

Two-sex demography, sexual niche differentiation, and range limits

Tom E.X. Miller* and Aldo Compagnoni

Program in Ecology and Evolutionary Biology, Department of
BioSciences, Rice University, Houston, TX USA

*Corresponding author: tom.miller@rice.edu (1-713-348-4218)

Abstract

1 Keywords

2 Introduction

3 Understanding the processes that generate species' distributional limits is a foun-
4 dational objective of ecology. The niche concept is central to theory for range limits
5 (Hutchinson, 1958), and available evidence suggests that geographic distributions
6 may commonly be interpreted as ecological niches “writ large” (Lee-Yaw *et al.*,
7 2016; Hargreaves *et al.*, 2013). Species distribution modeling (or “environmen-
8 tal niche modeling”) has long capitalized on this idea to infer niche characteristics
9 from statistical associations between occurrence across environmental variables. In
10 contrast, there is growing interest in process-based models of range limits, where
11 individual-level demographic responses to environmental variation inform predic-
12 tions about the ecological niche and environmental limits of population viability
13 (i.e., at least replacement-level population growth, $\lambda \geq 1$) (Merow *et al.*, 2014,
14 2017; Diez *et al.*, 2014). The mechanistic understanding offered by process-based
15 models of range limits provides a potentially powerful vehicle for predicting range
16 shifts in response to current and future environmental change (Evans *et al.*, 2016;
17 Ehrlén & Morris, 2015).

18 The widespread idea that range limits reflect niche limits intersects awkwardly
19 with another pervasive concept in ecology: intra-specific niche heterogeneity. This
20 refers to the fact that individuals within a population or species may differ in
21 their interactions with and responses to the biotic and / or abiotic environment
22 (Bolnick *et al.*, 2002; Araújo *et al.*, 2011; Holt, 2009). Intra-specific niche differ-
23 ences may be based on demographic state variables such as life stage, size class or
24 other, unmeasured aspects of individual identity. If range limits are a geographic
25 manifestation of niche limits, but a single population or species may be comprised

26 of many niches, then whose niche is it that determines the geographic distribution
27 and how would we know?

28 Sexual niche differentiation is a common form of intra-specific niche heterogene-
29 ity (Bolnick *et al.*, 2002) and has been widely documented in animals (the vast
30 majority of which are dioecious) and plants (ca. 6% of angiosperms are dioecious:
31 Renner & Ricklefs 1995). The prevalence of sexual niche differentiation was rec-
32 ognized by Darwin (2019), who described “different habits of life, not related...to
33 the reproductive functions” of females and males. There are numerous examples of
34 sex differences in trophic position (Pekár *et al.*, 2011), habitat use (Bowyer, 2004;
35 Phillips *et al.*, 2004), and responses to climate (Petry *et al.*, 2016; Rozas *et al.*,
36 2009; Gianuca *et al.*, 2019), differences that may or may not be accompanied by
37 sexual dimorphism. It has been hypothesized that sex-specific niches may evolve
38 by natural selection when it reduces competitive or other antagonistic interactions
39 between the sexes (Bierzychudek & Eckhart, 1988; Bolnick & Doebeli, 2003) or as a
40 byproduct of naturally or sexually selected size dimorphism (Shine, 1989; Temeles
41 *et al.*, 2010). In an ecological context, sexual niche differentiation can translate to
42 sex-specific advantages in different environments, causing skew in the operational
43 sex ratio (OSR: relative abundance of females and males available for mating) even
44 if the primary sex ratio is unbiased (Veran & Beissinger, 2009; Shelton, 2010). En-
45 vironmental clines in OSR have been widely documented in plants and animals at
46 fine spatial scales (Eppley, 2001; Bertiller *et al.*, 2002; Groen *et al.*, 2010) as well as
47 broader climatic clines across altitudes or latitudes (Petry *et al.*, 2016; Ketterson
48 & Nolan Jr, 1976; Caruso & Case, 2007). At range margins, where environments
49 are extreme relative to the range core, demographic differences between the sexes,
50 and hence skew in the OSR, may be greatest. In dioecious plants, for example,

51 populations at the upper altitudes and latitudes and in the more xeric margins of
52 species' ranges tend to be male-biased (Field *et al.*, 2013).

53 Returning to the question of whose niche determines range limits given poten-
54 tial for sexual niche differentiation, classic ecological theory provides a straight-
55 forward answer. "Female dominance" is a pervasive feature of population-dynamic
56 models whereby male availability is assumed to have no influence on female fertility.
57 This assumption is, of course, wrong but it may often be a useful approximation
58 when the sex ratio is balanced or does not vary. The female-dominant perspec-
59 tive predicts that female responses to environmental variation should govern range
60 limits. However, females may be male-limited in environments in which they are
61 favored, which could reduce population viability in marginal environmental. This
62 creates an additional, "two-sex" pathway by which environmental drivers may set
63 distributional limits, via perturbations to the mating pool that arise from sex-
64 specific responses. Here we ask whether female demographic responses to environ-
65 mental variation, alone, are sufficient to understand the ecological origins of range
66 limits, or whether the additional role (and especially lack) of males contributes to
67 range boundary formation.

68 P3: Sexual niche differentiation in dioecious species is a widespread form of
69 intra-specific niche heterogeneity. Sex-specific responses to environmental drivers
70 can generate geographic clines in operational sex ratio and strongly biased sex
71 ratios especially at range limits. While this pattern is well documented in a variety
72 of taxa, the role of sex ratio bias in limiting species' ranges is poorly understood.

73 P4: Most ecological theory assumes female dominance. In this case, there is a
74 straightforward answer to the question above – the female niche is the relevant set
75 of constraints for understanding range limits. However, while female dominance is

76 often a reasonable and useful assumption, it may break down under extreme sex
77 ratio bias, where mates may be limiting. If so, this creates an additional, two-sex
78 pathway by which males could limit the position of species ranges.

79 P5: In this study we used a dioecious grass species as a focal species to quan-
80 tify the relative importance of female-dominant and two-sex mechanisms of range
81 limitation.

82 Materials and methods

83 Study system and natural population surveys

84 *Poa arachnifera* is a perennial, cool-season grass endemic to the southern Great
85 Plains. This species occurs almost exclusively in central Texas, Oklahoma, and
86 Kansas (Fig. 1) though there are occasional records of adventive populations in
87 other states¹. Like all grasses, *P. arachnifera* is wind-pollinated. Individuals can
88 be sexed only when flowering, in early spring, based on the presence of stigmas
89 (females) or anthers (males) in the inflorescence. Following inflorescence and seed
90 production, plants go dormant for the hot summer months and vegetative growth
91 resumes in fall. Individuals grow via rhizomes to form patches that may be as large
92 as 50m² in area. Sex in *P. arachnifera* is genetically based (Renganayaki *et al.*,
93 2001, 2005) and the primary sex ratio is 1:1 (J. Goldman, USDA-ARS, *personal*
94 *communication*). The rhizomatous growth habit allowed us to clonally propagate
95 large numbers of known-sex individuals for experiments, as we describe below.

96 We surveyed *P. arachnifera* across its range to establish whether natural pop-

¹<http://bonap.net/Napa/TaxonMaps/Genus/County/Poa>

97 ulations exhibited geographic clines in sex ratio corresponding to the longitudinal
98 aridity gradient. We visited 14 populations in spring 2012 and 8 in spring 2013
99 (Table A1). At each location, we searched for *P. arachnifera* along roads, trails,
100 or creek drainages and recorded the number of female and male inflorescences that
101 we encountered.

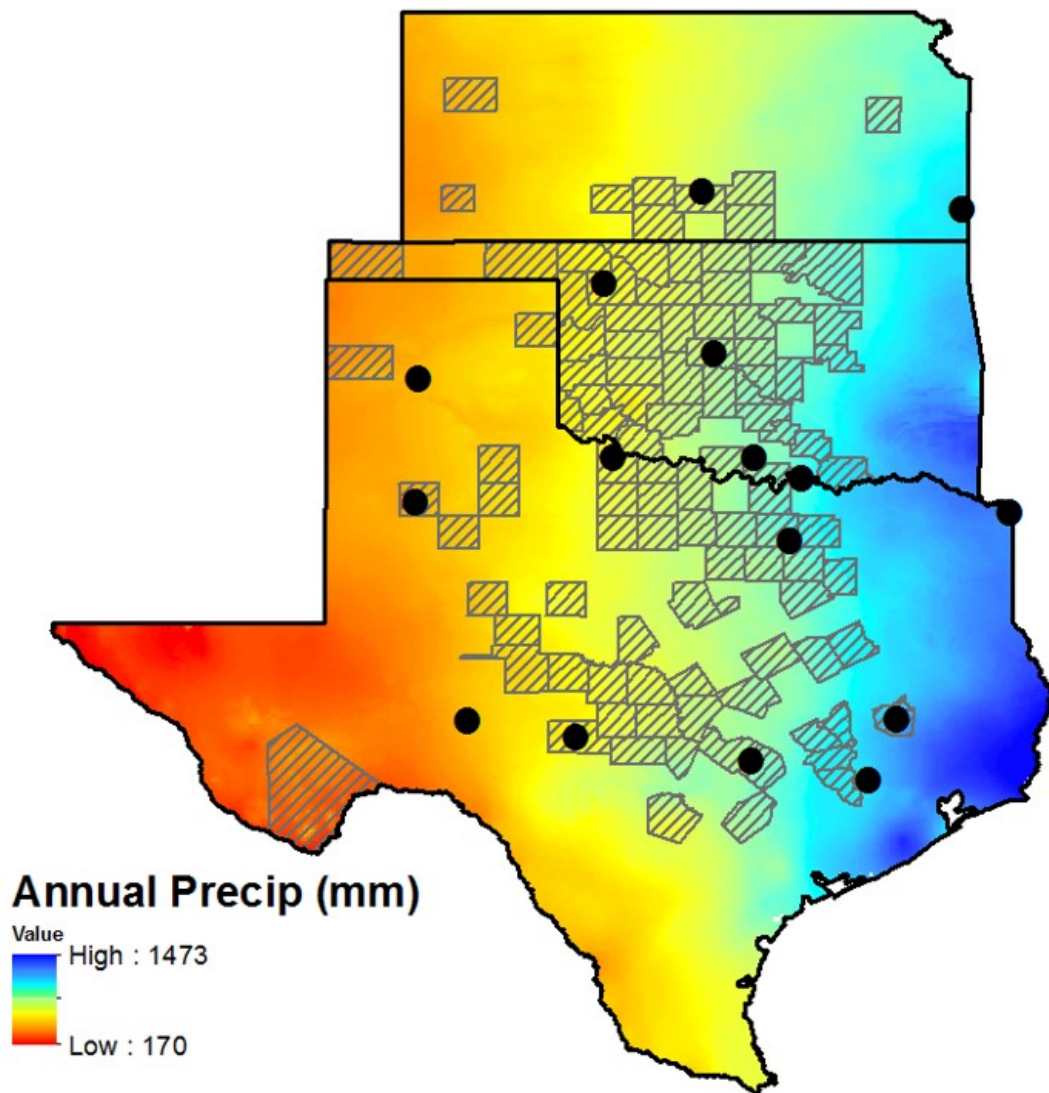


Figure 1: Caption.

102 **Statistical analysis of natural population surveys**

103 We fit a binomial generalized linear model (glm), where females were “successes”
 104 and total inflorescences was the number of “trials”, to test whether the operational

sex ratio (OSR) varied systematically with respect to longitude. Here and in the experiment that follows we use longitude as a proxy variable that captures all east-west environmental variation, notably precipitation (map figure) but also factors that co-vary with precipitation, such as productivity. This statistical model and all those that follow were fit in a Bayesian statistical framework using Stan (Carpenter *et al.*, 2017) and rstan (Team *et al.*, 2018) with vague priors on all parameters. In all cases, model fit was assessed with posterior predictive checks (Conn *et al.*, 2018).

Common garden experiment

Source material and experimental design

We established a common garden experiment at 14 sites throughout and beyond the geographic distribution of *P. arachnifera* (MAP). Experimental sites spanned latitudinal and longitudinal variation, though we focus here on longitude. During the three years of this experiment, total precipitation at each site closely tracked longitude (Fig. A1), as expected based on longer-term climate trends (map). Source material for this experiment came from 8 sites, which were of subset of the sites that were visited for the natural population survey (Table). For a subset of the natural populations we visited, we collected tillers from flowering individuals of each sex (mean: 11.6 individuals per site, range: 2–18) (Table A1). These were brought back to the Rice University greenhouse, where they were clonally propagated in ProMix potting soil and supplemental Osmocote fertilizer at 78–80°F under natural light.

Common gardens were set up in Fall (October–December) 2014. At each site,

128 we established 14 experimental blocks, which typically corresponded to a tree or
129 woodland edge, providing partial shade that mimics this species' natural micro-
130 environment. We planted 3 females and 3 males in each block, for a total of 42
131 individuals per sex per site and 1176 total plants across sites, with all source collec-
132 tions represented at all sites. Individuals were spaced within blocks to allow space
133 for rhizomatous growth that could be clearly attributed to individual transplants.
134 To promote establishment, we cleared vegetation immediately surrounding trans-
135 plants and provided ca. 1 L of water at the time of transplanting but provided no
136 subsequent watering, fertilization, or competitor removal.

137 We visited each site during May of 2015, 2016, and 2017. For each individual in
138 each year, we recorded data for four demographic vital rates: survival status (alive
139 or dead), size (number of tillers), flowering status (reproductive or vegetative), the
140 number of panicles produced by flowering plants.

141 **Statistical analysis of common garden experiment**

142 We analyzed the demographic vital rates with generalized linear mixed models in
143 a hierarchical Bayesian framework. All the vital rates shared a common linear
144 predictor for the expected value that included fixed effects of size, sex, linear and
145 quadratic terms for longitude, and all 2- and 3-way interactions. We included
146 quadratic effects of longitude to account for the possibility of non-monotonic re-
147 sponses, following the hypothesis that fitness may peak in the center of the range.
148 The linear predictor also included random effects of site, block, and source pop-
149 ulation of the transplant; the corresponding variance terms were used in the de-
150 mographic model (below) to capture process error in demography. We pooled all
151 three years of observations for analysis so we do not explicitly model temporal

152 variation but our results are implicitly averaged over years.

153 The survival and flowering data were Bernoulli distributed, and these models
154 applied the logit link function. We modeled tiller and panicle counts as zero-
155 truncated negative binomial using the log link. For flowering and panicle pro-
156 duction in year t , the size covariate was the natural logarithm of tiller number
157 in year t . For survival and size in year t , the size covariate was the natural
158 logarithm of tiller number in year $t - 1$ (for 2015 data, size in year $t - 1$ was
159 transplant size at the time of planting). Code for these analyses is available at
160 <https://github.com/texmiller/POAR-range-limits>.

161 Sex ratio experiment

162 At one site near the center of the range (Lake Lewisville Environmental Learning
163 Area: star in MAP), we established a separate experiment to quantify how sex ra-
164 tio variation affects female reproductive success. Details of this experiment, which
165 was conducted in 2014–2015, are described in Compagnoni et al. 2017. Briefly, we
166 established 124 experimental populations in $0.4m \times 0.4m$ plots that varied in popu-
167 lation density (1–48 plants/plot) and sex ratio (0–100%female), with 2–4 replicates
168 each of 34 density-sex ratio combinations. The experiment was established ca. 1
169 km from a natural population at this site and plots were situated with a minimum
170 of 15 m spacing, a buffer that was intended to limit pollen movement between
171 plots (pilot data indicated that $\geq 90\%$ of wind pollination occurred within 13m).
172 We measured female reproductive success in different density and sex ratio envi-
173 ronments by collecting panicles from a subset of females in each plot at the end of
174 the reproductive season. In the lab, we counted the total number of seeds on each

175 panicle and assessed seed viability with tetrazolium assays of 25 seeds per panicle.
176 We also conducted germination trials in the greenhouse (17–57 seeds per panicle,
177 mode: 30).

178 Statistical analysis of sex ratio experiment

179 Our previous study examined how interactions between density and frequency (sex
180 ratio) dependence contributed to female reproductive success (Compagnoni *et al.*,
181 2017). Here we focus solely on sex ratio variation, averaging over variation in
182 density. Our goal was to estimate a ‘mating function’ that defines how availability
183 of male panicles affects the viability of seeds on female panicles. We modeled the
184 seed viability data with a binomial distribution where the probability of viability
185 (p_v) was given by:

$$v = v_0 * (1 - OSR^\alpha) \quad (1)$$

186 where OSR is the operational sex ratio (fraction of panicles that were female) in
187 our experimental populations. This function has the properties, supported by our
188 previous work, that seed viability is maximized at v_0 as OSR approaches zero and
189 goes to zero as OSR approaches 1. Parameter α controls how viability declines
190 with increasing female bias.

191 We modeled germination data from greenhouse trials similarly, where counts of
192 germinants were modeled as binomial successes. Since germination was conditional
193 on seed viability, the probability of success was given by the product $v * g$, where
194 v is a function of OSR (Eq. 1) and g is assumed to be constant. The germination

195 trials alone do not provide enough information to independently estimate v and
 196 g but the combination of viability and germination data allowed us to do so. For
 197 both viability and germination, we found that accounting for overdispersion with
 198 a beta-binomial response distribution improved model fit.

199 Demographic model of range limits

200 The statistical models for the common garden and sex ratio experiments provided
 201 the backbone of the full demographic model, a matrix projection model (MPM)
 202 structured by size (tiller number) and sex. Following the statistical modeling, the
 203 MPM accommodates longitude as a predictor variable, allowing us to identify the
 204 longitudinal limits of population viability ($\lambda \geq 1$) and investigate the underlying
 205 drivers of population decline at range limits. For a given longitude, let $F_{x,t}$ and $M_{x,t}$
 206 be the number of female and male plants of size x in year t , where $x \in \{1, 2, \dots, U\}$
 207 and U is the maximum number of tillers a plant can attain (assumed to be the
 208 same for females and males). For a pre-breeding census, the coupled dynamics of
 209 both sexes are given by:

$$F_{y,t+1} = \sum_{x=1}^U [s^F(x) \cdot g^F(y, x) + r^F(x, \mathbf{F}_t, \mathbf{M}_t) \cdot \rho \cdot e(y)] F_{x,t} \quad (2)$$

$$M_{y,t+1} = \sum_{x=1}^U [s^M(x) \cdot g^M(y, x)] M_{x,t} + [r^F(x, \mathbf{F}_t, \mathbf{M}_t) \cdot (1 - \rho) \cdot e(y)] F_{x,t} \quad (3)$$

210 s and g give the probabilities of surviving at size x and growing from sizes x to
 211 y , respectively, and superscripts indicate that these functions may be unique to
 212 females (F) and males (M). r^F gives fertility of size- x females (number of new

213 recruits per female per year) and is dependent on population structure (the $U \times 1$
 214 vectors \mathbf{F}_t and \mathbf{M}_t). Parameter ρ is the primary sex ratio (proportion of recruits
 215 that are female) and $e(y)$ is the probability distribution of recruit size. We assume
 216 that all recruits first appear in the population at 1 tiller in size regardless of sex,
 217 so $e(y = 1) = 1$ and $e(y > 1) = 0$. The female fertility function is further defined
 218 as:

$$r^F(x, \mathbf{F}_t, \mathbf{M}_t) = p^F(x) \cdot c^F(x) \cdot d \cdot v(\mathbf{F}_t, \mathbf{M}_t) \cdot g \quad (4)$$

219 where p and c are flowering probability and panicle production for females of size
 220 x , d is the number of seeds (fertilized or unfertilized) per female panicle, v is the
 221 probability that a seed is fertilized, and g is the probability that a fertilized seed
 222 germinates. Lastly, and following Eq. 1, seed fertilization depends on the OSR
 223 according to:

$$v(\mathbf{F}_t, \mathbf{M}_t) = v_0 * \left[1 - \left(\frac{\sum_{x=1}^U p^F(x) c^F(x) F_{x,t}}{\sum_{x=1}^U p^F(x) c^F(x) F_{x,t} + p^M(x) c^M(x) M_{x,t}} \right)^\alpha \right] \quad (5)$$

224 Results

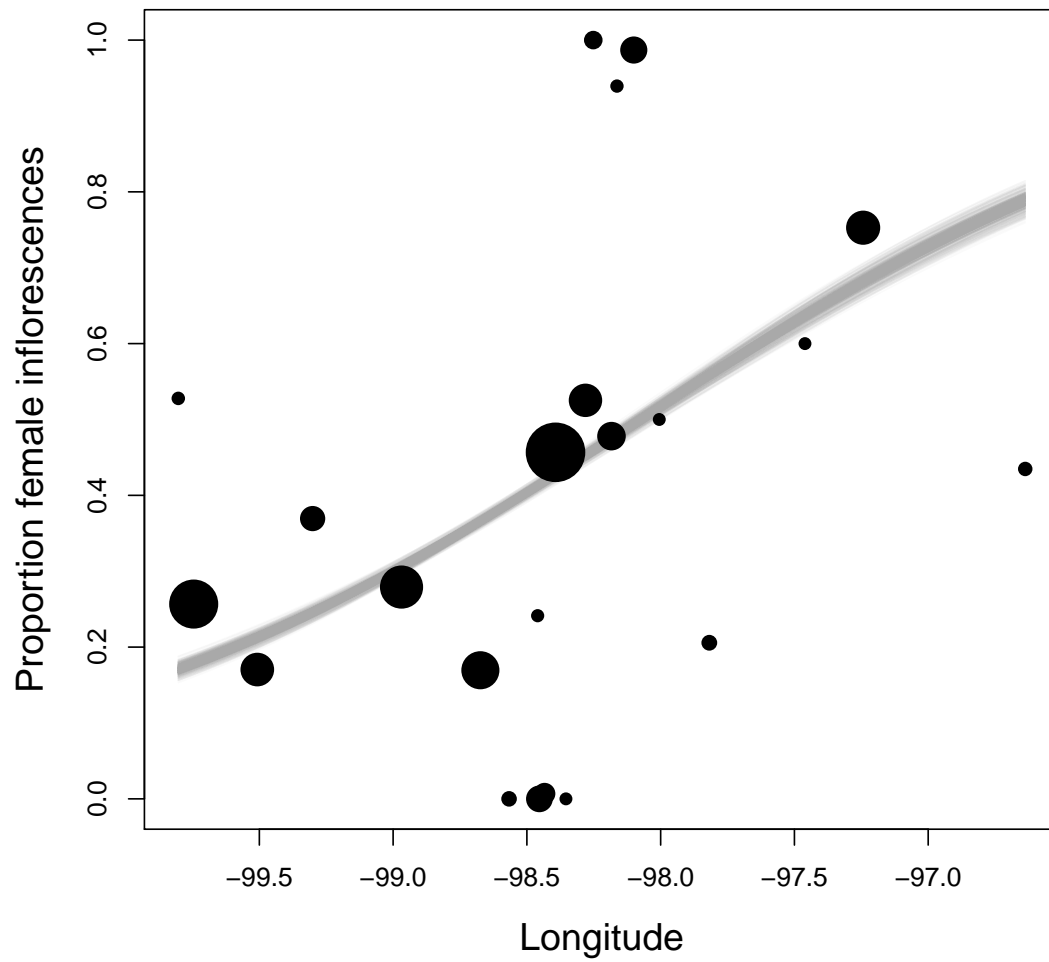


Figure 2: Caption.

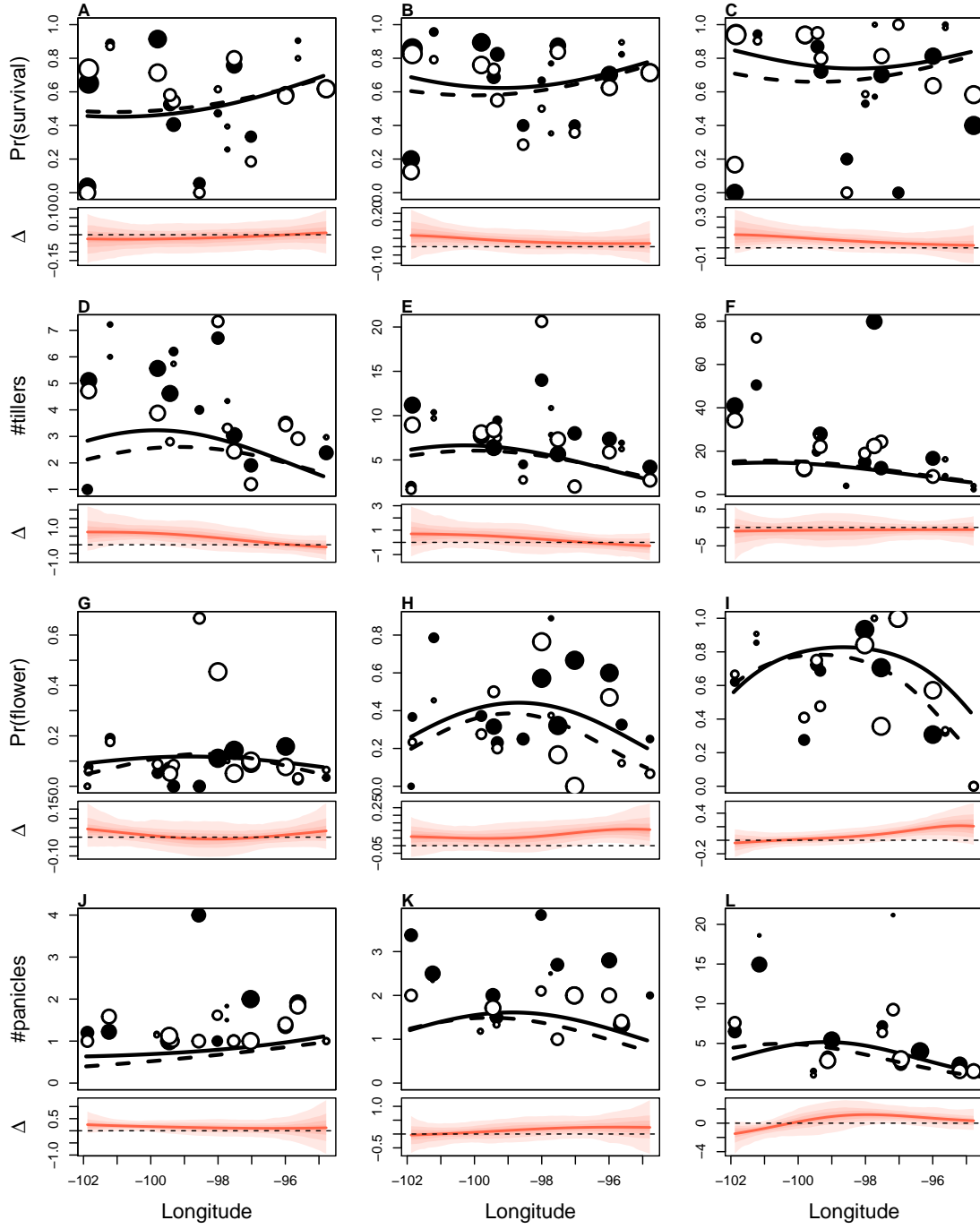


Figure 3: Caption.

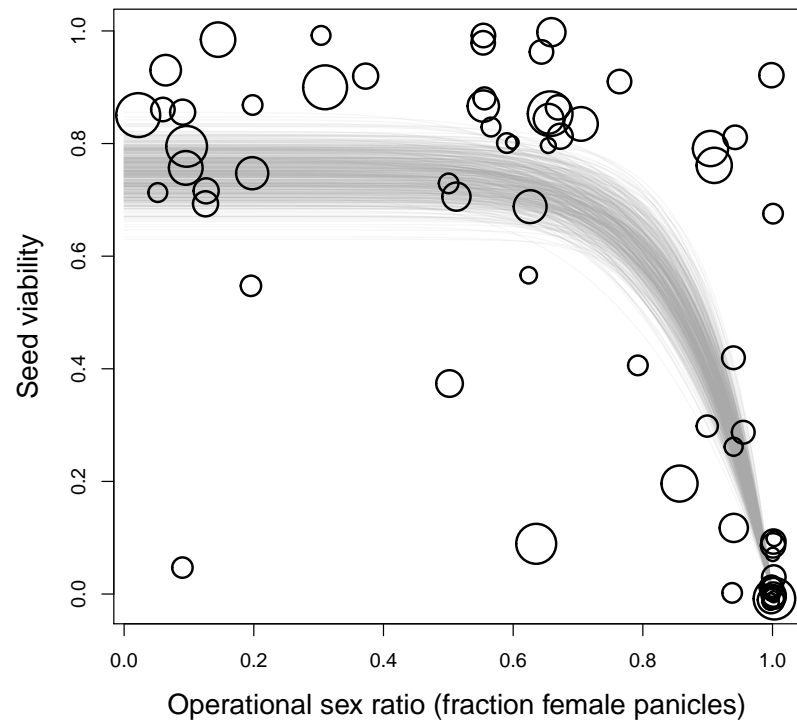


Figure 4: Caption.

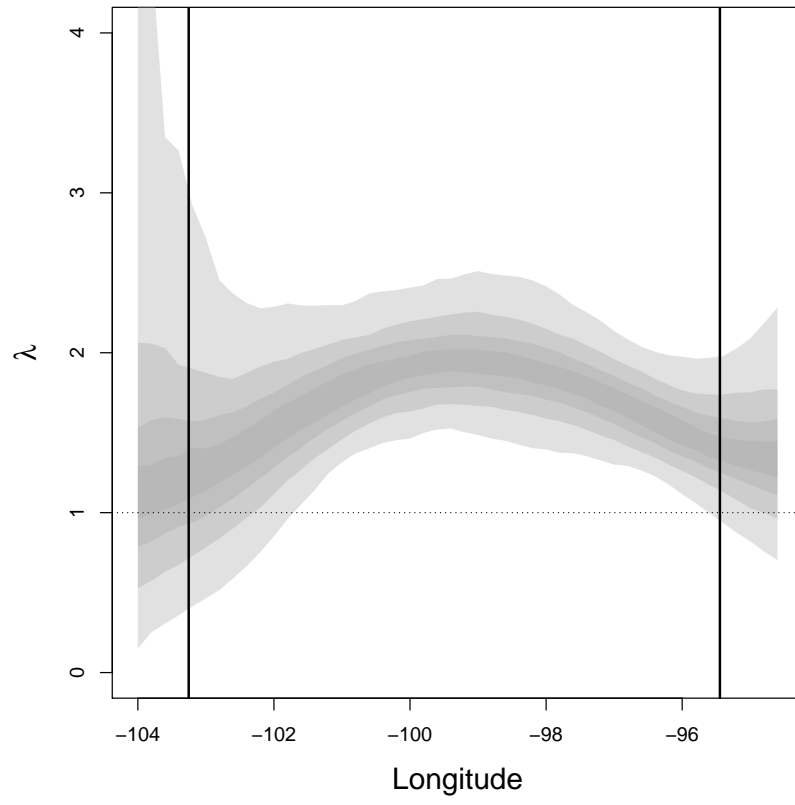


Figure 5: Caption.

225 Discussion

226 Acknowledgements

227 Author contributions

228 Data accessibility

229 References

- 230 Araújo MS, Bolnick DI, Layman CA (2011) The ecological causes of individual
231 specialisation. *Ecology letters*, **14**, 948–958.
- 232 Bertiller MB, Sain CL, Bisigato AJ, Coronato FR, Aries JO, Graff P (2002) Spatial
233 sex segregation in the dioecious grass *Poa ligularis* in northern patagonia: the
234 role of environmental patchiness. *Biodiversity & Conservation*, **11**, 69–84.
- 235 Bierzychudek P, Eckhart V (1988) Spatial segregation of the sexes of dioecious
236 plants. *The American Naturalist*, **132**, 34–43.
- 237 Bolnick DI, Doebeli M (2003) Sexual dimorphism and adaptive speciation: two
238 sides of the same ecological coin. *Evolution*, **57**, 2433–2449.
- 239 Bolnick DI, Svanbäck R, Fordyce JA, Yang LH, Davis JM, Hulsey CD, Forister
240 ML (2002) The ecology of individuals: incidence and implications of individual
241 specialization. *The American Naturalist*, **161**, 1–28.
- 242 Bowyer RT (2004) Sexual segregation in ruminants: definitions, hypotheses, and

243 implications for conservation and management. *Journal of Mammalogy*, **85**,
244 1039–1052.

245 Carpenter B, Gelman A, Hoffman MD, *et al.* (2017) Stan: A probabilistic pro-
246 gramming language. *Journal of statistical software*, **76**.

247 Caruso C, Case A (2007) Sex ratio variation in gynodioecious lobelia siphilitica:
248 effects of population size and geographic location. *Journal of Evolutionary Bi-*
249 *ology*, **20**, 1396–1405.

250 Compagnoni A, Steigman K, Miller TE (2017) Can’t live with them, can’t live
251 without them? balancing mating and competition in two-sex populations. *Pro-*
252 *ceedings of the Royal Society B: Biological Sciences*, **284**, 20171999.

253 Conn PB, Johnson DS, Williams PJ, Melin SR, Hooten MB (2018) A guide to
254 bayesian model checking for ecologists. *Ecological Monographs*, **88**, 526–542.

255 Darwin C (2019) *The descent of man*. BoD–Books on Demand.

256 Diez JM, Giladi I, Warren R, Pulliam HR (2014) Probabilistic and spatially vari-
257 able niches inferred from demography. *Journal of ecology*, **102**, 544–554.

258 Ehrlén J, Morris WF (2015) Predicting changes in the distribution and abundance
259 of species under environmental change. *Ecology Letters*, **18**, 303–314.

260 Eppley S (2001) Gender-specific selection during early life history stages in the
261 dioecious grass *distichlis spicata*. *Ecology*, **82**, 2022–2031.

262 Evans ME, Merow C, Record S, McMahon SM, Enquist BJ (2016) Towards
263 process-based range modeling of many species. *Trends in Ecology & Evolution*,
264 **31**, 860–871.

- 265 Field DL, Pickup M, Barrett SC (2013) Ecological context and metapopulation
266 dynamics affect sex-ratio variation among dioecious plant populations. *Annals*
267 *of botany*, **111**, 917–923.
- 268 Gianuca D, Votier SC, Pardo D, *et al.* (2019) Sex-specific effects of fisheries and
269 climate on the demography of sexually dimorphic seabirds. *Journal of Animal*
270 *Ecology*.
- 271 Groen KE, Stieha CR, Crowley PH, McLetchie DN (2010) Sex-specific plant re-
272 sponses to light intensity and canopy openness: implications for spatial segre-
273 gation of the sexes. *Oecologia*, **162**, 561–570.
- 274 Hargreaves AL, Samis KE, Eckert CG (2013) Are species' range limits simply niche
275 limits writ large? a review of transplant experiments beyond the range. *The*
276 *American Naturalist*, **183**, 157–173.
- 277 Holt RD (2009) Bringing the hutchinsonian niche into the 21st century: ecological
278 and evolutionary perspectives. *Proceedings of the National Academy of Sciences*,
279 **106**, 19659–19665.
- 280 Hutchinson GE (1958) Concluding remarks. In: *Cold Spring Harbour Symposium*
281 *on Quantitative Biology*, vol. 22, pp. 415—427.
- 282 Ketterson ED, Nolan Jr V (1976) Geographic variation and its climatic correlates
283 in the sex ratio of eastern-wintering dark-eyed juncos (*Junco hyemalis hyemalis*).
284 *Ecology*, **57**, 679–693.
- 285 Lee-Yaw JA, Kharouba HM, Bontrager M, *et al.* (2016) A synthesis of transplant

286 experiments and ecological niche models suggests that range limits are often
 287 niche limits. *Ecology letters*, **19**, 710–722.

288 Merow C, Bois ST, Allen JM, Xie Y, Silander JA (2017) Climate change both
 289 facilitates and inhibits invasive plant ranges in new england. *Proceedings of the*
 290 *National Academy of Sciences*, **114**, E3276–E3284.

291 Merow C, Latimer AM, Wilson AM, McMahon SM, Rebelo AG, Silander Jr JA
 292 (2014) On using integral projection models to generate demographically driven
 293 predictions of species’ distributions: development and validation using sparse
 294 data. *Ecography*, **37**, 1167–1183.

295 Pekár S, Martišová M, Bilde T (2011) Intersexual trophic niche partitioning in an
 296 ant-eating spider (araneae: Zodariidae). *PloS one*, **6**, e14603.

297 Petry WK, Soule JD, Iler AM, Chicas-Mosier A, Inouye DW, Miller TE, Mooney
 298 KA (2016) Sex-specific responses to climate change in plants alter population
 299 sex ratio and performance. *Science*, **353**, 69–71.

300 Phillips R, Silk J, Phalan B, Catry P, Croxall J (2004) Seasonal sexual segregation
 301 in two thalassarche albatross species: competitive exclusion, reproductive role
 302 specialization or foraging niche divergence? *Proceedings of the Royal Society of*
 303 *London. Series B: Biological Sciences*, **271**, 1283–1291.

304 Renganayaki K, Jessup R, Burson B, Hussey M, Read J (2005) Identification of
 305 male-specific aflp markers in dioecious texas bluegrass. *Crop science*, **45**, 2529–
 306 2539.

307 Renganayaki K, Read J, Fritz A (2001) Genetic diversity among texas bluegrass

308 genotypes (poa arachnifera torr.) revealed by aflu and rapd markers. *Theoretical*
309 *and Applied Genetics*, **102**, 1037–1045.

310 Renner SS, Ricklefs RE (1995) Dioecy and its correlates in the flowering plants.
311 *American journal of botany*, **82**, 596–606.

312 Rozas V, DeSoto L, Olano JM (2009) Sex-specific, age-dependent sensitivity of
313 tree-ring growth to climate in the dioecious tree juniperus thurifera. *New Phy-*
314 *tologist*, **182**, 687–697.

315 Shelton AO (2010) The ecological and evolutionary drivers of female-biased sex
316 ratios: two-sex models of perennial seagrasses. *The American Naturalist*, **175**,
317 302–315.

318 Shine R (1989) Ecological causes for the evolution of sexual dimorphism: a review
319 of the evidence. *The Quarterly Review of Biology*, **64**, 419–461.

320 Team SD, *et al.* (2018) Rstan: the r interface to stan. r package version 2.17. 3.

321 Temeles EJ, Miller JS, Rifkin JL (2010) Evolution of sexual dimorphism in bill size
322 and shape of hermit hummingbirds (phaethornithinae): a role for ecological cau-
323 sation. *Philosophical Transactions of the Royal Society B: Biological Sciences*,
324 **365**, 1053–1063.

325 Veran S, Beissinger SR (2009) Demographic origins of skewed operational and
326 adult sex ratios: perturbation analyses of two-sex models. *Ecology Letters*, **12**,
327 129–143.

Appendix A: Site locations and climate

	Population	Latitude	Longitude
1	Canyon_of_Eagles	30.88	-98.43
2	ClearBay-Thunderbird	35.23	-97.24
3	CooperWMA	36.60	-99.51
4	Copper Breaks	34.10	-99.75
5	Dinosaur_Valley	32.25	-97.82
6	Fort_Worth_Nature_Center	32.83	-97.46
7	Ft Cobb	35.18	-98.45
8	Ft Richardson	33.20	-98.16
9	Great Plains	34.74	-98.97
10	Great_Salt_Plains	36.79	-98.18
11	Horn_Hill_Cemetery	31.56	-96.64
12	Kingman_Fishing_Lake	37.65	-98.28
13	Lake Arrowhead	33.75	-98.39
14	Mineral_Wells	32.89	-98.01
15	Pedernales_Falls	30.33	-98.25
16	Possum Kingdom	32.87	-98.57
17	Quartz_Mountain	34.89	-99.30
18	Red Rock Canyon	35.44	-98.35
19	Red_River	34.13	-98.10
20	South_Llano	30.45	-99.80
21	Sulfur_Springs	31.08	-98.46
22	Wichita_Mountains	34.70	-98.67

Table A1: Sites of natural population surveys corresponding to Figure

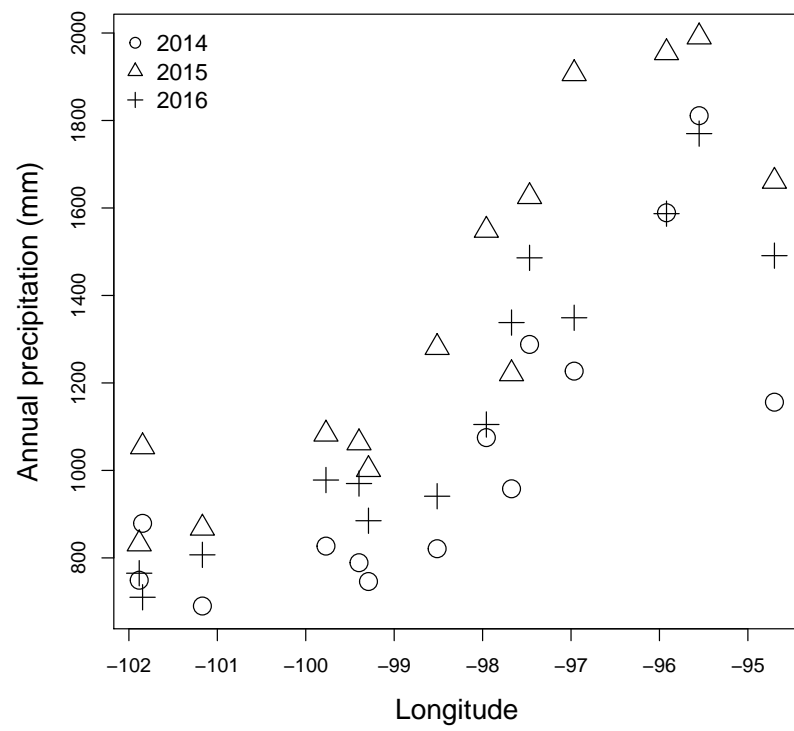


Figure A1: Caption.