## Statisitcal Thinking in Biology Research Chapter 1

Terry Neeman and Timothée Bonnet

November 27, 2018

### Acknowledgemnts and warning

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- Statistical ideas about biological variation inform the design of experiments
- Statistical ideas about biological variation inform the analysis of experiments
- Statistical thinking is an essential component of scientific thinking

### A bit of history of statistical methods

R.A. Fisher: 1890-1962



Statistical Principles for Research Workers (1925)

### A bit of history of statistical methods

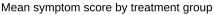
R.A. Fisher: 1890-1962

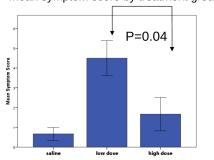


Statistical Principles for Research Workers (1925)

- Cautionary tales from the front
- 2 Introduction to Statistical Modelling
- 3 Another look at essential steps

# Message 1: A small p-value is not always evidence of a treatment effect



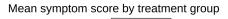


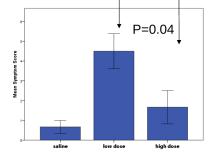
#### Vaccine challenge experiment:

- 6 mice/group (saline/low dose/high dose)
- All mice challenged with Shigella
- Followed for 14 days
- Outcome: Symptom score average Days 2 - 8

One-way ANOVA (post-hoc Bonferroni) p=0.04

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Do you think the vaccine works? What is strange?



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#### Experimental design

The observed difference in outcome could be the result of:

- Cage effects
- Mouse strain effects

These effects are CONFOUNDED with treatment effect



Cage 1: saline

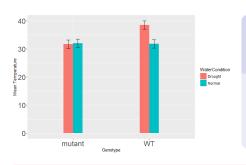


Cage 3: High Dose



Message 2: p-values from simple comparisons cannot tell us when differences are "different"

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Are temperature mechanisms modified in a genetically modified tomato plant?

- Genotypes: WT/mutant
- Water condition: Normal/Drought
- Leaf temperature measured

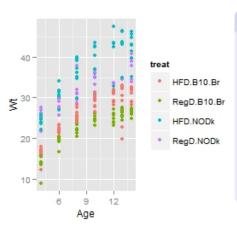
#### Comparisons made using t-tests

Evidence of difference + No evidence of difference  $\neq$  Evidence that differences are different.

Message 3: Interpreting experimental results needs more than t-tests

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Research question: Are mice susceptible to obesity when exposed to a high fat diet?



#### Experimental set-up:

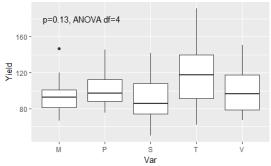
- 37 mice: 16 NODk /21 WT
- Randomised to either regular or high fat diet
- Monitored for 14 weeks
- Outcome measure: Body weight (g)
- Experimental factors: Diet (2), Strain (2), Time (8)

Acknowledgements: Ainy Hussain, PhD student 2013

Comparing yield in five barley varieties (1930s)

Experimental factors: 5 varieties of barley, 6 locations, 2 time points.

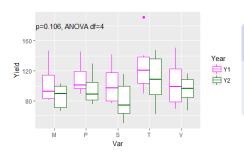
Outcome measure: yield



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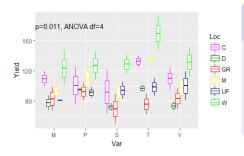
### Controlling for other sources of variation:

 Controlling for year = comparing yield WITHIN years and combining these

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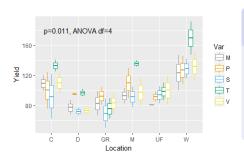
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Message 5: Knowing what factors contribute to the variation in outcome helps design experiments and analyses

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Research question: How does cold duration impact upon germination in alpine plant A. glacialis?



### Experimental set-up:

- Seed collections from alpine region in Australia
- 3 Regions- low/high altitude
- 4 sets of Petri dishes
- 4 cabinet shelves
- Response % germinated

What other factors are important to consider when comparing cold duration?

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- Interpreting experimental results needs more than t-tests. Need a statistical model of the experiment, matching scientific question.
- Combining information across subgroups can improve inference. A statistical model enables accumulation of evidence across experiments.
- Knowing what factors contribute to the variation in outcome matters. A statistical model allows one to incorporate effect of other factors in the analysis.

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### Introduction to Statistical Modelling

- What is a statistical model?
- Modelling outcomes:
  - a summary of data
  - a prediction model
  - an explanatory model
- Model may take many different functional forms
- Model a conceptualization of the experiment

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#### ALWAYS BEGIN WITH A RESEARCH QUESTION

### Key components of a statistical model of an experiment

- Outcome measure
  - Response variable
  - Measure of interest
- Experimental factors
  - Conditions that can be manipulated
  - Conditions of interest (e.g. genotype, gender)
  - ▶ Main questions: do the conditions impact upon the outcome measure?
- Blocking factors
  - Conditions (not of interest) that may impact upon the outcome measure
  - Sources of variation in the experiment that need to be controlled for
  - Clustering of experimental units

#### ALWAYS BEGIN WITH A RESEARCH QUESTION



### Key Objectives of a statistical model of an experiment

- To compare the mean response of an organism/system to a set of different experimental conditions.
  - Obtain estimate of "Treatment effect"
  - ▶ Is this "effect" different in subgroups of interest?
- What are the most important factors influencing the mean response?
- Subsidiary question: how can we design our experiment in future to more efficiently test our hypotheses?

# Example 1: Does dark respiration differ between C3 and C4 plants?

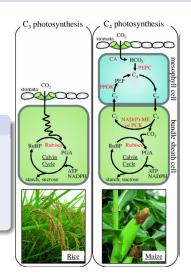
Outcome measure: dark respiration

Experimental factor: Plant type (C4/C3)

Data: 6 plants each of C4, C3

#### Can calculate

- Observed overall mean
- Observed mean C3 plants
- Observed mean C4 plants
- Variation around each mean



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#### Statistical model

Respiration = Mean for C3 + Difference C4-C3 \* (is C4?) + Noise

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- Observed mean C3 plants
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- Variation around each mean

#### Statistical model

```
Respiration = Mean for C3 + Difference C4-C3 * (is C4?) + Noise response = A + D \times predictor + \epsilon
```

A and D are the model PARAMETERS.

We want to infer whether D is different from 0

response =  $A + D \times predictor + \epsilon$ Can we separate the signal D from the noise  $\epsilon$ ?

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#### T-test

- Outcome is a continuous variable
- Experimental factor is one factor with 2 conditions
- No blocking factor / corrections

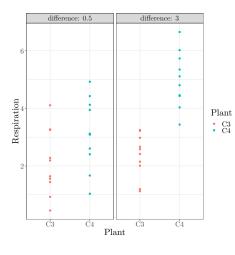
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#### T-test

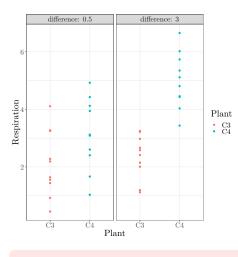
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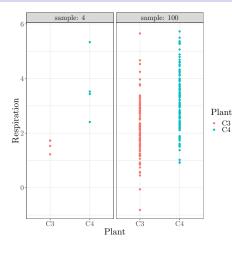
Is it easier when the true difference is 0.5 or when it is 3 ?



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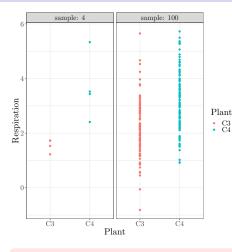
Is it easier when the true difference is 0.5 or when it is 3 ?

Large true difference between the means



$$t = rac{ extstyle D}{ extstyle Variation of } extstyle extstyle rac{ extstyle Sample Size}{\sqrt{2}}$$

Is it easier when sample size is 4 or when it is 100?

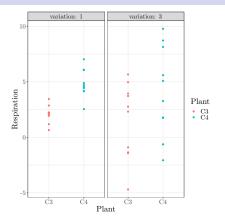


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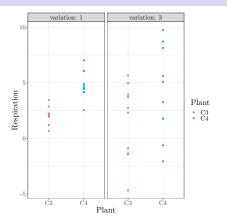
- Large true difference between the means
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$$t = rac{ extstyle D}{ extstyle Variation of } extstyle extstyle rac{ extstyle Sample Size}{\sqrt{2}}$$

Is it easier when unexplained variation is 1 or when it is 3?

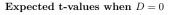


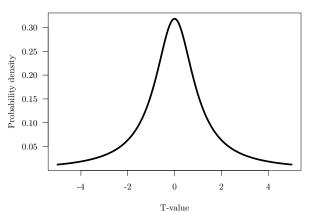
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## What makes *t* large:

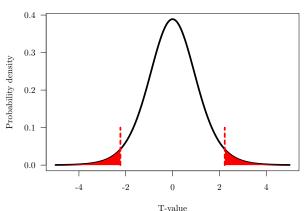
- Large true difference between the means
- 2 Large sample size
- Small unexplained variation





p-value: probability (area under curve) of getting a value as extreme as what you observed, when D=0

#### Expected t-values when D=0



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#### How to?

Draw 5 candies out of the pack

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- Estimate the p-value for the test "candies have same frequency"
- Redo the experiment in R, using random sampling (rbinom)
- What is the correct null-distribution?

# Back to C3/C4 plants. Analyse real data in R

- 1. Set working directory (setwd('' / '')) or create a R-project
- 2. Load and check data

```
resp <- read.csv("d_respiration.csv")
str(resp)
View(resp)</pre>
```

#### 3. Visualize data

```
library(ggplot2)
ggplot(resp,aes(Plant_type,rrarea,colour=Plant_type))+
    geom_point()+facet_wrap(~Variation)
```

#### Subset data by Variation (High and Low)

```
resp_H <- subset(resp, Variation == "High")
resp_L <- subset(resp, Variation == "Low")</pre>
```

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#### Compare C3 and C4 plants in "High Variation" subset

```
t.test(rrarea~Plant_type, data=resp_H, var.equal=TRUE)
```

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```
Two Sample t-test
data: rrarea by Plant_type
t = -0.93776, df = 10, p-value = 0.3705
alternative hypothesis: true difference in means is not equal to 0
95 percent confidence interval:
-1.7619349 0.7181446
sample estimates:
mean in group C3 mean in group C4
2.720021 3.241916
```

#### Subset data by Variation (High and Low)

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resp_H <- subset(resp, Variation == "High")
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```

#### Compare C3 and C4 plants in "Low Variation" subset

t.test(rrarea~Plant\_type, data=resp\_L, var.equal=TRUE)

## Fit an anova in R: aov()

```
aov1 <- aov(rrarea~Plant_type, data=resp_H)
summary(aov1)</pre>
```

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aov1 <- aov(rrarea~Plant_type, data=resp_H)
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```
Df Sum Sq Mean Sq F value Pr(>F)
Plant_type 1 0.817 0.8171 0.879 0.37
Residuals 10 9.292 0.9292
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## Fit an anova in R: aov()

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aov1 <- aov(rrarea~Plant_type, data=resp_H)
summary(aov1)</pre>
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$$response = A + D \times predictor + \epsilon$$

## Fit a linear model in R: lm()

```
lm1<-lm(rrarea ~ Plant_type, data = resp_L)
summary(lm1)</pre>
```

## Fit a linear model in R: lm()

lm1<-lm(rrarea ~ Plant\_type, data = resp\_L)</pre>

```
summary(lm1)
lm(formula = rrarea ~ Plant_type, data = resp_H)
Residuals:
   Min 1Q Median 3Q Max
-1.7380 -0.4201 -0.1437 0.6706 1.6754
Coefficients:
            Estimate Std. Error t value Pr(>|t|)
(Intercept) 2.7200 0.3935 6.912 4.13e-05 ***
Plant_typeC4 0.5219 0.5565 0.938 0.37
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '. '0.1 ' 1
Residual standard error: 0.9639 on 10 degrees of freedom
Multiple R-squared: 0.08083, Adjusted R-squared: -0.01109
F-statistic: 0.8794 on 1 and 10 DF, p-value: 0.3705
```

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lm1<-lm(rrarea ~ Plant\_type, data = resp\_L)</pre>

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#### Fit a linear model in R: lm()

```
library(emmeans)
emmeans(lm1, ~Plant_type)
```

```
Plant_type emmean SE df lower.CL upper.CL
C3 2.720021 0.3935305 10 1.843180 3.596861
C4 3.241916 0.3935305 10 2.365076 4.118757

Confidence level used: 0.95
```

#### Fit a linear model in R: 1m()

```
library(emmeans)
emmeans(lm1, ~Plant_type)
```

```
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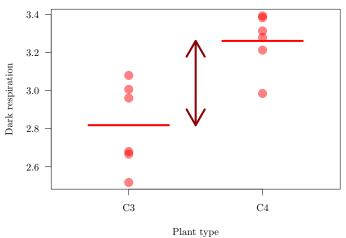
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```

$$response = A + D \times predictor + \epsilon$$

Compare the output from t.test, aov and Im

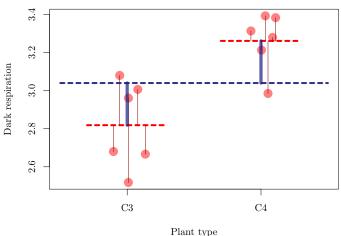
### Three equivalent ways to look at data

T-test, focus on difference between two means



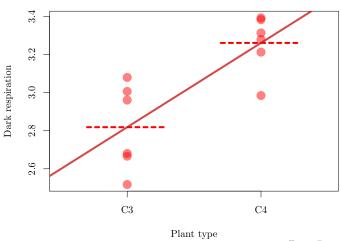
#### Three equivalent ways to look at data

ANOVA, focus on variation within VS. between



#### Three equivalent ways to look at data

Linear regression, focus on rate of change



All is one...

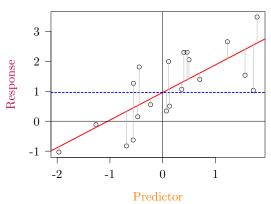
#### All is one...

#### ...but lm() rules (IMH)

- t-test, ANOVA, regression and others can be mathematically equivalent
- In R, lm() and related functions can do them all...
- ...and much more!

#### Focus on linear models

#### $Response = Intercept + Slope \times Predictor + Error$



#### A simple linear model

#### Response = Intercept + Slope $\times$ Predictor + Error

```
lm(response ~ 1 + predictor1 + predictor2, data=data)
```

#### equivalent to

```
lm(response ~ predictor1 + predictor2, data=data)
```

#### equivalent to

```
lm(response ~ predictor2 + predictor1, data=data)
```

- Intercept can be explicit or implicit
- Can remove intercept with  $\ldots \sim 0 + \ldots$
- Error is implicit
- Feed the option data= to keep code short, reliable and flexible
- Order of predictors do not matter



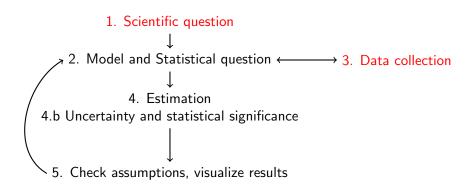
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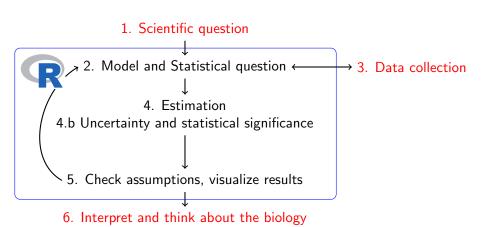
1. Scientific question

- 1. Scientific question
- 2. Model and Statistical question

- 1. Scientific question
- 2. Model and Statistical question ← → 3. Data collection

- 1. Scientific question
- 2. Model and Statistical question  $\longleftrightarrow$  3. Data collection
  - 4. Estimation
- 4.b Uncertainty and statistical significance





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summary(lmL)</pre>
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Estimate Std. Error t value Pr(>|t|)
(Intercept) 2.81857 0.07856 35.878 6.72e-12 ***
Plant_typeC4 0.44235 0.11110 3.982 0.00259 ** ---
...
```

#### **Estimation:**

response = 
$$A + D \times predictor + \epsilon$$
  
 $A = ?, D = ?$ 

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#### What do we do next?



#### Linear model basic assumptions

Predictor not perfectly correlated
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 Risk: Over-optimistic uncertainty, unreliable predictions

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Independence of error
 Risk: Bias and over-optimistic uncertainty

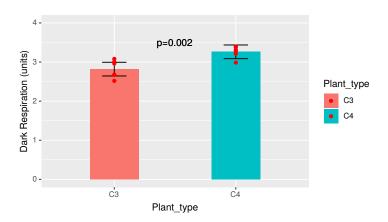
#### Assessing model assumptions in R:

```
lmL<-lm(rrarea ~ Plant_type, data = resp_L)
plot(lmL)
summary(lmL)</pre>
```

#### Visualize and report results

```
lm1.results<-summary(emmeans(lm1,~Plant_type))

ggplot(lm1.results,aes(Plant_type,emmean, fill=Plant_type))+
    geom_bar(stat="identity", width=.4)+
    geom_errorbar(aes(ymin =lm1.results$lower.CL,
    ymax = lm1.results$upper.CL), width=.2)+
    ylim(0,4)+
    geom_point(data=resp_L, aes(x=Plant_type, y=rrarea), color="red")+
    labs(y = "Dark Respiration (units)")+
    geom_text(aes(x=1.5, y=3.5, label="p=0.002"))</pre>
```



### Another example

#### Compare wheat yields between 3 varieties

• Outcome measure: Tonnes/hectare

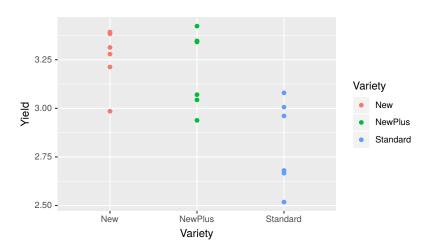
Experimental factor: Variety (new/newPLUS/standard)

Data: 6 plots/ variety



How many parameters in this model?

## Results from Wheat Yield Experiment with 3 Varieties



## Analyse these data in R

# Sample analysis in R: 1-way ANOVA

```
aov1=aov(Yield~Variety, data = wheat2)
summary(aov1)
```

```
Df Sum Sq Mean Sq F value Pr(>F)

Variety 2 0.6820 0.3410 8.951 0.00276 **

Residuals 15 0.5714 0.0381
```

#### **ANOVA**

- Compares means between TWO or MORE GROUPS
- Relies on F-statistic =  $\frac{\text{Between-groups variance}}{\text{Within groups variance}} = \frac{\text{Explained}}{\text{Unexplained}}$
- One test for significance of all groups

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But which groups are different???



## Using emmeans to extract group estimates

```
emmeans(aov1, ~Variety)
```

```
Variety emmean SE df lower.CL upper.CL Standard 2.81 0.079 15 2.64 2.98
New 3.26 0.079 15 3.09 3.43
NewPlus 3.19 0.079 15 3.02 3.36

Confidence level used: 0.95
```

### Using emmeans to compare groups

emmeans(aov1, pairwise~Variety)

### Equivalent with lm()

```
lm2<-lm(Yield ~ Variety, data = wheat2)
anova(lm2)
summary(lm2)</pre>
```

```
Analysis of Variance Table

Response: Yield

Df Sum Sq Mean Sq F value Pr(>F)

Variety 2 0.68203 0.34101 8.9513 0.002764 **

Residuals 15 0.57145 0.03810
```

```
Coefficients:

Estimate Std. Error t value Pr(>|t|)

(Intercept) 2.81857 0.07968 35.372 7.26e-16 ***

VarietyNew 0.44235 0.11269 3.925 0.00135 **

VarietyNewPlus 0.37529 0.11269 3.330 0.00457 **
```

### Equivalent with lm()

#### emmeans(lm2, pairwise~Variety)

```
        $emmeans

        Variety
        emmean
        SE
        df
        lower.CL upper.CL

        Standard
        2.81
        0.0796
        15
        2.64
        2.98

        New
        3.26
        0.0796
        15
        3.09
        3.43

        NewPlus
        3.19
        0.0796
        15
        3.02
        3.36
```

Confidence level used: 0.95

#### \$contrasts

 contrast
 estimate
 SE
 df
 t.ratio p.value

 Standard - New
 -0.44
 0.112
 15
 -3.925
 0.0036

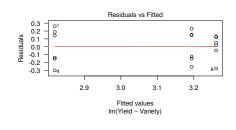
 Standard - NewPlus
 -0.37
 0.112
 15
 -3.330
 0.0120

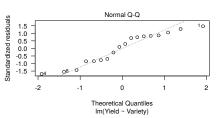
 New - NewPlus
 0.067
 0.112
 15
 0.595
 0.8248

P value adjustment: tukey method for comparing a family of 3 estimates

# Assessing model assumptions for lm()

plot(lm2)





# Summary of results with post hoc comparisons:

