class12

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Comparative analysis of protein structures

Using the bio3D package. Note: We used a different file1 (1hel not 1hsg) than in the lab walk-through.

```
library(bio3d)
pdb <- read.pdb("1hel")</pre>
##
     Note: Accessing on-line PDB file
pdb
##
    Call:
          read.pdb(file = "1hel")
##
##
##
      Total Models#: 1
        Total Atoms#: 1186, XYZs#: 3558 Chains#: 1 (values: A)
##
##
        Protein Atoms#: 1001 (residues/Calpha atoms#: 129)
##
##
        Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
##
##
        Non-protein/nucleic Atoms#: 185 (residues: 185)
        Non-protein/nucleic resid values: [ HOH (185) ]
##
##
##
      Protein sequence:
         KVFGRCELAAAMKRHGLDNYRGYSLGNWVCAAKFESNFNTQATNRNTDGSTDYGILQINS
##
##
         RWWCNDGRTPGSRNLCNIPCSALLSSDITASVNCAKKIVSDGNGMNAWVAWRNRCKGTDV
         QAWIRGCRL
##
##
## + attr: atom, xyz, seqres, helix, sheet,
##
           calpha, remark, call
```

Q7: How many amino acid residues are there in this pdb object?

There are 129 residues.

Q8: Name one of the non-protein residues?

HOH (185)

Q9: How many protein chains are in this structure?

1 chain

Setup

```
#install.packages("bio3d")
#install.packages("ggplot2")
#install.packages("ggrepel")
#install.packages("devtools")
#install.packages("BiocManager")

#BiocManager::install("msa")
#devtools::install_bitbucket("Grantlab/bio3d-view")
```

Q10. Which of the packages above is found only on BioConductor and not CRAN?

msa

Q11. Which of the above packages is not found on BioConductor or CRAN?

bio3d-view found in this line: devtools::install_bitbucket("Grantlab/bio3d-view")

**Q12. True or False? Functions from the devtools package can be used to install packages from GitHub and BitBucket?*

True

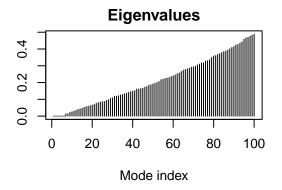
Let's use bioinformatics method called NMA (Normal Mode Analysis) to predict the dynamics (flexibility) of this enzyme.

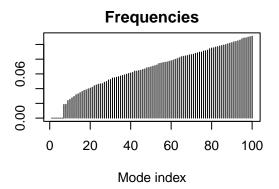
```
modes <- nma(pdb)

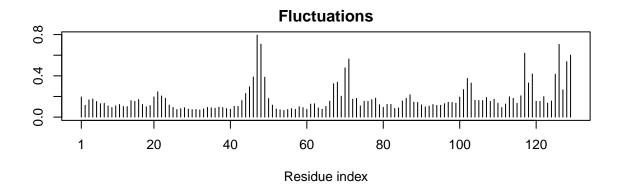
## Building Hessian... Done in 0.016 seconds.

## Diagonalizing Hessian... Done in 0.111 seconds.

plot(modes)</pre>
```

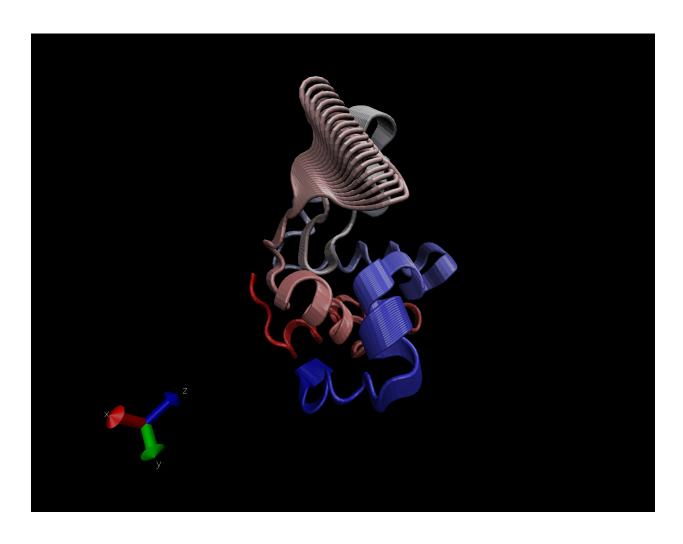






Make a "movie" of its predicted motion. We often call this a "trajectory".

mktrj(modes, file="nma.pdb")



Analysis of ADK

```
##
                                                                              180
               VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
## pdb|1AKE|A
##
              121
                                                                              180
##
##
              181
                                                   214
               YYSKEAEAGNTKYAKVDGTKPVAEVRADLEKILG
## pdb | 1AKE | A
##
              181
##
## Call:
     read.fasta(file = outfile)
##
##
## Class:
##
     fasta
##
## Alignment dimensions:
     1 sequence rows; 214 position columns (214 non-gap, 0 gap)
##
## + attr: id, ali, call
#run BLAST from R; may time out so we can use the other command
# blast <- blast.pdb(aa)</pre>
hits <- NULL
hits$pdb.id <- c('1AKE A','4X8M A','6S36 A','6RZE A','4X8H A','3HPR A','1E4V A','5EJE A','1E4Y A','3X2S
```

Q13. How many amino acids are in this sequence, i.e. how long is this sequence?

214

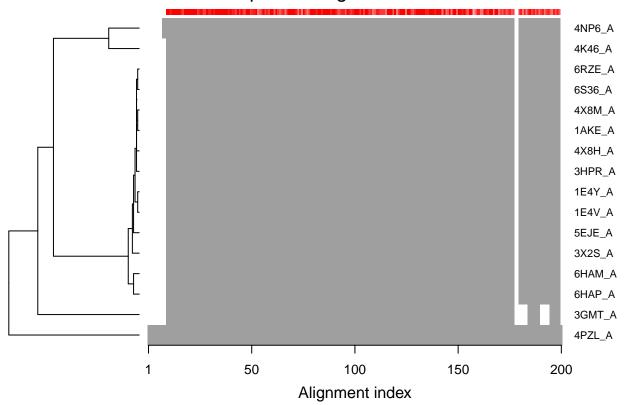
```
#hits <- plot(blast)
hits$pdb.id
   [1] "1AKE A" "4X8M A" "6S36 A" "6RZE A" "4X8H A" "3HPR A" "1E4V A" "5EJE A"
   [9] "1E4Y A" "3X2S A" "6HAP A" "6HAM A" "4K46 A" "4NP6 A" "3GMT A" "4PZL A"
#Download related PDB files, commenting out so I don't waste timer rerunning this
files <-get.pdb(hits$pdb.id, path="pdbs", split = TRUE, gzip = TRUE)</pre>
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 1AKE.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 4X8M.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 6S36.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 6RZE.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 4X8H.pdb.gz exists. Skipping download
```

```
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 3HPR.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 1E4V.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 5EJE.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 1E4Y.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 3X2S.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 6HAP.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 6HAM.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 4K46.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 4NP6.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 3GMT.pdb.gz exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 4PZL.pdb.gz exists. Skipping download
##
Multiple structure alignment
pdbs <- pdbaln(files, fit = TRUE)
## Reading PDB files:
## pdbs/split_chain/1AKE_A.pdb
## pdbs/split_chain/4X8M_A.pdb
## pdbs/split_chain/6S36_A.pdb
## pdbs/split_chain/6RZE_A.pdb
## pdbs/split chain/4X8H A.pdb
## pdbs/split_chain/3HPR_A.pdb
## pdbs/split_chain/1E4V_A.pdb
```

pdbs/split_chain/5EJE_A.pdb
pdbs/split_chain/1E4Y_A.pdb
pdbs/split_chain/3X2S_A.pdb

```
## pdbs/split chain/6HAP A.pdb
## pdbs/split_chain/6HAM_A.pdb
## pdbs/split chain/4K46 A.pdb
## pdbs/split_chain/4NP6_A.pdb
## pdbs/split_chain/3GMT_A.pdb
## pdbs/split chain/4PZL A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
        PDB has ALT records, taking A only, rm.alt=TRUE
## ..
##
       PDB has ALT records, taking A only, rm.alt=TRUE
       PDB has ALT records, taking A only, rm.alt=TRUE
       PDB has ALT records, taking A only, rm.alt=TRUE
         PDB has ALT records, taking A only, rm.alt=TRUE
## . PDB has ALT records, taking A only, rm.alt=TRUE
##
##
## Extracting sequences
##
  pdb/seq: 1
                name: pdbs/split chain/1AKE A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
## pdb/seq: 2
                name: pdbs/split_chain/4X8M_A.pdb
  pdb/seq: 3
                name: pdbs/split_chain/6S36_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
                name: pdbs/split_chain/6RZE_A.pdb
  pdb/seq: 4
      PDB has ALT records, taking A only, rm.alt=TRUE
                name: pdbs/split_chain/4X8H_A.pdb
## pdb/seq: 5
  pdb/seq: 6
                name: pdbs/split_chain/3HPR_A.pdb
##
      PDB has ALT records, taking A only, rm.alt=TRUE
                name: pdbs/split_chain/1E4V_A.pdb
  pdb/seq: 7
  pdb/seq: 8
                name: pdbs/split_chain/5EJE_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
## pdb/seq: 9
                name: pdbs/split_chain/1E4Y_A.pdb
## pdb/seq: 10
                 name: pdbs/split_chain/3X2S_A.pdb
## pdb/seq: 11
                 name: pdbs/split_chain/6HAP_A.pdb
## pdb/seq: 12
                 name: pdbs/split_chain/6HAM_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
## pdb/seq: 13
                 name: pdbs/split_chain/4K46_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
## pdb/seq: 14
                 name: pdbs/split_chain/4NP6_A.pdb
## pdb/seq: 15
                 name: pdbs/split_chain/3GMT_A.pdb
## pdb/seq: 16
                 name: pdbs/split_chain/4PZL_A.pdb
ids <- basename.pdb(pdbs$id)</pre>
plot(pdbs, labels=ids)
```

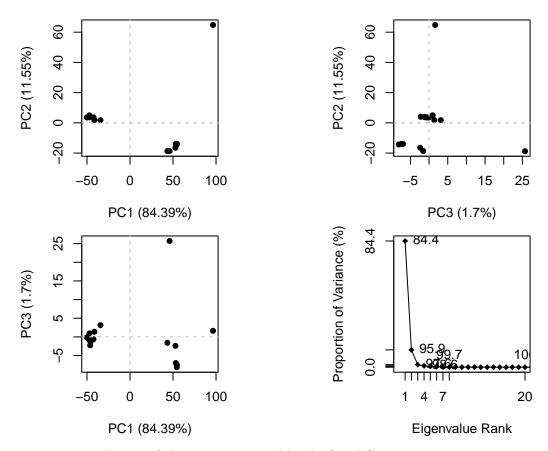
Sequence Alignment Overview



```
library(bio3d.view)
library(rg1)
view.pdbs(pdbs)
```

PCA

```
pc.xray <- pca(pdbs)
plot(pc.xray)</pre>
```



Make a trajectory visualizaitn of the motion captured by the first PC.

```
#visualize first PC
pc1 <- mktrj(pc.xray, pc=1, file="pc_1.pdb")</pre>
```

