
91 *constant variance.*

92 This step is familiar to most IPM users, as it is the start and end of the traditional
93 workflow. A well-fitted Gaussian model accurately describes the mean and variance
94 of future size conditional on current size and possibly on other measured covariates
95 or random effects. This step may include model selection to identify which treat-
96 ment effects or environmental drivers affect the mean and/or variance of future size.
97 Non-constant variance is often fitted in a two-stage process, first fitting mean growth
98 assuming constant variance, then doing a regression relating the squared residuals
99 from the initial fit to the fitted mean. It is sometimes better to fit size-dependence
100 in the mean and variance simultaneously, as can be done with the R packages **mgcv**
101 and **nmle**, because incorrectly assuming constant variance can affect the outcome of
102 model selection for the mean. One-step fitting is straightforward for simple models
103 in which initial size is the only factor that can influence growth variance. However,
104 the two-step process fitting residuals to the fitted value (expected future size) may
105 be convenient when there are multiple fixed and random effects, all of which may
106 contribute to non-constant variance, since the expected value implicitly accounts for
107 all of them. We illustrate both one-step and two-step approaches in the examples
108 below.

²Need to commit to case study choices - Steve wanted to include corals for contrast with Peterson et al.

109 Allowing non-constant variance means that it is not necessary to transform the
110 data in a way that stabilizes the growth variance. Transformation remains an option
111 when it does not create new problems (see Discussion), and it may have advantages
112 besides variance stabilization. In particular log-transformation is often appropriate
113 for size data (Ellner et al., 2016), and it helps avoid eviction at small sizes.

- 114 2. *Use statistical and graphical diagnostics to identify if and how the standardized residuals*
115 *deviate from Gaussian, and to identify a more appropriate distribution.*

116 If the Gaussian pilot model is valid, the set of standardized residuals (standardized
117 by the standard deviation) should be Gaussian with mean zero and unit variance,
118 with no skew or excess kurtosis. This criterion provides a straightforward test for
119 whether to accept a Gaussian growth model or explore alternatives. If the standard-
120 ized residuals are satisfactorily Gaussian, skip to the final step of the workflow.

121 There are many ways that growth data may deviate from Gaussian, and the
122 nature of those deviations can guide the search for a better distribution. Frequentist
123 tests such as the D'Agostino test of skewness (D'Agostino, 1970) and the Anscombe-
124 Glynn test of kurtosis (Anscombe and Glynn, 1983) could be used to diagnose
125 whether the aggregate distribution of standardized residuals deviates from normal-
126 ity (R package **moments** (Komsta and Novomestky, 2015)). However, the aggregate
127 distribution of standardized residuals may be misleading if properties such as skew
128 and kurtosis vary with size. For example, a change in the direction of skewness from
129 small to large sizes would require a distribution flexible enough to accommodate
130 both positive and negative skew, such as the skewed normal or Johnson S_U distri-
131 butions. Alternatively, growth data may lack skew but may exhibit leptokurtosis (in
132 which case the t distribution may be a good choice) or may shift from platykurtos-
133 sis to leptokurtosis depending on initial size (in which case the power exponential
134 distribution may be a good choice). It is therefore essential to visualize trends in dis-
135 tribution properties with respect to size, either initial size (for simple models with
136 only size-dependence) or expected future size (for models with multiple fixed ef-
137 fects). In the case studies below, we rely on quantile regression of the standardized
138 residuals to visualize skew and kurtosis as continuous functions of size or expected
139 value. Fig. 1 includes guidance on how the skew and kurtosis properties of the stan-
140 dardized residuals suggest options for an appropriate growth distribution. In our
141 case studies we take advantage of the many distributions provided in the **gamlss** R
142 package (Stasinopoulos et al., 2007), but any other distributions with the necessary
143 properties can be used.

144 3. *Refit the growth model using the chosen distribution.*

145 In models with multiple covariates and/or random effects, each potentially affecting
146 several distribution parameters (location, scale, skew, kurtosis) in different ways,
147 “refit the model” could entail a massive model selection process to identify the
148 “right” or “best” non-Gaussian model. And with so many options, model uncer-
149 tainty may be overwhelming and over-fitting becomes a significant risk even if pre-
150 cautions against it are taken. We therefore argue for adopting the more modest
151 goal of remedying the apparent defects in the Gaussian model. Conveniently, as
152 we demonstrate below, the functional forms for the mean and standard deviation
153 (or location and scale parameters) could be carried over from the pilot Gaussian
154 model into a non-Gaussian distribution, leaving skew and kurtosis as the targets for
155 improvement. This step exploits the fact that parameter estimation from a Gaus-
156 sian model is generally robust to deviations from normality (Schielzeth et al., 2020),
157 meaning that the mean of the Gaussian model is probably a good proxy for the mean
158 of the non-Gaussian model (and in case it is not, the next step in the workflow would
159 catch that). The functional forms for skew and kurtosis of the non-Gaussian model
160 can be guided by the qualitative features of the graphical diagnostics (e.g., skewness
161 switches from positive to negative with size).

162 4. *Test the final model through graphical diagnostics comparing simulated and real growth data.*

163 A good model will generate simulated data that look like the real data. Again, it is
164 important to inspect the properties of simulated data conditional on present size or
165 expected future size, rather than examining the entire distribution. We provide ex-
166 amples below of informative comparisons between simulated and real data, based
167 mainly on quantiles. If the simulated data do not correspond well with real data,
168 alternative (possibly more flexible) growth distributions should be explored, or more
169 complex functions relating distribution parameters to current size and other covari-
170 ates. However, we again caution against launching a full-blown model selection
171 exercise. Instead, possible alternative models could be chosen primarily to remedy
172 observable discrepancies between the real and simulated size transition data, and at
173 most slightly modified based on final diagnostic and statistical tests.

174 How should skewness and kurtosis be measured?

175 “Improvement” of a Gaussian model will always involve scrutiny of skewness and kur-
176 tosis, so measurement of these properties warrants some attention. The standard mea-



Figure 1: General workflow of recommendations for IPM growth modeling (left) and guide to common non-Gaussian distributions of size x for $x \in \mathbb{R}$ that can accommodate different combinations of skewness and kurtosis (right). All of these distributions (often including multiple versions or parameterizations of each) are available in the package **gamlss.dist**, except for the skewed generalized *t*, which is available in the package **sgt** (Davis, 2015).

¹⁷⁷ sures of skewness and kurtosis (tail thickness) are based on the third and fourth central
¹⁷⁸ moments, respectively, of the distribution:

$$\text{Skewness} = \frac{m_3}{\sigma^3}, \quad \text{Excess kurtosis} = \frac{m_4}{\sigma^4} - 3 \quad (1)$$

¹⁸⁰ where $m_k = \mathbb{E}(X - \bar{X})^k$ is the k^{th} central moment of a random quantity X and σ^2 is the
¹⁸¹ variance (second central moment). A Gaussian distribution has zero skewness and zero
¹⁸² excess kurtosis.

¹⁸³ The standard measures are easy to calculate but their use for choosing and eval-
¹⁸⁴ uating growth models is hindered by their poor sampling properties. Because empirical
¹⁸⁵ estimates involve high powers of data values, it only takes a few outliers to produce



Figure 2: Histograms of skewness and kurtosis estimates using moment-based definitions, compared with the nonparametric measures. Histograms are based on 5000 replicate draws of a sample of 200 independent values from a t distribution with 8 degrees of freedom. Dotted red vertical lines mark the minimum and maximum of sample estimates, and dashed blue lines show the 5th and 95th percentiles. The true value is indicated by a black dot on the x -axis. Figure drawn by script `NPmoments.R`

186 a very inaccurate estimate. Figure 2 shows a simulated example, where the underlying
 187 “data” are a sample of size 200 from a t distribution with 8 degrees of freedom; the true
 188 skew is 0, and the true excess kurtosis is 1.5. The distance between the largest and small-
 189 est estimates (indicated by the dotted red vertical lines), relative to the distance between
 190 the 5th and 95th percentiles, shows the broad extent of extreme values that can occur
 191 even with a good size sample, especially for kurtosis.

192 We therefore use “nonparametric” (NP) measures of skew and kurtosis that are
 193 based on quantiles and thus less sensitive to a few extreme data values. Let q_α denote
 194 the α quantile of a distribution or sample (e.g., $q_{0.05}$ is the 5th percentile). For any
 195 $0 < \alpha < 0.5$, a quantile-based measure of skewness is given by (McGillivray, 1986)

$$196 \text{NP Skewness} = \frac{q_\alpha + q_{1-\alpha} - 2q_{0.5}}{q_{1-\alpha} - q_\alpha}. \quad (2)$$

197 NP Skewness is a measure of asymmetry between the tails of the distribution above and
 198 below the median. The size of the upper tail can be measured (for any $0 < \alpha < 0.5$) by
 199 $\tau_U = q_{1-\alpha} - q_{0.5}$; for $\alpha = 0.05$ this is the difference between the 95th percentile and the
 200 median. The lower tail size is $\tau_L = q_{0.5} - q_\alpha$. The definition above is equivalent to

$$201 \quad \text{NP Skewness} = \frac{\tau_U - \tau_L}{(\tau_U + \tau_L)}. \quad (3)$$

202 So an NP Skewness of ± 0.2 says that the difference in tail sizes is 20% of their total. The
 203 range of possible values is -1 to 1. Both $\alpha = 0.25$ (sometimes called “Kelly’s skewness”)
 204 and $\alpha = 0.1$ (“Bowley’s skewness”) are common choices. We used $\alpha = 0.1$, unless
 205 otherwise stated.

206 An analogous quantile-based measure of kurtosis (Jones et al., 2011) is

$$207 \quad \text{NP Kurtosis} = \frac{q_{1-\alpha} - q_\alpha}{q_{0.75} - q_{0.25}}. \quad (4)$$

208 For $\alpha = 0.05$, NP Kurtosis is the difference between the 95th and 5th percentiles, relative
 209 to the interquartile range. To facilitate interpretation, we scale NP Kurtosis relative to
 210 its value for Gaussian distribution, and subtract 1. We call this “NP Excess Kurtosis”.
 211 The value for a Gaussian distribution is zero. A value of 0.2 means that the tails are (on
 212 average) 20% heavier than those of a Gaussian with the same interquartile range, and
 213 a value of -0.2 means that the tails are (on average) 20% lighter than a Gaussian with
 214 the same interquartile range. We calculate NP Kurtosis using $\alpha = 0.05$ unless otherwise
 215 stated, to focus on the tail edges, but again this is somewhat arbitrary.

216 Figure 2C,D illustrate how, applied to exactly the same simulated samples, the non-
 217 parametric measures of skewness and kurtosis produce a smaller fraction of highly in-
 218 accurate estimates caused by a few extreme values in the sample. But also note that, in
 219 contrast to the moment-based measures, numerically small values of the NP measures
 220 (e.g., 0.1 or 0.2) should not be disregarded, because they are both scaled so that a value
 221 of 1 indicates extremely large departures from a Gaussian distribution.

222 Quantile-based estimation of skewness and kurtosis carries the added value that
 223 quantile regression methods may be used to derive these properties of size transitions
 224 as continuous functions of initial size or expected future size. In the examples below,
 225 we use the **qgam** package to fit smooth additive quantile regression models, which have
 226 the flexibility to accommodate non-linear size-dependence in skewness and kurtosis.
 227 One risk of a gam-based approach is that fitted quantiles may be too “wiggly” without
 228 constraints on their complexity (in the examples below, we specify $k = 4$ to constrain the

dimension of the basis function). For the gam-averse, other quantile regression models may be equally suitable. For consistency with non-parametric skewness and kurtosis, we similarly use quantile-based measures of mean and variance and quantile regression to visualize these as functions of size. Specifically, following Wan et al. (2014),

$$\text{NP Mean} = \frac{q_{0.25} + q_{0.5} + q_{0.75}}{3} \quad (5)$$

and

$$\text{NP SD} = \frac{q_{0.75} - q_{0.25}}{1.35}. \quad (6)$$

1 Case study: Sea fan corals, *Gorgonia ventalina*

We begin with a simple example where current size is the only predictor of future size. Bruno et al. (2011) developed an IPM to understand the rise and fall of a fungal pathogen *Aspergillus sydowii* in Caribbean sea fan corals *G. ventalina*. The model was based on repeated observations of marked corals in permanent transects at several sites near Akumal, Mexico, recording disease status (infected/uninfected) and the area of uninfected tissue. The epidemic peak had passed and disease incidence was already low, so infected fans were relatively infrequent. We therefore limit the analysis here to uninfected individuals. Bruno et al. (2011) found statistically significant year and site effects, but as those explained a very small fraction of the variation in growth increments, they fitted a single growth model to data pooled across years and sites. We do the same here. The pooled data set consists of 358 observed size transitions. The data exhibited size-dependent variance in growth (change in area, cm^2), which Bruno et al. (2011) chose to stabilize by transforming size, using the cube-root of total fan area as the size measure (fig. ??B), and then fitting the standard model with Gaussian growth increments. Here we take a different approach, modeling size-dependent variance explicitly rather than trying to transform it away.

We develop a model using natural log transformation of area. With initial size as the only predictor, a simple way to fit a Gaussian model with nonconstant variance is the `gam` function in `mgcv` library (Wood, 2017) using the `gaulss` family. The mean and standard deviation are both fitted as smoothing spline functions of initial size, and the `predict` function returns the fitted mean and also the inverse of the fitted standard deviations with which we can compute the scaled residuals:

```
# XH is a data frame holding the data
# logarea.t0, .t1 denote initial and final values of log-transformed area
```

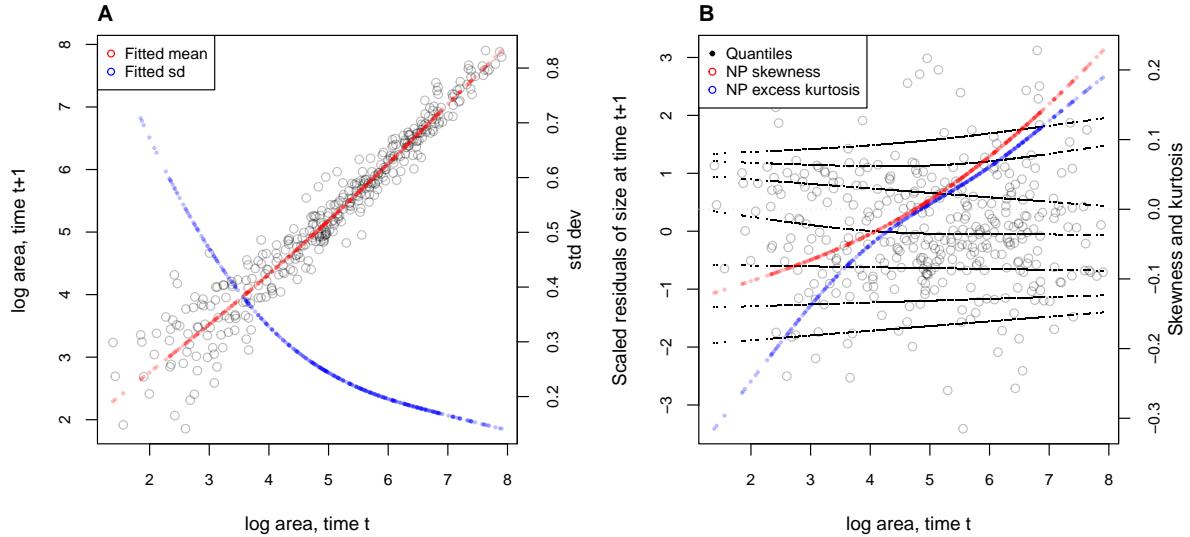


Figure 3: **A**, Size transition data for sea fan corals, *Gorgonia ventalina*, and fitted gam with mean (red) and standard deviation (blue) of future size conditional on current size. **B**, Quantile regressions of scaled residuals on size and nonparametric estimates of skewness (red) and excess kurtosis (blue) derived from them. Black lines in **B** show the 5th, 10th, 25th, 50th, 75th, 90th, and 95th quantiles. Figure made by script AkumalCorals_qgam.R.

```

261 fitGAU <- gam(list(logarea.t1~ s(logarea.t0), ~ s(logarea.t0)),
262   data=XH, gamma=1.4, family=gaulss())
263 fitted_all = predict(fitGAU,type="response");
264 fitted_sd = 1/fitted_all[,2];
265 scaledResids = residuals(fitGAU,type='response')/fitted_sd;

```

Fig. 3A shows the log-transformed data and Gaussian model. The mean function (solid blue curve) is visually nearly linear, but the fitted nonlinear spline is strongly favored over a linear model for the mean ($\Delta AIC \approx 9$). The spline for standard deviation σ versus initial size shows that smaller individuals exhibit greater variability in future size.

There are no blatant signs of trouble in the pilot Gaussian model, but quantile regressions on the scaled residuals, and the NP Skewness and Kurtosis metrics derived from them (Eq. 3 and 4), suggest deviations from normality (Fig. 3B). Specifically, skewness switches from negative to positive across the size distribution, with smaller corals more likely to shrink than grow and larger corals more likely to grow than shrink. Kurtosis also changes direction over the size distribution, with smaller initial sizes having thinner tails and larger initial sizes having fatter tails than Gaussian. The fitted nonparametric moments suggest that the upper and lower tails of size transition proba-

278 bilities may differ by up to 20%, and the weight of the tails may be 20% greater or less
279 than Gaussian, depending on initial size – not overwhelming deficiencies, but not trivial
280 either. Are these deviations from normality severe enough to warrant a second, non-
281 Gaussian iteration of growth modeling? This question may be answered by simulating
282 data from the Gaussian model and examining whether key properties of the simulated
283 data are consistent with those of the real data – this is the ultimate litmus test for a
284 growth model’s adequacy and should be a standard element of IPM construction, in our
285 opinion. If the simulated data are not consistent with the real data, it is time to choose
286 a better distribution (Fig. 1). In this case, the negative skew at small sizes and excess
287 kurtosis observed at large sizes are more extreme than what occurs across 100 random
288 iterations of data simulation (Fig. 4), suggesting that, for at least some parts of the size
289 distribution, a non-Gaussian model would better capture size transitions.

290 We sought a distribution that could accommodate the properties of the scaled resid-
291 uals, specifically changes in the sign of skewness and excess kurtosis across initial sizes.
292 We chose the sinh-arcsinh (SHASH) distribution, a four-parameter distribution that, con-
293 veniently, is included in **mcmc**’s **gam()** function:

```
294 fitSHASH <- gam(list(logarea.t1 ~ s(logarea.t0,k=4), # <- location  
295 ~ s(logarea.t0,k=4), # <- log-scale  
296 ~ s(logarea.t0,k=4), # <- skewness  
297 ~ s(logarea.t0,k=4)), # <- log-kurtosis  
298 data = XH, family = shash, optimizer = "efs")
```

299 Data simulated from this model are more consistent with the real data than the Gaussian
300 model: many of the 100 simulated SHASH data sets exhibited negative skew at small
301 sizes and positive excess kurtosis at large sizes that were as strong or stronger than
302 observed in the real data (Fig. 4). If one cared to quantify the difference between models,
303 the SHASH is clearly favored by AIC despite having twice as many parameters as the
304 Gaussian ($\Delta AIC = 7.04$).

305 What, then, have we gained by fitting a better growth model? Fig. 5A compares
306 the predicted distributions of subsequent size in the fitted model and Gaussian pilot
307 models, for the median size of a new recruit (leftmost pair of curves), the median initial
308 size (central curves), and the 95th percentile of initial size in the data (rightmost curves).
309 The differences are small, and most pronounced for the smallest size, where recruits
310 are predicted to grow slightly larger under the SHASH model than the Gaussian model.
311 The direction of this difference was surprising, since the SHASH accommodates negative
312 skew at small sizes in the data. However, in modeling skew appropriately, the SHASH

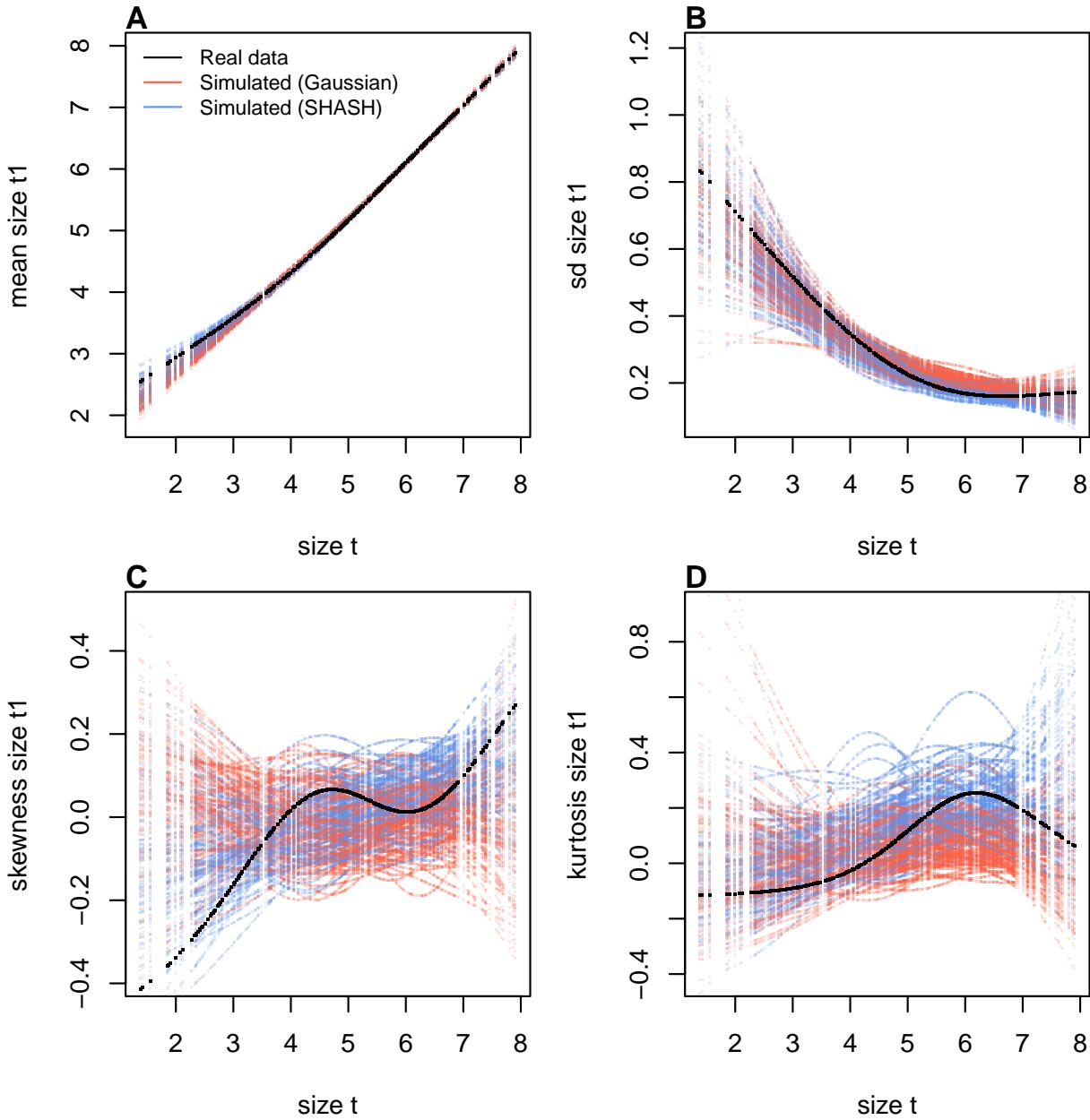


Figure 4: Comparisons among real coral data and data simulated from Gaussian and SHASH growth models for mean, standard deviation, NP skewness, and NP kurtosis of future size conditional on current size. Figure made by script `AkumalCorals_qgam.R`.

model also gives a better prediction for mean growth at small sizes than the Gaussian model, whose mean is biased downward by negative skew (Fig. 4A)³. Something similar happens in the standard deviation at large sizes (log size 5–7), where excess kurtosis in the data biased the SD upward (Fig. 4B). Fig. 5B shows the predicted steady-state size

³...Contradicting the earlier assertion that parameter estimates from Gaussian models are robust to deviations from normality!

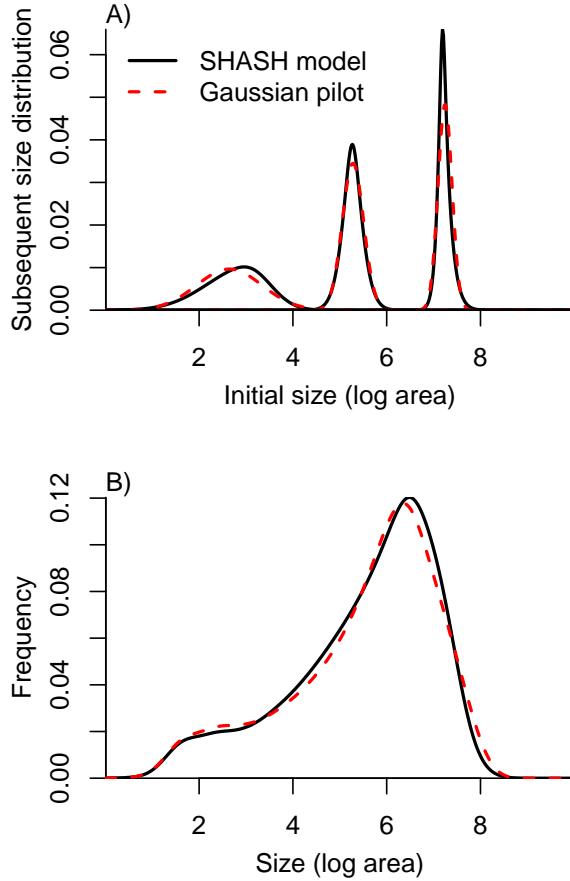


Figure 5: Comparisons between the fitted SEP1 growth model (solid black curves) and the Gaussian pilot model (dashed red curves) for sea fans *G. ventalina*. A) Predicted frequency distributions of size in year $t + 1$ for three different values of size in year t . The leftmost pair of curves are for initial size equal to the median size of a new recruit (data from (Bruno et al., 2011)); central pair is for the median initial size of uninfected individuals; rightmost pair are for the 95th percentile initial size for uninfected individuals. B) Steady-state size distributions resulting from a constant unit input of new recruits. As in Bruno et al. (2011) we assume that larvae are very widely dispersed, so that the number of recruits arriving at any one location is independent of the local population abundance or structure. Size distribution of recruits was described by a kernel density estimate based on the (sadly, only $n = 9$) measured sizes of known new recruits. Figure made by script AkumalCoralsIPMs.R.

317 distributions resulting from a constant unit input of recruits. Again, the differences are
 318 very subtle. Finally, the Gaussian and SHASH growth models predict very similar mean
 319 life span (17.7 and 17.9 years, respectively). From these outputs, there is little evidence
 320 that improved modeling of coral growth meaningfully improved biological inferences
 321 from the IPM; one could argue that it was not worth the trouble.

322 In this case study we used `gam` to fit both the Gaussian and SHASH models because
323 that obviated model selection on functions for mean, variance, and higher moments.
324 However, `gam` should be used with caution. Nonparametric regression models notori-
325 ously “wag their tails” because the ends of the fitted curve can be pulled close to the
326 outermost data points. This is especially problematic for growth modeling, because data
327 are typically sparse near the bounds of the size distribution. To minimize the risk of
328 overfitting we specified the number of “knots” (`k=4`) and used `gamma=1.4` to overweight
329 model degrees of freedom, as suggested by Gu (2013, sec. 3.2). But it is always impor-
330 tant to plot the fitted splines and make sure they do not wag unrealistically. If they do,
331 parametric regression may be a better choice.

332 2 Case study: tree cholla cactus, *Cylindriopuntia imbricata*

333 The next case study, focusing on the tree cholla cactus *Cylindriopuntia imbricata* at the
334 Sevilleta Long-Term Ecological Research site in central New Mexico, adds a new feature
335 on top of the simple size-dependent regressions in the previous study: random effects
336 associated with temporal (year) and spatial (plot) environmental heterogeneity. This
337 long-term study of cactus demography was initiated in 2004 and different subsets of
338 the data have been analyzed in various IPM studies, all using Gaussian growth kernels
339 (Compagnoni et al., 2016; Czachura and Miller, 2020; Elderd and Miller, 2016; Miller
340 et al., 2009; Ohm and Miller, 2014). In fact, (Elderd and Miller, 2016) presented a Gaus-
341 sian growth model fit to the cactus data as an example of a well fit growth function,
342 based on a marginal distribution of residuals that appeared approximately Gaussian
343 and posterior predictive checks (PPCs) of a Bayesian model that suggested consistency
344 between the real data and data simulated from the fitted model (Fig. 4 in (Elderd and
345 Miller, 2016)). While PPCs and the associated “Bayesian P-value” are popular diagnostic
346 tools, they are often considered to be too conservative (Conn et al., 2018; Zhang, 2014),
347 failing to reject marginally bad models even though they are very effective in rejecting
348 models that are terrible. The choice of discrepancy function (the statistic used to com-
349 pare real and simulated data) can also be limiting: in our previous work, we used a
350 discrepancy function focused on variance (the sum of the squared residuals), so we had
351 a built-in blind-spot for mismatches in higher moments. In the clarity of hindsight, the
352 PPC gave a false sense of security; the Gaussian was a poor choice all along.

353 The data for this new analysis include 4844 size transition observations from 929 in-
354 dividuals spanning 13 transition years (2004–2018) and 11 spatial replicates (three spatial
355 blocks in years 2004–2008 and eight 30m-by-30m plots in years 2009–2018). The data are
356 provided in Miller (2020). Following previous studies, we quantified size as the natural
357 logarithm of plant volume (cm^3), derived from height and width measurements.

358 We begin the growth modeling workflow, as above, with a generalized additive
359 model with the mean and standard deviation of size in year $t + 1$ modeled as function
360 of size in year t , with random intercepts for year and plot and assuming normally dis-
361 tributed residuals (`family=gaulss()`). The standardized residuals, accounting for size-
362 dependent residual variance (Fig. 6A), show clear signals of negative skew and positive
363 excess kurtosis across most of the size distribution but strongest in the middle of the size
364 distribution (Fig. 6B).

365 To better capture size transitions, we need a distribution with negative skew and
366 positive excess kurtosis, but both of which may be negligible at some sizes. We first tried

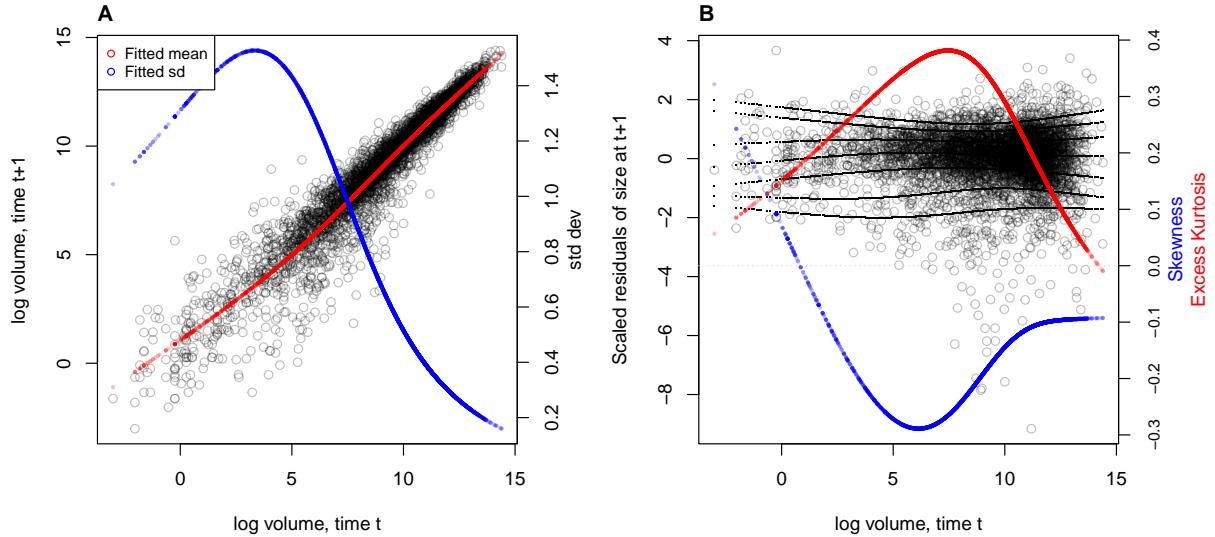


Figure 6: **A**, Size transition data for tree cholla cacti, *Cylindropuntia imbricata*, and fitted gam with mean (red) and standard deviation (blue) of future size conditional on current size. **B**, Quantile regressions of scaled residuals on size and nonparametric estimates of skewness (red) and excess kurtosis (blue) derived from them. Black lines in **B** show the 5th, 10th, 25th, 50th, 75th, 90th, and 95th quantiles. Figure made by script `cactus_growth_modeling_qgam.R`.

367 Johnson's S_U and then the skewed t distributions, both of which are limited to positive
 368 excess kurtosis. Both distributions provided some improvement over the Gaussian, but
 369 were not happy with the fit of either. Iterating through the workflow (Fig. 1), we ar-
 370 rived, again, at the SHASH distribution, which is more flexible than either the JSU or
 371 skewed t , capable of capturing a greater range of kurtosis for a given amount of skew,
 372 and vice versa (Steve's NPSkewKurtosisRanges.pdf). Furthermore, fitting the SHASH
 373 as a generalized additive model with `mgcv` allowed for flexible, non-monotonic size-
 374 dependence in skewness and kurtosis without the need for model selection on specific
 375 size-dependent functions; through iterations of trial and error, we found this flexibility
 376 was necessary to generate simulated data that compared favorably to the real data. The
 377 other distributions that we tried are not available as `mgcv` families, so we fit these with
 378 custom maximum likelihood functions, an approach we illustrate in the next case study.
 379 The final growth model was similar to the SHASH gam in the coral case study, but
 380 with random intercepts for the location parameter, representing spatial and temporal
 381 heterogeneity:

```
382 fit_shash <- gam(list(logvol_t1 ~ s(logvol_t,k=4) +  

  383 s(plot,bs="re") + s(year_t,bs="re")), # <- model for locat
```

```

384 ~ s(logvol_t,k=4), # <- model for log-scale
385 ~ s(logvol_t,k=4), # <- model for skewness
386 ~ s(logvol_t,k=4)), # <- model for log-kurtosis
387 data = CYIM_grow,
388 family = shash,
389 optimizer = "efs")

```

390 The final SHASH model provided good correspondence between simulated and
391 real data, and provided more compelling improvement over the Gaussian model than
392 we saw in the coral case study (Fig. 7). The SHASH model over-estimated negative
393 skew at some sizes relative to the signal of skewness in the data (Fig. 7C), but the nature
394 of size-dependent skew in the data is not very biologically plausible and may instead
395 be driven by the tail-wagging tendency of gams. As in the coral case study, we see
396 that correctly modeling skewness and kurtosis improved estimation of the mean and
397 standard deviation (Fig. 7A,B), yielding a growth model that is clearly truer to the data
398 than the pilot Gaussian fit.

399 We explored how improved growth modeling influenced IPM results, leveraging
400 the plot and year structure of the study design to quantify spatial and temporal vari-
401 ance in fitness. The structure and parameterization of the full IPM are described in
402 Appendix XX. We used the fitted random effects from the vital rate models to estimate
403 the asymptotic growth rate for each year (λ_t), centered on the average plot, and for
404 each plot (λ_p), centered on the average year. This allowed us to quantify demographic
405 variance associated with temporal and spatial heterogeneity. We found that the Gaus-
406 sian growth model tended to over-estimate λ_t , particularly in the harshest years (Fig.
407 8A), and thus under-estimated temporal variance in fitness ($Var(\lambda_{t(Gaussian)}) = 0.0018$,
408 $Var(\lambda_{t(SHASH)}) = 0.0023$). The opposite was true for plot-to-plot variation (Fig. 8B),
409 where the Gaussian model under-estimated λ_p and over-estimated spatial variance in
410 fitness ($Var(\lambda_{p(Gaussian)}) = 0.00015$, $Var(\lambda_{p(SHASH)}) = 0.000088$). Across both growth
411 models, fluctuations in fitness were stronger through time than across space. The
412 difference in temporal variance would suggest that Gaussian growth modeling would
413 lead to over-estimation of the stochastic growth rate λ_S , since temporal variance has
414 a negative influence on λ_S . However, this was not the case: stochastic IPMs based
415 on Gaussian and SHASH growth models had nearly identical stochastic growth rates
416 ($\lambda_S(Gaussian) = 0.9906$, $\lambda_S(SHASH) = 0.9909$). This is likely because temporal fluctu-
417 ations in vital rates, which is where the SHASH growth model would make a difference,
418 have a weaker influence on λ_S than the temporal fluctuations in size structure that they

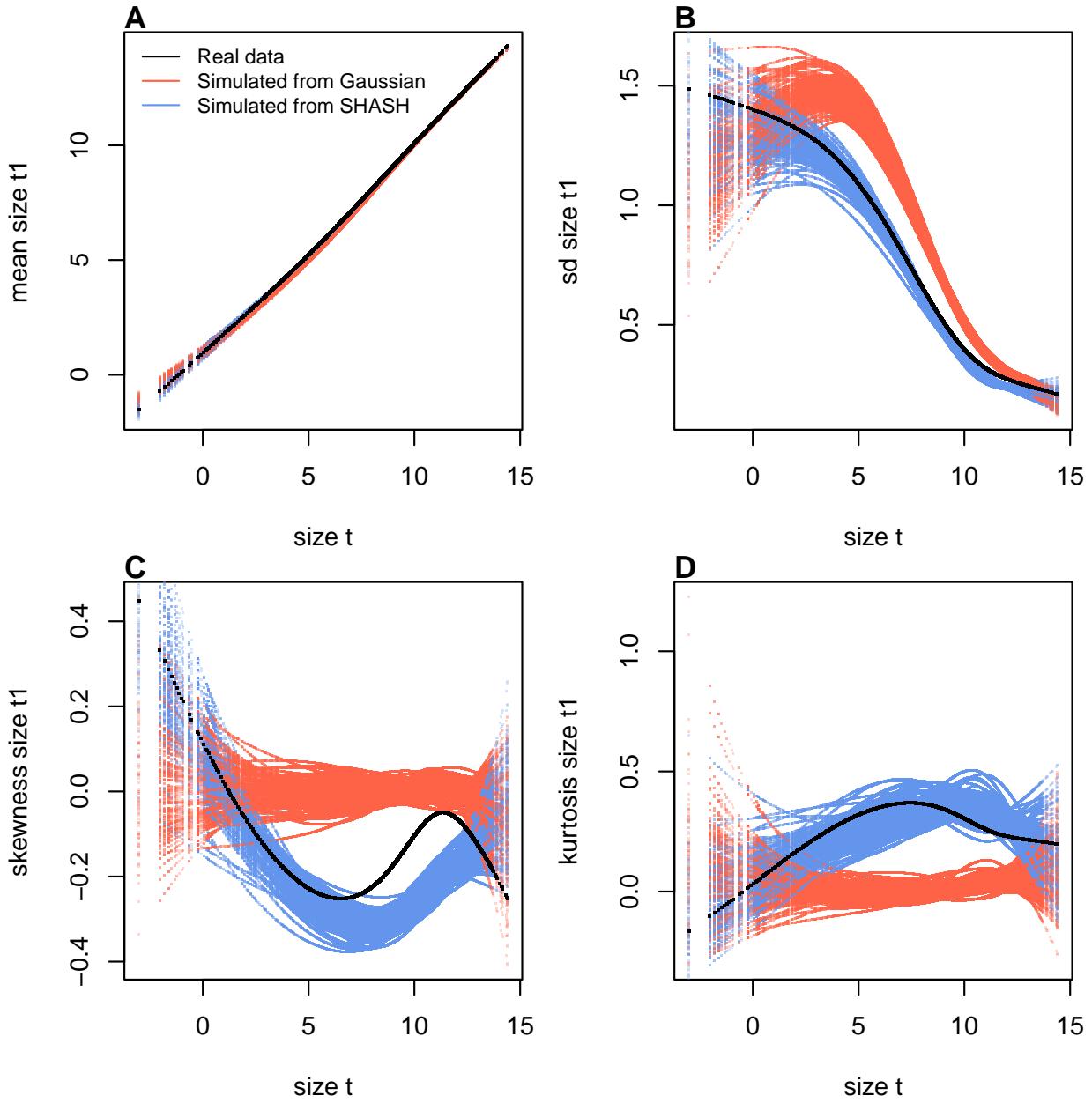


Figure 7: Comparisons among real cactus data and data simulated from Gaussian and SHASH growth models for mean, standard deviation, NP skewness, and NP kurtosis of future size conditional on current size. Figure made by script `cactus_growth_modeling_qgam.R`.

⁴¹⁹ generate (Compagnoni et al., 2016; Ellis and Crone, 2013). Thus, depending on the target
⁴²⁰ of one's analysis, modeling non-Gaussian size transitions with a Gaussian growth model
⁴²¹ could bias results in either direction, or make no difference at all.

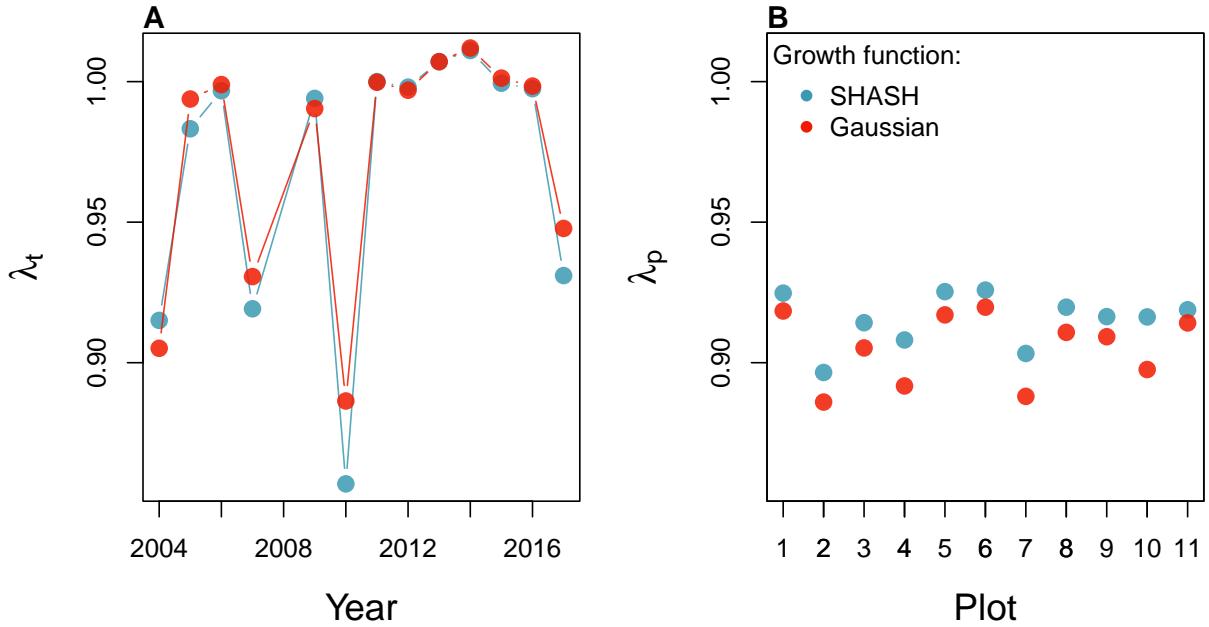


Figure 8: .

⁴²² 3 Case study: creosotebush, *Larrea tridentata*

⁴²³ Our next case study comes from our studies of the woody shrub creosotebush (*Larrea tri-*
⁴²⁴ *dentata*) at the Sevilleta Long-Term Ecological Research (LTER) site in central New Mex-
⁴²⁵ ico, US. At this site as elsewhere in the Southwest US, creosotebush is encroaching into
⁴²⁶ desert grassland habitats. The data described here were collected along transects span-
⁴²⁷ ning grass-shrub ecotones to understand patterns of density dependence in creosotebush
⁴²⁸ demography. Specifically, we asked whether fitness is maximized approaching zero den-
⁴²⁹ sity at the leading edge of the expansion front (consistent with ‘pulled’ expansion), or
⁴³⁰ whether there is a demographic advantage for shrubs at higher density due to positive
⁴³¹ feedbacks expected for ecosystem engineers (leading to ‘pushed’ expansion). Our pub-
⁴³² lished study (Drees et al., 2023) used a spatial integral projection model (SIPM) to predict
⁴³³ the speed of shrub encroachment, assuming normally-distributed size transitions. Here
⁴³⁴ we step through our suggested workflow to ask whether a non-Gaussian model would
⁴³⁵ have been more faithful to the data, and how such an improvement would influence
⁴³⁶ predictions for the speed of encroachment. We use this case study to illustrate several
⁴³⁷ new elements and challenges, including modeling skewness and kurtosis as functions
⁴³⁸ of expected future size (instead of initial size) and using distributions that are not cur-
⁴³⁹ rently available as **mgcv** families. In fact, to diversify our use of software and illustrate
⁴⁴⁰ alternatives, we do not use gam’s for any element of this case study.

441 Growth data come from 522 shrubs censused longitudinally over four years (2013-
442 2017). Census individuals occurred along 12 replicate transects (200 to 600 m in length)
443 that spanned gradients of shrub density along shrub-grass ecotones. Size was measured
444 as volume of an elliptical cone based on height and width measurements; the size vari-
445 able of the IPM was the natural logarithm of volume (cm^3). For each census individual,
446 we recorded the size and density of all conspecifics within the five-meter transect “win-
447 dow” in which it occurred, and took the sum of all sizes within the window as a measure
448 of local density. The data are available in Ochocki et al. (2023).

449 As an initial Gaussian approach, we first fit a set of candidate generalized linear
450 mixed models, including transect as a random effect, that represented competing hy-
451 potheses for how size, density, and their interaction influence growth. Specifically, we fit
452 five candidate Gaussian models that included fixed effects of initial size only (model 1),
453 size and density (model 2), and size, density, and their interaction (model 3), allowing
454 for shrubs of different sizes to have different growth responses to local density. Models
455 4 and 5 mirrored models 2 and 3 but included second-order terms for density, allowing
456 for the possibility of non-monotonic density dependence.

```
457 LATR_GAU<-list()  
458 LATR_GAU[[1]] <- lmer(log_volume_t1 ~ log_volume_t + (1|unique.transect), data=LATR_g  
459 LATR_GAU[[2]] <- lmer(log_volume_t1 ~ log_volume_t + dens_scaled + (1|unique.transect),  
460 LATR_GAU[[3]] <- lmer(log_volume_t1 ~ log_volume_t*dens_scaled + (1|unique.transect),  
461 LATR_GAU[[4]] <- lmer(log_volume_t1 ~ log_volume_t + dens_scaled + I(dens_scaled^2) +  
462 LATR_GAU[[5]] <- lmer(log_volume_t1 ~ log_volume_t*dens_scaled + log_volume_t*I(dens_
```

463 As in (Drees et al., 2023) we pooled data across three transition years. Initial AIC rank-
464 ings of these pilot models favor model 4 slightly over model 5 ($\Delta AIC = 0.8$) and sig-
465 nificantly over all other models ($\Delta AIC > 2$). However, these models were fit assuming
466 constant variance, and inspection of the residuals of the best model indicate this is not a
467 safe assumption.

468 Unlike our previous case studies, here we have multiple fixed effects that may in-
469 fluence the variance of future size conditional on current size. In cases such as this,
470 we recommend modeling variance as a function of expected future size rather than ini-
471 tial size, as we did with the corals and cacti. The expected (or “fitted”) values reflect
472 the combined influence of all fixed and random effects, and are therefore a good op-
473 tion when there are multiple sources of variation in the variance, as we knew to be the
474 case for creosotebush (Drees et al., 2023). While there are several convenient software
475 packages for simultaneously modeling Gaussian mean and variance as functions of inde-

476 pendent variables (**mgcv** for additive models as we saw above, **nmle** for linear models),
477 modeling variance as a function of the mean is trickier because they cannot easily be fit
478 simultaneously.

479 4 Discussion

480 Here are some of the issues to be discussed.

- 481 • Modeling the mean with gam vs glm
- 482 • Modeling variance and higher moments as functions of covariates vs fitted values
- 483 • Choosing a better distribution – how to make the choice
- 484 • Comparison of our method with beta regression
- 485 • We have emphasize growth but same principles apply to other continuous state transitions, eg disease IPMs.

487 Acknowledgements

488 This research was supported by US NSF grants DEB-1933497 (SPE) and

489 5 Authorship statement

490 All authors discussed all aspects of the research and contributed to developing methods,
491 analyzing data, and writing and revising the paper.

492 6 Data accessibility statement

493 No original data appear in this paper. Should the paper be accepted, all computer scripts
494 supporting the results will be archived in a Zenodo package, with the DOI included at
495 the end of the article. During peer review, our data and code are available at [496 https://github.com/texmiller/IPM_size_transitions](https://github.com/texmiller/IPM_size_transitions).

497 **Literature Cited**

- 498 Anscombe, F. J. and Glynn, W. J. (1983). Distribution of the kurtosis statistic b_2 for
499 normal samples. *Biometrika*, 70(1):227–234.
- 500 Bates, D., Sarkar, D., Bates, M. D., and Matrix, L. (2007). The lme4 package. *R package
501 version*, 2(1):74.
- 502 Bruno, J. F., Ellner, S. P., Vu, I., Kim, K., and Harvell, C. D. (2011). Impacts of aspergillosis
503 on sea fan coral demography: modeling a moving target. *Ecological Monographs*,
504 81(1):123–139.
- 505 Compagnoni, A., Bibian, A. J., Ochocki, B. M., Rogers, H. S., Schultz, E. L., Sneck, M. E.,
506 Elderd, B. D., Iler, A. M., Inouye, D. W., Jacquemyn, H., et al. (2016). The effect of
507 demographic correlations on the stochastic population dynamics of perennial plants.
508 *Ecological Monographs*, 86(4):480–494.
- 509 Conn, P. B., Johnson, D. S., Williams, P. J., Melin, S. R., and Hooten, M. B. (2018). A guide
510 to bayesian model checking for ecologists. *Ecological Monographs*, 88(4):526–542.
- 511 Cooch, E. G. and White, G. C. (2020, accessed 5/17/2020). *Program MARK - a 'gentle
512 introduction'*. Available at phidot.org.
- 513 Coulson, T. (2012). Integral projections models, their construction and use in posing
514 hypotheses in ecology. *Oikos*, 121(9):1337–1350.
- 515 Crone, E. E. (2016). Contrasting effects of spatial heterogeneity and environmental
516 stochasticity on population dynamics of a perennial wildflower. *Journal of Ecology*,
517 104(2):281–291.
- 518 Czachura, K. and Miller, T. E. (2020). Demographic back-casting reveals that subtle
519 dimensions of climate change have strong effects on population viability. *Journal of
520 Ecology*.
- 521 D'Agostino, R. B. (1970). Transformation to normality of the null distribution of g_1 .
522 *Biometrika*, pages 679–681.
- 523 Davis, C. (2015). *sgt: Skewed Generalized T Distribution Tree*. R package version 2.0.
- 524 Drees, T., Ochocki, B. M., Collins, S. L., and Miller, T. E. (2023). Demography and
525 dispersal at a grass-shrub ecotone: a spatial integral projection model for woody plant
526 encroachment. *Ecological Monographs*, page e1574.

- 527 Easterling, M. R., Ellner, S. P., and Dixon, P. M. (2000). Size-specific sensitivity: applying
528 a new structured population model. *Ecology*, 81(3):694–708.
- 529 Elderd, B. D. and Miller, T. E. (2016). Quantifying demographic uncertainty: Bayesian
530 methods for integral projection models. *Ecological Monographs*, 86(1):125–144.
- 531 Ellis, M. M. and Crone, E. E. (2013). The role of transient dynamics in stochastic popula-
532 tion growth for nine perennial plants. *Ecology*, 94(8):1681–1686.
- 533 Ellner, S. P., Adler, P. B., Childs, D. Z., Hooker, G., Miller, T. E., and Rees, M. (2022).
534 A critical comparison of integral projection and matrix projection models for demo-
535 graphic analysis: Comment. *Ecology*, 103(10):e3605.
- 536 Ellner, S. P., Childs, D. Z., and Rees, M. (2016). *Data-driven Modeling of Structured Popula-
537 tions: A Practical Guide to the Integral Projection Model*. Springer, New York.
- 538 Gould, W. R. and Nichols, J. D. (1998). Estimation of temporal variability of survival in
539 animal populations. *Ecology*, 79:2531 – 2538.
- 540 Gu, C. (2013). *Smoothing Spline ANOVA Models*. Springer Science+Business Media, New
541 York, 2 edition.
- 542 Hadfield, J. D. et al. (2010). Mcmc methods for multi-response generalized linear mixed
543 models: the mcmcglmm r package. *Journal of Statistical Software*, 33(2):1–22.
- 544 Hérault, B., Bachelot, B., Poorter, L., Rossi, V., Bongers, F., Chave, J., Paine, C. T., Wagner,
545 F., and Baraloto, C. (2011). Functional traits shape ontogenetic growth trajectories of
546 rain forest tree species. *Journal of ecology*, 99(6):1431–1440.
- 547 Jones, M. and Pewsey, A. (2009). *Biometrika*, 96:761 – 780.
- 548 Jones, M. C., Rosco, J. F., and Pewsey, A. (2011). Skewness-invariant measures of kurtosis.
549 *The American Statistician*, 65(2):89 – 95.
- 550 Komsta, L. and Novomestky, F. (2015). Moments, cumulants, skewness, kurtosis and
551 related tests. *R package version*, 14(1).
- 552 Link, W. A. and Nichols, J. D. (1994). On the importance of sampling variance to inves-
553 tigations of temporal variation in animal population size. *Oikos*, 69(3):539 – 544.
- 554 Louthan, A. M., Keighron, M., Kiekebusch, E., Cayton, H., Terando, A., and Morris, W. F.
555 (2022). Climate change weakens the impact of disturbance interval on the growth rate
556 of natural populations of venus flytrap. *Ecological Monographs*, 92(4):e1528.

- 557 McGillivray, H. (1986). Skewness and asymmetry: measures and orderings. *Annals of*
558 *Statistics*, 14:994–1011.
- 559 Metcalf, C. J. E., Ellner, S. P., Childs, D. Z., Salguero-Gómez, R., Merow, C., McMahon,
560 S. M., Jongejans, E., and Rees, M. (2015). Statistical modelling of annual variation for
561 inference on stochastic population dynamics using Integral Projection Models. *Methods*
562 *in Ecology and Evolution*, 6:1007–1017.
- 563 Miller, T. E. (2020). Long-term study of tree cholla demography in the los pinos
564 mountains, sevilleta national wildlife refuge. [https://doi.org/10.6073/pasta/
565 dd06df3f950afe4a4642306182237d13](https://doi.org/10.6073/pasta/dd06df3f950afe4a4642306182237d13).
- 566 Miller, T. E., Louda, S. M., Rose, K. A., and Eckberg, J. O. (2009). Impacts of insect
567 herbivory on cactus population dynamics: experimental demography across an envi-
568 ronmental gradient. *Ecological Monographs*, 79(1):155–172.
- 569 Ochocki, B. M., Drees, T., and Miller, T. E. (2023). Density-dependent demography of
570 creosote bush (*Larrea tridentata*) along grass-shrub ecotones. [https://doi.org/10.
571 6073/pasta/ca53c16f16dcf9fb11f3ee99ea5445ac](https://doi.org/10.6073/pasta/ca53c16f16dcf9fb11f3ee99ea5445ac).
- 572 Ohm, J. R. and Miller, T. E. (2014). Balancing anti-herbivore benefits and anti-pollinator
573 costs of defensive mutualists. *Ecology*, 95(10):2924–2935.
- 574 Ozgul, A., Childs, D. Z., Oli, M. K., Armitage, K. B., Blumstein, D. T., Olson, L. E., Tul-
575 japurkar, S., and Coulson, T. (2010). Coupled dynamics of body mass and population
576 growth in response to environmental change. *Nature*, 466(7305):482–485.
- 577 Peterson, M. L., Morris, W., Linares, C., and Doak, D. (2019). Improving structured
578 population models with more realistic representations of non-normal growth. *Methods*
579 *in Ecology and Evolution*, 10(9):1431–1444.
- 580 Plard, F., Schindler, S., Arlettaz, R., and Schaub, M. (2018). Sex-specific heterogene-
581 ity in fixed morphological traits influences individual fitness in a monogamous bird
582 population. *The American Naturalist*, 191(1):106–119.
- 583 Rees, M., Childs, D. Z., and Ellner, S. P. (2014). Building integral projection models: a
584 user's guide. *Journal of Animal Ecology*, 83(3):528–545.
- 585 Salguero-Gómez, R. and Casper, B. B. (2010). Keeping plant shrinkage in the demo-
586 graphic loop. *Journal of Ecology*, 98(2):312–323.

- 587 Schielzeth, H., Dingemanse, N. J., Nakagawa, S., Westneat, D. F., Allegue, H., Teplitsky,
588 C., Réale, D., Dochtermann, N. A., Garamszegi, L. Z., and Araya-Ajoy, Y. G. (2020).
589 Robustness of linear mixed-effects models to violations of distributional assumptions.
590 *Methods in ecology and evolution*, 11(9):1141–1152.
- 591 Schultz, E. L., Eckberg, J. O., Berg, S. S., Louda, S. M., and Miller, T. E. (2017). Native
592 insect herbivory overwhelms context dependence to limit complex invasion dynamics
593 of exotic weeds. *Ecology letters*, 20(11):1374–1384.
- 594 Stasinopoulos, D. M., Rigby, R. A., et al. (2007). Generalized additive models for location
595 scale and shape (gamlss) in r. *Journal of Statistical Software*, 23(7):1–46.
- 596 Stubberud, M. W., Vindenes, Y., Vøllestad, L. A., Winfield, I. J., Stenseth, N. C., and Lan-
597 gangen, Ø. (2019). Effects of size-and sex-selective harvesting: An integral projection
598 model approach. *Ecology and Evolution*, 9(22):12556–12570.
- 599 Wan, X., Wang, W., Liu, J., and Tong, T. (2014). Estimating the sample mean and stan-
600 dard deviation from the sample size, median, range and/or interquartile range. *BMC*
601 *medical research methodology*, 14:1–13.
- 602 Williams, J. L., Miller, T. E., and Ellner, S. P. (2012). Avoiding unintentional eviction from
603 integral projection models. *Ecology*, 93(9):2008–2014.
- 604 Wood, S. (2017). *Generalized Additive Models: An Introduction with R*. Chapman and
605 Hall/CRC, 2 edition.
- 606 Zhang, J. L. (2014). Comparative investigation of three bayesian p values. *Computational*
607 *Statistics & Data Analysis*, 79:277–291.

Appendices

S.1 The Jones-Pewsey distribution

Jones and Pewsey (2009) introduced a simple, tractable generalization of the Normal distribution with two additional parameters determining asymmetry (skewness), and tail weight (kurtosis) which can be either lighter or heavier than the Gaussian. It is defined as a transformation of a $\text{Normal}(0,1)$ random variable using the hyperbolic sine function (\sinh) and its inverse (asinh), as follows. The distribution family's base probability density $f_{\epsilon,\delta}$ is the probability density of the random variable $X_{\epsilon,\delta}$ where

$$Z = \sinh(\delta \text{ asinh}(X_{\epsilon,\delta}) - \epsilon) \quad (\text{S.1})$$

and Z has a $\text{Normal}(0,1)$ distribution. Equivalently,

$$X_{\epsilon,\delta} = \sinh\left(\frac{1}{\delta} \text{ asinh}(Z) + \frac{\epsilon}{\delta}\right). \quad (\text{S.2})$$

Parameters $\delta = 1, \epsilon = 0$ give the $\text{Normal}(0,1)$ distribution. Skewness has the sign of ϵ , and $\delta > 0$ controls tail weight, with heavier than Gaussian tails for $\delta < 1$ and lighter than Gaussian tails for $\delta > 1$. A formula for the density $f_{\epsilon,\delta}$ is given by Jones and Pewsey (2009, eqn. 2). The general four-parameter family with location parameter μ and scale parameter σ is defined as the probability densities of $\mu + \sigma X_{\epsilon,\delta}$. We refer to this as the JP distribution family.

As is unfortunately the case for most four-parameter distributions μ is not the mean, σ is not the standard deviation, ϵ is not the skew and δ is not the kurtosis. All else being equal, larger μ gives a larger mean, larger σ gives a higher standard deviation, higher ϵ gives higher asymmetry, and higher δ gives heavier tail weight. But each moment is jointly determined by all four parameters.

The main advantage of the JP distribution is that the attainable combinations of skewness and kurtosis are very broad, compared to other four-parameter families, and come very close to the theoretical limits on kurtosis as a function of skewness (Jones and Pewsey, 2009, Fig. 2). Additionally, being a transformation of the Normal makes it very simple to generate random numbers from the distribution, and to compute probability density, cumulative distribution, and quantile functions. There are also simple analytic formulas for the first four moments (Jones and Pewsey, 2009, p. 764) which we use below

636 to define a centered and scaled version in which μ and σ are the mean and standard
637 deviation.

638 The definition (S.2) shows that the distribution depends on ϵ only through the ratio
639 ϵ/δ . We have found that this property can be problematic for estimating distribution
640 parameters. Even with good sized ($n = 250$ or 500) data sets generated from the distri-
641 bution with known parameters, both maximum likelihood and Bayesian estimation were
642 unstable for some values of ϵ and δ , occasionally yielding estimates far from the truth.
643 One cause was a ridge in the (ϵ, δ) likelihood surface with a constant of ϵ/δ . Another is
644 that when δ is large, changes in ϵ have little effect.

645 To avoid that problems, we reparameterize the distribution as follows:

646
$$X_{\lambda, \tau} = \sinh(e^{-\tau} \operatorname{asinh}(Z) + \lambda). \quad (\text{S.3})$$

647 Thus, the two parameterizations are related by

648
$$\delta = e^\tau, \epsilon = \delta\lambda = e^\tau\lambda. \quad (\text{S.4})$$

649 The definition of τ allows it to take any real value, with negative values giving thinner
650 than Gaussian tails and positive values giving fatter than Gaussian tails. λ also can take
651 any real value, and the distribution's skew has the same sign as λ . Because the \sinh
652 function is nonlinear, it is still the case that the skew depends on τ as well as λ , but the
653 "crosstalk" between the kurtosis and skew parameters is weaker. As a result, we found
654 that maximum likelihood estimation of parameter values was generally more reliable if
655 the distribution is parameterized in terms of τ and λ .

656 S.2 Estimating mixed-effects models using shrinkage

657 Ecologists often fit demographic and other statistical models that include random effects
658 terms to quantify variation among years, spatial locations, individuals, etc. Random
659 effects are a natural choice when interest centers on the magnitude of variation (e.g., how
660 much does mortality vary among years?) rather than individual values (e.g., mortality
661 in 2013). They also allow each estimate to "borrows strength" from others, so that (for
662 example) the estimate from a year with small sample size (and thus large sampling
663 variability) is shifted towards the center of the overall distribution.

664 Specialized software is often used to fit such models, such as the **nlme**, **lme4**, **mgee**
665 and **gamm4** libraries in R, but these only allow a small subset of the distribution families

666 we want to consider for modeling growth increments (the **gamlss** package allows many
667 distribution families, but in our experience, even when random effects are simple in
668 structure the fitting algorithms often fail to converge or fail to find the global optimum).

669 One way past this limitation is Bayesian estimation, using STAN with user-written
670 (or borrowed) code for the chosen growth distribution (see section XX for an example).
671 In this appendix we describe another option, introduced by Link and Nichols (1994)
672 and Gould and Nichols (1998): fitting a fixed-effects model by Maximum Likelihood,
673 followed by shrinkage of coefficient estimates. None of the ideas here are original. The
674 material overlaps Appendix S1 of Metcalf et al. (2015), but for completeness we make
675 it self-contained. Appendix D of Cooch and White (2020) (written by K.D. Burnham)
676 provides more details and examples in the context of capture-recapture analysis.

677 Here we explain shrinkage using a simple model based on our analysis of *Pseu-*
678 *doroegneria spicata*. That model includes random effects for between-year variation in
679 the slope and intercept of future size (log area) as a function of initial size. To keep
680 the example simple, we assume that initial size and year are the only covariates, and
681 we assume that growth increments follow a skew-Normal distribution with noncon-
682 stant variance and constant skew parameter. Code for this example is in the script
683 `SimpleShrinkageExample.R`. The first part of the script generates an artificial data set
684 by fitting the model to a subset of the growth data (20th century Control plots), and
685 randomly generating new “size next year” values for each individual in the actual data
686 set. The second part contains the “data” analysis.

687 As in our *P. spicata* analysis, we assumed that that the skew and kurtosis parameters
688 were functions of the location parameter; this dominated ($\Delta AIC \approx 30$) the alternate
689 model with skew and kurtosis depending on initial size. The analogous Gaussian model,
690 with constant variance, could be fitted as follows using `lmer`:

691 `lmer(new.size ~ init.size + (init.size|year), data=growthData, REML=TRUE);`
692 where `growthData` is a data frame holding the data with `year` as an unordered factor.
693 For our skew-Normal model, we instead use maximum likelihood with all between-year
694 variation included as fixed effects. The appropriate design matrix is easily constructed
695 using the `model.matrix` function:

696 `U = model.matrix(~ year + init.size:year - 1, data=growthData)`

697 If there are T years, the matrix `U` specified in this way has $2T$ columns corresponding to
698 n annual intercepts and T annual slopes.

699 Using this design matrix, we can readily write a log likelihood function for use with
 700 the **maxLik** package, with a log link function for the variance because it is necessarily
 701 positive:

```
702 LogLik=function(pars,new.size,U){  

  703   pars1 = pars[1:ncol(U)]; pars2=pars[-(1:ncol(U))];  

  704   mu = U%*%pars1;  

  705   sigma = exp(pars2[1]+pars2[2]*mu);  

  706   dSN1(new.size, mu=mu, sigma=sigma, nu=pars2[3], log=TRUE)  

  707 }
```

708 Parameters and their standard errors can then be estimated with **maxLik**, starting
 709 from a random guess:

```
710 start=c(runif(ncol(U)), rep(0,3))  

  711 out=maxLik(logLik=LogLik,start=start, new.size=simData$new.size,U=U,  

  712   method="BHHH",control=list(iterlim=5000,printLevel=1),finalHessian=TRUE);  

  713 coefs = out$estimate; # parameters  

  714 V = vcov(out); SEs = sqrt(diag(V)); # standard errors
```

715 In real life we would repeat the optimization several times with several different starting
 716 values, to be confident that the optimal parameter values had been found.

717 Focus now on the year-specific intercept parameters $\hat{a}_t, t = 1, 2, \dots, T$. We can view
 718 the year-specific estimates \hat{a}_t as consisting of unobserved true values a_t plus sampling
 719 error:

$$720 \hat{a}_t = a_t + \varepsilon_t \quad (S.5)$$

721 Because of the sampling errors, the sample variance of the estimates \hat{a}_t is an upward-
 722 biased estimate of the true across-year variance in the parameter. That is undesirable if
 723 the model will be used to project how temporal variability affects population dynamics.
 724 However, maximum likelihood estimation gives us an approximate variance-covariance
 725 matrix \hat{V} of the sampling errors, V in the code above. With that information, we can
 726 estimate the parameters of a random effects model for the intercept parameters, and
 727 thereby improve the year-specific estimates and the estimate of the across-year variance.

728 The model is as follows. We make the standard mixed-models assumptions that the
 729 a_t are drawn independently from some fixed distribution with unknown variance σ^2 .
 730 We also assume that the estimates \hat{a}_t are unbiased, that is

$$731 \mathbb{E}(\varepsilon_t | a_t) = 0. \quad (S.6)$$

732 These are optimistic assumptions, but not excessively optimistic. Some degree of tem-
 733 poral correlation will often be present, and as we explain at the end, it is theoretically
 734 possible to account for it. Maximum likelihood parameter estimates are not unbiased,
 735 but if the assumptions of maximum likelihood are satisfied the bias is asymptotically
 736 negligible compared to the standard error (the bias scales as the inverse of sample size,
 737 the standard error as the square root of the inverse of sample size).

738 Let S^2 denote the sample variance of the estimates \hat{a}_t . It can then be shown that

$$739 \mathbb{E}(S^2) = \sigma^2 + \frac{1}{T} \sum_{t=1}^T \mathbb{E}Var(\varepsilon_t) - \frac{1}{T(T-1)} \sum_{i=1}^{j-1} \sum_{j=1}^T \mathbb{E}Cov(\varepsilon_i, \varepsilon_j). \quad (\text{S.7})$$

740 This is eqn. (1) in Gould and Nichols (1998) in our notation, without the term that results
 741 from temporal autocorrelation.

742 The terms besides σ^2 on the right-hand are the expected impact of sampling error
 743 on the across-year variance of the parameter estimates; their presence makes S^2 a biased
 744 estimate of σ^2 . However, all of those terms correspond to entries in the variance-
 745 covariance matrix V . We can therefore use our estimated variance-covariance matrix \hat{V}
 746 to remove the bias due to sampling variability:

$$747 \hat{\sigma}^2 = S^2 - \frac{1}{T} \sum_{t=1}^T \hat{V}_{t,t} + \frac{1}{T(T-1)} \sum_{i=1}^{j-1} \sum_{j=1}^T \hat{V}_{i,j}. \quad (\text{S.8})$$

748 $\hat{\sigma}^2$ estimates the variance of the distribution from which the a_t are assumed to be drawn.

749 Using that estimate, we can adjust the year-specific estimates to reduce the ex-
 750 pected impact of sampling error. Depending on your purposes, there are two possible
 751 adjustments. The first option is the one used in the popular capture-recapture analysis
 752 software Mark Cooch and White (2020),

$$753 \tilde{a}_t = \bar{a}_t + \sqrt{\frac{\hat{\sigma}^2}{\hat{\sigma}^2 + \hat{V}_{t,t}}} (\hat{a}_t - \bar{a}_t). \quad (\text{S.9})$$

754 The name “shrinkage” comes from the fact that each estimate is adjusted towards the
 755 overall mean, with larger adjustments of values that have higher estimated sampling
 756 error variance, $\hat{V}_{t,t}$. This shrinkage estimate has the property that the expected sample
 757 variance of the adjusted estimates \tilde{a}_t is very close to $\hat{\sigma}^2$, so the \tilde{a}_t approximate the actual
 758 amount of parameter variation.

759 The second is to replace \hat{a}_t by the least-squares estimate of a_t under the additional
 760 assumption that the a_t are drawn from a Gaussian distribution; this is given by

$$761 \quad \tilde{a}_t = \bar{a}_t + \frac{\hat{\sigma}^2}{\hat{\sigma}^2 + \hat{V}_{t,t}} (\hat{a}_t - \bar{a}_t). \quad (S.10)$$

762 This option is theoretically preferable if the Gaussian assumption is reasonable, and you
 763 are more interested in year-specific values rather than across-year variance. However,
 764 Metcalf et al. (2015) found that even (S.9), which does less shrinkage, resulted in a small
 765 downward bias in the temporal variance of population growth rates. This argues for
 766 always using the first option, and we do the same here.

767 We differ from MARK, however, in using (S.8) rather than an iterative method
 768 that takes (S.8) as its starting estimate and refines the estimate by using weighted least
 769 squares based on the current estimate. Metcalf et al. (2015) found, in simulation studies,
 770 that the iterative method was either slightly beneficial or wildly inaccurate. We therefore
 771 advise against it.

772 Finally, as mentioned above, the estimate of σ^2 can account for temporal autocor-
 773 relation in the a_t . When present, those correlations add a term to eqn. (S.7) (see eqn.
 774 (1) in Gould and Nichols (1998)), which can be estimated from the sample autocorre-
 775 lation of the \hat{a}_t . We do not recommend doing this (and therefore omit the formulas)
 776 because the autocorrelations can only be reliably estimated if they fall to nearly zero
 777 within lag $m \ll T$, in which case the autocorrelation term is small (specifically, $O(m/T)$).
 778 Otherwise, the random error from using poorly estimated autocorrelations is likely to
 779 outweigh the small bias from omitting that term.

780 The take-home message is that estimating random effects from the regression coef-
 781 ficients is very simple:

```
782 # Variance-covariance matrices for intercepts and slopes
783 V1 = V[1:T,1:T]; V2 = V[(T+1):(2*T),(T+1):(2*T)];
784 # Extract year-specific intercepts, center them to zero
785 fixed.fx = coefs[1:T]; fixed.fx = fixed.fx-mean(fixed.fx);
786
787 # Estimate sigma^2
788 var.hat = mean(fixed.fx^2) - mean(diag(V1)) +
789             (sum(V1)-sum(diag(V1)))/(2*T*(T-1));
790
791 # Shrink deviations from the mean
```

```

792 shrinkRanIntercept = fixed.fx*sqrt(var.hat/(var.hat + diag(V1)));
793
794 # Do it all again for the slopes
795 fixed.fx2 = coefs[(T+1):(2*T)]; fixed.fx2 = fixed.fx2-mean(fixed.fx2);
796 var2.hat = mean(fixed.fx2^2) - mean(diag(V2)) +
797   (sum(V2)-sum(diag(V2)))/(2*T*(T-1));
798 shrinkRanSlope = fixed.fx2*sqrt(var2.hat/(var2.hat + diag(V2)));

```

799 The figure below shows the results for one artificial PSSP “data” set, having $T = 22$
 800 years and growth measurements on about 175 individuals/year on average. The true
 801 random year effects (the ones used to generate the data) are recovered with good accu-
 802 racy and no bias. In particular there is no sign of extreme values being pulled in too
 803 far towards the mean, which would cause an S-shaped graph of estimated versus true
 804 values.

