

# Principles of differential expression analysis

Charlotte Soneson

Friedrich Miescher Institute for Biomedical Research &  
SIB Swiss Institute of Bioinformatics



Swiss Institute of  
Bioinformatics



Friedrich Miescher Institute  
for Biomedical Research

“To call in the statistician after the experiment is done may be no more than asking him to perform a postmortem examination: he may be able to say what the experiment died of.”

**Sir Ronald Fisher, Indian Statistical Congress, Sankhya, around 1938**



**Stephen John Senn**

@stephensenn



 Follow

Statisticians are the bad fairies of research.  
People forget to invite them until it's too late, at  
which point they send everyone to sleep.

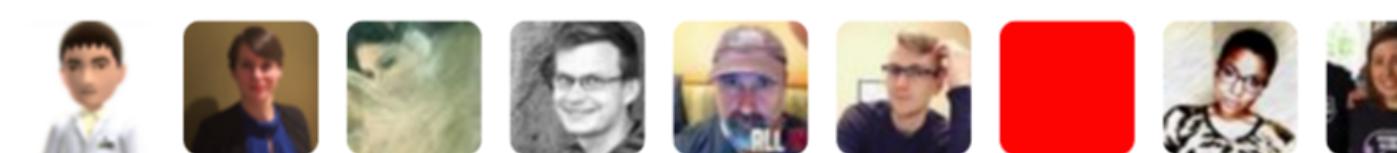
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RETWEETS

92

LIKES

93



11:22 AM - 21 Feb 2016

# Different types of experiments

Learning experiment questions	Confirming experiment questions
<ul style="list-style-type: none"><li>• Does the drug have toxic side effects (at what dose, given for how long, in which tissue)?</li><li>• Does stress affect rodent behaviour (what kind of stress, for how long, on what behavioural tasks)?</li><li>• How does exercise affect cognitive functioning of older people (what type of exercise, how much, which aspect of cognition)?</li></ul>	<ul style="list-style-type: none"><li>• Does 5 mg/kg of the drug given once a day for 5 days increase blood creatinine<sup>a</sup> concentration?</li><li>• Does fox urine odour (a stressor) affect the amount of food Wistar rats consume during the first 24 hours after exposure?</li><li>• Does 30 min of aerobic activity (treadmill running) at 60% VO<sub>2</sub> max<sup>b</sup>, 3 days a week for 6 weeks, in males between 55–70 years of age, improve performance on a mental rotation task?</li></ul>

<sup>a</sup> Increased creatinine indicates kidney damage.

<sup>b</sup> VO<sub>2</sub> max is the maximal oxygen uptake and is a measure of a person's aerobic fitness.

# What is experimental design?

The organization of an experiment, to ensure that the **right type** of data, and **enough** of it, is available to answer the **questions of interest** as clearly and efficiently as possible.

# What affects the outcome of an experiment?

$$\text{Outcome} = \underbrace{\text{Treatment effects}}_{\begin{array}{l} \text{Environment} \\ \text{Compound} \\ \text{Inhibitor} \\ \text{siRNA} \\ \text{Dose} \\ \text{Time} \end{array}} + \underbrace{\text{Biological effects}}_{\begin{array}{l} \text{Sex} \\ \text{Age} \\ \text{Weight} \\ \text{Litter} \\ \text{Genotype} \\ \text{Species} \\ \text{Cell line} \end{array}} + \underbrace{\text{Technical effects}}_{\begin{array}{l} \text{Technician} \\ \text{Batch} \\ \text{Plate} \\ \text{Cage} \\ \text{Array} \\ \text{Day} \\ \text{Order} \\ \text{Source} \end{array}} + \underbrace{\text{Error}}_{\begin{array}{l} \text{Experimental} \\ \text{Treatment} \\ \text{Sampling} \\ \text{Measurement} \end{array}}$$

# What is **bad** experimental design?

Analysis batch I / Study center I / Processing protocol I ...

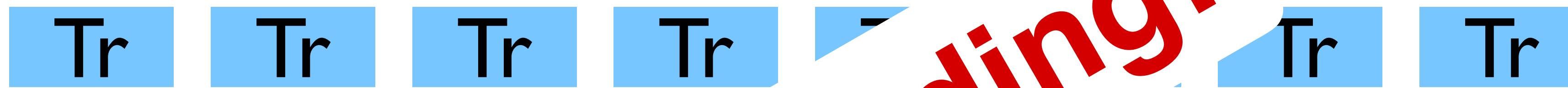
Tr Tr Tr Tr Tr Tr Tr Tr

Analysis batch II / Study center II / Processing protocol II ...

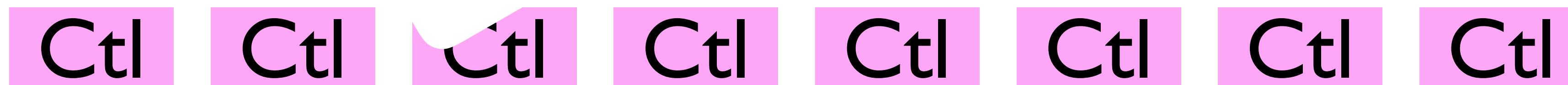
Ctl Ctl Ctl Ctl Ctl Ctl Ctl Ctl

# What is **bad** experimental design?

Analysis batch I / Study center I / Processing protocol I ...



Analysis batch II / Study center II / Processing protocol II ...



Confounding!

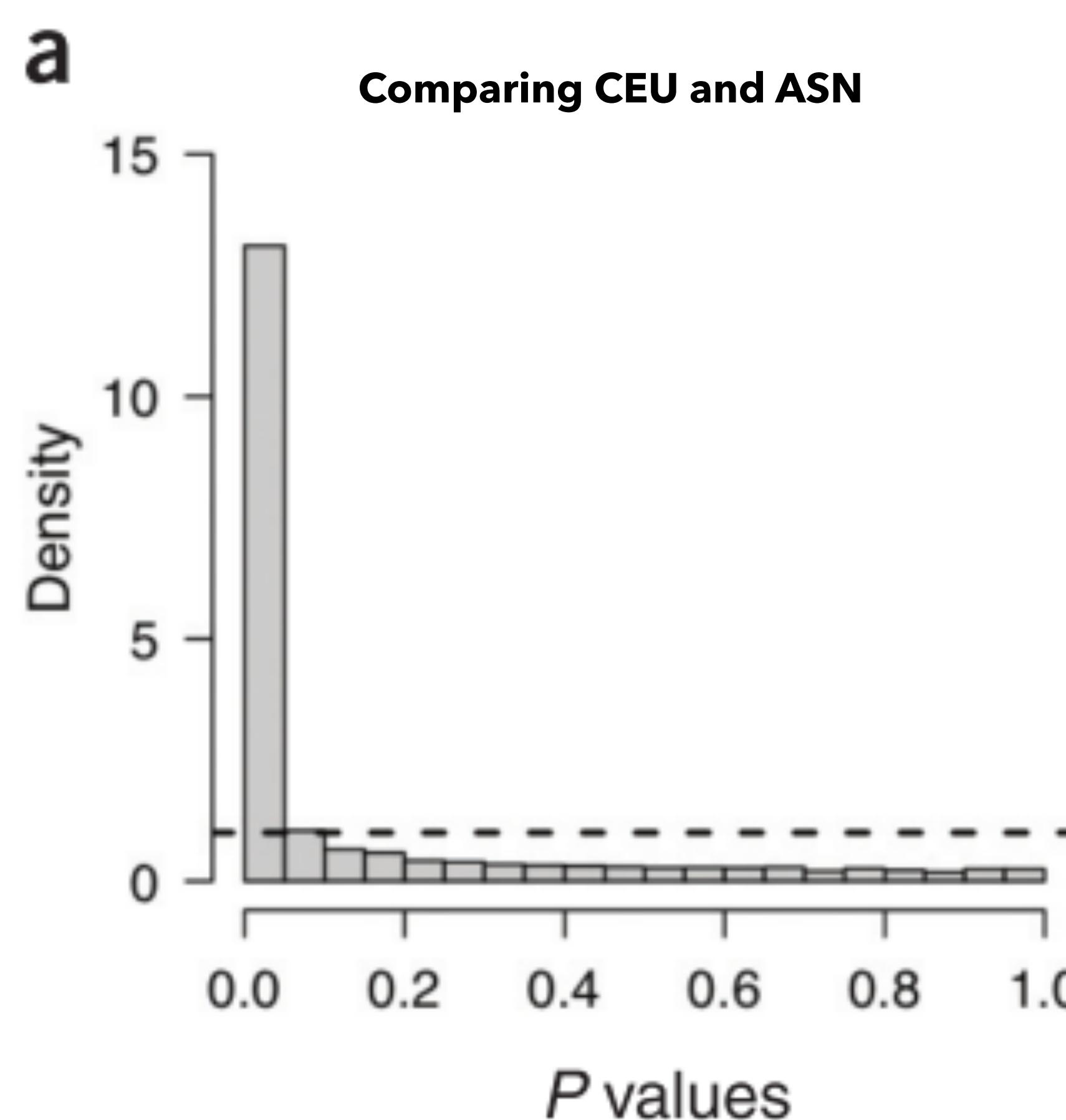
# What can happen with bad experimental design?

- Example: gene expression study comparing 60 CEU and 82 ASN HapMap individuals
- 26% of the genes were found to be significantly differentially expressed (78% with less restrictive multiple testing correction)
- **But:** all CEU samples were processed (sometimes years) before all the ASN samples!

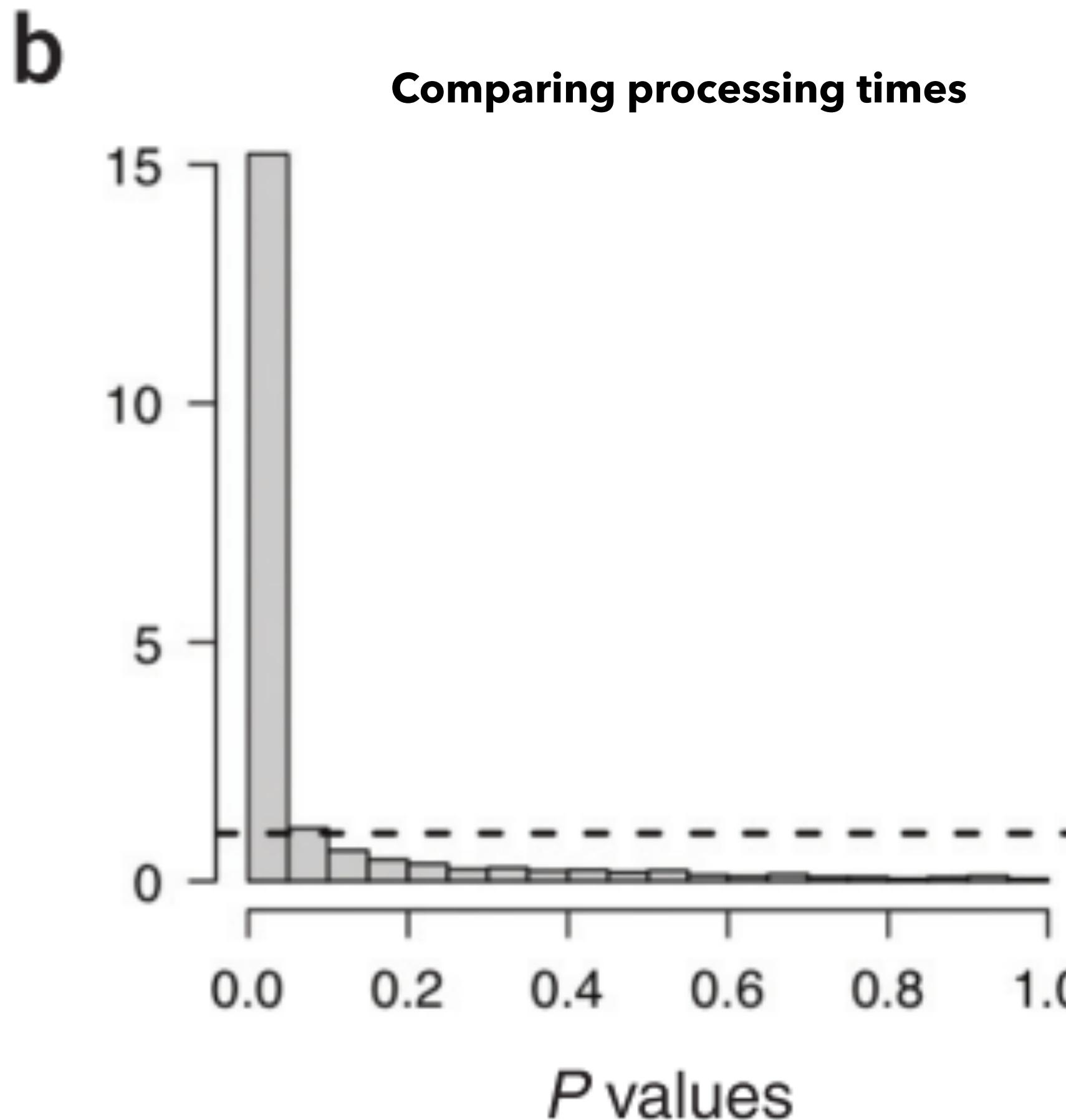
# What can happen with bad experimental design?

- Example: gene expression study comparing 111 CEU and 82 ASN HapMap individuals
  - 26% of the genes were significantly differentially expressed (78% with less resampling after FDR multiple testing correction)
  - **But:** all CEU samples were processed (sometimes years) before all the ASN samples!
- Confounding!**

# What can happen with bad experimental design?



**78% differentially expressed**



**96% differentially expressed**

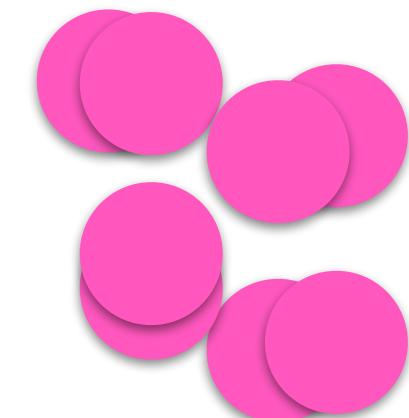
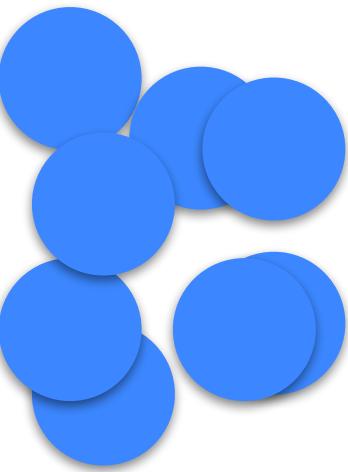
# What would be a better experimental design?

- Process all samples at the same time/in one batch (not always feasible)
- Minimize confounding as much as possible through
  - blocking
  - randomization
- Batch effects may still be present, but with an appropriate design we can account for them

Nonzero batch effect  
Zero treatment effect

**X** Confounded design

Gene expression



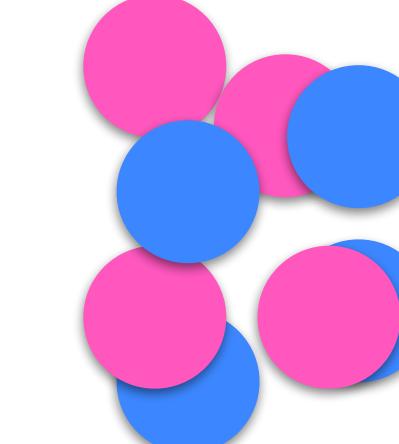
Batch A

Batch B

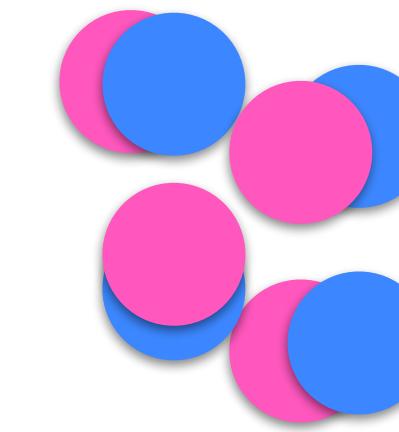
Treated  
Untreated

**✓** Non-confounded design

Gene expression



Batch A

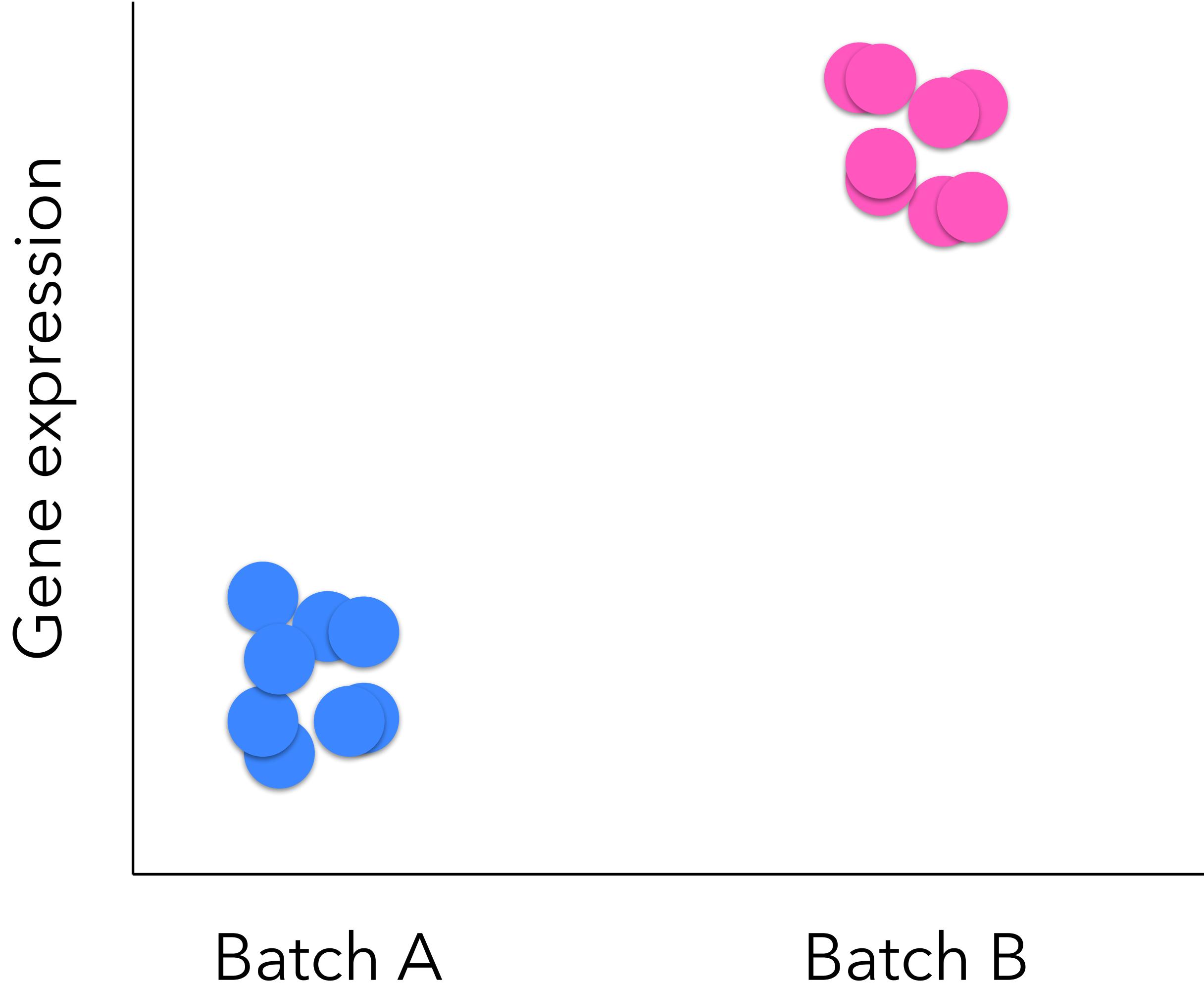


Batch B

Nonzero batch effect

Nonzero treatment effect

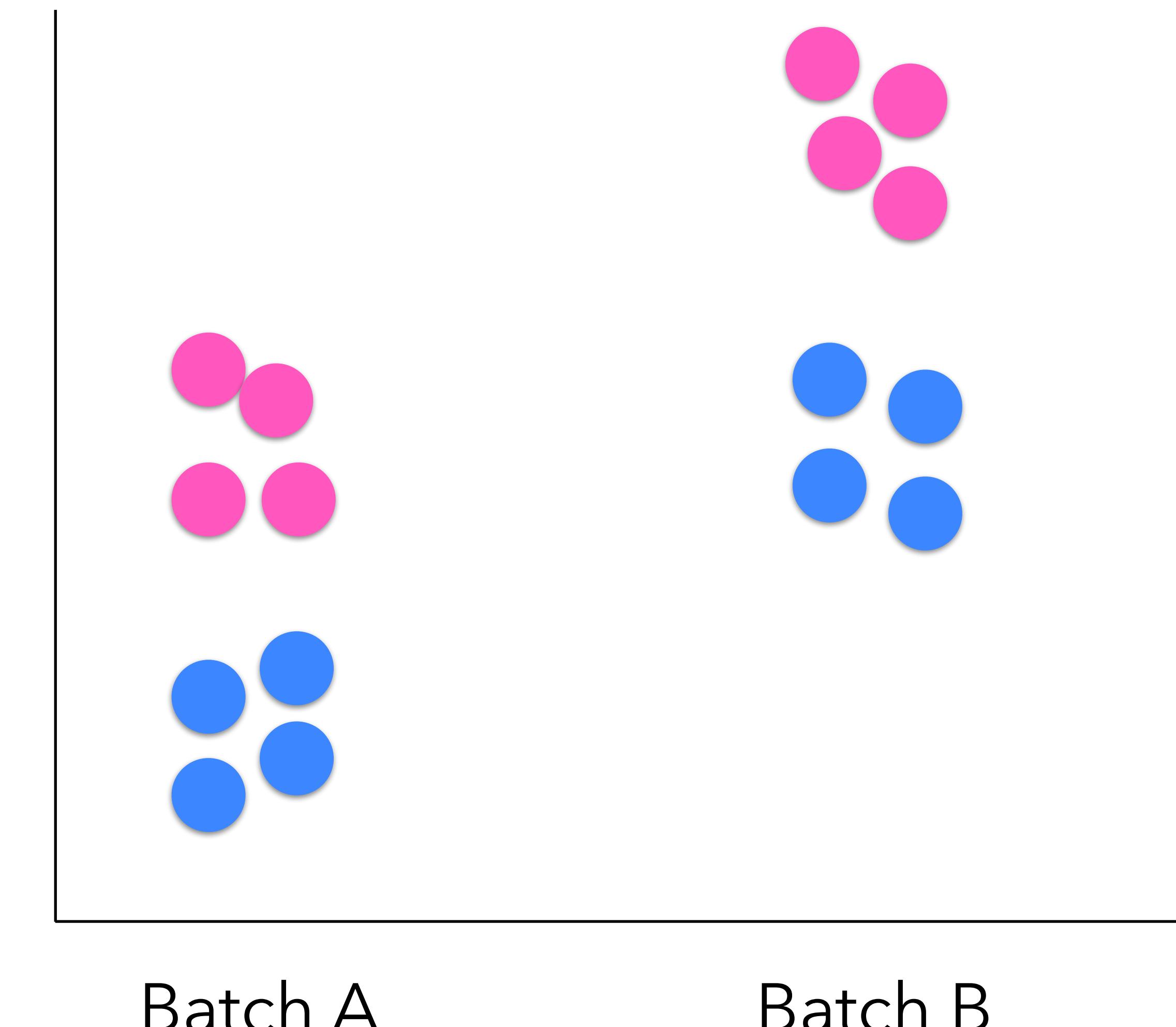
**X** Confounded design



Treated

Untreated

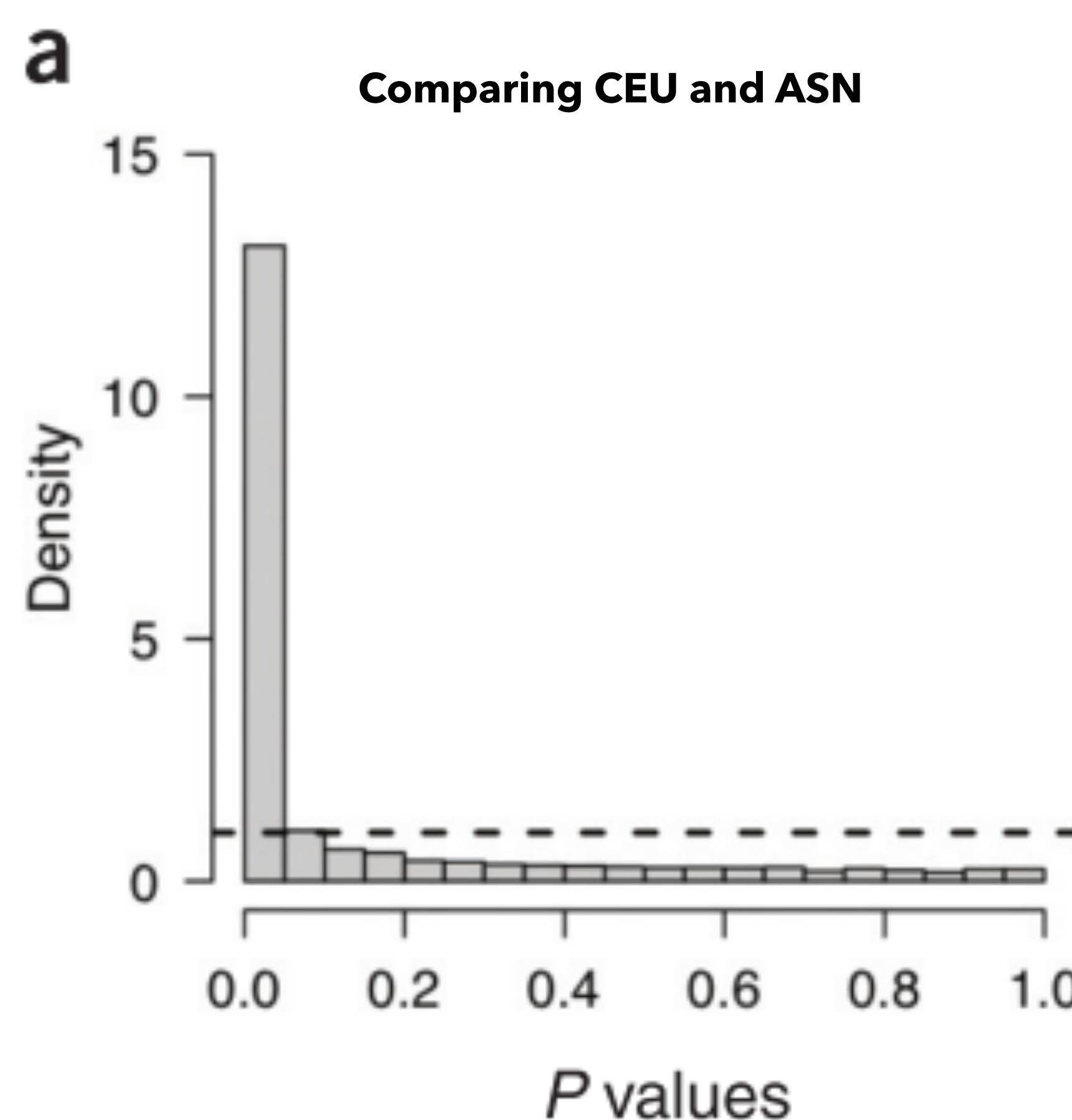
✓ Non-confounded design



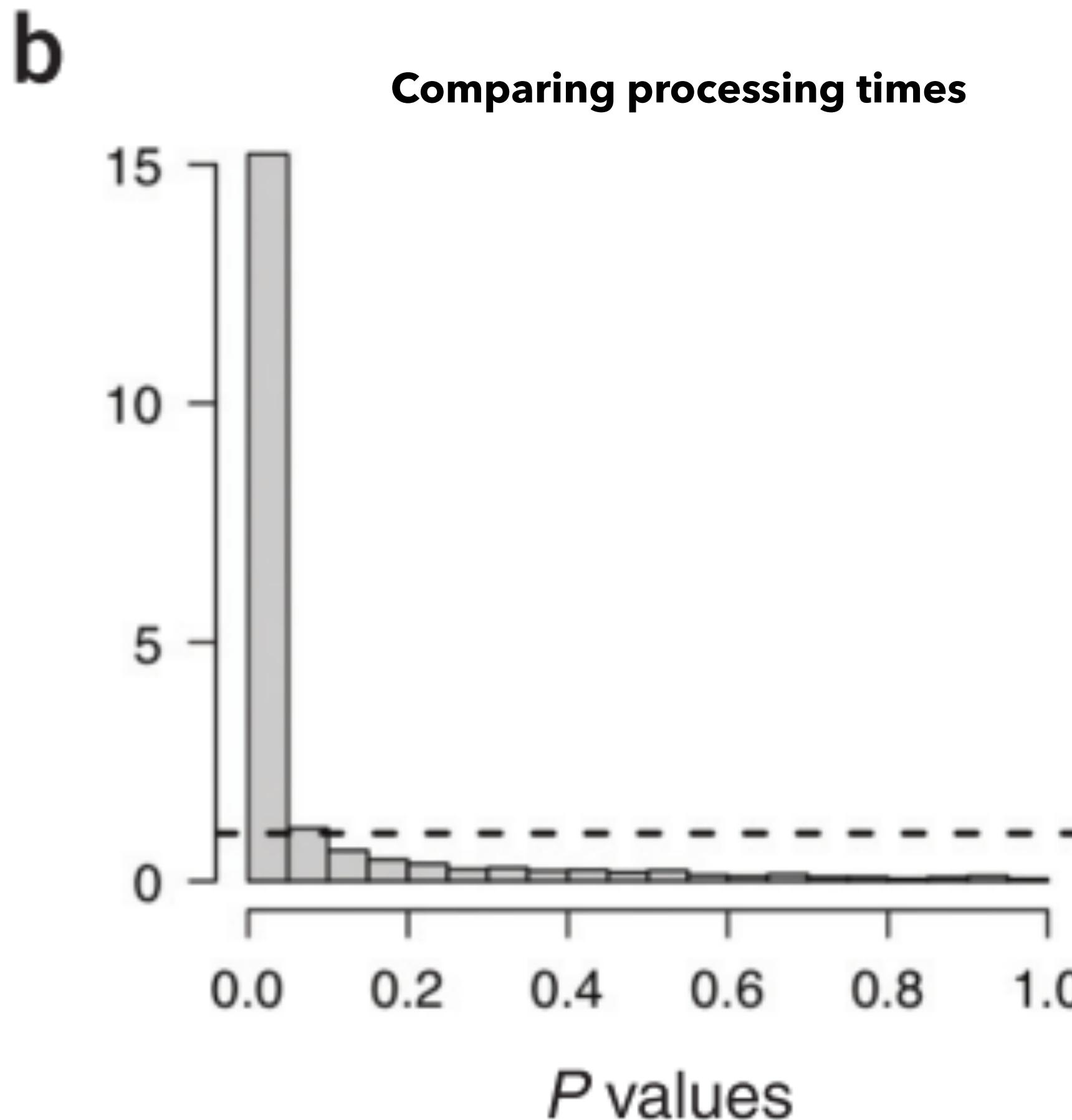
# Dealing with batch effects

- In statistical modeling, batch effects can be included as **covariates** (additional predictors) in the model.
- For exploratory analysis, we often attempt to “eliminate” or “adjust for” such unwanted variation in advance, by subtracting the estimated effect from each variable (e.g. the expression of a gene).
- Even partial confounding between batch and signal of interest can lead to problems.

# What can happen with bad experimental design?



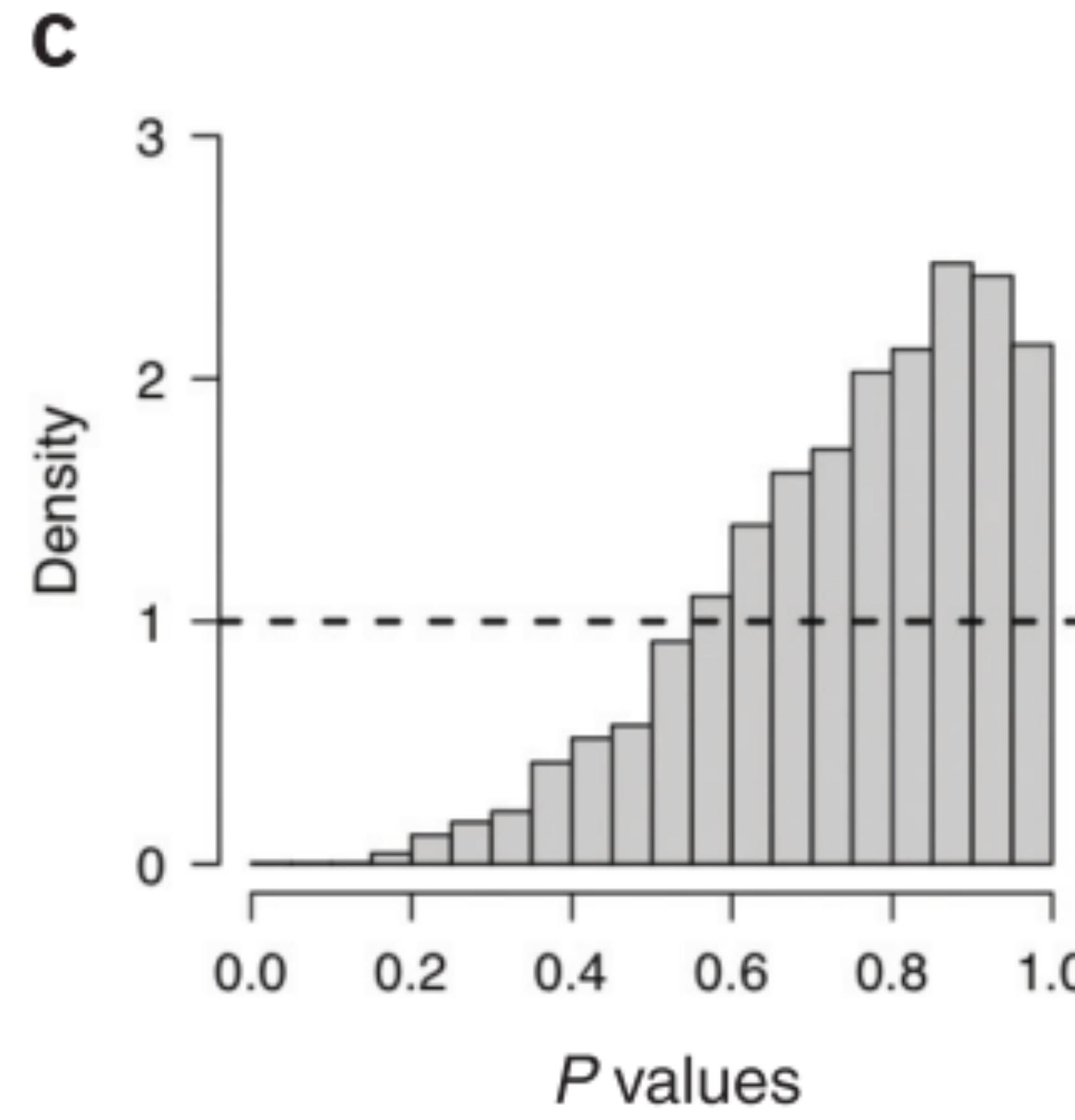
**78% differentially expressed**



**96% differentially expressed**

# “Batch effect correction” won’t work here

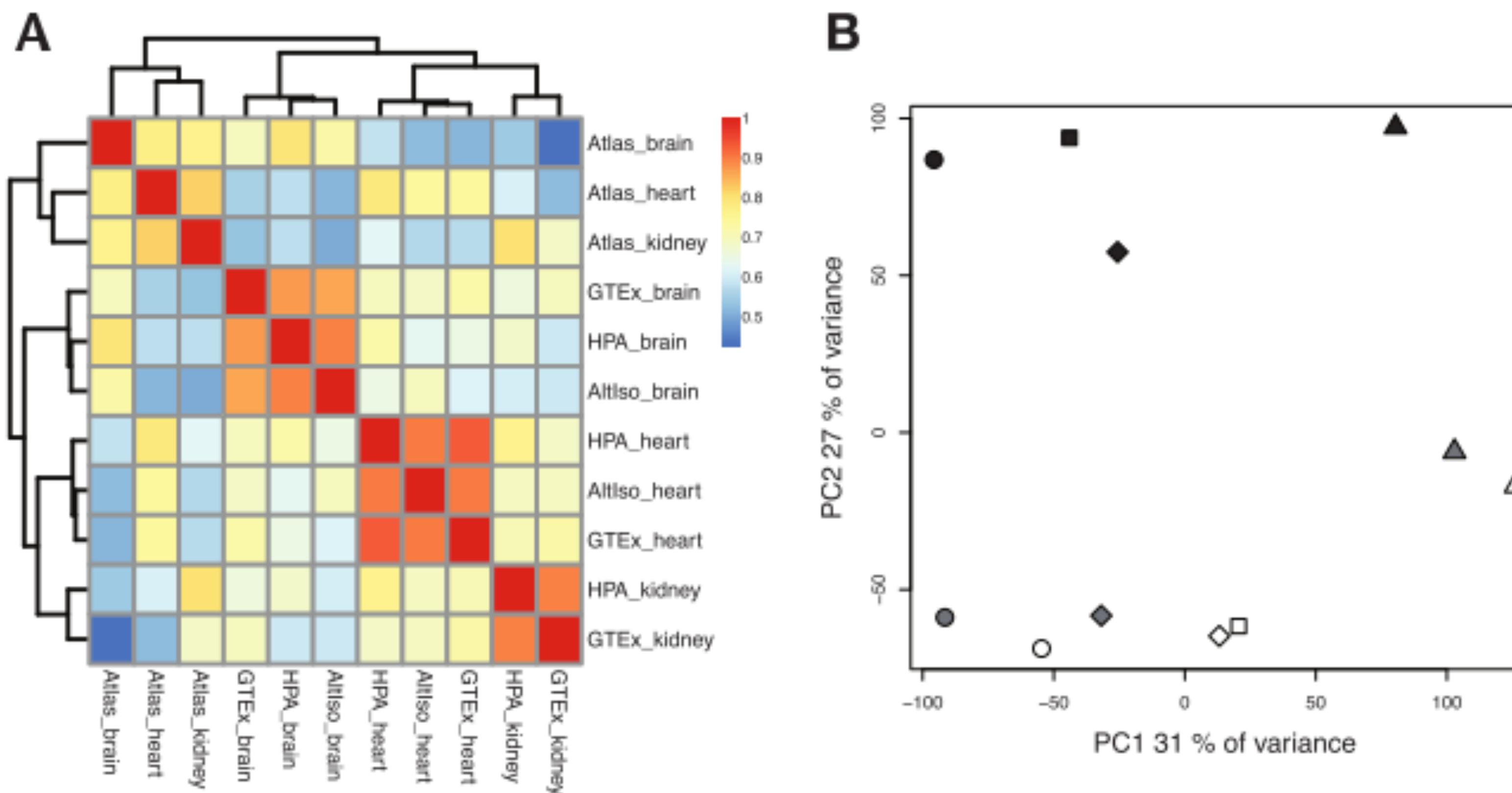
p-values from test comparing CEU and ASN, after controlling for the processing year



**0% differentially expressed**

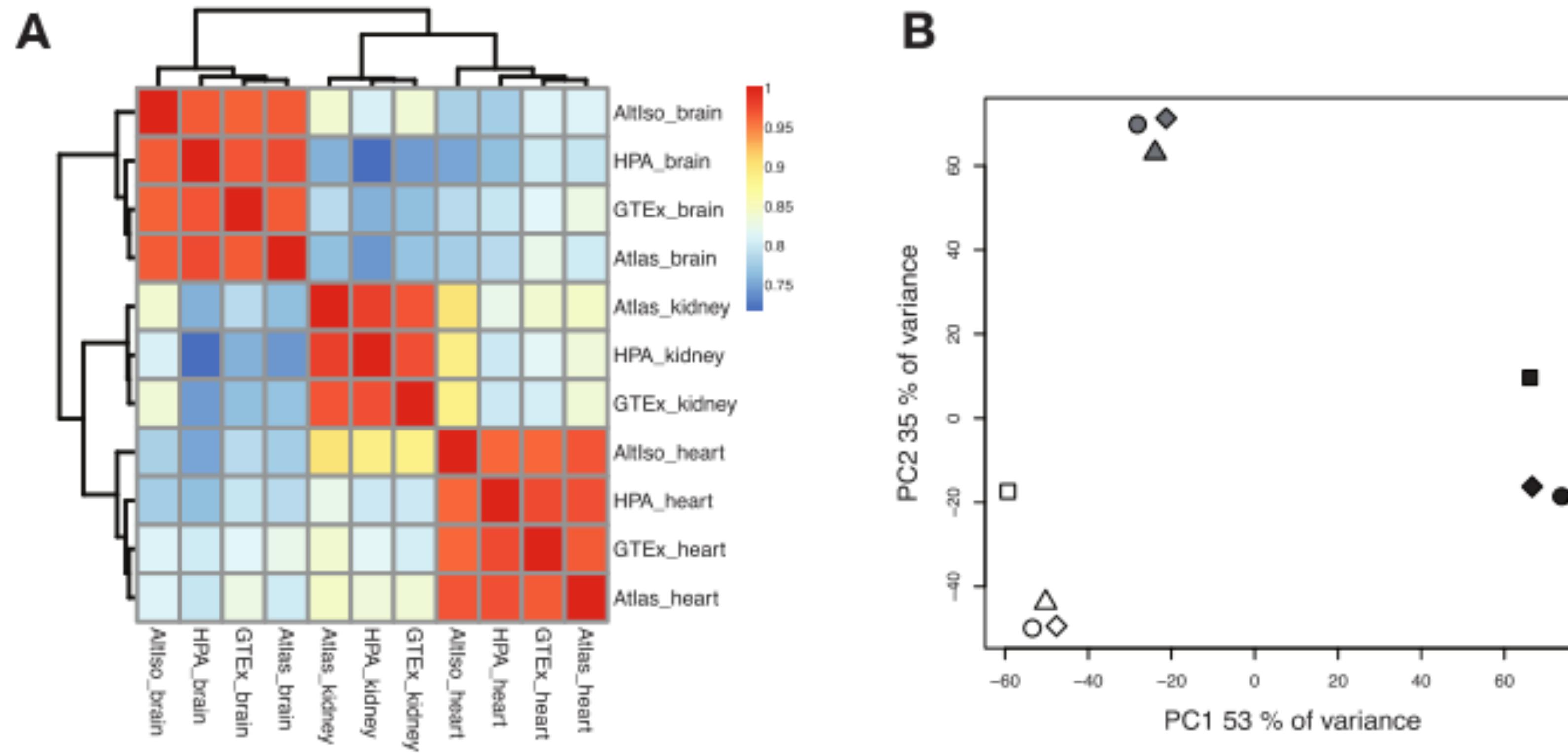
# Accounting for batch effects in practice

Public, processed RNA-seq data from 3 tissues, 4 studies show strong “study” (=batch) signal



# Accounting for batch effects in practice

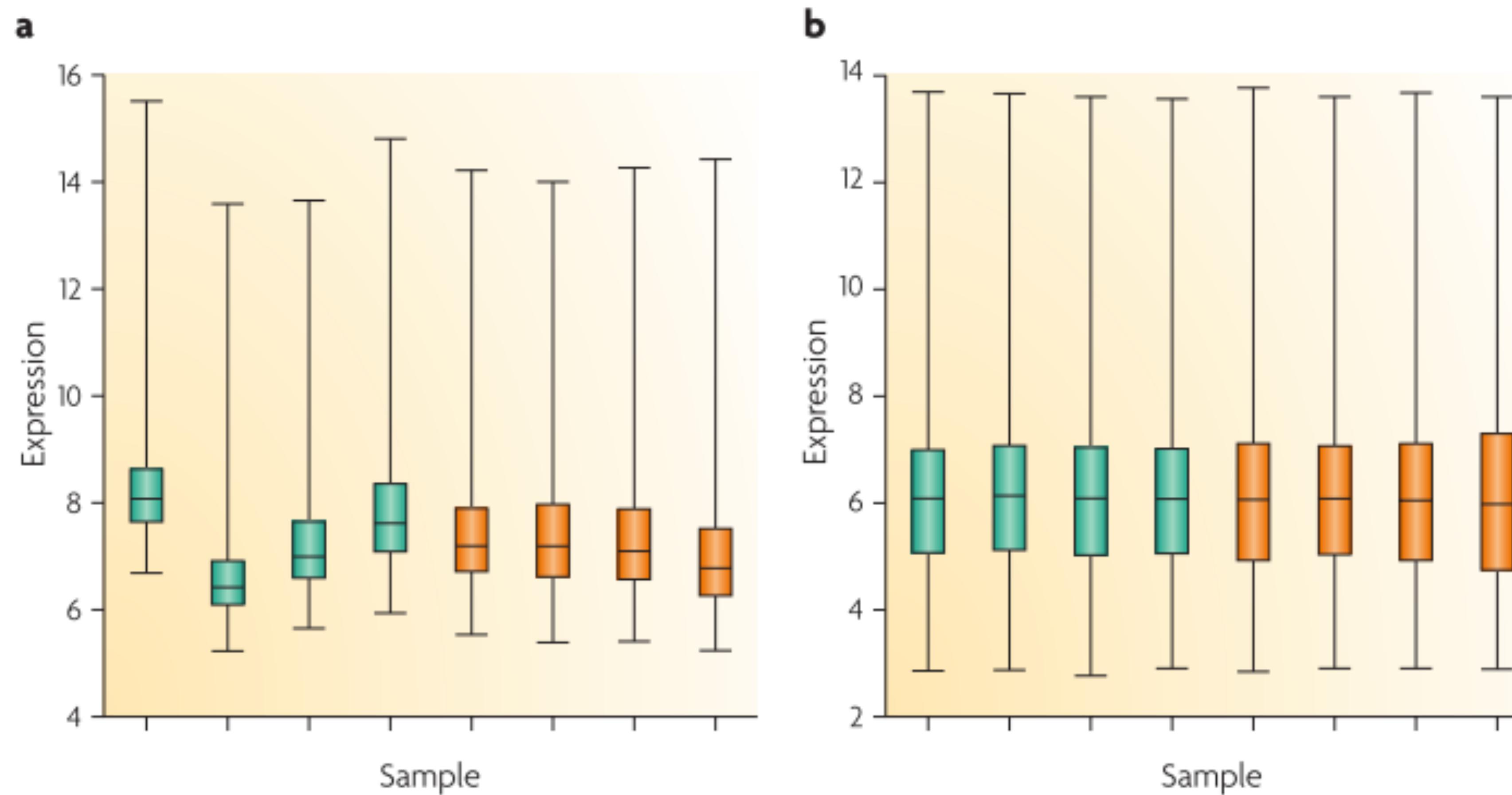
Accounting for the batch effect brings out the signal of interest



color = tissue; symbol = study (batch)

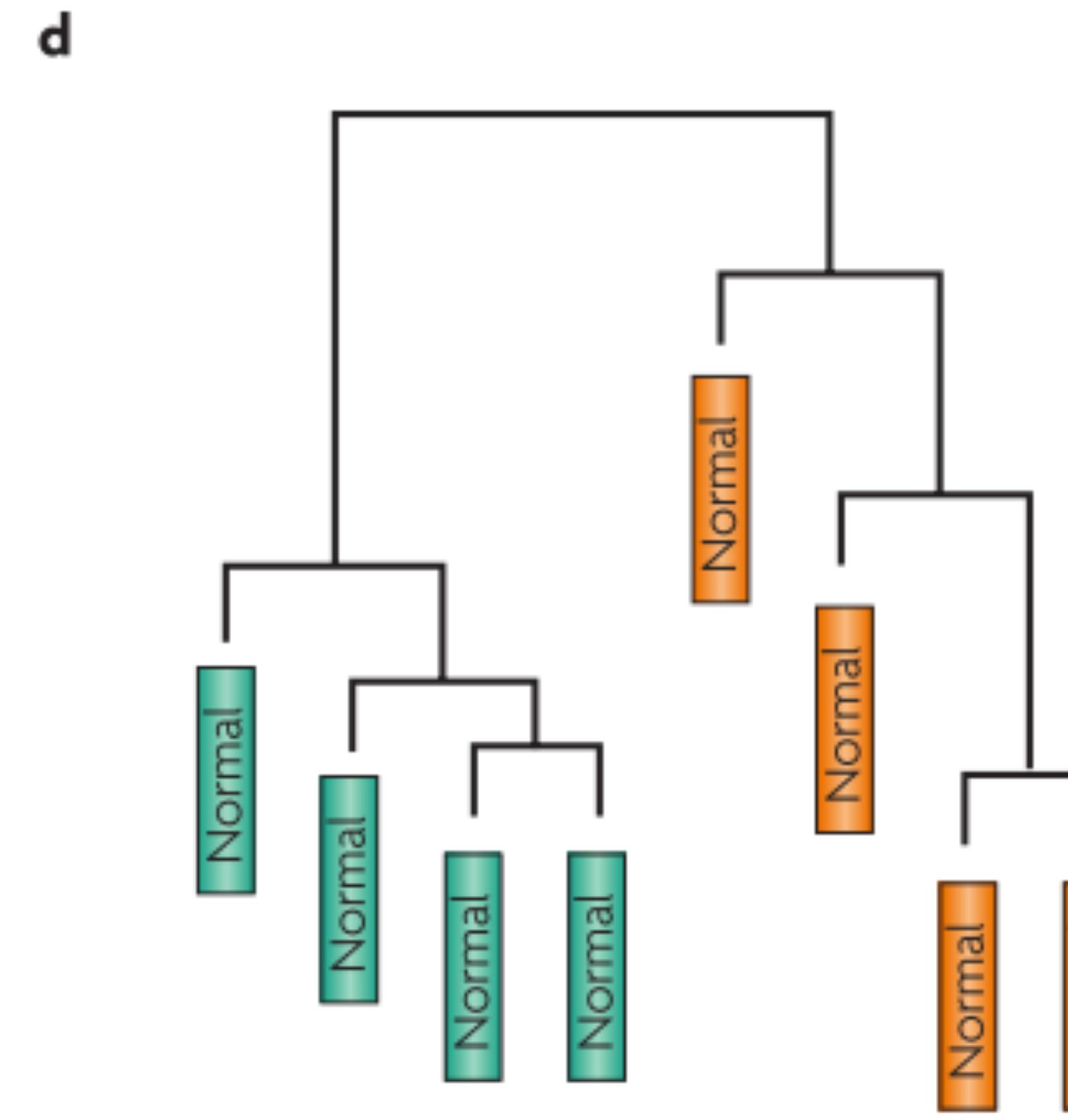
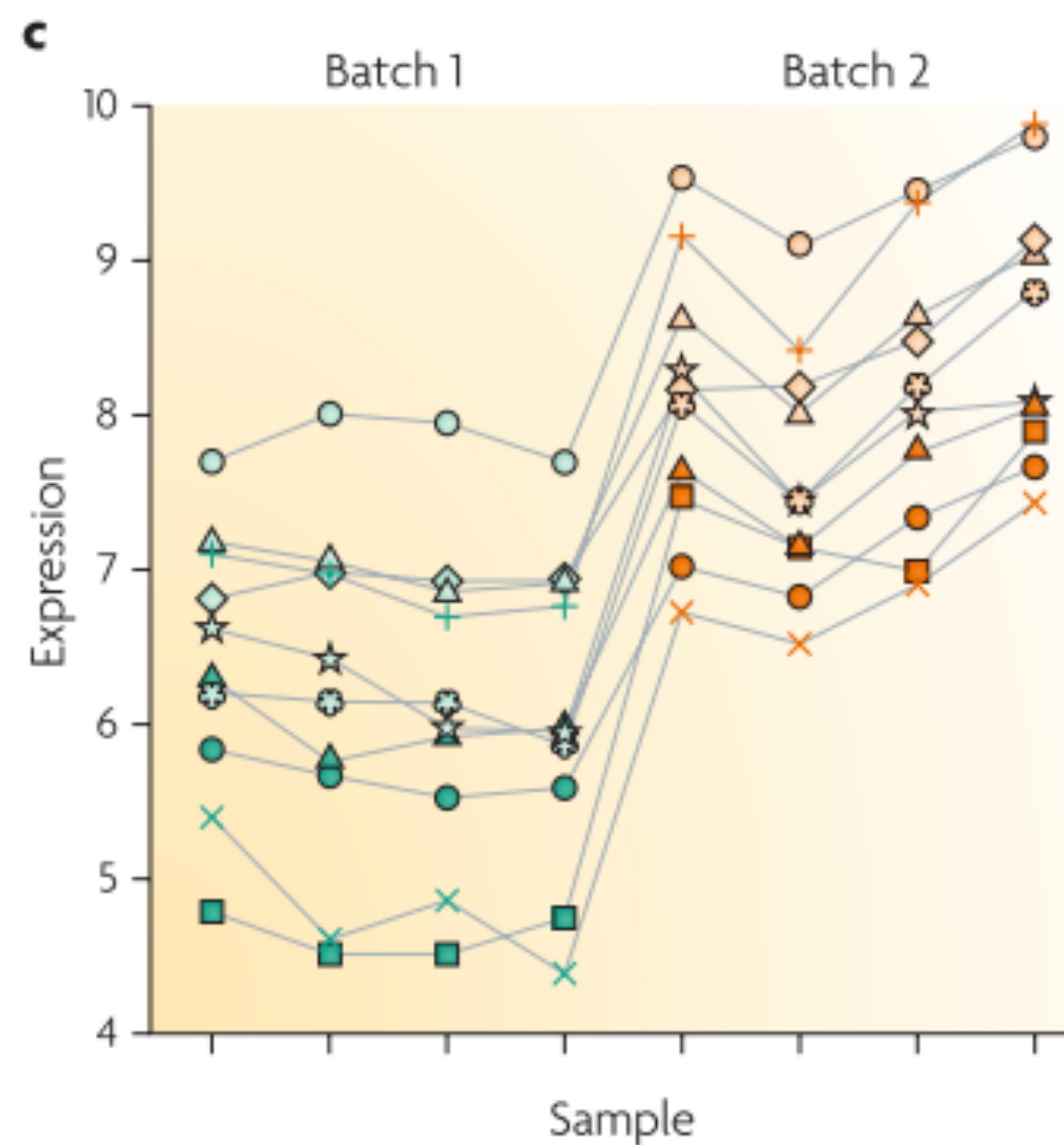
# Batch effect adjustment vs normalization

Batch effect adjustment goes *beyond* the “global” between-sample normalization methods



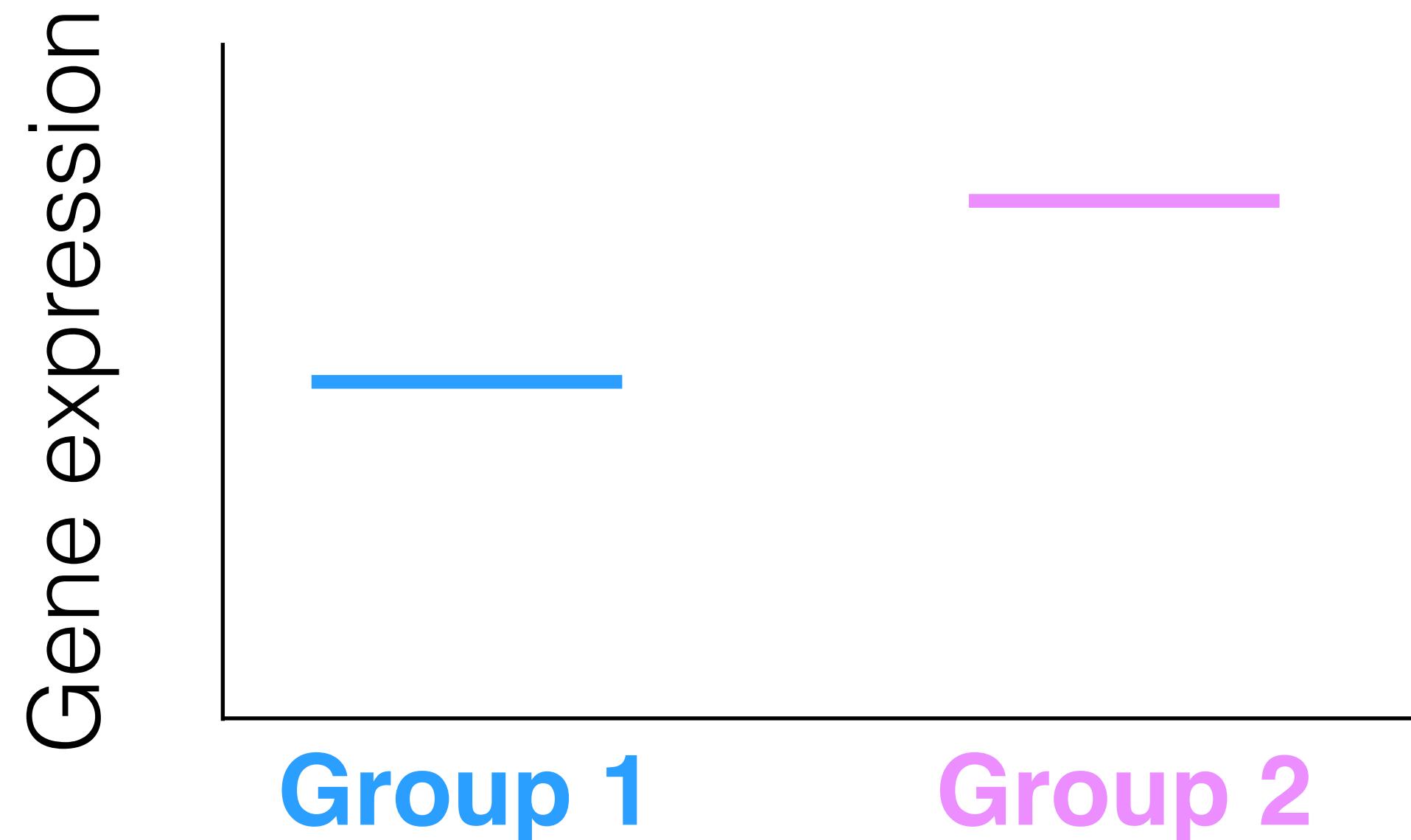
# Batch effect adjustment vs normalization

Batch effect adjustment goes beyond the “global” between-sample normalization methods



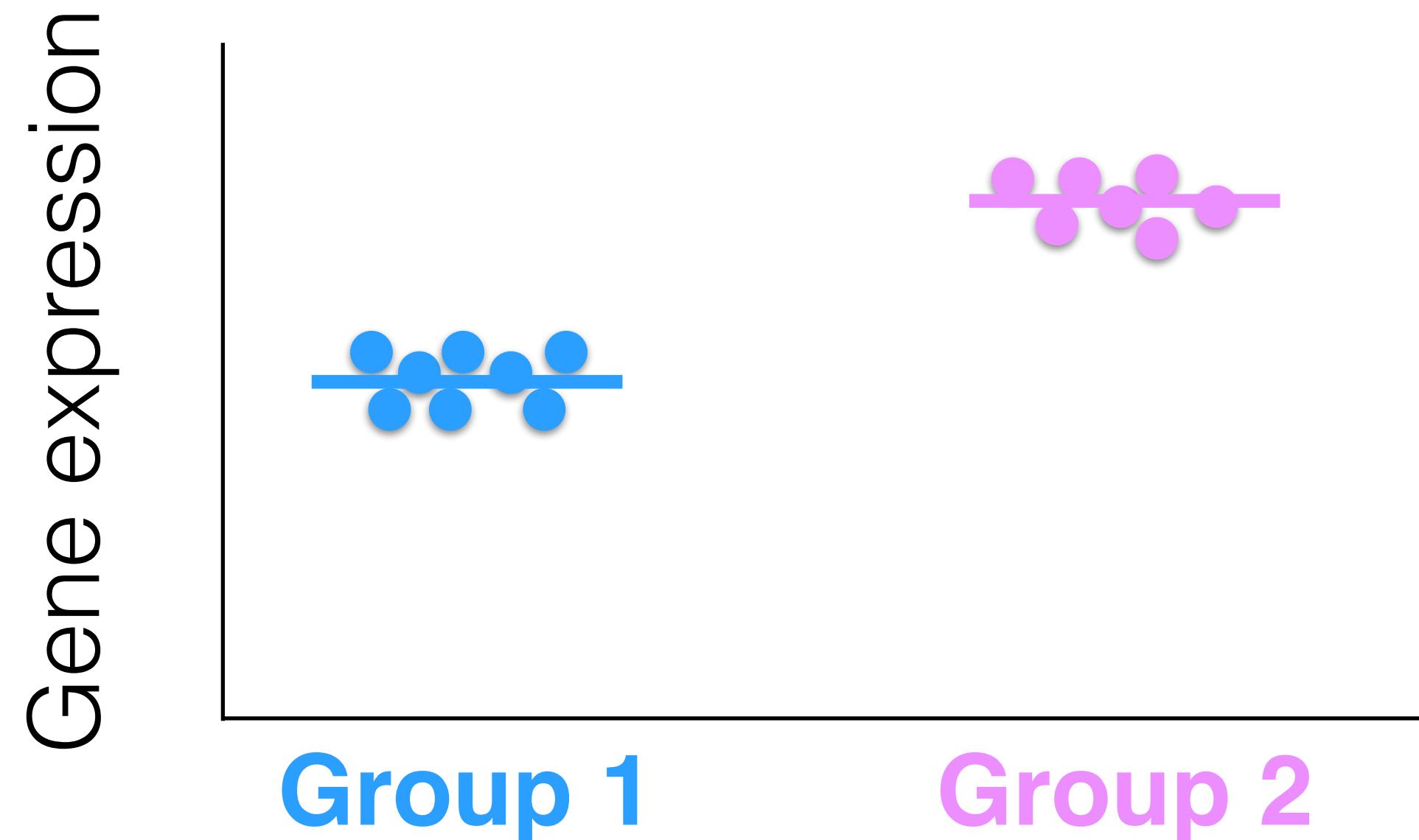
# Other design issues: replication

- Replicates are **necessary** to estimate within-condition variability.
- Variability estimates are, in turn, **vital** for statistical testing.



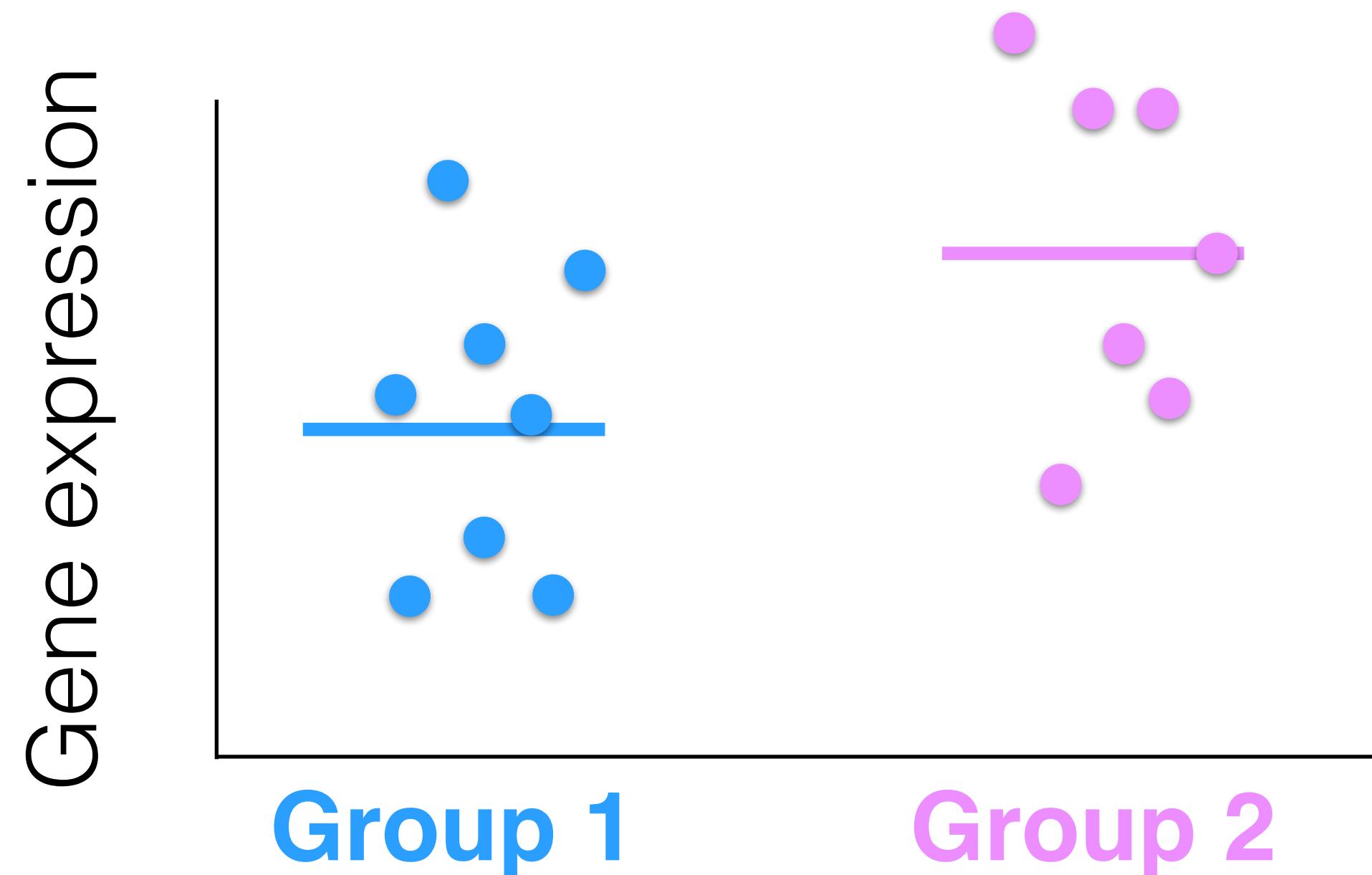
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# Other design issues: replication

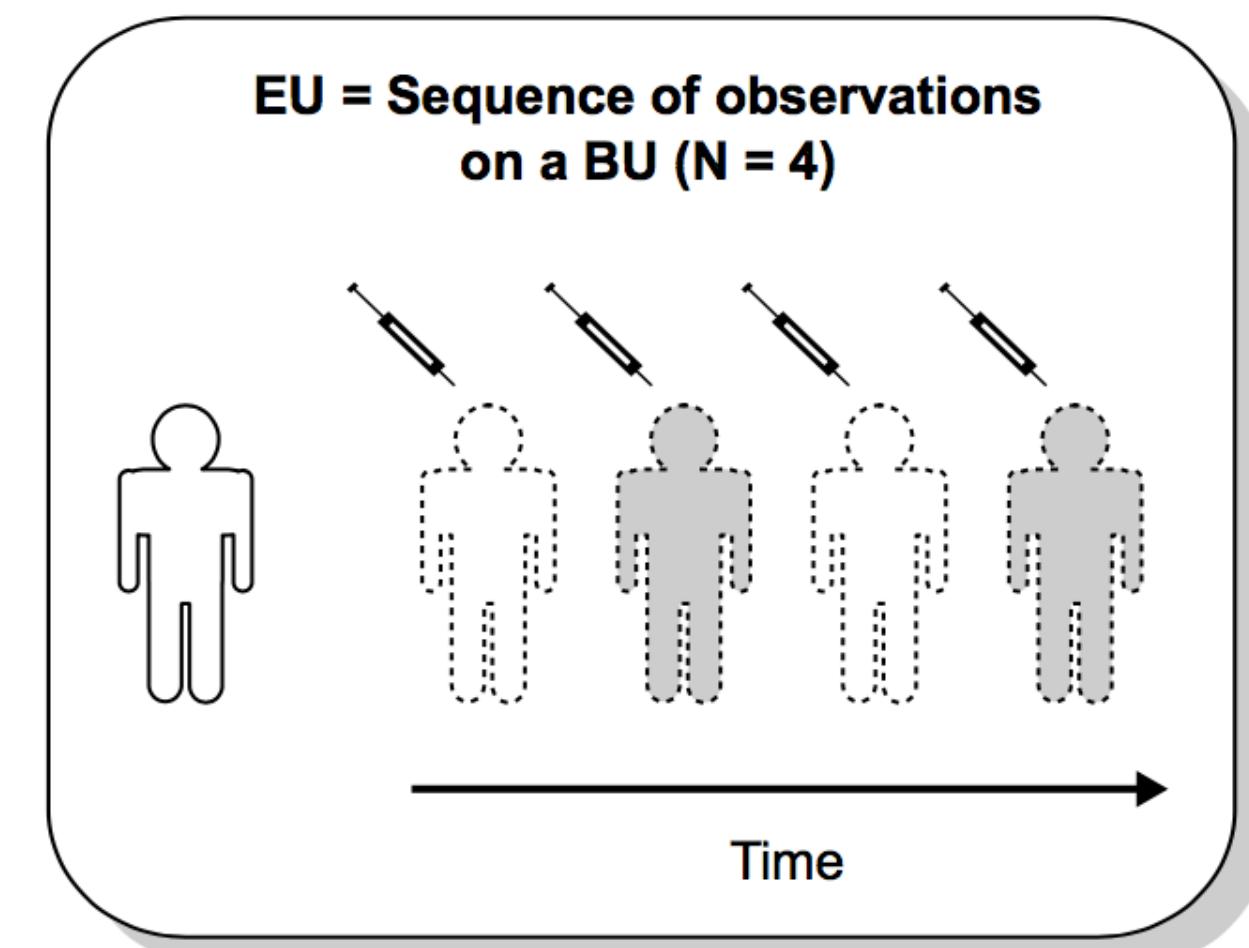
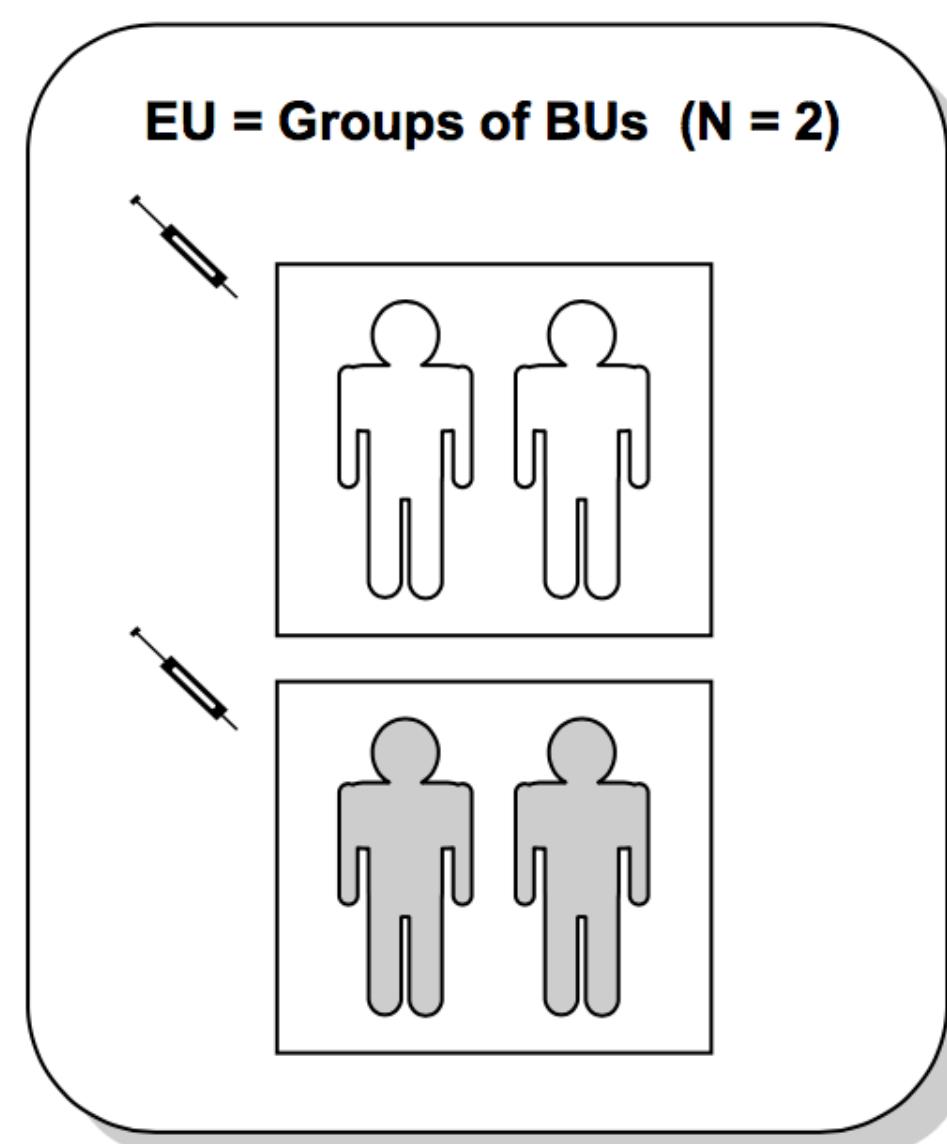
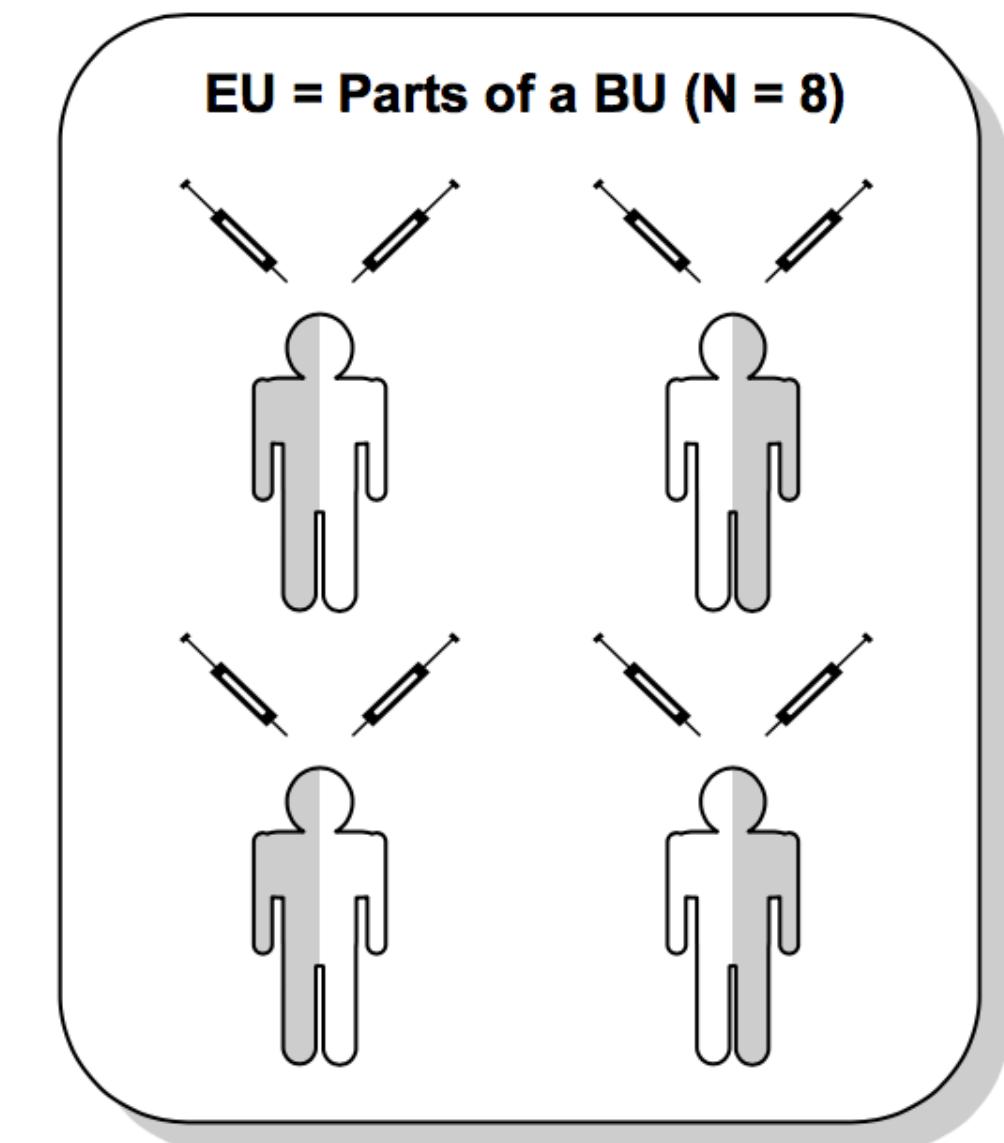
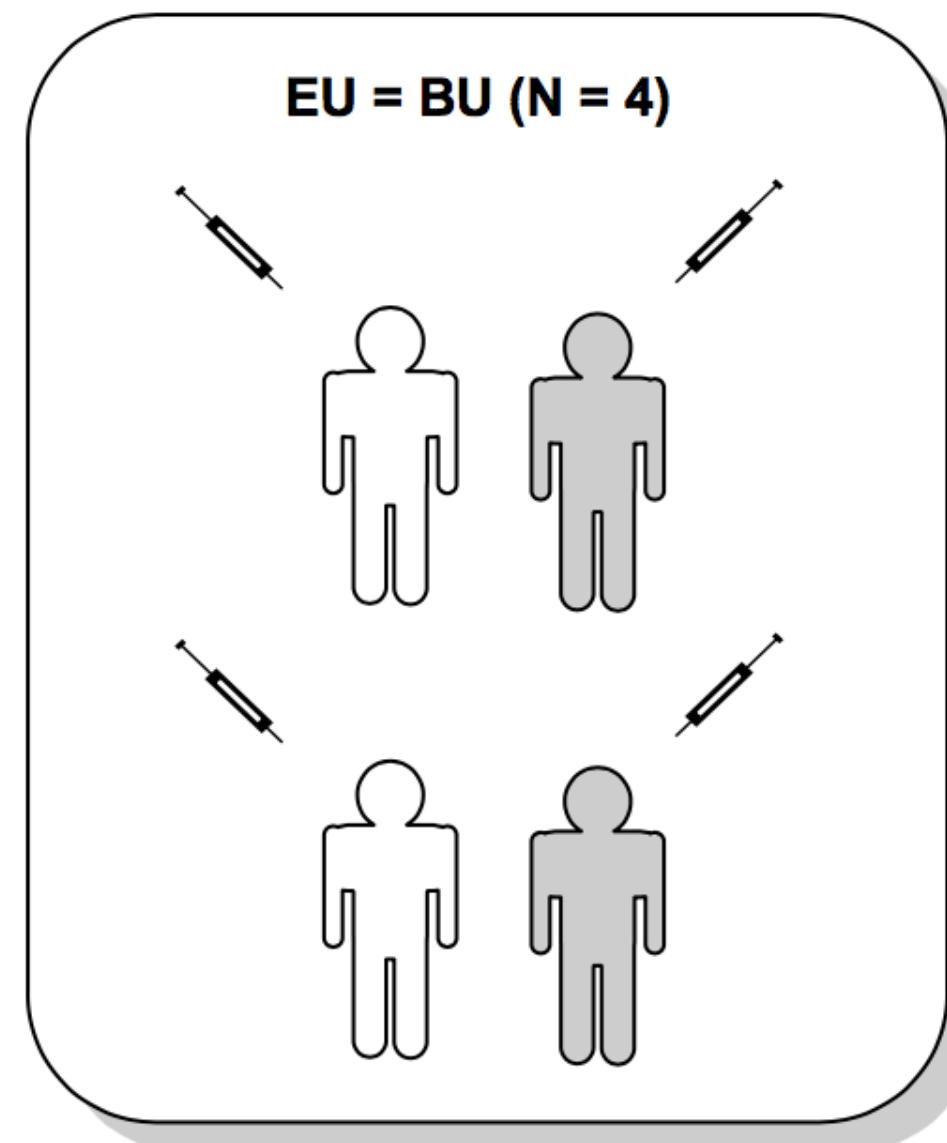
- Replicates are **necessary** to estimate within-condition variability.
- Variability estimates are, in turn, **vital** for statistical testing.



# Different types of units

- Biological units (BU) - entities we want to make inferences about (e.g., animal, person)
- Experimental units (EU) - smallest entities that can be independently assigned to a treatment (e.g., animal, litter, cage, well)
- Observational units (OU) - entities at which measurements are made

# Biological vs experimental units



# Pseudoreplication

- “**Artificial inflation** of the sample size, that usually occurs when the biological unit of interest differs from the experimental unit or observational unit.”
- Only replication of experimental units is true replication

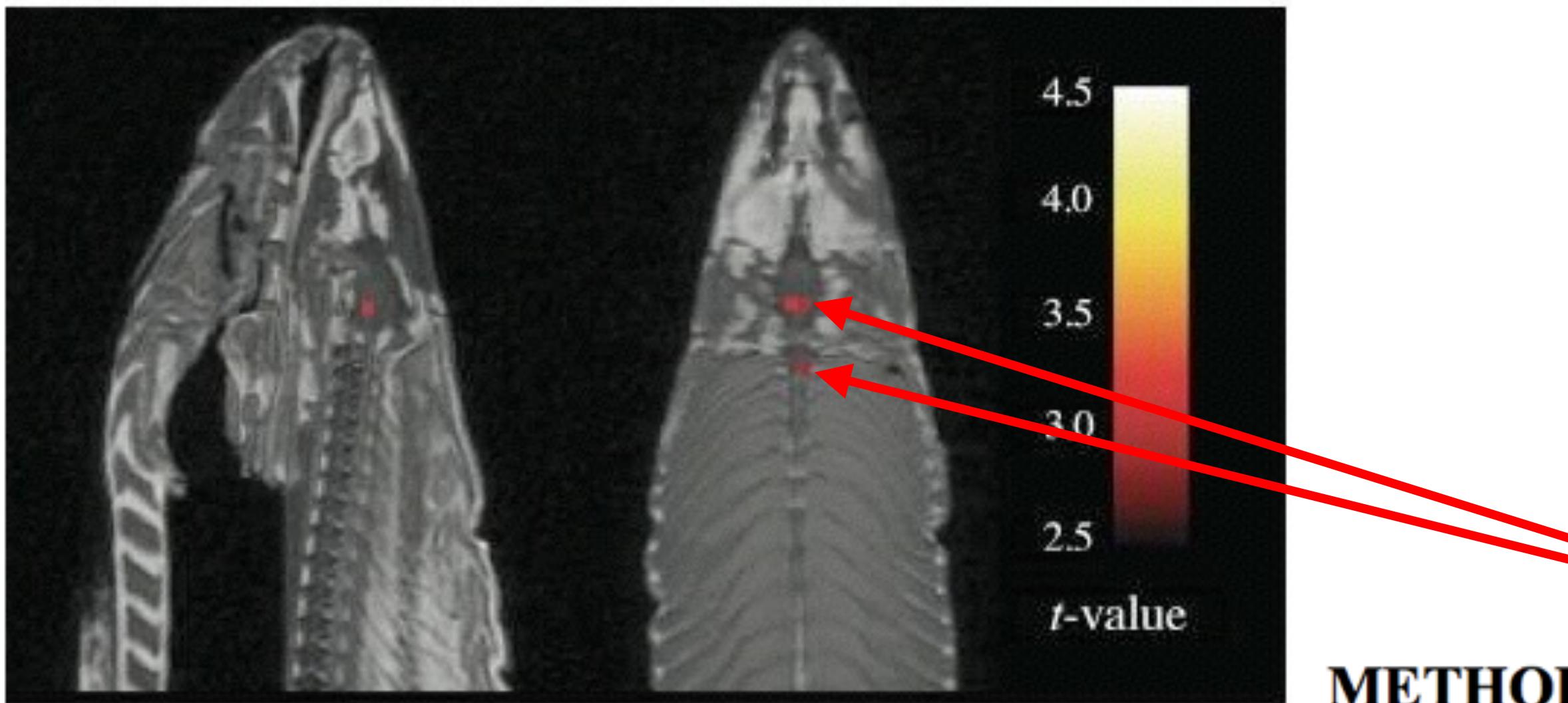
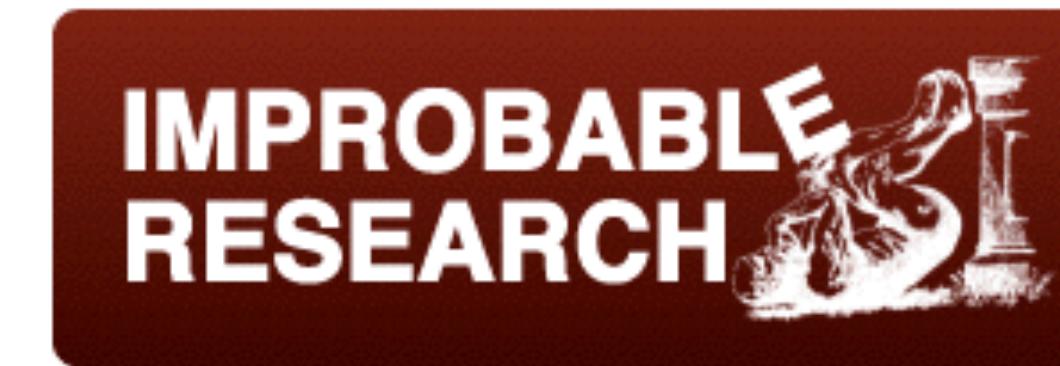
# What is a p-value?

- The p-value is the probability of obtaining a test statistic *at least as extreme* as the one observed, *if the null hypothesis is true* (i.e., if there is no true signal in the data)
- Hence, if we get a p-value of **0.05**, it means that there is a **5%** chance of getting that extreme results even in the absence of real signal!

# What does this mean for high-throughput studies?

- Assume that we perform 10,000 tests (one for each gene)...
- ... and that there is no true signal at all in the data
- Then we would expect to get around 500 p-values below 0.05
- Relying solely on p-values would be misleading!

**NEUROSCIENCE PRIZE:** Craig Bennett, Abigail Baird, Michael Miller, and George Wolford [USA], for demonstrating that brain researchers, by using complicated instruments and simple statistics, can see meaningful brain activity anywhere — even in a dead salmon.



## METHODS

**Subject.** One mature Atlantic Salmon (*Salmo salar*) participated in the fMRI study. The salmon was approximately 18 inches long, weighed 3.8 lbs, and was not alive at the time of scanning.

**Task.** The task administered to the salmon involved completing an open-ended mentalizing task. The salmon was shown a series of photographs depicting human individuals in social situations with a specified emotional valence. The salmon was asked to determine what emotion the individual in the photo must have been experiencing.

**Design.** Stimuli were presented in a block design with each photo presented for 10 seconds followed by 12 seconds of rest. A total of 15 photos were displayed. Total scan time was 5.5 minutes.

# We need to change perspective

- Instead of limiting the false positive probability for each *individual test*, try to limit
  - the probability of obtaining *any* false positives (FWER)
  - the fraction of false positives among the significant genes (FDR)

# Benjamini-Hochberg correction - controlling the FDR

- Assume we are performing  $N$  tests
- Intuition:
  - for each threshold  $\alpha$ , we can estimate the expected number of false discoveries by  $\alpha N$
  - Compare this to the actual number of discoveries at that threshold ( $N_\alpha$ )
  - Choose  $\alpha$  so that  $\alpha N / N_\alpha \leq 0.05$  (or another desired threshold)

# Interpreting the FDR

- The FDR is a measure for a set of genes
- In a set of genes with  $\text{FDR} = 0.05$ , approximately 5% can be expected to be false discoveries
- However, we don't know *which* ones! It could be the most significant!
- *q-values* are gene-wise significance measures ("adjusted p-values") - the smallest FDR we have to accept in order to call the gene significant

# Model formulas and design matrices

- Testing is done separately for each gene
- We must tell the packages **which model** to fit (e.g. which predictors to use)
- The design does *not* follow “automatically” from having the sample annotation table - many different designs are often possible
- Model formulas in R:

response variable ~ predictors

- Fit a separate model for each gene - response variable changes. Specify only predictors

## Examples

```
## Linear model, mtcars data  
lm(mpg ~ cyl, data = mtcars)
```

```
## Linear model (limma), gene expression data  
lmFit(object = y, design = model.matrix(~ group))
```

```
## GLM (edgeR), RNA-seq data  
fit <- glmFit(y = d, design = model.matrix(~ time))
```

```
## DESeq2, RNA-seq data  
dds <- DESeqDataSetFromMatrix(countData = countData,  
                                colData = DataFrame(condition),  
                                design = ~ condition)
```

# Testing and contrasts

- After fitting the model(s), we must decide *which* coefficient (or combination thereof) we want to apply a hypothesis test for.
- Combinations of coefficients are called *contrasts*.
- Design matrices can often be defined in many equivalent ways - important that the contrast is defined accordingly!

## Examples

```
## GLM (edgeR), RNA-seq data
glmLRT(fit, coef = 2)
glmLRT(fit, contrast = c(-1, 1))

## DESeq2, RNA-seq data
results(dds, contrast = c("condition", "B", "A"))
results(dds, contrast = c(0, -1, 1))
results(dds)
```

# Model formulas and design matrices

- A **design matrix** contains the values of the predictor variables for each sample

coefficients

$$\begin{pmatrix} y_1 \\ y_2 \\ y_3 \\ y_4 \\ y_5 \\ y_6 \end{pmatrix} = \boxed{\begin{pmatrix} 1 & 0 \\ 1 & 0 \\ 1 & 0 \\ 1 & 1 \\ 1 & 1 \\ 1 & 1 \end{pmatrix}} \begin{pmatrix} \beta_0 \\ \beta_1 \end{pmatrix} + \begin{pmatrix} \varepsilon_1 \\ \varepsilon_2 \\ \varepsilon_3 \\ \varepsilon_4 \\ \varepsilon_5 \\ \varepsilon_6 \end{pmatrix} = \boxed{X}\beta + \varepsilon$$

e.g.: (log) expression values for a given gene

$$y_i = \beta_0 + \beta_1 x_i + \varepsilon_i$$

# Many ways of modeling the same expected values

- 1 predictor, 2 groups

	group 1	group 2
X	b0	b0 + b1
	~X	

	group 1	group 2
X	b0	b1
	~0 + X	

the coefficients mean different things in the different cases!

- 2 predictors, 2\*2 groups

	Y	
X	b0	b0+b1
	b0+b2	b0+b1+ b2+b3

$$\begin{aligned} &\sim X^*Y \\ &\sim X + Y + X:Y \end{aligned}$$

	Y	
X	b0	b1
	b2	b3

New variable,  
combining X  
and Y

$$\sim 0 + XY$$

	Y	
X	b0	b0+b1
	b0+b2	b0+b2+ b3

$$\sim X + X:Y$$

# Model formulas and design matrices - example 1

## One predictor, two levels (without intercept)

### Sample table:

	sample	treatment
1	s1	control
2	s2	control
3	s3	control
4	s4	treated
5	s5	treated
6	s6	treated

### Design matrix:

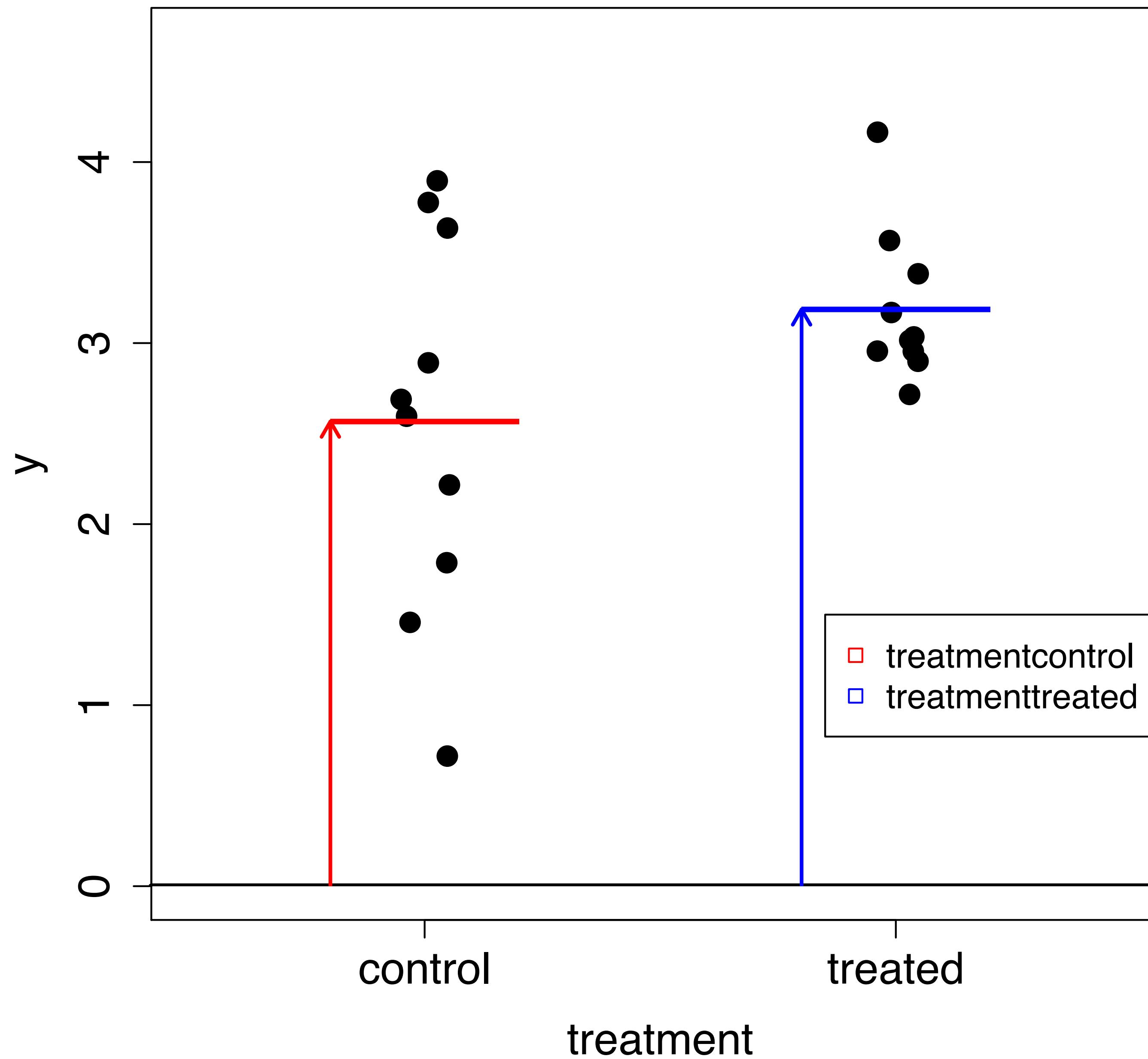
	<u>treatmentcontrol</u>	<u>treatmenttreated</u>
1	1	0
2	1	0
3	1	0
4	0	1
5	0	1
6	0	1

### Formula:

$\sim 0 + \text{treatment}$

### Modeled values:

	control	treated
<b>treatmentcontrol</b>		<b>treatmenttreated</b>



# Model formulas and design matrices - example 1

One predictor, two levels (with intercept)

## Sample table:

	sample	treatment
1	s1	control
2	s2	control
3	s3	control
4	s4	treated
5	s5	treated
6	s6	treated

## Design matrix:

	(Intercept)	treatmenttreated
1	1	0
2	1	0
3	1	0
4	1	1
5	1	1
6	1	1

## Formula:

$\sim$  treatment

## Modeled values:

control	treated
$1 * \text{Intercept} + 0 * \text{treatmenttreated}$	$1 * \text{Intercept} + 1 * \text{treatmenttreated}$

# Model formulas and design matrices - example 1

One predictor, two levels (with intercept)

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	sample	treatment
1	s1	control
2	s2	control
3	s3	control
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5	s5	treated
6	s6	treated

## Design matrix:

	(Intercept)	treatmenttreated
1	1	0
2	1	0
3	1	0
4	1	1
5	1	1
6	1	1

## Formula:

$\sim$  treatment

## Modeled values:

	control	treated
	$1 * \text{Intercept} + 0 * \text{treatmenttreated}$	$1 * \text{Intercept} + 1 * \text{treatmenttreated}$

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1	s1	control
2	s2	control
3	s3	control
4	s4	treated
5	s5	treated
6	s6	treated

## Design matrix:

	(Intercept)	treatmenttreated
1	1	0
2	1	0
3	1	0
4	1	1
5	1	1
6	1	1

## Formula:

$\sim$  treatment

## Modeled values:

	control	treated
	$1 * \text{Intercept} + 0 * \text{treatmenttreated}$	$1 * \text{Intercept} + 1 * \text{treatmenttreated}$

# Model formulas and design matrices - example 1

One predictor, two levels (with intercept)

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	sample	treatment
1	s1	control
2	s2	control
3	s3	control
4	s4	treated
5	s5	treated
6	s6	treated

## Design matrix:

	(Intercept)	treatmenttreated
1	1	0
2	1	0
3	1	0
4	1	1
5	1	1
6	1	1

## Formula:

$\sim$  treatment

## Modeled values:

	control	treated
	$1 * \text{Intercept} + 0 * \text{treatmenttreated}$	$1 * \text{Intercept} + 1 * \text{treatmenttreated}$

# Model formulas and design matrices - example 1

One predictor, two levels (with intercept)

## Sample table:

	sample	treatment
1	s1	control
2	s2	control
3	s3	control
4	s4	treated
5	s5	treated
6	s6	treated

## Design matrix:

	(Intercept)	treatmenttreated
1	1	0
2	1	0
3	1	0
4	1	1
5	1	1
6	1	1

## Formula:

$\sim$  treatment

## Modeled values:

	control	treated
	$1 * \text{Intercept} + 0 * \text{treatmenttreated}$	$1 * \text{Intercept} + 1 * \text{treatmenttreated}$

# Model formulas and design matrices - example 1

One predictor, two levels (with intercept)

## Sample table:

	sample	treatment
1	s1	control
2	s2	control
3	s3	control
4	s4	treated
5	s5	treated
6	s6	treated

## Design matrix:

	(Intercept)	treatmenttreated
1	1	0
2	1	0
3	1	0
4	1	1
5	1	1
6	1	1

## Formula:

$\sim$  treatment

## Modeled values:

	control	treated
	$1 * \text{Intercept} + 0 * \text{treatmenttreated}$	$1 * \text{Intercept} + 1 * \text{treatmenttreated}$

# Model formulas and design matrices - example 1

One predictor, two levels (with intercept)

## Sample table:

	sample	treatment
1	s1	control
2	s2	control
3	s3	control
4	s4	treated
5	s5	treated
6	s6	treated

## Design matrix:

	(Intercept)	treatmenttreated
1	1	0
2	1	0
3	1	0
4	1	1
5	1	1
6	1	1

## Formula:

$\sim$  treatment

## Modeled values:

	control	treated
	$1 * \text{Intercept} + 0 * \text{treatmenttreated}$	$1 * \text{Intercept} + 1 * \text{treatmenttreated}$

# Model formulas and design matrices - example 1

One predictor, two levels (with intercept)

## Sample table:

	sample	treatment
1	s1	control
2	s2	control
3	s3	control
4	s4	treated
5	s5	treated
6	s6	treated

## Design matrix:

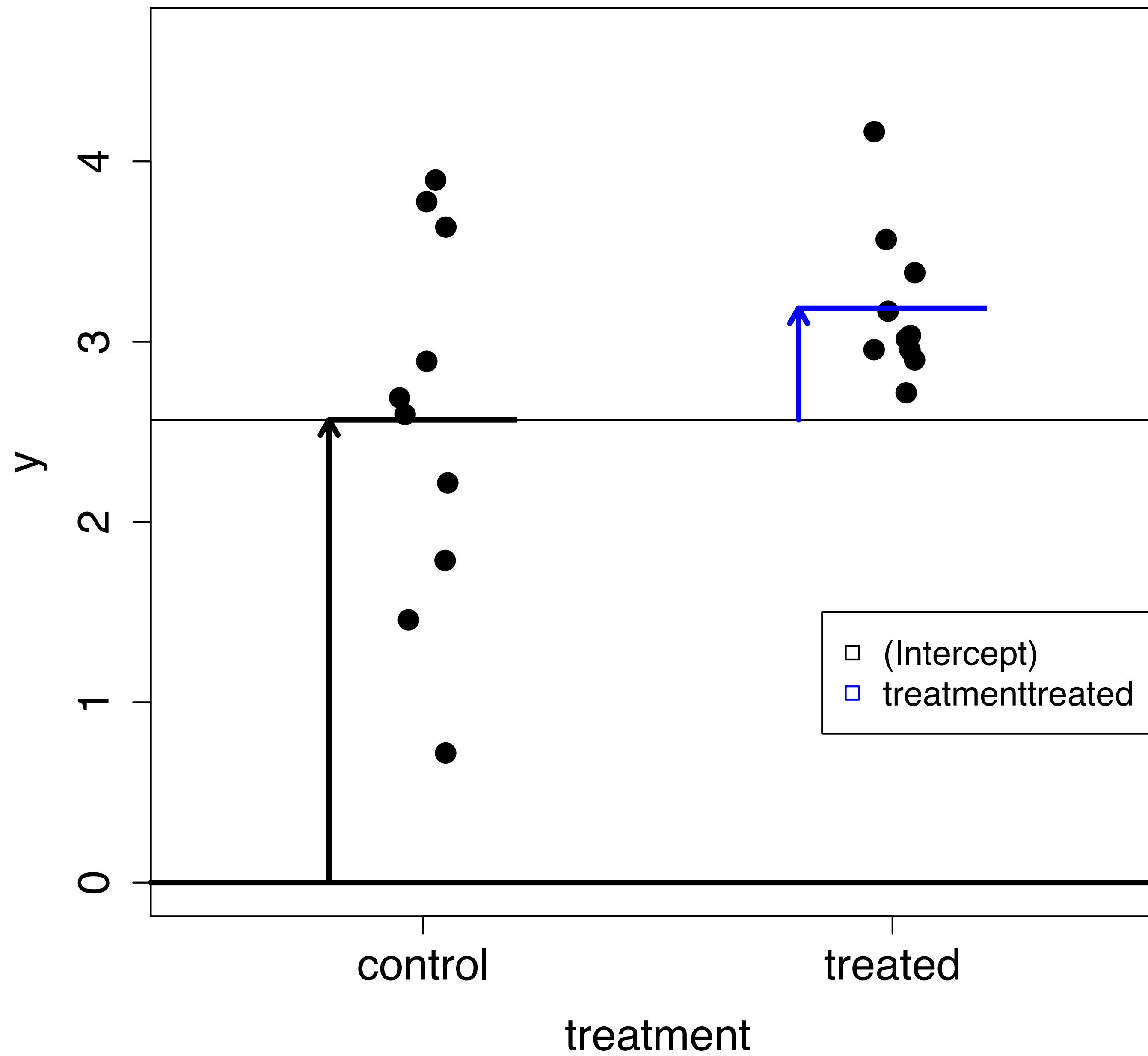
	(Intercept)	treatmenttreated
1	1	0
2	1	0
3	1	0
4	1	1
5	1	1
6	1	1

## Formula:

$\sim$  treatment

## Modeled values:

	control	treated
	Intercept	Intercept + treatmenttreated



# Model formulas and design matrices - example 2

One continuous predictor

## Sample table:

	sample	age
1	s1	21
2	s2	12
3	s3	64
4	s4	44
5	s5	19
6	s6	26

## Design matrix:

	(Intercept)	age
1	1	21
2	1	12
3	1	64
4	1	44
5	1	19
6	1	26

## Formula:

$\sim \text{age}$

## Modeled values:

	s1	s2	s3	s4	s5	s6
	Intercept + 21 * age	Intercept + 12 * age	Intercept + 64 * age	Intercept + 44 * age	Intercept + 19 * age	Intercept + 26 * age

# Model formulas and design matrices - example 3

One predictor, three levels

## Sample table:

	sample	treatment
1	s1	control
2	s2	control
3	s3	treatA
4	s4	treatA
5	s5	treatB
6	s6	treatB

## Design matrix:

	(Intercept)	treatmenttreatA	treatmenttreatB
1	1	1	0
2	2	1	0
3	3	1	1
4	4	1	1
5	5	1	0
6	6	1	0

## Formula:

$\sim$  treatment

## Modeled values:

	control	treatA	treatB
Intercept	Intercept	Intercept + treatmenttreatA	Intercept + treatmenttreatB

# Model formulas and design matrices - example 4

## One predictor, paired data (or two predictors)

### Sample table:

	sample	treatment
1	s1	control
2	s1	treated
3	s2	control
4	s2	treated
5	s3	control
6	s3	treated

### Design matrix:

	(Intercept)	samples2	samples3	treatmenttreated
1	1	1	0	0
2	2	1	0	1
3	3	1	1	0
4	4	1	1	1
5	5	1	0	1
6	6	1	0	1

### Formula:

$\sim \text{sample} + \text{treatment}$

### Modeled values:

	s1	s2	s3
control	Intercept	Intercept + samples2	Intercept + samples3
treated	Intercept + treatmenttreated	Intercept + samples2 + treatmenttreated	Intercept + samples3 + treatmenttreated

# Model formulas and design matrices - example 4

## One predictor, paired data (or two predictors)

### Sample table:

	genotype	treatment
1	A	control
2	A	control
3	A	treated
4	A	treated
5	B	control
6	B	control
7	B	treated
8	B	treated

### Design matrix:

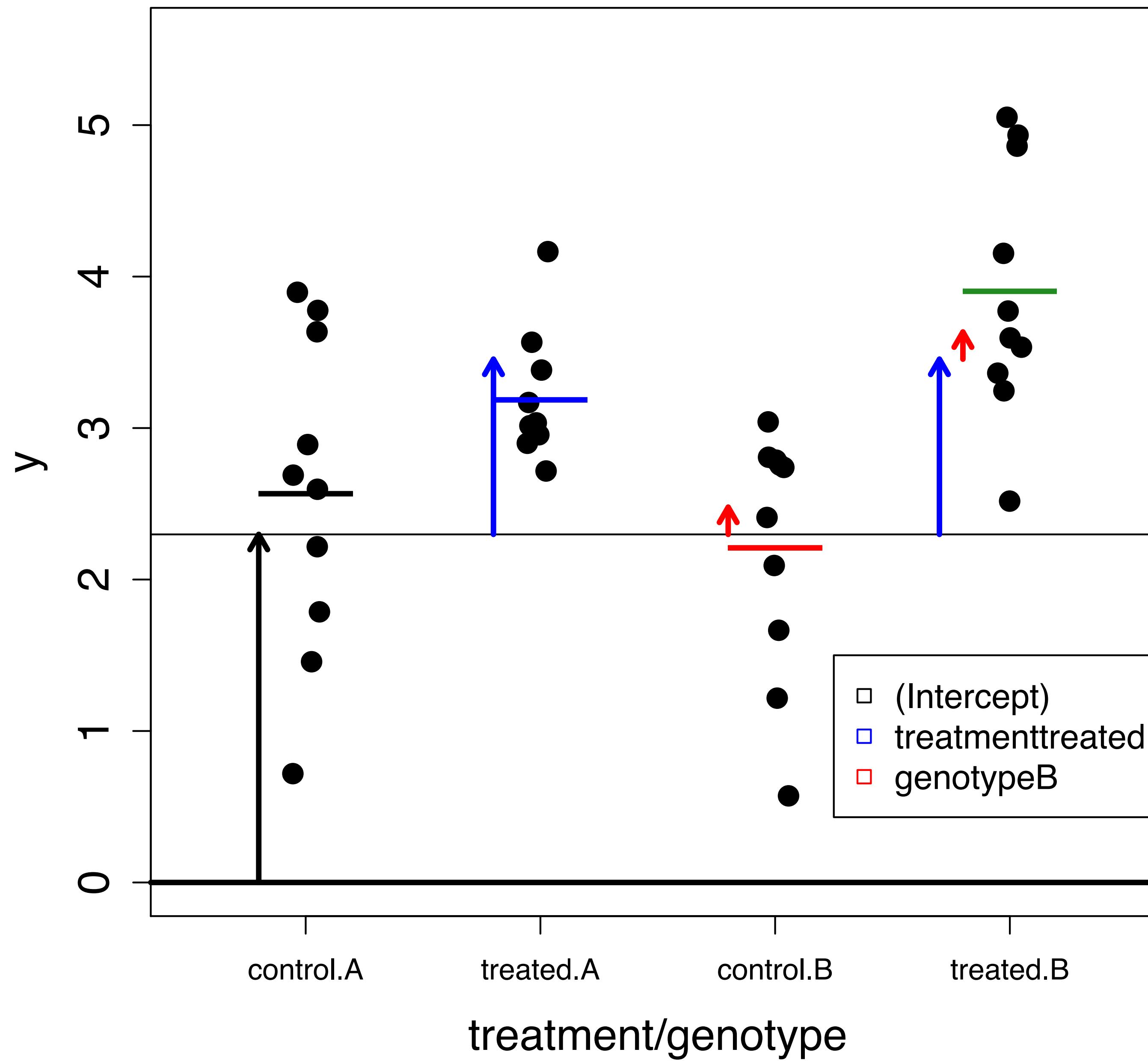
	(Intercept)	genotypeB	treatmenttreated
1	1	0	0
2	2	0	0
3	3	0	1
4	4	0	1
5	5	1	0
6	6	1	0
7	7	1	1
8	8	1	1

### Formula:

$\sim \text{genotype} + \text{treatment}$

### Modeled values:

		genotype A	genotype B
		Intercept	Intercept + genotypeB
		Intercept + treatmenttreated	Intercept + genotypeB + treatmenttreated
control	genotype A	Intercept	Intercept + genotypeB
	genotype B	Intercept + treatmenttreated	Intercept + genotypeB + treatmenttreated
treated	genotype A	Intercept + treatmenttreated	Intercept + genotypeB + treatmenttreated
treated	genotype B	Intercept + treatmenttreated	Intercept + genotypeB + treatmenttreated



# Model formulas and design matrices - example 5

Two predictors, with interaction

## Sample table:

	genotype	treatment
1	A	control
2	A	control
3	A	treated
4	A	treated
5	B	control
6	B	control
7	B	treated
8	B	treated

## Design matrix:

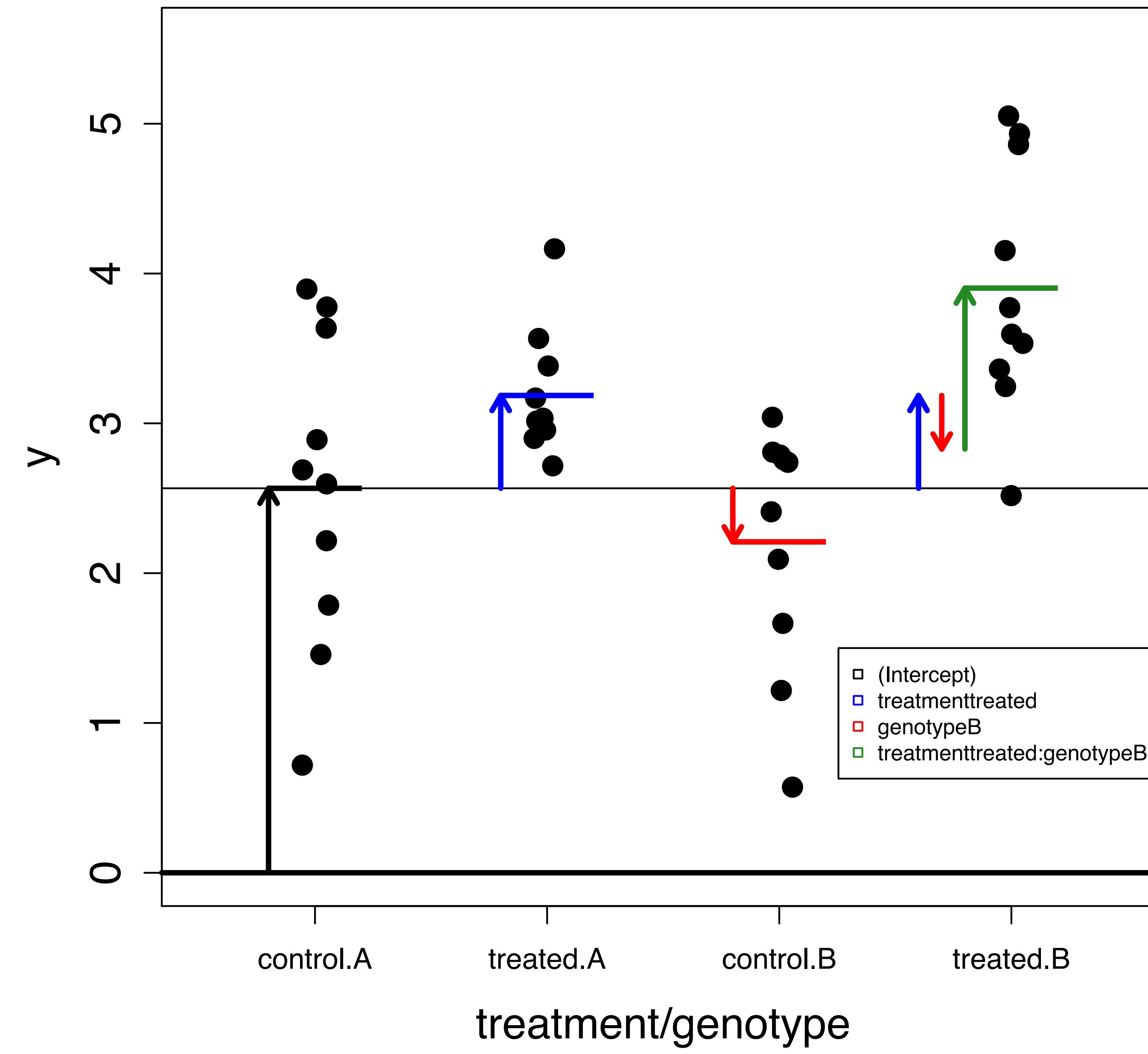
	(Intercept)	genotypeB	treatmenttreated	genotypeB:treatmenttreated
1	1	0	0	0
2	2	0	0	0
3	3	0	1	0
4	4	0	1	0
5	5	1	0	0
6	6	1	1	0
7	7	1	1	1
8	8	1	1	1

## Formula:

~ genotype \* treatment  
~ genotype + treatment + genotype:treatment

## Modeled values:

		genotype A	genotype B
		control	treated
control	control	Intercept	Intercept + genotypeB
	treated	Intercept + treatmenttreated	Intercept + genotypeB + treatmenttreated + genotypeB:treatmenttreated



# Model formulas and design matrices - example 6

Two predictors, with interaction

## Sample table:

```
treat.gt  
1 control.A  
2 control.A  
3 treated.A  
4 treated.A  
5 control.B  
6 control.B  
7 treated.B  
8 treated.B
```

## Design matrix:

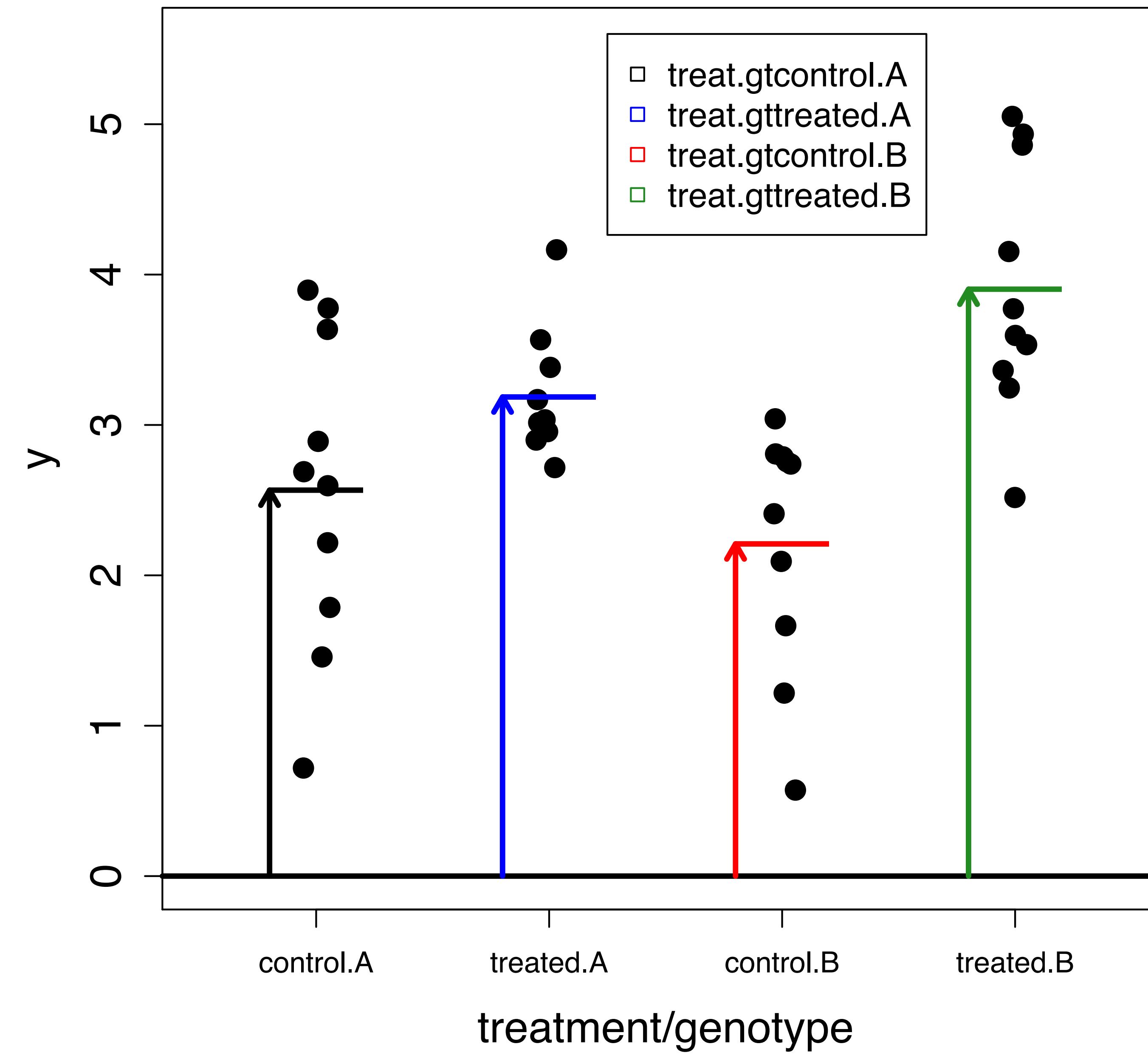
	treat.gtcontrol.A	treat.gttreated.A	treat.gtcontrol.B	treat.gttreated.B
1 control.A	1	0	0	0
2 control.A	1	0	0	0
3 treated.A	1	0	0	0
4 treated.A	0	1	0	0
5 control.B	0	1	0	0
6 control.B	0	0	1	0
7 treated.B	0	0	1	0
8 treated.B	0	0	0	1

## Formula:

$\sim 0 + \text{treat.gt}$

## Modeled values:

		genotype A	genotype B
		treat.gtcontrol.A	treat.gtcontrol.B
control	treat	treat.gttreated.A	treat.gttreated.B
	treated	treat.gttreated.A	treat.gttreated.B



# References

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