

# Comparison of Classification and Clustering Algorithms on PimaIndiansDiabetes Dataset Using R

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```
library(caret)

## Loading required package: lattice
## Loading required package: ggplot2
library(mclust)

## Package 'mclust' version 5.4.1
## Type 'citation("mclust")' for citing this R package in publications.
library(fpc)
library(cluster)
library(clusteval)
library(factoextra)

## Welcome! Related Books: `Practical Guide To Cluster Analysis in R` at https://goo.gl/13EFCZ
library(ggplot2)
library(kmed)
library(mlbench)
```

## Loading Pima Indians Diabetes Dataset

```
# attach the Pima Indians Diabetes Database to the environment
data("PimaIndiansDiabetes")
# rename the dataset
dataset <- PimaIndiansDiabetes
```

## Partitioning Data for Validation

```
# create a list of 80% of the rows in the original dataset we can use for training
validation_index <- createDataPartition(dataset$diabetes, p=0.80, list=FALSE)
# select 20% of the data for validation
validation <- dataset[-validation_index,]
# use the remaining 80% of data to training and testing the models
dataset <- dataset[validation_index,]
```

## Getting Insights from Data

```
# dimensions of dataset
dim(dataset)

## [1] 615  9
```

```

# list types for each attribute
sapply(dataset, class)

## pregnant glucose pressure triceps insulin mass pedigree
## "numeric" "numeric" "numeric" "numeric" "numeric" "numeric" "numeric"
## age diabetes
## "numeric" "factor"

# take a peek at the first 6 rows of the data
head(dataset)

## pregnant glucose pressure triceps insulin mass pedigree age diabetes
## 1 6 148 72 35 0 33.6 0.627 50 pos
## 2 1 85 66 29 0 26.6 0.351 31 neg
## 3 8 183 64 0 0 23.3 0.672 32 pos
## 4 1 89 66 23 94 28.1 0.167 21 neg
## 5 0 137 40 35 168 43.1 2.288 33 pos
## 6 5 116 74 0 0 25.6 0.201 30 neg

# list the levels for the class
levels(dataset$diabetes)

## [1] "neg" "pos"

# summarize the class distribution
percentage <- prop.table(table(dataset$diabetes)) * 100
cbind(freq=table(dataset$diabetes), percentage=percentage)

## freq percentage
## neg 400 65.04065
## pos 215 34.95935

# summarize attribute distributions
summary(dataset)

## pregnant glucose pressure triceps
## Min. : 0.000 Min. : 0.0 Min. : 0.00 Min. : 0.00
## 1st Qu.: 1.000 1st Qu.:100.0 1st Qu.: 62.00 1st Qu.: 0.00
## Median : 3.000 Median :117.0 Median : 72.00 Median :23.00
## Mean : 3.979 Mean :121.0 Mean : 68.66 Mean :20.27
## 3rd Qu.: 6.000 3rd Qu.:140.5 3rd Qu.: 80.00 3rd Qu.:32.00
## Max. :17.000 Max. :199.0 Max. :114.00 Max. :99.00
## insulin mass pedigree age
## Min. : 0.00 Min. : 0.00 Min. :0.0780 Min. :21.00
## 1st Qu.: 0.00 1st Qu.:27.15 1st Qu.:0.2415 1st Qu.:24.00
## Median : 0.00 Median :31.90 Median :0.3780 Median :29.00
## Mean : 77.91 Mean :31.72 Mean :0.4727 Mean :33.71
## 3rd Qu.:127.50 3rd Qu.:36.30 3rd Qu.:0.6340 3rd Qu.:41.00
## Max. :744.00 Max. :59.40 Max. :2.4200 Max. :81.00
## diabetes
## neg:400
## pos:215
##
##
##
##

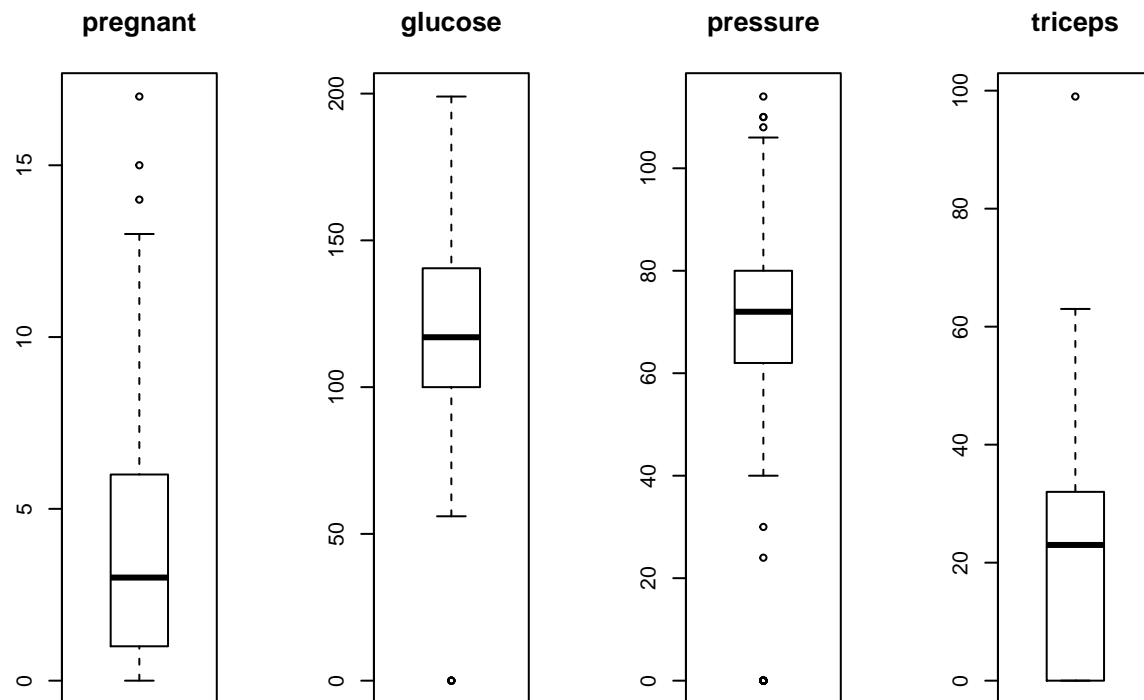
```

```

# split input and output
x <- dataset[,1:8]
y <- dataset[,9]

# boxplot for each attribute on one image
par(mfrow=c(1,4))
for(i in 1:4) {
  boxplot(x[,i], main=names(PimaIndiansDiabetes)[i])
}

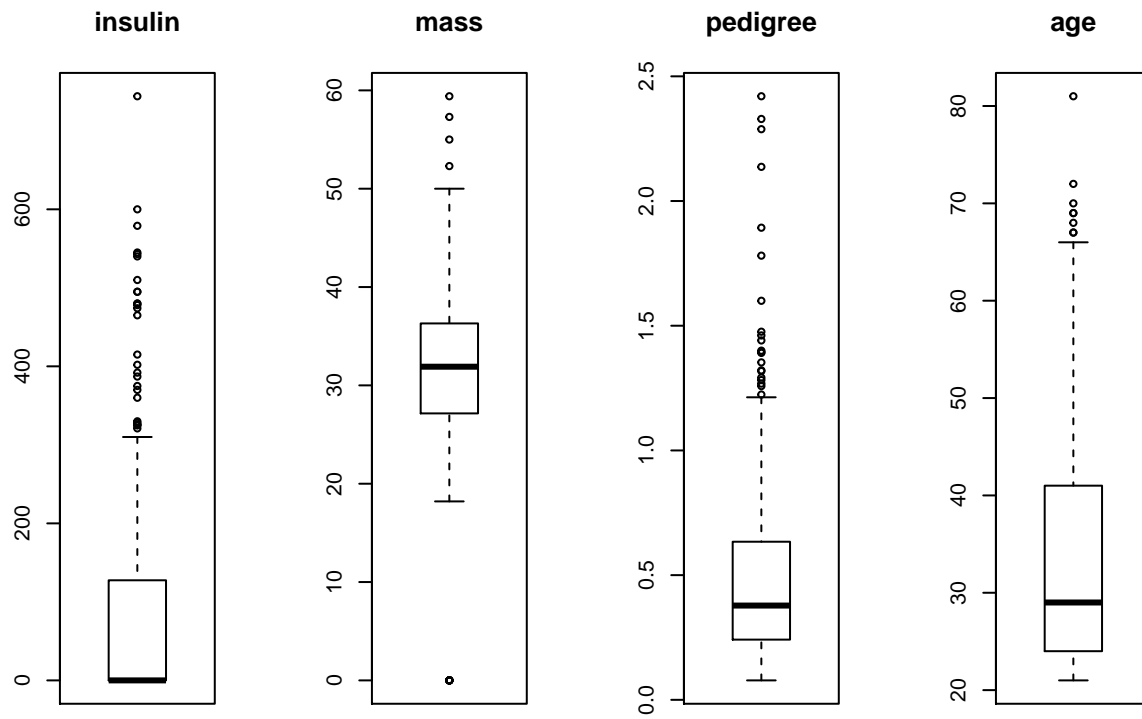
```



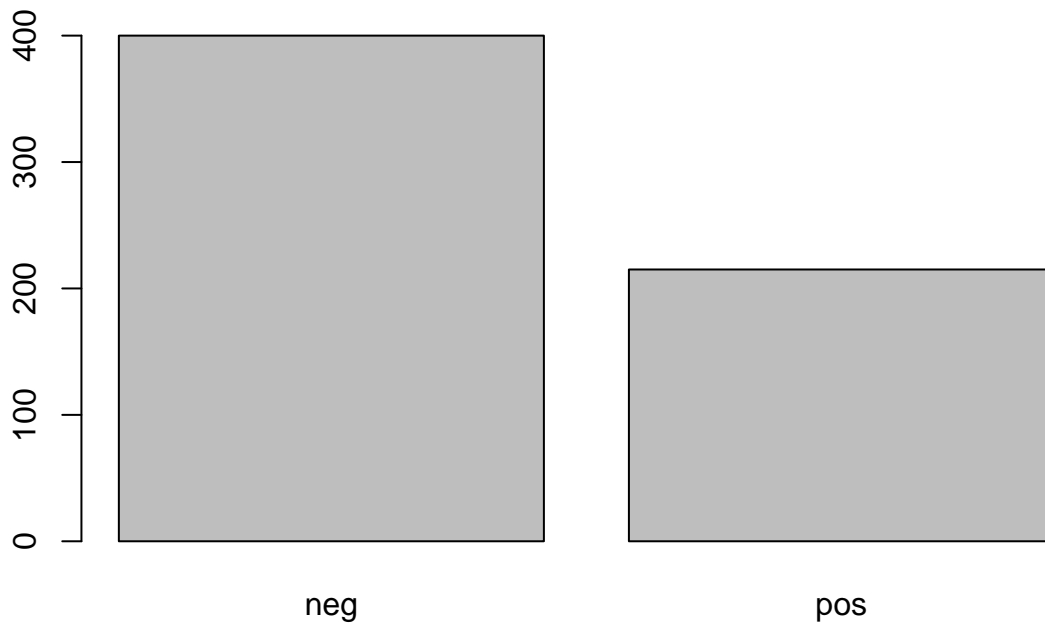
```

# boxplot for each attribute on one image
par(mfrow=c(1,4))
for(i in 5:8) {
  boxplot(x[,i], main=names(PimaIndiansDiabetes)[i])
}

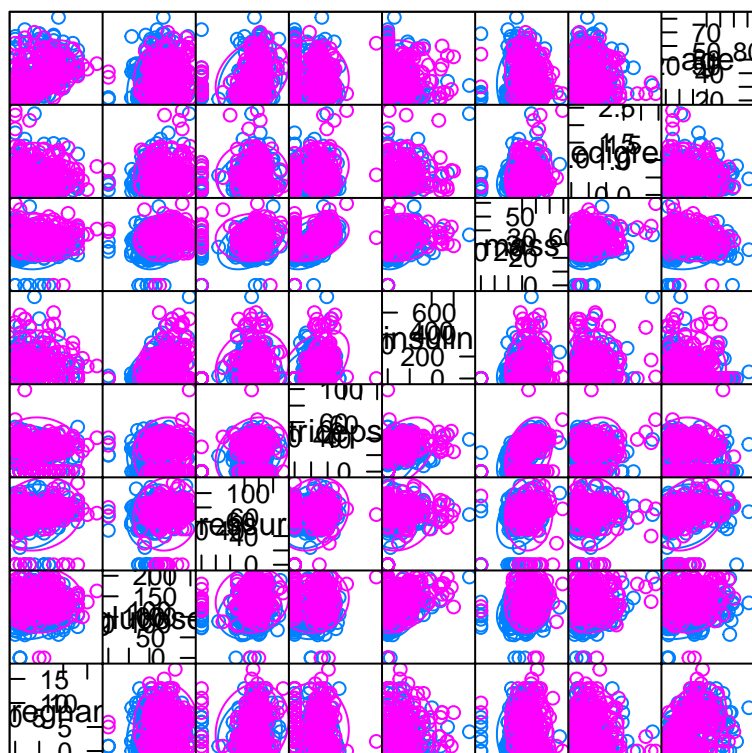
```



```
# barplot for class breakdown
plot(y)
```

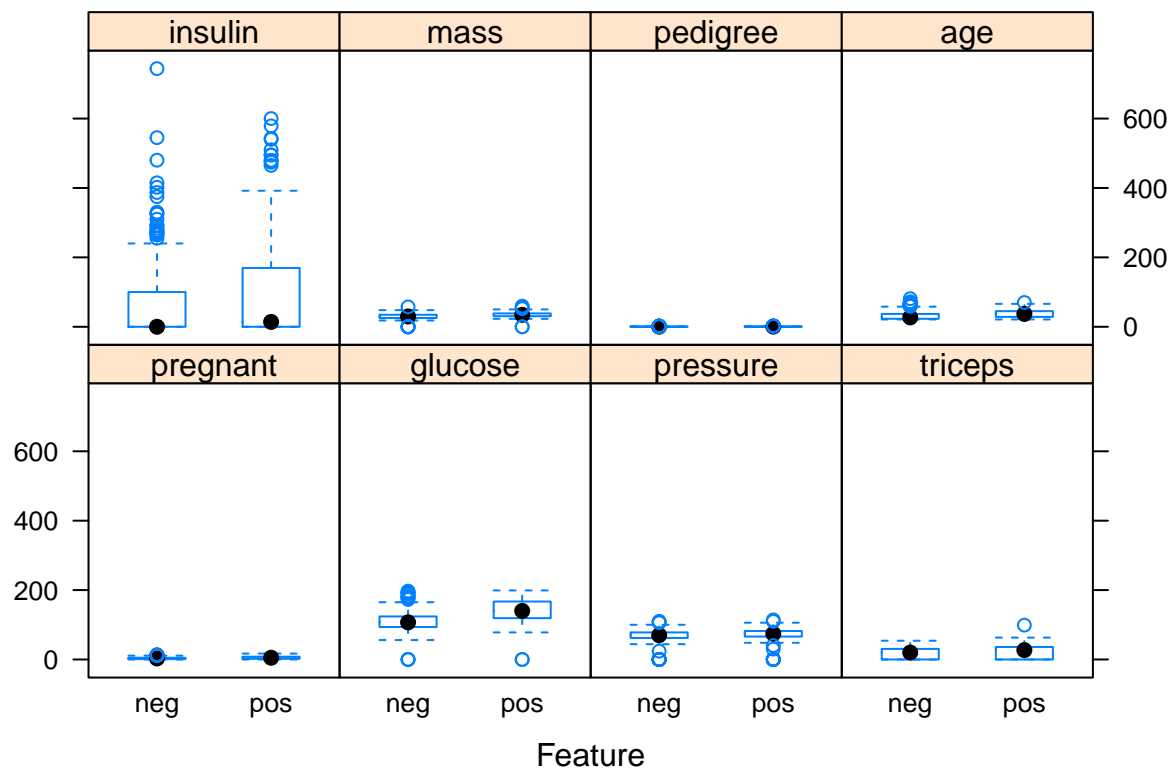


```
# scatterplot matrix
featurePlot(x=x, y=y, plot="ellipse")
```

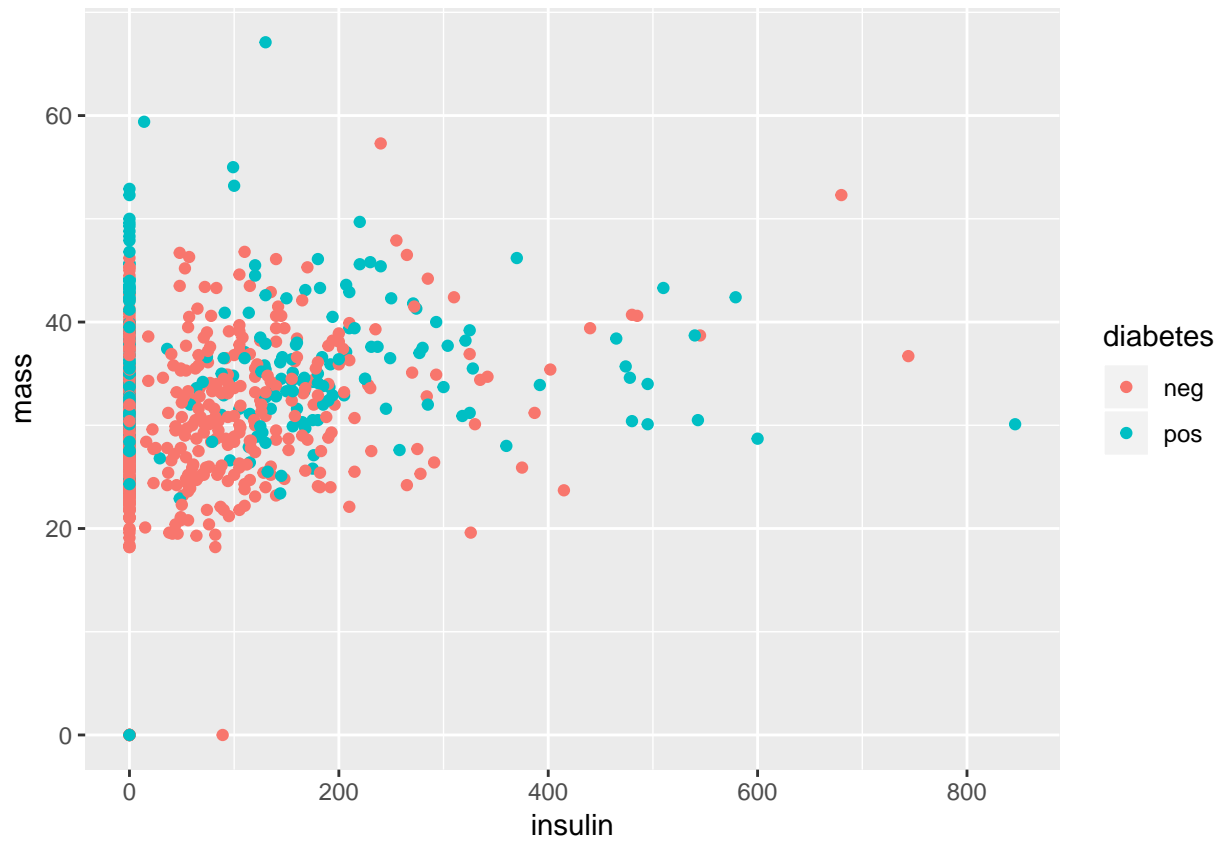


Scatter Plot Matrix

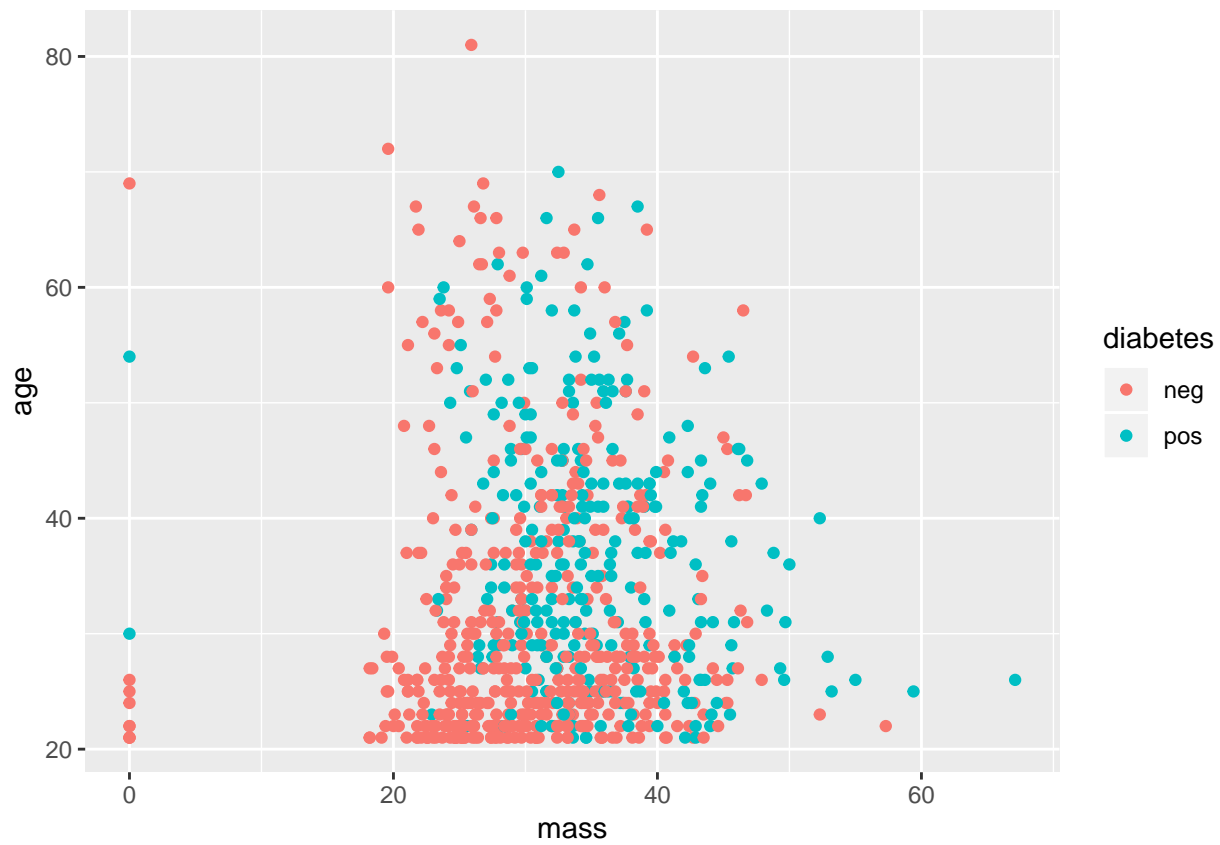
```
# box and whisker plots for each attribute
featurePlot(x=x, y=y, plot="box")
```



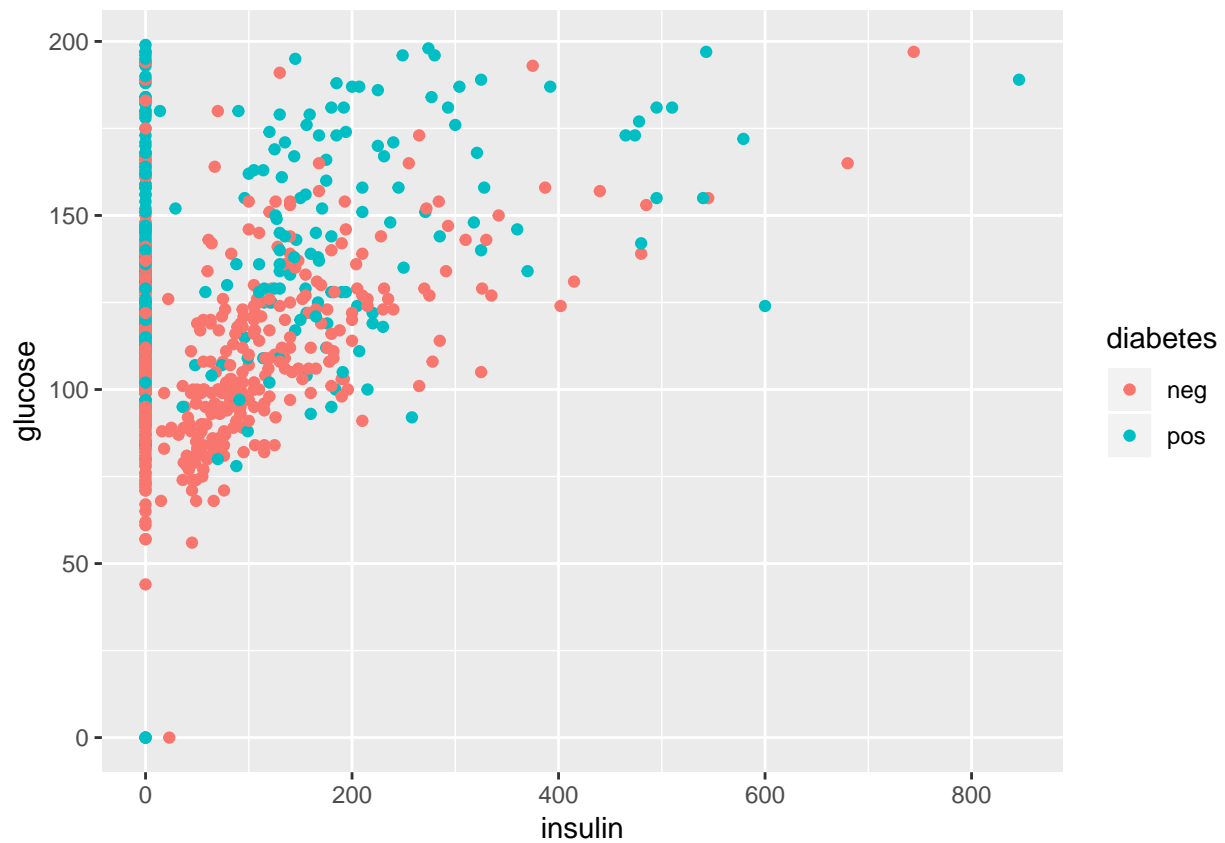
```
ggplot(PimaIndiansDiabetes, aes(insulin,mass, color = diabetes)) + geom_point()
```



```
ggplot(PimaIndiansDiabetes, aes(mass,age, color = diabetes)) + geom_point()
```

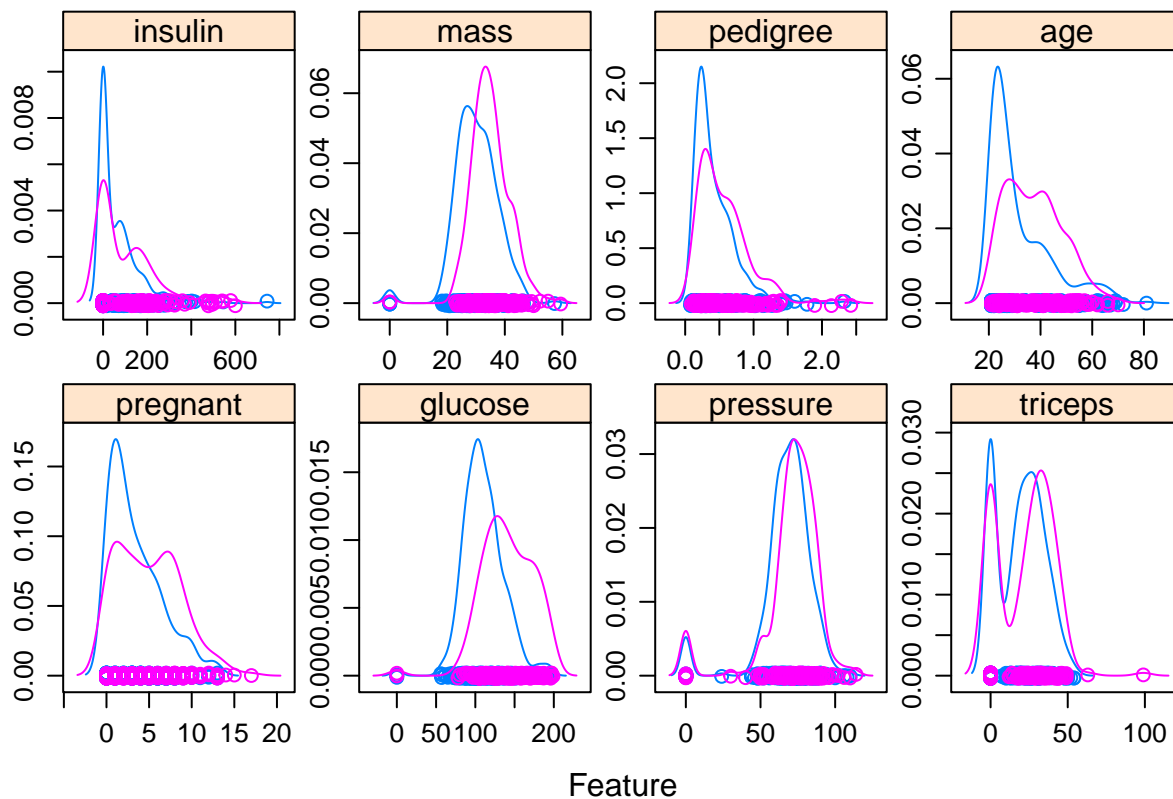


```
ggplot(PimaIndiansDiabetes, aes(insulin, glucose, color = diabetes)) + geom_point()
```



```
# density plots for each attribute by class value
scales <- list(x=list(relation="free"), y=list(relation="free"))
featurePlot(x=x, y=y, plot="density", scales=scales)
```





## Applying Classification Algorithms

```
# Run algorithms using 10-fold cross validation
control <- trainControl(method="cv", number=10)
metric <- "Accuracy"

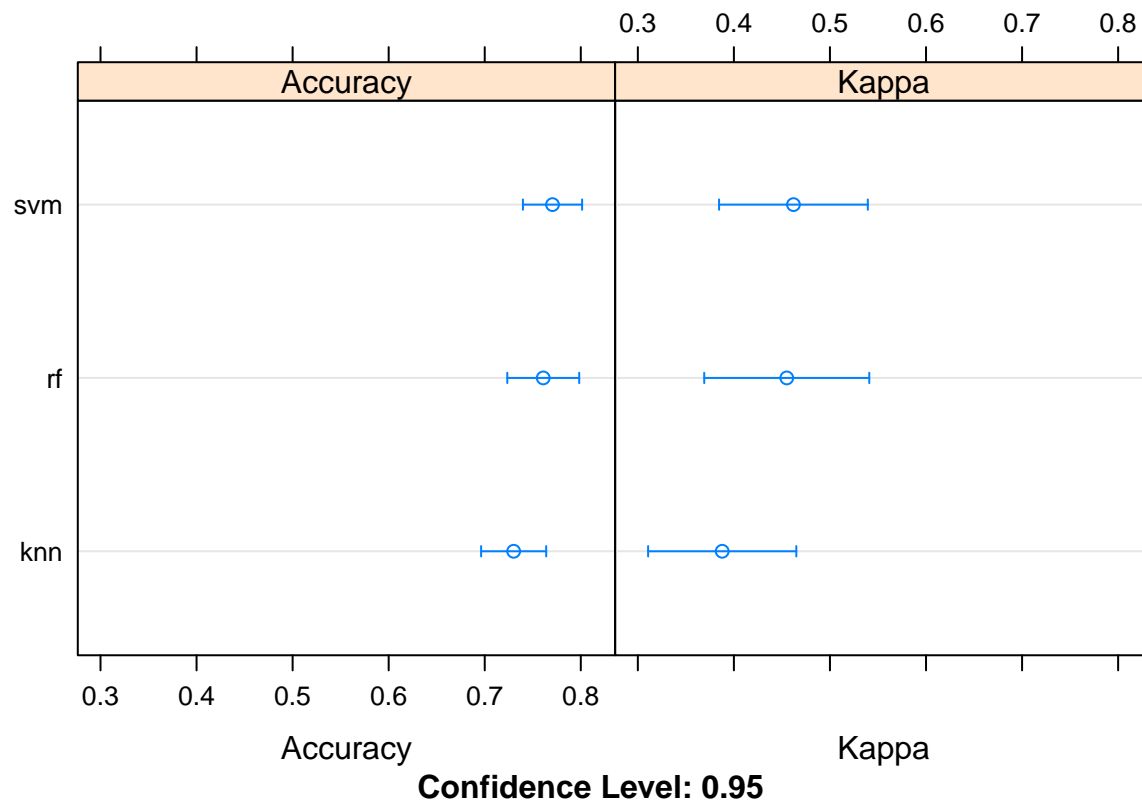
# kNN
set.seed(7)
fit.knn <- train(diabetes~., data=dataset, method="knn", metric=metric, trControl=control)
# SVM
set.seed(7)
fit.svm <- train(diabetes~., data=dataset, method="svmRadial", metric=metric, trControl=control)
# Random Forest
set.seed(7)
fit.rf <- train(diabetes~., data=dataset, method="rf", metric=metric, trControl=control)
```

## Comparison of the Classification Algorithms

```
# summarize accuracy of models
results <- resamples(list(knn=fit.knn, svm=fit.svm, rf=fit.rf))
summary(results)

##
## Call:
## summary.resamples(object = results)
##
```

```
## Models: knn, svm, rf
## Number of resamples: 10
##
## Accuracy
##      Min.      1st Qu.      Median      Mean      3rd Qu.      Max. NA's
## knn 0.6557377 0.6895161 0.7419355 0.7301163 0.7633527 0.7868852    0
## svm 0.7258065 0.7377049 0.7559492 0.7705711 0.7960074 0.8387097    0
## rf  0.6935484 0.7213115 0.7562136 0.7609201 0.8024194 0.8524590    0
##
## Kappa
##      Min.      1st Qu.      Median      Mean      3rd Qu.      Max. NA's
## knn 0.2287778 0.2955868 0.4097712 0.3878377 0.4778409 0.5113986    0
## svm 0.3612565 0.3767561 0.4210771 0.4619966 0.5342957 0.6327014    0
## rf  0.3154613 0.3610598 0.4267109 0.4550382 0.5487726 0.6768687    0
# compare accuracy of models
dotplot(results)
```



## Insights from the best model

```
# summarize Best Model
print(fit.svm)

## Support Vector Machines with Radial Basis Function Kernel
##
## 615 samples
## 8 predictors
## 2 classes: 'neg', 'pos'
```

```
##
## No pre-processing
## Resampling: Cross-Validated (10 fold)
## Summary of sample sizes: 554, 553, 554, 553, 554, 553, ...
## Resampling results across tuning parameters:
##
##      C      Accuracy   Kappa
##  0.25  0.7641460  0.4374239
##  0.50  0.7705711  0.4619966
##  1.00  0.7608408  0.4482180
##
## Tuning parameter 'sigma' was held constant at a value of 0.1351037
## Accuracy was used to select the optimal model using the largest value.
## The final values used for the model were sigma = 0.1351037 and C = 0.5.

# estimate skill of SVM on the validation dataset
predictions <- predict(fit.svm, validation)
confusionMatrix(predictions, validation$diabetes)

## Confusion Matrix and Statistics
##
##              Reference
## Prediction neg pos
##          neg  88  22
##          pos  12  31
##
##              Accuracy : 0.7778
##              95% CI : (0.7036, 0.8409)
##      No Information Rate : 0.6536
##      P-Value [Acc > NIR] : 0.000586
##
##              Kappa : 0.4865
##  McNemar's Test P-Value : 0.122713
##
##              Sensitivity : 0.8800
##              Specificity : 0.5849
##              Pos Pred Value : 0.8000
##              Neg Pred Value : 0.7209
##              Prevalence : 0.6536
##              Detection Rate : 0.5752
##      Detection Prevalence : 0.7190
##              Balanced Accuracy : 0.7325
##
##              'Positive' Class : neg
##
```

## Applying Clustering Algorithms

```
# K-means
set.seed(20)
fit.kmeans <- kmeans(PimaIndiansDiabetes[, 1:8], 2, nstart = 20)
# Hierarchical Agglomerative
set.seed(20)
```

```
d <- dist(PimaIndiansDiabetes[,1:8], method = "euclidean") # distance matrix
fit.ha <- hclust(d, method="ward.D")
# K-Medoids Clustering
num <- as.matrix(PimaIndiansDiabetes[,1:8])
mrwdist <- distNumeric(num, num, method = "mrw")
fit.kmedoids <- fastkmed(mrwdist, ncluster = 2, iterate = 50)
```

## Getting insights from Hierarchical Agglomerative Clustering

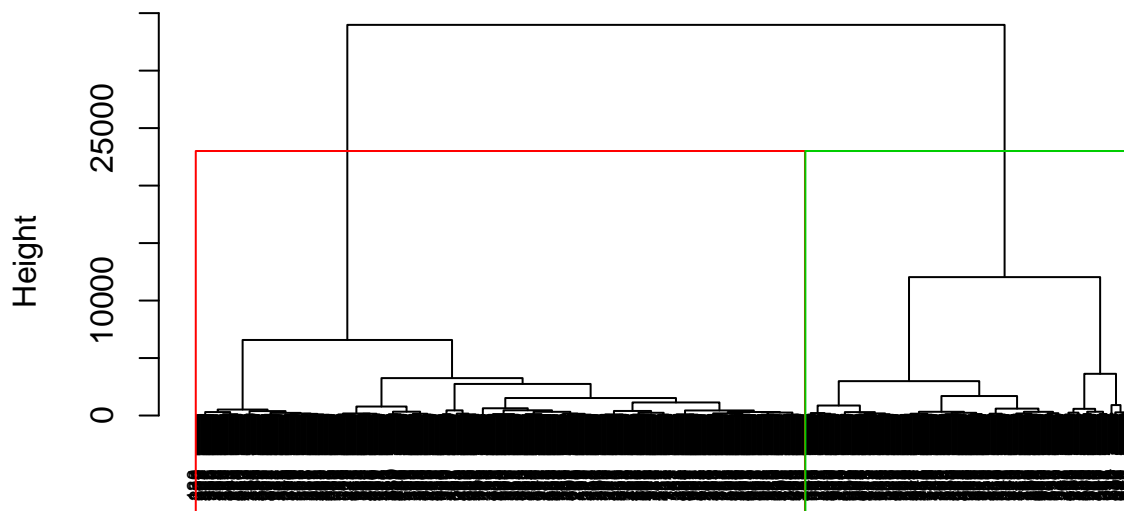
```
# Cut tree into 4 groups
sub_grp <- cutree(fit.ha, k = 2)

# Number of members in each cluster
table(sub_grp)
```

```
## sub_grp
## 1 2
## 500 268
## sub_grp
```

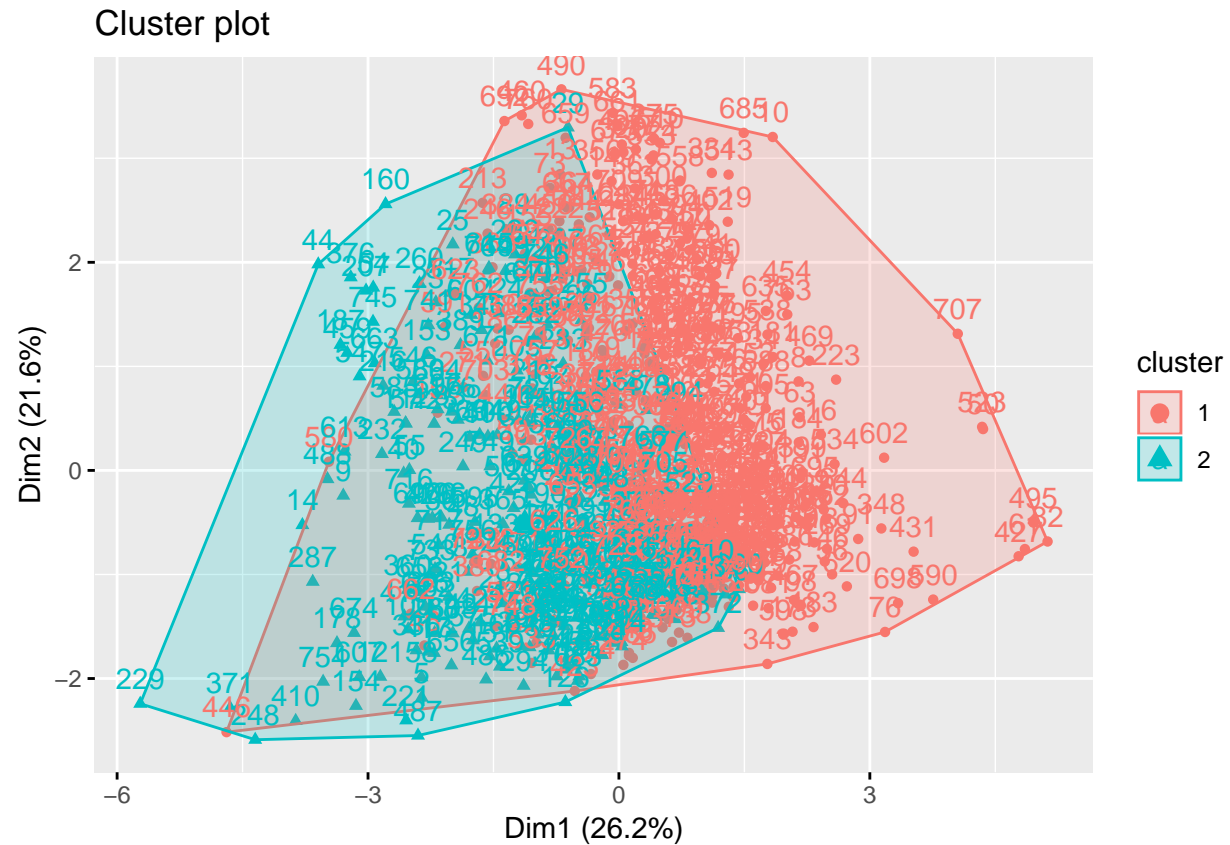
```
plot(fit.ha, cex = 0.6)
rect.hclust(fit.ha, k = 2, border = 2:5)
```

### Cluster Dendrogram



d  
hclust (\*, "ward.D")

```
fviz_cluster(list(data = PimaIndiansDiabetes[,1:8], cluster = sub_grp))
```

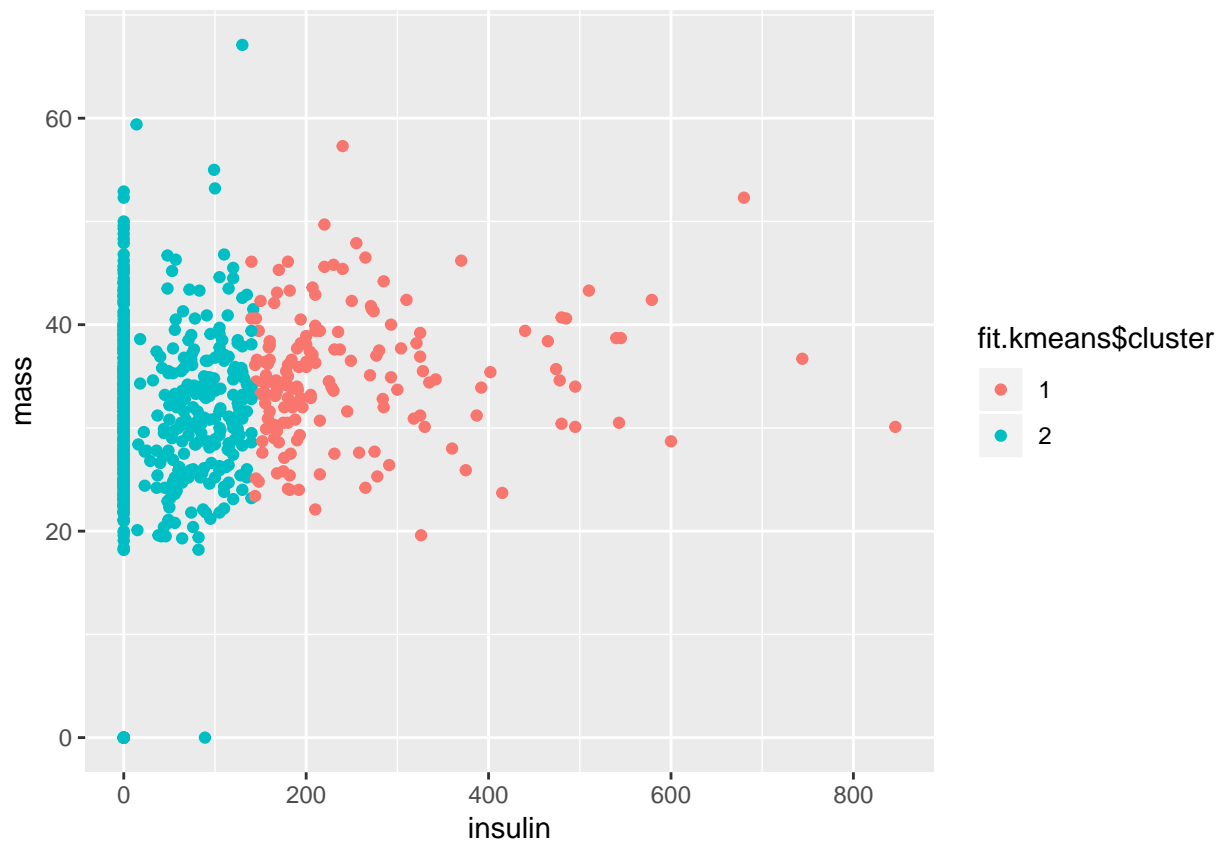


## Getting insights from K-Means Clustering

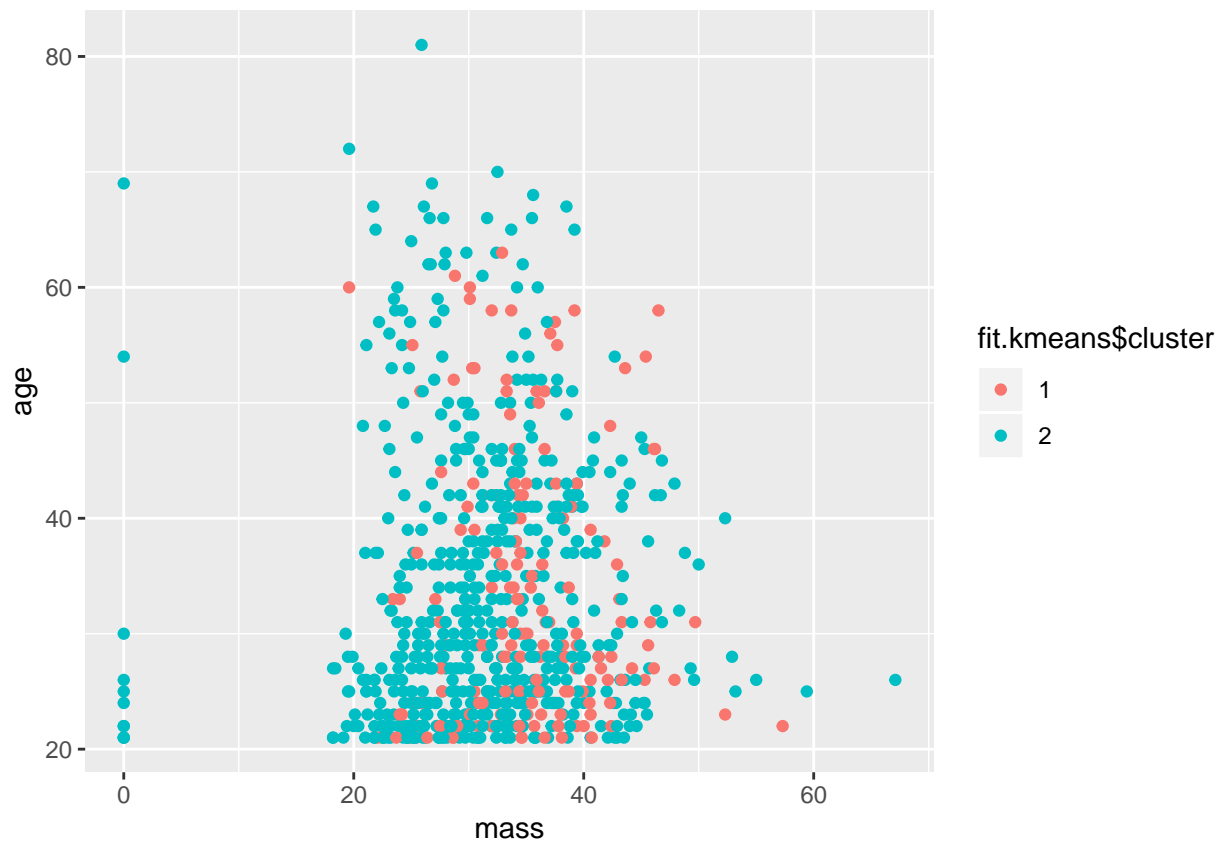
```
table(fit.kmeans$cluster, PimaIndiansDiabetes$diabetes)
```

```
##
##      neg pos
## 1    79  86
## 2   421 182
```

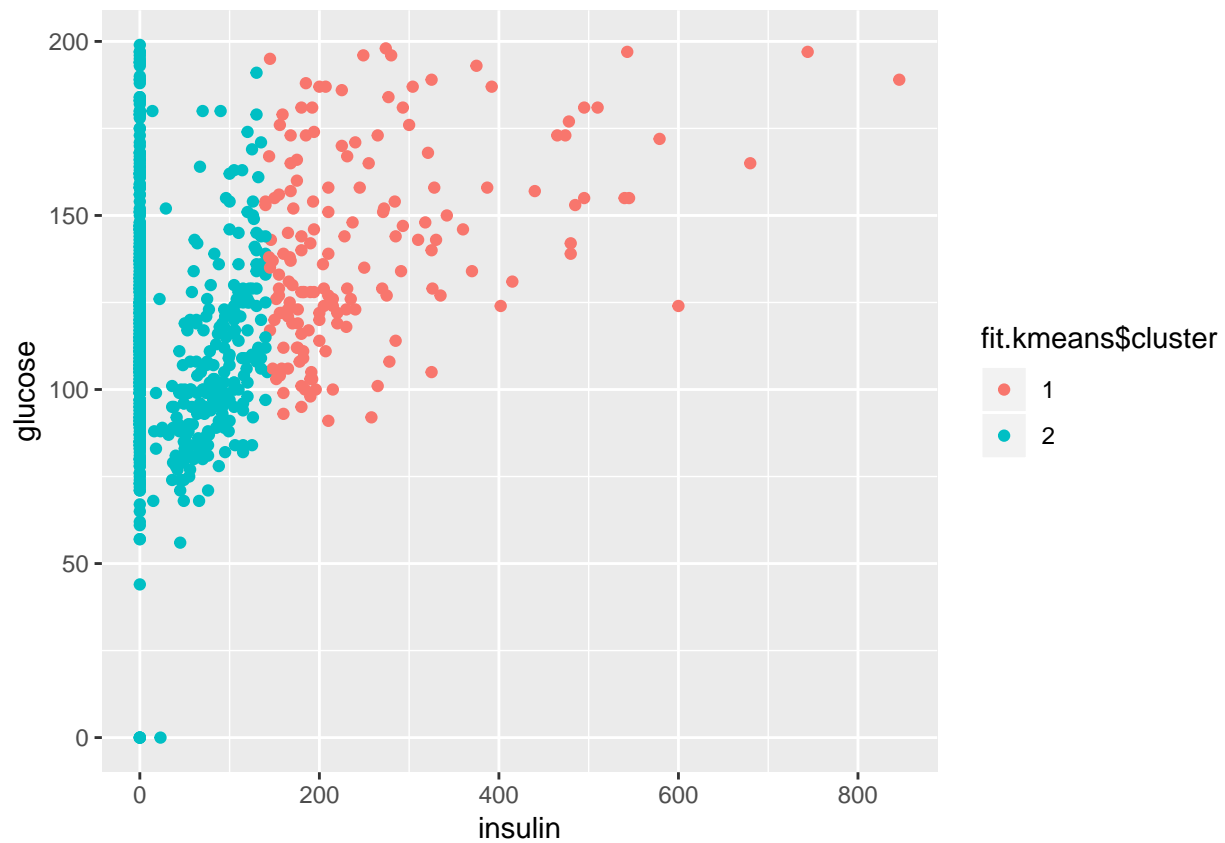
```
fit.kmeans$cluster <- as.factor(fit.kmeans$cluster)
ggplot(PimaIndiansDiabetes, aes(insulin, mass, color = fit.kmeans$cluster)) + geom_point()
```



```
fit.kmeans$cluster <- as.factor(fit.kmeans$cluster)
ggplot(PimaIndiansDiabetes, aes(mass, age, color = fit.kmeans$cluster)) + geom_point()
```

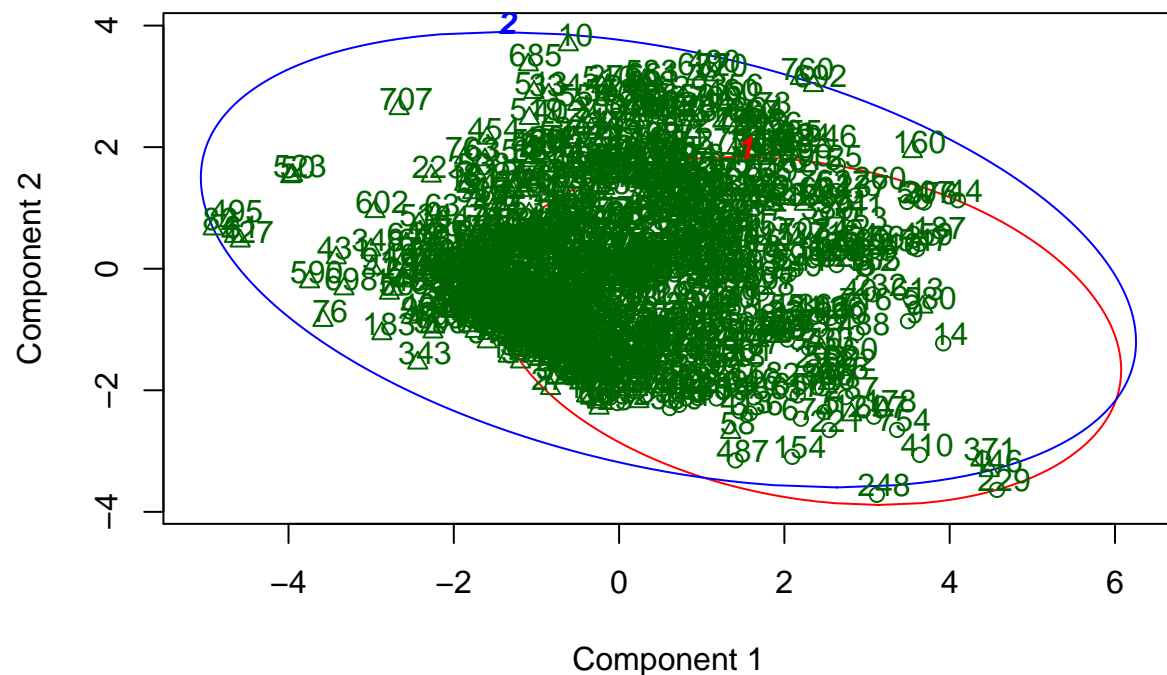


```
fit.kmeans$cluster <- as.factor(fit.kmeans$cluster)
ggplot(PimaIndiansDiabetes, aes(insulin, glucose, color = fit.kmeans$cluster)) + geom_point()
```



```
clusplot(PimaIndiansDiabetes, fit.kmeans$cluster, color=TRUE, shade=FALSE, labels=2, lines=0)
```

### CLUSPLOT( PimaIndiansDiabetes )



These two components explain 45.85 % of the point variability.

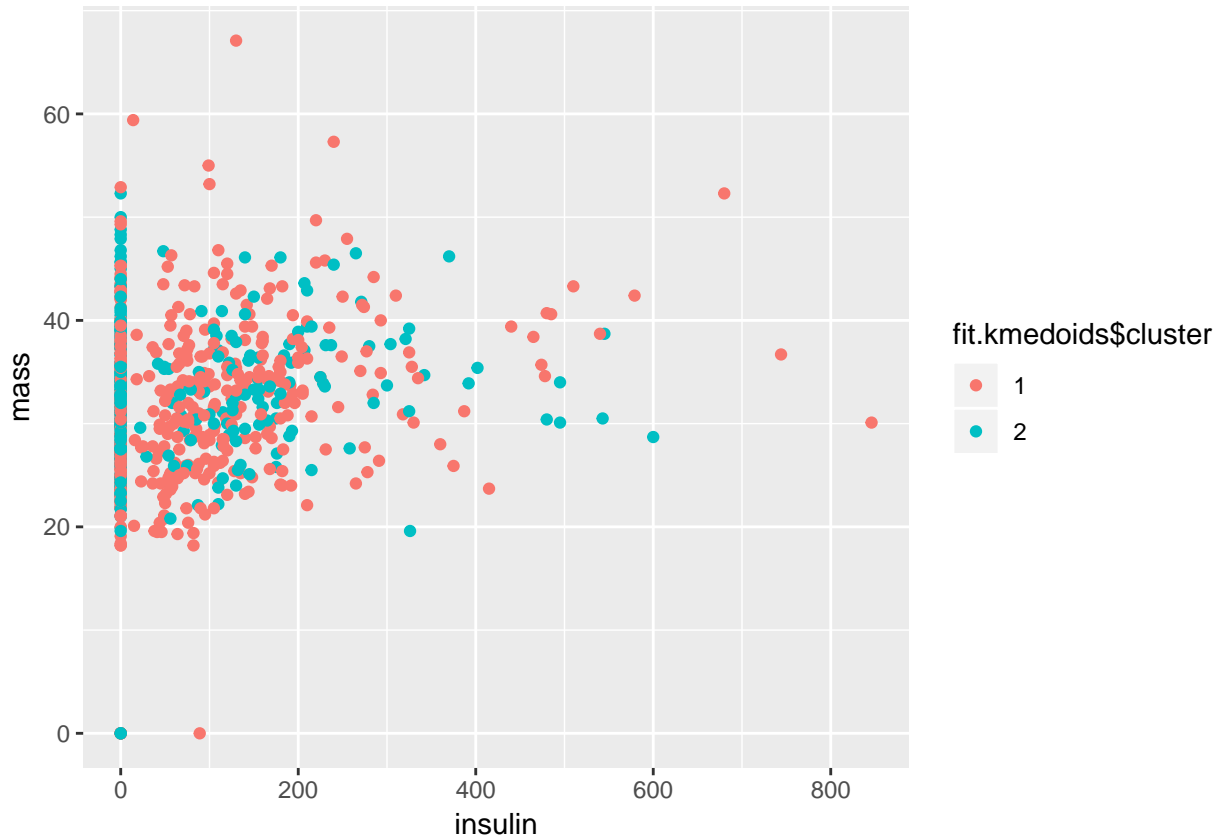


## Getting insights from K-Medoids Clustering

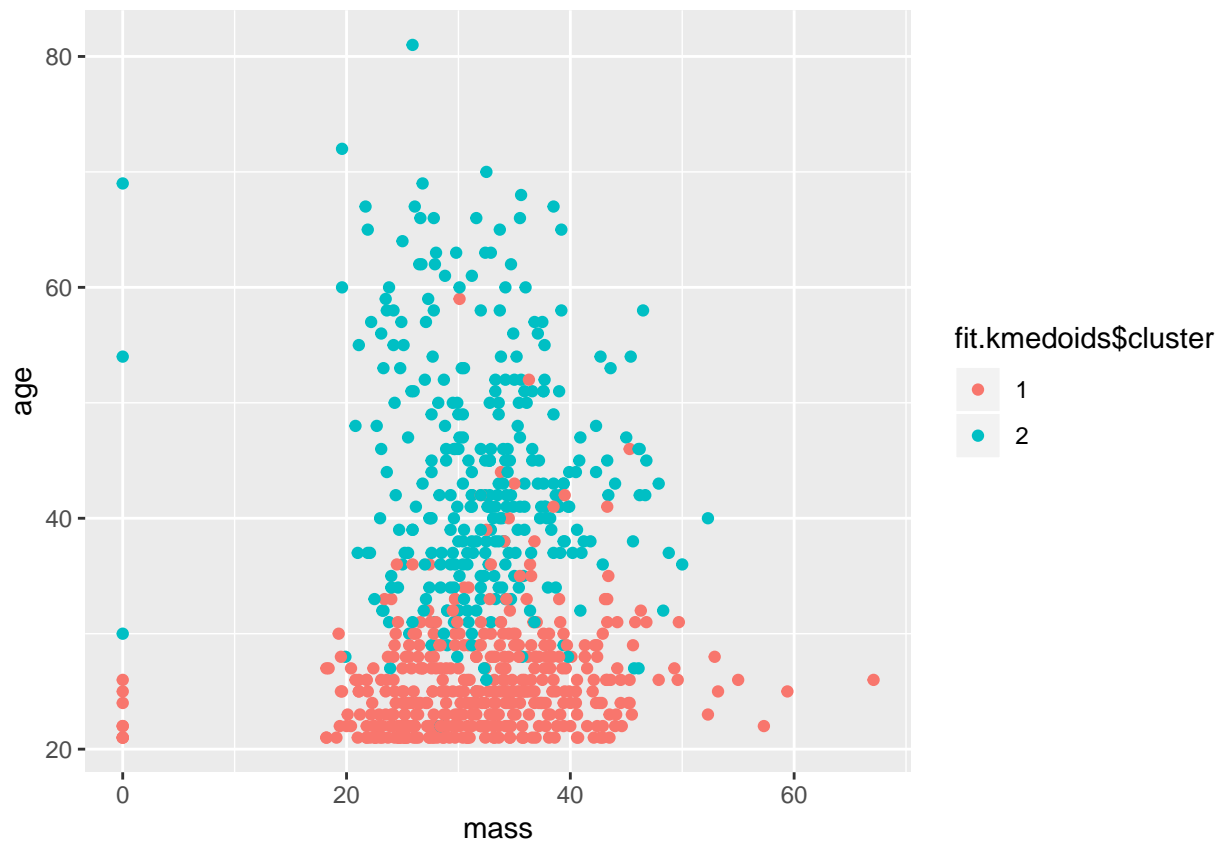
```
table(fit.kmedoids$cluster, PimaIndiansDiabetes[,9])
```

```
##  
##      neg pos  
##    1 342 109  
##    2 158 159
```

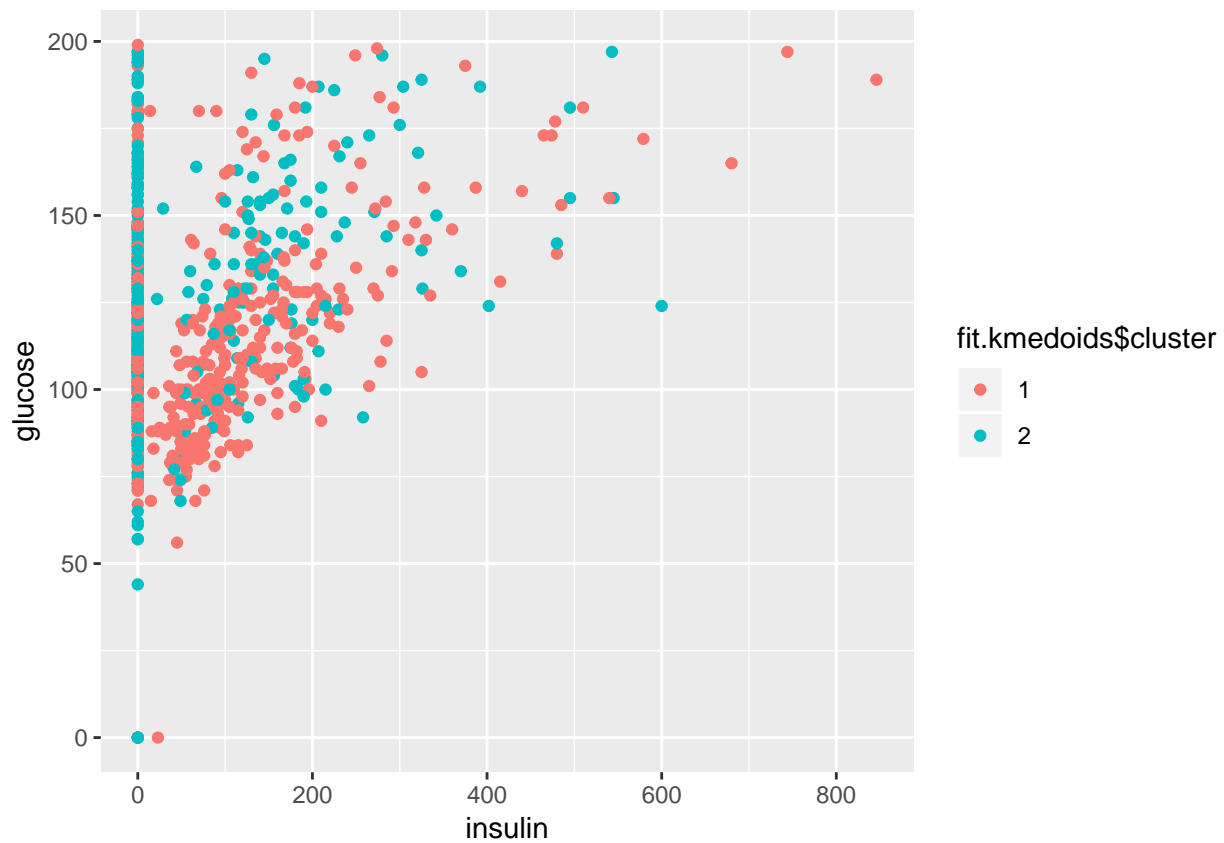
```
fit.kmedoids$cluster <- as.factor(fit.kmedoids$cluster)  
ggplot(PimaIndiansDiabetes, aes(insulin, mass, color = fit.kmedoids$cluster)) + geom_point()
```



```
fit.kmedoids$cluster <- as.factor(fit.kmedoids$cluster)  
ggplot(PimaIndiansDiabetes, aes(mass, age, color = fit.kmedoids$cluster)) + geom_point()
```

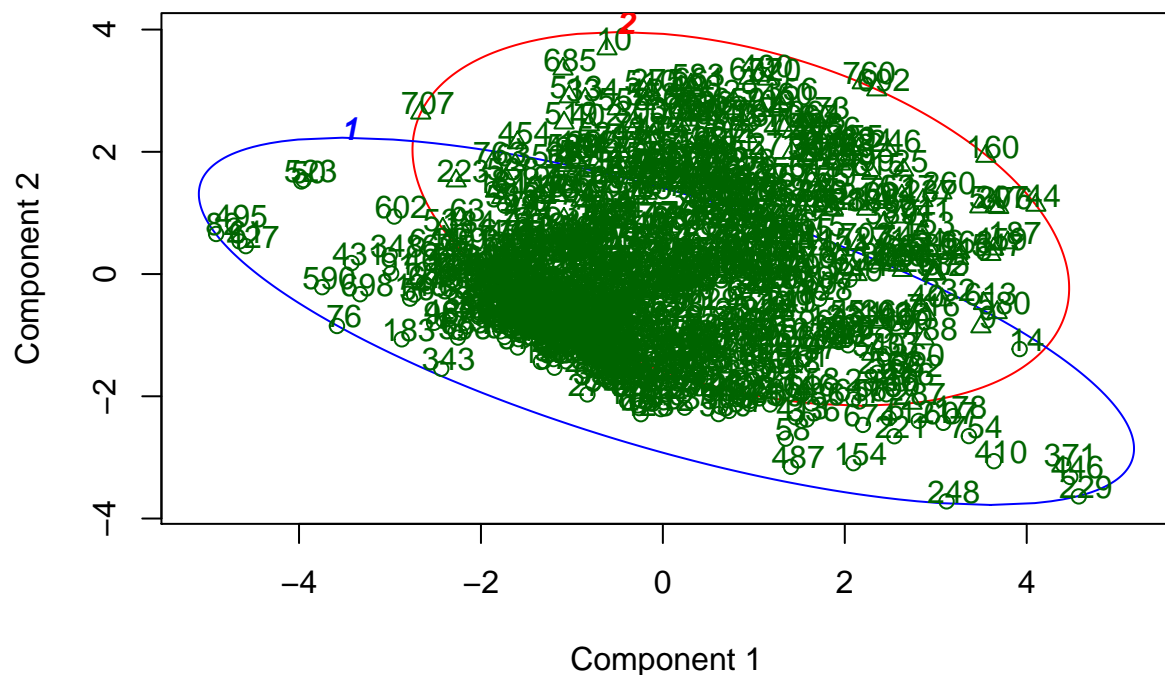


```
fit.kmedoids$cluster <- as.factor(fit.kmedoids$cluster)
ggplot(PimaIndiansDiabetes, aes(insulin, glucose, color = fit.kmedoids$cluster)) + geom_point()
```



```
clusplot(PimaIndiansDiabetes, fit.kmedoids$cluster, color=TRUE, shade=FALSE, labels=2, lines=0)
```

### CLUSPLOT( PimaIndiansDiabetes )



These two components explain 45.85 % of the point variability.

## Conclusion

With better accuracy and kappa measures, SVM has outperformed other competitors on Glass Dataset while Hierarchical Agglomerative Clustering is the winner when compared with K-Means and K-Medoids Clustering on Glass Dataset as it has clustered data better evident from the Cluster Plot and Cluster Dendrogram.