

# Two-sex demography, sexual niche differentiation, and range limits

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## Abstract

## 1 Keywords

## 2 Introduction

3 P1: importance of understanding range limits in basic and applied ecology, theory  
4 for proximate causes of range limits, demographic failure at range edges and the  
5 idea that range limits are niche limits

6 P2: The idea that range limits are niche limits intersects awkwardly with  
7 another prevalent concept in ecology: intraspecific niche heterogeneity. If a species'  
8 range limits reflect its niche limits, and a single species contains many niches, then  
9 whose niche is it that determines the geographic distribution.

10 P3: Sexual niche differentiation in dioecious species is a widespread form of  
11 intra-specific niche heterogeneity. Sex-specific responses to environmental drivers  
12 can generate geographic clines in operational sex ratio and strongly biased sex  
13 ratios especially at range limits. While this pattern is well documented in a variety  
14 of taxa, the role of sex ratio bias in limiting species' ranges is poorly understood.

15 P4: Most ecological theory assumes female dominance. In this case, there is a  
16 straightforward answer to the question above – the female niche is the relevant set  
17 of constraints for understanding range limits. However, while female dominance is  
18 often a reasonable and useful assumption, it may break down under extreme sex  
19 ratio bias, where mates may be limiting. If so, this creates an additional, two-sex  
20 pathway by which males could limit the position of species ranges.

21 P5: In this study we used a dioecious grass species as a focal species to quan-  
22 tify the relative importance of female-dominant and two-sex mechanisms of range  
23 limitation.

## 24 Materials and methods

### 25 Study system and natural population surveys

26 *Poa arachnifera* is a perennial, cool-season grass native to the southern Great  
27 Plains (Fig). Individuals can be sexed only when flowering, in early spring, based  
28 on the presence of stigmas (females) or anthers (males) in the inflorescence. Fol-  
29 lowing inflorescence and seed production, plants go dormant for the hot summer  
30 months and vegetative growth resumes in fall. Individuals grow via rhizomes to  
31 form patches that may be as large as  $50m^2$  in area. Sex is genetically based in  
32 *P. arachnifera* (Renganayaki *et al.*, 2001, 2005) and the primary sex ratio is 1:1  
33 (J. Goldman, USDA-ARS, *personal communication*). The rhizomatous growth  
34 habit allowed us to clonally propagate large numbers of known-sex individuals for  
35 experiments, as we describe below.

36 We surveyed *P. arachnifera* across its range to establish whether natural pop-  
37 ulations exhibited geographic clines in sex ratio corresponding to the longitudinal  
38 aridity gradient. We visited 14 populations in spring 2012 and 8 in spring 2013.  
39 Survey locations are shown in **map** and coordinates are provided in Table A1. At  
40 each location, we searched for *P. arachnifera* along roads, trails, or creek drainages  
41 and recorded the number of female and male inflorescences that we encountered.  
42 We fit a binomial generalized linear model (glm), where females were “successes”  
43 and total inflorescences was the number of “trials”, to test whether the operational  
44 sex ratio (OSR) varied systematically with respect to longitude. Here and in the  
45 experiment that follows we use longitude as a proxy variable that captures all  
46 east-west environmental variation, notably precipitation (map figure) but also fac-  
47 tors that co-vary with precipitation, such as productivity. This statistical model

48 and all those that follow were fit in a Bayesian statistical framework using Stan  
49 (Carpenter *et al.*, 2017) and rstan (Team *et al.*, 2018) with vague priors on all  
50 parameters.

## 51 Common garden experiment

### 52 Source material and experimental sites

53 We established a common garden experiment at 14 sites throughout and beyond  
54 the geographic distribution of *P. arachnifera* (MAP). Experimental sites spanned  
55 latitudinal and longitudinal variation, though we focus here on longitude. During  
56 the three years of this experiment, total precipitation at each site closely tracked  
57 longitude (Fig. A1), as expected based on longer-term climate trends (map).  
58 Source material for this experiment came from 8 sites, which were of subset of the  
59 sites that were visited for the natural population survey (Table). At each of these  
60 site visits in 2013 and 2014, we collected tillers from flowering individuals of each  
61 sex (mean: 11.6 individuals per site, range: 2–18). These were brought back to  
62 the Rice University greenhouse, where they were clonally propagated in cylidrical  
63 cone-tainers with ProMix potting soil and supplemental Osmocote fertilizer at  
64 78–80°F under natural light.

65 Common gardens were set up in Fall (October–December) 2014. At each site,  
66 we established **N** experimental blocks, which typically corresponded to a tree or  
67 woodland edge, providing partial shade that mimics this species’ natural micro-  
68 environment. We planted **N** females and **N** males in each block, for a total of **N**  
69 individuals per sex per site and **N** total plants across sites. To promote establish-  
70 ment, we cleared vegetation immediately surrounding transplants and provided ca.

71 1 L of water at the time of transplanting but provided no subsequent watering,  
72 fertilization, or competitor removal.

### 73 **Data collection**

74 We visited each site during May of 2015, 2016, and 2017. For each individual  
75 in each year, we recorded survival status (alive or dead), size (number of tillers),  
76 flowering status (reproductive or vegetative), the number of panicles produced by  
77 flowering plants, and panicle length, which is proportional to production of seeds  
78 (in females) and pollen (in males).

### 79 **Pollination experiment**

### 80 **Mechanistic model of range limits**

### 81 **Statistical modeling**

### 82 **Demographic modeling**

### 83 **Results**

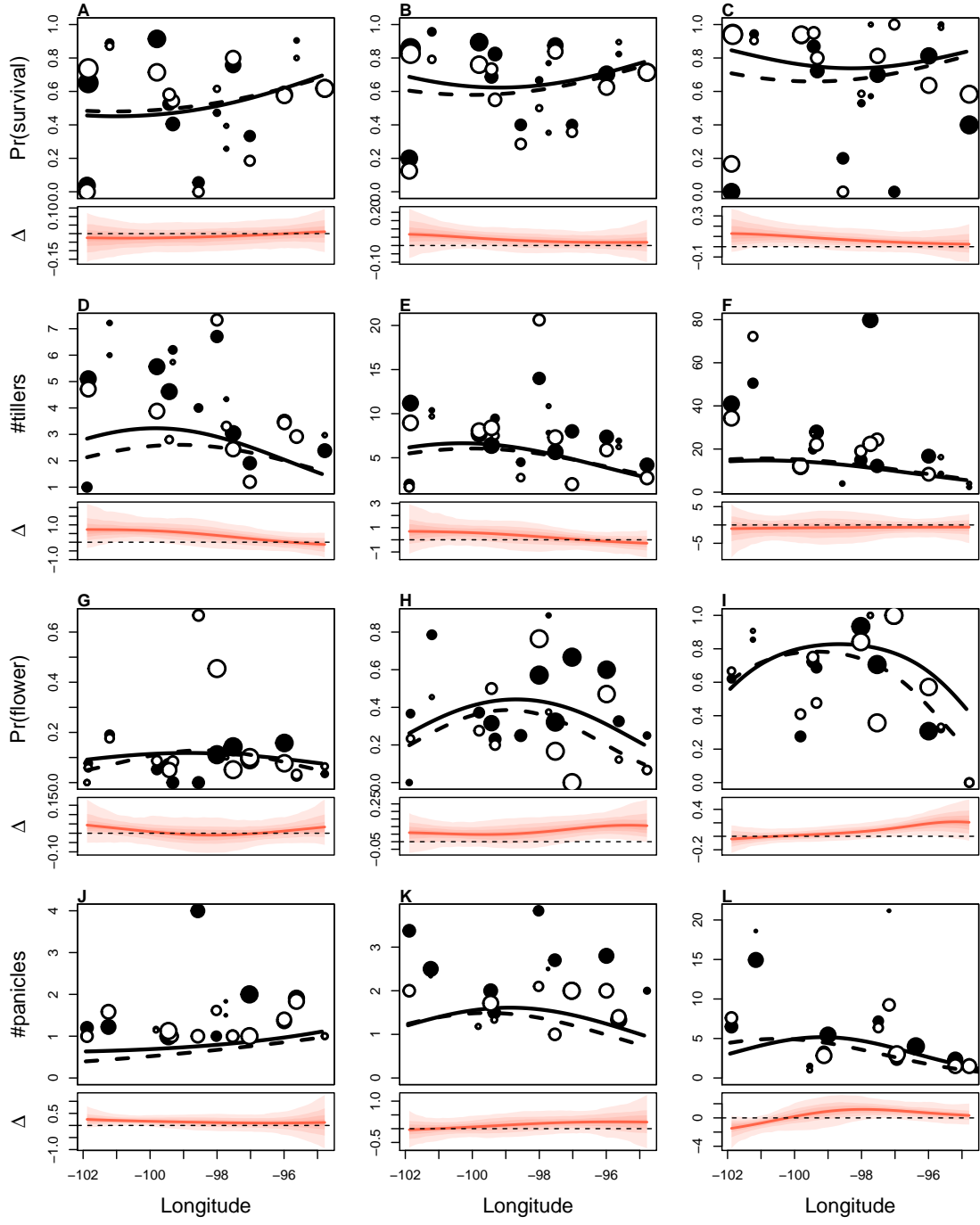


Figure 1: Caption.

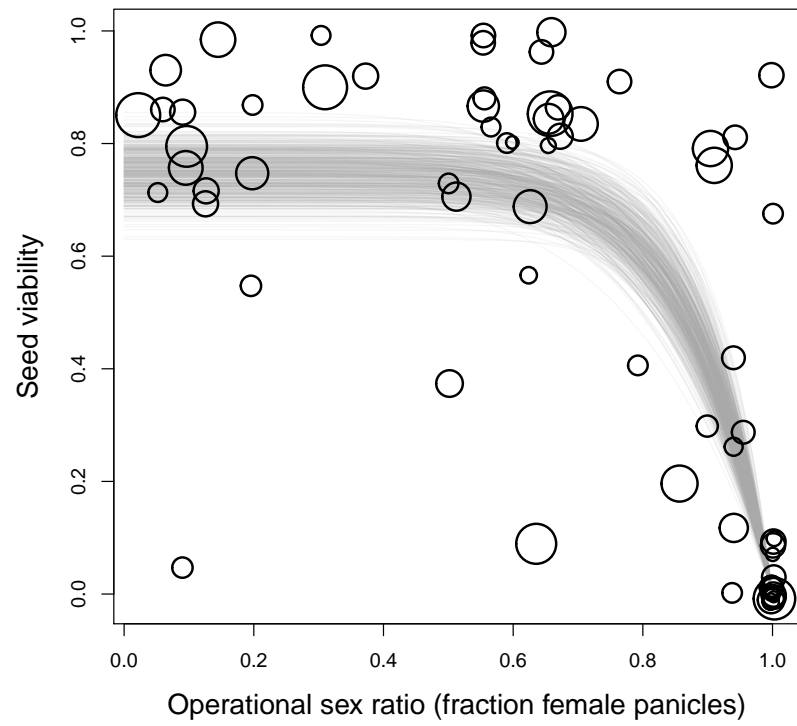


Figure 2: Caption.



## 84 Discussion

## 85 Acknowledgements

## 86 Author contributions

## 87 Data accessibility

## 88 References

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90 gramming language. *Journal of statistical software*, **76**.
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- 94 Renganayaki K, Read J, Fritz A (2001) Genetic diversity among texas bluegrass  
95 genotypes (*poa arachnifera* torr.) revealed by aflu and rapd markers. *Theoretical*  
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- 97 Team SD, *et al.* (2018) Rstan: the r interface to stan. r package version 2.17. 3.

## Appendix A: Site locations and climate

	Population	Latitude	Longitude
1	Canyon_of_Eagles	30.88	-98.43
2	ClearBay-Thunderbird	35.23	-97.24
3	CooperWMA	36.60	-99.51
4	Copper Breaks	34.10	-99.75
5	Dinosaur_Valley	32.25	-97.82
6	Fort_Worth_Nature_Center	32.83	-97.46
7	Ft Cobb	35.18	-98.45
8	Ft Richardson	33.20	-98.16
9	Great Plains	34.74	-98.97
10	Great_Salt_Plains	36.79	-98.18
11	Horn_Hill_Cemetery	31.56	-96.64
12	Kingman_Fishing_Lake	37.65	-98.28
13	Lake Arrowhead	33.75	-98.39
14	Mineral_Wells	32.89	-98.01
15	Pedernales_Falls	30.33	-98.25
16	Possum Kingdom	32.87	-98.57
17	Quartz_Mountain	34.89	-99.30
18	Red Rock Canyon	35.44	-98.35
19	Red_River	34.13	-98.10
20	South_Llano	30.45	-99.80
21	Sulfur_Springs	31.08	-98.46
22	Wichita_Mountains	34.70	-98.67

Table A1: Sites of natural population surveys corresponding to Figure

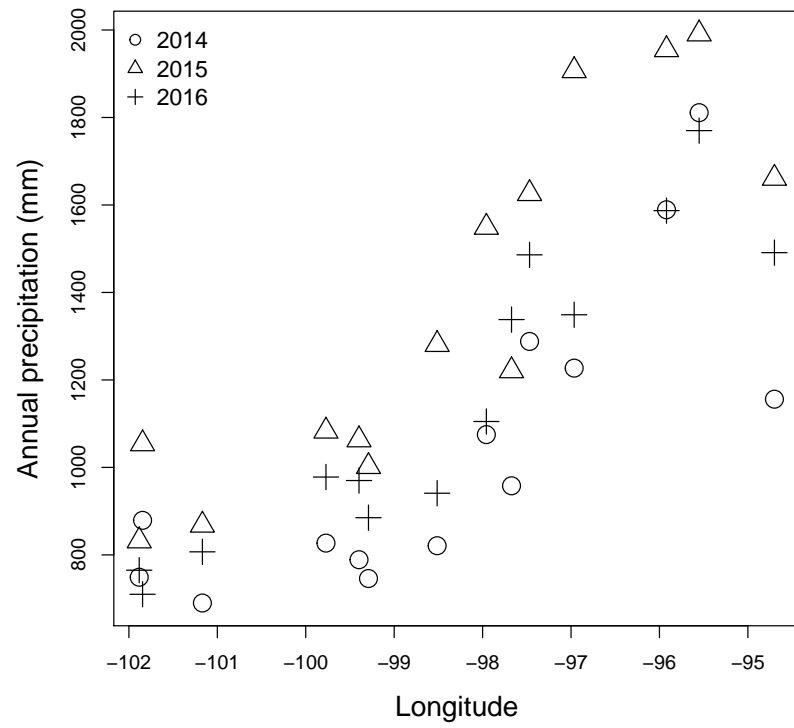


Figure A1: Caption.