hochschule mannheim



Extension of two dimensions morphogenesis simulation models of the urothelium into three dimensions within the moduro simulation environment

Thorsten Mueller

Bachelor Thesis

for the acquisition of the academic degree Bachelor of Science (B.Sc.)

Course of Studies: Computer Science

Department of Computer Science
University of Applied Sciences Mannheim

11.11.2015

Tutors

Prof. Dr. Markus Gumbel, Hochschule Mannheim Erika Mustermann, Paukenschlag GmbH

Mueller, Thorsten:

Extension of two dimensions morphogenesis simulation models of the urothelium into three dimensions within the moduro simulation environment / Thorsten Mueller. — Bachelor Thesis, Mannheim: University of Applied Sciences Mannheim, 2018. 44 pages.

Mueller, Thorsten:

Einsatz eines Flux-Kompensators für Zeitreisen mit einer maximalen Höchstgeschwindigkeit von WARP 7 / Thorsten Mueller.

Bachelor-Thesis, Mannheim: Hochschule Mannheim, 2018. 44 Seiten.

Erklärung

Hiermit erkläre ich, dass ich die vorliegende Arbeit selbstständig verfasst und keine anderen als die angegebenen Quellen und Hilfsmittel benutzt habe.

Ich bin damit einverstanden, dass meine Arbeit veröffentlicht wird, d. h. dass die Arbeit elektronisch gespeichert, in andere Formate konvertiert, auf den Servern der Hochschule Mannheim öffentlich zugänglich gemacht und über das Internet verbreitet werden darf.

Mannheim, 11.11.2015

Thorsten Mueller

Abstract

Extension of two dimensions morphogenesis simulation models of the urothelium into three dimensions within the moduro simulation environment

The European languages are members of the same family. Their separate existence is a myth. For science, music, sport, etc, Europe uses the same vocabulary. The languages only differ in their grammar, their pronunciation and their most common words. Everyone realizes why a new common language would be desirable: one could refuse to pay expensive translators. To achieve this, it would be necessary to have uniform grammar, pronunciation and more common words. If several languages coalesce, the grammar of the resulting language is more simple and regular than that of the individual languages. The new common language will be more simple and regular than the existing European languages. It will be as simple as Occidental; in fact, it will be Occidental. To an English person, it will seem like simplified English, as a skeptical Cambridge friend of mine told me what Occidental is.

Einsatz eines Flux-Kompensators für Zeitreisen mit einer maximalen Höchstgeschwindigkeit von WARP 7

Jemand musste Josef K. verleumdet haben, denn ohne dass er etwas Böses getan hätte, wurde er eines Morgens verhaftet. Wie ein Hund! sagte er, es war, als sollte die Scham ihn überleben. Als Gregor Samsa eines Morgens aus unruhigen Träumen erwachte, fand er sich in seinem Bett zu einem ungeheueren Ungeziefer verwandelt. Und es war ihnen wie eine Bestätigung ihrer neuen Träume und guten Absichten, als am Ziele ihrer Fahrt die Tochter als erste sich erhob und ihren jungen Körper dehnte. Es ist ein eigentümlicher Apparat, sagte der Offizier zu dem Forschungsreisenden und überblickte mit einem gewissermaßen bewundernden Blick den ihm doch wohl bekannten Apparat. Sie hätten noch ins Boot springen können, aber der Reisende hob ein schweres, geknotetes Tau vom Boden, drohte ihnen damit und hielt sie dadurch von dem Sprunge ab. In den letzten Jahrzehnten ist das Interesse an Künstlern sehr zurückgegangen. Aber sie überwanden sich, umdrängten den Käfig und wollten sich gar nicht fortrühren.

Contents

1	Intro	oduction 1
	1.1	Motivation
	1.2	Background
		1.2.1 Biology of the Urothelium
		1.2.2 CompuCell3D
		1.2.3 Glazier Graner Hogeweg Model
	1.3	Objective
	1.4	Outline
2	Stat	e of the Art
	2.1	Moduro
	2.2	Display and Simulation of the Urothelium
	2.3	Moduro Toolbox
	2.4	Models
	2.5	Adhesion
	2.6	Cell properties
	2.7	Events in the simulation
		2.7.1 Cell Growth
		2.7.2 Mitosis
		2.7.3 Necrosis
		2.7.4 Mutation
		2.7.5 Transformation
		2.7.6 Urination
	2.8	Fitness functions
		2.8.1 Arrangement fitness function
		2.8.2 Volume fitness function
		2.8.3 Overall fitness function
	2.9	3D functionalaties
3	Metl	nods 18
	3.1	Lambda multipliers
	3.2	Abstract methods
	3.3	Calculation steps until urination
	3.4	Area of stem cells on the basal membrane

Contents

	3.5 3.6 3.7 3.8 3.9	Target volume and target surface after mitosis	22 23 25 28 31
4	Res : 4.1	Ults Draw sphere cells	33
	4.2	Grow sphere cells	36
	4.3	Simulations	36
5	Disc	ussion	39
	5.1	Draw Sphere Cells	41
	5.2	Calculate Surface Variation Pixels to Sphere	41
	5.3	lambda values	42
	5.4	Approximation & calculation Errors	42
	5.5	CC3D	42
6	Futu	re work	43
7	Con	clusion	44
Lis	st of A	Abbreviations	V
Lis	st of 1	Tables Tables	vi
Lis	st of F	Figures	vii
Lis	stings	3	ix
Bil	bliogi	aphy	X

Chapter 1

Introduction

1.1 Motivation

One of several requirements regarding complex life is cell adhesion. If different cells would not stick to each other the only living things would be cells. In humans, organs are made of several cells as well as epithels are made of several cells and several layers of cells. This is also the case for the urothelium, which is an epithelium, i.e. a membranous tissue which consists of one or several layers. For the urothelium it is necessary that the cells stick to each other. Otherwise the functions of the urothelium could not be executed and also it would not be able to grow.

There are two types of tumors. One is the benign and the other one is the malignant tumor [1]. The benign tumor is self limited. Thus, it does not invade surrounding tissues as well as it does not spread into other body parts [1]. The malignant tumor on the other hand is not limited in its growth and is able to invade other body parts [1].

Since bladder cancer is one of the most common cancer types among men it is important to understand how and why the cancer is able to grow.

Bladder cancer starts to grow in the urothelium. With the grow and spread, the structure of cells sticking together is changed. In this case, the urothelium is no longer able to completely perform its tasks. In order to understand the urothelium, how and when bladder cancer appears observations of this epithel is necessary.

To understand the functionalities of organs and epithels, in general organisms, observations are essential. For the urothelium this is already done, as there are a lot of different in vitro experiments about the methodology of the urothelium. After an

observation of an epithel or organ is complete, researches are able to predict how the observed organism will react in different situations. To verify these predictions a simulation is necessary. A simulation is an illustration of the reality **REF** but it can also be used to change reality in a for the research specific way to get more knowledge of the epithel, or an organism in general.

A simulation should always be as simple as possible but also not too simple **REF** Otherwise the simulation does not represent the reality. There are several programs with different algorithms for cell simulation. A popular algorithm is the Glazier-Graner-Hogeweg (GGH) model. This model is popular because it is easy to describe how cells interact with each other and it is possible to define constraints for the volume and surface of each cell.

The program CompuCell3D (CC3D) is a simulation program, which uses the GGH algorithm in its simulation. In the moduro project we use CC3D, and with the program we use the GGH algorithm.

The target of the moduro project is to predict under which circumstances bladder cancer occurs and when it is able to grow. Therefore, 16 different morphogenesis models of the urothelium were created. An overview of these models is displayed in table 2.1 at page 12. So far, all 16 models were simulated in 2D for 720 days. The results reveal that some of the models are more realistic and others are less realistic.

Because a cell is a three dimensional organism, a 2D simulation of the urothelium might not give as many aspects as a 3D simulation could do. Thus, the aim of this bachelor thesis is to create a 3D simulation of these 16 different models. With this 3D simulation we hope to receive new insights into the urothelium and how bladder cancer occurs.

1.2 Background

1.2.1 Biology of the Urothelium

Bladder cancer is the 4th most common cancer type in men regarding to everyday-health.com, where every 36st out of 100.000 men gets it [2]. Bladder cancer usually starts with some cells in the bladder, which grow uncontrolled. From these cells, the tumor can spread further into surrounding areas [3]. The most common bladder cancer type is the urothelial carcinoma [3].

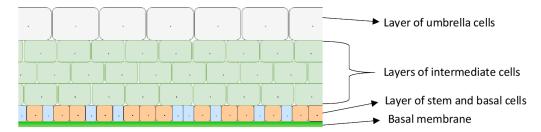


Figure 1.1: A simplified illustration of the urothelium. At the very bottom there is the basal membrane. Above the membrane the stem and basal cells are displayed. The blue cells represent the stem cells, the red cells display the basal cells. Above the layer out of these two cell types, the urothelium contains several layers of intermediate cells. At the top of the urothelium there is an layer of umbrella cells.

The bladder is located in the lower urinary tract and consists of several parts, where the urothelium is one part and coats the bladder [4]. More specifically, it covers the bladder from the renal pelvis to the proximal urethra [5], [6].

Two important tasks of the bladder are the storage and release of urine. To do so the bladder will extend, during the storage, and then shrink again [7]. One task of the urothelium is to form a distensible barrier [4], [8]–[11], which prevents unregulated exchange of ions, solutes, and toxic metabolites between the bladder and the blood [4], [8]–[10]. That the urothelium ensures its barrier function, it has to enlarge and downsize its size. This is done by the largest cells of the urothelium, the umbrella cells. Since the umbrella cells are in direct contact with the bladder, it is their task to change size and form during the grow and shrink process of the bladder. Birder described the urothelium as "... a responsive structure capable of detecting physiological and chemical stimuli and releasing a number of signaling mole-cules." [6]. Another task of the urothelium is to control the movement and passage of macromolecules, ions, water, toxic metabolites and solutes [8], [9]. If the urothelium is damaged, it rapidly generates new cells, to ensure full functionality [5], [8], [9].

To receive a better overview the different cell types are explained in the following paragraph. In figure 1.1 at page 3 a simplified illustration of the urothelium with its different cell types and cell layers is provided.

The umbrella cells, also called superficial cells, are connected directly with the bladder and have an average diameter of 25 up to $250 \,\mu\mathrm{m}$ [5], [9].

Below these cells the intermediate cells are located. With an average diameter of $10 \text{ up to } 20 \,\mu\text{m}$ [5], [9], they are smaller then the umbrella cells. There are at least three and up to five layers of the intermediate cells **REF**

1	1	1	3	3	5	5	5
1	1	1	3	3	3	5	5
2	2	1	3	4	5	5	5
2	2	2	4	4	4	7	7
6	2	4	4	4	4	4	7
6	6	4	4	4	7	7	7
6	6	9	9	9	9	8	7
6	6	9	9	9	8	8	8

Figure 1.2: A square lattice in 2D. The same digits represent one cell $(\sigma(\vec{i}))$, whereas the different colors represent different cell types $\tau(\sigma)$.

The smallest and the most common cells in the urothelium are the basal and stem cells. Those cells have a diameter of up to $10 \, \mu m$ [4], [9].

The urothelium consits of several layers. In the first layer, there are the basal and stem cells. Above them, there are several layers of intermediate cells. On top of the epithelium there is one layer of umbrella cells.

1.2.2 CompuCell3D

CC3D is an open-source program, which provides a simulation environment for multi- or single-cell-based modeling of tissues, organs and organisms [12]. To do so, CC3D uses the GGH model in its simulation. CC3D provides the possibility to create programs for the simulation, e.g. cell growth, mitosis, apoptosis or necrosis scripts, in python, C++ or in CC3DML, which is their own Markup Language. With such programs CC3D allows the user to modify the behavior of the simulation for a specific purpose. CC3D uses the GGH approach, explained in section 1.2.3 at page 5. It allows the user to choose between two cell-lattice types, i.e. a presentation of the pixels or voxels of a cell at a specific position in the simulation field. By default it uses a square-lattice of single pixels for each dimension, an example therefore is displayed in figure 1.2 at page 4. There is also the possibility to use a hexagonal-lattice, where the pixels would be hexagons in two dimensions, or rhombic dodecahedrons in three dimensions. Since the core of a GGH simulation is the effective energy [13], CC3D tries to minimize this effective energy every Monte

Carlo Step (MCS), i.e. a calculation step in the simulation. The basic form for the effective energy is:

$$\mathcal{H}_{boundary} = \sum_{\vec{i}, \vec{j}} J(\tau(\sigma(\vec{i})), (\tau(\sigma(\vec{j})))(1 - \delta(\sigma(\vec{i}), (\sigma(\vec{j}))))$$
(1.1)

This equation is a part of the equation of the GGH model. The formula of the GGH model, and with it this equation, is explained at section 1.2.3 at page 5.

There are two ways to extend this form, it is possible to either add a volume or a surface constraint for each cell. During each MCS an index-copy attempt takes place [13]. Therefore, a pixel is selected, and it is be tried to overwrite a randomly chosen pixel, next to the current pixel, in order to minimize the effective energy. The index copy attempt succeeds and takes place if this index copy attempt decreases the effective energy [13]. Each MCS the program tries to minimize the effective energy with index copy attempts.

1.2.3 Glazier Graner Hogeweg Model

Since there are several formulas and models developed from Glazier and Graner, this subsection briefly describes these models and explains the Cellular Potts Model (CPM) and GGH model.

The GGH model is widely used in biological simulations, since it provides a good flexibility, extensibility and it is easy to use [14]. Glazier and Graner developed a model as an extension of the large-q Potts model, which itself is an extension of the Ising Model, and called it first the Extended Potts Model (EPM). Nowadays, this model is called CPM [14]–[16]. Glazier and Graner extended the CPM in a way that also volume constraints are considered for the hamiltonian, see following form:

$$\mathcal{H}_{CPM} = \sum_{\vec{i}, \vec{j}} J(\tau(\sigma(\vec{i})), (\tau(\sigma(\vec{j})))(1 - \delta(\sigma(\vec{i}), (\sigma(\vec{j}))))$$

$$+ \sum_{\sigma} \lambda_{vol}((\tau)v(\sigma) - V_{target}(\tau(\sigma)))^{2}$$
(1.2)

The hamiltonian of equation 1.2 describes the effective energy for the extension of the CPM model. The first sum describes J of all cells, the adhesion energy between different cells. Therefor, every cell has a specific cell type $\tau(\sigma)$ [15], [16]. Each cell is placed onto a lattice with a spin $(\sigma(\vec{i}))$ for every given dimension [14], [15].

The adhesion energy between cells is only considered if the kroenecker delta is 0. Thus, the surface energy between cells is considered if $\delta(\sigma, \sigma') = 0$ [14]–[19].

With the second sum over all cells the volume of each cell is now considered within the effective energy. The user is now able to set a target volume $V_{target}(\tau(\sigma))$ for each cell, which the cell should have, and a multiplier λ_{vol} for the deviation between the current volume $(\tau)v(\sigma)$ and the target volume. During the simulation this deviation is tried to be kept as small as possible for every cell in order to keep the effective energy as small as possible.

Together with Hogeweg they further developed the created extension of the CPM. The further developed model is called GGH model. The main extension is that the user is now able to add surface area constraints [14]–[16] as well as to use a negative boundary energy [14]. With the surface constraint the equation for the effective energy of the GGH model is:

$$\mathcal{H}_{GGH} = \sum_{\vec{i}, \vec{j}} J(\tau(\sigma(\vec{i})), (\tau(\sigma(\vec{j})))(1 - \delta(\sigma(\vec{i}), (\sigma(\vec{j}))))$$

$$+ \sum_{\sigma} \lambda_{vol}(\tau)v(\sigma) - V_{target}(\tau(\sigma)))^{2}$$

$$+ \sum_{\sigma} \lambda_{sur}(\tau)s(\sigma) - S_{target}(\tau(\sigma)))^{2}$$
(1.3)

In addition to the hamiltonian of equation 1.2 is the surface constraint. It has the same principle as the volume constraint. Thus, the user is able to define a target surface $S_{target}(\tau(\sigma))$ for each cell and a multiplier λ_{sur} for the deviation between the target and the actual surface $(\tau)s(\sigma)$ of each cell. Since the volume and surface constraint are included in the effective energy, it should be possible to use these two parts of the effective energy to shape the cells.

Beside the surface constraint the new model allows the user to model (a): cell growth and proliferation (b): mitosis, i.e. cell division (c): fields, forces and diffusion and (d): chemotaxis and haptotaxis [14].

Glazier et. al. describe their model as:

GGH models define biological structure consisting of the configuration of a set of *generalized cells*, each represented on a *cell lattice* as a domain of lattice sites sharing the same cell index [...], a set of *internal cell states* for each cell [...], and a set of *auxiliary fields* ..." [14].

The GGH model has the advantage that "Initial conditions emulating a particular biological configuration rather than random initial conditions." [14] and it has now biologically motivated properties instead of physically motivated properties [14].

1.3 Objective

The aim of this bachelor thesis is to create a 3D morphogenesis simulation of the urothelium using CC3D. Since the simulation models and python program for a 2D simulation are given and the simulation is done by CC3D the task is to modify the current application, of the 2D simulation, in a way that this program can be used for a 3D simulation of the different models.

Therefore, some parts of the program have to be modified. Some functionalities have to be invented and developed whereas for other functionalities it is enough to modify these.

The result of this bachelor thesis will be presented with an realistic model of the 2D simulations. This model is chosen from the given models in the 2D simulation. The question how much more time the 3D simulation need than the 2D simulation will be covered in this thesis as well. If it is possible to provide a calculation of how much more effort a simulation in three dimensions need it will be included, otherwise an estimation of the more effort is presented.

1.4 Outline

In this chapter the knowledge to understand this bachelor thesis is provided. The next chapter provides the status at which the project was at the beginning of this bachelor thesis. Once the basic knowledge and the state of the art are explained, necessary modifications of the program are revealed. After these modifications are presented, the result of this bachelor thesis is shown. In the last section of this thesis, this work is discussed from different points of view and the conclusion out of this work is presented.

Chapter 2

State of the Art

In this chapter all information about the current project are revealed. At the beginning the properties of the project are explained and later in this chapter the simulation program is presented. The sections 2.5 to 2.9 are explained based on the program of the project, wheras the other sections are presented based on previous work of the project.

2.1 Moduro

In the project moduro stands for 'Modeling of the Urothelium with the GGH approach'.

The department of medical informatics at the university of applied sciences Mannheim and the clinic of urology in cooperation with the medical faculty Mannheim at the university of Heidelberg participate in this project. The aim is to predict how and when bladder cancer arises.

The current moduro project has a stable 2D simulation of 16 different models using CC3D. The simulations are performed by CC3D. During the simulation statistics about the current simulation are written in text files and can be read out by the moduro toolbox.

The project consists of a program and several models, both are written in python. The models include properties of cell behavior. Therefore, the adhesion energy between cells and the possibilities of the new cell types after mitosis is defined in the different models. The application modifies the cell behavior, e.g. it checks when a mitosis takes place, how fast the cell will growth, etc..

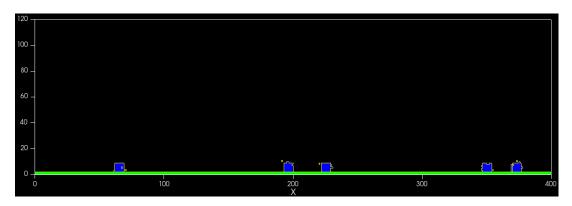


Figure 2.1: Initial state of an 2D simulation of the model SPA/BPCD/IPCD

2.2 Display and Simulation of the Urothelium

In the 2D simulation, the urothel is simulated with a size of $500\,\mu m$ for the x-axis and $150\,\mu m$ for the y-axis. Because, the voxel density, i.e. it describes how many pixels displaying $1\,\mu m$, is 0.8, the size of the urothelium is represented with 400 pixels at the x-axis and 120 pixels at the y-axis. CC3D allows the user to use any simulation size she/he wants, as long as the required hardware can handle the simulation.

It is possible to use CC3D on several cores of the CPU. In the project one core for the simulation is used, because otherwise CC3D splits the simulation field into different grids. As a result race conditions can occur at the edges of these grids [13]. This means it is possible that a cell is in both grids. Thus, one half is in one grid and the other half is in another grid. During the simulation both grids calculate the volume of the cell and both will apply the new volume of their calculation but it is not checked which result has to be used. Therefore, in order to reduce wrong results, it is necessary to avoid race conditions.

The 2D simulations of the urothelium covered 720 days. In the simulations one day is breakdown into 500 MCS. Therefore, the simulation covers 360000 MCS. The simulation duration can be set in the program. Thus, the user define how many MCS are one day and how long the simulation runs. At the first calculation step, MCS 0, the simulation is initialized, i.e. the cells are drawn and placed on the basal membrane. An illustration of an initial simulation is displayed in figure 2.1. Since morphogenesis is simulated, the urothelium is proposed to growth and to proliferate in the given area. An illustration therefor is presented in figure 2.2.

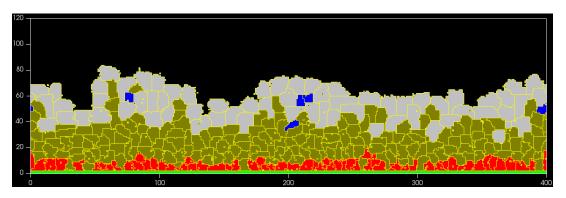


Figure 2.2: 2D Simulation of the model SPA/BPCD/IPCD after 33 days

2.3 Moduro Toolbox

The moduro toolbox is a software tool, which is able to visualize data in order to evaluate a simulation.

Every 12 hours the simulation is checked for its reality. This is done with the fitness functions of section 2.8. The results of these calculations are written in several text files, in a by CC3D created directory of the simulation. Moreover, the amount of cells of the different cell types and the overall amount of cells is saved as well. The moduro toolbox is able to read out the data of the text files and then visualize these. This visualization is done with charts and with a table.

CC3D saves some screenshots, of the simulation field, during the simulation. With these screenshots the moduro toolbox is able to create a video out of these screenshots. To create the video an extra program has to be installed which the toolbox would use.

2.4 Models

The 2D simulation of the urothel provides 16 different models. These models differ mostly in which cell types after the mitosis are created. The project divides the models into two domains, one has the identifier 'SSD' and means that every time a stem cell divides there will be one stem and one basal cell. The second domain is called 'SPA'. In the second domain the stem cell has a probability of 90% that it will be one stem and one basal cells after mitosis. There is also the chance with a probability of 5% that after mitosis of a stem cell there are two stem cells or two basal cells.

For the mitosis of basal cells there are 4 different models. The mitosis model 'BSD'

describes that each basal cell which undergoes mitosis will create one basal and one intermediate cell. The model 'BPA' describes that there is a 5% chance that the basal cell will become two basal or two intermediate cells. There is a 90% probability that the cell become one basal and one intermediate cell. The model 'BPCD', describes that always a basal cell becomes two basal cells during mitosis and if the basal cell is not on the basal membrane it transform into an intermediate cell. The model 'BCD' has the behavior that a basal cell immediately transforms into an intermediate cell if it is not at the basal membrane.

There are two different models how intermediate cells split. One is the 'IPCD', therefor in mitosis an intermediate cell becomes two intermediate cells during. Each MCS it is checked if there is a cell around the intermediate cell. If this is not the case it is transformed into an umbrella cell. The second model is the 'ICD'. In this one, only transformation of the intermediate cells into the umbrella cells happens if the intermediate cell is not enclosed by other cells.

With these mitosis concepts there are 16 different models in the project. These models were created by an earlier version of the project [20] and are displayed in table 2.1 at page 12.

2.5 Adhesion

During morphogenesis the cells not only growth, they also sort themselves. In order for cell sorting there have to be different adhesion values **REF** i.e. how strong two different cells are holding on each other. In the project this is done with a matrix, which every of the 16 model has. Such a matrix was created in an earlier version of the project [20] and is displayed in table 2.2 at page 13.

2.6 Cell properties

In the project the physiology constraints of a cell are included. In an earlier version of the project these were evidenced [20] and are displayed in table 2.3 at page 13. In the program these constraints are converted into pixels and then applied.

In the simulation every cell has several attributes. Moreover, each cell has an cell dictionary, in which additional attributes are stored. The properties regarding the cell type are likely what every cell in general has, e.g. a min- max diameter, a min-

Type	ID	Description	Model
IIs	SSD	Stem cell-like division	SSS
Stem cells			S S S B
	SPA	Stem cell population asymmetry	$p_s = 0.05$ $p_a = 0.90$ $p_s = 0.05$
	BSD	Stem cell-like division in basal cell	B
Basal cells	BPA	Basal cell population asymmetry	$\begin{array}{cccccccccccccccccccccccccccccccccccc$
B	BPCD	Proliferation and contact differentiation of basal cells	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$
	BCD	Only contact differentiation of basal cells	¬ BM B> 1
liate cells	IPCD	Proliferation and contact differentiation of intermediate cells	$ \begin{array}{cccc} & & & M \\ & & & & & \\ & & & & & \\ & & & & & $
Intermediate c	ICD	Only contact differentiation of intermediate cells	M 1> U

Table 2.1: 16 different models derived in the project as there are different ways of proliferation and mitosis are simulated for the different cell types.

Types			BM	S	В	I	U
Medium	Medium M		14	14	14	14	4
Basal membrane	BM		-1	1	3	12	12
Stem cell	S			6	4	8	14
Basal cell	В				5	8	12
Intermediate cell	I					6	4
Umbrella cell	U						2

Table 2.2: Adhesion matrix for a model in the simulation. Smaller values refer to more adhesion and higher values mean less adhesion. The cell type M is the medium cell type, it is by CC3D a specific cell type which is every in the available space in the simulation, and BM is the basal membrane.

Cell type		V_{min}	d_{min}	V_{max}	d_{max}	Volume	Surface
Stem	S	268	8	523	10	perfect	average
Basal	В	381	9	523	10	important	average
Intermediate	I	905	12	1767	15	important	poor
Umbrella	U	1767	15	3591	19	important	poor

Table 2.3: Constraints of a cell. Volumes V in μ m³, diameters d in μ m. The 'Volume' and 'Surface' column describe how the λ_{vol} and the λ_{sur} should be set for each cell type.

max volume, growth in µm per day or the time until apoptosis, i.e. cell death.

The attributes in the cell dictionary are mainly used for decision making. Some values are the current and expected live time, a flag for necrosis and if a cell is inhibited, i.e. enclosed by other cells.

2.7 Events in the simulation

In the simulation program there are several events modeled. These events can occur every calculation step or at a specific MCS in the simulation.

The events which are performed every MCS, except for MCS 0 and every MCS of factor 250, are a) cell growth and b) the check for mitosis c) cell death d) cell transformation and e) cell mutation. The event which occurs every 12 hours, every 250 MCS, is f) urination. These events are presented in detail in the following subsections.

2.7.1 Cell Growth

In the project the maximum possible growth of a cell is calculated and applied. This calculation is used for the relation volume and target volume as well as for the relation volume, surface and target surface. Because, the volume and the surface of a cell is calculated by CC3D and can only be read out, the target volume and target surface are calculated in the program and set.

2.7.2 Mitosis

The verification if a cell divides or growth further is done in every simulation step, except for these with a factor of 250. Therefore, it is possible to define a very specific calculation step at which a cell divides, as there a 500 MCS per day. In order that a cell divides it grows over its maximal size before it divides. The cells which are able to grow and as a result divide are a) stem cells b) basal cells and c) intermediate cells. The umbrella cells grow as well, but they do not split.

2.7.3 Necrosis

If a cell dies, necrosis takes place. In the program a flag in the cell dictionary is set. Every calculation step, except for the MCS of factor 250, the program checks if a cell dies or not. If so, the cell will shrink and as a result disappear.

2.7.4 Mutation

After 2 days, 1000MCS, it is possible that a cell mutates, i.e. it becomes malignant. We simulate the possibility for cells to mutate after 2 days, because otherwise there would not be much cells around the mutated cell. Each cell type has its own probability to become malignant. In the current simulation it is not considered that cells mutate since the probability for each cell type to mutate is 0%.

If a cell mutates, there is also the flag for necrosis set. Thus, the cell shrinks and disappears.

2.7.5 Transformation

A transformation can take place if a basal cell divides and at least one intermediate cell is created. If the intermediate cell is not enclosed by other cells, it immediately will be transformed to an umbrella cell.

2.7.6 Urination

Every 12 hours, every 500 MCS, a urination takes place. The first urination event takes place at MCS 375. This is simulated in a way that randomly 2% of the cells in direct contact with the bladder are washed out. In the program a flag for necrosis is set and the cells will disappear because of the necrosis event.

2.8 Fitness functions

In order to validate the simulated models, there are several functionalities which check if the model is realistic or not. These functions, i.e. part of a program, check every 12 hours, every 250 MCS, if the model is realistic or not. The result of these fitness functions is written into several files, and can be read out by the moduro toolbox.

2.8.1 Arrangement fitness function

The arrangement fitness function ensures that the strata of the simulated urothelium has the correct order [20], i.e. that the first layer on the basal membrane consists only of stem and basal cells **REFS** the next three to five layers consists only of intermediate cells **REFS** and that there is one layer of umbrella cells **REFS**

$$lib = \frac{L - lib}{L} \tag{2.1}$$

$$f_a = 1 - \frac{((1 - L_B) + (1 - L_U) + lib + (1 - L_O)}{4}$$
 (2.2)

In equation 2.1 L refers to the amount of layers in the urothelium. lib describe the amount of layers between the stem and basal cell layer and the umbrella cell layer

at the top of the urotehlium, which include not only intermediate cells. Thus, if the layers between the first and last layer consist only of intermediate cells, the equation 2.1 has the result 0.

In equation $2.2 L_B$ and L_U are boolean values, i.e. they have the value 0 or 1 and represent a false or true value. They are 1 if the first layer of cells consists only of basal or stem cells and if the most upper layer consists only of umbrella cells, otherwise they will be 0. lib is calculated in equation 2.1. L_O presents the optimum amount of layers. It is 1, if the amount of layers in the simulated urothelium is between 3 and 7, otherwise it is 0. The result of equation 2.2 is between 0 and 1, where 0 refers to no reality in the simulation and 1 refers to a perfect simulated urothelium.

2.8.2 Volume fitness function

This function calculates the relative volume regarding the current volume of the different cell types in the urothel. The relative amount of the different cell types should be: stem and basal cell = 10%, intermediate cells = 67% and umbrella cells = 23% considering an average thickness of $85 \mu m$ [20]. Therefor the formula is:

$$f_{V_i} = \frac{1}{4(\frac{V_{Si} - V_{Ii}}{V_{Si}})^2 + 1}$$
 (2.3)

 V_{Si} and V_{Ii} describes the *should* and the actual *is* volume of a specific cell type i [20]. This calculation is done three times. One time for the stem and basal cells, one time for the intermediate cells and one time for the umbrella cells. The results of the three calculations are then further used to determine the relative volume overall.

$$f_V = \frac{f_{V_B} + f_{V_I} + f_{V_U}}{3} \tag{2.4}$$

In this equation f_{V_B} refers to the realtive volume of the stem and basal cells, f_{V_I} presents the relative volume of the intermediate cells and f_{V_U} include the realtive volume of the umbrella cells.

The result of all four calculations are written into a text file and can be read out by the moduro toolbox later.

2.8.3 Overall fitness function

The overall fitness function calculates the total fitness out of the volume and the arrangement fitness function. Therefor the average of both functions is calculated [20]. This calculation is done by the moduro toolbox only. For every calculation step, the arrangement and volume fitness function are calculated the overall fitness is calculated. Therefor the formula is:

$$f(t_i) = \frac{f_V(t_i) + f_a(t_i)}{2}$$
 (2.5)

 t_i describes a specific time point, in MCS, at which this calculation is done. At the end of the simulation the average of the overall fitness function is calculated to determine the reality of this simulation. Therefor, the formula is:

$$f = \frac{1}{e+1} + \sum_{i=0}^{e} f(t_i)$$
 (2.6)

The result of the calculation is between 0 and 1. Where 0 describes no reality at all and 1 presents a perfect realistic simulated urothelium.

2.9 3D functionalaties

The functionalaties of the program are able to be used in 3D. Whenever a part of the program is required to be used in 2D as well as in 3D, it is checked if the simulation field has a third dimension or not. Examples for such functionalities are the fitness functions of the section before or the decision how many stem cells are placed on the basal membrane, which is modified and explained in section 3.4.

Because so far only the 2D simulations were progressed, it is important to keep the functionalaties of the 2D simulation in the program. Doing so it is possible to switch between a 3D and a 2D simulation, as well as the benefits of both simulation types can be observed.

Chapter 3

Methods

This section has the purpose to give an overview over all changes in the project which occurred during this bachelor thesis. These changes include some small improvements of the application as well as how to draw a sphere cell. First minor changes are briefly described, deeper in this section the more effortful changes are explained.

3.1 Lambda multipliers

In the project there were several places, where the multipliers λ_{vol} and λ_{sur} are calculated, but they are only set when a cell is initialized. The multiplier for the volume is set a second time in the necrosis event, if the program models that a cell dies.

There are two additional multipliers, for the surface and volume constraints, saved in within each cell object. Since we are not using these values in the program and they do not influence or set the λ_{vol} or λ_{sur} which CC3D uses, these two values are deleted. Moreover, in one place there were methods to calculate the lambda values. Since these methods are not used in the project and they had a multiplier itself to calculate the multiplier for the specific part of the effective energy, they are deleted as well.

In the project the multipliers λ_{vol} and λ_{sur} are now set each time when a cell is initialized. A second time the multiplier λ_{vol} is set, is if necrosis takes place. Because now the unnecessary methods and values for the multipliers values are removed, no confusion about which constraint values in the program are used will come up.

3.2 Abstract methods

In the program, in several classes there were methods, which are not used. These methods are not used in the class, in which they are written, due to polymorphism. They are used in classes, which inherit from the class where they are written. Polymorphism is a method out of Object Oriented Programming (OOP). Another technique out of OOP is the use of abstract classes and abstract methods. To explain polymorphism, abstract classes or abstract methods in detail, would take too much space out of this bachelor thesis. Thus, the change in the program is explained in the following.

In the project there a several classes which inherit from each other. In these classes there were the same methods implemented. Since it is not required that all classes have to have the same methods, these are redundant methods, as long as they do not differ in their functionality. For such methods it is possible to declare them as an abstract method. An abstract method contains only the method construct, but no functionality. Due to polymorphism the class in which such an abstract method is implemented is used to call this method.

In python there is a library 'abc'. With this library it is possible to declare an abstract method. To do so every class with one or more abstract methods needs to initialize a special class variable. With this variable python is able to recognize that there are abstract methods included.

```
class ModelConfig(object):
    __metaclass__ = ABCMeta
```

Listing 3.1: The initialization of a class variable which is required by python in order to detect abstract methods and abstract classes.

The listing above displays how the class variable has to be initialized in order that abstract methods are recognized. If a class has at least one abstract method, it is an abstract class. In python abstract classes can include implemented methods as well as abstract methods.

```
@abstractmethod
def _createExecConfig(self):
    pass
```

Listing 3.2: Declaration of an abstract method.

The listing above is an example of an abstract method in python. Using abstract methods creates more structure and clarity within the project. Therefore, the ab-

stract declarations in this project created a much better clarity of the structure of the project and of the methods, which were used in the classes.

3.3 Calculation steps until urination

To simulate the urination in the program it was checked if the current calculation step is larger than 250 and a factor of 125. Because every 250 MCS the volume- and arrangement fitness functions are calculated and no other events in the simulation take place, the urination was simulated every 12 hours. The check if the current MCS is a factor of 125 happens with the modulo operator. Thus, the current calculation step is divided by 125 and if the result, as a whole number, is 0 it is a factor. The check when the urination event takes place is modified. The urination event now takes place if the current calculation step is larger than 250 and if the current MCS modulo 125 equals 1. Therefore, the urination event is used every 6 hours, at MCS 376, 501, 626, 751, etc..

3.4 Area of stem cells on the basal membrane

In an earlier version of the project it was evidenced that around 12% of the area of the basal membrane are required to be filled with stem cells in order to have an optimal proliferation during the morphogenesis of the cells [20]. In the project the calculation of the amount of stem cells for two dimensions were correct but without an mathematical evidence.

Since the y-axis is negligible in the calculation of an area, the calculation for two dimensions considers the x-axis and the calculation for three dimensions considers the x- and z-axis. Therefore, in two dimensions the area of the stem cells should be calculated by using the cell diameter. An illustration therefor is displayed in figure 3.1. For three dimensions it is possible to calculate the area of a circle with the formula $A = \pi \cdot r^2$, because a sphere in 2D is a circle, and use this calculation to further determine the amount of stem cells on the basal membrane. An example for the basal membrane and a stem cell in three dimensions is displayed in figure 3.2.

With the following two equations it is possible to calculate the amount of stem cells in two dimensions. $A_{stemcells}$ refers to the area which can be used for stem cells.



Figure 3.1: Considered area to spread the stem cells in 2D with an example of one stem cell placed on the basal membrane. Because only the x-axis is displayed we need to calculate the diameter of a cell.

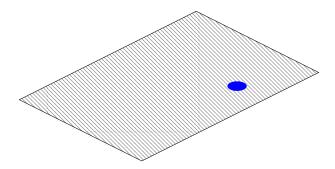


Figure 3.2: Considered area to spread the stem cells in 3D with an example of one stem cell placed on the basal membrane. The hatched area displays the basal membrane and the circle represents one stem cell. This stem cell is a circle, because in the calculation of the amount of stem cells the y-axis is negligible.

Therefore, it is 12% of the basal membrane. C is a constant, which describes 12% of an object. Thus, c = 0.12.

$$A_{stemcells} = xLength \cdot c \tag{3.1}$$

$$Amount_{StemCells} = \frac{A_{stemcells}}{d_{cell}}$$
 (3.2)

Equation 3.1 ensures that only 12% of the given area is used. Formula 3.2 then calculates the amount of stem cells on this given area. Because the result of the calculation is often not a whole number, the result is checked if the first decimal digit is larger or equal than 5 and then it is rounded up or casted, i.e. the decimal digits are cut off.

As table 3.2 displays, it is important to round the result. Otherwise there would be an approximation error and as a result a calculation error.

For three dimensions the equation 3.1 has to be extended, because the z-axis is now also considered in the calculation of the amount of stem cells on the basal membrane. An illustration therefor is figure 3.2. Equation 3.2 has to be modified, because now the area of an circle, instead of the diameter, is used for the calculation. Therefore, the equations to calculate the amount of stem in three dimensions are:

$$A_{stemcells} = (xLength \cdot zLength) \cdot c \tag{3.3}$$

Table 3.1: Possible approximation error by not rounding the result of equation 3.2. The first column describes the length of the basal membrane, the values are in μm . In the second column the result of equation 3.2 is displayed. In the third column the result of the second column is rounded up in the first row. In the second row this result is casted. The fourth column displays how much space the stem cells, in μm , of the rounded or casted result of eqataion 3.2 require. The last column displays the physical space required in percentage to the basal membrane.

	Result of equation 3.2		Area used of stem cells in µm	
200	~ 2.66	3	27	13.5%
200	~ 2.66	2	18	9%

$$Amount_{StemCells} = \frac{A_{stemcells}}{\pi \cdot r^2}$$
 (3.4)

The result of formula 3.4 has to be rounded as well, otherwise the program would include rounding errors. These calculations are now included in the program.

3.5 Target volume and target surface after mitosis

Mitosis of the cell is simulated by CC3D. In the simulation mitosis is simulated as the following: one cell splits into two cells. The cell which splits dies and then two new cells out of the died cell are created.

In the program we specify and check if and when a cell splits. CC3D decides where the cell splits and calculates the volume and surface of the two new cells. In our program we calculate and set attributes of the new cells, e.g. the target volume or the target surface.

The target volume is calculated by dividing the target volume of the cell before mitosis by 2. This value is applied for both new created cells. The problem with this technique is that it is possible that the cell might not split in the middle. In the case of mitosis, setting the target volume of the two created cells without the knowledge of the volume is a source of error. Because the target volume of the two new cells are set without considering the current volume of the cell, it occurred that one of the new cells, after mitosis, had a target volume which was smaller than the current volume.

After mitosis both cells are initialized. Thus, the target volume and target surface is calculated and set. After the initialization of the cells is done, the target volume of

the new cells is set to the target volume divided by 2 of the cell before mitosis.

Because the target volume is calculated out of the current volume of the cell during the initialization process, the calculation of the target volume is removed out of the mitosis event. Now the target volume and surface after mitosis is set only during the initialization process of the new cells.

With this change of the program the case that some cells have a smaller target volume than the current volume disappeared. In the program, the target volume and target surface after mitosis is calculated and set dependent on the of CC3D given volume.

3.6 Approximation Error

Because the project includes conversions from μm into voxels and the amount of pixels, e.g. for the surface of a cell, has to be set as data type integer, i.e. a whole number, it is possible that in some places in the program there are approximation errors. In the project, the values of unit μm are saved with the data type float, i.e. a number with decimal digits. To set these values as a whole number the values are casted into the data type integer.

Whenever a conversion from μm into voxels is done the complete calculation is calculated with decimal digits. After the calculation is done it is verified if the first decimal digit is larger or equal to 5. If this condition is true the result is increased by 1, otherwise not. As a last step the result is casted. This technique has the advantage that calculation errors due to casting are removed, because the cast is the very last step. It is possible that in the program still include some rounding errors, but these can not be removed because CC3D requires the amount of voxels as a whole number and the calculation is with decimal digits.

In the function displayed below, the cast and the rounding of the result are done in the last step of the calculation.

```
def calcVoxelVolumeFromVolume(self, volume):
    r = (3 * volume / (4.0 * PI)) ** (1.0 / 3.0) # Radius of a sphere with
        known volume.
    rDimension = r * self.voxelDensity
    if self.dimensions == 2:
        return int(self.__truncate(PI * (rDimension ** 2))) # Area of a circle.
    else:
        result = 4.0 / 3.0 * PI * (rDimension ** 3)
        if result % 1.0 >= 0.5:
            result += 1
```

return int(result)

Listing 3.3: Function to calculate the volume of a sphere in voxels out of a given physical volume. First out of the given physical volume the radius is calculated. Then it is converted into the voxel unit. Next the volume of the voxel sphere is calculated and as last step the result is rounded and casted.

One use of the displayed function is to calculate the voxel volume out of the physical volume of a cell. In the simulation a minimum and a maximum volume for each cell type is calculated. These values are used in the calculation to determine if mitosis takes place or not.

For the basal cell the minimum volume is $381\,\mu\mathrm{m}$ and the maximal volume is $523\,\mu\mathrm{m}$. In table 2.3 at page 13 the constraints of the different cell types in $\mu\mathrm{m}$ are displayed. In the table 3.2 three different possible calculations of the conversion of a volume in $\mu\mathrm{m}$ into a volume in voxels are presented.

Table 3.2: Three different ways to calculate the voxel volume out of a given physical volume. The first column describes the physical volume in μm . In the second column the radius in μm out of the volume is calculated. Next, the radius is used as it is, casted or rounded up. In the fourth column the exact result of the volume in voxel is presented and in the last column the rounded result of the voxel volume is displayed.

Volume µm	in	radius in µm	radius used in further calcu- lation	not rounded result in vx	rounded result in vx
381		4.49	4.497	1285.67	1286
381		4.49	4	904.77	905
381		4.49	5	1767.15	1767

In the table, the first row calculates the voxel volume as it is done in project right now. Thus, rounding and casting is the last step in the calculation. In the second raw the radius is calculated and casted. Then this casted radius is used for the further calculation. The third row calculates the radius and rounds it immediately. This rounded radius is then used for the further calculation.

As the table displays, there is a significant difference in all three results. Because such a calculation is used to determine when a cell splits as well as it calculates the growth per MCS, it influences the result of a simulation. Thus, it is important to round and cast the result as very last step in the calculation.

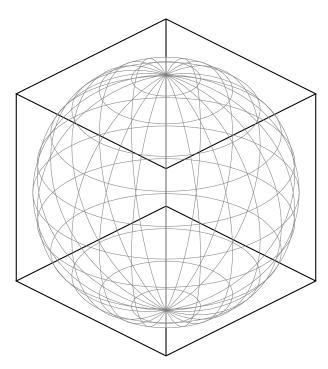


Figure 3.3: A cuboid layed around a sphere

3.7 Draw Sphere Cells

Since a sphere as a cell is an approximation to a cell in the urothelium, it is a possible technique to draw and further simulate a cell as a sphere.

To be able to draw a sphere out of voxels it is required that a cuboid lays around the sphere, as it is displayed in 3.3. The cuboid has to be at least as large as $2 \cdot radius$ of the sphere. In addition, the cuboid should not be larger than necessary, otherwise there would be unnecessary computable cost. The illustration in figure 3.4 at page 27 display the perfect size of a square and a circle. These 2D objects are chosen to display the boundaries of a circle in a square as well as the boundaries would be in three dimensions.

To be able to draw a cell as a sphere, CC3D has to allow the user to draw several different voxels in the simulation field, all containing to one cell. Since CC3D allows the user to draw several pixels containing to one cell, a solution for this problem is possible.

Because we are using the square lattice, the cuboid is filled with voxels. To be able to calculate every point within the sphere, the cuboid and the sphere are required to

have the same center **REF** In this case the equation 3.5 provides a mechanism in which every point within the sphere can be calculated.

$$\sqrt{(x_r - x_0)^2 + (y_r - y_0)^2 + (z_r - z_0)^2} <= radius$$
 (3.5)

In the equation above x_r , y_r and z_r describe the current point of each of the three axis and x_0 , y_0 and z_0 describe the center of the sphere. If the distance of the current x, y, z coordinate is smaller or equal to the radius the current point is within the sphere, otherwise it would be outside of the sphere. Because a voxel itself contains several points, this equation, in this case, has the weakness that only one point is considered. Thus, only one point of the voxel is considered in the decision if the complete voxel is within the sphere or not. Since every voxel is a cuboid itself, the length of all corners are the same. This fact can be used to increase the accuracy of the equation above.

In the program it is possible to calculate the corner length of a voxel. Thus, it is possible to decide if the center of a voxel is at the inside or outside of a sphere. How the center of a voxel is calculated is explained in the following.

Since the radius of the sphere is known, whether it is known or it is calculated out of the volume or the area of a sphere, the diameter of the cuboid can be calculated. Since the voxel density is known in the simulation, the corner length of each voxel can be calculated as it is displayed in equation 3.6.

$$c = \frac{2 \cdot radius}{voxeldensity} \tag{3.6}$$

In this equation c describes the corner length of a voxel. With this corner length it is now possible to calculate the center of a voxel and use this center further to decide if the voxel is inside or outside of the sphere. To receive the center of a voxel the calculation has to be done for every of the three axes. The following three formulas in equation 3.7 display such calculations.

$$x_{c} = x_{r} + \frac{x_{r+c} - x_{r}}{2}$$

$$y_{c} = y_{r} + \frac{y_{r+c} - x_{y}}{2}$$

$$z_{c} = z_{r} + \frac{z_{r+c} - x_{z}}{2}$$
(3.7)

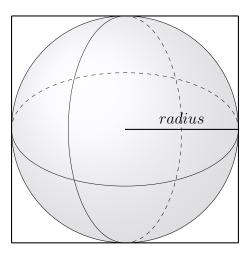


Figure 3.4: A cuboid with minimal size layed around a sphere

In this equation x_r , y_r and z_r describe the start point and x_{r+c} , y_{r+c} and z_{r+c} describe the end point of the voxel. With the calculations of equation 3.7 it is now possible to consider the center of a voxel in the decision if the voxel is in- or outside the sphere, as it is displayed in equation 3.8.

$$\sqrt{((x_c - x_0)^2 + (y_c - y_0)^2 + (z_c - z_0)^2} \le radius$$
 (3.8)

To draw a cell as a sphere a new function had to be created. This function is named addSphereCell as it is displayed below. In the method all required points, the startand end points as well as the centers, of all three axis are given in μm . Thus, these points as well as the radius, which is also given to the function, and the steplength, i.e. the corner length of a voxel, are converted into the voxel unit.

With all necessary points in the voxel unit it is now possible to iterate over all three axis and check for every voxel, if it is included in the sphere or not.

```
def _addSphereCell(self, typename, xPos, yPos, zPos, radius, steppable):
    cell = steppable.newCell(typename)
    xStart = self.execConfig.calcPixelFromMuMeter(xPos - radius)
    x0 = self.execConfig.calcPixelFromMuMeter(xPos)
    xEnd = self.execConfig.calcPixelFromMuMeter(xPos + radius)
    yStart = self.execConfig.calcPixelFromMuMeter(yPos - radius)
    y0 = self.execConfig.calcPixelFromMuMeter(yPos)
    yEnd = self.execConfig.calcPixelFromMuMeter(yPos + radius)
    zStart = self.execConfig.calcPixelFromMuMeter(zPos - radius)
    z0 = self.execConfig.calcPixelFromMuMeter(zPos)
    zEnd = self.execConfig.calcPixelFromMuMeter(zPos + radius)
    radiusPx = self.execConfig.calcPixelFromMuMeter(zPos + radius)
```

Listing 3.4: Function to draw a cell as a sphere. First all required points for the calculation are converted into the voxel unit. Then over each axis of the cuboid it is iterated. During these iterations for each voxel the distance to the center of the cuboid and sphere is calculated and then it is checked if the voxel is within the sphere or not. If the voxel is a part of the sphere it will be added to the sphere.

With this created function it is now possible to draw a cell as a sphere, as it is displayed at figure xy.

To draw the cell as a sphere is the first step to have sphere cells in the simulation. Two major parts of the simulation are the growth and the mitosis of the simulation. In order that the cells are able to stay as a sphere it is required to adjust the volume and surface calculation as well.

3.8 Growth of a sphere cell

Since in the project morphogenesis of the urothelium is simulated, the drawn sphere cells are required to grow.

In the simulation the current volume and surface of a cell is calculated by CC3D. Every user is able to influence these values by setting the target volume and target surface and the proper multiplier, λ_{vol} and λ_{sur} , for the specific part of the effective energy. Because the goal of the simulation is to minimize the effective energy, the cell will change the current volume and surface in the direction of the set target volume and target surface.

To model the growth of a cell, the growth per day of the volume for the different cell types is set. Each calculation step the growth of one MCS is calculated and applied, 500 calculations steps are one day. After the growth of the volume is calculated and set as new target volume of the specific cell, a new target surface depending on the target volume is calculated and set.

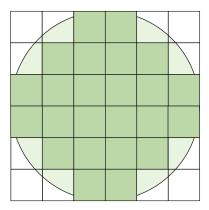


Figure 3.5: A circle in a possible pixel presentation. All small squares present pixels. The colored squares present the pixels, which are included in the pixel presentation of the circle.

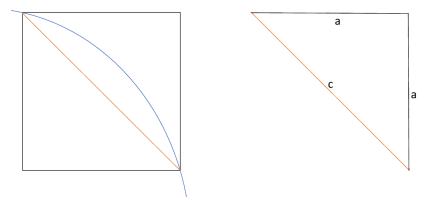


Figure 3.6: The left part of the illustration displays a pixel at the surface, in which the circle goes through. The blue line represents the circle. As an approximation to the surface line a diagonal line in the square is drawn. With this diagonal line an isosceles, right ancular triangle appears and it is possible to calculate the additional surface of one pixel. This isosceles, right ancular triangle is displayed at the right side of this figure.

In the program the target surface is calculated out of a real sphere and then multiplied by a factor. Since the surface of a sphere of voxels is larger than the surface of a real sphere, it is necessary to find the correct factor, by which the surface of a sphere has to be multiplied, in order to calculate the surface of a sphere of voxels.

For simplicity reasons, a first approximation of the factor is calculated in 2D. Thus, a circle and a square filled with pixels are used. An example therefor is displayed in figure 3.5.

Figure 3.6 displays a approximation, with which it is possible to calculate the factor for the surface.

In figure 3.6 the left square is considered as a pixel, it could be any pixel of the figure 3.5 in which the circle goes through. Thus, the sides of the square are the same. The blue line represents the circle, in a way it could go through the pixel. An

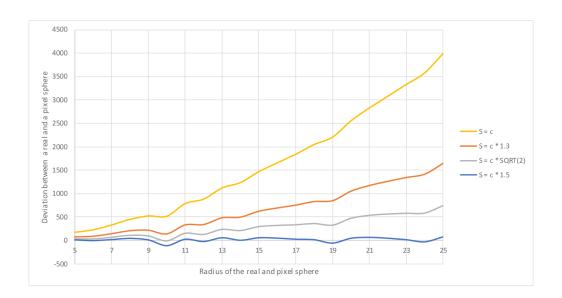


Figure 3.7: Deviation between the surface of sphere of voxels and a real sphere. Each deviation is calculated with a different factor of the surface of the real sphere. S refers to the surface of the sphere of voxels and c refers to the surface of a real sphere. Thus, $c=4\cdot\pi\cdot r^2$. At the x-axis the radius of the sphere and the sphere of voxels is displayed. At the y-axis the deviation between both surfaces is represented.

approximation is to insert a diagonal line. With this diagonal line an isosceles, right ancular triangle is created as it is displayed at the right side of figure 3.6.

The following formula provides a way to calculate the sides of the isosceles, right ancular triangle.

$$c^2 = 2 \cdot a^2 \tag{3.9}$$

This formula can be used to calculate the side a, as it is displayed in the following equation.

$$a = \sqrt{\frac{c^2}{2}}$$

$$a = \frac{c \cdot \sqrt{2}}{2}$$
(3.10)

Since the surface of a pixel has two corner sides, the equation to calculate the surface of a pixel sphere is

$$S = 2a$$

$$S = c \cdot \sqrt{2}$$
(3.11)

This formula applies to all pixels, which are at the surface of the pixel sphere, c is considered as the surface of the circle. Thus $c = \pi \cdot r^2$.

The calculated factor $\sqrt{2}$ was tested for a sphere surface. Because, this is an approximation to the surface of a pixel sphere, the tenth part beside this factor are also tested mathematically. As figure 3.7 displays the approximation with factor $\sqrt{2}$ still has some deviation. Almost no deviation between the surface of a sphere and a surface of a sphere of voxels is with a factor of 1.5. Thus, this factor is applied for the calculation of the target surface.

3.9 Calculation of the volume and surface sites of a voxel sphere

In order to calculate the voxel volume and the surface sites of a voxels sphere on our own, I created an algorithm for this problem. Otherwise for every measurement of the volume and surface of the sphere of voxels a new simulation in CC3D had to be started.

The algorithm requires the radius of the sphere, this radius has to be a whole number. For the tenths part between two whole numbers the result of the algorithm and CC3D deviates. Since a sphere and a cuboid are both symmetrical, it is possible to split both into 8 pieces. Doing so only one eighth of the of the cuboid has to be calculated in order to determine the volume and the surface of the sphere.

To calculate which voxels are in the voxel sphere, the algorithm creates a x, z, y matrix. In the following calculations and decisions the center of each voxel is used. For every x,z voxel in the eighth of the cuboid the y constraint of the circle is calculated. Next, every voxel at the x,z coordinate is checked if the center of the voxel is within this constraint or not. Thus, every voxel at a x,z coordinate is checked if its center is within the sphere or not. This is repeated until every x,z coordinate of the eighth of the cuboid is calculated. Every time a voxel is within the sphere, it will be marked in the matrix.

To receive the volume of the voxel sphere all marked entries in the created matrix are count. Then, this result is mirrored for all three axis. Thus, the amount of marked voxels in the matrix is multiplied by 8, which equals to one multiply by 2 for each axis. Therefore $V = ((Voxels \cdot 2) \cdot 2) \cdot 2$. To receive the surface sites of the voxels at the surface, every marked entry is checked if the voxel at x+1, z+1 or the y+1 is either out of bounce, i.e. if it is outside the cuboid, or outside the sphere. Every of the three conditions is checked for every voxel within the surface. If one condition is true, the amount of surface sites is increased by 1. In the end the result,

like the volume, has to multiplied by 8, in order to mirror the result for all three axis.

Results

This chapter present the results of the bachelor thesis. Because a lot of changes influence the simulation but are not observable, only visible results are mentioned in this chapter. These visible results are presented with a voxel density of 1. Therefore $1\,\mu m$ equals 1 voxel.

4.1 Draw sphere cells

With the created method of section 3.7 the program is now able to draw a cell as sphere out of voxel, as it is displayed in figure 4.1 and 4.2.

Since voxels, cuboids, are used in the 3D simulation to draw a cell as a sphere, it is not possible to draw a perfectly round sphere. This problem is displayed with an circle and a square in picture 3.5 at page 29. The drawn cells are as spherish as possible in the simulation with the use of voxels.

Figure 4.1 displays two independent drawn cells with a radius of $5\,\mu\mathrm{m}$ and $9\,\mu\mathrm{m}$. These cells show that there are a lot of edges in the sphere. As the radius increases, the sphere shape of the cell gets more detailed, as it is displayed in figure 4.2. In this figure two cells are drawn idepently with a radius of $14\,\mu\mathrm{m}$ and $23\,\mu\mathrm{m}$. With the increase of the radius, the deviation of the surface increases as well, as it is shown in figure 3.7. This might be a result as the surface of a sphere and a cuboid, with the daimeter $2 \cdot r$, deviates more as r increases.

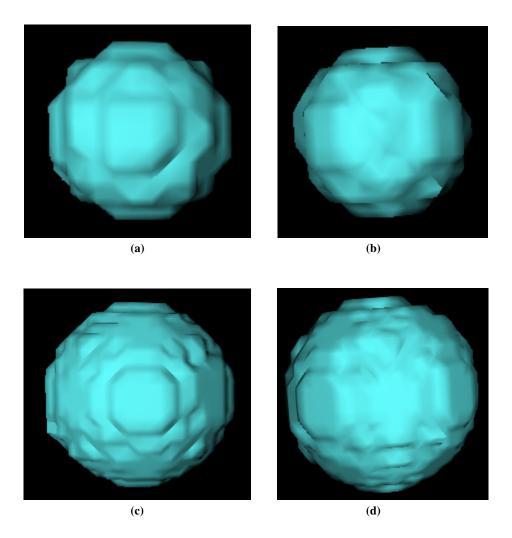


Figure 4.1: A single cell drawn into the simulation field. The radius of the cell of (a) and (b) is 5 and the radius of the cell of (c) and (d) is 9. Pictures (a) and (c) are with the front view, whereas the pictures (b) and (d) have an view angle of around 45 degree. The color of the cell is chosen in a way that more details are visible.

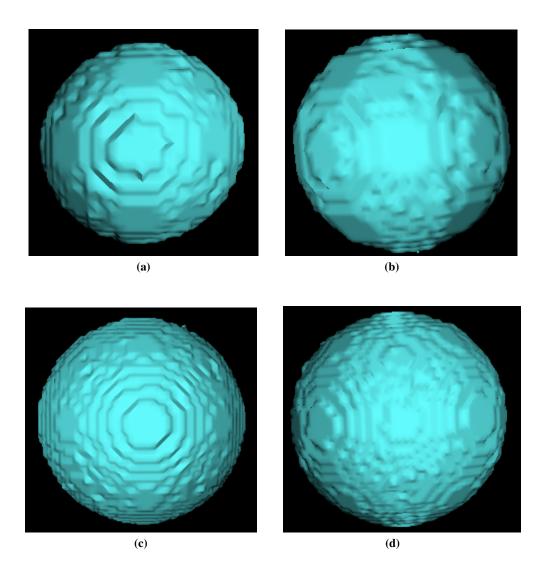


Figure 4.2: A single cell drawn into the simulation field. The radius of the cell of (a) and (b) is 14 and the radius of the cell of (c) and (d) is 23. Pictures (a) and (c) are with the front view, whereas the pictures (b) and (d) have an view angle of around 45 degree. The color of the cell is chosen in a way that more details are visible.

4.2 Grow sphere cells

In section 3.8 the factor for the calculation of the surface of the cell was evidenced best to be 1.5. With this factor it should be possible to let the sphere cell grow as a sphere. To test the growth of the cell, one single cell was placed in the simulation field. As the cell growed during the simulation the volume and surface values were rad out of the command line and compared to the documented values of a drawn cell.

To let a cell grow as a sphere does not work. Even the volume and surface values, calculated by CC3D, and the target volume and target surface values, calculated by the program, meet the values of drawn sphere cells with a small deviation *of an maximum deviation up to 50*.

Figures 4.3 and 4.4 display two examples of a single cell placed in the simulation field. It is observable that drawn sphere cells with a smaller radius become a cubish shape faster. In figure 4.3 the cell has a cubish form after 50 MCS where the cell in figure 4.4 becomes cubish after 750 MCS. This might be a result that the larger sphere cell has more detail than the cell with a smaller radius.

4.3 Simulations

Since no simulations are completed with three dimensions, several 3D simulations are presented. The models, which are simulated, are the most realistic models which were evidenced in an earlier version of the project [20]. The specific models are...

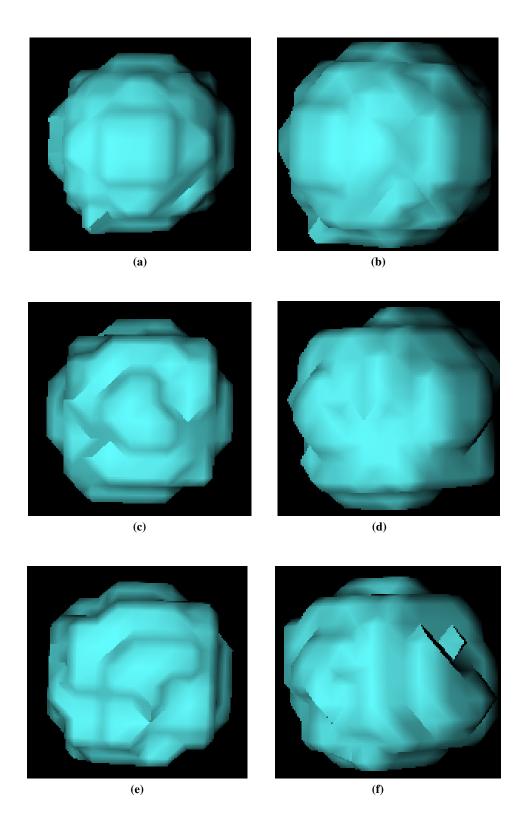


Figure 4.3: A sphere cell, with a radius of $5\,\mu\mathrm{m}$ and a voxel density of 1, as it grows. Images (a), (c) and (e) are the front view of the cell and figures (b), (d) and(f) have around a 45 degree angle of the front. Figure (a) and (b) are at MCS 0, images (c) and (d) at calculation step 50 and figures (e) and (f) present MCS 250.

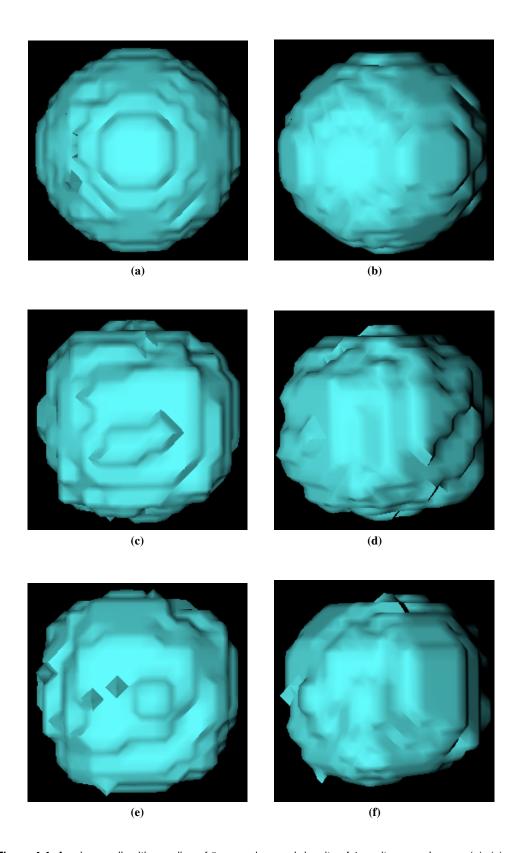


Figure 4.4: A sphere cell, with a radius of $9\,\mu\mathrm{m}$ and a voxel density of 1, as it grows. Images (a), (c) and (e) are the front view of the cell and figures (b), (d) and(f) have around a 45 degree angle of the front. Figure (a) and (b) are at MCS 0, images (c) and (d) at calculation step 750 and figures (e) and (f) present MCS 1250.

Discussion

This chapter servers to discuss the developed solutions and the results from different aspects. Problems, which occured during the bachlor thesis, are presented as well.

To draw a cell as sphere out of voxels is one way to get the desired spherish shape of the cell. Sadly to keep the shape of the cell during the growth process does not work. Since for a given volume and a given surface there are all kinds of shapes CC3D does not know which shape the cell should have. It does not explain why every time the cells get a cubish form during the growth process. It might be a result of the square lattice, which is used in the simulation. There are several research papers, which use CC3D for their simulation, sadly it is never explained how the desired cell form is achieved neither which lattice type is used. So far, it is not possible to know if there is an mistake in the simulation or if the cells get a cubish form due to the square lattice.

It is possible that a cell reaches a desired volume and surface, by only setting the terms for the volume and surface of the effective energy. The desired values can be reached faster if a high multiplier, λ_{vol} and λ_{sur} , is set. This has the disadvantage that a volume or an surface constraint might weigh more than the term of the adhesion in the effective energy. If the specific multiplier has a small value, it might be possible that it takes a long time until the desired shape is reached. In addition to the volume and surface constraint comes the adhesion. Adhesion influences the shape of two or more cells, as it describes how strong the stick to each other. If the adhesion is set too high the cells infiltrate each other but if it is too low the cells may not stick to each other as they change their shape. During the simulations it was observable that some cells of different cell types want to infiltrate each other. This is a result of to high adhesion. Which adhesion values has which effects is

not explained in the literature. This might be a result that it is possible to simulte a lot of different cell types, which may have different behavior at different adhesion values. Since there is no literature about the adhesion of cells by using CC3D it has to be tried out during the simulations. To find the correct adhesion values between cell types has a huge effect on the simulation. Therefore, it might is the only chance to find the correct values, for the desired behavior of the simulation, by trying different adhesion values in different simulation runs. This does not explain why the used adhesion values are not explained. For further research it is important that informations like the adhesion values or about the shape of a cell are shared. It could be result that the published papers are constrained in the length.

In the simulations the voxel density should be set to 1. In this case, in CC3D, one 1 µm is presented by one voxel. If the voxel density is higher than one voxel represents less µm. Since the simulations requires a lot of more time if the voxel density is higher and the deviation between a cell as a sphere out of voxels and a real sphere is larger it should be tried to keep to voxel density as small as possible but also not to small. Why the deviation of a cell and a real sphere grows so much more for a higher voxel density is a puzzle. On one hand the deviation grows with the radius, with more voxels. Since with a higher voxel density there are also more voxels to display it might is a result that CC3D uses more voxels to display the cell. On the other hand the volume and surface values of the drawn cell are that high that for the surface of a real sphere a factor of 6 would give an approximation to the surface of the drawn cell. This maybe is result that CC3D does not know what voxel density is used. Thus, it does not know that a voxel represent not as much physical distance as a voxel used with lower voxel density. As it is displayed in figure 5.1 the volume of a real sphere and of a drawn sphere cell in CC3D, drawn in a simulation with a voxel density of 2, is way to far apart as this voxel density could be used for a realistic simulation of the urothelium. It is interesting to observe that the volume of a sphere cell increases this fast, whereas the surface of a sphere grows in an almost linear way for an increasing radius. Since in CC3D in a 3D simulation the volume refers to a physical volume and the surface refers to a physical surface it might be that one voxel still represent 1 µm, even it should be only 0.5 µm. This might explain the results, as they are similar to the results of $2 \cdot r$ if the voxel densitive equals 1.

- take a look a biocellion (no race conditions) and morphois
- use chemical field for growth

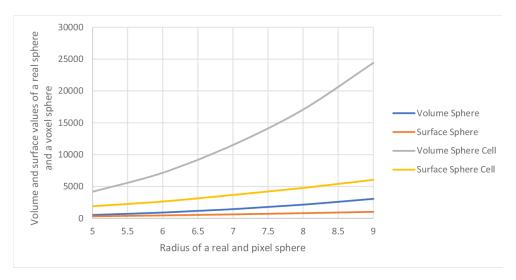


Figure 5.1

- Random Walker algorithm
- kugel oberfläche mit pyramiden berechnen (problem wir haben immernoch pixel und square lattice)

archimedes volumen eines kugelschnittes

5.1 Draw Sphere Cells

It is also possible to use only the effective energy

5.2 Calculate Surface Variation Pixels to Sphere

Two possibilities to design the algorithm

- check the center of each pixel for the radius, is within the radius or not
- calculate the y-values of the radius for an x-value of the center of several pixels -> decide dependen on the result of the y-values if the specific pixel is counted or not (if the center of the pixel is within or outside the radius)

5.3 lambda values

This was the only way to do it

5.4 Approximation & calculation Errors

Find a more elegant way to do it. Floating point arithmetic

5.5 CC3D

- CC3D sucks
- 400vx · 400vx · 400vx = 107171875vx CC3D has its problem and crashes sometimes on 4GB RAM
- CC3D does not empty the used RAM after a simulation -> after several simulations the computer will run out of RAM an CC3D crashes
- no Debug -> testing with print commands at command line

Future work

Conclusion

List of Abbreviations

GGH Glazier-Graner-Hogeweg

CC3D CompuCell3D

MCS Monte Carlo Step

CPM Cellular Potts Model

EPM Extended Potts Model

OOP Object Oriented Programming

List of Tables

2.1	16 different models derived in the project as there are different ways of proliferation and mitosis are simulated for the different cell types.	12
2.2	Adhesion matrix for a model in the simulation. Smaller values refer to more adhesion and higher values mean less adhesion. The cell type M is the medium cell type, it is by CC3D a specific cell type which is every in the available space in the simulation, and BM is	
	the basal membrane	13
2.3	Constraints of a cell. Volumes V in μm^3 , diameters d in μm . The 'Volume' and 'Surface' column describe how the λ_{vol} and the λ_{sur}	
	should be set for each cell type	13
3.1	Possible approximation error by not rounding the result of equation 3.2. The first column describes the length of the basal membrane, the values are in μm . In the second column the result of equation 3.2 is displayed. In the third column the result of the second column is rounded up in the first row. In the second row this result is casted. The fourth column displays how much space the stem cells, in μm , of the rounded or casted result of equation 3.2 require. The last column displays the physical space required in percentage to the	
	basal membrane.	22
3.2	Three different ways to calculate the voxel volume out of a given physical volume. The first column describes the physical volume in μm . In the second column the radius in μm out of the volume is calculated. Next, the radius is used as it is, casted or rounded up. In the fourth column the exact result of the volume in voxel is presented and in the last column the rounded result of the voxel	
	volume is displayed.	24

List of Figures

1.1	A simplified illustration of the urothelium. At the very bottom there is the basal membrane. Above the membrane the stem and basal cells are displayed. The blue cells represent the stem cells, the red cells display the basal cells. Above the layer out of these two cell types, the urothelium contains several layers of intermediate cells. At the top of the urothelium there is an layer of umbrella cells A square lattice in 2D. The same digits represent one cell $(\sigma(\vec{i}))$, whereas the different colors represent different cell types $\tau(\sigma)$	3
2.1 2.2	Initial state of an 2D simulation of the model SPA/BPCD/IPCD 2D Simulation of the model SPA/BPCD/IPCD after 33 days	9 10
3.1	Considered area to spread the stem cells in 2D with an example of one stem cell placed on the basal membrane. Because only the x-axis is displayed we need to calculate the diameter of a cell Considered area to spread the stem cells in 3D with an example of one stem cell placed on the basal membrane. The hatched area displays the basal membrane and the circle represents one stem cell.	21
	This stem cell is a circle, because in the calculation of the amount of stem cells the y-axis is negligible	21
3.3 3.4	A cuboid layed around a sphere	25 27
3.5	A circle in a possible pixel presentation. All small squares present pixels. The colored squares present the pixels, which are included	
3.6	in the pixel presentation of the circle	29
	side of this figure	29

List of Figures

3.7	Deviation between the surface of sphere of voxels and a real sphere. Each deviation is calculated with a different factor of the surface of the real sphere. S refers to the surface of the sphere of voxels and c refers to the surface of a real sphere. Thus, $c = 4 \cdot \pi \cdot r^2$. At the x-axis the radius of the sphere and the sphere of voxels is displayed. At the y-axis the deviation between both surfaces is represented	30
4.1	A single cell drawn into the simulation field. The radius of the cell of (a) and (b) is 5 and the radius of the cell of (c) and (d) is 9. Pictures (a) and (c) are with the front view, whereas the pictures (b) and (d) have an view angle of around 45 degree. The color of the cell is chosen in a way that more details are visible	34
4.2	A single cell drawn into the simulation field. The radius of the cell of (a) and (b) is 14 and the radius of the cell of (c) and (d) is 23. Pictures (a) and (c) are with the front view, whereas the pictures (b) and (d) have an view angle of around 45 degree. The color of the	J- 1
4.3	cell is chosen in a way that more details are visible A sphere cell, with a radius of $5\mu m$ and a voxel density of 1, as it grows. Images (a), (c) and (e) are the front view of the cell and figures (b), (d) and(f) have around a 45 degree angle of the front. Figure (a) and (b) are at MCS 0, images (c) and (d) at calculation	35
4.4	step 50 and figures (e) and (f) present MCS 250	37
5.1		41

Listings

3.1	The initialization of a class variable which is required by python in	
	order to detect abstract methods and abstract classes	19
3.2	Declaration of an abstract method	19
3.3	Function to calculate the volume of a sphere in voxels out of a given	
	physical volume. First out of the given physical volume the radius	
	is calculated. Then it is converted into the voxel unit. Next the	
	volume of the voxel sphere is calculated and as last step the result	
	is rounded and casted	23
3.4	Function to draw a cell as a sphere. First all required points for the	
	calculation are converted into the voxel unit. Then over each axis	
	of the cuboid it is iterated. During these iterations for each voxel	
	the distance to the center of the cuboid and sphere is calculated and	
	then it is checked if the voxel is within the sphere or not. If the	
	voxel is a part of the sphere it will be added to the sphere	27

Bibliography

- [1] N. J. Poplawski, U. Agero, J. S. Gens, M. Swat, J. A. Glazier, and A. R. A. Anderson, "Front instabilities and invasiveness of simulated avascular tumors", *Bulletin of Mathematical Biology*, vol. 71, no. 5, pp. 1189–1227, Jul. 2009. DOI: 10.1007/s11538-009-9399-5. [Online]. Available: https://doi.org/10.1007/s11538-009-9399-5.
- [2] [Online]. Available: https://www.everydayhealth.com/bladder-cancer/guide/.
- [3] [Online]. Available: https://www.cancer.org/cancer/bladder-cancer/about/what-is-bladder-cancer.html.
- [4] M. Lazzeri, "The physiological function of the urothelium—more than a simple barrier", *Urologia internationalis*, vol. 76, no. 4, pp. 289–295, 2006. DOI: https://doi.org/10.1159/000092049.
- [5] T. Yamany, J. Van Batavia, and C. Mendelsohn, "Formation and regeneration of the urothelium", *Curr Opin Organ Transplant*, vol. 19, no. 3, pp. 323–330, Jun. 2014. DOI: 10.1097/MOT.000000000000084.
- [6] L. A. Birder, "More than just a barrier: Urothelium as a drug target for urinary bladder pain", *American Journal of Physiology-Renal Physiology*, vol. 289, no. 3, F489–F495, 2005, PMID: 16093424. DOI: 10.1152/ajprenal.00467.2004. eprint: http://www.physiology.org/doi/pdf/10.1152/ajprenal.00467.2004. [Online]. Available: http://www.physiology.org/doi/abs/10.1152/ajprenal.00467.2004.
- [7] A. A. Karl-Erik Andersson, "Urinary bladder contraction and relaxation: Physiology and pathophysiology", *Physiological Reviews*, vol. 84, no. 3, pp. 935–986, 2004, PMID: 15269341. DOI: 10.1152/physrev.00038.2003. eprint: http://www.physiology.org/doi/pdf/10.1152/physrev.00038.2003. [Online]. Available: http://www.physiology.org/doi/abs/10.1152/physrev.00038.2003.

- [8] G. Apodaca, "The uroepithelium: Not just a passive barrier", *Traffic*, vol. 5, no. 3, pp. 117–128, 2004. DOI: 10.1046/j.1600-0854.2003.00156.x. [Online]. Available: http://dx.doi.org/10.1046/j.1600-0854.2003.00156.x.
- [9] S. N. A. Puneet Khandelwal and G. Apodaca, "Cell biology and physiology of the uroepithelium", *American Journal of Physiology-Renal Physiology*, vol. 297, no. 6, F1477–F1501, 2009, PMID: 19587142. DOI: 10.1152/ajprenal.00327.2009. eprint: http://www.physiology.org/doi/pdf/10.1152/ajprenal.00327.2009. [Online]. Available: http://www.physiology.org/doi/abs/10.1152/ajprenal.00327.2009.
- [10] S. A. Lewis, "Everything you wanted to know about the bladder epithelium but were afraid to ask", *American Journal of Physiology-Renal Physiology*, vol. 278, no. 6, F867–F874, 2000, PMID: 10836974. DOI: 10.1152/ajprenal.2000.278.6.F867. eprint: https://doi.org/10.1152/ajprenal.2000.278.6.F867. [Online]. Available: https://doi.org/10.1152/ajprenal.2000.278.6.F867.
- [11] H. J. L. W. R. Cross I. Eardley and J. Southgate, "A biomimetic tissue from cultured normal human urothelial cells: Analysis of physiological function", *American Journal of Physiology-Renal Physiology*, vol. 289, no. 2, F459–F468, 2005, PMID: 15784840. DOI: 10.1152/ajprenal.00040.2005. eprint: http://www.physiology.org/doi/pdf/10.1152/ajprenal.00040.2005. [Online]. Available: http://www.physiology.org/doi/abs/10.1152/ajprenal.00040.2005.
- [12] [Online]. Available: http://www.compucell3d.org/.
- [13] M. H. Swat, J. Belmonte, R. W. Heiland, B. L. Zaitlen, J. A. Glazier, and A. Shirinifard, *Introduction to computell3d v3.7.4*, 2017. [Online]. Available: www.computell3d.org/BinDoc/cc3 d_binaries/Manuals/Introduction_To_CompuCell3D_v.3.7.4.pdf.
- [14] J. A. Glazier, A. Balter, and N. Poplawski, "Magnetization to morphogenesis: A brief history of the glazier-graner-hogeweg model", Single-Cell-Based Models in Biology and Medicine, p. 79, 2007. [Online]. Available: https://www.researchgate.net/profile/Ariel_Balter/publication/227073495 _Magnetization_to_Morphogenesis_A_Brief_History_of_the_Glazier-Graner-Hogeweg_Model/links/00b7d52d79e94eadc7000000.pdf.
- [15] F. Graner and J. A. Glazier, "Simulation of biological cell sorting using a two-dimensional extended potts model", *Phys. Rev. Lett.*, vol. 69, pp. 2013–2016, 13 Sep. 1992. DOI: 10.1103/PhysRevLett.69.2013. [Online]. Available: https://link.aps.org/doi/10.1103/PhysRevLett.69.2013.

- [16] J. A. Glazier and F. Graner, "Simulation of the differential adhesion driven rearrangement of biological cells", *Phys. Rev. E*, vol. 47, pp. 2128–2154, 3 Mar. 1993. DOI: 10.1103/PhysRevE.47.2128. [Online]. Available: https://link.aps.org/doi/10.1103/PhysRevE.47.2128.
- [17] E. Stott, N. Britton, J. Glazier, and M. Zajac, "Stochastic simulation of benign avascular tumour growth using the potts model", *Mathematical and Computer Modelling*, vol. 30, no. 5, pp. 183–198, 1999. DOI: https://doi.org/10.1016/S0895-7177(99)00156-9. [Online]. Available: http://www.sciencedirect.com/science/article/pii/S0895717799001569.
- [18] N. Chen, J. A. Glazier, J. A. Izaguirre, and M. S. Alber, "A parallel implementation of the cellular potts model for simulation of cell-based morphogenesis", *Computer Physics Communications*, vol. 176, no. 11, pp. 670–681, 2007. DOI: https://doi.org/10.1016/j.cpc.2007.03.007. [Online]. Available: http://www.sciencedirect.com/science/article/pii/S0010465507002044.
- [19] T. M. Cickovski, C. Huang, R. Chaturvedi, T. Glimm, H. G. E. Hentschel, M. S. Alber, J. A. Glazier, S. A. Newman, and J. A. Izaguirre, "A framework for three-dimensional simulation of morphogenesis", *IEEE/ACM Transactions on Computational Biology and Bioinformatics*, vol. 2, no. 4, pp. 273–288, Oct. 2005. DOI: 10.1109/TCBB.2005.46.
- [20] A. Torelli, P. Erben, J. Debatin, and M. Gumbel, "Proliferation and regeneration of the healthy human urothelium: A multi-scale simulation approach with 16 hypotheses of cell differentiation".