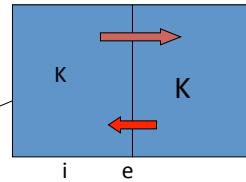


What are the flows (j' 's) if $V_{rest} = -80\text{mV}$



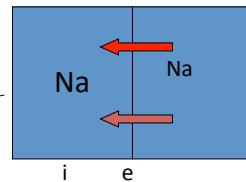
$$E_K = -104\text{mV}$$

$$j_{diff} > j_{Efield}$$

net influx

$$j_{diff} \& j_{Efield}$$

point inward



$$E_{Na} = +85.3\text{mV}$$

Neither Nernst Potential is in equilibrium with the rest potential.

We can have another kind of equilibrium

$$\sum_p j_p = 0$$

The Goldman Equation was developed to consider this type of equilibrium. It is based on the Nernst Planck Equation, but under certain conditions.

Let's look at the total current

$$J = J_K + J_{Na} + J_{Cl}$$

From Goldman eqn we can write the following

$$J = \frac{V_m F^2 P_K}{RT} \left(\frac{w - ye^{V_m F / RT}}{1 - e^{V_m F / RT}} \right)$$

where

$$w = [K]_e + \frac{P_{Na}}{P_K} [Na]_e + \frac{P_{Cl}}{P_K} [Cl]_i$$

$$y = [K]_i + \frac{P_{Na}}{P_K} [Na]_i + \frac{P_{Cl}}{P_K} [Cl]_e$$

At Equilibrium: net current $J=0$

$$J = \frac{V_m F^2 P_K}{RT} \left(\frac{w - ye^{V_m F / RT}}{1 - e^{V_m F / RT}} \right) = 0$$

or

$$w - ye^{V_m F / RT} = 0 \implies e^{V_m F / RT} = \frac{w}{y}$$

$$V_m = \frac{RT}{F} \ln \frac{w}{y}$$

Goldman Eqn for rest (equil)
potential

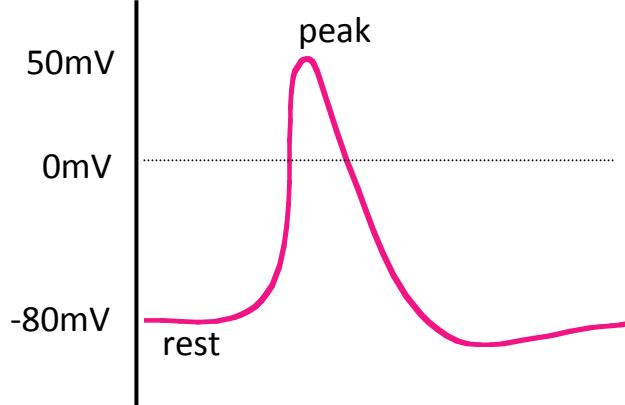
From Goldman Eqn- Equilibrium is a weighted average of all the Nernst Potentials.
The weights are the permeabilities.

Under rest conditions, what permeability is greatest? Na? K?

$$P_K \gg P_{Na}$$

How do we know this?

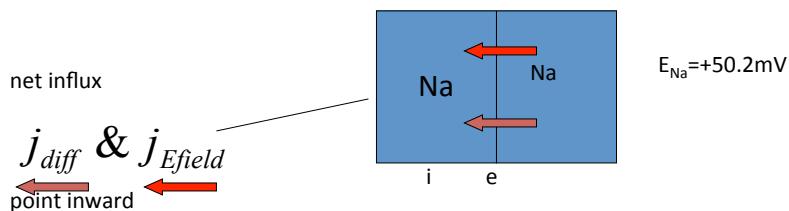
$$V_{m_rest} \approx E_K$$



What's true about permeabilities at peak?

$$P_{Na} \gg P_K \quad \text{since} \quad V_{m_peak} \approx E_{Na}$$

Recall that at rest ($V_m = -80\text{mV}$)



$$E_{Na} = +50.2\text{mV}$$

Everything is primed for Na to enter the cell at rest!

Let's define $(V_m - E_{Na})$ as the driving force for Na

At rest, the driving force is negative signifying an inward current

To get the current density, J_{Na} , we need multiply driving force by?

g_{Na} Conductance per unit area

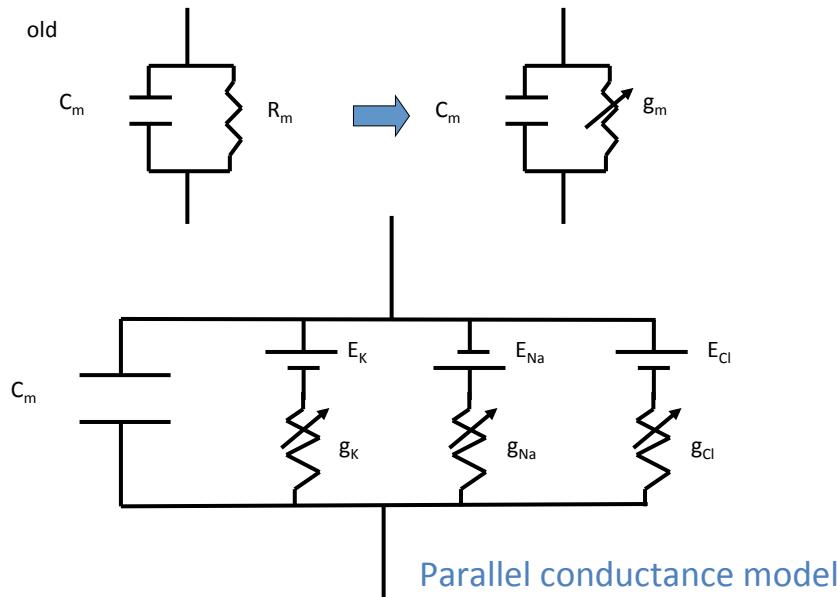
Units?

has units S/cm^2 , is positive, and varies during action potential

g_{Na}

$$J_{Na} = g_{Na} (V_m - E_{Na})$$

Improved model of excitable membrane



Total membrane current given by

$$J_m = g_K(V_m - E_K) + g_{Na}(V_m - E_{Na}) + g_{Cl}(V_m - E_{Cl}) + C_m \frac{dV_m}{dt}$$

At rest, $J_m=0$, $dV_m/dt=0$

capacitive current

$$V_m = \frac{g_k E_K + g_{Na} E_{Na} + g_{Cl} E_{Cl}}{g_k + g_{Na} + g_{Cl}}$$

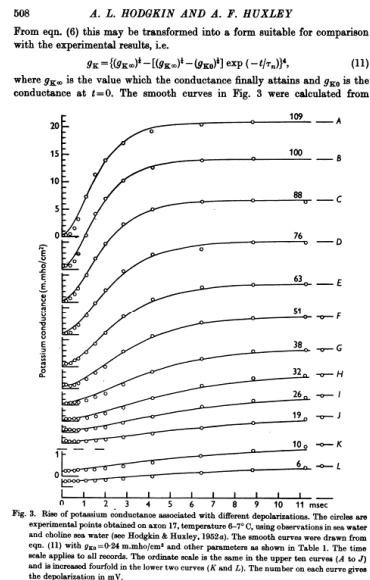
How can the ionic conductances be determined as a function of V_m ?

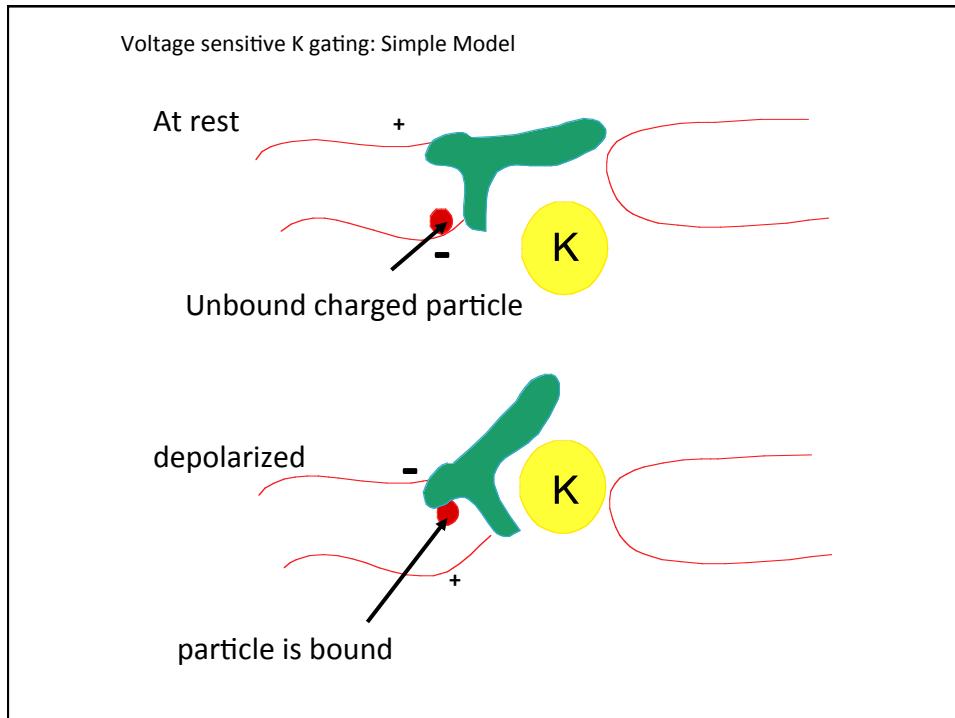
If V_m can be held constant, then since driving force ($V_m - E_{Na}$) is constant. As a result,

$$I_{Na} \propto g_{Na} \quad g_{Na} = I_{Na} / (V_m - E_{Na})$$

Might be able to measure total membrane current. How can the individual components be separated?

First build a system to hold V_m constant.





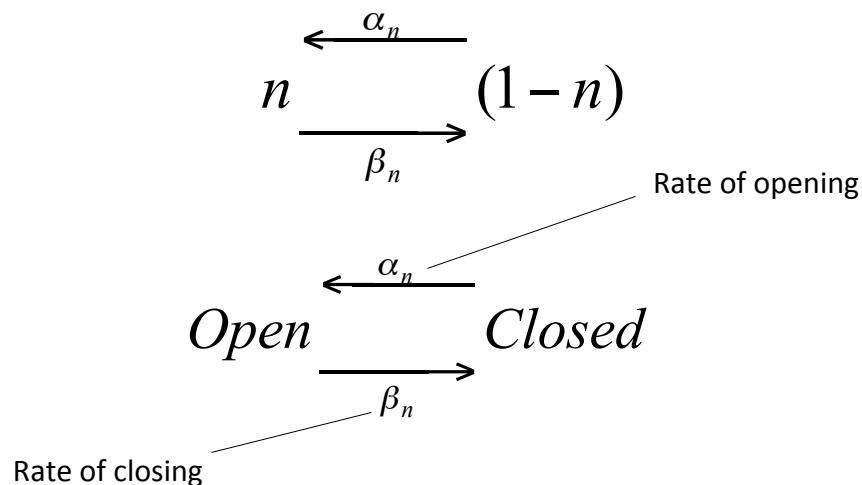
HH Gating Model

- Movement of gating particle is rate dependent
- At any given voltage, some particles are inside and some are outside. They move at some rate, depending on the membrane potential
- Binding can be explained with first order kinetics.

HH gating Model for K channel

- n - fraction of particles that are bound to the outside
- $(1-n)$ - fraction of particles that are unbound on inside
- β_n - rate constant for going from outside to inside. Open to closed state. Rate cnst of closing
- α_n - rate constant for going from inside to outside. Closed to open. Rate cnst of opening

Binding Kinetics can be written as:



The net rate of change of the fraction of particles that become bound (net rate of opening) is

$$\frac{dn}{dt} = \alpha_n(1-n) - \beta_n n$$

Net rate of change depends on the **difference** of the rate from going from closed to open and the rate going from open to closed.

At any given voltage clamp, alphas and betas are constant. Equation for n is a first order ODE with constant coefficients.

The analytical solution is

$$n(t) = n_\infty - (n_\infty - n_0)e^{-t/\tau_n}$$

n_0 is the fraction of particles that are bound before voltage clamp is applied.

n_∞ is fraction of particles that are bound at steady state after clamp is applied

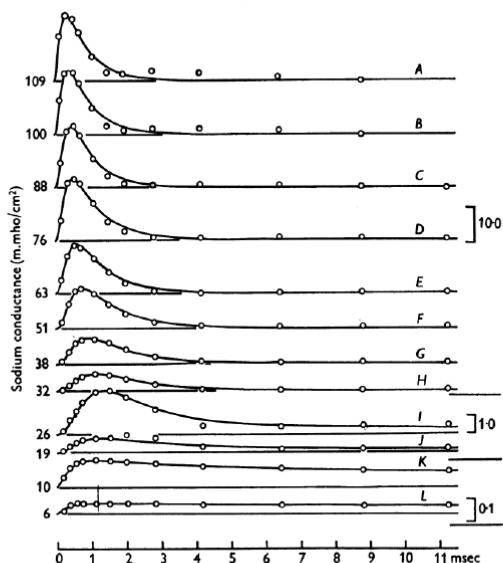
Both are obtained by setting $dn/dt = 0$!

$$\frac{dn}{dt} = 0 = \alpha_n(1 - n) - \beta_n n$$

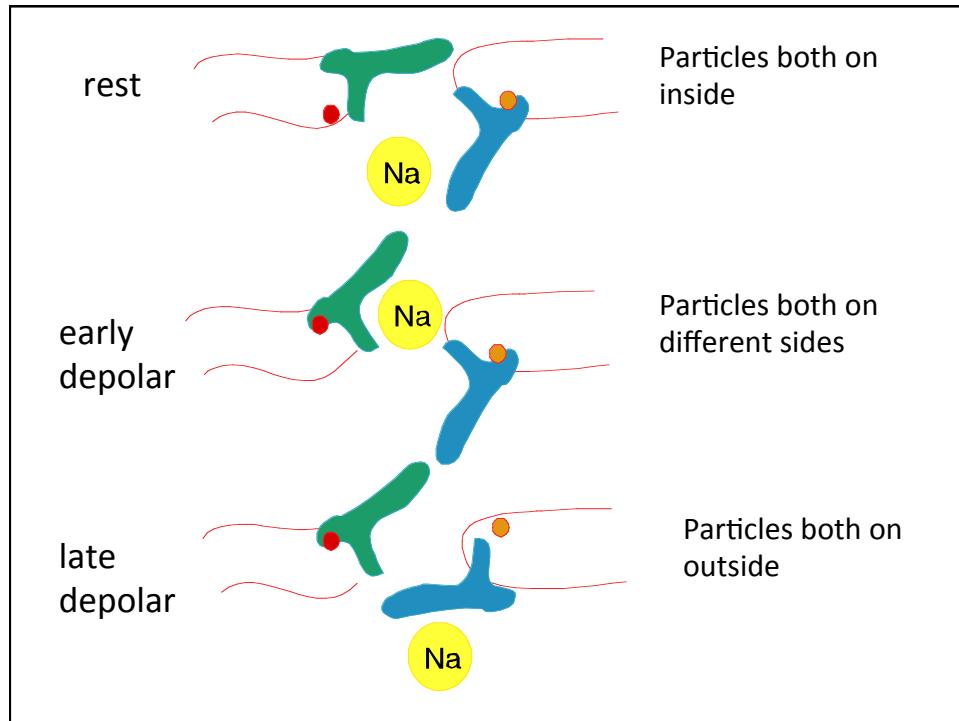
Where n was $n_0 = \frac{\alpha_n(V_0)}{\alpha_n(V_0) + \beta_n(V_0)}$

Where n wants
to be $n_\infty = \frac{\alpha_n(V_{Clamp})}{\alpha_n(V_{Clamp}) + \beta_n(V_{Clamp})}$

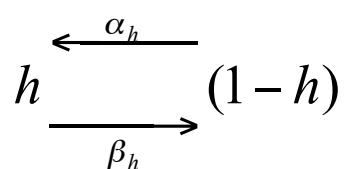
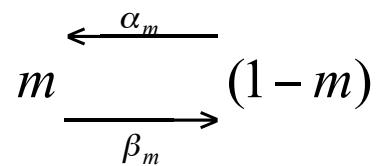
How fast n wants
to get there $\tau_n = \frac{1}{\alpha_n(V_{Clamp}) + \beta_n(V_{Clamp})}$



from Hodgkin & Huxley, J. Physiol. (1952)



Same binding Kinetics as used for K:



The net rate of change of the fraction of particles that become bound is

$$\frac{dm}{dt} = \alpha_m(1-m) - \beta_m m$$

$$\frac{dh}{dt} = \alpha_h(1-h) - \beta_h h$$

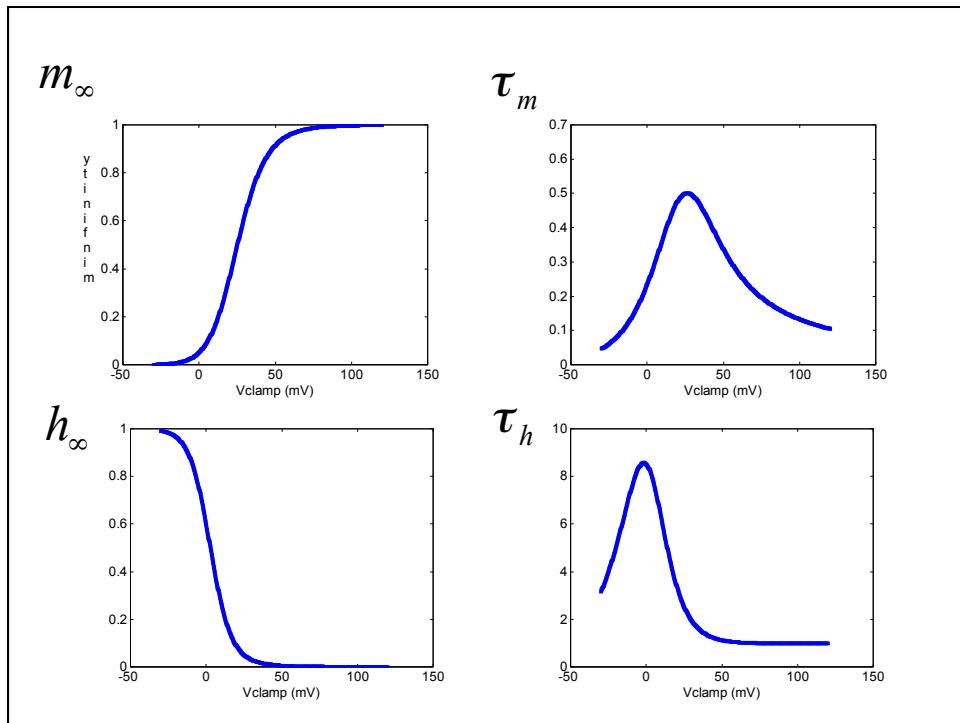
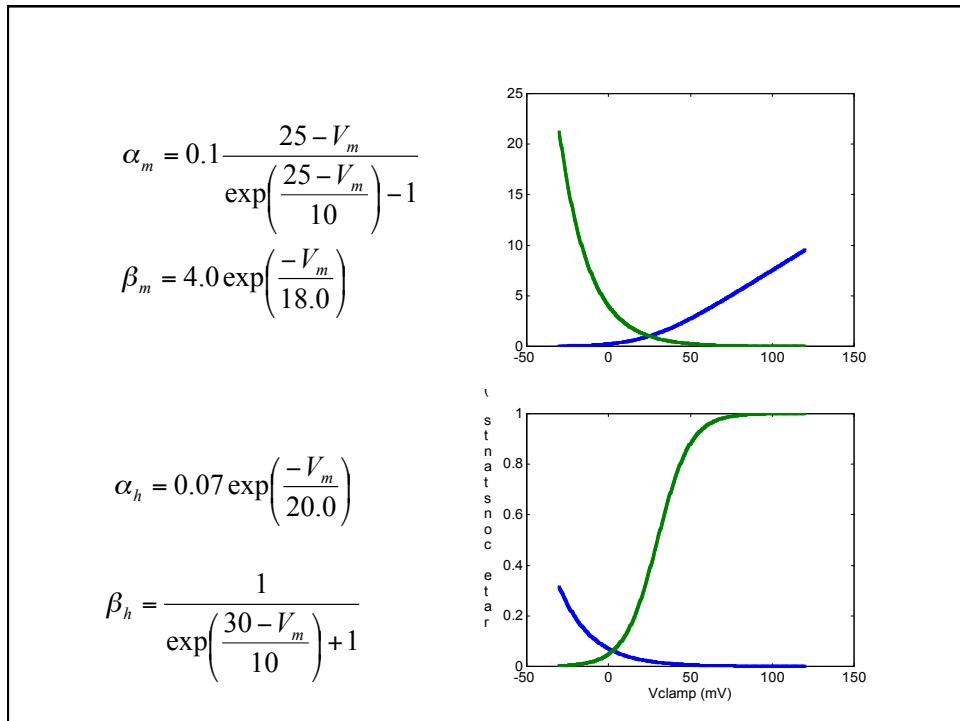
For activation and inactivation to work, α_m and α_h must have opposite behavior as a function of V_m

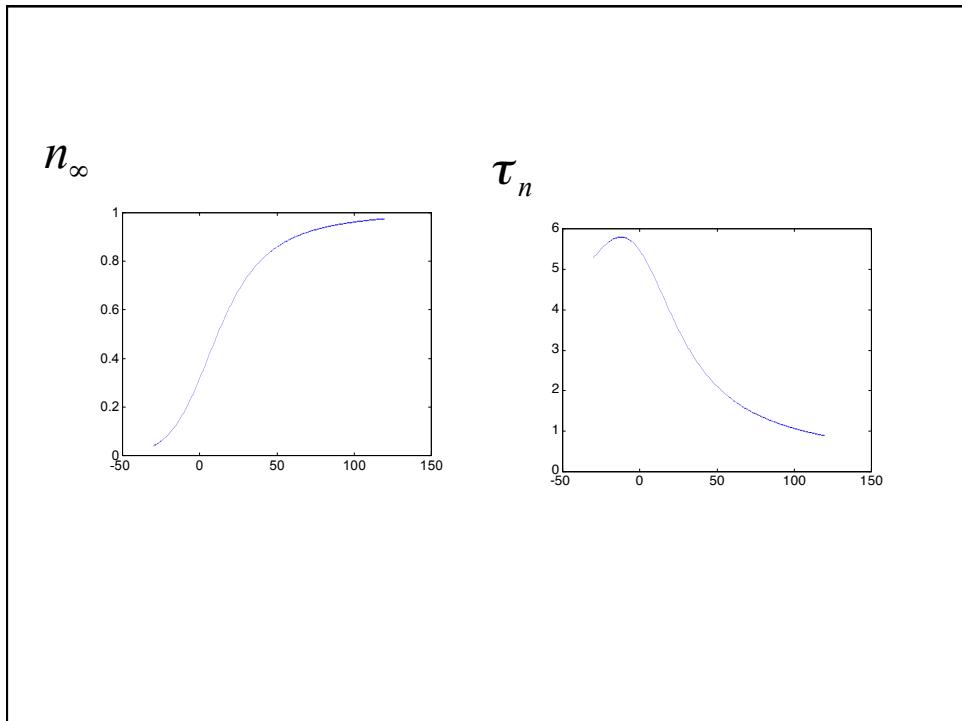
The analytical solutions are

$$m(t) = m_\infty - (m_\infty - m_0)e^{-t/\tau_m}$$

$$h(t) = h_\infty - (h_\infty - h_0)e^{-t/\tau_h}$$

Parameters have analogous forms to those obtained for n.





Putting it all together

$$\frac{dv_m}{dt} = -\frac{1}{C_m} (I_{Na} + I_K + I_L) \quad I_{Na} = \bar{g}_{Na} m^3 h (v_m - e_{Na})$$

$$\frac{dm}{dt} = \alpha_m (1 - m) - \beta_m m \quad I_K = \bar{g}_K n^4 (v_m - e_K)$$

$$\frac{dh}{dt} = \alpha_h (1 - h) - \beta_h h \quad I_L = \bar{g}_L (v_m - e_L)$$

$$\frac{dn}{dt} = \alpha_n (1 - n) - \beta_n n$$

No analytical solution! Must solve numerically.

Alphas and Betas

$$\alpha_m(V_m) = 0.1 \frac{25 - V_m}{\exp\left(\frac{25 - V_m}{10}\right) - 1},$$

$$\beta_m(V_m) = 4 \exp\left(-\frac{V_m}{18}\right);$$

$$\alpha_h(V_m) = 0.07 \exp\left(\frac{-V_m}{20}\right),$$

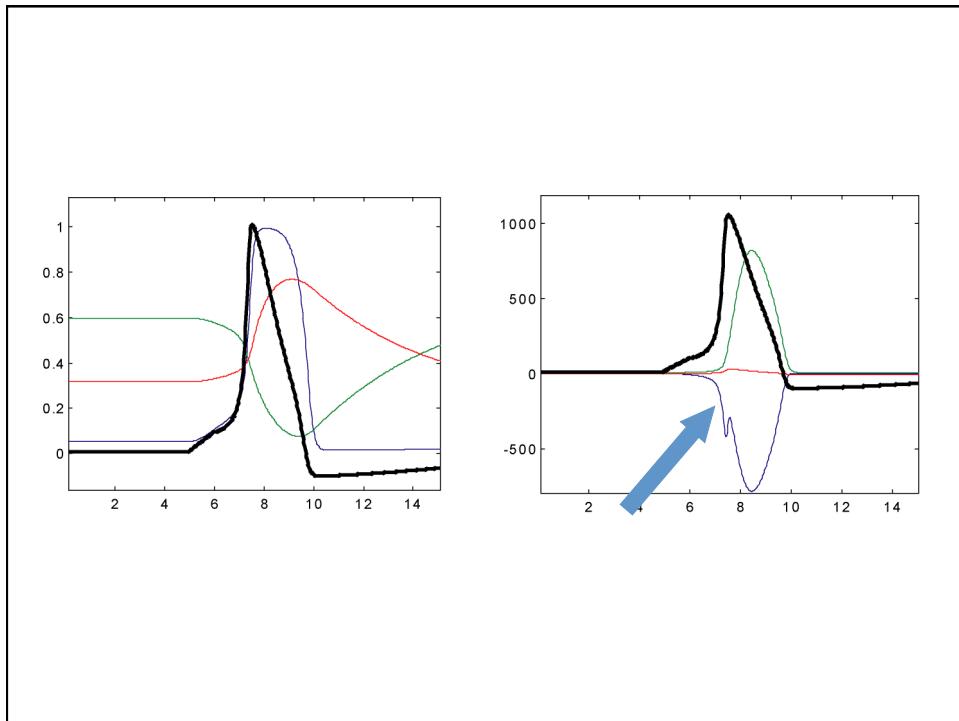
$$\beta_h(V_m) = \frac{1}{\exp\left(\frac{30 - V_m}{10}\right) + 1};$$

$$\alpha_n(V_m) = 0.01 \frac{10 - V_m}{\exp\left(\frac{10 - V_m}{10}\right) - 1},$$

$$\beta_n(V_m) = 0.125 \exp\left(-\frac{V_m}{80}\right);$$

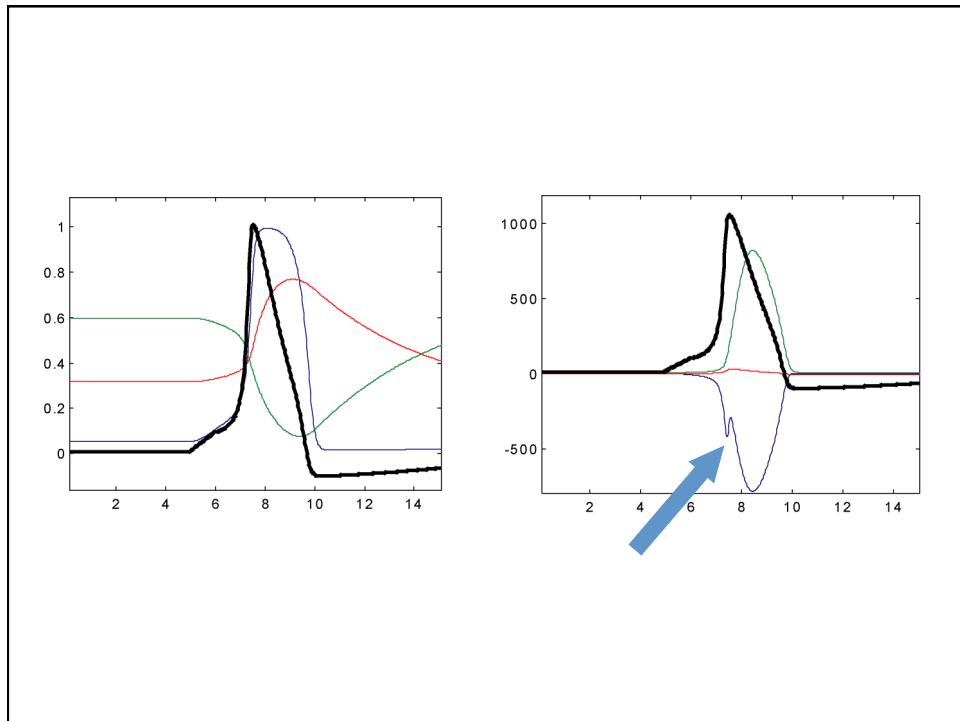
Qualitative view of an AP

- Stimulus causes membrane potential to rise.
- m increases, n increases and h decreases as V_m increases
- Number of bound m particles increases faster than number of bound n and unbound h since $\tau_m < \tau_n$ and τ_h
- I_{Na} increases since more channels are open and this acts to further increase V_m



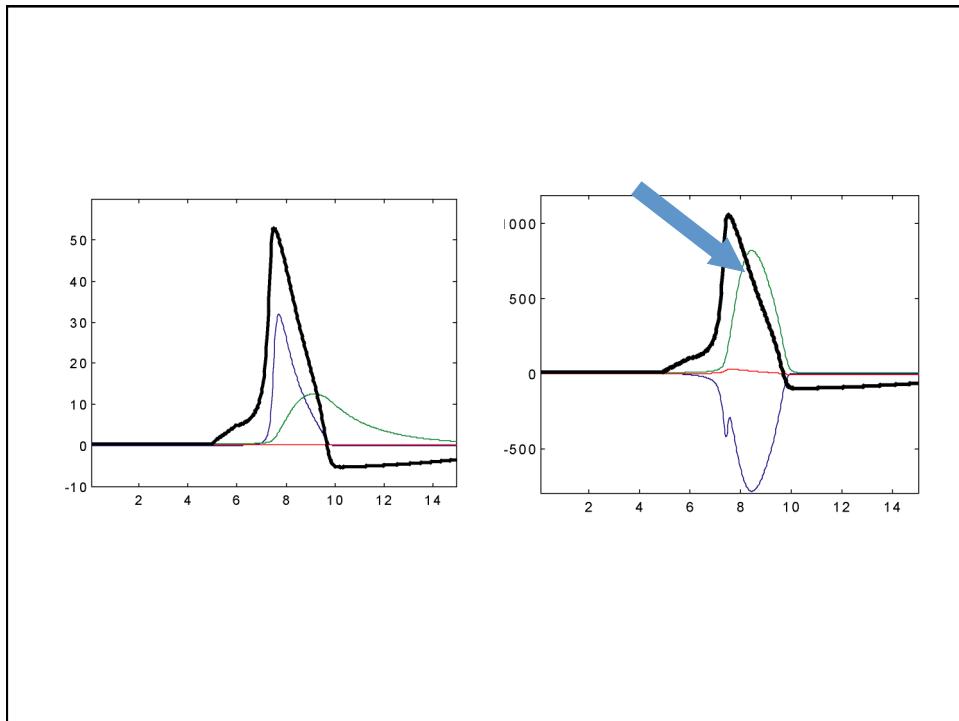
Qualitative view of an AP

- As V_m increases, the driving force (d.f.) for Na decreases. For I_{Na} to increase, the increase in g_{Na} must dominate the decrease in d.f.
- Eventually decreasing d.f. overwhelms increase in g_{Na} and I_{Na} decreases. The h particles are also tending to unbind as V_m increases but time constant is relatively slow.



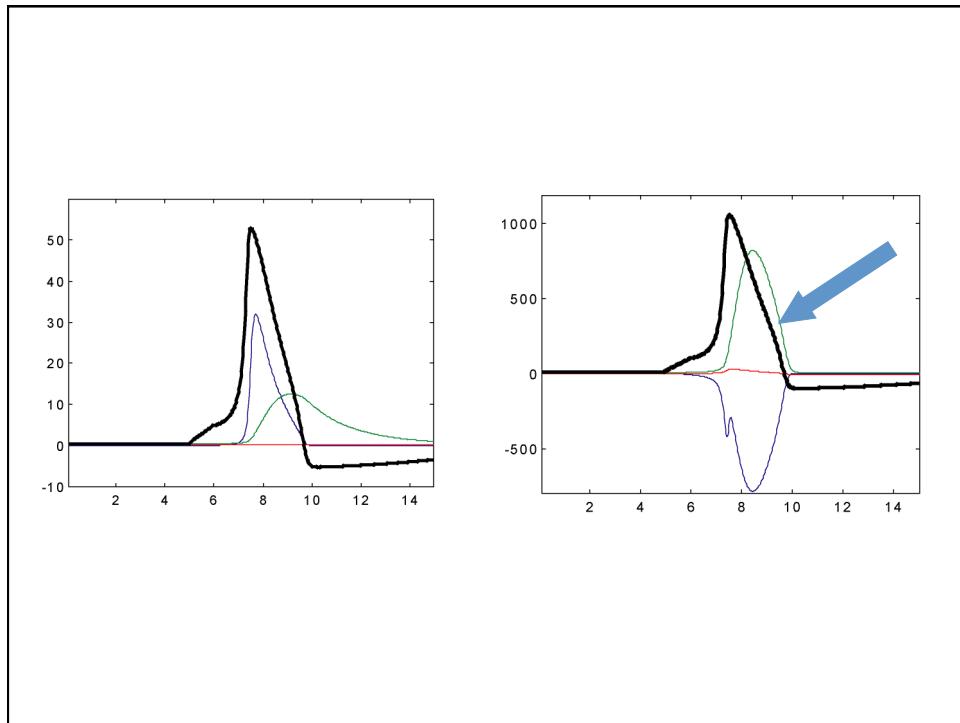
Qualitative view of an AP

- As V_m increases driving force (d.f.) for K increases. g_K increases slowly due to n^4 and large τ_n . I_K increases acting to push V_m toward E_K .
- At peak of A.P., $I_{ion}=0$ since no capacitive current at peak since $dV_m/dt=0$.

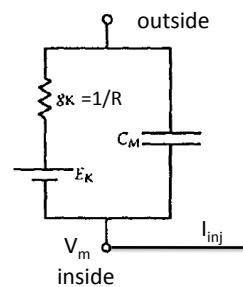
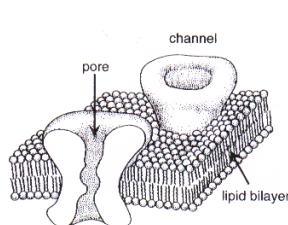


Qualitative view of an AP

- After peak, g_{Na} decreases due to inactivation (h unbinding). As V_m moves toward E_K , d.f. for Na increases but decrease in g_{Na} overwhelms and I_{Na} decreases.
- Increases in g_K overwhelms decrease in d.f. for K. Increase I_K dominates I_{Na} and V_m returns toward E_K and eventually equilibrates to V_{rest} .



The membrane equation

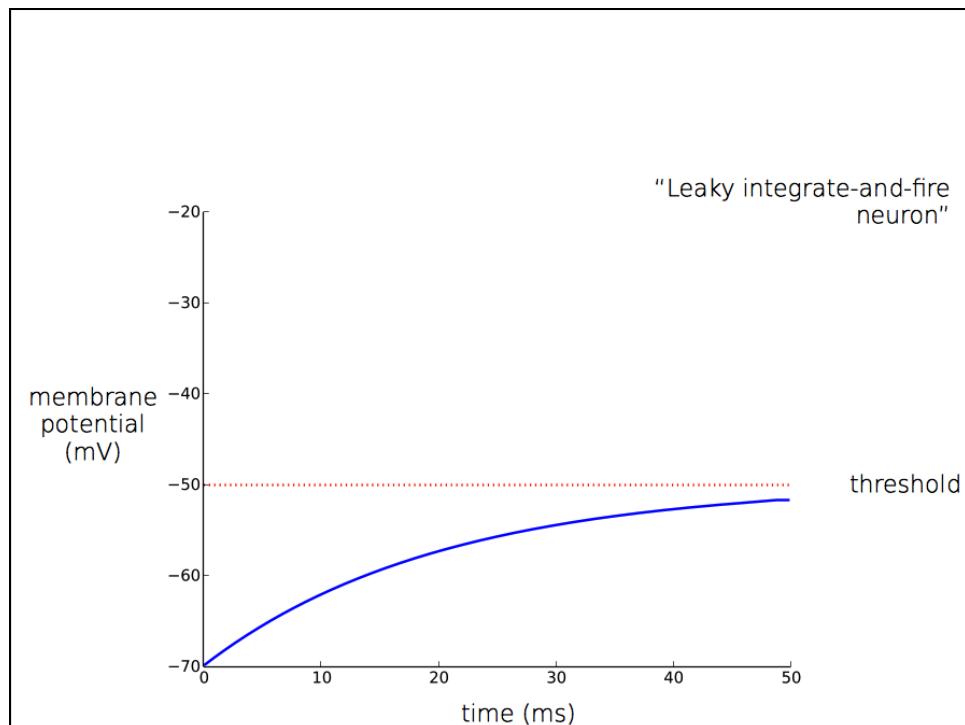
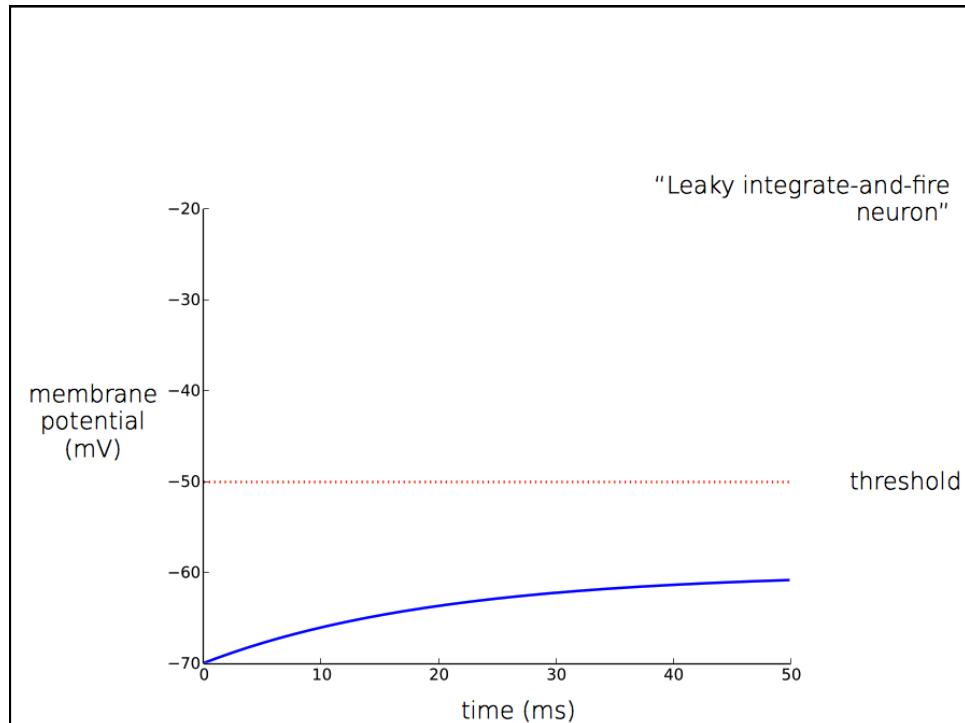


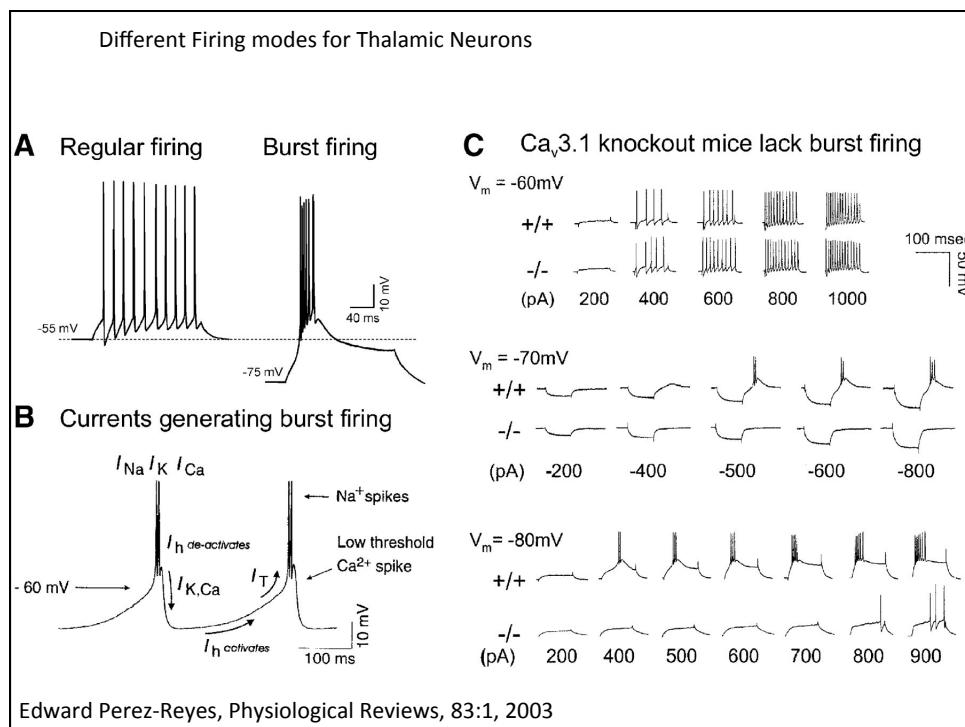
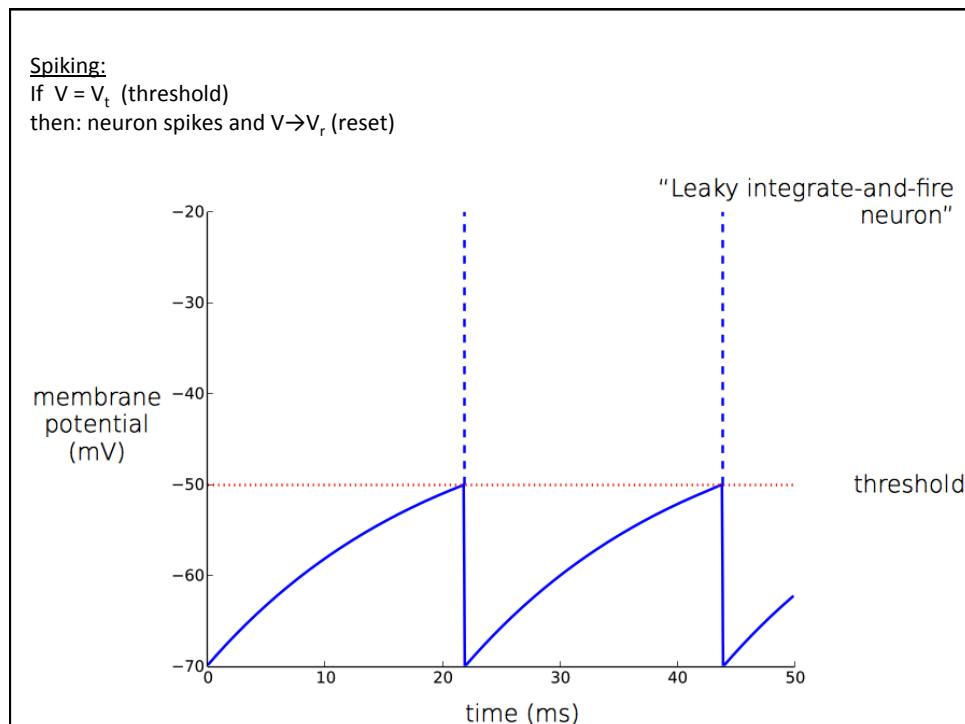
$$C \frac{dV_m}{dt} + \frac{(V_m - E_K)}{R} = I_{inj}$$

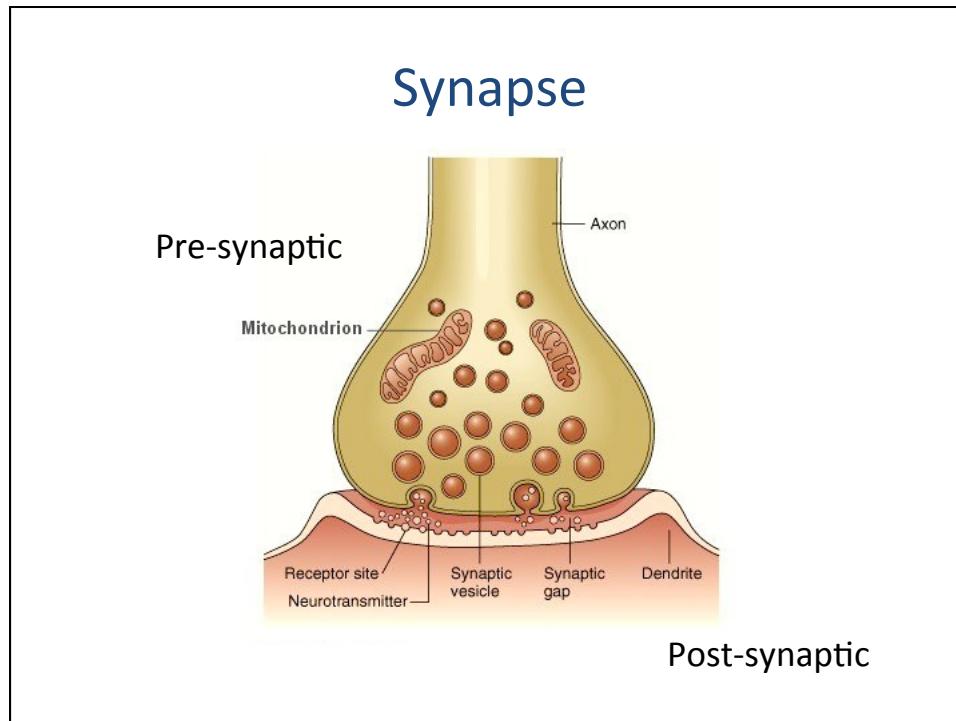
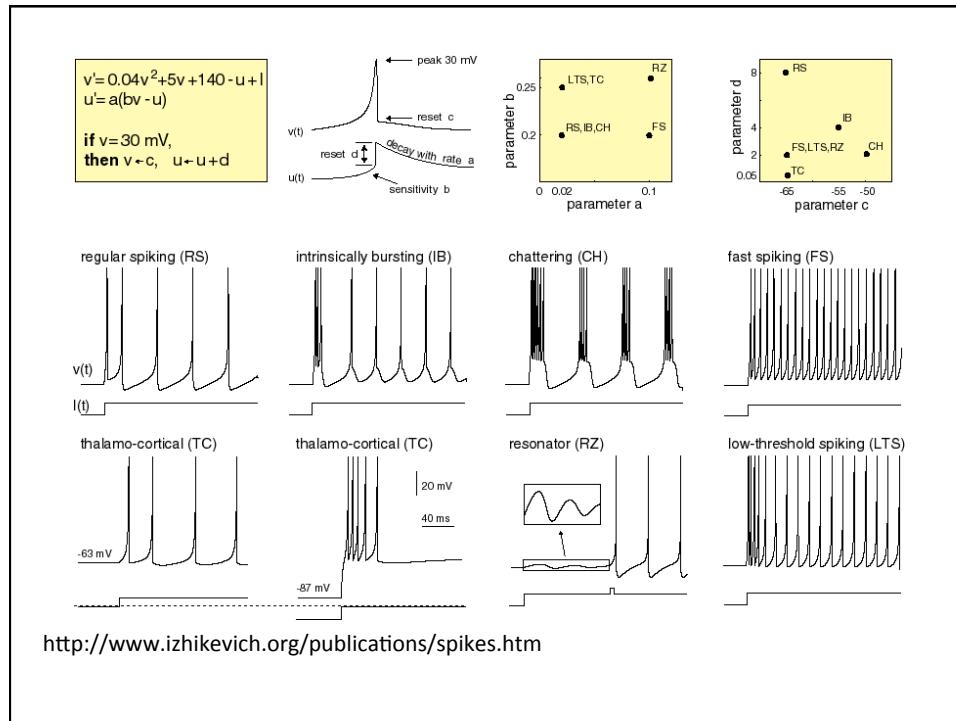
$$\tau \frac{dV_m}{dt} = E_K - V_m + RI_{inj}$$

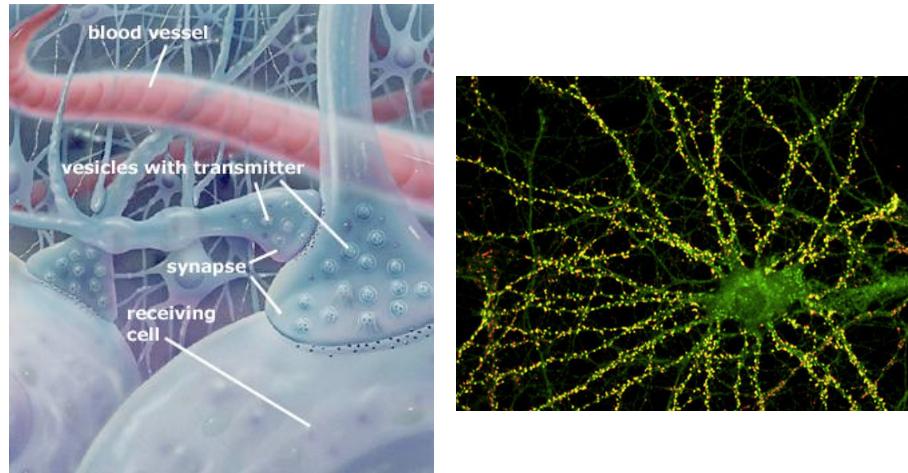
$$\tau = RC$$

membrane time constant
(typically 3-10 ms)









Synapse

- Functional chemical connection between neurons through close contact (20-50 nm)
- to dendrite; to soma; to hillock : to dendrite.
- Neurons may have 1000's of synapses
- vesicles containing neurotransmitter 40 nm

Synapse- Transmission

- Action potential reaches pre-synaptic terminal
- pre-synaptic potential opens voltage dependent Ca channels
- Ca causes vesicle migration, fusion to pre-synaptic membrane
- transmitter release via exocytosis - release in quantized amounts
- transmitter substance diffuses across cleft toward post-synaptic membrane

Synapse- Transmission (cont)

- Transmitter binds to receptor sites, causing membrane permeability change to ions
- post-synaptic potential is generated; can be EPSP (excitatory) or IPSP (inhibitory), depending on synapse type
- if excitatory and EPSP exceeds threshold, action potential is generated
- transmitter is broken down by enzyme in cleft
- products of hydrolysis are taken up by presynaptic site

Synapse - characteristics

- Delay time for vesicle release, diffusion and binding (approx 0.5 msec)
- unidirectional action- propagation is from pre to post-synaptic fiber
- response is graded- proportional to rate of release of neurotransmitter, firing rate of neurons
- shows summation: temporal (same synapse) or spatial (different synapses at same time)
- shows fatigue: depletion of neurotransmitter

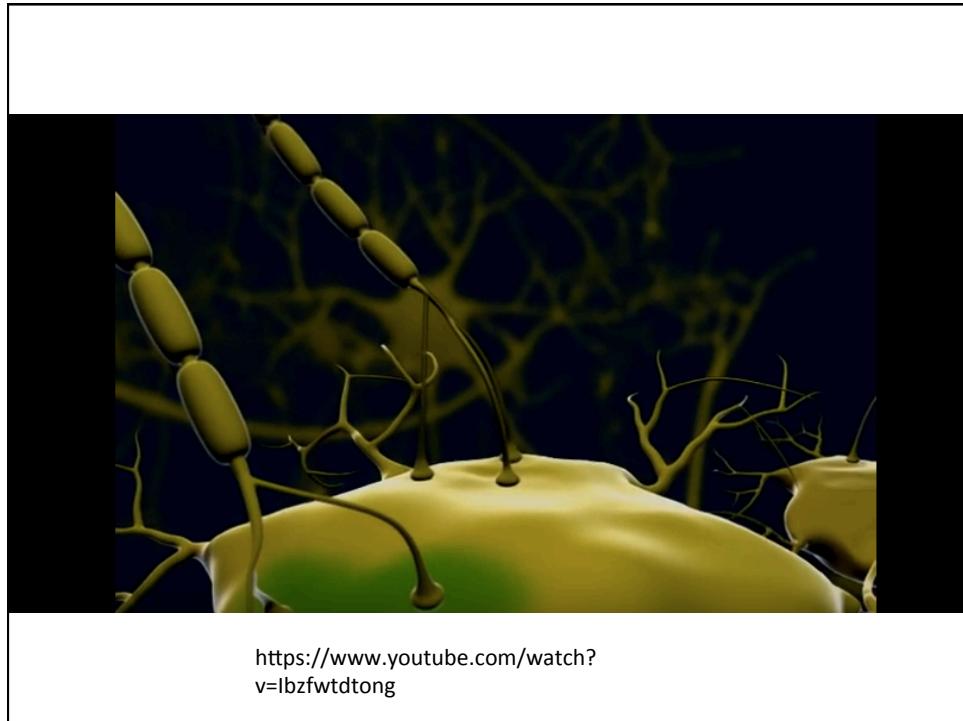
Synapse - neurotransmitters

- Glutamate- excitatory -
 - AMPA/kainate (fast, 1msec rise, 5 msec fall) receptors
 - NMDA (slow, 20 msec rise and 25-125 msec fall) receptors
- γ - aminobutyric acid (GABA) is primary inhibitory neurotransmitter
 - GABA_A - fast inhibition receptors
 - GABA_B - uses second messenger (slow) receptors

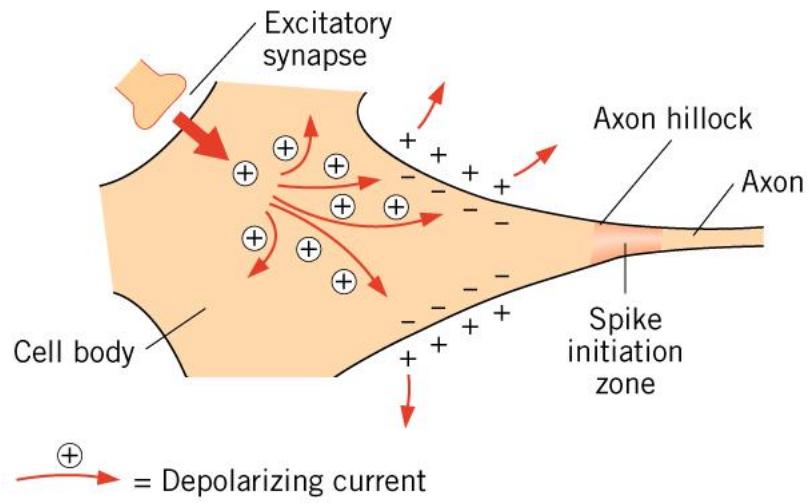
Synapse - neuromodulators

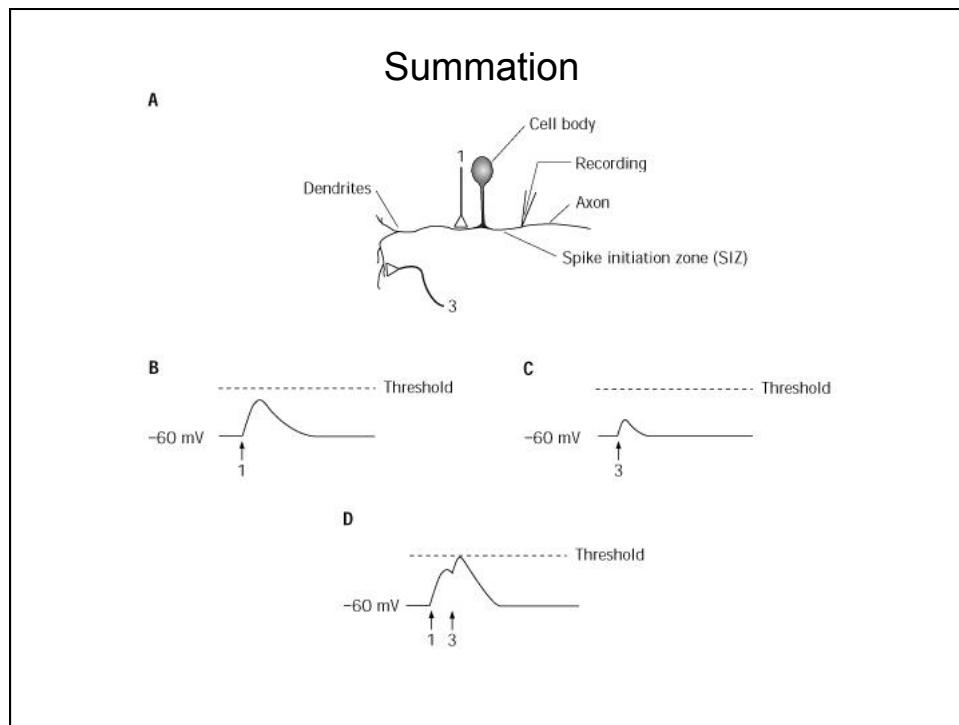
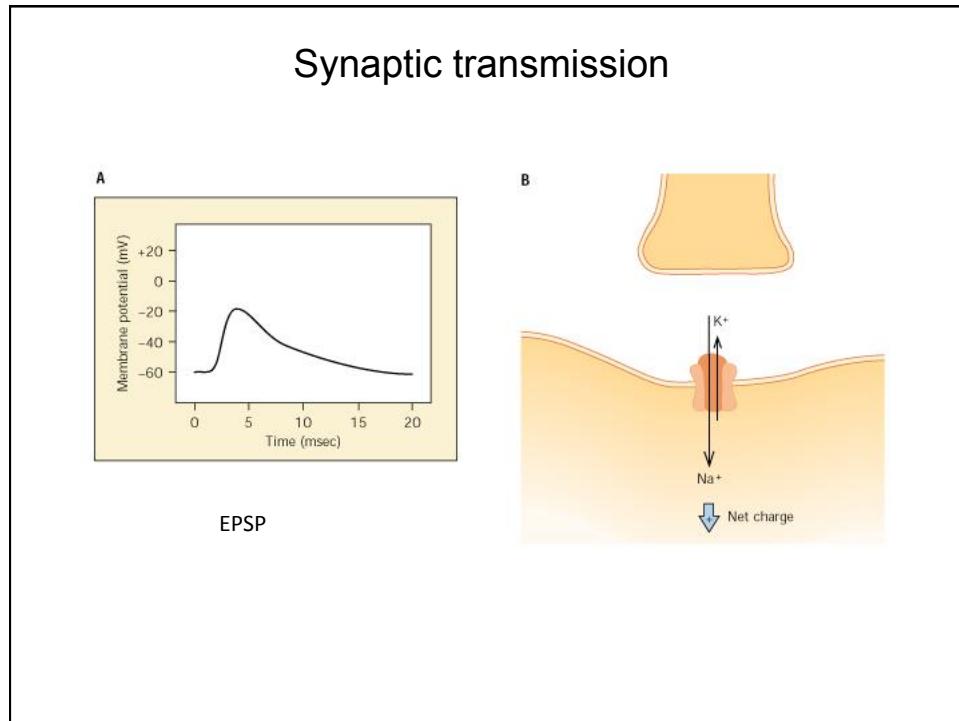
- neuromodulatory transmitters are secreted by a small group of neurons and diffuse through large areas of the nervous system, having an effect on multiple neurons. Examples of neuromodulators include dopamine, serotonin, acetylcholine, histamine and others.

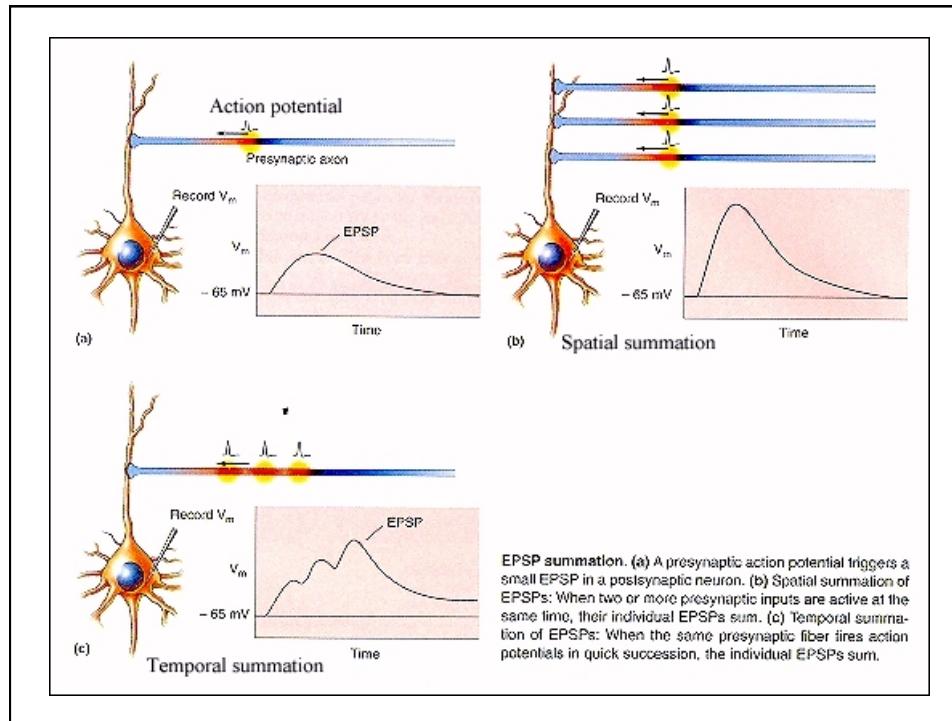




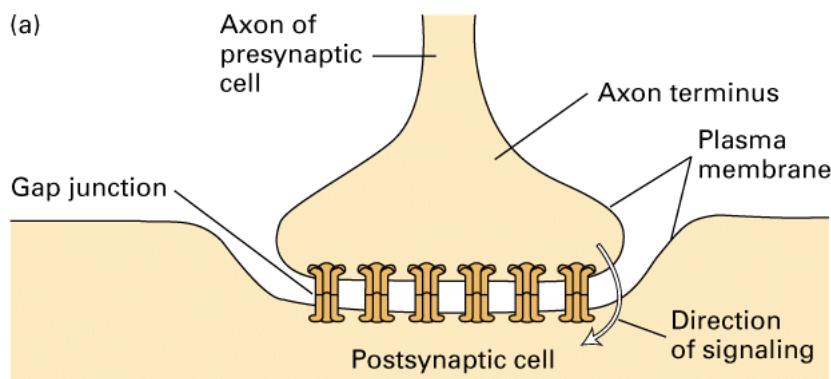
Synaptic currents drive spike generation







Electrical Connections- Gap Junctions



Neuroplasticity

- Neuroplasticity (sometimes called brain plasticity, cortical plasticity or cortical remapping) refers to the changes that occur in the organization of the brain as a result of experience or learning. Can be a change in the wiring. Can be a change in the strength of connections.

CNS Tutorial T3

Modelling of spiking neural networks with the Brian simulator

Marcel Stimberg

Dan Goodman

An example

```

from brian2 import *
tau_m = 20*ms
tau_e = 5*ms
tau_i = 10*ms
V_th = -50*mV
E_L = -60*mV
I_const = 11*mV
eqs = '''
dv/dt = (-v-E_L) + I_const + g_e + g_i : volt
dg_e/dt = -g_e/tau_e : volt
dg_i/dt = -g_i/tau_i : volt
'''
P = NeuronGroup(4000, model=eqs,
                 threshold='v>V_th', reset='v=E_L')
P.v = 'E_L+10*mV*rand()'
Pe = P[3200]
Pi = P[3200:]

w_e = 1.62*mV
w_i = 9*mV
Ce = Synapses(Pe, P, pre='g_e+=w_e')
Ci = Synapses(Pi, P, pre='g_i-=w_i')
Ce.connect(True, p=0.02)
Ci.connect(True, p=0.02)

M = SpikeMonitor(P)

run(1*second)
plot(M.t/ms, M.i, '.')
xlabel('Time (in ms)'); ylabel('Neuron number')
show()

```

Stimberg M, Goodman DFM, Benichoux V, Brette R (2014). **Equation-oriented specification of neural models for simulations**. Frontiers Neuroinf, doi: 10.3389/fninf.2014.00006.

$$\begin{aligned}\tau_m \frac{dV}{dt} &= -(V - E_L) + g_e + g_i \\ \tau_e \frac{dg_e}{dt} &= -g_e \\ \tau_i \frac{dg_i}{dt} &= -g_i\end{aligned}$$

Website

briansimulator.org

About

* Brian 2.0 fourth beta release: [release notes](#) *
* Brian tutorial at the CNS 2015 meeting in Prague: [more information](#) *

Brian is a simulator for spiking neural networks available on almost all platforms. The motivation for this project is that a simulator should not only save the time of processors, but also the time of scientists.

Brian is easy to learn and use, highly flexible and easily extensible. The Brian package itself and simulations using it are all written in the Python programming language, which is an easy, concise and highly developed language with many advanced features and development tools, excellent documentation and a large community of users providing support and extension packages.

The following code defines a randomly connected network of integrate and fire neurons with exponential inhibitory and excitatory currents, runs the simulation and makes the raster plot on the right.

```

1 | from brian2 import *
2 | eqs = '''
3 | dv/dt = (ge+gi-(v+49*mV))/(20*ms) : volt
4 | dge/dt = -ge/(5*ms) : volt
5 | dgi/dt = -gi/(10*ms) : volt
6 |
7 | P = NeuronGroup(4000, eqs, threshold='v>-50*mV', reset='v=-60*mV')
8 | P.v = -60*mV
9 | Pi = P[3200]
10 | Pe = P[3200]
11 | Ce = Synapses(Pe, Pi, pre='ge+=1.62*mV')
12 | Ci = Synapses(Pi, Pe, pre='gi-=9*mV')
13 | Ce.connect(True, p=0.02)
14 | Ci.connect(True, p=0.02)
15 | M = SpikeMonitor(P)
16 |
17 | run(1*second)
18 | plot(M.t/ms, M.i, '.')
19 | show()

```

See the manuals for more examples.

Documentation

brian2.readthedocs.org

Brian 2 documentation

Contents:

- [Introduction](#)
 - Installation
 - Release notes
 - Changes from Brian 1
 - Known issues
- [Tutorials](#)
 - Introduction to Brian part 1: Neurons
 - Introduction to Brian part 2: Synapses
- [User's guide](#)
 - Importing Brian
 - Physical units
 - Models and neuron groups
 - Equations
 - Refractoriness
 - Synapses
 - Input stimuli
 - Recording during a simulation
 - Running a simulation
 - Computational methods and efficiency
 - Functions
 - Devices
 - Brian 1 Hears bridge

Mailing lists

briansupport@googlegroups.com brian-development@googlegroups.com

Groups

Google Groups Search for topics NEW TOPIC C Mark all as read Actions Filters

◀ Brian Shared publicly 29 of many topics ★ [Ann]

This group does not have a welcome message.

Add welcome message

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<input type="checkbox"/>	★ [Ann] Fourth Brian 2.0 beta release	By me - 1 post - 0 views - updated 17 Jul
<input type="checkbox"/>	★ Constrain voltage-dependant rate constants	By AmelieND - 3 posts - 2 views - updated 15 Jul
<input type="checkbox"/>	★ m...	By Roberto Mulet - 5 posts - 3 views - updated 10 Jul
<input type="checkbox"/>	★ Real-time plotting in latest Brian or Brian2?	By Mohan - 9 posts - 10 views - updated 15 Jul
<input type="checkbox"/>	★ Converting code from Brian1 to Brian2	By Marjan R - 3 posts - 6 views - updated 14 Jul
<input type="checkbox"/>	★ Referencing linked variable in post-synaptic neuron	By sharbatani44@gmail.com - 2 posts - 7 views - updated 9 Jul
<input type="checkbox"/>	★ Explanation required for parameters x & y	By Himanshu Rajmne - 2 posts - 4 views - updated 7 Jul
<input type="checkbox"/>	★ Learning in Brian	By Maria Kesa - 4 posts - 5 views - updated 6 Jul
<input type="checkbox"/>	★ Re: [Brian] Thick spikes in HH model	By me - 1 post - 8 views - updated 30 Jun

Anatomy of a Brian script

```

from brian2 import *
tau_m = 20*ms
tau_e = 5*ms
tau_i = 10*ms
V_th = -50*mV
E_L = -60*mV
I_const = 11*mV
eqs = '''
dv/dt = (-v-E_L) + I_const + g_e + g_i : volt
dg_e/dt = -g_e/tau_e : volt
dg_i/dt = -g_i/tau_i : volt
'''
P = NeuronGroup(4000, model=eqs,
                 threshold='v>V_th', reset='v=E_L')
P.v = 'E_L+10*mV*rand()'
Pe = P[3200]
Pi = P[3200:]

w_e = 1.62*mV
w_i = 9*mV
Ce = Synapses(Pe, P, pre='g_e+=w_e')
Ci = Synapses(Pi, P, pre='g_i-=w_i')
Ce.connect(True, p=0.02)
Ci.connect(True, p=0.02)

M = SpikeMonitor(P)

run(1*second)
plot(M.t/ms, M.i, '.')
xlabel('Time (in ms)'); ylabel('Neuron number')
show()

```

Import Brian library
Set constants
Specify the neuronal model
Initialise state variables
Specify subgroups of neurons
Specify synaptic model
Connect neurons
Record activity
Run the simulation
Plot the activity

Constants

- Constants defined outside of strings can be used inside strings (only scalar values):

```

tau = 10*ms
G = NeuronGroup(1, 'dv/dt = -v / tau : volt')

```

- Values of constants are resolved at the **run** call:

```

tau = 10*ms
G = NeuronGroup(1, 'dv/dt = -v / tau : volt')
run(100*ms)                                will be used for second run
tau = 20*ms
run(100*ms)

```

Variables and equations

Equations define *state variables* of an object:

```
tau = 10*ms
G = NeuronGroup(5, '''dv/dt = (-v + inp) / tau : volt (unless refractory)
                    inp = a * clip(sin(2*pi*freq*t), 0, inf) : volt
                    freq : Hz (constant)
                    a : volt (shared)
                ''', threshold='v>20*mV', reset='v = 0*mV',
                refractory=2*ms)
G.freq = [100, 200, 300, 400, 500] * Hz
G.a = 100*mV
G.v = 10*mV
print G.inp # not a state variable, but can be accessed like one
```

Additional variables are defined automatically

```
>>> print(G.variables.keys())
['a', '_spikespace', 'i', 'inp', 'N', 't', 'v', 'dt', 'lastspike', 'freq', 'not_refractory']
```

neuron index number of neurons time step time of last spike refractory?

Variables and equations

A Hodgkin-Huxley equations

```
G = NeuronGroup(number_of_neurons,
    '''dv/dt = (I_L+I_Na+I_K)/C_m : volt # membrane potential
        I_L = g_L*(-v+E_L) : amp # passive current
        I_Na = g_Na*(m**3)*h*(-v+E_Na) : amp # sodium current
        I_K = g_K*(n**4)*(v-E_K) : amp # potassium current
    ...# equations for n, m, h'''')
```

special variable
provided for noise

B Noisy membrane

```
G = NeuronGroup(number_of_neurons,
    'dv/dt = -(v-v_0)/tau_m +
        tau_m=0.5*3*mV*x1 : volt # membrane potential')
```

C Leaky integrate-and-fire neuron

```
G = NeuronGroup(number_of_neurons,
    'dv/dt = -(v-v_0)/tau_m : volt # membrane potential',
    threshold='v > v_th', reset='v = v_0')
```

D Leaky integrate-and-fire neuron with adaptive threshold

```
G = NeuronGroup(number_of_neurons,
    '''dv/dt = -(v-v_0)/tau_m : volt # membrane potential
        dv_th/dt = -(v_th-v_th0)/tau_th : volt # threshold',
        threshold='v > v_th', reset='v = v_0
            v_th += 3*mV'''')
```

Expressions

- Can be used to
 - Specify conditions (threshold, refractoriness, synaptic connection)

```
G = NeuronGroup(10, 'dv/dt = -v / tau : 1 (unless refractory)',
                 threshold='v > 1', refractory='(t-lastspike) < 3*ms')
```

- Index and set state variables

```
min_freq = 100*Hz
print G.v['freq > min_freq']
G.v = '0*mV + rand()*10*mV'
```

- Can refer to state variables, constants, units

Brian's unit system

- Brian allows to use units for scalars and vectors

```
>>> E_L = -70*mV
>>> print E_L
-70.0 mV
>>> freqs = [100, 200, 300] * Hz
>>> print freqs
[ 100.  200.  300.] Hz
```

- Most numpy functions work correctly with units
(make sure to not import from numpy directly)

```
>>> mean(freqs)
200.0 * hertz
>>> diff(freqs)
array([ 100.,  100.]) * hertz
```

Brian's unit system

- To remove units, use `numpy.asarray` or divide by the unit

```
>>> print freqs/Hz  
[ 100. 200. 300.]  
>>> print asarray(freqs)  
[ 100. 200. 300.]
```

- For state variables: adding an underscore returns unitless value

```
>>> print G.v  
<neurongroup.v: array([-70., -70., -70., -70., -70.]) * mvolt>  
>>> print G.v_  
<neurongroup_1.v: array([-0.07, -0.07, -0.07, -0.07, -0.07])>
```

- Unit consistency is also checked in equations, expressions and abstract code statements