

**ELECTRONIC SUPPLEMENTARY MATERIAL: A MECHANISTIC MODEL TO
COMPARE THE IMPORTANCE OF INTERRELATED POPULATION-LEVEL
MEASURES ON PATHOGEN RICHNESS**

TIM C.D. LUCAS^{1*†}, HILDE M. WILKINSON-HERBOTS² AND KATE E. JONES^{3*}

¹Centre for Biodiversity and Environment Research, Department of Genetics, Evolution and Environment, University College London, Gower Street, London, WC1E 6BT, United Kingdom. 0000-0003-4694-8107

²Department of Statistical Science, University College London, Gower Street, London, WC1E 6BT, United Kingdom.

³Centre for Biodiversity and Environment Research, Department of Genetics, Evolution and Environment, University College London, Gower Street, London, WC1E 6BT, United Kingdom and Institute of Zoology, Zoological Society of London, Regent's Park, London, NW1 4RY, United Kingdom. 0000-0001-5231-3293

[†]Current address: Oxford Big Data Institute, Li Ka Shing Centre for Health Information and Discovery, University of Oxford, Oxford, OX3 7FZ, United Kingdom.

*Corresponding authors: Tim C.D. Lucas: timcdlucas@gmail.com, Kate E. Jones: kate.e.jones@ucl.ac.uk

TABLE S1. A summary of all symbols used along with their units and default values. The justifications for parameter values are given in the main manuscript.

Symbol	Explanation	Units	Value
ρ	Number of pathogens		2
x, y	Colony index		
p	Pathogen index i.e. $p \in \{1, 2\}$ for pathogens 1 and 2		
q	Disease class i.e. $q \in \{1, 2, 12\}$		
S_x	Number of susceptible individuals in colony x		
I_{qx}	Number of individuals infected with disease(s) $q \in \{1, 2, 12\}$ in colony x		
R_x	Number of individuals in colony x in the recovered with immunity class		
N	Total Population size		2,000 – 32,000
m	Number of colonies		5 – 80
n	Colony size		100 – 600
a	Area	km ²	2,500 – 40,000
d	Density	individuals.km ⁻¹	0.2 – 3.2
β	Transmission rate		0.1 – 0.4
α	Coinfection adjustment factor		0.1
γ	Recovery rate	year ⁻¹ .individual ⁻¹	1
ξ	Dispersal rate	year ⁻¹ .individual ⁻¹	0.01
Λ	Birth rate	year ⁻¹ .individual ⁻¹	0.05
μ	Death rate	year ⁻¹ .individual ⁻¹	0.05
g	Population growth rate		1
k_y	Degree of node y (number of colonies that individuals from colony y can disperse to)		
δ	Waiting time until next event	years	
e_i	The rate at which event i occurs	year ⁻¹	
R_0	The (single subpopulation) basic reproduction number	infections	
b_i	Regression coefficient		
c	Regression intercept		
ϵ	Binomially distributed regression error term		

S1. SUPPLEMENTARY MATERIALS AND METHODS: STOCHASTIC SIMULATIONS

We examined the model using stochastic, continuous-time simulations implemented in R [1]. The implementation is available as an R package on GitHub [2]. The model can be written as a continuous-time Markov chain. The Markov chain contains the random variables $((S_x)_{x=1\dots m}, (I_{x,q})_{x=1\dots m, q \in \{1,2,12\}}, (R_x)_{x=1\dots m})$. Here, $(S_x)_{x=1\dots m}$ is a length m vector of the number of susceptibles in each colony. $(I_{x,q})_{x=1\dots m, q \in \{1,2,12\}}$ is a length $m \times 3$ vector describing the number of individuals of each disease class ($q \in \{1,2,12\}$) in each colony. Finally, $(R_x)_{x=1\dots m}$ is a length m vector of the number of individuals in the recovered class. The model is a Markov chain where extinction of both pathogen species and extinction of the host species are absorbing states. The expected time for either host to go extinct is much larger than the duration of the simulations.

At any time, suppose the system is in state $((s_x), (i_{x,q}), (r_x))$. At each step in the simulation we calculate the rate at which each possible event might occur. One event is then randomly chosen, weighted by its rate

$$p(\text{event } i) = \frac{e_i}{\sum_j e_j}, \quad (1)$$

where e_i is the rate at which event i occurs and $\sum_j e_j$ is the sum of the rates of all possible events. Finally, the length of the time step, δ , is drawn from an exponential distribution

$$\delta \sim \text{Exp} \left(\sum_j e_j \right). \quad (2)$$

We can now write down the rates of all events. Assuming asexual reproduction, that all classes reproduce at the same rate and that individuals are born into the susceptible class we get

$$s_x \rightarrow s_x + 1 \quad \text{at a rate of } \Lambda \left(s_x + \sum_q i_{qx} + r_x \right) \quad (3)$$

where $s_x \rightarrow s_x + 1$ is the event that the number of susceptibles in colony x will increase by 1 (a single birth) and $\sum_q i_{qx}$ is the sum of all infection classes $q \in \{1,2,12\}$. The rates of death, given a death rate μ , and no increased mortality due to infection, are given by

$$s_x \rightarrow s_x - 1 \quad \text{at a rate of } \mu s_x, \quad (4)$$

$$i_{qx} \rightarrow i_{qx} - 1 \quad \text{at a rate of } \mu i_{qx}, \quad (5)$$

$$r_x \rightarrow r_x - 1 \quad \text{at a rate of } \mu r_x. \quad (6)$$

We modelled transmission as being density-dependent. This assumption was more suitable than frequency-dependent transmission as we were modelling a disease transmitted by saliva or urine in highly dense populations confined to caves, buildings or potentially a small number of tree roosts. We were notably not modelling a sexually transmitted disease (STD) as spillover of STDs from bats to humans is likely to be rare. Infection of a susceptible with either Pathogen 1 or 2 is therefore given by

$$i_{1x} \rightarrow i_{1x} + 1, \quad s_x \rightarrow s_x - 1 \quad \text{at a rate of } \beta s_x (i_{1x} + i_{12x}), \quad (7)$$

$$i_{2x} \rightarrow i_{2x} + 1, \quad s_x \rightarrow s_x - 1 \quad \text{at a rate of } \beta s_x (i_{2x} + i_{12x}), \quad (8)$$

while coinfection, given the coinfection adjustment factor α , is given by

$$i_{12,x} \rightarrow i_{12,x} + 1, \quad i_{1x} \rightarrow i_{1x} - 1 \quad \text{at a rate of } \alpha \beta i_{1x} (i_{2x} + i_{12x}), \quad (9)$$

$$i_{12,x} \rightarrow i_{12,x} + 1, \quad i_{2x} \rightarrow i_{2x} - 1 \quad \text{at a rate of } \alpha \beta i_{2x} (i_{1x} + i_{12x}). \quad (10)$$

Note that lower values of α give lower rates of coinfection as in Castillo-Chavez et al. [3].

The rate of migration from colony y (with degree k_y) to colony x , given a dispersal rate ξ is given by

$$s_x \rightarrow s_x + 1, \quad s_y \rightarrow s_y - 1 \quad \text{at a rate of } \frac{\xi s_y}{k_y}, \quad (11)$$

$$i_{qx} \rightarrow i_{qx} + 1, \quad i_{qy} \rightarrow i_{qy} - 1 \quad \text{at a rate of } \frac{\xi i_{qy}}{k_y}, \quad (12)$$

$$r_x \rightarrow r_x + 1, \quad r_y \rightarrow r_y - 1 \quad \text{at a rate of } \frac{\xi r_y}{k_y}. \quad (13)$$

Note that the dispersal rate does not change with infection. As above, this is due to the low virulence of bat viruses. Finally, recovery from any infectious class occurs at a rate γ

$$i_{qx} \rightarrow i_{qx} - 1, \quad r_x \rightarrow r_x + 1 \quad \text{at a rate of } \gamma i_{qx}. \quad (14)$$

S2. SUPPLEMENTARY MATERIALS AND METHODS: DETERMINISTIC MODEL

We can study the model analytically if we restrict the population to a single subpopulation. We first study the endemic pathogen. We then have a typical SIR model with vital dynamics [4].

$$\frac{dS}{dt} = \Lambda n - \mu S - \beta S I_1, \quad (15)$$

$$\frac{dI_1}{dt} = \beta S I_1 - (\gamma + \mu) I_1, \quad (16)$$

$$\frac{dR}{dt} = \gamma I_1 - \mu R. \quad (17)$$

S2.0.1. *Equilibrium values of single pathogen model.* To find the equilibrium number of susceptibles, S^* , we set $\frac{dI_1}{dt} = 0$.

$$\beta S^* I_1 = (\gamma + \mu) I_1, \quad (18)$$

$$S^* = \frac{\gamma + \mu}{\beta}. \quad (19)$$

To find I^* we substitute Equation 19 into Equation 15 and set $\frac{dI_1}{dt} = 0$ giving

$$0 = \Lambda n - \mu S^* - \beta S^* I_1^*, \quad (20)$$

$$0 = \Lambda n - \frac{\mu(\gamma + \mu)}{\beta} - (\gamma + \mu) I_1^*, \quad (21)$$

$$I_1^* = \frac{\Lambda n}{\gamma + \mu} - \frac{\mu}{\beta}. \quad (22)$$

S2.0.2. *Second pathogen.* If we assume that Pathogen 2 is introduced when the endemic pathogen is at equilibrium, the dynamics of Pathogen 2 are governed by

$$\frac{dI_2}{dt} = \beta S^* I_2 + \alpha \beta I_1^* I_2 - (\gamma + \mu) I_2 \quad (23)$$

where $\beta S^* I_2$ is the number of infections, $\alpha \beta I_1^* I_2$ is the number of coinfections and $(\gamma + \mu) I_2$ is the number of deaths and recoveries. I_2 is therefore the number of individuals infected with Pathogen 2 regardless of whether they are infected with Pathogen 1. In this sense it is the combined size of classes I_2 and $I_1 I_2$ as used in the stochastic model (Section S1). For simplicity here we simply label this group I_2 . Substituting in Equations 19 and 22, and noting that $R_0 = \frac{\beta n}{\gamma + \mu}$ we get

$$\frac{dI_2}{dt} = (\gamma + \mu) I_2 + \alpha \beta \left(\frac{\Lambda n}{\gamma + \mu} - \frac{\mu}{\beta} \right) I_2 - (\gamma + \mu) I_2, \quad (24)$$

$$\frac{dI_2}{dt} = \alpha \left(\frac{\beta \Lambda n}{\gamma + \mu} - \mu \right) I_2, \quad (25)$$

$$\frac{dI_2}{dt} = \alpha (\Lambda R_0 - \mu) I_2. \quad (26)$$

This is greater than zero when

$$\alpha (\Lambda R_0 - \mu) I_2 > 0 \quad (27)$$

As I_2 is positive, this is greater than zero when $\Lambda R_0 - \mu > 0$ and $\alpha > 0$. As $\Lambda = \mu$ due to the assumption of stable population size, $\Lambda R_0 - \mu > 0$ is true as long as $R_0 > 1$. R_0 is greater than one (as proved by the fact that Pathogen 1 is endemic), Therefore, $\frac{dI_2}{dt}$ is greater than zero as long as α is greater than zero.

TABLE S2. Raw data for range size simulations. The population parameters are shown along with the number of invasions and the number of simulations. Note that simulations where both pathogens went extinct have been removed (100 simulations were originally run for each parameter set). β is the transmission rate, n is colony size, m is the number of colonies and N is the total population size.

β	n	m	Area ($\times 1000 \text{ km}^2$)	N ($\times 1000$)	Density (km^{-2})	Invasions	Sims
0.1	400	20	2.5	8	3.2	2	100
0.1	400	20	5.0	8	1.6	3	100
0.1	400	20	10.0	8	0.8	2	100
0.1	400	20	20.0	8	0.4	3	100
0.1	400	20	40.0	8	0.2	2	100
0.2	400	20	2.5	8	3.2	3	100
0.2	400	20	5.0	8	1.6	3	100
0.2	400	20	10.0	8	0.8	1	100
0.2	400	20	20.0	8	0.4	4	100
0.2	400	20	40.0	8	0.2	1	100
0.3	400	20	2.5	8	3.2	3	100
0.3	400	20	5.0	8	1.6	3	100
0.3	400	20	10.0	8	0.8	3	100
0.3	400	20	20.0	8	0.4	5	100
0.3	400	20	40.0	8	0.2	9	100

TABLE S3. Raw data for colony size simulations. The population parameters are shown along with the number of invasions and the number of simulations. Note that simulations where both pathogens went extinct have been removed (100 simulations were originally run for each parameter set). β is the transmission rate, n is colony size, m is the number of colonies and N is the total population size.

β	n	m	Area ($\times 1000 \text{ km}^2$)	N ($\times 1000$)	Density (km^{-2})	Invasions	Sims
0.1	100	20	2.5	2	0.8	4	88
0.1	200	20	5.0	4	0.8	5	100
0.1	400	20	10.0	8	0.8	2	100
0.1	800	20	20.0	16	0.8	0	100
0.1	1600	20	40.0	32	0.8	55	100
0.2	100	20	2.5	2	0.8	3	92
0.2	200	20	5.0	4	0.8	6	100
0.2	400	20	10.0	8	0.8	0	100
0.2	800	20	20.0	16	0.8	39	100
0.2	1600	20	40.0	32	0.8	95	100
0.3	100	20	2.5	2	0.8	1	91
0.3	200	20	5.0	4	0.8	4	100
0.3	400	20	10.0	8	0.8	7	100
0.3	800	20	20.0	16	0.8	67	100
0.3	1600	20	40.0	32	0.8	100	100

TABLE S4. Raw data for number of colonies simulations. The population parameters are shown along with the number of invasions and the number of simulations. Note that simulations where both pathogens went extinct have been removed (100 simulations were originally run for each parameter set). β is the transmission rate, n is colony size, m is the number of colonies and N is the total population size.

β	n	m	Area ($\times 1000 \text{ km}^2$)	N ($\times 1000$)	Density (km^{-2})	Invasions	Sims
0.1	400	5	2.5	2	0.8	0	97
0.1	400	10	5.0	4	0.8	0	100
0.1	400	20	10.0	8	0.8	2	100
0.1	400	40	20.0	16	0.8	2	100
0.1	400	80	40.0	32	0.8	7	100
0.2	400	5	2.5	2	0.8	2	99
0.2	400	10	5.0	4	0.8	1	100
0.2	400	20	10.0	8	0.8	0	100
0.2	400	40	20.0	16	0.8	3	100
0.2	400	80	40.0	32	0.8	11	100
0.3	400	5	2.5	2	0.8	1	96
0.3	400	10	5.0	4	0.8	2	100
0.3	400	20	10.0	8	0.8	7	100
0.3	400	40	20.0	16	0.8	15	100
0.3	400	80	40.0	32	0.8	17	100

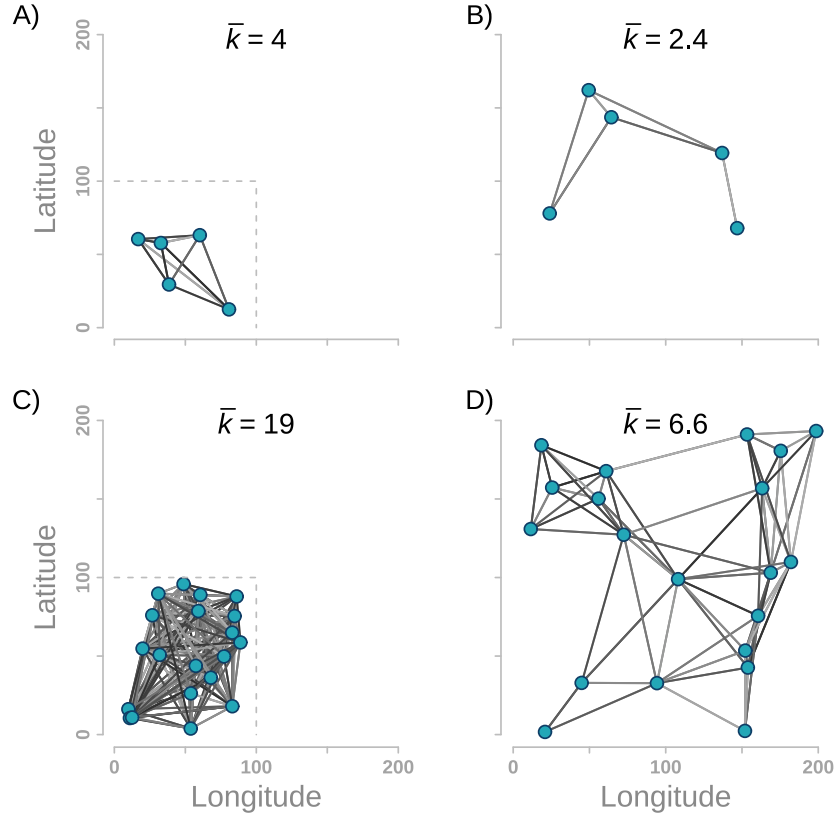


FIGURE S1. The relationship between range size and metapopulation network structure. Colonies are shown by circles. Colonies that are close enough for animals to disperse between (i.e. within 100 km of each other) are joined by a line. Colonies are placed randomly in spaces of various sizes (grey dashed lines). A and C) the default range size (10000 km²). B and D) the largest range size (40000 km²). A and B) the smallest number of colonies (five). C and D) the default number of colonies (20). The mean number of connections per subpopulation, \bar{k} , is shown for each metapopulation.

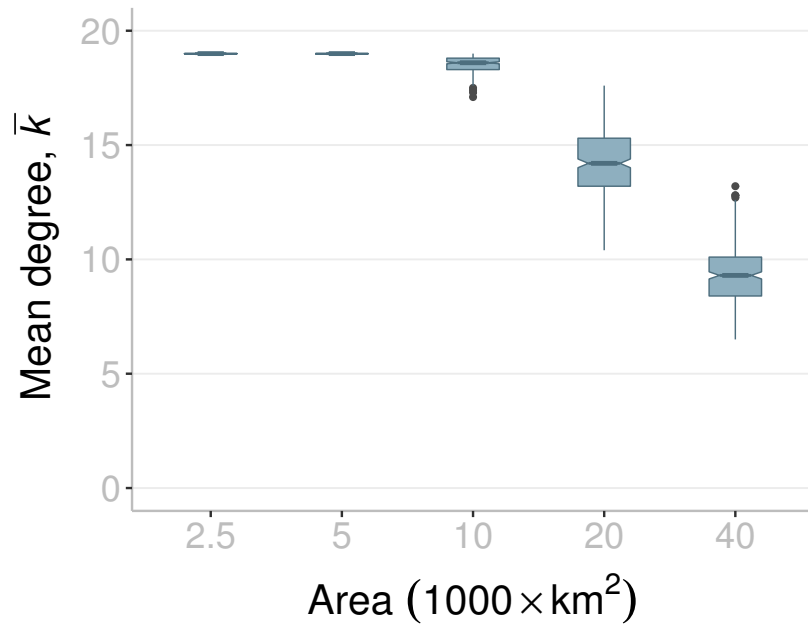


FIGURE S2. Change in average metapopulation network degree (\bar{k}) with increasing range size. Bars show the median, boxes show the interquartile range, vertical lines show the range and grey dots indicate outlier values. Notches indicate the 95% confidence interval of the median. All simulations had 20 colonies, meaning 19 is the maximum value of \bar{k} .

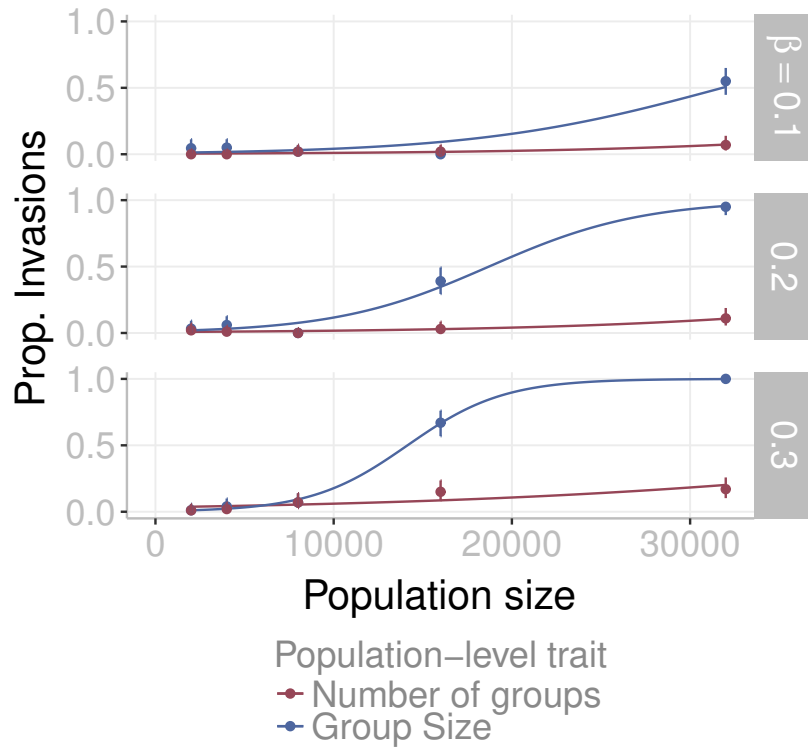


FIGURE S3. Comparison of the effect of host population size on probability of invasion when population size is altered by changing colony size or colony number. Relationships are shown separately for each transmission value, β . It can be seen that changes in colony size give a much greater increase in invasion probability than changes in colony number. Note that this is the same data as Figure 3 but with the x -axis scaled by population size, rather than relative parameter change.

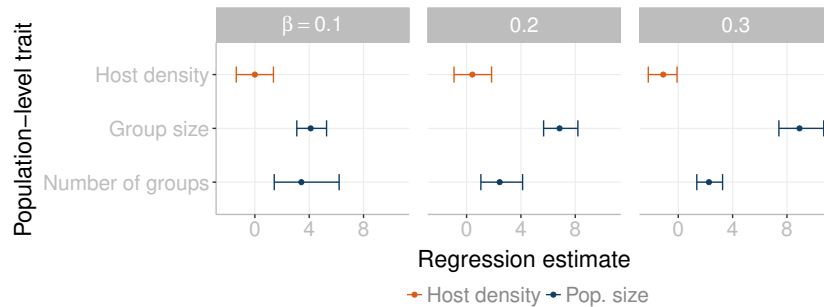


FIGURE S4. GLM regression coefficients for for three sets of simulations at three transmission, β values. To examine whether host density or population size affects pathogen invasion more strongly we compare the orange and blue bars. To examine whether group size or number of groups affects pathogen invasion more strongly we compare the second and third bar in each facet.

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