

# Plate reader analysis using NEMO

## Part 1

# Plate reformatting

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#### Aim:

Reformat the kinetics data from the plate reader to make it useful for any further required analysis.

#### Introduction:

In order to analyse the data we need to reformat the data file that is generated by the plate reader. We have used: Plate Reader EnSight Multimode Plate Reader (LOCATION SW511 L5-2419 (QM Elizabeth Li)).

Data file is a csv file that records absorbance results, then fluorescence and finally luminescence results. Plate repeat numbers correspond to the cycle number or time points. This SOP guides you to reformat the data using the script accessible in the share folder.

### Prerequisite:

- 1) Shared pathway to be used for Data processing
- 2) Connect to nemo on Terminal:

```
ssh -Y vour name@login.nemo.thecrick.org
```

3) Copy the template folder with scripts from the share folder to your user folder:

cp -rp /camp/lab/debenedictise/home/shared/microplatereader template

/camp/lab/debenedictise/home/users/your\_name/your\_folder\_name

4) Put your plate reader output data under <a href="your\_folder\_name">your\_folder\_name</a> as a csv file <a href="(your\_csv\_file.csv">(your\_csv\_file.csv</a>) and your plate content information as <a href="Manifest.csv">Manifest.csv</a>. For this you can use any of these two methods:

#### Method 1:

Open another Terminal window (not connected to nemo) and use scp command on your terminal to transfer your csv file to nemo:

```
scp -rp '/Users/path_to_your_file/your_csv_file.csv'
```

vour\_name@login.nemo.thecrick.org:/camp/lab/debenedictise/home/users/your\_name/your\_folder\_name/.

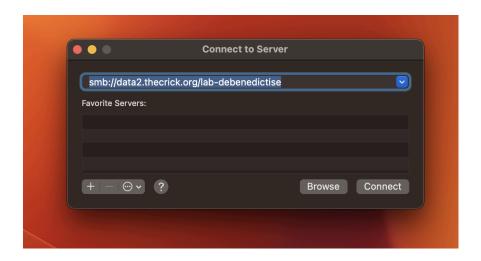
scp -rp '/Users/path\_to\_your\_file/Manifest.csv'

your\_name@login.nemo.thecrick.org:/camp/lab/debenedictise/home/users/your\_name/your\_folder\_name/.

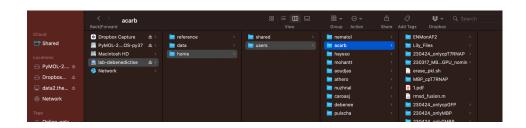
### Method 2:

Use smb file transfer which treats nemo as an external memory drive on your computer.

a. Press [command + K] when Finder is open or Finder>Go>Network. The following screen will appear. Write the address as given and click Connect.



- b. Type your username and password for Crick (nemo)
- c. Once connected you will see the folders:



d. Drag your file into the your\_folder\_name/ under lab-debenedictise/home/user/your\_name

- 5) Now you have both your csv file to be formatted and analysed, and the required formatting and analysis scripts in your folder name/.
- 6) Go back to Terminal with nemo connected and go to your folder including the csv and script files with the following command:

```
cd /camp/lab/debenedictise/home/users/your name/your folder name/
```

7) To perform calculations, we need to go to vis partition (has more computational capacity) instead of login, with:

```
srun -N 1 -p vis --gres=gpu:1 -t 05:00:00 --pty --x11 bash
```

For x11 to work, XQuartz must be downloaded on your computer if you are using Mac:

#### https://www.xquartz.org/

```
[[acarb@login002] shared]$ history | grep srun
292 history | grep srun
293 srun -N 1 -p vis --gres=gpu:1 -t 05:00:00 --pty --x11 bash
331 history | grep srun
332 srun -N 1 -p vis --gres=gpu:1 -t 05:00:00 --pty --x11 bash
335 srun -N 1 -p vis --gres=gpu:1 -t 05:00:00 --pty --x11 bash
478 srun -N 1 -p vis --gres=gpu:1 -t 05:00:00 --pty --x11 bash
501 srun -N 1 -p vis --gres=gpu:1 -t 05:00:00 --pty --x11 bash
624 srun -N 1 -p vis --gres=gpu:1 -t 05:00:00 --pty --x11 bash
1079 history | grep srun
[[acarb@login002] shared]$ srun -N 1 -p vis --gres=gpu:1 -t 05:00:00 --pty --x11 bash
[acarb@login002] shared]$
```

Now you are ready to go:) Call function callreader.sh to format the csv file:

```
./callreader.sh your_csv_file.csv
```

This will create two separate folders for Fluorescence and Luminescence with the exact names, both including OD data. You can use the formatted csv files (20200530\_erika\_taga\_00001\_reader\_plate\_1\_kinetic\_supp\_abs.csv for OD and 20200530\_erika\_taga\_00001\_reader\_plate\_1\_kinetic\_supp\_3\_high.csv for Fluorescence or Luminescence, depending on the parent folder) under

these folders directly for your own analysis or continue with Erika's analysis with the following:

o Go to Fluorescence folder

cd Fluorescence

o Call analysis function

./callanalysis.sh

o Repeat these two steps for Luminescence