

December 3, 2023

Initial selection:  $rAMP \text{ diff} > 2$  |  $\text{phase diff} > 6$  - since  $rAMP$  values are  $< 1$ , not filtered by  $rAMP$  here

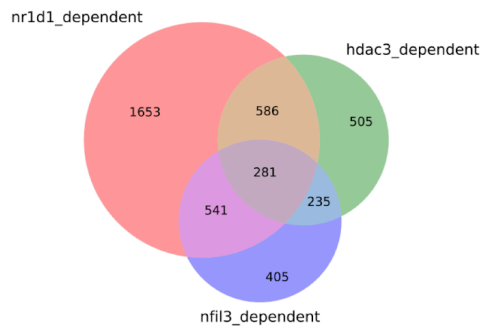


Figure 1. Phase diff > 6 only

Modifying the parameters in dependent.py below

```
import pandas as pd

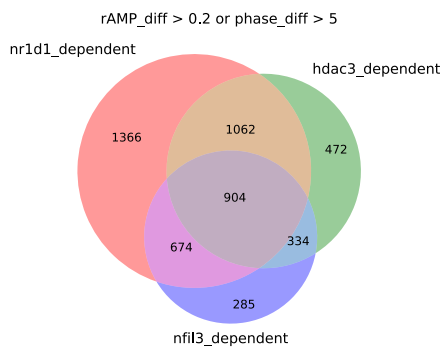
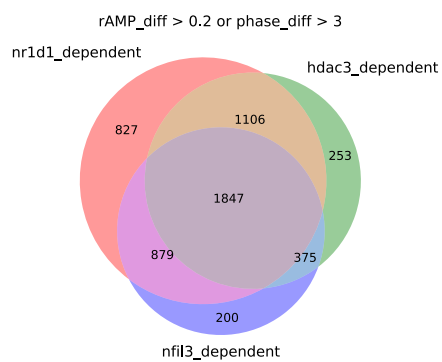
gene = ["hdac3", "nfil3", "nr1d1"]

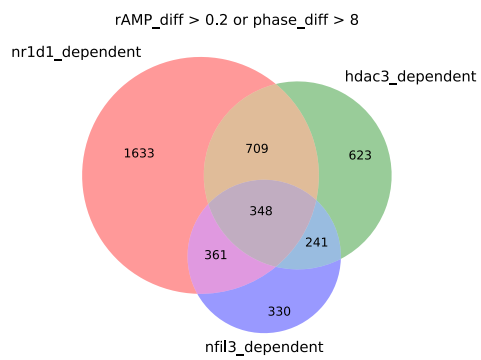
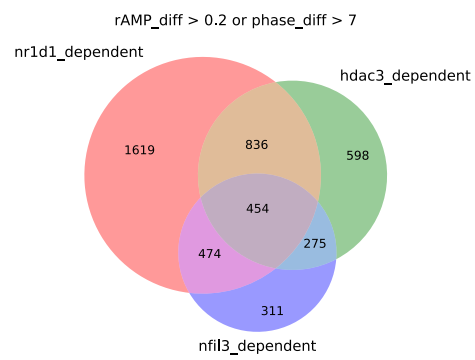
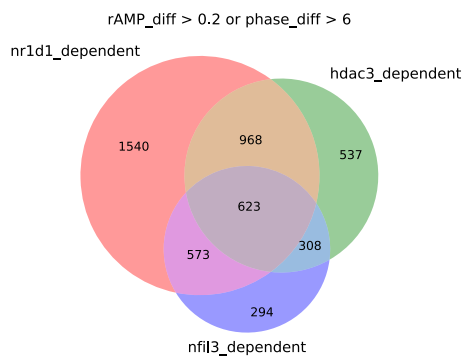
for g in gene:
    difference_df = pd.read_csv(g+"_difference_modify.txt", sep='\t')
    dependent_df = difference_df[(difference_df['meta2d_rAMP_diff'] > 0.3) | (difference_df['meta2d_phase_diff'] > 7)]

    print(len(dependent_df))

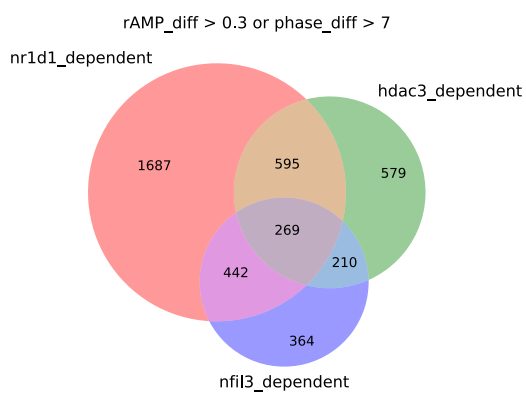
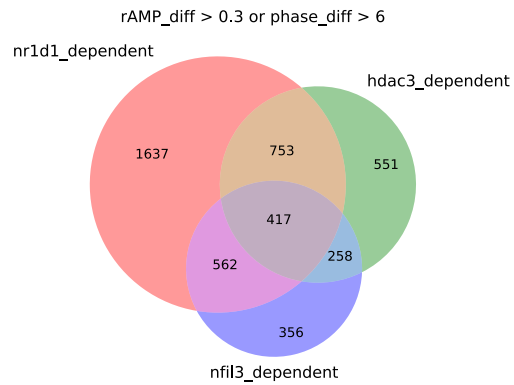
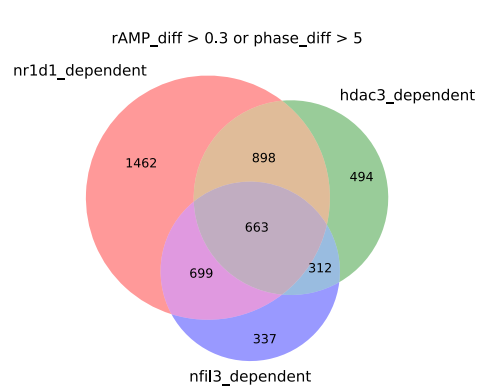
    dependent_df.to_csv(g+"_dependent_modify_0.3_7.txt", sep='\t', index=False)
```

### Results with different parameters in OR setting



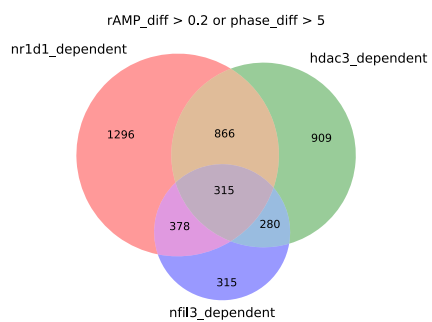
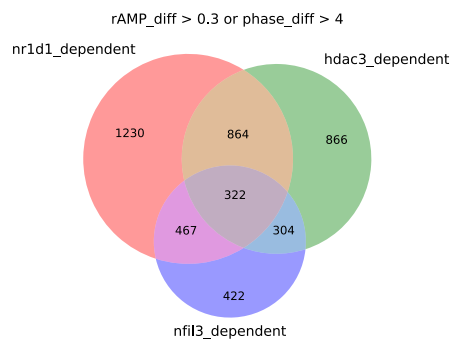
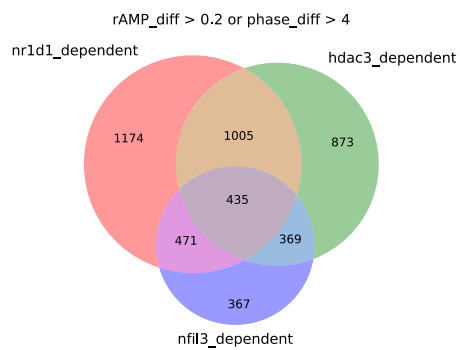
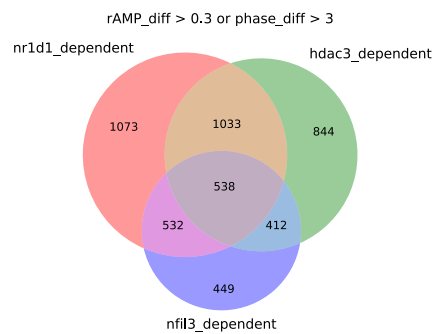
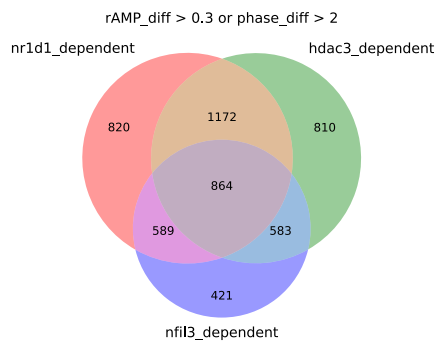
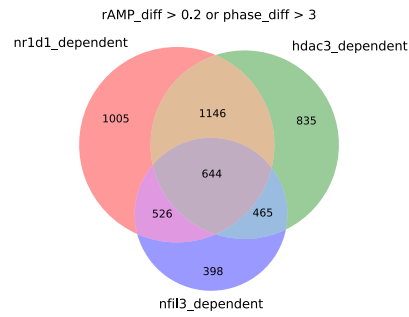
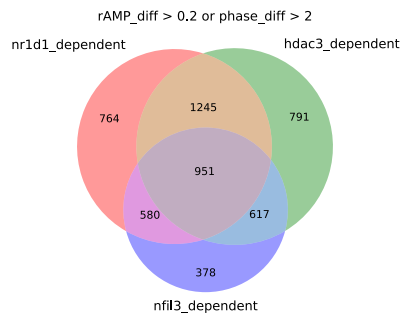


skip 7

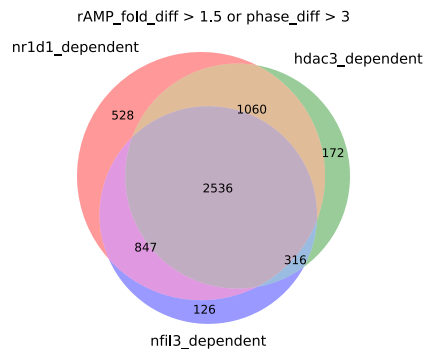
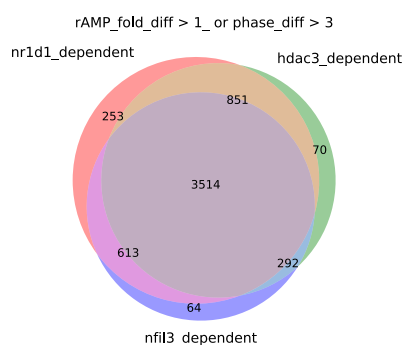


December 11, 2023

From meta2d data, select genes that have p val < 0.05 in WT (filter out the rest in both WT and KO). P-value determines if the genes are cycling (are circadian genes)



Modified rAMP difference calculation from absolute ( $\text{abs}(\text{KO}-\text{WO})$ ) to relative ( $\text{WO}/\text{KO}$ )



Feb 3<sup>rd</sup> 2024

Tophat – align -> cuffdiff to calculate FPKM values

Dr. Kuang ran cuffdiff with 32 files total – 4 GF, 8 Hdac3, 8 Nr1d1, 12 Nfil3

```
cuffdiff -o ZK_4group -p 30 -u genes.gtf ./GF_mm10/ZK1_tophat_out/accepted_hits.bam ./GF_mm10/ZK2_tophat_out/accepted_hits.bam ./GF_mm10/ZK3_tophat_out/accepted_hits.bam ./GF_mm10/ZK4_tophat_out/accepted_hits.bam ./Hdac3_mm10/ZK1_tophat_out/accepted_hits.bam ./Hdac3_mm10/ZK2_tophat_out/accepted_hits.bam ./Hdac3_mm10/ZK3_tophat_out/accepted_hits.bam ./Hdac3_mm10/ZK4_tophat_out/accepted_hits.bam ./Hdac3_mm10/ZK5_tophat_out/accepted_hits.bam ./Hdac3_mm10/ZK6_tophat_out/accepted_hits.bam ./Hdac3_mm10/ZK7_tophat_out/accepted_hits.bam ./Hdac3_mm10/ZK8_tophat_out/accepted_hits.bam ./Nr1d1_mm10/ZK1_tophat_out/accepted_hits.bam ./Nr1d1_mm10/ZK2_tophat_out/accepted_hits.bam ./Nr1d1_mm10/ZK3_tophat_out/accepted_hits.bam ./Nr1d1_mm10/ZK4_tophat_out/accepted_hits.bam ./Nr1d1_mm10/ZK5_tophat_out/accepted_hits.bam ./Nr1d1_mm10/ZK6_tophat_out/accepted_hits.bam ./Nr1d1_mm10/ZK7_tophat_out/accepted_hits.bam ./Nr1d1_mm10/ZK8_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK1_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK2_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK3_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK4_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK5_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK6_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK7_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK8_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK9_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK10_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK11_tophat_out/accepted_hits.bam ./Nfil3_mm10/ZK12_tophat_out/accepted_hits.bam
```

output file to XueK/ZK\_4group -> genes.fpkms\_tracking

remove rows with FAIL in status column > genesFailRemoved.txt

1	tracking_id	class_code	nearest_ref_gene_id	gene_short_name	tss_id	locus	length	coverage	q1_FPKM	q1_conf_lo	q1_conf_hi	q1_status
5809	Cyp4f13	-	-	Cyp4f13	TSS10638	chr17:32924-	-	-	51.2917	11.4555	91.1279	OK
5810	Cyp4f14	-	-	Cyp4f14	TSS18626	chr17:32905-	-	-	1002.84	0	1036.21	FAIL
5811	Cyp4f15	-	-	Cyp4f15	TSS25797	chr17:32685-	-	-	0.111026	0	0.337876	OK
5812	Cyp4f16	-	-	Cyp4f16	TSS12179	chr17:32536-	-	-	218.487	41.3726	395.602	OK
5813	Cyp4f17	-	-	Cyp4f17	TSS2923	chr17:32506-	-	-	0.239663	0	0.694555	OK
5814	Cyp4f18	-	-	Cyp4f18	TSS15053	chr8:719884-	-	-	1.76286	0	4.23868	OK
5815	Cyp4f39	-	-	Cyp4f39	TSS16876	chr17:32452-	-	-	0.443671	0	1.07173	OK
NR161	Rundf1a	-	-	Rundf1a	TSS5038	chr17:32609-	-	-	30.1799	8.79535	70.0644	OK
(base) tinaryu@Tinas-MacBook-Pro-3 KuangLab % grep FAIL genes.txt   cu												
492865	SLNDRB1	-	-	SLNDRB1	-	-	-	-	-	-	-	-
Actr3	-	-	-	Actr3	-	-	-	-	-	-	-	-
B230312A22R1k	-	-	-	B230312A22R1k	-	-	-	-	-	-	-	-
BC065397	-	-	-	BC065397	-	-	-	-	-	-	-	-
Ccdc93	-	-	-	Ccdc93	-	-	-	-	-	-	-	-
Cnnt1	-	-	-	Cnnt1	-	-	-	-	-	-	-	-
Ctag65	-	-	-	Ctag65	-	-	-	-	-	-	-	-
Cyp4f14	-	-	-	Cyp4f14	-	-	-	-	-	-	-	-
Etv3	-	-	-	Etv3	-	-	-	-	-	-	-	-
Gaint6	-	-	-	Gaint6	-	-	-	-	-	-	-	-
GLRX	-	-	-	GLRX	-	-	-	-	-	-	-	-
Gm11602	-	-	-	Gm11602	-	-	-	-	-	-	-	-
Gng12	-	-	-	Gng12	-	-	-	-	-	-	-	-
Heph	-	-	-	Heph	-	-	-	-	-	-	-	-
Ldlr	-	-	-	Ldlr	-	-	-	-	-	-	-	-
Mir1942	-	-	-	Mir1942	-	-	-	-	-	-	-	-
Mox1412	-	-	-	Mox1412	-	-	-	-	-	-	-	-
Ndufs1	-	-	-	Ndufs1	-	-	-	-	-	-	-	-
Ogdn	-	-	-	Ogdn	-	-	-	-	-	-	-	-
Oxdk	-	-	-	Oxdk	-	-	-	-	-	-	-	-
Plec	-	-	-	Plec	-	-	-	-	-	-	-	-
P1khg2	-	-	-	P1khg2	-	-	-	-	-	-	-	-
Rp13a	-	-	-	Rp13a	-	-	-	-	-	-	-	-
Rp13a-ps1	-	-	-	Rp13a-ps1	-	-	-	-	-	-	-	-
Sepp7	-	-	-	Sepp7	-	-	-	-	-	-	-	-
(base) tinaryu@Tinas-MacBook-Pro-3 KuangLab % grep -c FAIL genes.txt												
25	-	-	-	-	-	-	-	-	-	-	-	-

Extract FPKM values for each group (7 files) > GF.txt, hdac3\_WT.txt, hdac3\_KO.txt, nr1d1\_WT.txt, nr1d1\_KO.txt, nfli3\_WT.txt, nfli3\_KO.txt

2 versions: one original and one replicated

gene_id	q1_FPKM	q2_FPKM	q3_FPKM	q4_FPKM	gene_id	q1_FPKM	q2_FPKM	q3_FPKM	q4_FPKM	q1_FPKM_re	q2_FPKM_re	q3_FPKM_re	q4_FPKM_re
0610005C131	42.3248	90.0548	58.472	33.0997	0610005C131	42.3248	90.0548	58.472	33.0997	42.3248	90.0548	58.472	33.0997
0610007C211	69.5387	65.9222	64.5194	53.7944	0610007C211	69.5387	65.9222	64.5194	53.7944	69.5387	65.9222	64.5194	53.7944
0610007L01F	9.0729	9.29366	8.62519	7.60749	0610007L01F	9.0729	9.29366	8.62519	7.60749	9.0729	9.29366	8.62519	7.60749
0610007N19I	6.30459	9.80456	7.29856	6.4871	0610007N19I	6.30459	9.80456	7.29856	6.4871	6.30459	9.80456	7.29856	6.4871
0610007P08F	1.01314	1.1118	1.01785	1.14966	0610007P08F	1.01314	1.1118	1.01785	1.14966	1.01314	1.1118	1.01785	1.14966

Feb 23<sup>rd</sup> 2024

1. Filter out genes that are not expressed well at all time points  $\text{fpkm val} < 1$

Before 23263 genes `nfil3_KO.txt`

After 11249 genes `nfil3_KO_expressed.txt`

```
(base) tinaryu@Tinas-MacBook-Pro-3 replicated % awk '$2 > 1 || $3 > 1 || $4 > 1 || $5 > 1'
GF.txt > GF_expressed.txt
```

2. Run Metacycle using the `groupname_expressed.txt` files above > `meta2d_GF_expressed.txt`
3. Filter out genes with meta2d  $\text{pval} > 0.05$ . From the Metacycle output file with LS, ARS, JTK, and meta2d values, extract the columns with meta2d data only

```
((base) tinaryu@Tinas-MacBook-Pro-3 meta2d_rep_expressed % _cut -f 1,17-23 meta2d_GF_expressed.txt | awk '$2 < 0.05' > meta2d_GF_sig.txt
```

2 types of files:

file 1 – list of gene names with meta2d  $\text{pval} < 0.05$

file 2 – meta2d  $\text{pval} < 0.05$  genes and meta2d values

Before 11249 genes `meta2d_nfil3_KO_expressed.txt`

After 6111 genes `meta2d_sig/meta2d_nfil3_KO_sig.txt`

4. Extract geneIDs of genes that are dependent on `hdac3`, `nr1d1`, or `nfil3` by subtracting gene list of WT – KO using unix

```
((base) tinaryu@Tinas-MacBook-Pro-3 meta2d_sig_genelist % grep -v -xFf meta2d_hdac3_KO_sig_g
enelist.txt meta2d_hdac3_WT_sig_genelist.txt > hdac3_dependent_genelist.txt
```

6230 genes in `hdac3_WT` -> 3355 genes which are `hdac3` dependent

Compare the gene overlap between 3 groups

