

Investigations into Chalcogen Bonding

by

Thomas Fellowes

[ORCID: 0000-0002-6389-6049](#)

A thesis submitted in total fulfillment for the
degree of Doctor of Philosophy

in the
Faculty of Science
School of Chemistry and Bio21 Institute
THE UNIVERSITY OF MELBOURNE

June 2021

Contents

List of Figures	ix
List of Tables	xv
Compounds	xvii
Symbols	xxi
I Background	1
1 Introduction	3
1.1 Chalcogen Bonding	5
1.1.1 Mechanisms	5
1.2 Applications of Ch-Bonding	7
1.2.1 Materials	7
1.2.2 Catalysis and bond activation	11
1.2.3 Biological systems	13
1.2.3.1 Proteins	13
1.2.3.2 Nucleic acids	15
1.3 Experimental methods	17
1.3.1 Computational methods	17
1.3.1.1 Natural Bond Orbital theory	17
1.3.1.2 Quantum Theory of Atoms In Molecules	21
1.3.2 X-ray crystallography	25
1.3.2.1 Charge density refinement	25
1.3.3 Nuclear Magnetic Resonance	32
1.3.3.1 Solid State NMR	32
2 Simulating chalcogen bonding using molecular mechanics	47
2.1 Abstract	47
2.2 Introduction	48
2.3 Results and Discussion	51
2.3.1 Classical bonding parameters	52

2.3.2 Energy decomposition analysis	54
2.3.3 Incorporation of pseudoatom	56
2.3.4 Electrostatic potential map	58
2.3.5 Validation against DFT geometries	58
2.3.6 Validation against experimental density	59
2.3.7 Validation against SOD1 binding	60
2.4 Conclusion	61
II The strength and nature of chalcogen bonding	69
3 Insights from co-crystal structures	71
3.1 Abstract	71
3.2 Introduction	72
3.3 Results and Discussion	73
3.3.1 Synthesis of 1 , 1a and 3	73
3.3.2 Co-crystal structures of benzisoselenazolinones and Lewis bases	74
3.3.3 Effects of the benzisoselenazolinone on Ch-bond strength	75
3.3.4 Endocyclic bond lengthening associated with stronger complexes	75
3.3.5 H-bond enhanced Ch-bonding	76
3.3.6 Effects of the Lewis base on Ch-bond strength	77
3.3.7 DFT interaction energies and NBO analysis	78
3.4 Conclusion	80
3.5 Supplementary materials	80
3.5.1 Synthetic procedures	80
3.5.1.1 Preparation of 1	80
3.5.1.2 Preparation of 1a	81
3.5.1.3 Preparation of 3	82
3.5.2 Crystallographic data	82
3.5.2.1 Crystal data for 1 ·DMAP	83
3.5.2.2 Crystal data for 1a	83
3.5.2.3 Crystal data for 1a ·DMAP·H ₂ O	84
3.5.2.4 Crystal data for 1a ·DMAP	84
3.5.2.5 Crystal data for 1a ·quinuclidine	85
3.5.2.6 Crystal data for 1a ·DABCO	85
3.5.2.7 Crystal data for 3	86
3.5.2.8 Crystal data for 3 ·pyridine	87
3.5.2.9 Crystal data for 3 ·DMAP	87
4 Further investigations into Ch-bonded complexes	93
4.1 Introduction	93
4.2 Results and discussion	94

4.2.1	Hammett plots of crystallographic data	96
4.2.1.1	Co-crystals where $Z' = 2$	105
4.2.2	Measurement of chemical shift anisotropy	111
4.2.3	Calculation of chemical shift anisotropy	112
4.2.3.1	Measurement of CSA in a single crystal	118
4.2.4	Solution-phase studies	123
4.2.4.1	NMR titration experiments	123
4.2.4.2	UV-Vis titration experiment	127
4.2.5	Ch-bonding in the gas phase	128
4.3	Conclusion	131
4.4	Supplementary materials	132
4.4.1	Synthetic procedures	132
4.4.1.1	Preparation of 5	132
4.4.1.2	Preparation of 6	133
4.4.1.3	General procedure A	133
4.4.1.4	General procedure B	134
4.4.1.5	General procedure C	134
4.4.1.6	General procedure for substituted pyridines	135
4.4.2	Crystallisation methods	135
4.4.3	Characterisation data	135
4.4.3.1	Ebselen 1	135
4.4.3.2	4-Nitro ebselen 1NO₂	136
4.4.3.3	4-Cyano ebselen 1CN	136
4.4.3.4	4-Trifluoromethyl ebselen 1CF₃	136
4.4.3.5	4-Bromo ebselen 1Br	137
4.4.3.6	4-Carboxyethyl ebselen 1CO₂Et	137
4.4.3.7	4-Methyl ebselen 1Me	137
4.4.3.8	4-Methoxy ebselen 1OMe	138
4.4.3.9	4-Ethoxy ebselen 1OEt	138
4.4.4	Crystallographic Data	138
4.4.4.1	Crystal data for 1	138
4.4.4.2	Crystal data for 1·DMAP	139
4.4.4.3	Crystal data for 1·4a	140
4.4.4.4	Crystal data for 1·4b	140
4.4.4.5	Crystal data for 1NO₂·DMAP	141
4.4.4.6	Crystal data for 1NO₂·DMF	141
4.4.4.7	Crystal data for 1CN	142
4.4.4.8	Crystal data for 1CN·DMAP	143
4.4.4.9	Crystal data for 1CN·DMAP·DCM	143
4.4.4.10	Crystal data for 1CN·4a	144
4.4.4.11	Crystal data for 1CN·4b·DCM	144

4.4.4.12 Crystal data for 1CF₃	145
4.4.4.13 Crystal data for 1CF₃·DMAP	146
4.4.4.14 Crystal data for 1Br	146
4.4.4.15 Crystal data for 1Br·DMAP	146
4.4.4.16 Crystal data for 1Br·4a	147
4.4.4.17 Crystal data for 1Br·4b	148
4.4.4.18 Crystal data for 1CO₂Et	148
4.4.4.19 Crystal data for 1CO₂Et·DMAP·DCM	149
4.4.4.20 Crystal data for 1CO₂Et·4a	150
4.4.4.21 Crystal data for 1CO₂Et·4b	150
4.4.4.22 Crystal data for 1Me	150
4.4.4.23 Crystal data for 1Me·DMAP	151
4.4.4.24 Crystal data for 1Me·4a	152
4.4.4.25 Crystal data for 1Me·4b	152
4.4.4.26 Crystal data for 1OMe	153
4.4.4.27 Crystal data for 1OMe·DMAP	154
4.4.4.28 Crystal data for 1OEt	154
4.4.4.29 Crystal data for 1OEt·DMAP	155
4.4.4.30 Crystal data for 1OEt·4a	156
4.4.4.31 Crystal data for 1OEt·4b	156
 III Chalcogen bonding at oxygen	 161
 5 Experimental evidence of Chalcogen bonding at oxygen.	 163
5.1 Abstract	163
5.2 Introduction	164
5.3 Results and Discussion	165
5.3.1 Structural effects attributable to the Ch-bond	165
5.3.2 NBO calculations	166
5.3.3 QTAIM analysis of electron density	167
5.3.4 CSD search for similar structures	168
5.3.5 Analysis of analogues and determination of experimental electron density	169
5.4 Supplementary material	172
5.4.1 Synthesis	172
5.4.1.1 Preparation of 8 and 9	172
5.4.2 Crystallographic data	173
5.4.2.1 Crystal data for 7	173
5.4.2.2 Crystal data for 8	173
5.4.2.3 Crystal data for 9	174

IV Applications of Ch-bonding	179
6 Thermal rearrangement of a Ch-bonded solvate	181
6.1 Abstract	181
6.2 Introduction	182
6.3 Results and discussion	183
6.3.1 Synthesis	183
6.3.2 Structural characterisation	184
6.3.3 Variable temperature studies	189
6.4 Conclusion	191
6.5 Experimental	192
6.5.1 Synthesis	192
6.5.1.1 Preparation of 14	192
6.5.1.2 Preparation of 13	193
6.5.2 Crystallography	194
6.5.2.1 Crystal data for 13 (ex.DMF)	194
6.5.2.2 Crystal data for 13 ·pyridine	194
6.6 Acknowledgements	195
7 Development of a Ch-bonding DNA binder	201
7.1 Introduction	201
7.1.1 Cancer and treatment	201
7.1.2 Radiation therapy	202
7.1.3 Radioprotection	203
7.1.4 Minor groove binding	204
7.1.5 Bis-benzimidazole derived radioprotectors	205
7.1.6 Proton-coupled electron transfer (PCET)	206
7.1.7 Chalcogens as nitrogen bioisosteres	206
7.1.8 Ebselen as an antioxidant	207
7.2 Synthesis of analogues	207
7.2.1 Preparation of benzisoselenazolinone-benzimidazole 11	208
7.2.1.1 Ullmann coupling	208
7.2.1.2 Benzisoselenazolinone-first route	210
7.2.1.3 Benzisoselenazoline-last route	212
7.3 DNA binding studies	214
7.3.1 Co-crystallisation results	214
7.3.2 UV-Vis titration	216
7.4 Conclusions	219
7.5 Experimental procedures	219
7.5.1 DNA titration procedure	219
7.5.2 Synthetic methods	222
7.5.2.1 Preparation of 1b	222

7.5.2.2 Preparation of 1·S	222
7.5.2.3 Preparation of 16	223
7.5.2.4 Preparation of 15	224
7.5.2.5 Preparation of 15·PMB	224
7.5.2.6 Preparation of 18	225
7.5.2.7 Preparation of 19·PMB·S	226
7.5.2.8 Preparation of 21	227
7.5.2.9 Preparation of 22	227
7.5.2.10 Preparation of 23	228
7.5.2.11 Preparation of 19	229
7.5.2.12 Preparation of 14	230
7.5.2.13 Preparation of 13	231
8 Concluding remarks	235

List of Figures

1.1	Bromine-dioxane complex.	4
1.2	Chalcogen bonding model, and similarity to H-bonding.	5
1.3	“Molecular balances” designed by Pascoe2017 for the measurement of Ch-bond energies. ^{Pascoe2017}	6
1.4	One-dimensional chains of 4-(4-iodobenzylidene)-TEMPO structured by X-bonds. ^{Metrangolo2008}	8
1.5	Anion binding through anion- π interactions and X-bonding. ^{VargasJentzsch2013}	8
1.6	Ch-bonding in thiaboranes. ^{Fanfrlik2014}	9
1.7	A tetrameric macrocycle held together by Ch-bonds. ^{Ho2016}	9
1.8	A bidentate Ch-bonding anion binder. ^{Garrett2016}	10
1.9	One dimensional chains formed in the crystal packing of ebselen. . .	10
1.10	Proposed mechanism for the catalytic reduction of quinoline by a Ch-bonding catalyst. ^{Benz2017}	12
1.11	The structure of mebroqualone.	12
1.12	Directional preference for Ch-bonded interactions in proteins.	14
1.13	Holliday junction formed by X-bonding. ^{Voth2007}	16
1.14	Canonical Watson-Crick U/A base pair, and the “wobble” U:G base pair in RNA.	16
1.15	MO diagram for dihydrogen. Reproduced from reference [h-orbital-diagram].	18
1.16	Molecular orbitals of benzene. Reproduced from reference [Luhmann2015].	19
1.17	Representative Natural Bond Orbitals of a DMF molecule.	20
1.18	Electron density $\rho(\mathbf{r})$ in the plane of a DMF molecule.	22
1.19	Laplacian of the electron density $\nabla^2\rho(\mathbf{r})$ of DMF	22
1.20	Electron density gradient $\nabla\rho(\mathbf{r})$ vector field of DMF.	23
1.21	Critical points of DMF.	23
1.22	Scattering factors for some atoms as a function of $(\sin\theta)/\lambda$	27
1.23	Structure of 1,3,5-triazine.	28
1.24	Correlation of thermal and dipolar functions.	31
1.25	Principal axis system with respect to laboratory reference.	34
1.26	SS-NMR powder line shape	35
1.27	^{31}P SS-NMR spinning at the magic angle.	36
2.1	Catalytic cycle of ebselen 1 <i>in vivo</i>	49

2.2 Potential energy surfaces of ebselen.	53
2.3 Structures of complexes used for SAPT(DFT) analysis.	56
2.4 SAPT(DFT) analysis of complexes with four Lewis bases.	56
2.5 ESP of ebselen generated from DFT density, and atom centred charges.	58
2.6 Average binding geometry of ebselen in the SOD1 groove.	61
3.1 Structure of 1a ·DMAP, showing the two distinct geometries.	76
3.2 Structure of 1a ·DMAP·H ₂ O.	76
3.3 Pyridine and DMAP adducts of Se-tetracycle 3	77
3.4 Adducts of benzisoselenazolinone 1a with quinuclidine and DABCO.	78
3.5 Interaction energies for various complexes.	79
3.6 Orbital overlap for 1 ·DMAP.	79
3.7 X-ray crystal structure of 1 ·DMAP.	83
3.8 X-ray crystal structure of 1a	84
3.9 X-ray crystal structure of 1a ·DMAP·H ₂ O.	84
3.10 X-ray crystal structure of 1a ·DMAP.	85
3.11 X-ray crystal structure of 1a ·quinuclidine.	86
3.12 X-ray crystal structure of 1a ·DABCO.	86
3.13 X-ray crystal structure of 3	87
3.14 X-ray crystal structure of 3 ·pyridine.	87
3.15 X-ray crystal structure of 3 ·DMAP.	88
4.1 Nucleophilic substitution at selenium.	94
4.2 Hammett plot of endocyclic Se–N bond length of uncomplexed ebselen derivatives.	98
4.3 One dimensional chains formed by Ch-bonding between the selenium and carbonyl oxygen in 1Me	99
4.4 Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with DMAP.	99
4.5 Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with 4a	100
4.6 Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with 4b	101
4.7 Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with DMAP.	102
4.8 Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with 4a	103
4.9 Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with 4b	104
4.10 Hammett plots of QTAIM parameters ρ_{BCP} and $\nabla^2\rho_{BCP}$ of ebselen derivatives complexed with DMAP.	104
4.11 Hammett plots of QTAIM parameters ρ_{BCP} and $\nabla^2\rho_{BCP}$ of ebselen derivatives complexed with 4a	105

4.12 Hammett plots of QTAIM parameters ρ_{BCP} and $\nabla^2\rho_{BCP}$ of ebselen derivatives complexed with 4b	105
4.13 Solid state IR spectrum of 1OEt ·DMAP	106
4.14 Single crystal x-ray structure of 1OMe ·DMAP	107
4.15 Calculated vs measured powder diffraction pattern for 1OMe ·DMAP	108
4.16 Solution NMR spectra of 1OMe ·DMAP	109
4.17 Solid state ^{13}C -NMR spectrum of 1OMe ·DMAP	110
4.18 Solid state ^{77}Se -NMR spectrum of 1OMe ·DMAP	111
4.19 ^{77}Se CPMAS NMR spectra for ebselen derivatives	113
4.20 Calculated chemical shift tensors for ebselen derivatives	117
4.21 Single crystal of 1 ·DMAP used for SS-NMR	118
4.22 Indexed faces of 1 ·DMAP	119
4.23 ^{77}Se NMR spectrum of a single crystal of 1 ·DMAP in the orientation depicted in the inset	119
4.24 ^{77}Se NMR spectrum of a single crystal of 1 ·DMAP in the orientation depicted in the inset	120
4.25 Experimentally determined chemical shift tensor for 1 ·DMAP	122
4.26 NMR titration binding isotherms	126
4.27 Hammett plot of binding energies between ebselen derivatives and DMAP	127
4.28 UV-Vis titration of 1 with DMAP	128
4.29 Positive mode ESI of 1 ·DMAP·H ⁺	129
4.30 Negative mode ESI of 1 ·isonicotinate	130
4.31 TCID experiment of 1 ·isonicotinate	131
4.32 TCID experiment of 1a ·isonicotinate	131
4.33 X-ray crystal structure of 1	139
4.34 X-ray crystal structure of 1 ·DMAP	139
4.35 X-ray crystal structure of 1 · 4a	140
4.36 X-ray crystal structure of 1 · 4b	141
4.37 X-ray crystal structure of 1NO₂ ·DMAP	141
4.38 X-ray crystal structure of 1NO₂ ·DMF	142
4.39 X-ray crystal structure of 1CN	142
4.40 X-ray crystal structure of 1CN ·DMAP	143
4.41 X-ray crystal structure of 1CN ·DMAP·DCM	144
4.42 X-ray crystal structure of 1CN · 4a	144
4.43 X-ray crystal structure of 1CN · 4b ·DCM	145
4.44 X-ray crystal structure of 1CF₃	145
4.45 X-ray crystal structure of 1CF₃ ·DMAP	146
4.46 X-ray crystal structure of 1Br	147
4.47 X-ray crystal structure of 1Br ·DMAP	147
4.48 X-ray crystal structure of 1Br · 4a	148

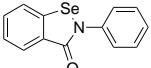
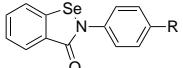
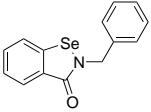
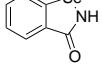
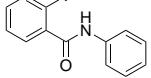
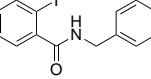
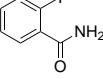
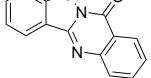
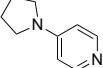
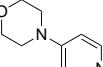
4.49 X-ray crystal structure of 1Br·4b	148
4.50 X-ray crystal structure of 1CO₂Et	149
4.51 X-ray crystal structure of 1CO₂Et·DMAP-DCM	149
4.52 X-ray crystal structure of 1CO₂Et·4a	150
4.53 X-ray crystal structure of 1CO₂Et·4b	151
4.54 X-ray crystal structure of 1Me	151
4.55 X-ray crystal structure of 1Me·DMAP	152
4.56 X-ray crystal structure of 1Me·4a	152
4.57 X-ray crystal structure of 1Me·4b	153
4.58 X-ray crystal structure of 1OMe	153
4.59 X-ray crystal structure of 1OMe·DMAP , polymorph A.	154
4.60 X-ray crystal structure of 1OMe·DMAP , polymorph B.	155
4.61 X-ray crystal structure of 1OEt	155
4.62 X-ray crystal structure of 1OEt·DMAP	156
4.63 X-ray crystal structure of 1OEt·4a	156
4.64 X-ray crystal structure of 1OEt·4b	157
 5.1 Oxime 7 displays a close O ··· O contact.	164
5.2 Structures of CSD matches for the <i>o</i> -nitro-O-aryl oxime motif.	168
5.3 Oxime 8 adopts a similar conformation to 7	169
5.4 Deformation and Laplacian maps of the electron density for 8	170
5.5 NCI maps for 8 and 9	171
5.6 Electrostatic potential mapped onto a plane through the oxime bond. .	172
 6.1 Chalcogen bonding model, and similarity to H-bonding.	182
6.2 Thermal ellipsoid plot of 13(ex.DMF)	184
6.3 Offset π-stacking of 13(ex.DMF) extending down the <i>b</i> -axis.	185
6.4 O ··· Se chalcogen bonding interactions in 13(ex.DMF)	186
6.5 2-D layers of 13(ex.DMF)	186
6.6 Extension of the 2-D layers of 13(ex.DMF) into 3-D by van der Waals interactions.	187
6.7 Thermal ellipsoid plot of 13·pyridine	188
6.8 Sheets of compound 13 viewed from orthogonal and parallel directions. .	188
6.9 Two orthogonal views of parallel sheets of compound 13	189
6.10 TGA analysis of 13·pyridine	190
6.11 Variable temperature powder XRD patterns from 13·pyridine	190
6.12 Rearrangement of Ch-bonds upon desolvation.	192
 7.1 Amifostine, and its active metabolite.	203
7.2 Hoechst stains, featuring the characteristic bis-benzimidazole motif. .	204
7.3 Methylproamine, a lead compound identified for radioprotection. .	205

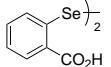
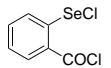
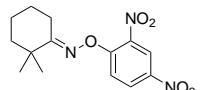
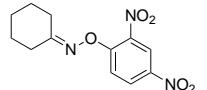
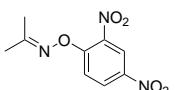
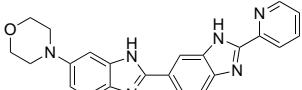
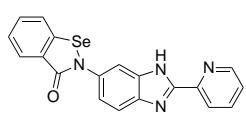
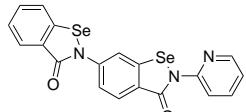
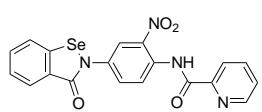
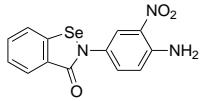
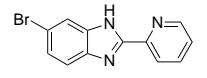
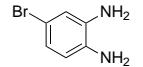
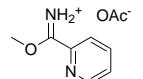
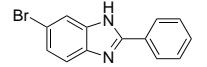
7.4 Single crystal structure of methylproamine complexed with an AT rich DNA oligomer.	205
7.5 M2PB 10 , an optimised lead compound for radioprotection.	206
7.6 Lead bis-benzimidazole 10 and initial target compounds 11 and 12	208
7.7 X-ray crystal structure of one isomer of 19·PMB·S	211
7.8 X-ray crystal structure of 20	212
7.9 X-ray crystal structure of 19	214
7.10 Crystal packing of the A2T2 self-complimentary DNA 16mer, showing the open channels between the DNA molecules.	216
7.11 Calibration curve for 19	218
7.12 UV-vis titration of 19 with ct-DNA at low DNA stock concentration.	218
7.13 UV-vis titration of 19 with ct-DNA at high DNA stock concentration.	218
7.14 ^1H NMR spectra of the adduct of 19 with ct-DNA, and the pure ligand (blue) for comparison.	220
7.15 X-ray crystal structure of 1b	223
7.16 X-ray crystal structure of 15·PMB	225
7.17 X-ray crystal structure of 22 as the tetrafluoroborate salt.	228
7.18 X-ray crystal structure of 23 as the hexachlorostannate salt.	229

List of Tables

2.1	Classical bonding parameters for ebselen.	52
2.2	Dihedral parameters for ebselen.	54
2.3	Selected atomic charges for the pseudoatom and no pseudoatom models.	57
2.4	Median geometric parameters for complexes with 1	59
3.1	Selected structural parameters of Ch-bonded complexes.	74
3.2	Hydrogen bond basicities of bases studied.	78
4.1	Selected structural and electron density parameters of Ch-bonded complexes.	97
4.2	Linear regression results for Hammett plots in figs. 4.4 to 4.6.	100
4.3	Linear regression results for Hammett plots in figs. 4.7 to 4.9.	102
4.4	Principal values of the chemical shift tensor extracted from powder spectra in fig. 4.19.	112
4.5	Principal values of the chemical shielding tensor calculated from optimised structures.	116
4.6	Principal values of the chemical shift tensor calculated from optimised structures.	116
4.7	NMR titration binding energies.	127
5.1	Selected geometric parameters of oximes 7 , 8 and 9	165
5.2	Selected geometric parameters of structures HEPMEV , HUGSUX , QOLXOE , and IQADOT	168

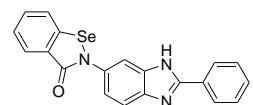
Compounds

Compound number	Compound code	Structure
1	ebs	
1R	ebs.4R	
1a	ebs.bn	
1b	ebs.h	
2	amide	
2a	amide.bn	
2b	amide.h	
3	tetracycle	
4a	py.pyrrol	
4b	py.morph	

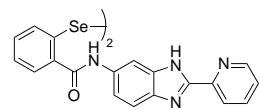
5	diselenide	
6	dichloride	
7	dimethylcyclohexanone-oxime-dnp	
8	cyclohexanone-oxime-dnp	
9	acetone-oxime-dnp	
10	m2pb	
11	ebs-rhs-2py	
12	ebs-ebs-2py	
13	ebs-nitroamide-2py	
14	ebs-nitroaniline	
15	rhs-bromo-2py	
16	bromodiamine	
17	2py-carboximidate	
18	rhs-bromo-ph	

19

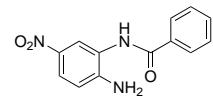
ebs-rhs-ph

**20**

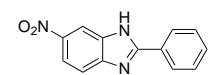
ebs-rhs-2py-diselenide

**21**

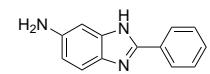
rhs-nitro-amide

**22**

rhs-nitro

**23**

rhs-amine



Symbols

σ chemical shielding

σ estimated standard deviation

Part I

Background

Chapter 1

Introduction

The importance of non-bonding interactions in our world simply cannot be overstated. While we may think of the more familiar covalent and ionic bonds as the core of chemistry, more often than not it is non-bonding interactions that dictate the nature of substances, from bulk properties such as tensile strength, to microscopic phenomena like protein-ligand interactions.

At a fundamental level, all non-bonding interactions (indeed, conventional bonding too) are attributable to the electrostatic force. However, it is more convenient to broadly categorise interactions based on strength and structural motifs that are common to each class. Such classes include dispersion forces, dipole-dipole interactions, and hydrogen bonding (H-bonding). In the 19th century, it became apparent that these classes paint an incomplete picture of intermolecular bonding. As part of their investigation into solvent effects on the colour of iodine solutions in 1949, Bensei and Hildebrand invoked the concept of “charge-transfer” complexes to explain the association of molecular iodine with nucleophilic solvent molecules.¹ Although the electrophilic nature of molecular halogens was already well appreciated with respect to reactivity, this appears to be the first acknowledgement that this understanding could be applied to ground state complexes. In support of this, Hassel and Hvoslef elucidated the structure of a 1:1 complex of molecular bromine

and 1,4-dioxane in 1954, and observed an unusually strong interaction between the two molecules.² The crystal was comprised of long chains of monomers (fig. 1.1), exhibiting a very short O...Br contact (2.71Å, the sum of the van der Waals (VDW) radii is 3.37Å), and a slightly lengthened Br–Br bond (2.31Å vs 2.28Å in gaseous bromine).

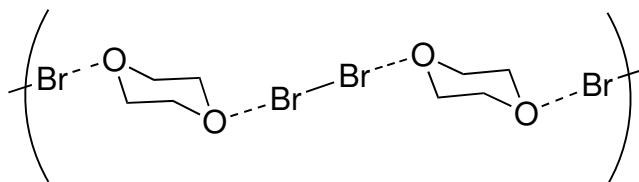


FIGURE 1.1: Long chains of molecular bromine and 1,4-dioxane, observed by Hassel and Hvoslef.

In the subsequent years, a wide variety of names and labels were applied to the “charge-transfer” phenomenon, including “lock and key”, “donor-acceptor”, and “filling of anti-bonding orbitals”, as summarised in Bent’s excellent 1968 review.³ The term “halogen bonding” (X-bonding) only gained widespread acceptance in 1983 after the publication of the review of Dumas, Gomel, and Guerin.⁴ The term deliberately evokes similarity to the better known concept of hydrogen bonding, as the two phenomena are of comparable strength and, importantly, directionality. In 1998 Anthony C. Legon again used the term to describe the attractive interaction between halogens and Lewis bases in pre-reactive complexes, harking back to the original understanding of electrophilic halogens as a primarily reactive phenomenon.^{5,6} In the following years, the potential of X-bonding was more fully recognised, with applications in supramolecular chemistry and self-assembly,^{7–9} catalysis and bond activation,^{10–12} and molecular sensing and recognition.^{13–15}

From X-bonding grew the related concepts of chalcogen (Ch-), pnictogen (Pn-), and tetrel bonding (T-bonding), as the electrophilic nature of these elements was discovered.¹⁶ Similar applications have also been found for these classes of non-covalent interactions, Ch-bonding in particular.^{17–24}

1.1 Chalcogen Bonding

Ch-bonding is the attractive interaction between a Lewis base and a chalcogen atom bearing an electron withdrawing substituent (fig. 1.2). The terminology of Ch-bonding partners is perhaps counter-intuitive, as donors and acceptors are named by analogy with H-bonded equivalents. Note this is contrary to the formal flow of electron density; Ch-bond *donors* bear vacant *acceptor* orbitals, while Ch-bond *acceptors* are *donors* of electron density. For the purposes of this thesis, “donor” will refer to the Lewis acidic chalcogen species.

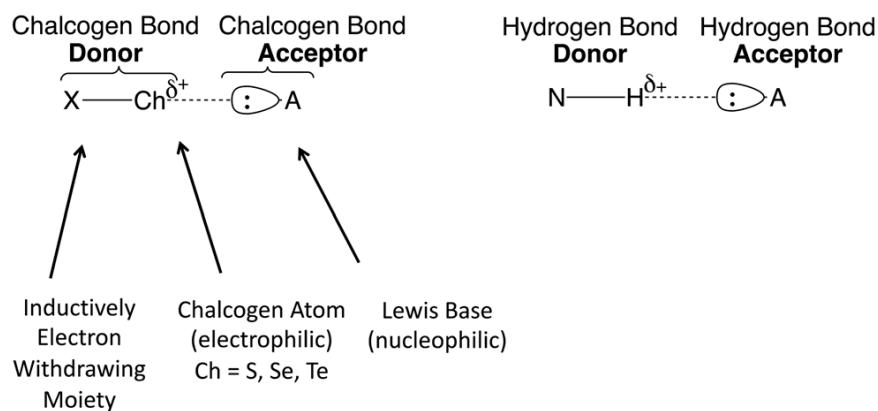


FIGURE 1.2: Chalcogen bonding model, and similarity to H-bonding.

1.1.1 Mechanisms

As was discussed in the previous section, all intermolecular (and indeed *intramolecular*) forces are manifestations of the electromagnetic force. Nonetheless, it is more convenient to categorise energetic contributions to any given interaction as orbital, electrostatic, or dispersion mediated, as there are marked differences in strength and directionality between them. Historically, X- and Ch-bond interactions were understood to be primarily due to orbital overlap and resulting charge transfer. Evidence for this included the lengthening of the X-X bond corresponding to increased occupation of the $\sigma^*(X-X)$ orbital due to the incoming lone pair (this

was observed in Hassel and Hvoslef's original work²). Characteristic charge transfer bands are also observed in the UV-vis spectra of halogen solutions.²⁵

Such evidence of orbital contributions is not, however, common to all systems displaying X- or Ch-bond interactions. The iodoperfluoroalkanes and arenes studied by Resnati et al. show no charge transfer bands in UV-vis spectra, yet are exceptionally strong X-bond donors.²⁶ Jane Murray and Peter Politzer have advocated for an alternative, primarily electrostatic explanation of X- and Ch-bond interactions.^{27,28} They propose that a positively charged σ -hole is generated along the extension of a σ bond to an electron withdrawing substituent, due to polarisation of the bonding orbital. This also adequately explains the strength and directionality of these interactions. An interpretation which has not attracted as much attention is that of dispersion forces.

While the varying contributions from each of these factors can be dissected computationally via perturbation methods, experimental evidence is surprisingly sparse. Pascoe, Ling and Cockroft, however, devised an elegant experiment to quantitatively determine energetic contributions.²⁹ ^{19}F NMR was used to determine the relative populations of two conformers of a "molecular balance", one bearing a Ch-bond interaction, one not (fig. 1.3). Interaction energies were thus derived, and found to be more or less invariant with respect to the solvent. This appeared to rule out electrostatic contributions, as solvent dipole moment and bulk polarisability would otherwise have a large effect on the position of the equilibrium. These results also suggest that dispersion plays only a minor role, for similar reasons.

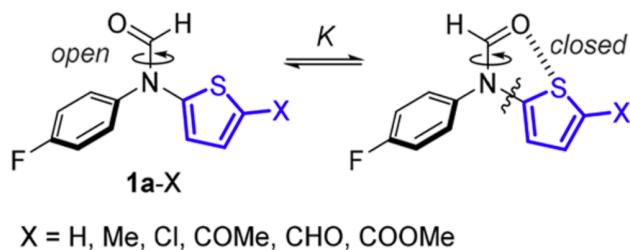


FIGURE 1.3: "Molecular balances" designed by Pascoe et al. for the measurement of Ch-bond energies.²⁹

DFT was used to further probe the dispersion contribution. Free energies were calculated using functionals which both did, and did not include dispersion corrections, and the values compared to the experimental results. The non-dispersion corrected B3LYP functional gave superior correlation with experiment than did either M06-2X and ω B97-D, both of which include dispersion. This provided additional evidence that, in the systems studied, the major contributor to the interaction is orbital overlap and charge transfer.

Although current evidence points towards these interactions being primarily orbital related, the electrostatic “ σ -hole” terminology of Politzer and Murray has stuck, and the term is now used to encompass the whole gamut of X- Ch- Pn- and T-bonding interactions.

1.2 Applications of Ch-Bonding

Ch-bonding, and by extension, all σ -hole interactions can theoretically be applied to any formal Lewis acid-base system. They are especially attractive as a *hydrophobic* complement to H-bonding interactions, which are generally considered to be *hydrophilic*. The following are brief summaries of existing applications in the literature.

1.2.1 Materials

The major applications of σ -hole interactions have so far been in the realm of crystal engineering. Early work by Corradi et al. showed that halogen bonding was able to outcompete H-bonding in the formation of supramolecular architectures.⁷ A review by Metrangolo et al. summarised the forms that are accessible using X-bonding to direct crystal growth.⁸ 1D, 2D, and 3D architectures are able to be generated using appropriate X-bond donors, and these show potential in the design of liquid crystals,

organic semiconductors and paramagnetic materials (fig. 1.4) A more recent review by the same group described applications in anion transport, and luminescent and photo-responsive crystals.⁹ The group of Stefan Matile has further explored anion transport, and has published a review comparing X-bonding with other hydrophobic interactions such as anion- π and anion-macro dipole interactions (fig. 1.5).¹⁴

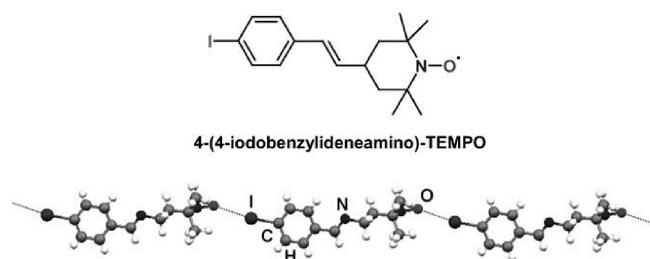


FIGURE 1.4: One-dimensional chains of 4-(4-iodobenzylideneamino)-TEMPO structured by X-bonds.⁸

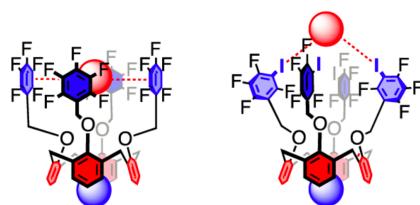
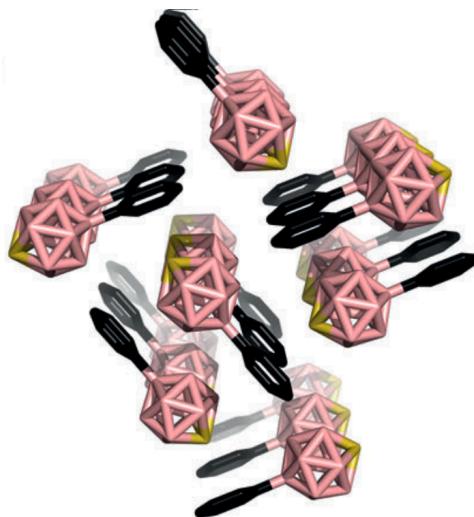


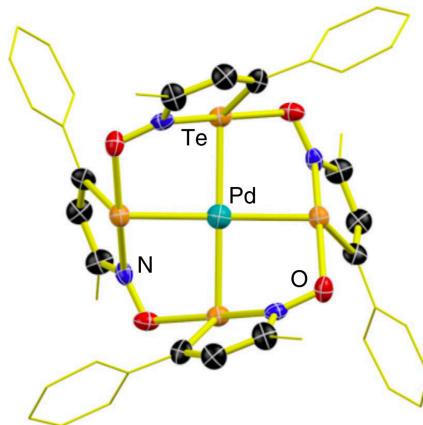
FIGURE 1.5: Anion binding through anion- π interactions and X-bonding.¹⁴

Ch-bonding, too, has been investigated with respect to materials chemistry. Fanfrlík et al. demonstrated the importance of Ch-bonding on the crystal packing of thiaboranes.¹⁷ They found that the sulfur-based σ -hole was sufficiently strong to interact with the weakly basic π electrons of a phenyl group, with contacts as short as 3.2 Å being observed (fig. 1.6).

In 2016, Ho et al. published their work into tellurium-based Ch-bonding.¹⁹ Their scaffolds are based on an iso-tellurazole N-oxide, which reversibly forms macrocyclic structures that persist in both gas and solution phase (fig. 1.7). The macrocycles were found to coordinate Pd_2^+ . This is particularly interesting, as the tellurium atoms are simultaneously behaving as a Lewis acid and base. The authors point out that such soft macrocycles are quite rare, and their work could facilitate further

FIGURE 1.6: Ch-bonding in thiaboranes.¹⁷

studies of transition metals in a soft coordination environment. They went on to investigate benzo-fused derivatives of iso-tellurazoles, as well as selenium analogues, which crystallised to form macromolecular pores and voids.²⁴

FIGURE 1.7: A tetrameric macrocycle held together by Ch-bonds.¹⁹

The Taylor group has been active in the development of X- and Ch-bonding molecular sensors. Early work demonstrated that X-bonding tridentate ligands (reminiscent of enterobactin) showed moderate selectivity for Cl^- .³⁰ They later developed bidentate Ch-bonding ligands which exhibited a tenfold increase in association constant with respect to chloride (fig. 1.8).^{18,31}

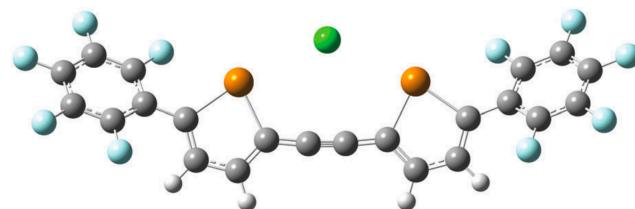


FIGURE 1.8: A bidentate Ch-bonding anion binder.¹⁸

Similar results have been achieved by the Beer group, who have developed X-bonding sensors for the perrhenate anion.¹³ These sensors are based on functionalised cyclodextrins, and are even more sensitive than the corresponding H-bonding analogues. The group has also used iodotriazole scaffolds to chelate anions.¹⁵ Incorporation of a chiral binaphthol moiety was shown to differentiate between enantiomers of chiral anions.

The role of selenium-based Ch-bonding on the crystal structure and mechanism of the drug ebselen was demonstrated by Thomas et al.³² Ch-bonding between the selenium atom and carbonyl oxygen directs the crystal packing of this compound, and leads to the formation of one-dimensional chains of molecules (fig. 1.9). Interestingly, the Lewis acidity of the σ -hole was invoked as an explanation of the antioxidant properties of the drug.

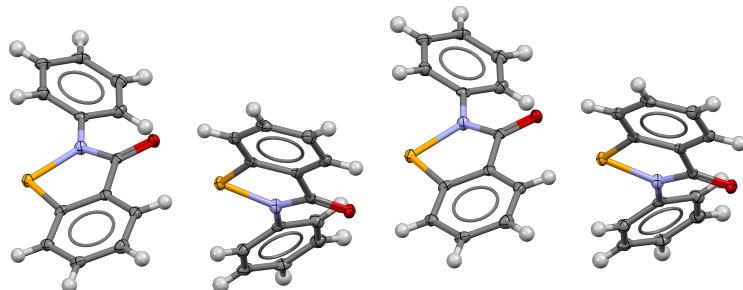


FIGURE 1.9: One dimensional chains formed in the crystal packing of ebselen.

1.2.2 Catalysis and bond activation

In 2008, halogen bonding was first applied to a Hantzsch ester reduction of a quinoline derivative.³³ This reaction is well characterised and understood, and has been catalysed with a variety of Brønsted and Lewis acids. The X-bond donors chosen were perfluorooiodoalkanes, and high conversions were achieved with modest catalyst loadings of 10%.

In 2011, a modified Ritter reaction was devised wherein benzhydryl bromide was activated by a dicationic imidazolium-based X-bond donor to give the carbocation intermediate, which was then captured by acetonitrile and then hydrolysed to afford the amide product.¹⁰ This pioneering work was limited by the necessity of stoichiometric amounts of the X-bond donor, as it is consumed in the course of the reaction.

A similar alkylation of 1-chloroisochroman was achieved by the same group using a neutral perfluorooiodoarene X-bond donor in catalytic quantities.³⁴ The proposed mechanism is similar to the thiourea-catalysed reaction of Reisman et al.,³⁵ which has shown promise in asymmetric induction. The authors noted issues with solubility of the perfluorooiodoarene catalysts, which is expected of such highly fluorinated compounds.

The reduction of quinoline was successfully catalysed by a dithiophene system by Benz et al. in 2017,²² and then again by the same group with a benzodiselenazole (fig. 1.10).³⁶ With the increased selectivity and strength of the Se-based Ch-bonding catalyst compared to the X-bonding perfluorooiodoalkane, the authors were able to reduce the catalyst loading to 1%.

Wonner et al. developed a selenated bis-benzimidazolium Ch-bonding catalyst for the alkylation of 1-chloroisochroman, and solvolysis of benzhydryl bromide.^{20,37} Although the best results were observed with the dicationic catalysts, conversion was also achieved with a neutral bis-benzimidazole catalyst, providing further evidence



FIGURE 1.10: Proposed mechanism for the catalytic reduction of quinoline by a Ch-bonding catalyst.²²

that the catalytic Lewis-acid site is indeed the σ -hole. For a given row in the periodic table (i.e. comparing a Se-based donor to a Br-based donor), Ch-bonding appeared to give superior results to X-bonding, as measured by % yield.

An unusual manifestation of X-bonding is in the self-disproportionation of enantiomers, as reported by Terada et al.¹¹ They observed spontaneous enrichment of one enantiomer of mebroqualone (fig. 1.11) upon chromatography using an achiral solid phase, which they attributed to the formation of diastereomeric X-bonded oligomers. This phenomenon has been observed in compounds capable of interacting through H-bonding or strong dipole-dipole interactions.³⁸

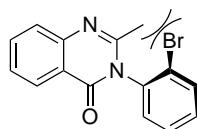


FIGURE 1.11: The structure of mebroqualone. Rotation is hindered about the biaryl bond, allowing the isolation of two atropisomers.

1.2.3 Biological systems

1.2.3.1 Proteins

While we usually think of the tertiary structure of proteins as being dominated by H-bonding and hydrophobic effects, there is increasing evidence that Ch-bonding plays an important role as well. This is not unexpected, as sulfur, a component of both cysteine and methionine, is known to form Ch-bonds in small molecules. In the early days of Ch-bonding interactions (2001, before the name had come into common use) Iwaoka et al. published an analysis of 604 protein structures in the Protein Data Bank (PDB).³⁹

A remarkable number of close contacts (sum of Van der Waals radii plus 0.5Å) between sulfur and Lewis basic (X) atoms were identified in the structures, with 33% of cysteine residues and 22% of methionine residues showing a close contact. Furthermore, the geometric parameters of these contacts were studied, with more than half of all contacts having a S–S–X (X=O,N) of 150–180°. The observed contacts were ascribed to a $\pi(\text{C}=\text{O}) \rightarrow \sigma^*(\text{S}-\text{S})$ interaction, in contrast to the $n(\text{X}) \rightarrow \sigma^*(\text{S}-\text{X})$ interaction which dominates in small molecules. Also contrary to the case of small molecules, the authors suggested that the orbital component of the interaction is small, though important for establishing the directionality of the interaction. Dispersion appears to be the major stabilising force, as optimization using a non-dispersion corrected level of theory gave unrealistically large distances.

The differences between Ch-bonding in proteins and small molecules is likely due to variation in orbital energies in the functional groups which are found in each class. While small molecule Ch-bond donors are characterised by easily accessible, low energy $\sigma^*(\text{Ch}-\text{X})$ orbitals, these are simply not found in proteins. Instead, donors are characterised by $\sigma^*(\text{S}-\text{S})$ or $\sigma^*(\text{S}-\text{C})$ orbitals, which are much higher in energy and less accessible to acceptors. Ch-bond acceptors, too, are markedly different between small molecules and proteins. In general, the HOMO of a system

(the most Lewis basic site) is dominated by lone pairs. This is observed in most small molecules, as they form Ch-bonds through these lone pairs. However, the amide bond, which is ubiquitous in proteins, shows an unusual inversion in orbital energies. The $\pi(\text{C}=\text{O})$ orbital is elevated with respect to the lone pair according to MP2 calculations, making it the more basic site. Structural data supports this assertion, as the Ch-bond donor usually approaches the top of the C=O bond in proteins, rather than the usual approach towards the oxygen lone pair (fig. 1.12).⁴⁰

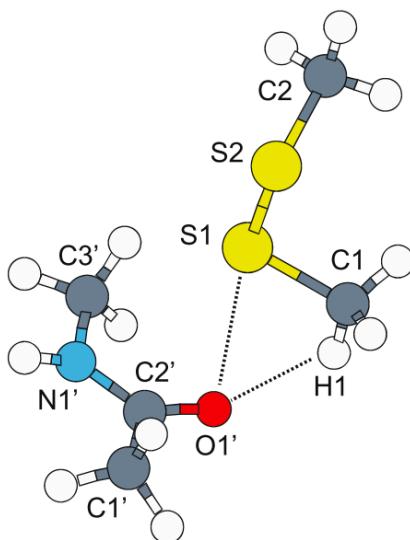


FIGURE 1.12: Directional preference for Ch-bonded interactions in proteins.

It is worth noting that selenium is also found in proteins as selenocysteine and selenomethionine. This would be expected to be an even stronger Ch-bond donor. However, selenoproteins are relatively few in number, precluding such extensive statistical analysis.⁴¹ They are only mentioned here for the sake of completeness.

Intramolecular interactions are just one instance of Ch-bonding in biological systems. Proteins often interact with ligands or substrates through H-bonds, so it is reasonable to propose that Ch-bonding could be applied in this field as well. Indeed, a protein-ligand Ch- and X-bonding interaction was used to target the gatekeeper methionine (MET146) residue of c-Jun N-terminal kinase 3 (JNK3) in a model study by Lange et al.⁴² In this work, a protein→ligand Ch-bond was used to stabilise the interaction. Inhibition of a cysteine protease using a variety of sulfur-containing

heterocyclic ligands was also investigated.⁴³ These ligands formed ligand→protein Ch-bonds, complementary to the work by Lange. Non-conventional protein-ligand interactions are summarised in a comprehensive review by Beno et al.⁴⁴

1.2.3.2 Nucleic acids

Nucleic acids represent another application of Ch-bonding in biology. In addition to their crucial role in the storage of genetic information, they have also been investigated as a structural material in nanotechnology. The ubiquity of H-bonds in nucleic acid complexes suggests that σ -hole interactions may also be used to direct formation of these complexes. X-bonding was indeed able to be used to direct formation of a Holliday junction between two DNA strands (fig. 1.13).⁴⁵ Holliday junctions are branched nucleic acid structures that appear in many biological processes including recombination and double-strand break repair. They are also useful as building blocks for DNA nanotechnology, where their self assembly and predictable geometry are exploited. The authors estimated that the X-bonding interaction (mediated through a bromo-substituent) was 2–5 kcal/mol stronger than the corresponding H-bond. A 2017 review identified a further 21 X-bonded nucleic acid structures.⁴⁶ X-bonding was in fact able to partially replace inter-strand H-bonding interaction in a DNA oligomer.⁴⁷

Ch-bonding, too, has been investigated in the context of nucleic acids. Sharma et al. recently investigated the possibility of forming Ch-bonded analogues of the canonical A:T/G:C base pairs in order to expand the genetic alphabet.⁴⁸ A G_{SeF}:C dimer was found to be more stable than the canonical G:C pair, where the SeF subscript indicates that one of the H-bond donors on the guanine was replaced by a selenenyl fluoride. Farrell et al. showed that introducing heavy chalcogens into nucleobases has profound impacts on their photochemistry, with implications for their use in photodynamic therapy.⁴⁹ The photophysics of selenonucleobases has been further explored, showing that they are uniquely photo-stable while efficiently populating

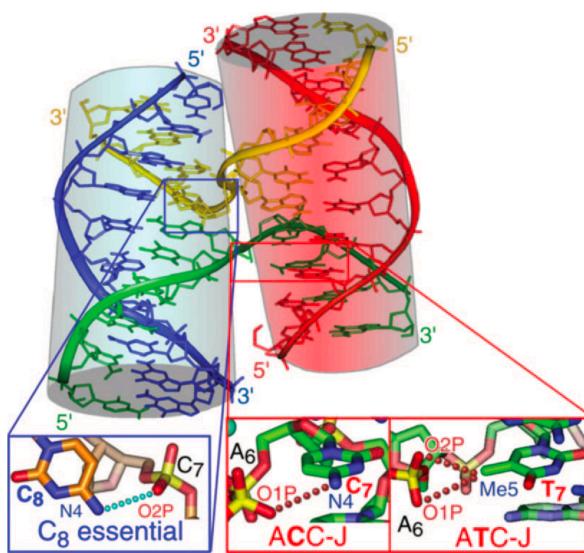


FIGURE 1.13: Holliday junction formed by X-bonding.⁴⁵

and depopulating their excited states.^{50–53} While none of these investigations specifically mentioned Ch-bonding, it is highly likely that it would come into play, which may provide a means of directing the activity of these photo-sensitisers.

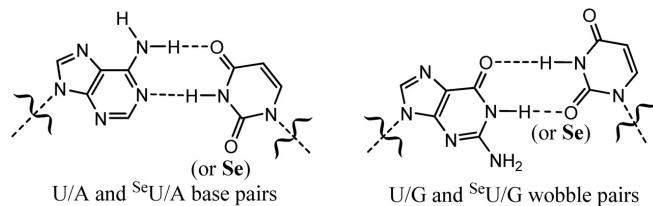


FIGURE 1.14: Canonical Watson-Crick U:A base pair, and the “wobble” U:G base pair in RNA. The latter is disfavoured when an oxygen atom in uracil is substituted by selenium.

Selenium has also been incorporated into nucleobases as a H-bond acceptor, in which it was shown that selenonucleobases can effectively discriminate against the formation of a so-called “wobble” base pair (U:G as opposed to U:A in RNA, or T:G/T:A in DNA, fig. 1.14).^{54,55} Another advantage of selenonucleobase incorporation is the ability to use the heavy selenium atom for MAD/SAD phasing of macromolecular crystal structures.⁵⁶ This was also the motivation for a 2019 study in which the selenium was incorporated into the phosphate backbone.⁵⁷ A major advantage of this approach is that the bulk of the selenium atom does not need

to be accommodated in the interior of the double helix, so structural distortions are minimised. To date, there have not been any drugs which target nucleic acids through Ch-bonding, which is an area we explore in chapter 7.

1.3 Experimental methods

As mentioned above, X- and Ch-bonded interactions were originally identified crystallographically, with additional spectroscopic evidence being present. These pioneering methods have not changed a great deal, and x-ray crystallography is still the most powerful tool we have to study these interactions. We also use a number of spectroscopic and computational techniques to probe the nature of Ch-bonding. Some of the methods used in this thesis are explained below.

1.3.1 Computational methods

Computational methods are an indispensable tool for chemists, allowing us to study molecules too fragile to isolate in the laboratory, and extract further insight from real-world systems. A number of the techniques used in this thesis are briefly explained below.

1.3.1.1 Natural Bond Orbital theory

The SCF procedure, which is at the heart of almost all quantum chemistry calculations, affords a set of solution wave functions Ψ to the Schrödinger equation (eq. (1.1)) using a Hamiltonian which is approximated to some degree (e.g. the Fock operator in the Hartree-Fock method).

$$\hat{H}\Psi = E\Psi \tag{1.1}$$

Each one electron wave function, or molecular orbital (MO), describes the distribution of an electron in the field of the nuclear potentials, and is generally constructed from a Linear Combination of Atomic Orbitals (LCAO)*, that is, sums and differences of atom centred basis functions of various symmetries. The Pauli exclusion principle states that a (many-particle) wave function must be antisymmetric with respect to exchange of two fermionic particles such as electrons. This means that for every MO resulting from the sum of two atomic orbitals, there must be a corresponding MO resulting from the difference of the same two atomic orbitals, from which we obtain the familiar MO construction for simple diatomics (fig. 1.15).

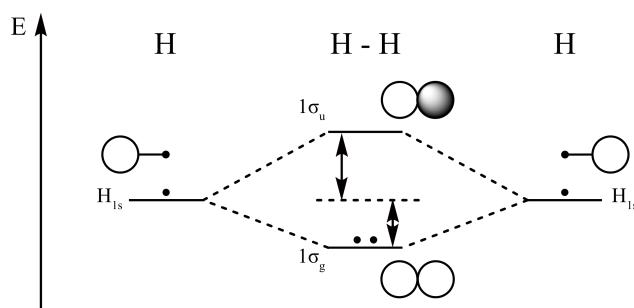


FIGURE 1.15: MO diagram for dihydrogen. Reproduced from reference [58].

In more complex polyatomic species, these MOs are most easily interpreted as eigenfunctions of the molecular Hamiltonian, and their energies are the corresponding eigenvalues. These orbitals describe the distribution of each electron throughout the molecule and hence the bonding within the molecule, but do not translate well into the more intuitive valence bond description of bonding that is most useful to chemists. From a series of MOs, there is not an easy way to draw a Lewis structure of the molecule at hand. This is because MOs tend to be highly delocalised throughout the molecule, which is obvious even in the relatively simple case of benzene (fig. 1.16).

The Natural Bond Orbital (NBO) scheme attempts to ameliorate this delocalisation, by transforming the molecular orbitals into NBOs which correspond to features

*An alternative approach is that of plane-wave basis sets, which are better suited to infinitely periodic systems.

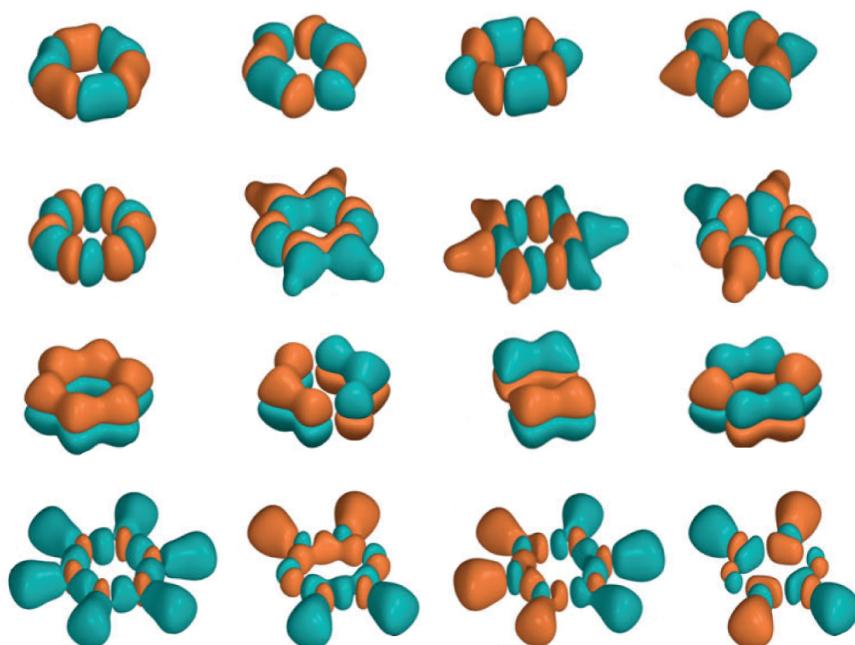


FIGURE 1.16: Molecular orbitals of benzene. Reproduced from reference [59].

found in a Lewis structure.⁶⁰ These take the form of one centre and two centre elements (lone pairs and bonding orbitals respectively), which together form the optimal Lewis structure for a molecule. The NBO scheme also provides a framework for non-Lewis features, such as non-valence electrons which are found in core NBOs. As a consequence of the transformation from atomic orbitals to NBOs, higher energy two centre elements arise, which correspond to anti-bonding orbitals whose partial occupancy leads to reduction of bond order. Rydberg NBOs complete the span of the NBO space, which are high energy, highly diffuse orbitals which typically contribute little to the overall chemistry of the system. Examples of NBOs in a simple molecule are shown in fig. 1.17

We use NBO theory to rationalise the hyperconjugative effects present in Ch-bonded complexes, namely, the $n \rightarrow \sigma^*$ overlap which appears to be present. NBO analysis allows a numerical value to be assigned to the degree of this overlap, in terms of the formal number of electrons which are “transferred” from one orbital to the other. The energies of each NBO can then be used to calculate the stabilisation afforded by a given delocalisation.

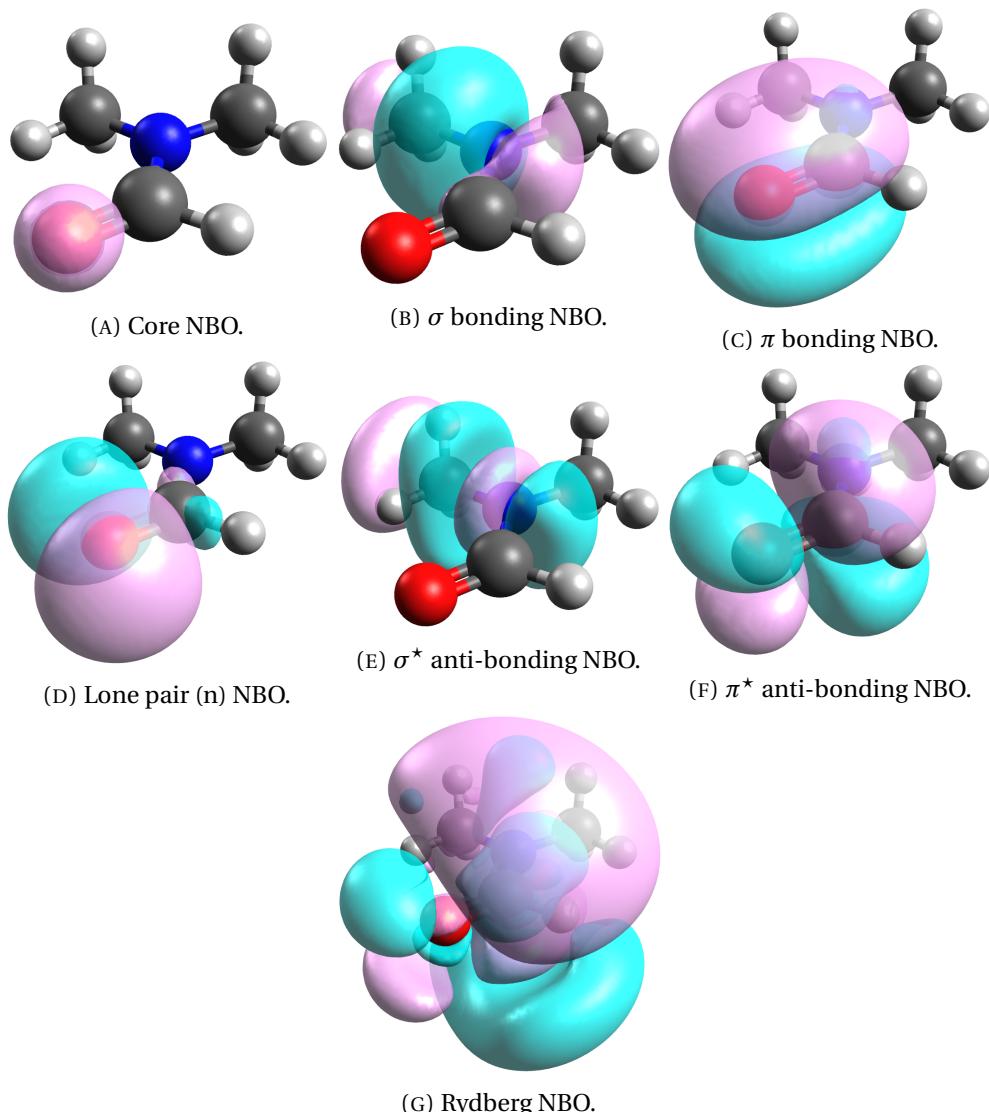


FIGURE 1.17: Representative Natural Bond Orbitals of a DMF molecule.

In summary, NBOs are a mathematical transformation of highly delocalised MOs into functions that correspond to the Lewis structure interpretation of chemical bonding. Although this generates an ideal Lewis structure of a species, departures from this ideal (in the form of delocalisation and hyperconjugative effects) can be seen in decreased occupancy of bonding orbitals and increased occupancy of anti-bonding orbitals. This allows for a mathematically rigorous and quantitative treatment of bonding, using familiar chemical concepts.

1.3.1.2 Quantum Theory of Atoms In Molecules

The Quantum Theory of Atoms In Molecules (QTAIM) is a description of bonding that is complimentary to that of NBO theory.⁶¹ Rather than describing bonds in terms of orbitals, it uses features of the electron density to define bonding. This is particularly attractive as orbitals and wave functions are not observable quantities, whereas electron density is. Indeed electron density can be measured directly through diffraction experiments (see section 1.3.2.1). It can also be calculated from a molecular wave function, which can itself be calculated by any number of methods.

To discuss QTAIM, we must first discuss some mathematical aspects of topology. The fundamental quantity in QTAIM is the electron density $\rho(\mathbf{r})$, which constitutes a scalar field over the molecule. That is, every point in space has a single number representing the electron density at that point (see fig. 1.18 for an example). Differentiating $\rho(\mathbf{r})$ affords the gradient $\nabla\rho(\mathbf{r})$, a measure of the local rate of change (in space) of the electron density. $\nabla\rho(\mathbf{r})$ is a vector quantity, with a magnitude and direction, which thus forms a vector field. Differentiating $\nabla\rho(\mathbf{r})$ again affords the second derivative of $\rho(\mathbf{r})$, also called the Hessian \mathbf{H} . \mathbf{H} is a tensor quantity, which is most commonly represented as a 3×3 matrix. This is not easily represented as a field, but the three eigenvalues (components of the diagonalised Hessian) are useful quantities, which each represent the rate of change of $\nabla\rho(\mathbf{r})$ at a given point. These are given the symbols λ_1 , λ_2 , and λ_3 . The sum of these eigenvalues is called the Laplacian of the electron density, which is given the symbol $\nabla^2\rho(\mathbf{r})$. Maps of $\nabla^2\rho(\mathbf{r})$ can recover some chemical features of molecules that are reminiscent of the orbital structure, even though it is derived completely from the $\rho(\mathbf{r})$ scalar field (fig. 1.19).

A QTAIM atom is defined by a volume bounded by a surface of zero electron density gradient flux (fig. 1.20). That is, there are no field lines of $\nabla\rho(\mathbf{r})$ crossing the surface. A molecule can thus be partitioned into atoms, each of which is strictly additive and transferable. Furthermore, atomic contributions to molecular properties can

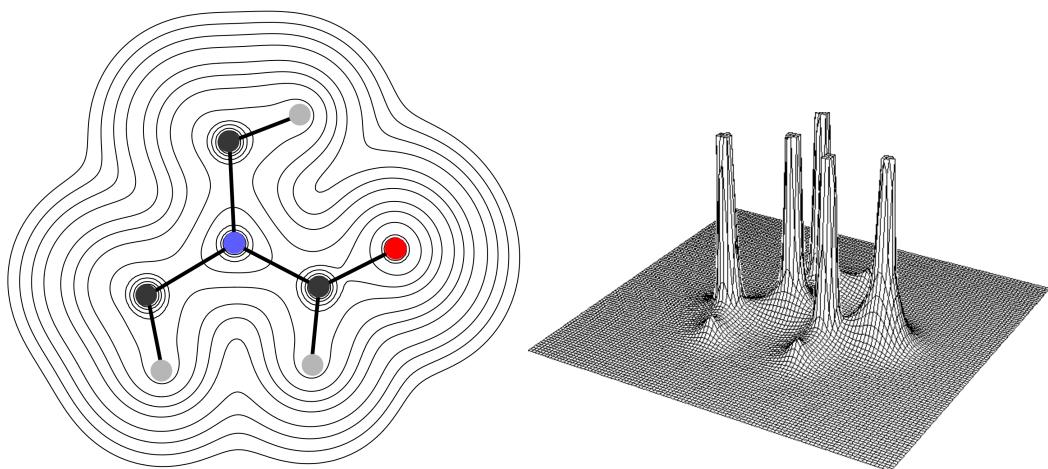


FIGURE 1.18: Electron density in the plane of a DMF molecule. The nuclei correspond to the large peaks, and bonds correspond to the ridges linking them.

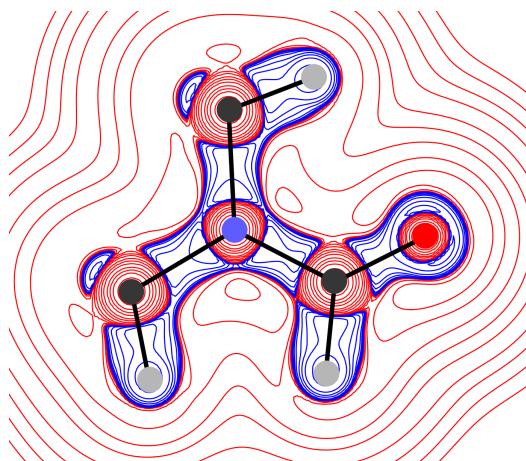


FIGURE 1.19: Laplacian of the electron density $\nabla^2 \rho(\mathbf{r})$ of DMF. Positive values are shown in red and negative values in blue. $\nabla^2 \rho(\mathbf{r})$ can be seen to recover the shell structure of atoms, as well as chemical features such as bonds and lone pairs.

be uniquely defined. This is similar to the idea of functional groups in organic chemistry. In the same way a carboxylic acid functional group can be transferred between molecules while retaining its characteristic properties (its acidity), a QTAIM atom can also be transferred, as its properties depend *only* on the topology of the electron density.

Each QTAIM atom is associated with a local maximum of electron density which is located at the nucleus. At this point $\nabla \rho(\mathbf{r})$ is zero, so it is termed a critical point. As it is a maximum, $\rho(\mathbf{r})$ will decrease in all directions, therefore all three eigenvalues of

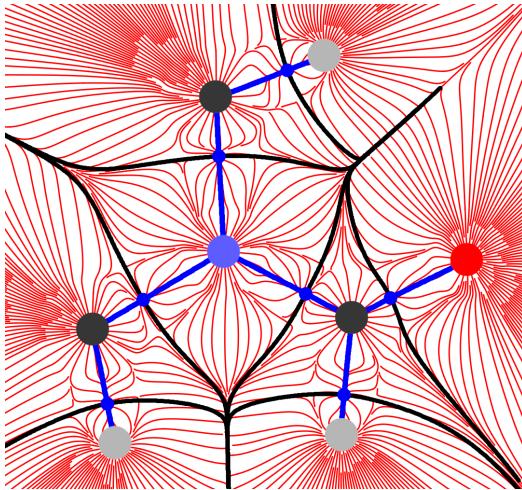


FIGURE 1.20: Electron density gradient $\nabla\rho(\mathbf{r})$ vector field of DMF. All field lines (red) converge on the nuclei, and asymptotically approach the zero flux surfaces (black lines) which bound the QTAIM atoms. Bond paths and critical points are shown in blue.

\mathbf{H} will be negative. These atomic critical points are therefore termed $(3, -3)$ critical points, as they exist in three dimensional space and have three negative eigenvalues.

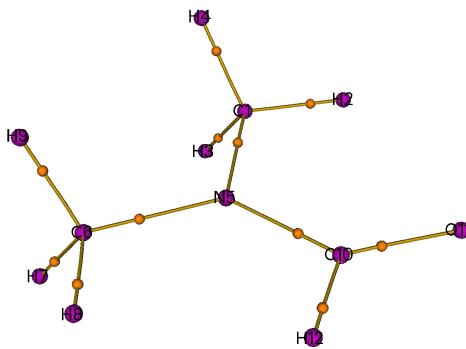


FIGURE 1.21: $(3,-3)$ (pink), and $(3,-1)$ (orange) critical points of DMF. Bond paths are shown in yellow.

Nuclei are not the only place critical points are found (where $\nabla\rho(\mathbf{r})$ is zero). Atomic critical points contained within neighbouring QTAIM atoms (sharing a common zero gradient flux surface) are linked by lines along which $\rho(\mathbf{r})$ decreases in the two directions perpendicular to the line. In two dimensions this can be visualised as a ridge connecting two mountain peaks. This line is termed a bond path, as the presence of such a line is a prerequisite for bonding to occur (although the reverse is

not necessarily true). At the point where the bond path crosses the zero gradient flux surface there will be another critical point where $\nabla\rho(\mathbf{r})$ is zero. This type of critical point is called a bond critical point (BCP) or (3, -1) critical point, as $\rho(\mathbf{r})$ decreases in two directions but increases in one direction (along the bond path). \mathbf{H} at a (3, -1) critical point therefore has two negative eigenvalues and one positive eigenvalue. An example of a molecular structure derived from the critical points and bond paths is shown in fig. 1.21. There are also critical points found at the centres of rings (ring or (3, 1) critical points) or at the centres of cage-like structures (cage or (3, 3) critical points), however these are not discussed in this thesis.

As stated before, the presence of a bond path and BCP is necessary but not sufficient to say there is chemical bonding occurring. In this work we therefore use the presence of a bond path, as measured from experimental electron density, to support our claim that Ch-bonding is occurring. We also use the values of properties related to $\rho(\mathbf{r})$ at the BCP to characterise the type of bonding that we observe. Although this does not replace a proper energy decomposition analysis, certain types of interaction have characteristic values of $\rho(\mathbf{r})$ and $\nabla^2\rho(\mathbf{r})$ at the BCP. Covalent bonds are characterised by accumulation of electron density between the nuclei, and this is reflected in relatively large values for $\rho(\mathbf{r})$, and a large and negative value of $\nabla^2\rho(\mathbf{r})$. Closed shell interactions, on the other hand, do not show this accumulation of charge, so at the BCP $\rho(\mathbf{r})$ is small, and $\nabla^2\rho(\mathbf{r})$ is positive.

In contrast to NBO theory, QTAIM is a complementary, but equally quantitative and mathematically sound, interpretation of bonding in chemical species. It has the unique benefit of being grounded solely in the experimentally observable property of the electron density, while still being able to recover chemically intuitive aspects of bonding.

1.3.2 X-ray crystallography

Single crystal x-ray crystallography is the main experimental technique used in this work. It allows unambiguous determination of the structure of compounds (including stereochemistry), and extremely accurate measurement of geometric parameters such as bond lengths. Practically speaking, a structure is determined by mounting a single crystal of a compound on a goniometer which can rotate the crystal through any orientation, then observing the position and intensity of diffracted x-ray beams. Rotating the crystal allows different diffraction spots to be observed, and a more complete dataset to be collected. The position of the diffraction spots provides information about the size and shape of the unit cell, and the intensities provide information about the internal structure i.e. the locations of the atoms in the unit cell. A full discussion of the mathematics of diffraction is outside the scope of this thesis, but the interested reader is referred to *X-Ray Structure Determination: A Practical Guide* by Stout and Jensen.⁶²

1.3.2.1 Charge density refinement

Fundamental to the technique of x-ray structure determination is the process of refinement, by which parameters of the model (typically the molecular geometry) are optimised to reproduce the observed intensity data as well as possible. Mathematically, this is performed as a least-squares optimisation, where the function

$$\sum_{r=1}^m w_r (f_{\text{obs},r} - f_{\text{calc},r})^2 \quad (1.2)$$

is minimised, where $f_{\text{obs},r}$ corresponds to the observed intensity, and $f_{\text{calc},r}$ corresponds to the intensity calculated from the model by Fourier transform. w_r is a weighting function used to reflect the confidence in any given observed data point.

Assuming the measured intensities are free of any error,* this function could theoretically be minimised to zero i.e. the model is a complete and true representation of the crystal. The question then becomes, how do we determine $f_{\text{calc},r}$ from our model? To answer this, we must consider what our model actually *is*.

The simplest refinement model consists of atomic coordinates for each atom, plus an overall scale factor (for now we will neglect the anisotropic displacement parameters and chemical occupancies). However, in order to calculate $f_{\text{calc},r}$ by Fourier transform, we must introduce some description of what the atoms which we are locating “look like”. This forms a hidden, but still crucially important, part of the model. Typically, atoms are described in reciprocal space as spherically symmetric functions whose contribution to $f_{\text{calc},r}$ is only dependent on $(\sin\theta)/\lambda$ (fig. 1.22). This is a consequence of the fact that x-rays are scattered by the *electron cloud* of the atom.[†] At $(\sin\theta)/\lambda = 0$ the scattering factor is equal to the number of electrons in the atom. However, the finite size of the electron cloud causes destructive interference and attenuation of the scattering factor as $(\sin\theta)/\lambda$ increases, as x-rays scattered from one side of the electron cloud will be out of phase with those scattered from the other side. Contrast this with neutron diffraction, where the neutrons are scattered predominantly by the point-like nucleus, affording a flat scattering function in reciprocal space. There is almost no destructive interference, and thus neutron crystallography is characterised by an abundance of strong high angle data. The scattering factor functions for all elements of the periodic table (and many ions) have been calculated based on theoretical electron densities from Hartree-Fock wave functions, and the results are available in the International Tables.⁶³ These functions are mercifully incorporated in crystallographic refinement software, shielding them from the user.

*This is practically never the case, but careful measurement and modern corrections can approach this limit.

[†]X-rays have much less energy than the mass-energy of subatomic particles, and so are scattered in the low energy limit of Compton scattering, known as Thompson scattering. The scattering power of a particle is inversely proportional to the square of its mass, so the massive nuclei contribute negligibly to scattering compared to electrons.

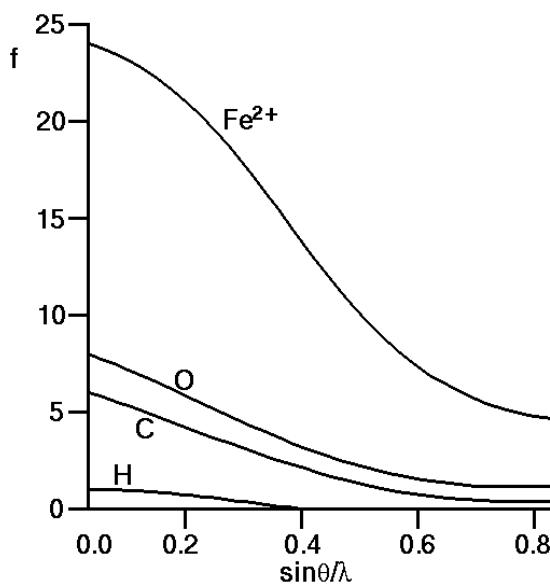


FIGURE 1.22: Scattering factors for some atoms as a function of $(\sin \theta)/\lambda$.

From the above discussion, one major shortcoming of this approach should be clear. Namely, that the electron density of bonded atoms is *not* spherically symmetric. Perturbations such as bonding electron density, lone pairs, and indeed σ -holes can, and do, affect the observed scattering factors of atoms, which are not taken into account in typical refinement software. An extreme case of this, and one that is familiar to any crystallographer, is that of the hydrogen atom. As the lightest element with only one electron, perturbations due to bonding have an enormous effect, such that the typical C–H bond length is underestimated by some 0.13 Å.⁶⁴ A subtler but more interesting manifestation of this was identified in 1968 by Coppens.⁶⁵ X-ray and neutron structures were determined for a crystal of 1,3,5-triazine, and inspection of the difference (a so-called X – N map) revealed unusual systematic variations in the anisotropic displacement parameters. The nitrogen atoms were deformed radially to the ring, while the carbon atoms were deformed tangentially (fig. 1.23). This is chemically intuitive, as we would expect the lone pairs of the nitrogens to extend radially, while the π -electrons of the carbon atoms would form a ring. The fact that these deformations were incorporated into the anisotropic displacement parameters in the x-ray structure highlights a deficiency of the typical method of refinement.

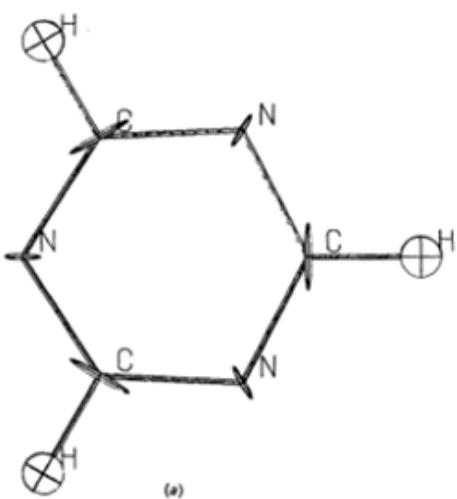


FIGURE 1.23: X – N map of 1,3,5-triazine, reproduced from Coppens.⁶⁵

Evidence for aspherical atoms can even be traced back to seminal studies performed by Bragg, in which the famous 222 reflection of a diamond crystal was observed. The 222 reflection is a systematic extinction of the $Fd\bar{3}m$ space group to which diamond belongs, but its absence is based on the assumption of spherical atoms. In diamond, the carbon atoms are actually tetrahedral in form due to the bonding electron density, which lowers the space group symmetry and causes the 222 reflection to be present, though very weak.⁶⁶ Though cutting edge for the time, Bragg's experiment was rather primitive, and multiple scattering likely contributed some intensity to the observed 222 reflection. The observation was nonetheless correct, and the tetrahedral nature of the carbon atoms in diamond has subsequently been proven time and time again.

Although such perturbations generally only manifest in very high quality structures (aspherical atoms are typically the least of a crystallographer's worries), a number of methods have been proposed to improve refinements, and indeed gain further chemical knowledge from the experimentally determined aspherical electron density. A simple approach is the use of tabulated aspherical scattering factors for atoms of distinct chemical types. An example of this is the ELMAM2 database,

developed by Jelsch and co-workers.⁶⁷ The ELMAM2 database consists of experimentally derived scattering factors for all common atom types, with a focus on those found in amino acids and peptides. A refinement using this database would begin by assigning a type to each atom, which can be done relatively easily using bonding geometries from a more primitive refinement. This can afford substantial improvement in refinement statistics by providing a more accurate model, at a very modest computational cost. The obvious drawback of this approach is that the scattering factors must already be tabulated, and the enormous diversity beyond the biochemically important C, H, N, and O precludes the assembly of a truly comprehensive database. Furthermore, as the scattering factors are pre-tabulated, there is little chemical information that can be extracted from the model beyond somewhat improved precision in geometric parameters.

An improved approach is to compute scattering factors on a case-by-case basis. This is the rationale behind Hirshfeld Atom Refinement (HARt), first developed by Jayatilaka and Dittrich.⁶⁸ HARt is an iterative procedure in which the atomic coordinates from a spherical atom refinement (perhaps after correcting C–H bond distances) are used to variationally calculate a wave function, from which is calculated an electron density and thus scattering factors. These new scattering factors improve the model after refinement, from which is calculated a *new* wave function, and so on until the cycles of least squares refinement and wave function calculations converge. Clearly this repeated calculation of a wave function is computationally expensive, with even relatively simple molecules taking minutes. Fortunately, however, even very inexpensive levels of theory tend to generate sensible densities for the ground state. An advantage over the database approach is that one can capture unusual effects in the electron density (such as various non-bonding interaction), provided that they are correctly modelled by the underlying quantum chemistry theory used to generate the wave function. However this raises a philosophical question: what exactly we are hoping to get out of this procedure? It can certainly provide improved refinement statistics, including anisotropic refinement of hydrogens in agreement

with neutron data, but if we are seeking chemical information about the system, we might as well simply use a gas-phase optimised wave function and dispense with the crystallography.

Related to this is the field of quantum crystallography, which differs somewhat in motivation, if not in methodology. Quantum crystallography is concerned with extracting a wave function from diffraction data, rather than using a wave function to improve a crystallographic model. Clearly there is significant overlap. One method takes inspiration from perturbation theory, where, as in MP2, the Hartree-Fock wave function is taken as the zeroth order approximation. The diffraction data is then used to construct a perturbative term, which introduces a degree of correlation to the wave function, bringing it closer to the truth (at least within the Born-Oppenheimer approximation).⁶⁹ Even with the power of modern computers, this is not an easy task, and quantum crystallography remains an active and exciting area of research. It is also worth mentioning the x-ray constrained wave function method developed by Grimwood et al.⁷⁰ This is simply a Hartree-Fock wave function calculations that is constrained via a Lagrangian multiplier to reproduce the observed intensities in the experimental data, assuming there are no crystallographic artifacts in the data such as disorder or twinning. That is to say, the crystal is simply comprised of infinitely repeating symmetry related molecular units. The x-ray constrained wave function can be used to calculate a density, which can be used in a HARt-like procedure, which has been named x-ray wave function refinement (XWR).⁷¹ Both quantum chemistry and crystallographic refinement codes have been implemented in the software package TONTO.⁷²

Perhaps the most powerful approach for experimental crystallographers is that of multipole refinement using the formalism developed by Hansen and Coppens.⁷³ While superficially similar to quantum crystallographic approaches, no chemical restraint* is placed on the resulting electron density, allowing for the modelling of

*This is not strictly true, as electroneutrality is routinely imposed as a constraint on multipole models.

phenomena which are not described by *ab initio* calculations. The electron density is expanded in a series of atom-centred spherical harmonic basis functions, the local coordinate systems of which are often chosen to align with chemical features. Each basis function requires two parameters for the occupancy and expansion, in addition to the position and anisotropic displacement parameters for each atom. We therefore require a large number of observations to ensure the system is over-determined, and can be treated in a least-squares manner. Furthermore, the ADPs and dipolar basis terms are highly correlated, leading to an unstable solution if refined simultaneously (fig. 1.24).

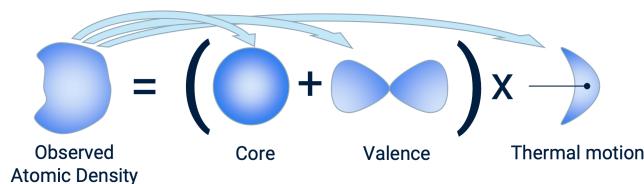


FIGURE 1.24: Thermal motion (described by anisotropic displacement parameters) and dipolar basis terms are highly correlated.

For this reason, it is critical to establish accurate atomic positions and ADPs before refining the multipole parameters and charge density. Traditionally this was done using neutron data which, as mentioned above, is insensitive to electron density perturbations as neutrons are scattered almost exclusively by the nucleus. Unfortunately neutron sources are notoriously expensive and low-flux, requiring extremely large crystals which introduces further errors due to extinction. With the advent of modern x-ray diffractometer technology, we can luckily determine atomic coordinates and ADPs using x-ray data alone. This is done using high angle reflections, which are dominated by core scattering and thus contain little data from the diffuse bonding electron density. Core electron density is almost perfectly spherical, allowing ADPs to be correctly refined from this data. With accurate atomic coordinates and ADPs in hand, hydrogen atoms can be located in the low angle data, and then the multipole parameters refined to convergence. Multipole refinement can benefit enormously from an initial database model, particularly for hydrogen atoms, so it is no surprise that the techniques are often used sequentially.^{74,75} The

resulting electron density can be used to characterise molecules and predict reactivity, particularly when interpreted within the powerful Quantum Theory of Atoms In Molecules framework of Bader.⁶¹ The obvious attraction of this method is that it is almost entirely experimental. However, the atom centred multipole functions and their resulting sum, while superficially resembling a wave function, is not anti-symmetric and is therefore in violation of the Pauli exclusion principle. It therefore cannot be used to determine *all* properties of a system.

To summarise, electron density can be measured in a crystal by x-ray diffraction, and various models used to fit it. The “gold standard” would be the derivation of a wave function, but at present this has not been done without the introduction of chemical constraints. In practise, the most accurate and most informative fitting of electron density to diffraction data is done with a series of atom-centred spherical harmonic functions (the Hansen-Coppens formalism). The resulting electron density can be used in QTAIM analysis, or used to construct electrostatic potential maps, which can guide chemists in their understanding of molecules in crystals.

1.3.3 Nuclear Magnetic Resonance

Nuclear magnetic resonance spectroscopy (NMR) is an essential tool for all organic chemists, as it is able to give relatively detailed structural information quickly and cheaply. It is based on the magnetic moment of certain nuclei, which are usually randomly oriented but spontaneously align in the presence of a strong magnetic field.

1.3.3.1 Solid State NMR

The chemical shift of a nucleus is not a simple scalar quantity, as solution phase NMR experiments might suggest. The shielding, and hence the effective magnetic field felt by the nucleus depends on the electronic environment around the nucleus,

which is decidedly *anisotropic*, therefore incapable of being expressed as a scalar. Depending on the orientation of the electron cloud (and other shielding/deshielding influences) around the nucleus with respect to the magnetic field, different chemical shifts will be observed for the same nucleus. This phenomenon is the chemical shift anisotropy of a nucleus, and it is described mathematically by a second rank tensor, which is geometrically depicted as an ellipsoid.

In a solution phase NMR experiment, rapid and random tumbling of the molecules averages out the anisotropy of each signal, affording a sharp peak at the isotropic chemical shift. This can be simulated to some degree by magic angle spinning in solids experiments, but it is often the case that the chemical shift anisotropy is more interesting than the isotropic shift, so a solid phase NMR experiment is the only way to examine it.

The shape of the chemical shift tensor can provide insight into the electronic environment of a nucleus, with large shielding components often being aligned with electronic features such as bonds or lone pairs. It is often convenient to define a principal axis system (PAS) for the tensor, consisting of two components which describe the longest and shortest axes of the ellipsoid, and the axis perpendicular to both. The PAS of the chemical shielding tensor is denoted by the lowercase x , y and z , and the components of the tensor which are aligned with these axes are labelled σ_{xx} , σ_{yy} and σ_{zz} . The laboratory reference frame is denoted by the uppercase X , Y and Z , with the magnetic field B_0 being aligned with the Z axis. A common method for transforming one coordinate system to another is through the use of Euler angles α , β and γ , which define sequential right-handed rotations about the axes of the rotating frame. As the chemical shielding is necessarily invariant to rotation about Z , we can dispense with the first of these angles α , and simply refer to the latter two as the polar angles $\beta = \theta$ and $\gamma = \phi$. The chemical shielding tensor is thus depicted in fig. 1.25, and this will be the convention used in this work.

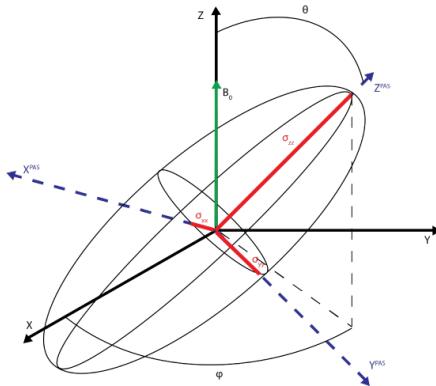


FIGURE 1.25: Representation of the principal axis system (PAS, blue dotted lines) with respect to the laboratory reference frame (black solid lines). The polar angles θ and ϕ relate the former to the latter. The principal components of the tensor are shown in red, and the overall shape is shown by the black ellipses.

In almost all experiments, the chemical shielding tensor σ is only indirectly measured, via the chemical shift δ , which is defined with respect to a reference nucleus such that $\delta = \sigma_{\text{ref}} - \sigma_{\text{sample}}$. As the coordinate system as described above is somewhat arbitrary, the IUPAC convention defines the three principal components of the chemical shift tensor δ_{11} , δ_{22} and δ_{33} such that $\delta_{11} \geq \delta_{22} \geq \delta_{33}$. The isotropic chemical shift is given by the average $\delta_{\text{iso}} = \frac{(\delta_{11} + \delta_{22} + \delta_{33})}{3}$. There are two other conventions for describing chemical shift anisotropy, which are useful in different contexts. The Herzfeld Berger convention represents the chemical shift in terms of the span of the signal $\Omega = \delta_{11} - \delta_{33}$ and the skew $\kappa = \frac{3(\delta_{22} - \delta_{\text{iso}})}{\Omega}$ where δ_{iso} is the same as in the IUPAC convention. This is particularly useful for intuitively describing the powder line shape formed by a given signal. The other convention is the Haeberlen convention, which defines the three principal components as $|\delta_{zz} - \delta_{\text{iso}}| \geq |\delta_{xx} - \delta_{\text{iso}}| \geq |\delta_{yy} - \delta_{\text{iso}}|$, and the parameters $\Delta_{\text{CSA}} = \delta_{zz} - \delta_{\text{iso}}$ and $\eta_{\text{CSA}} = \frac{\delta_{xx} - \delta_{yy}}{\delta_{zz} - \delta_{\text{iso}}}$. This convention is most useful for describing the angular dependence of a chemical shift on the polar angles θ and ϕ , which is given by the relationship⁷⁶

$$\delta_{\text{obs}} = \delta_{\text{iso}} + \Delta_{\text{CSA}} \left(\frac{3 \cos^2 \theta - 1 + \eta_{\text{CSA}} \sin^2 \theta \cos 2\phi}{2} \right) \quad (1.3)$$

In a non-spinning polycrystalline or amorphous powder, the above expression can be integrated over the range of chemical shifts to give the characteristic powder line shape see in fig. 1.26, reproduced from the work of Facelli, Grant, and Michl.⁷⁷ The sample consists of randomly oriented tensors, all of which contribute to the signal. The principal values of the tensor δ_{11} , δ_{22} and δ_{33} correspond to the two extrema of the line, and the central peak.

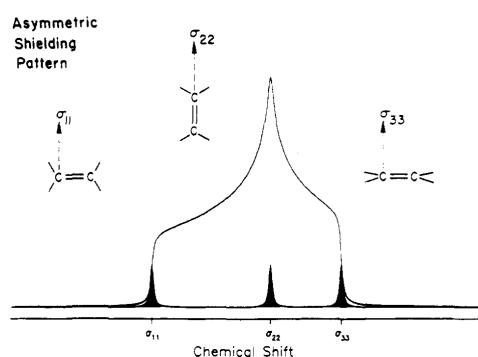


FIGURE 1.26: Characteristic solid-state NMR powder line shape for an asymmetric tensor e.g. in an alkene. The positions of the three principal values are shown.

Spinning at the magic angle can improve the S/N ratio by partially averaging out the chemical shift anisotropy and dipolar relaxation. Instead of the powder pattern, we instead observe a sharp isotropic chemical shift and a series of spinning sidebands which are separated from the isotropic peak by multiples of the spinning frequency. The radiated power is thus concentrated in narrower bands, leading to a much improved S/N ratio.

An example is presented in fig. 1.27 of proton decoupled solid state ^{31}P spectra of barium diethyl phosphate, reproduced from the work of Herzfeld and Berger.⁷⁸ In the first spectrum, no magic angle spinning was used, and the line shape is characteristically broadened into an asymmetric peak (the powder line shape). Note the poor S/N ratio due to the broad peak. The S/N ratio can be seen to improve as the spinning speed increases, and the sidebands become more sparse, affording a stronger signal overall.

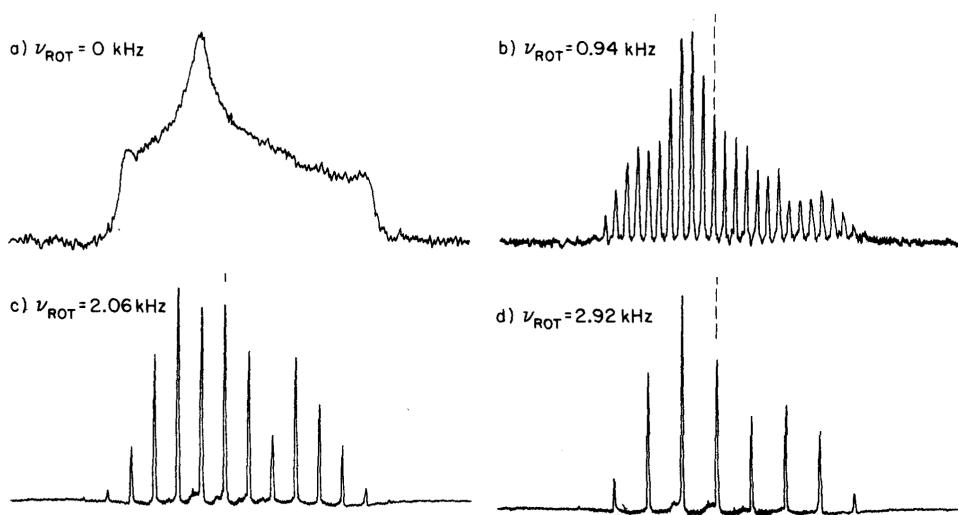


FIGURE 1.27: Proton decoupled solid state ^{31}P spectra of barium diethyl phosphate spinning at the magic angle, at the speed specified. The isotropic chemical shift is shown by the vertical dotted line.

The principal values of the tensor are clearly easily obtained from a non-spinning sample, as they can practically be read off the spectrum. However, the extremely poor S/N ratio (especially for a relatively insensitive nucleus like ^{77}Se) limits the utility of this method. Spinning at the magic angle is clearly necessary to improve the S/N ratio, however this obscures the true locations of the extrema of the signal, therefore the values of δ_{11} and δ_{33} , as well as modifying the position of the maximum (δ_{22}).

There exist programs (such as SOLA, integrated within TopSpin by Bruker BioSpin) which can fit an experimental line shape or sideband manifold affording the principal values of the tensor, however there is clearly a trade-off between obtaining a good S/N ratio and having enough sidebands within the limits of the signal for a meaningful fit. Another issue is encountered with preferred orientation of crystallites, as in powder diffraction, which is not modelled in the fitting routine, leading to a poorer fit. Perhaps the most fundamental issue stems from the fact that the sample is a powder of randomly oriented particles. This means that the absolute orientation of the tensor with respect to the molecule cannot be determined from this kind of

experiment. Nonetheless, the *shape* of the tensor is still valuable information that may be able to shed light on Ch-bonding in the solid phase.

References

- (1) Benesi, H. A.; Hildebrand, J. H. *Journal of the American Chemical Society* **1949**, *71*, 2703–2707,
DOI: [10.1021/ja01176a030](https://doi.org/10.1021/ja01176a030).
- (2) Hassel, O.; Hvoslef, J.; Vihovde, E. H.; Sørensen, N. A. The Structure of Bromine 1,4-Dioxanate. 1954,
DOI: [10.3891/acta.chem.scand.08-0873](https://doi.org/10.3891/acta.chem.scand.08-0873).
- (3) Bent, H. A. *Chemical Reviews* **1968**, *68*, 587–648,
DOI: [10.1021/cr60255a003](https://doi.org/10.1021/cr60255a003).
- (4) Dumas, J.-M.; Gomel, M.; Guerin, M., *Molecular interactions involving organic halides*, 1983, pp 985–1020,
DOI: [10.1002/9780470771723.ch2](https://doi.org/10.1002/9780470771723.ch2).
- (5) Legon, A. C. *Chemistry - A European Journal* **1998**, *4*, 1890–1897,
DOI: [10.1002/\(SICI\)1521-3765\(19981002\)4:10<1890::AID-CHEM1890>3.0.CO;2-4](https://doi.org/10.1002/(SICI)1521-3765(19981002)4:10<1890::AID-CHEM1890>3.0.CO;2-4).
- (6) Legon, A. C. *Angewandte Chemie - International Edition* **1999**, *38*, 2686–2714,
DOI: [10.1002/\(SICI\)1521-3773\(19990917\)38:18<2686::AID-ANIE2686>3.0.CO;2-6](https://doi.org/10.1002/(SICI)1521-3773(19990917)38:18<2686::AID-ANIE2686>3.0.CO;2-6).
- (7) Corradi, E.; Meille, S. V.; Messina, M. T.; Metrangolo, P.; Resnati, G. *Angewandte Chemie - International Edition* **2000**, *39*, 1782–1786,
DOI: [10.1002/\(SICI\)1521-3773\(20000515\)39:10<1782::AID-ANIE1782>3.0.CO;2-5](https://doi.org/10.1002/(SICI)1521-3773(20000515)39:10<1782::AID-ANIE1782>3.0.CO;2-5).

- (8) Metrangolo, P.; Meyer, F.; Pilati, T.; Resnati, G.; Terraneo, G. *Angewandte Chemie - International Edition* **2008**, *47*, 6114–6127,
DOI: [10.1002/anie.200800128](https://doi.org/10.1002/anie.200800128).
- (9) Priimagi, A.; Cavallo, G.; Metrangolo, P.; Resnati, G. *Accounts of Chemical Research* **2013**, *46*, 2686–2695,
DOI: [10.1021/ar400103r](https://doi.org/10.1021/ar400103r).
- (10) Walter, S. M.; Kniep, F.; Herdtweck, E.; Huber, S. M. *Angewandte Chemie - International Edition* **2011**, *50*, 7187–7191,
DOI: [10.1002/anie.201101672](https://doi.org/10.1002/anie.201101672).
- (11) Terada, S.; Hirai, M.; Honzawa, A.; Kitagawa, O.; Kamizela, A.; Wzorek, A.; Soloshonok, V. A. *Chemistry - A European Journal* **2017**, *23*, 14631–14638,
DOI: [10.1002/chem.201703308](https://doi.org/10.1002/chem.201703308).
- (12) Takagi, K.; Yamauchi, K.; Murakata, H. *Chemistry – A European Journal* **2017**, *23*, 9495–9500,
DOI: [10.1002/chem.201702455](https://doi.org/10.1002/chem.201702455).
- (13) Cornes, S. P.; Sambrook, M. R.; Beer, P. D. *Chemical Communications* **2017**, *53*, 3866–3869,
DOI: [10.1039/C7CC01605K](https://doi.org/10.1039/C7CC01605K).
- (14) Vargas Jentzsch, A.; Hennig, A.; Mareda, J.; Matile, S. *Accounts of Chemical Research* **2013**, *46*, 2791–2800,
DOI: [10.1021/ar400014r](https://doi.org/10.1021/ar400014r).
- (15) Borissov, A.; Lim, J. Y. C.; Brown, A.; Christensen, K. E.; Thompson, A. L.; Smith, M. D.; Beer, P. D. *Chemical Communications* **2017**, *00*, 1–3,
DOI: [10.1039/x0xx00000x](https://doi.org/10.1039/x0xx00000x).
- (16) Murray, J. S.; Lane, P.; Clark, T.; Politzer, P. *Journal of Molecular Modeling* **2007**, *13*, 1033–1038,
DOI: [10.1007/s00894-007-0225-4](https://doi.org/10.1007/s00894-007-0225-4).

- (17) Fanfrlík, J.; Přáda, A.; Padělková, Z.; Pecina, A.; Macháček, J.; Lepšík, M.; Holub, J.; Růžička, A.; Hnyk, D.; Hobza, P. *Angewandte Chemie - International Edition* **2014**, *53*, 10139–10142,
DOI: [10.1002/anie.201405901](https://doi.org/10.1002/anie.201405901).
- (18) Garrett, G. E.; Carrera, E. I.; Seferos, D. S.; Taylor, M. S. *Chem. Commun.* **2016**, *52*, 9881–9884,
DOI: [10.1039/C6CC04818H](https://doi.org/10.1039/C6CC04818H).
- (19) Ho, P. C.; Szydlowski, P.; Sinclair, J.; Elder, P. J. W.; Kübel, J.; Gendy, C.; Lee, L. M.; Jenkins, H.; Britten, J. F.; Morim, D. R.; Vargas-Baca, I. *Nature Communications* **2016**, *7*, 11299,
DOI: [10.1038/ncomms11299](https://doi.org/10.1038/ncomms11299).
- (20) Wonner, P.; Vogel, L.; Kniep, F.; Huber, S. M. *Chemistry - A European Journal* **2017**, *23*, 16972–16975,
DOI: [10.1002/chem.201704502](https://doi.org/10.1002/chem.201704502).
- (21) Mitchell, M. O. *Journal of Molecular Modeling* **2017**, *23*, 287,
DOI: [10.1007/s00894-017-3452-3](https://doi.org/10.1007/s00894-017-3452-3).
- (22) Benz, S.; López-Andarias, J.; Mareda, J.; Sakai, N.; Matile, S. *Angewandte Chemie International Edition* **2017**, *56*, 812–815,
DOI: [10.1002/anie.201611019](https://doi.org/10.1002/anie.201611019).
- (23) Biot, N.; Bonifazi, D. *Chemistry - A European Journal* **2018**, *24*, 5439–5443,
DOI: [10.1002/chem.201705428](https://doi.org/10.1002/chem.201705428).
- (24) Ho, P. C.; Rafique, J.; Lee, J.; Lee, L. M.; Jenkins, H. A.; Britten, J. F.; Braga, A. L.; Vargas-Baca, I. *Dalton Trans.* **2017**, *46*, 6570–6579,
DOI: [10.1039/C7DT00612H](https://doi.org/10.1039/C7DT00612H).
- (25) Blackstock, S. C.; Lorand, J. P.; Kochi, J. K. *Journal of Organic Chemistry* **1987**, *52*, 1451–1460,
DOI: [10.1021/jo00384a013](https://doi.org/10.1021/jo00384a013).

- (26) Yan, X. Q.; Zhao, X. R.; Wang, H.; Jin, W. J. *The Journal of Physical Chemistry B* **2014**, *118*, 1080–1087,
DOI: [10.1021/jp4097869](https://doi.org/10.1021/jp4097869).
- (27) Murray, J. S.; Lane, P.; Politzer, P. *International Journal of Quantum Chemistry* **2008**, *108*, 2770–2781,
DOI: [10.1002/qua.21753](https://doi.org/10.1002/qua.21753).
- (28) Murray, J. S.; Lane, P.; Politzer, P. *Journal of Molecular Modeling* **2009**, *15*, 723–729,
DOI: [10.1007/s00894-008-0386-9](https://doi.org/10.1007/s00894-008-0386-9).
- (29) Pascoe, D. J.; Ling, K. B.; Cockroft, S. L. *Journal of the American Chemical Society* **2017**, *139*, 15160–15167,
DOI: [10.1021/jacs.7b08511](https://doi.org/10.1021/jacs.7b08511).
- (30) Dimitrijević, E.; Kvak, O.; Taylor, M. S. *Chemical Communications* **2010**, *46*, 9025–9027,
DOI: [10.1039/c0cc03347b](https://doi.org/10.1039/c0cc03347b).
- (31) Garrett, G. E.; Gibson, G. L.; Straus, R. N.; Seferos, D. S.; Taylor, M. S. *Journal of the American Chemical Society* **2015**, *137*, 4126–4133,
DOI: [10.1021/ja512183e](https://doi.org/10.1021/ja512183e).
- (32) Thomas, S. P.; Satheeshkumar, K.; Mugesh, G.; Guruow, T. N. *Chemistry - A European Journal* **2015**, *21*, 6793–6800,
DOI: [10.1002/chem.201405998](https://doi.org/10.1002/chem.201405998).
- (33) Bruckmann, A.; Pena, M. A.; Bolm, C. *Synlett* **2008**, 900–902,
DOI: [10.1055/s-2008-1042935](https://doi.org/10.1055/s-2008-1042935).
- (34) Kniep, F.; Jungbauer, S. H.; Zhang, Q.; Walter, S. M.; Schindler, S.; Schnapperelle, I.; Herdtweck, E.; Huber, S. M. *Angewandte Chemie - International Edition* **2013**, *52*, 7028–7032,
DOI: [10.1002/anie.201301351](https://doi.org/10.1002/anie.201301351).

- (35) Reisman, S. E.; Doyle, A. G.; Jacobsen, E. N. *Journal of the American Chemical Society* **2008**, *130*, 7198–7199,
DOI: [10.1021/ja801514m](https://doi.org/10.1021/ja801514m).
- (36) Benz, S.; Mareda, J.; Besnard, C.; Sakai, N.; Matile, S. *Chemical Science* **2017**,
8, 8164–8169,
DOI: [10.1039/C7SC03866F](https://doi.org/10.1039/C7SC03866F).
- (37) Wonner, P.; Vogel, L.; Düser, M.; Gomes, L.; Kniep, F.; Mallick, B.; Werz, D. B.; Huber, S. M. *Angewandte Chemie International Edition* **2017**, *56*, 12009–12012,
DOI: [10.1002/anie.201704816](https://doi.org/10.1002/anie.201704816).
- (38) Cundy, K.; Crooks, P. *Journal of Chromatography A* **1983**, *281*, 17–33,
DOI: [10.1016/S0021-9673\(01\)87863-8](https://doi.org/10.1016/S0021-9673(01)87863-8).
- (39) Iwaoka, M.; Takemoto, S.; Okada, M.; Tomoda, S. *Chemistry Letters* **2001**, 132–133,
DOI: [10.1246/cl.2001.132](https://doi.org/10.1246/cl.2001.132).
- (40) Iwaoka, M.; Isozumi, N. *Molecules* **2012**, *17*, 7266–7283,
DOI: [10.3390/molecules17067266](https://doi.org/10.3390/molecules17067266).
- (41) Iwaoka, M. In *Noncovalent Forces*, Scheiner, S., Ed.; Springer International Publishing: Cham, 2015, pp 265–289,
DOI: [10.1007/978-3-319-14163-3_9](https://doi.org/10.1007/978-3-319-14163-3_9).
- (42) Lange, A.; Günther, M.; Büttner, F. M.; Zimmermann, M. O.; Heidrich, J.; Hennig, S.; Zahn, S.; Schall, C.; Sievers-Engler, A.; Ansideri, F.; Koch, P.; Laemmerhofer, M.; Stehle, T.; Laufer, S. A.; Boeckler, F. M. *Journal of the American Chemical Society* **2015**, *137*, 14640–14652,
DOI: [10.1021/jacs.5b07090](https://doi.org/10.1021/jacs.5b07090).

- (43) Giroud, M.; Ivkovic, J.; Martignoni, M.; Fleuti, M.; Trapp, N.; Haap, W.; Kuglstatter, A.; Benz, J.; Kuhn, B.; Schirmeister, T.; Diederich, F. *ChemMedChem* **2017**, *12*, 257–270,
DOI: [10.1002/cmdc.201600563](https://doi.org/10.1002/cmdc.201600563).
- (44) Beno, B. R.; Yeung, K. S.; Bartberger, M. D.; Pennington, L. D.; Meanwell, N. A. *Journal of Medicinal Chemistry* **2015**, *58*, 4383–4438,
DOI: [10.1021/jm501853m](https://doi.org/10.1021/jm501853m).
- (45) Voth, A. R.; Hays, F. A.; Ho, P. S. *Proceedings of the National Academy of Sciences* **2007**, *104*, 6188–6193,
DOI: [10.1073/pnas.0610531104](https://doi.org/10.1073/pnas.0610531104).
- (46) Kolář, M. H.; Tabarrini, O. *Journal of Medicinal Chemistry* **2017**, *60*, 8681–8690,
DOI: [10.1021/acs.jmedchem.7b00329](https://doi.org/10.1021/acs.jmedchem.7b00329).
- (47) Parker, A. J.; Stewart, J.; Donald, K. J.; Parish, C. A. *Journal of the American Chemical Society* **2012**, *134*, 5165–5172,
DOI: [10.1021/ja2105027](https://doi.org/10.1021/ja2105027).
- (48) Sharma, K. D.; Kathuria, P.; Wetmore, S. D.; Sharma, P. *Physical Chemistry Chemical Physics* **2020**, *22*, 23754–23765,
DOI: [10.1039/d0cp04921b](https://doi.org/10.1039/d0cp04921b).
- (49) Farrell, K. M.; Brister, M. M.; Pittelkow, M.; Sølling, T. I.; Crespo-Hernández, C. E. *Journal of the American Chemical Society* **2018**, *140*, 11214–11218,
DOI: [10.1021/jacs.8b07665](https://doi.org/10.1021/jacs.8b07665).
- (50) Mai, S.; Wolf, A. P.; González, L. *Journal of Chemical Theory and Computation* **2019**, *15*, 3730–3742,
DOI: [10.1021/acs.jctc.9b00208](https://doi.org/10.1021/acs.jctc.9b00208).
- (51) Peng, Q.; Zhu, Y. H.; Zhang, T. S.; Liu, X. Y.; Fang, W. H.; Cui, G. *Physical Chemistry Chemical Physics* **2020**, *22*, 12120–12128,
DOI: [10.1039/d0cp01369b](https://doi.org/10.1039/d0cp01369b).

- (52) Fang, Y. G.; Peng, Q.; Fang, Q.; Fang, W.; Cui, G. *ACS Omega* **2019**, *4*, 9769–9777, DOI: [10.1021/acsomega.9b01276](https://doi.org/10.1021/acsomega.9b01276).
- (53) Uleany, K. O.; Cercola, R.; Nikolova, M.; Matthews, E.; Wong, N. G.; Dessent, C. E. *Molecules* **2020**, *25*, 15–17, DOI: [10.3390/molecules25143157](https://doi.org/10.3390/molecules25143157).
- (54) Hassan, A. E.; Sheng, J.; Zhang, W.; Huang, Z. *Journal of the American Chemical Society* **2010**, *132*, 2120–2121, DOI: [10.1021/ja909330m](https://doi.org/10.1021/ja909330m).
- (55) Sun, H.; Sheng, J.; Hassan, A. E.; Jiang, S.; Gan, J.; Huang, Z. *Nucleic Acids Research* **2012**, *40*, 5171–5179, DOI: [10.1093/nar/gks010](https://doi.org/10.1093/nar/gks010).
- (56) Salon, J.; Sheng, J.; Jiang, J.; Chen, G.; Caton-Williams, J.; Huang, Z. *Journal of the American Chemical Society* **2007**, *129*, 4862–4863, DOI: [10.1021/ja0680919](https://doi.org/10.1021/ja0680919).
- (57) Conlon, P. F.; Eguaogie, O.; Wilson, J. J.; Sweet, J. S.; Steinhoegl, J.; Englert, K.; Hancox, O. G.; Law, C. J.; Allman, S. A.; Tucker, J. H.; Hall, J. P.; Vyle, J. S. *Chemical Science* **2019**, *10*, 10948–10957, DOI: [10.1039/c9sc04098f](https://doi.org/10.1039/c9sc04098f).
- (58) CCoil Molecular orbital diagram — Wikipedia, The Free Encyclopedia, <https://commons.wikimedia.org/w/index.php?curid=6083186>, [Online; accessed 27-May-2021, CC BY-SA 3.0,] 2021.
- (59) Lühmann, D.-S.; Weitenberg, C.; Sengstock, K. *Physical Review X* **2015**, *5*, 031016, DOI: [10.1103/PhysRevX.5.031016](https://doi.org/10.1103/PhysRevX.5.031016).
- (60) Reed, A. E.; Curtiss, L. A.; Weinhold, F. *Chemical Reviews* **1988**, *88*, 899–926, DOI: [10.1021/cr00088a005](https://doi.org/10.1021/cr00088a005).
- (61) Bader, R. F. *Chemical Reviews* **1991**, *91*, 893–928, DOI: [10.1021/cr00005a013](https://doi.org/10.1021/cr00005a013).

- (62) Stout, G.; Jensen, L., *X-Ray Structure Determination: A Practical Guide*; A Wiley-Interscience publication; Wiley: 1989.
- (63) Brown, P. J.; Fox, A. G.; Maslen, E. N.; O'Keefe, M. A.; Willis, B. T. M. In *International Tables for Crystallography*; International Union of Crystallography: Chester, England, 2006; Vol. C, pp 554–595,
DOI: [10.1107/97809553602060000600](https://doi.org/10.1107/97809553602060000600).
- (64) Cooper, R. I.; Thompson, A. L.; Watkin, D. J. *Journal of Applied Crystallography* **2010**, *43*, 1100–1107,
DOI: [10.1107/S0021889810025598](https://doi.org/10.1107/S0021889810025598).
- (65) Coppens, P. *Acta Crystallographica Section B Structural Crystallography and Crystal Chemistry* **1968**, *24*, 1272–1274,
DOI: [10.1107/s0567740868004115](https://doi.org/10.1107/s0567740868004115).
- (66) Bragg, W. H. *Proceedings of the Physical Society of London* **1920**, *33*, 304–311,
DOI: [10.1088/1478-7814/33/1/331](https://doi.org/10.1088/1478-7814/33/1/331).
- (67) Domagała, S.; Fournier, B.; Liebschner, D.; Guillot, B.; Jelsch, C. *Acta Crystallographica Section A: Foundations of Crystallography* **2012**, *68*, 337–351,
DOI: [10.1107/S0108767312008197](https://doi.org/10.1107/S0108767312008197).
- (68) Jayatilaka, D.; Dittrich, B. *Acta Crystallographica Section A Foundations of Crystallography* **2008**, *64*, 383–393,
DOI: [10.1107/S0108767308005709](https://doi.org/10.1107/S0108767308005709).
- (69) Weiss, R. J., *X-ray determination of electron distributions*; Elsevier North-Holland: 1962; Vol. 6.
- (70) Grimwood, D. J.; Bytheway, I.; Jayatilaka, D. *J. Comput. Chem.* **2003**, *24*, 470–483,
DOI: [10.1002/jcc.10238](https://doi.org/10.1002/jcc.10238).
- (71) Woińska, M.; Jayatilaka, D.; Dittrich, B.; Flaig, R.; Luger, P.; Woźniak, K.; Dominiak, P. M.; Grabowsky, S. *ChemPhysChem* **2017**, *18*, 3334–3351,
DOI: [10.1002/cphc.201700810](https://doi.org/10.1002/cphc.201700810).

- (72) Jayatilaka, D.; Grimwood, D. J. In *Lect. Notes Comput. Sci. (including Subser. Lect. Notes Artif. Intell. Lect. Notes Bioinformatics)*, 2003; Vol. 2660, pp 142–151,
DOI: [10.1007/3-540-44864-0_15](https://doi.org/10.1007/3-540-44864-0_15).
- (73) Hansen, N. K.; Coppens, P. *Acta Crystallographica Section A* **1978**, *34*, 909–921,
DOI: [10.1107/S0567739478001886](https://doi.org/10.1107/S0567739478001886).
- (74) Guillot, B.; Viry, L.; Guillot, R.; Lecomte, C.; Jelsch, C. *J. Appl. Crystallogr.* **2001**, *34*, 214–223,
DOI: [10.1107/S0021889801001753](https://doi.org/10.1107/S0021889801001753).
- (75) Volkov, A.; Macchi, P; Farrugia, L. J.; Gatti, C.; Mallinson, P.; Richter, T.; Koritsanszky, T. XD2006 - A Computer Program Package for Multipole Refinement, Topological Analysis of Charge Densities and Evaluation of Intermolecular Energies from Experimental and Theoretical Structure Factors, 2006.
- (76) Haeberlen, U. In *High Resolut. Nmr Solids Sel. Averaging*, Haeberlen, U. B. T., Ed.; Elsevier: 1976, pp 17–35,
DOI: [10.1016/B978-0-12-025561-0.50009-5](https://doi.org/10.1016/B978-0-12-025561-0.50009-5).
- (77) Facelli, J. C.; Grant, D. M.; Michl, J. *Accounts of Chemical Research* **1987**, *20*, 152–158,
DOI: [10.1021/ar00136a005](https://doi.org/10.1021/ar00136a005).
- (78) Herzfeld, J.; Berger, A. E. *The Journal of Chemical Physics* **1980**, *73*, 6021–6030,
DOI: [10.1063/1.440136](https://doi.org/10.1063/1.440136).

Chapter 2

Simulating chalcogen bonding using molecular mechanics: A pseudoatom approach to model ebselen.

This chapter was submitted to *chemRxiv*, 21 Mar 2020.¹

2.1 Abstract

The organoselenium compound ebselen has recently been investigated as a treatment for COVID-19, however efforts to model ebselen *in silico* have been hampered by the lack of an efficient and accurate method to assess its binding to biological macromolecules. We present here a Generalized Amber Force Field modification which incorporates classical parameters for the selenium atom in ebselen, as well as a positively charged pseudoatom to simulate the σ -hole, a quantum mechanical phenomenon that dominates the chemistry of ebselen. Our approach is justified using an energy decomposition analysis of a number of DFT optimized structures, which show that the σ -hole interaction is primarily electrostatic in origin. Finally,

our model is verified by conducting MD simulations on a number of simple complexes, as well as the clinically relevant enzyme SOD1, which is known to bind to ebselen.

2.2 Introduction

Ebselen (**1**) is a molecule that has piqued the interest of many medicinal chemists, in no small part due to its decidedly non-drug like appearance. First synthesized in 1924, its unusual properties went more or less uninvestigated for more than 50 years.² Interest in ebselen boomed in the early 1980s, and since then it has been the subject of several studies into its synthesis, biological properties, and metabolism.^{3–11} Its biological activity can be broadly attributed to its ability to neutralize reactive oxygen species (ROS), reducing the level of oxidative stress to which cells are subjected.¹² To this end, ebselen has been investigated for its neuroprotective, mood-stabilizing, anti-inflammatory, and anti-cancer properties.^{13–18} Recently it was identified as a compound of interest for the treatment of COVID-19, showing promising inhibition of the viral M^{pro} protease enzyme.¹⁹

The *in vivo* antioxidant ability of ebselen is believed to be mediated through a catalytic cycle analogous to that of glutathione peroxidase (a selenoenzyme).²⁰ The selenium-containing heterocycle is reductively opened to afford the free selenol, which is the active catalyst. This is rapidly oxidised by ROS to a selenenic acid, which is then reduced back to the selenol by glutathione (GSH) via a selenenyl sulfide (fig. 2.1). Its activity against a number of other targets appears to also be mediated through formation of a covalent complex via nucleophilic attack at the selenium. There is also evidence that ebselen interacts with targets non-covalently.¹⁹ These interactions may include association with aromatic or hydrophobic residues, or H-bonding through the carbonyl. Ebselen can also form non-covalent complexes

with Lewis bases through an electrophilic σ -hole on the selenium atom, similarly to electron-deficient sulfur-containing molecules.^{21–23}

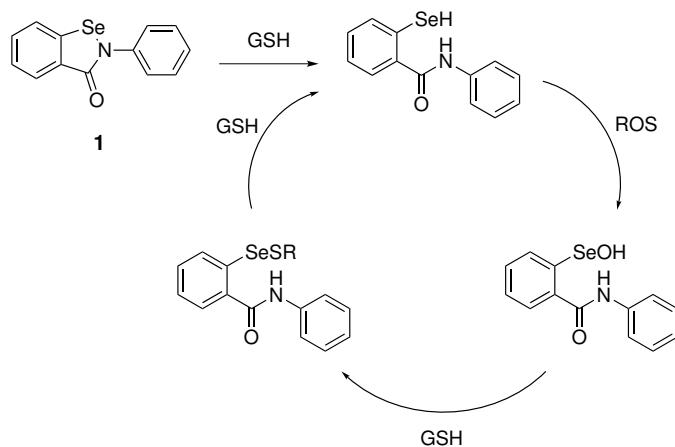


FIGURE 2.1: Catalytic cycle of ebselen **1** *in vivo*.

Molecular modelling is a vital tool in drug development, allowing for rapid and broad-reaching screening of drug candidates against likely substrates at minimal cost and risk. *Ab initio* quantum methods (QM) are widely used to model small molecules (generally smaller than a few hundred atoms), where their accuracy and ability to describe quantum effects underlying photochemical properties and bond breaking/formation processes are critical. They are however, computationally costly. Molecular mechanics (MM), where systems are treated strictly classically, is a viable alternative for large systems. The drawbacks are that MM relies on having an extensive parameter set (a “force field”) to describe the system (which is not necessarily available, or applicable to the system at hand), and that descriptions of quantum effects often fail spectacularly.

Both of these issues are encountered when attempting to model ebselen using MM. Firstly, parameters to describe selenium-containing small molecules are simply not available in most popular force fields, including GAFF, Gromos, and CGenFF. This has been substantially addressed in the work of Torsello et al., where extensive parametrization of a series of diaryl diselenides and diaryl ditellurides was

performed, and they provided a methodology to extend this to general chalcogen-containing molecules.²⁴ They did not, however, address the second issue of quantum effects, which is reasonable given that they do not play a large role in diselenides. The chemistry of ebselen, on the other hand, is dominated by the σ -hole, which is a quantum effect.²¹ The σ -hole is a region of positive electrostatic potential situated opposite to the Se–N bond, caused by a strongly anisotropic electron distribution around the selenium atom. This causes the selenium to adopt a highly directional electrophilic character, which can lead to the formation of “chalcogen-bonds” with electron pair donors (named by analogy to the ubiquitous hydrogen bond).²⁵

The presence of a σ -hole is not a new problem in MM, nor are they exclusive to chalcogens, as they are also found on the heavier halogens, where they give rise to halogen bonding.²⁶ Perhaps due to the higher prevalence of halogens in drug-like molecules, a number of approaches have been proposed to account for σ -holes in halogenated molecules. The common theme in these methods is the inclusion of a pseudoatom with positive electrostatic potential attached to the halogen atom. This pseudoatom is variously called an extra point (EP), explicit σ -hole (ESH), or virtual or off-atom centred point, and the approaches differ in the location of the pseudoatom and method used to derive its charge.^{27–30} Lone pairs can also be described using negatively charged pseudoatoms, and this approach has been used for some time.^{30–32}

It is worth noting an alternative approach, used by Cozzolino and Vargas-Baca, which treats secondary bonding interactions as true bonds, with an explicitly parametrized potential.³³ This was used to parametrize the supramolecular synthon 1,2,5-telluradizazole, which is known to self assemble into a range of interesting structures. This approach, however, relies on describing the bond using an *anharmonic* potential, which is not easily implemented in most software.

The inability to model ebselen in biological systems is a major hurdle in understanding the mechanism of its action. Furthermore, recent high-profile work has

highlighted the need for an accurate force field which takes quantum effects into account.^{34,35} In this work we develop a parameter set for the selenium atom in ebselen, including a pseudoatom to simulate the σ -hole. We have based the force field on GAFF, due to its popularity and ability to describe most drug like molecules in a way which is compatible with biomolecular force fields. Although GAFF is designed to work in the AMBER molecular dynamics system, we used GROMACS to develop the parameters, as it natively supports massless pseudoatoms which were critical for the description of the σ -hole. In principle, the work could be translated to any program that supports massless points, or the pseudoatom could even be given a mass and associated harmonic parameters which would enable some level of polarisability. However we have simply provided the GROMACS parameters with the massless point for its numerical stability and speed. Using these parameters, we show that this model accurately reproduces experimental geometries and energies, and compares favourably to *ab initio* calculations. This force field will prove useful in understanding the interactions between ebselen and current targets, and possibly lead to the discovery of new targets.

2.3 Results and Discussion

We began by deriving the classical bonding parameters involving selenium in ebselen, using the procedure of Torsello.²⁴ All quantum calculations were performed using Gaussian16, unless otherwise specified.³⁶ Electrostatic potentials were calculated using the cubegen program in the Gaussian suite, or mol2cub.³⁷ The ground state geometry of ebselen was optimized at the ω B97M-V/def2-TZVP level, followed by vibrational analysis to confirm the structure was minimized.^{38–40} The range-separated hybrid functional ω B97M-V has been shown to accurately describe Ch-bonding interactions at modest computational cost.⁴¹ Partial charges were assigned to the atoms using the RESP scheme, at the HF/6-31G* level.⁴² This was chosen for consistency with existing AMBER force fields.

2.3.1 Classical bonding parameters

Bond and angle force constants were derived by conducting a relaxed potential energy surface scan over a range of $\pm 0.3 \text{ \AA}$ for bonds and $\pm 10^\circ$ for angles. The resulting data was truncated to within 5 kcal/mol of the equilibrium energy (at larger distances the surfaces were appreciably anharmonic), and this surface was fitted with a classical harmonic oscillator model (eq. (2.1)) using the `nls` function in the R software package.⁴³ The equilibrium distance/angle x_0 was fixed to the value from the optimized geometry. The resulting potential energy surfaces and harmonic approximations are shown in fig. 2.2. Torsion angles were similarly scanned at the DFT and MM (with the torsion term set to zero) levels, and the difference between these surfaces was fitted using a periodic series (eq. (2.2)). The resulting parameters are presented in tables 2.1 and 2.2.

$$V(x) = \frac{1}{2}k(x - x_0)^2 \quad (2.1)$$

$$V(\phi) = \sum_{n=1} \left(\frac{V_{\max,n}}{2} \times (1 + \cos(n\phi + \gamma_n)) \right) \quad (2.2)$$

TABLE 2.1: Classical bonding parameters for ebselein. Bond lengths are given in \AA , and angles in degrees. Force constants for bonds and angles are given in kcal/mol $\cdot\text{\AA}^2$ and kcal/mol $\cdot\text{rad}^2$ respectively.

Parameter	x_0	k
r(Se–N)	1.8586	434.67
r(Se–C)	1.8829	422.33
$\angle(\text{C–Se–N})$	86.6	610.7
$\angle(\text{Se–N–C}_{\text{ar}})$	119.6	182.7
$\angle(\text{Se–N–C}_{\text{CO}})$	115.8	404.5
$\angle(\text{C–C–Se})$	119.4	329.2

Values of 2.12 and 0.2910 for the Lennard-Jones parameters σ and ε were used for selenium, to account for the polar flattening that is observed in strongly polarised

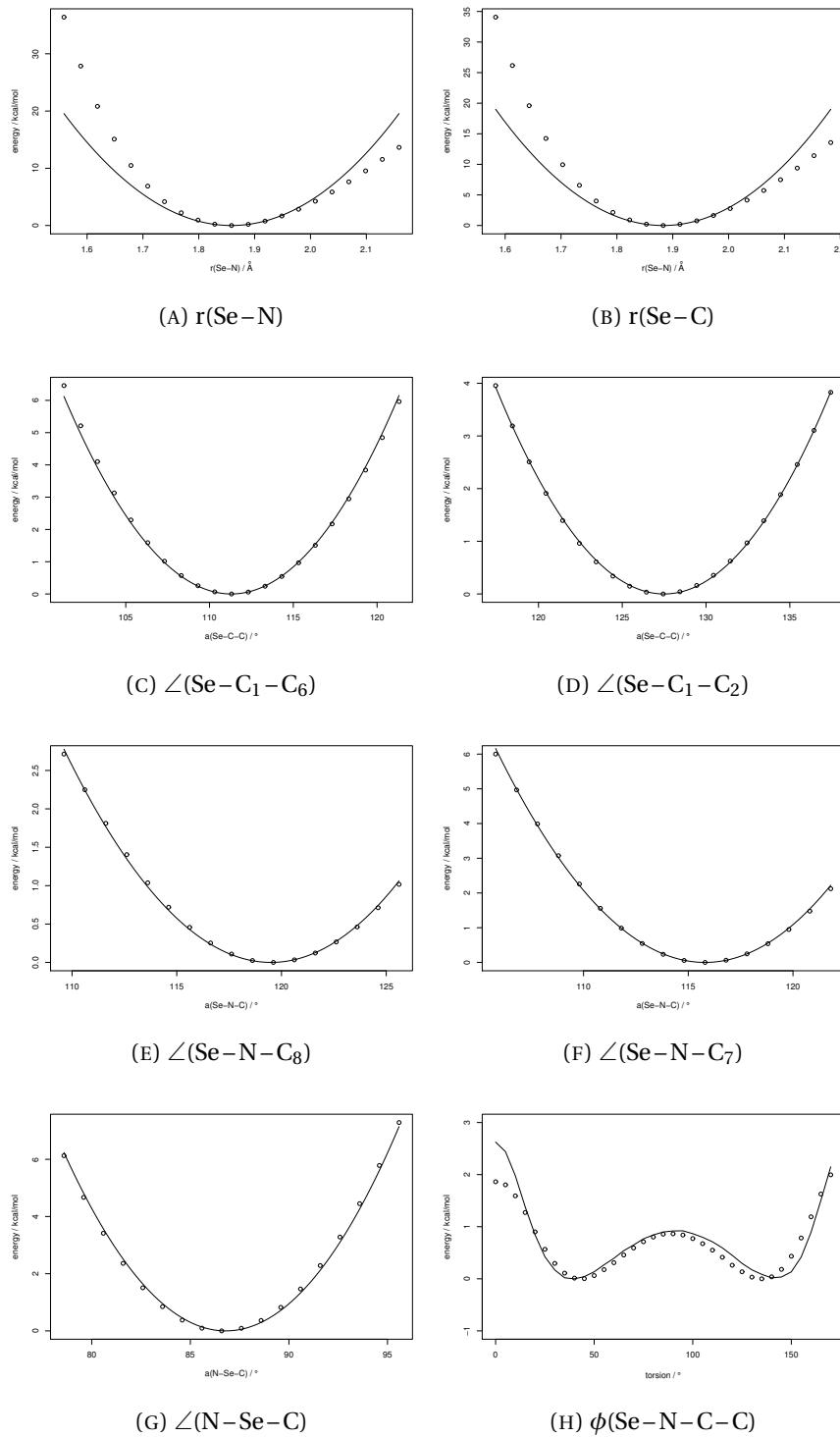


FIGURE 2.2: Potential energy surfaces for the indicated geometric parameter. The DFT surface is shown as points and the harmonic approximation is shown as a line.

TABLE 2.2: Dihedral parameters for ebselen.

Parameter	$V_{\max,1}$ kcal/mol	$V_{\max,2}$ kcal/mol	γ_1 °	γ_2 °
$\phi(\text{C}_{\text{ar}}-\text{C}_{\text{ar}}-\text{N}-\text{Se})$	-0.9653	0.5108	180	180

atoms. These values are lower than would be expected for a large atom like selenium (e.g. sulfur's parameters are only slightly lower at 1.9825 and 0.2824), however we found that they gave the most realistic bond energies and geometries. This is likely due to errors in the force field which are cancelled out by these artificially low values, which we believe is acceptable seeing as our goal is to provide an internally consistent system rather than absolute truth. The default GAFF Lennard-Jones parameters for the carbonyl oxygen were found to give an unreasonably high barrier to rotation about the central dihedral angle (due to steric repulsion between the oxygen and the aryl hydrogen), so they were changed to 1.5 and 0.08. This did not appear to have any negative effect on the rest of the model.

Default GAFF values were used for all other atoms, and Lorentz/Berthelot mixing rules were used to derive cross-terms.

2.3.2 Energy decomposition analysis

While attempting to model the σ -hole using molecular mechanics, we must remember that we are forcing a classical treatment onto an inherently quantum phenomenon. That said, some parts of the quantum phenomenon are easier than others to treat classically. There are thought to be three attractive energetic components which contribute to a σ -hole interaction. Namely, electrostatics, induction, and dispersion.^{44–46} The magnitudes of each component of σ -hole interactions has been the subject of heated debate in recent years. For many applications, these disagreements are fairly philosophical and of little consequence, however this is not the case when attempting to model σ -hole interactions using MM.

The electrostatic component generally refers to the interaction between two static (not distorted by each other) electric fields, which can be graphically represented by visualizing the electrostatic potential surfaces of the donor and acceptor moieties (fig. 2.5). This is already treated in MM (for the case of atom centred charges) as a sum of pairwise interactions. The accuracy of this component is only limited by the resolution of the electrostatic potential; it would appear that a pseudoatom approach could thus adequately describe the σ -hole. Dispersion is accounted for empirically within the r^{-6} term of the Lennard-Jones potential.

Issues arise when attempting to model the induction component of the σ -hole E_{ind} . This component refers to the redistribution of charge within (polarization) or between (charge-transfer) the donor and acceptor as they approach each other. Movement of charge is simply not accounted for within the most common AMBER force fields. This presents a large problem, as charge-transfer drives the strong directionality of σ -hole interactions, and may account for a significant proportion of their strength.

To ensure that this is not an insurmountable problem for this parametrization, we conducted energy decomposition analyses (EDA) on a variety of complexes containing ebselen. There are numerous EDA schemes available such as KM-EDA, NEDA, and ALMO, however we chose to use symmetry-adapted perturbation theory (SAPT).^{47,48} In contrast to several other schemes, SAPT explicitly includes dispersion (as opposed to adding it as an empirical correction), and contains no physically meaningless “catch-all” energy term. The total interaction energy E_{tot} is decomposed into an electrostatic component E_{elst} , an inductive component E_{ind} (this incorporates polarization and charge transfer, as they are not distinct phenomena within the SAPT framework), and a dispersive component E_{dis} . These attractive forces are balanced by a repulsive exchange component E_{exch} .

Four Lewis bases were chosen which are representative of those likely to be encountered in biological systems, and which span a wide range of basicity. Their structures

are given in fig. 2.3. SAPT(DFT) analyses were conducted using the Psi4 software package on geometries optimized at the ω B97M-V/def2-TZVP level, and the results are shown in fig. 2.4.⁴⁹

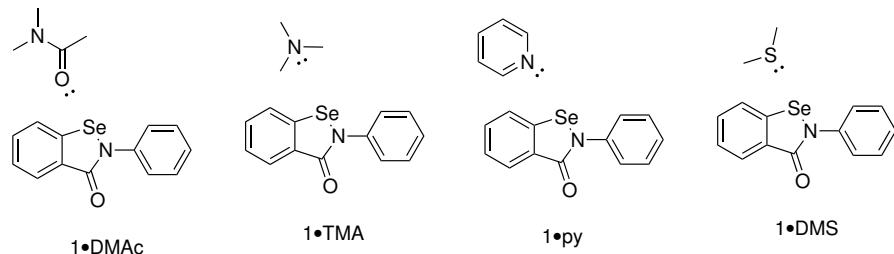


FIGURE 2.3: Structures of complexes used for SAPT(DFT) analysis.

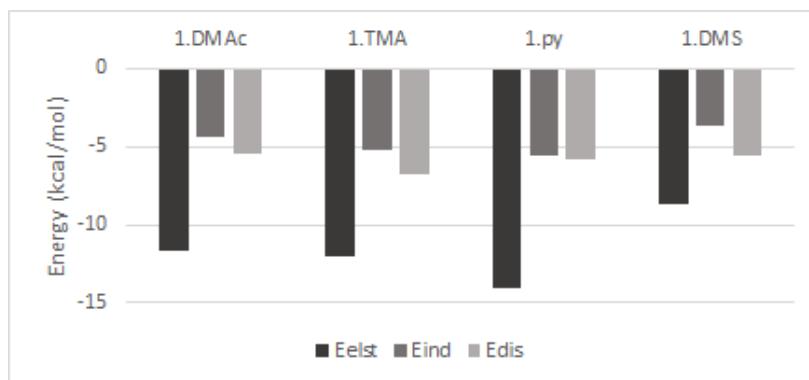


FIGURE 2.4: SAPT(DFT) analysis of complexes with four Lewis bases. All energies are given in kcal/mol.

The SAPT results indicate that the majority (around 80%) of the interaction can be described by electrostatics and dispersion. This suggests that the explicit σ -hole parametrization will be reliable, as electrostatics and dispersion are well described by MM.

2.3.3 Incorporation of pseudoatom

With classical parameters for ebselen in hand, as well as theoretical assurance that the system can be adequately described using electrostatics, we began to optimize parameters for the pseudoatom representing the σ -hole. The pseudoatom was modelled as a virtual site riding on the selenium atom along the extension of the

Se–N bond. In GROMACS this is assigned the type 2fd, which is described by only one parameter (the distance along the extension of the defining bond). This is very computationally efficient, although it is an approximation to reality, as the center of the σ -hole is actually slightly offset (see fig. 2.5). A more accurate description would be the three-center 3fd or 3fad type, however this would introduce a performance penalty.

The distance parameter for the pseudoatom was set to 1.189 Å which places the point charge on the VdW surface of the selenium atom. A number of approaches have been suggested to determine this distance, however it is important for numerical stability of the simulation that the charge lies on or within the VdW surface.²⁹ The charge of the pseudoatom was calculated using the RESP procedure, which fits a pre-calculated electrostatic potential to atom- (or pseudoatom-) centred point charges while enforcing symmetry restraints. The ESP was calculated at the HF/6-31G* level using Gaussian16, in order to be consistent with existing force field parameters. The pseudoatom was then introduced manually, and RESP applied using the ANTECHAMBER program. For comparison, we also calculated charges for the structure *without* a pseudoatom. Relevant charges are presented in table 2.3.

TABLE 2.3: Selected atomic charges for the pseudoatom and no pseudoatom models.

Atom	RESP charge (pseudoatom)	RESP charge (no pseudoatom)
E ₂₆	0.281382	—
Se ₁	−0.372631	0.056728
N ₂	−0.241674	−0.599430
C ₃	0.463064	0.827981
O ₄	−0.558076	−0.613468

2.3.4 Electrostatic potential map

With these parameters in hand, we were able to construct electrostatic potential maps (fig. 2.5), which show good qualitative agreement between the DFT and pseudoatom model potentials. Barely visible in the DFT ESP map is a second σ -hole, opposite the Se–C bond. Carbon is significantly less electronegative than nitrogen, so it doesn't polarize the selenium to the same degree, leading to a much smaller σ -hole. While it is conceivable that this σ -hole could form Ch-bonds as well, we have not observed any evidence of this in any of the derivatives we have studied.²² We therefore did not attempt to model it, although it could be modelled in the same way as the main σ -hole opposite the nitrogen.

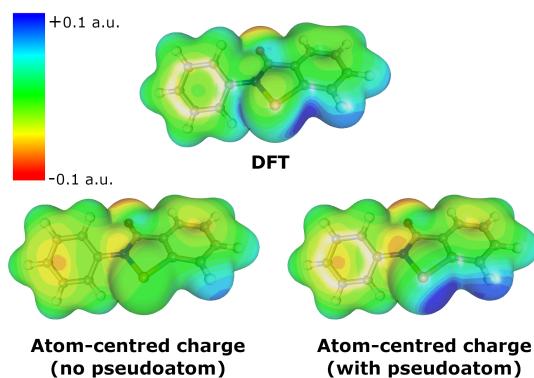


FIGURE 2.5: ESP mapped on the 0.005 a.u. electron density isosurface. The σ -hole is visible as the dark blue region on the DFT and atom-centred charge (with pseudoatom) surfaces.

2.3.5 Validation against DFT geometries

A preliminary verification of our model was conducted by comparing the geometries and energies calculated in the SAPT(DFT) analysis with the respective MM values. The Lewis bases chosen for the SAPT(DFT) analysis were constructed in AMBER. GAFF was used for all atoms, and an extra point was added to simulate the lone pair(s) of the Lewis bases per the method of Dixon and Kollman.³¹ Geometries were

assessed by minimizing the ebselen-Lewis base structure, and energies calculated relative to the unbound minimized monomers. The results are shown in table 2.4.

TABLE 2.4: Median geometric parameters for complexes with **1**. DFT equilibrium values are given in brackets for comparison. DFT energies are derived from SAPT(DFT).

Complex	$r(\text{Se} \cdots \text{B})$ Å	$\angle(\text{N}-\text{Se} \cdots \text{B})$ °	$\angle(\text{lone pair})$ °	Energy (kcal/mol)
1 ·py	2.879 Å (2.775 Å)	174.9° (176.2°)	171.1° (165.7°)	-7.548 (-7.093)
1 ·DMAc	2.741 Å (2.786 Å)	174.7° (172.4°)	129.1° (117.9°)	-7.068 (-7.551)
1 ·TMA	2.928 Å (2.857 Å)	170.1° (177.4°)	107.2° (113.6°)	-5.447 (-6.627)
1 ·DMS	3.253 Å (3.265 Å)	160.4° (177.4°)	81.2° (89.8°)	-2.510 (-5.646)

These results show that Ch-bonds can be adequately described by the inclusion of a positively charged pseudoatom. Almost all parameters and energies can be adequately reproduced, with the only anomaly being the substantially underestimated bond energy for the **1**·DMS complex. Interestingly, the geometry is modelled well in spite of this. This may be due to the increased role of dispersion in this complex, which has been deliberately de-emphasised in our parameters in order to emulate the polar flattening of the selenium. There is also a substantial charge-transfer component to this interaction which, when taken to completion, leads to formation of a Se–S bond and opening of the ring (fig. 2.1). This cannot be modelled in a harmonic force field, so it is perhaps not surprising that the interaction energy is underestimated. Nonetheless, we believe that the parameter set will still be very useful for molecular docking.

2.3.6 Validation against experimental density

We also sought to validate our model against the experimentally determined density of the crystal. An ebselen crystal (CSD code **SENGOH1**, $5 \times 5 \times 5$ unit cells, 1000 molecules) was constructed, and placed in a simulation box of the appropriate size at 1 atm. The crystal was heated to 290 K over 2 ns using a Berendsen thermostat and

barostat, and the density was calculated to be 1.548 g/cm^3 , which compares well to the experimental density of 1.529 g/cm^3 at the same temperature.⁵⁰ This procedure was repeated using the non-pseudoatom model, which afforded a density at 290 K of 1.468 g/cm^3 . The pseudoatom clearly represents an improvement when simulating the solid state structure of ebselen.

2.3.7 Validation against SOD1 binding

In order to show the utility of our model in a biological context, we conducted a binding simulation with a known ebselen target. Superoxide dismutase-1 (SOD1) forms a covalent complex with ebselen through the Cys111 residue, which appears to support correct folding of the protein, inhibiting aggregation and associated toxicity.⁵¹ Although formation of the covalent complex cannot be simulated using our model (as this is a bond-forming process), we are able to visualize the stabilized encounter complex which undergoes ring opening to form the final adduct. Indeed, the Ch-bond formed through the σ -hole can be thought of as the early stages of a nucleophilic attack at the selenium.²¹ SOD1 (PDB **2C9V**) was chosen because of the availability of an atomic resolution structure, demonstrated evidence of ebselen binding, and it's relatively small size.^{51,52} The structure was prepared by removing disorder, then removing water and ions (the Cu and Zn ions were retained). The ff14SB force field was used for the protein. The ebselen residue was introduced within the binding groove approximately halfway between the two units. The complex was then neutralized by addition of four Na^+ ions at the sites of most negative electrostatic potential, and solvated with a TIP3P explicit water model to give a final box size of $77.095 \times 96.253 \times 78.411 \text{ \AA}$. The structure was minimized over 1000 cycles to remove bad contacts, then heated to 300 K over 200 ps. A simulation of 2 ns at 300 K was then performed to assess the average binding geometry, which was found to exhibit a bifurcated Ch-bond between the expected Cys112 sulfur and the adjacent Ile114 backbone carbonyl (fig. 2.6). A similar experiment was

performed *without* the σ -hole, which failed to bind in a reproducible geometry, with the ebselen molecule wandering through the groove. This is presumably driven by hydrophobic interactions, and the entropic cost of desolvation.

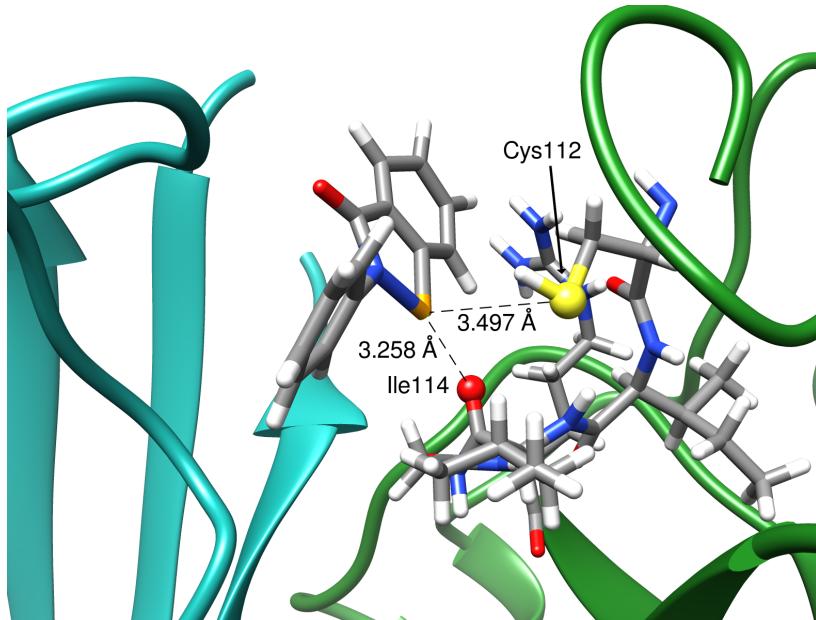


FIGURE 2.6: Average binding geometry of ebselen in the SOD1 groove.

2.4 Conclusion

In conclusion, we have developed a set of parameters which can greatly improve modelling of ebselen and its derivatives. Our model gives realistic geometries and energies of gas phase complexes, and reproduces the interaction between ebselen and a protein. Although this work is restricted to ebselen itself, the parameters will be generally applicable to derivatives of ebselen (with appropriate charge fitting). We hope that these results will be useful for the discovery of new targets.

References

- (1) Fellowes, T.; White, J. **2020**,
DOI: [10.26434/chemrxiv.12345434.v1](https://doi.org/10.26434/chemrxiv.12345434.v1).
- (2) Lesser, R.; Weiß, R. *Berichte der deutschen chemischen Gesellschaft (A and B Series)* **1924**, *57*, 1077–1082,
DOI: [10.1002/cber.19240570703](https://doi.org/10.1002/cber.19240570703).
- (3) Weber, R.; Renson, M. *Bulletin de la Societe Chimique de France* **1976**, *1124*–6.
- (4) Renson, M.; Etschenberg, E.; Winkelmann, J. 2-Phenyl-1,2-benzisoselenazol-3(2H)-one containing pharmaceutical preparations and process for the treatment of rheumatic diseases, 1981.
- (5) Müller, A.; Cadenas, E.; Graf, P.; Sies, H. *Biochemical Pharmacology* **1984**, *33*, 3235–3239,
DOI: [10.1016/0006-2952\(84\)90083-2](https://doi.org/10.1016/0006-2952(84)90083-2).
- (6) Wendel, A.; Fausel, M.; Safayhi, H.; Tiegs, G.; Otter, R. *Biochemical Pharmacology* **1984**, *33*, 3241–3245,
DOI: [10.1016/0006-2952\(84\)90084-4](https://doi.org/10.1016/0006-2952(84)90084-4).
- (7) Parnham, M. J.; Kindt, S. *Biochemical Pharmacology* **1984**, *33*, 3247–3250,
DOI: [10.1016/0006-2952\(84\)90085-6](https://doi.org/10.1016/0006-2952(84)90085-6).
- (8) Engman, L.; Hallberg, A. *Journal of Organic Chemistry* **1989**, *54*, 2966–2968,
DOI: [10.1017/CBO9781107415324.004](https://doi.org/10.1017/CBO9781107415324.004).
- (9) Schewe, T. *General Pharmacology* **1995**, *26*, 1153–1169,
DOI: [10.1016/0306-3623\(95\)00003-J](https://doi.org/10.1016/0306-3623(95)00003-J).
- (10) Balkrishna, S. J.; Bhakuni, B. S.; Chopra, D.; Kumar, S. *Organic Letters* **2010**, *12*, 5394–5397,
DOI: [10.1021/ol102027j](https://doi.org/10.1021/ol102027j).

- (11) Iwasaki, M.; Miki, N.; Tsuchiya, Y.; Nakajima, K.; Nishihara, Y. *Organic Letters* **2017**, *19*, 1092–1095,
DOI: [10.1021/acs.orglett.7b00116](https://doi.org/10.1021/acs.orglett.7b00116).
- (12) Mugesh, G.; Singh, H. B. *Chemical Society Reviews* **2000**, *29*, 347–357,
DOI: [10.1039/a908114c](https://doi.org/10.1039/a908114c).
- (13) Parnham, M. J.; Graf, E. *Biochemical Pharmacology* **1987**, *36*, 3095–3102,
DOI: [10.1016/0006-2952\(87\)90617-4](https://doi.org/10.1016/0006-2952(87)90617-4).
- (14) Kil, J.; Pierce, C.; Tran, H.; Gu, R.; Lynch, E. D. *Hearing Research* **2007**, *226*,
44–51,
DOI: [10.1016/j.heares.2006.08.006](https://doi.org/10.1016/j.heares.2006.08.006).
- (15) Singh, N.; Halliday, A. C.; Thomas, J. M.; Kuznetsova, O.; Baldwin, R.; Woon, E. C.; Aley, P. K.; Antoniadou, I.; Sharp, T.; Vasudevan, S. R.; Churchill, G. C. *Nature Communications* **2013**, *4*, 1332–1337,
DOI: [10.1038/ncomms2320](https://doi.org/10.1038/ncomms2320).
- (16) Azad, G. K.; Tomar, R. S. *Molecular Biology Reports* **2014**, *41*, 4865–4879,
DOI: [10.1007/s11033-014-3417-x](https://doi.org/10.1007/s11033-014-3417-x).
- (17) Parnham, M.; Sies, H. *Expert Opinion on Investigational Drugs* **2000**, *9*, 607–619,
DOI: [10.1517/13543784.9.3.607](https://doi.org/10.1517/13543784.9.3.607).
- (18) Chantadul, V.; Wright, G. S. A.; Amporndanai, K.; Shahid, M.; Antonyuk, S. V.; Washbourn, G.; Rogers, M.; Roberts, N.; Pye, M.; O'Neill, P. M.; Hasnain, S. S. *Communications Biology* **2020**, *3*, 97,
DOI: [10.1038/s42003-020-0826-3](https://doi.org/10.1038/s42003-020-0826-3).
- (19) Jin, Z. et al. *Nature* **2020**, *582*, 289–293,
DOI: [10.1038/s41586-020-2223-y](https://doi.org/10.1038/s41586-020-2223-y).
- (20) Antony, S.; Bayse, C. A. *Inorganic Chemistry* **2011**, *50*, 12075–12084,
DOI: [10.1021/ic201603v](https://doi.org/10.1021/ic201603v).

- (21) Thomas, S. P.; Satheeshkumar, K.; Mugesh, G.; Guruow, T. N. *Chemistry - A European Journal* **2015**, *21*, 6793–6800,
DOI: [10.1002/chem.201405998](https://doi.org/10.1002/chem.201405998).
- (22) Fellowes, T.; White, J. M. *CrystEngComm* **2019**, *21*, 1539–1542,
DOI: [10.1039/C8CE01853G](https://doi.org/10.1039/C8CE01853G).
- (23) Beno, B. R.; Yeung, K. S.; Barthberger, M. D.; Pennington, L. D.; Meanwell, N. A. *Journal of Medicinal Chemistry* **2015**, *58*, 4383–4438,
DOI: [10.1021/jm501853m](https://doi.org/10.1021/jm501853m).
- (24) Torsello, M.; Pimenta, A. C.; Wolters, L. P.; Moreira, I. S.; Orian, L.; Polimeno, A. *Journal of Physical Chemistry A* **2016**, *120*, 4389–4400,
DOI: [10.1021/acs.jpca.6b02250](https://doi.org/10.1021/acs.jpca.6b02250).
- (25) Murray, J. S.; Lane, P.; Politzer, P. *Journal of Molecular Modeling* **2009**, *15*, 723–729,
DOI: [10.1007/s00894-008-0386-9](https://doi.org/10.1007/s00894-008-0386-9).
- (26) Clark, T.; Hennemann, M.; Murray, J. S.; Politzer, P. *Journal of Molecular Modeling* **2007**, *13*, 291–296,
DOI: [10.1007/s00894-006-0130-2](https://doi.org/10.1007/s00894-006-0130-2).
- (27) Rendine, S.; Pieraccini, S.; Forni, A.; Sironi, M. *Physical Chemistry Chemical Physics* **2011**, *13*, 19508–19516,
DOI: [10.1039/c1cp22436k](https://doi.org/10.1039/c1cp22436k).
- (28) Ibrahim, M. A. A. *Journal of Computational Chemistry* **2011**, *32*, 2564–2574,
DOI: [10.1002/jcc.21836](https://doi.org/10.1002/jcc.21836).
- (29) Kolář, M.; Hobza, P. *Journal of Chemical Theory and Computation* **2012**, *8*, 1325–1333,
DOI: [10.1021/ct2008389](https://doi.org/10.1021/ct2008389).
- (30) Harder, E. et al. *Journal of Chemical Theory and Computation* **2016**, *12*, 281–296,
DOI: [10.1021/acs.jctc.5b00864](https://doi.org/10.1021/acs.jctc.5b00864).

- (31) Dixon, R. W.; Kollman, P. A. *Journal of Computational Chemistry* **1997**, *18*, 1632–1646,
DOI: [10.1002/\(SICI\)1096-987X\(199710\)18:13<1632::AID-JCC5>3.0.CO;2-S](https://doi.org/10.1002/(SICI)1096-987X(199710)18:13<1632::AID-JCC5>3.0.CO;2-S).
- (32) Cieplak, P.; Caldwell, J.; Kollman, P. *Journal of Computational Chemistry* **2001**, *22*, 1048–1057,
DOI: [10.1002/jcc.1065](https://doi.org/10.1002/jcc.1065).
- (33) Cozzolino, A. F.; Vargas-Baca, I. *Crystal Growth and Design* **2011**, *11*, 668–677,
DOI: [10.1021/cg100951y](https://doi.org/10.1021/cg100951y).
- (34) Weglarz-Tomczak, E.; Tomczak, J. M.; Talma, M.; Burda-Grabowska, M.; Giurg, M.; Brul, S. *Scientific Reports* **2021**, *11*, 3640,
DOI: [10.1038/s41598-021-83229-6](https://doi.org/10.1038/s41598-021-83229-6).
- (35) Menéndez, C. A.; Byléhn, F.; Perez-Lemus, G. R.; Alvarado, W.; de Pablo, J. J. *Science Advances* **2020**, *6*, eabd0345,
DOI: [10.1126/sciadv.abd0345](https://doi.org/10.1126/sciadv.abd0345).
- (36) Frisch, M. J. et al. Gaussian16 Revision C.01, 2016.
- (37) Fellowes, T. mol2cub <https://github.com/tjfellowes/mol2cub>.
- (38) Chai, J. D.; Head-Gordon, M. *Physical Chemistry Chemical Physics* **2008**, *10*, 6615–6620,
DOI: [10.1039/b810189b](https://doi.org/10.1039/b810189b).
- (39) Weigend, F.; Ahlrichs, R. *Physical Chemistry Chemical Physics* **2005**, *7*, 3297–3305,
DOI: [10.1039/b508541a](https://doi.org/10.1039/b508541a).
- (40) Weigend, F. *Physical Chemistry Chemical Physics* **2006**, *8*, 1057–1065,
DOI: [10.1039/b515623h](https://doi.org/10.1039/b515623h).
- (41) Mehta, N.; Fellowes, T.; White, J. M.; Goerigk, L. *Journal of Chemical Theory and Computation* **2021**, *17*, 2783–2806,
DOI: [10.1021/acs.jctc.1c00006](https://doi.org/10.1021/acs.jctc.1c00006).

- (42) Cornell, W. D.; Cieplak, P.; Bayly, C. I.; Kollman, P. A. *Journal of the American Chemical Society* **1993**, *115*, 9620–9631,
DOI: [10.1021/ja00074a030](https://doi.org/10.1021/ja00074a030).
- (43) R Core Team R: A Language and Environment for Statistical Computing, Vienna, Austria, 2019.
- (44) Bleiholder, C.; Werz, D. B.; Köppel, H.; Gleiter, R. *Journal of the American Chemical Society* **2006**, *128*, 2666–2674,
DOI: [10.1021/ja056827g](https://doi.org/10.1021/ja056827g).
- (45) Bleiholder, C.; Gleiter, R.; Werz, D. B.; Köppel, H. *Inorganic Chemistry* **2007**, *46*, 2249–2260,
DOI: [10.1021/ic062110y](https://doi.org/10.1021/ic062110y).
- (46) Pascoe, D. J.; Ling, K. B.; Cockroft, S. L. *Journal of the American Chemical Society* **2017**, *139*, 15160–15167,
DOI: [10.1021/jacs.7b08511](https://doi.org/10.1021/jacs.7b08511).
- (47) Bukowski, R. et al. SAPT2020: An Ab Initio Program for Many-Body Symmetry-Adapted Perturbation Theory Calculations of Intermolecular Interaction Energies, University of Delaware and University of Warsaw, 2020.
- (48) Jeziorski, B.; Moszynski, R.; Szalewicz, K. *Chemical Reviews* **1994**, *94*, 1887–1930,
DOI: [10.1021/cr00031a008](https://doi.org/10.1021/cr00031a008).
- (49) Parrish, R. M. et al. *Journal of Chemical Theory and Computation* **2017**, *13*, 3185–3197,
DOI: [10.1021/acs.jctc.7b00174](https://doi.org/10.1021/acs.jctc.7b00174).
- (50) Dupont, L.; Dideberg, O.; Jacquemin, P. *Acta Crystallographica Section C Crystal Structure Communications* **1990**, *46*, 484–486,
DOI: [10.1107/s0108270189007894](https://doi.org/10.1107/s0108270189007894).

- (51) Capper, M. J.; Wright, G. S.; Barbieri, L.; Luchinat, E.; Mercatelli, E.; McAlary, L.; Yerbury, J. J.; O'Neill, P. M.; Antonyuk, S. V.; Banci, L.; Hasnain, S. S. *Nature Communications* **2018**, *9*, 1–9,
DOI: [10.1038/s41467-018-04114-x](https://doi.org/10.1038/s41467-018-04114-x).
- (52) Strange, R. W.; Antonyuk, S. V.; Hough, M. A.; Doucette, P. A.; Valentine, J. S.; Hasnain, S. S. *Journal of Molecular Biology* **2006**, *356*, 1152–1162,
DOI: [10.1016/j.jmb.2005.11.081](https://doi.org/10.1016/j.jmb.2005.11.081).

Part II

The strength and nature of chalcogen bonding

Chapter 3

New insights into chalcogen bonding provided by co-crystal structures of benzisoselenazolinone derivatives and nitrogen bases.

This chapter was published in Cryst. Eng. Comm., 22 Jan 2019.^{1†}

3.1 Abstract

A number of derivatives of benzisoselenazolinones, including the drug ebselen, have been synthesized, and their interactions with various nitrogen bases characterized through x-ray crystallography. Structural studies revealed a strong interaction in all cases, with Se ··· N distances well within the Van der Waals radii of the constituent atoms. We suggest that there is a significant charge transfer component to this interaction, in contrast to some other interactions of similar strength and directionality.

[†]Compound numbering, section headings, and terminology have been updated to fit this thesis.

We have also found that this interaction can be enhanced via H-bonding to the carbonyl group of the benzisoselenazolinone moiety.

3.2 Introduction

Chalcogen bonding (Ch-bonding) is a class of non-covalent interaction which has recently piqued the interests of the chemical community, and potential applications in materials and medicinal chemistry are emerging.^{2–5} It bears similarities to the related concept of halogen bonding, and the ubiquitous phenomenon of hydrogen bonding, in that the result is a relatively strong and highly directional non-bonded interaction.⁶ This strength and directionality has been exploited in crystal engineering,^{7–9} anion recognition,^{10–12} and bond activation,^{13–15} and appears to play a critical role in protein folding.^{16,17} Studies on Te ··· N Ch-bonds in solution phase have shown they can be as strong as 2.7 kcal/mol.¹⁸ A number of interesting and potentially useful supramolecular polymers have been synthesised and characterised by Vargas-Baca et al., based on tellurium- and selenium-containing heterocycles.^{19,20}

In our efforts to apply the concept of Ch-bonding to biological systems, we turned to the benzisoselenazolinone scaffold of the antioxidant compound ebselen **1**. **1** has been known since the 1980s to effectively scavenge reactive oxygen species *in vivo*.²¹ It has remarkably low toxicity for an organoselenium compound, and is being investigated as a possible treatment for a number of conditions.^{22–25} It is also an ideal scaffold for Ch-bonding, bearing a selenium atom bonded to an electronegative amide nitrogen. Indeed, Ch-bonding in **1** has been investigated previously in the context of crystal packing of the pure compound, but there is a lack of experimental evidence for interactions with other acceptors.²⁶

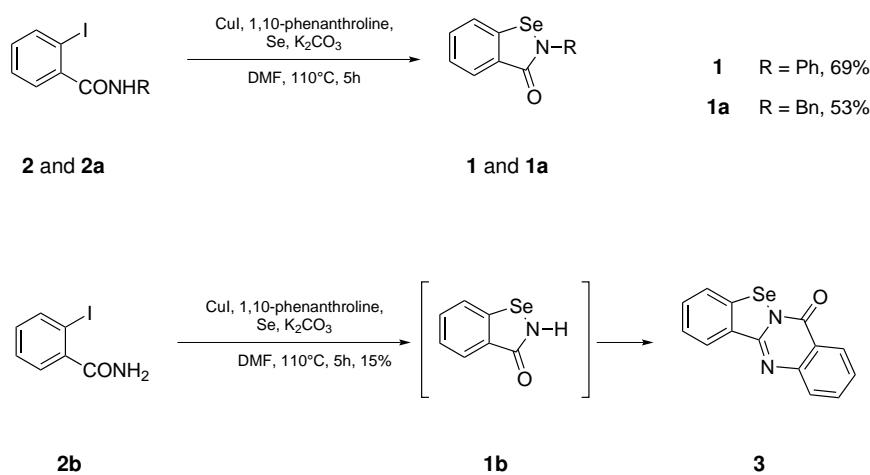
Numerous studies have examined the nature of the Ch-bond, in particular the balance between electrostatic effects (due to anisotropic electrostatic potential

around the chalcogen atom), covalent (orbital overlap and electron delocalisation), and dispersion forces.^{27–31} In the case of halogen bonding, these contributions are generally well characterized. Halogen bonds range from primarily electrostatic (as in the case of fluorinated iodobenzenes³²) to charge-transfer dominated (molecular halogens³³). In the case of Ch-bonding in derivatives of **1**, the contributions are less clear. We therefore sought to characterize Ch-bonding interactions between a number of derivatives of **1**, and a variety of Ch-bond acceptors.

3.3 Results and Discussion

3.3.1 Synthesis of benziselenazolinone derivatives **1**, **1a** and **3**

Compounds **1** and **1a** were synthesized via published procedures from amides **2** and **2a**.³⁴ The Se-tetracycle **3** was isolated in poor yield as the major product from the cyclization of primary amide **2b** in an attempt to form **1b**. It is noteworthy that the spectral characteristics of **3** are essentially identical to reported spectral data of **1b**.³⁴



SCHEME 3.1: Synthesis of Ch-bond donors **1**, **1a** and **3**.

TABLE 3.1: Selected structural parameters of Ch-bonded complexes.

Complex	r(N···Se) Å	r(Se–N ₁) Å	r(Se–C ₁) Å	r(N–C(O)) Å	∠(N···Se–N ₁) °	∠(C _{para} ···N···Se) °
1 only	—					
1 · DMAP	2.371(1)	1.9676(10)	1.8959(12)	1.2345(14)	174.18(4)	173.52(4)
1a only	—	1.8805(14)	1.8867(16)	1.350(2)	—	—
1a · DMAP M1	2.4276(14)	1.9297(13)	1.8984(15)	1.348(2)	173.93(5)	174.89(5)
1a · DMAP M2	2.4331(14)	1.9191(14)	1.8966(14)	1.349(2)	175.30(5)	158.14(5)
1a · DMAP · H ₂ O	2.4046(15)	1.9367(14)	1.9070(14)	1.330(2)	175.54(5)	167.43(5)
1a · quinuclidine	2.5874(17)	1.9077(17)	1.898(2)	1.354(3)	176.77(7)	161.32(7)
1a · DABCO	2.6166(15)	1.9019(14)	1.8967(17)	1.355(2)	175.76(6)	160.00(6)
3 only	—	1.883(2)	1.899(3)	1.393(3)	—	—
3 · pyridine	2.461(3)	1.926(2)	1.908(3)	1.373(4)	174.13(9)	169.1(1)
3 · DMAP	2.304(1)	1.9716(9)	1.918(1)	1.375(1)	173.81(4)	173.16(4)

3.3.2 Co-crystal structures of benzisoselenazolinones and Lewis bases

High quality low temperature crystal structures were obtained for the parent benzisoselenazolinone derivatives **1**,²⁶ **1a** and the Se-tetracycle **3** and the chalcogen-bonded co-crystals of these compounds with a variety of nitrogen bases, including pyridine, dimethylaminopyridine (DMAP), quinuclidine and 1,4-diazabicyclo[2.2.2]-octane (DABCO). Powder diffraction patterns were obtained of the bulk co-crystal material and compared with the single crystal data, with excellent agreement. This provides strong evidence of phase purity, with the exception of **1a**·DMAP, which indicated the presence of the unbound monomers in addition to the Ch-bonded adduct. Relevant structural parameters are presented in table 3.1, while all thermal ellipsoid plots are presented in the supplementary material (section 3.5.2). In the following discussion we begin by assessing the Ch-bond donor abilities of **1**, **1a**, and **3** by comparing the structural parameters with a common nitrogen base adduct (DMAP), followed by comparison of a single Ch-bond donor **3** with two different nitrogen bases with markedly different basicities (pyridine and DMAP).

3.3.3 Effects of the benziselenazolinone on Ch-bond strength

The DMAP adducts of **1**, **1a** and **3** are characterized by near linear N ··· Se–N(CO) angles (table 3.1) with N_{DMAP} ··· Se distances which are 2.4276(14) and 2.4331(16) Å for the two independent molecules of **1a**, 2.371(1) Å for **1**, and the strikingly short N ··· Se distance of 2.304(1) Å for **3**. All are well within the van der Waals radii of N and Se of 3.85 Å.³⁵ The antipodal Se–N₁ bond distance within these adducts is significantly lengthened compared to the free Ch-bond donors, 0.063 Å in **1**, 0.049 Å in **1a**, and 0.088 Å in **3**, with the degree of lengthening being related in an inverse sense to the N_{DMAP} ··· Se distance, in all cases the (non-antipodal) Se–C₁ bond is essentially unchanged. These structural parameters suggest an order of Ch-bond donor abilities **1a** < **1** < **3**, which is supported by theoretical calculations which are discussed below, as well as being consistent with the ⁷⁷Se NMR chemical shifts.

3.3.4 Endocyclic bond lengthening associated with stronger complexes

Further discussion is warranted on the **1a**·DMAP adduct. Firstly, the two independent molecules of **1a**·DMAP differ significantly with respect to the direction that the nitrogen base lone pair makes with the Se–N₁ bond. In molecule 1 this angle is close to collinear at 174.89(5)° while in molecule 2, this deviates significantly from linearity 158.14(5)°. Associated with this difference is a slightly longer N_{DMAP} ··· Se distance of 2.4331(14) Å in molecule 2 compared to 2.4276(14) Å in molecule 1 ($\Delta=0.0055$ Å; 3σ) and a shorter Se–N₁ distance 1.9191(14) Å vs 1.9297(14) Å ($\Delta=-0.10$ Å; 7.5σ) indicating a slightly weaker interaction. This can be seen in fig. 3.1.

The structural effects described, particularly the lengthening of the Se–N₁ bond are consistent with donation of electron density from the nitrogen lone pair into the Se–N₁ anti-bonding orbital being a significant component of these N ··· Se Ch-bonds, which has been described before.²⁸

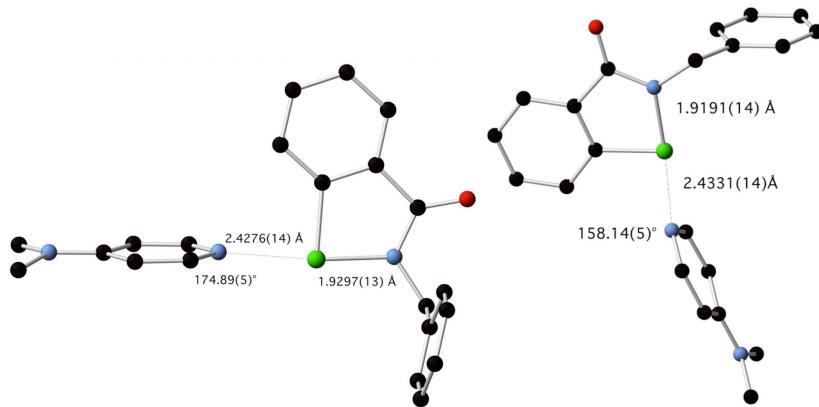


FIGURE 3.1: Structure of **1a**·DMAP, showing the two distinct geometries.

3.3.5 H-bond enhanced Ch-bonding

The second reason for further discussion of the **1a**·DMAP adduct is based on the structural parameters obtained for the hydrate structure **1a**·DMAP·H₂O which was serendipitously obtained by evaporation of a THF solution in an open flask. The structure is shown in fig. 3.2.

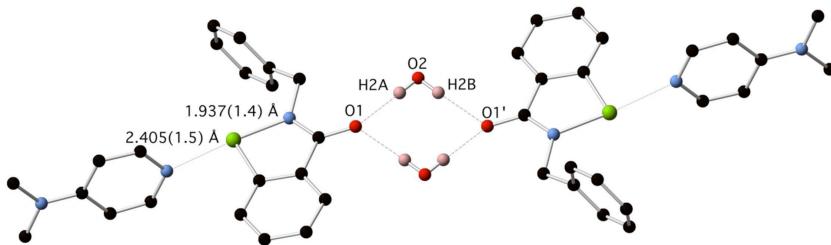


FIGURE 3.2: Structure of **1a**·DMAP·H₂O.

1a·DMAP·H₂O crystallizes as a centrosymmetric hydrogen-bonded dimer in which two water molecules bridge two molecules of **1a** across a crystallographic inversion centre. Of note, when comparison is made between the structural parameters for **1a**·DMAP molecule 1, which has a similar geometry about the N...Se moiety as for

1a·DMAP·H₂O, there is a significant contraction of the N_{DMAP} ··· Se distance from 2.4276(14) Å to 2.4046(14) Å ($\Delta=-0.023$ Å; 16 σ), and an increase in the Se–N₁ bond distance from 1.9297(13) to 1.9367(14) Å ($\Delta=0.007$ Å; 5 σ). We have coined the term ‘hydrogen-bond enhanced Ch-bonding’ to describe this interesting structural effect.

3.3.6 Effects of the Lewis base on Ch-bond strength

We were fortunate to obtain crystal structures of the Se-tetracycle **3** with both pyridine and DMAP, which gave us the opportunity to compare the structural effects arising from two Ch-bond acceptors with significantly different basicities. The pyridine adduct of **3** is characterized by a N_{PYR} ··· Se distance 2.461(3) Å and Se–N₁ distance of 1.926(2) Å and a near linear N_{PYR} ··· Se–N₁ angle, the Se–N₁ bond distance is significantly longer than the corresponding distance 1.883(2) Å in non-bound structure of **3**. The DMAP adduct of **3** is characterized by a significantly shorter N_{DMAP} ··· Se distance of 2.304(1) Å ($\Delta=-0.157$ Å) and longer Se–N₁ distance of 1.9716(9) Å ($\Delta=0.046$ Å), consistent with a significantly stronger interaction.

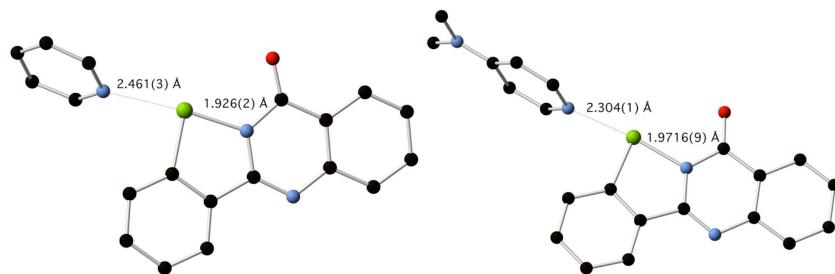


FIGURE 3.3: Pyridine and DMAP adducts of Se-tetracycle **3**.

Ch-bonded co-crystals of the benzisoselenazolinone derivative **1a** with the tertiary amines; quinuclidine and DABCO were obtained and structurally characterized. The adducts are presented in fig. 3.4. The N_{QUIN} ··· Se and N_{DABCO} ··· Se distances of 2.5874(17) Å and 2.6166(15) Å for **1a**·quinuclidine and **1a**·DABCO respectively are significantly longer than those observed for **1a**·DMAP suggesting an order of Ch-bond strengths with **1a** DABCO < Quinuclidine < DMAP which correlates well

with the hydrogen bond acceptor ability of these bases, as quantified by the pK_{HB} (table 3.2).

TABLE 3.2: Hydrogen bond basicities of bases studied.

	pyridine	DABCO	quinuclidine	DMAP
pK_{HB}	1.86 ³⁶	2.63 ³⁷	2.71 ³⁷	2.80 ³⁶
1a ·base $r(N \cdots Se) / \text{\AA}$	—	2.6166(15)	2.5874(17)	2.4276(14) ^a
1a ·base $\Delta H_f / \text{kcal/mol}$	-6.35	-7.82	—	-7.91

^aBond distance given is the shorter of the two coordination environments.

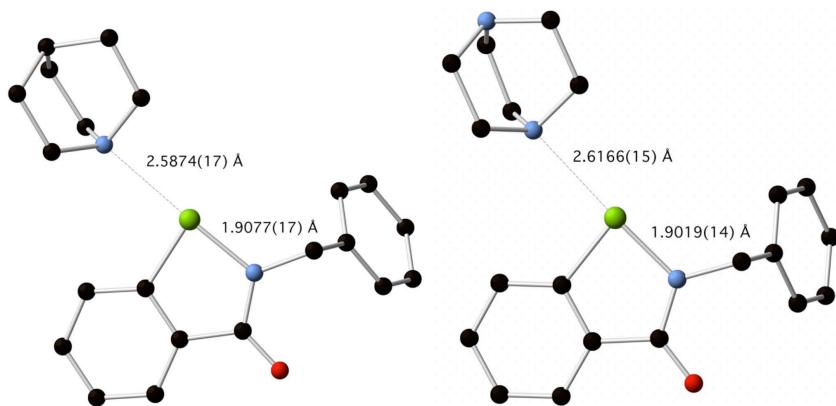


FIGURE 3.4: Adducts of benzisoselenazolinone **1a** with quinuclidine and DABCO.

3.3.7 DFT interaction energies and NBO analysis

Interaction energies for the complexes were calculated using the ω B97X-D dispersion corrected functional, which has been used to study similar systems with good agreement with coupled cluster methods.²⁷ All geometries were therefore optimized at ω B97X-D/def2-TZVP, and minima verified by frequency analysis.

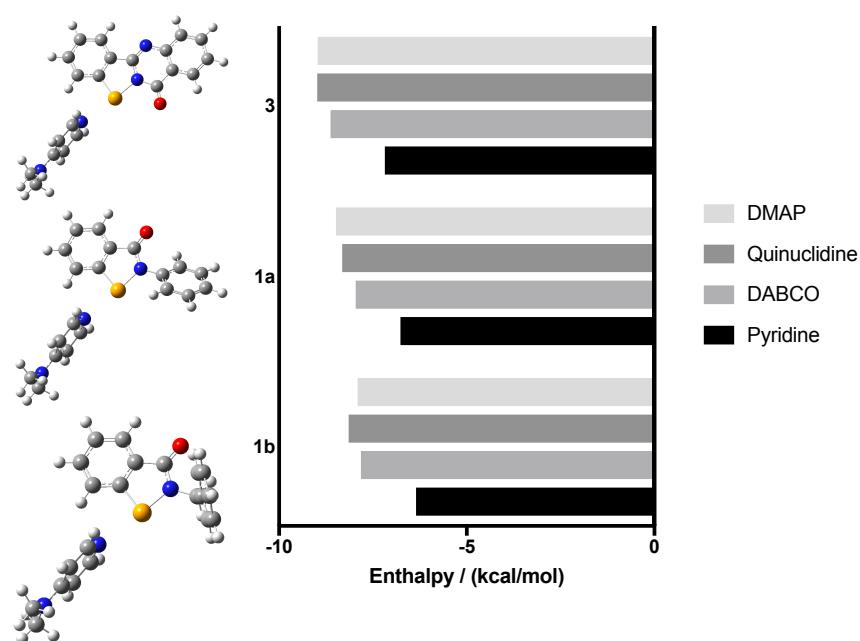


FIGURE 3.5: Interaction enthalpies calculated at ω B97X-D/def2-TZVP. Optimised geometries for the DMAP complexes are shown to the left.

NBO analysis was conducted on the optimized geometries, which supports our suggestion that there is a strong orbital component to Ch-bonding in these systems. Second order perturbation theory revealed that the energy associated with $n(N_{\text{base}}) \rightarrow \sigma^*(N_1-\text{Se})$ delocalisation was 12.79, 15.45, and 16.23 kcal/mol for **1a**·DMAP, **1**·DMAP, and **3**·DMAP respectively.

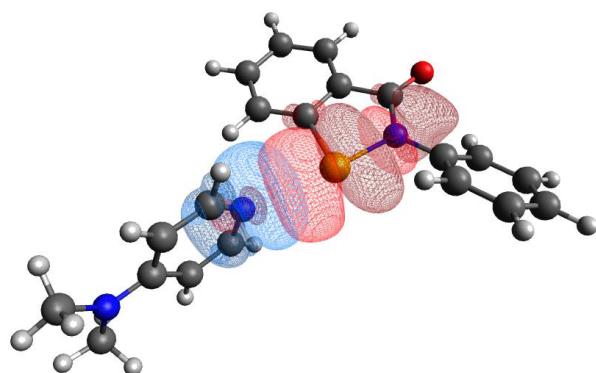


FIGURE 3.6: Overlap of nitrogen lone pair with $\sigma^*(\text{Se}-\text{N})$ in **1**·DMAP complex.

3.4 Conclusion

In summary, we have demonstrated the importance of Ch-bonding between derivatives of ebselen **1** and a variety of nitrogen bases. These selenium-containing heterocycles form close contacts with electron pair donors, well within the Van der Waals radii, with predictable geometries consistent with the Ch-bonding model. These interactions appear to be primarily due to orbital overlap as opposed to electrostatic or dispersion mediated effects, as evidenced by lengthening of the antipodal Se–N bond, and computational analysis, which is consistent with findings in related systems by Cockroft et al.²⁸ We have also found that the strength of a Ch-bond can be enhanced via a hydrogen bond to the carbonyl group of the heterocycle. We hope to exploit the strength and directionality of Ch-bonds in ebselen to target biomolecules such as nucleic acids and proteins using compounds containing the isoselenazolinone moiety.

3.5 Supplementary materials

3.5.1 Synthetic procedures

3.5.1.1 Preparation of 2-phenylbenzo[*d*][1,2]selenazol-3(2*H*)-one **1**.

Copper iodide (98.4 mg, 0.517 mmol) and 1,10-phenanthroline (83.1 mg, 0.461 mmol) were stirred in anhydrous DMF (3 mL) for 15 mins at r.t., then 2-iodo-N-phenylbenzamide (653.2 mg, 2.021 mmol), selenium (196.9 mg, 2.495 mmol) and potassium carbonate (627.3 mg, 4.539 mmol) were added sequentially under a flow of argon. The mixture was heated at 110°C for 8 h, when TLC showed consumption of starting material. The mixture was then tipped into brine (30 mL) and stirred to form a brown precipitate, which was extracted into DCM (40 mL) and washed with water (2 × 20 mL). The DCM solution was filtered through a silica plug, then evaporated,

and the residue applied to a SNAP 25 g silica cartridge, eluting with a petroleum ether/ethyl acetate gradient. The major peak was evaporated to afford colourless crystals of **1** (378.8 mg, 69%, m.p. 179.1–180.3°C, lit. mp 180–181°C).

^1H NMR (499 MHz, d_6 -DMSO) δ ppm 8.07 (1H, d, J = 8.02 Hz), 7.90 (1H, d, J = 7.45 Hz), 7.68 (1H, t, J = 7.25 Hz), 7.61 (2H, d, J = 7.68 Hz), 7.52–7.42 (3H, m), 7.26 (1H, t, J = 7.39 Hz).

^{77}Se NMR (95 MHz, d_6 -DMSO) δ 959.66.

3.5.1.2 Preparation of 2-benzylbenzo[*d*][1,2]selenazol-3(2*H*)-one **1a**.

Copper iodide (95.8 mg, 0.503 mmol) and 1,10-phenanthroline (93.8 mg, 0.521 mmol) were stirred in anhydrous DMF (3 mL) for 15 mins at r.t., then N-benzyl-2-iodobenzamide (860.9 mg, 2.553 mmol), selenium (256.6 mg, 3.249 mmol) and potassium carbonate (542.2 mg, 3.923 mmol) were added sequentially under a flow of argon. The mixture was heated at 110°C for 5 h, when TLC showed consumption of starting material. The mixture was then tipped into brine (30 mL) and stirred to form a solid mass, which was dissolved in DCM (40 mL) and washed with water (2×20 mL). The DCM solution was filtered through a silica plug, then evaporated, and the residue applied to a SNAP 50 g silica cartridge, eluting with a petroleum ether/ethyl acetate gradient. The major peak was evaporated to afford pale yellow crystals of **1a** (396.4 mg, 53%, m.p. 137.8–138.8°C).

^1H NMR (400 MHz, d_6 -DMSO) δ ppm 7.99 (1H, d, J = 8.04 Hz), 7.83 (1H, d, J = 7.7 Hz), 7.59 (1H, t, J = 7.56 Hz), 7.42 (1H, t, J = 7.44 Hz), 7.37–7.22 (5H, m), 4.9 (2H, s).

^{77}Se NMR (95 MHz, d_6 -DMSO) δ 884.02.

3.5.1.3 Preparation of 5*H*-benzo[4,5][1,2]selenazolo[2,3-*a*]quinazolin-5-one **3**.

Copper iodide (96.4 mg, 0.503 mmol) and 1,10-phenanthroline (85.7 mg, 0.476 mmol) were stirred in anhydrous DMF (4 mL) for 10 mins at r.t., then 2-iodobenzamide (510.6 mg, 2.067 mmol), selenium (209.5 mg, 2.653 mmol) and potassium carbonate (506.3 mg, 3.663 mmol) were added sequentially under a flow of argon. The mixture was heated at 110°C for 12 h, when TLC showed consumption of starting material. The mixture was then tipped into brine (30 mL) and stirred to form a solid mass, which was extracted into ethyl acetate (20 mL) and washed with water (2 × 20 mL). The DCM solution was filtered through a silica plug, then evaporated, and the residue applied to a SNAP 50 g silica cartridge, eluting with a petroleum ether/ethyl acetate gradient. The major peak was evaporated to afford pale yellow crystals of **3** (45.8 mg, 15%, m.p. 267–268°C).

¹H NMR (499 MHz, CDCl₃) δ ppm 8.49 (1H, d, *J* = 7.92 Hz), 8.39 (1H, dd, *J*₁ = 1.05 Hz, *J*₂ = 8.01 Hz), 7.91 (1H, d, *J* = 7.85 Hz), 7.88–7.84 (1H, m), 7.8 (1H, d, *J* = 7.98 Hz), 7.76–7.7 (1H, m), 7.63–7.54 (2H, m).

⁷⁷Se NMR (95 MHz, CDCl₃) δ 992.48.

3.5.2 Crystallographic data

Intensity data was collected on an Oxford Diffraction SuperNova CCD diffractometer using either Cu-Kα or Mo-Kα radiation at 130.0(1) K, or on a Rigaku XtalLAB Synergy at 100.0(1) K. Compound **1a**·DMAP·H₂O underwent a destructive phase change when cooling to 130 K, therefore data were collected at 200 K. Data for **3** was collected on the MX1 beam line at the Australian Synchrotron.³⁸ The temperature was maintained using an Oxford Cryostream cooling device. The structures were solved by direct methods and difference Fourier synthesis.³⁹ Thermal ellipsoid plot was generated using the program ORTEP-3⁴⁰ integrated within the WINGX⁴¹ suite of programs.

3.5.2.1 Crystal data for **1·DMAP**

$C_{20}H_{19}N_3OSe$, $M = 396.34$, $T = 130.0$ K, $\lambda = 0.71073$ Å, Triclinic, space group $P\bar{1}$, $a = 8.3674(3)$, $b = 9.8399(5)$, $c = 10.6622(5)$ Å, $\alpha = 93.296(4)^\circ$, $\beta = 93.021(4)^\circ$, $\gamma = 101.210(4)^\circ$, $V = 857.86(7)$ Å³, $Z = 2$. $D_c = 1.534$ mg M⁻³, $\mu(\text{Mo-K}\alpha) = 2.201$ mm⁻¹, $F(000) = 404$, crystal size $0.52 \times 0.34 \times 0.23$ mm. 11339 reflections measured, $\theta_{\max} = 36.66^\circ$, 7889 independent reflections, $R_{\text{int}} = 0.0163$, the final R was 0.0293 ($I > 2\theta(I)$, 6882 reflections) and $wR(F^2)$ was 0.0721 (all data), GOF 0.992. CCDC 1867205. From dichloromethane/pentane (70%) m.p. 111.3–112.1°C.

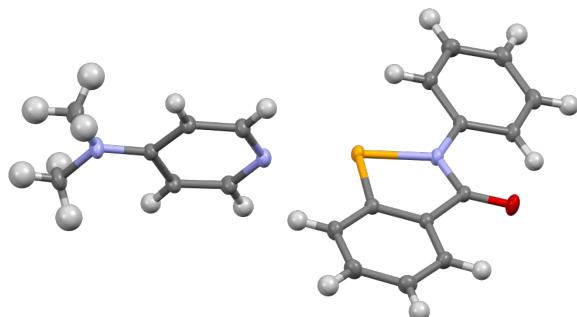
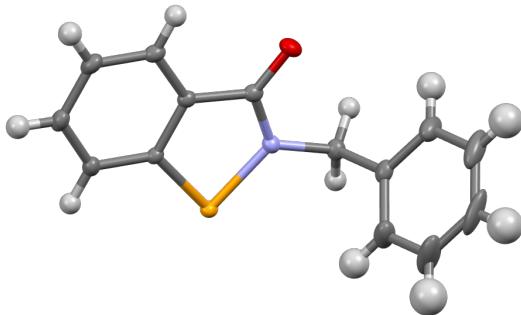


FIGURE 3.7: X-ray crystal structure of **1·DMAP**.

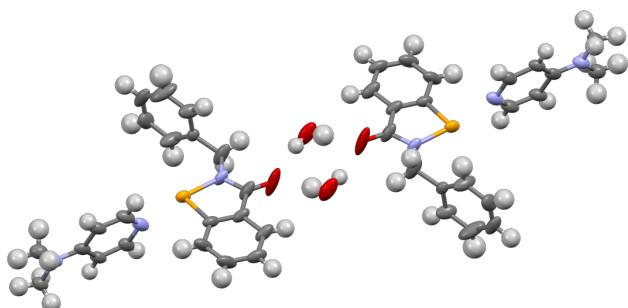
3.5.2.2 Crystal data for **1a**

$C_{14}H_{11}NOSe$, $M = 288.20$, $T = 100.0$ K, $\lambda = 0.71073$ Å, Orthorhombic, space group $Pca2_1$, $a = 11.7848(3)$, $b = 4.5869(1)$, $c = 21.3572(5)$ Å, $V = 1154.48(5)$ Å³, $Z = 4$. $D_c = 1.658$ mg M⁻³, $\mu(\text{Mo-K}\alpha) = 3.233$ mm⁻¹, $F(000) = 576$, crystal size $0.63 \times 0.54 \times 0.22$ mm. 44918 reflections measured, $\theta_{\max} = 45.38^\circ$, 9588 independent reflections, $R_{\text{int}} = 0.0481$, the final R was 0.0331 ($I > 2\theta(I)$, 7848 reflections) and $wR(F^2)$ was 0.0792 (all data), GOF 1.063. CCDC 1867211.

FIGURE 3.8: X-ray crystal structure of **1a**.

3.5.2.3 Crystal data for **1a·DMAP·H₂O**

$C_{21}H_{21}N_3OSe \cdot (H_2O)$, $M = 428.38$, $T = 200.0$ K, $\lambda = 0.71073$ Å, Triclinic, space group $P\bar{1}$, $a = 9.6254(2)$, $b = 10.2486(2)$, $c = 10.6505(2)$ Å, $\alpha = 83.660(2)^\circ$, $\beta = 76.398(2)^\circ$, $\gamma = 78.423(2)^\circ$, $V = 998.19(4)$ Å³, $Z = 2$. $D_c = 125$ mg M⁻³, $\mu(\text{Mo-K}\alpha) = 1.901$ mm⁻¹, $F(000) = 440$, crystal size $0.41 \times 0.32 \times 0.23$ mm. 30047 reflections measured, $\theta_{\max} = 41.06^\circ$, 12528 independent reflections, $R_{\text{int}} = 0.0267$, the final R was 0.0456 ($I > 2\theta(I)$, 6303 reflections) and $wR(F^2)$ was 0.1219 (all data), GOF 1.000. CCDC 1867213. From THF in an open flask (90%) m.p. 96–97°C.

FIGURE 3.9: X-ray crystal structure of **1a·DMAP·H₂O**.

3.5.2.4 Crystal data for **1a·DMAP**

$C_{21}H_{21}N_3OSe$, $M = 410.37$, $T = 130.0$ K, $\lambda = 0.71073$ Å, Triclinic, space group $P\bar{1}$, $a = 9.6002(4)$, $b = 10.2109(4)$, $c = 19.8380(7)$ Å, $\alpha = 78.710(3)^\circ$, $\beta = 84.901(3)^\circ$, $\gamma =$

$77.458(4)^\circ$, $V = 1859.33(13)$ Å 3 , $Z = 4$, $Zt = 2$. $D_c = 1.466$ mg M $^{-3}$, $\mu(\text{Mo-K}\alpha) = 2.034$ mm $^{-1}$, $F(000) = 840$, crystal size $0.65 \times 0.24 \times 0.37$ mm. 36541 reflections measured, $\theta_{\max} = 40.95^\circ$, 23437 independent reflections, $R_{\text{int}} = 0.0264$, the final R was 0.0448 ($I > 2\theta(I)$, 15177 reflections) and $wR(F^2)$ was 0.1120 (all data), GOF 1.044. CCDC 1867209. From dichloromethane/pentane (60%) m.p. 86.1–92.5°C.

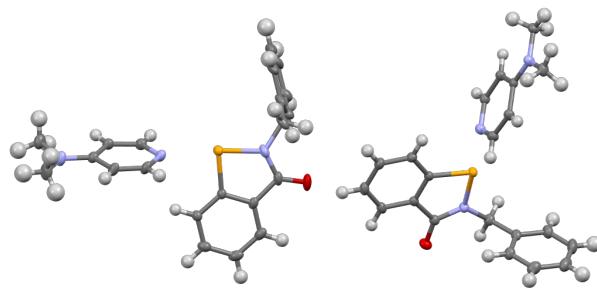


FIGURE 3.10: X-ray crystal structure of **1a**·DMAP.

3.5.2.5 Crystal data for **1a**·quinuclidine

$C_{21}H_{24}N_2OSe$, $M = 399.38$, $T = 130.0$ K, $\lambda = 1.54184$ Å, Monoclinic, space group P2₁/c, $a = 10.1610(2)$, $b = 16.0506(3)$, $c = 11.4300(2)$ Å, $\beta = 104.622(2)^\circ$, $V = 1803.75(6)$ Å 3 , $Z = 4$. $D_c = 1.471$ mg M $^{-3}$, $\mu(\text{Cu-K}\alpha) = 3.895$ mm $^{-1}$, $F(000) = 824$, crystal size $0.29 \times 0.10 \times 0.03$ mm. 12588 reflections measured, $\theta_{\max} = 77.19^\circ$, 3771 independent reflections, $R_{\text{int}} = 0.0379$, the final R was 0.0329 ($I > 2\theta(I)$, 3397 reflections) and $wR(F^2)$ was 0.0849 (all data), GOF 1.028. CCDC 1867207. From dichloromethane/pentane (50%) m.p. 135.2–137.4°C.

3.5.2.6 Crystal data for **1a**·DABCO

$C_{20}H_{23}N_3OSe$, $M = 400.37$, $T = 130.0$ K, $\lambda = 1.54184$ Å, Monoclinic, space group P2₁/c, $a = 10.1249(2)$, $b = 15.9246(3)$, $c = 11.4660(2)$ Å, $\beta = 106.572(2)^\circ$, $V = 1771.93(6)$ Å 3 , $Z = 4$. $D_c = 1.501$ mg M $^{-3}$, $\mu(\text{Cu-K}\alpha) = 2.965$ mm $^{-1}$, $F(000) = 824$, crystal

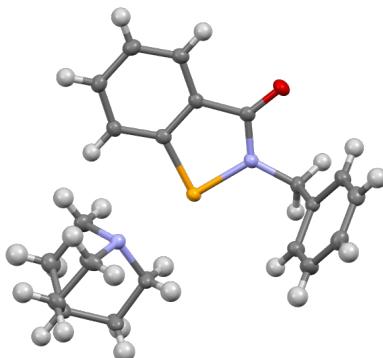


FIGURE 3.11: X-ray crystal structure of **1a**·quinuclidine.

size $0.37 \times 0.17 \times 0.04$ mm. 13121 reflections measured, $\theta_{\max} = 77.12^\circ$, 3711 independent reflections, $R_{\text{int}} = 0.0280$, the final R was 0.0258 ($I > 2\theta(I)$, 3333 reflections) and $wR(F^2)$ was 0.0657 (all data), GOF 1.056. CCDC 1867206. From dichloromethane/pentane (65%) m.p. 131.4–133.3°C.

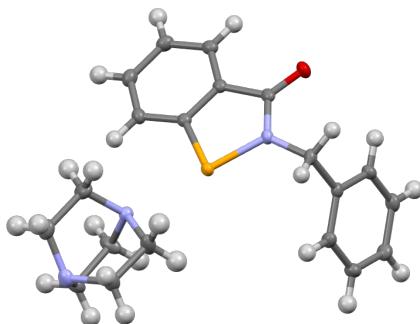
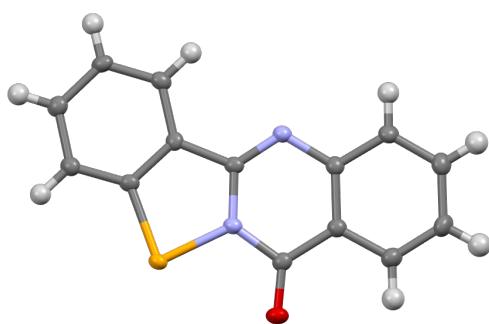


FIGURE 3.12: X-ray crystal structure of **1a**·DABCO.

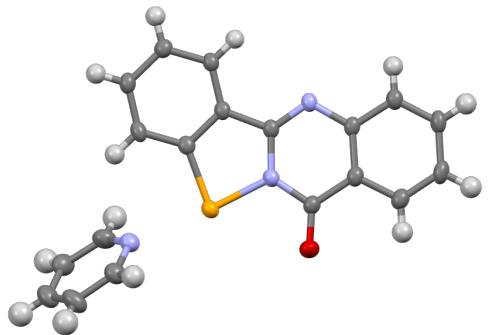
3.5.2.7 Crystal data for 3

$C_{14}H_8N_2OSe$, $M = 299.18$, $T = 100.0$ K, $\lambda = 0.71092$ Å, Orthorhombic, space group Pca2₁, $a = 17.371(4)$, $b = 5.3080(11)$, $c = 11.633(2)$ Å, $V = 1072.6(4)$ Å³, $Z = 4$. $D_c = 1.853$ mg M⁻³, $\mu(\text{Mo-}K\alpha) = 3.486$ mm⁻¹, $F(000) = 592$, crystal size $0.15 \times 0.10 \times 0.02$ mm. 16031 reflections measured, $\theta_{\max} = 31.56^\circ$, 2967 independent reflections, $R_{\text{int}} = 0.0363$, the final R was 0.0271 ($I > 2\theta(I)$, 2061 reflections) and $wR(F^2)$ was 0.0751 (all data), GOF 1.129. CCDC 1867208.

FIGURE 3.13: X-ray crystal structure of **3**.

3.5.2.8 Crystal data for **3**·pyridine

$C_{19}H_{13}N_3OSe$, $M = 378.28$, $T = 130.0$ K, $\lambda = 1.54184$ Å, Monoclinic, space group $P2_1/c$, $a = 20.7476(9)$, $b = 4.9407(2)$, $c = 17.6687(7)$ Å, $\beta = 107.376(4)^\circ$, $V = 1156.27(5)$ Å³, $Z = 4$. $D_c = 1.454$ mg M⁻³, $\mu(\text{Cu}-K\alpha) = 3.018$ mm⁻¹, $F(000) = 760$, crystal size $0.56 \times 0.05 \times 0.03$ mm. 5766 reflections measured, $\theta_{\max} = 75.76^\circ$, 3419 independent reflections, $R_{\text{int}} = 0.0301$, the final R was 0.0346 ($I > 2\theta(I)$, 2889 reflections) and $wR(F^2)$ was 0.0955 (all data), GOF 1.054. CCDC 1867211. From dichloromethane/pentane (70%) m.p. 247.5–248.4°C.

FIGURE 3.14: X-ray crystal structure of **3**·pyridine.

3.5.2.9 Crystal data for **3**·DMAP

$C_{21}H_{18}N_4OSe$, $M = 421.35$, $T = 100.0$ K, $\lambda = 0.71073$ Å, Triclinic, space group $P\bar{1}$, $a = 8.8093(2)$, $b = 10.7445(2)$, $c = 10.9812(2)$ Å, $\alpha = 111.687(2)^\circ$, $\beta = 109.283(2)^\circ$,

$\gamma = 96.631(2)^\circ$, $V = 877.57(3)$ Å 3 , $Z = 2$, $Zt = 2$. $D_c = 1.595$ mg M $^{-3}$, $\mu(\text{Mo-K}\alpha) = 2.159$ mm $^{-1}$, $F(000) = 428$, crystal size $0.18 \times 0.11 \times 0.06$ mm. 56053 reflections measured, $\theta_{\max} = 41.07^\circ$, 11273 independent reflections, $R_{\text{int}} = 0.0547$, the final R was 0.0358 ($I > 2\theta(I)$, 8667 reflections) and $wR(F^2)$ was 0.0872 (all data), GOF 1.048. CCDC 1867212. From dichloromethane/pentane (80%) m.p. 248.8–249.4°C.

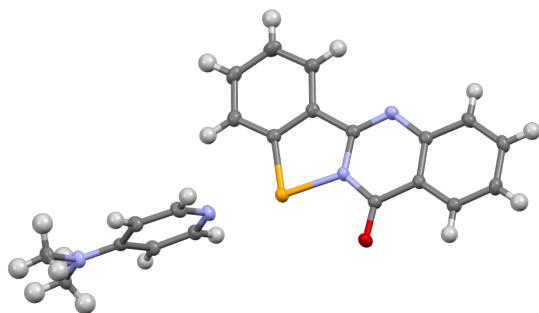


FIGURE 3.15: X-ray crystal structure of **3**·DMAP.

References

- (1) Fellowes, T.; White, J. M. *CrystEngComm* **2019**, *21*, 1539–1542,
DOI: [10.1039/C8CE01853G](https://doi.org/10.1039/C8CE01853G).
- (2) Mitchell, M. O. *Journal of Molecular Modeling* **2017**, *23*, 287,
DOI: [10.1007/s00894-017-3452-3](https://doi.org/10.1007/s00894-017-3452-3).
- (3) Wonner, P.; Vogel, L.; Düser, M.; Gomes, L.; Kniep, F.; Mallick, B.; Werz, D. B.; Huber, S. M. *Angewandte Chemie International Edition* **2017**, *56*, 12009–12012,
DOI: [10.1002/anie.201704816](https://doi.org/10.1002/anie.201704816).
- (4) Fanfrlík, J.; Přáda, A.; Padělková, Z.; Pecina, A.; Macháček, J.; Lepšík, M.; Holub, J.; Růžička, A.; Hnyk, D.; Hobza, P. *Angewandte Chemie - International Edition* **2014**, *53*, 10139–10142,
DOI: [10.1002/anie.201405901](https://doi.org/10.1002/anie.201405901).

- (5) Vogel, L.; Wonner, P.; Huber, S. M. *Angewandte Chemie International Edition* **2019**, *58*, 1880–1891,
DOI: [10.1002/anie.201809432](https://doi.org/10.1002/anie.201809432).
- (6) Paolo, T. D.; Sandorfy, C. *Canadian Journal of Chemistry* **1974**, *52*, 3612–3622,
DOI: [10.1139/v74-540](https://doi.org/10.1139/v74-540).
- (7) Gleiter, R.; Werz, D. B.; Rausch, B. J. *Chemistry - A European Journal* **2003**, *9*,
2676–2683,
DOI: [10.1002/chem.200204684](https://doi.org/10.1002/chem.200204684).
- (8) Kremer, A.; Fermi, A.; Biot, N.; Wouters, J.; Bonifazi, D. *Chemistry (Weinheim an der Bergstrasse, Germany)* **2016**, *22*, 5665–5675,
DOI: [10.1002/chem.201504328](https://doi.org/10.1002/chem.201504328).
- (9) Huynh, H.-T.; Jeannin, O.; Fourmigué, M. *Chem. Commun.* **2017**, 8467–8469,
DOI: [10.1039/C7CC04833E](https://doi.org/10.1039/C7CC04833E).
- (10) Lim, J. Y. C.; Marques, I.; Thompson, A. L.; Christensen, K. E.; Félix, V.; Beer, P. D. *Journal of the American Chemical Society* **2017**, *139*, 3122–3133,
DOI: [10.1021/jacs.6b12745](https://doi.org/10.1021/jacs.6b12745).
- (11) Lim, J. Y. C.; Liew, J. Y.; Beer, P. D. *Chem. - A Eur. J.* **2018**, *24*, 14560–14566,
DOI: [10.1002/chem.201803393](https://doi.org/10.1002/chem.201803393).
- (12) Garrett, G. E.; Carrera, E. I.; Seferos, D. S.; Taylor, M. S. *Chem. Commun.* **2016**,
52, 9881–9884,
DOI: [10.1039/C6CC04818H](https://doi.org/10.1039/C6CC04818H).
- (13) Wonner, P.; Vogel, L.; Kniep, F.; Huber, S. M. *Chemistry - A European Journal* **2017**, *23*, 16972–16975,
DOI: [10.1002/chem.201704502](https://doi.org/10.1002/chem.201704502).
- (14) Benz, S.; López-Andarias, J.; Mareda, J.; Sakai, N.; Matile, S. *Angewandte Chemie International Edition* **2017**, *56*, 812–815,
DOI: [10.1002/anie.201611019](https://doi.org/10.1002/anie.201611019).

- (15) Benz, S.; Mareda, J.; Besnard, C.; Sakai, N.; Matile, S. *Chemical Science* **2017**, *8*, 8164–8169,
DOI: [10.1039/C7SC03866F](https://doi.org/10.1039/C7SC03866F).
- (16) Iwaoka, M.; Takemoto, S.; Okada, M.; Tomoda, S. *Chemistry Letters* **2001**, 132–133,
DOI: [10.1246/cl.2001.132](https://doi.org/10.1246/cl.2001.132).
- (17) Iwaoka, M. In *Noncovalent Forces*, Scheiner, S., Ed.; Springer International Publishing: Cham, 2015, pp 265–289,
DOI: [10.1007/978-3-319-14163-3_9](https://doi.org/10.1007/978-3-319-14163-3_9).
- (18) Garrett, G. E.; Gibson, G. L.; Straus, R. N.; Seferos, D. S.; Taylor, M. S. *Journal of the American Chemical Society* **2015**, *137*, 4126–4133,
DOI: [10.1021/ja512183e](https://doi.org/10.1021/ja512183e).
- (19) Ho, P. C.; Szydlowski, P.; Sinclair, J.; Elder, P. J. W.; Kübel, J.; Gendy, C.; Lee, L. M.; Jenkins, H.; Britten, J. F.; Morim, D. R.; Vargas-Baca, I. *Nature Communications* **2016**, *7*, 11299,
DOI: [10.1038/ncomms11299](https://doi.org/10.1038/ncomms11299).
- (20) Ho, P. C.; Rafique, J.; Lee, J.; Lee, L. M.; Jenkins, H. A.; Britten, J. F.; Braga, A. L.; Vargas-Baca, I. *Dalton Trans.* **2017**, *46*, 6570–6579,
DOI: [10.1039/C7DT00612H](https://doi.org/10.1039/C7DT00612H).
- (21) Müller, A.; Cadenas, E.; Graf, P.; Sies, H. *Biochemical Pharmacology* **1984**, *33*, 3235–3239,
DOI: [10.1016/0006-2952\(84\)90083-2](https://doi.org/10.1016/0006-2952(84)90083-2).
- (22) Schewe, T. *General Pharmacology* **1995**, *26*, 1153–1169,
DOI: [10.1016/0306-3623\(95\)00003-J](https://doi.org/10.1016/0306-3623(95)00003-J).
- (23) Kil, J.; Pierce, C.; Tran, H.; Gu, R.; Lynch, E. D. *Hearing Research* **2007**, *226*, 44–51,
DOI: [10.1016/j.heares.2006.08.006](https://doi.org/10.1016/j.heares.2006.08.006).

- (24) Singh, N.; Halliday, A. C.; Thomas, J. M.; Kuznetsova, O.; Baldwin, R.; Woon, E. C.; Aley, P. K.; Antoniadou, I.; Sharp, T.; Vasudevan, S. R.; Churchill, G. C. *Nature Communications* **2013**, *4*, 1332–1337,
DOI: [10.1038/ncomms2320](https://doi.org/10.1038/ncomms2320).
- (25) Parnham, M.; Sies, H. *Expert Opinion on Investigational Drugs* **2000**, *9*, 607–619,
DOI: [10.1517/13543784.9.3.607](https://doi.org/10.1517/13543784.9.3.607).
- (26) Thomas, S. P.; Satheeshkumar, K.; Mugesh, G.; Gururow, T. N. *Chemistry - A European Journal* **2015**, *21*, 6793–6800,
DOI: [10.1002/chem.201405998](https://doi.org/10.1002/chem.201405998).
- (27) Oliveira, V.; Cremer, D.; Kraka, E. *Journal of Physical Chemistry A* **2017**, *121*, 6845–6862,
DOI: [10.1021/acs.jpca.7b06479](https://doi.org/10.1021/acs.jpca.7b06479).
- (28) Pascoe, D. J.; Ling, K. B.; Cockroft, S. L. *Journal of the American Chemical Society* **2017**, *139*, 15160–15167,
DOI: [10.1021/jacs.7b08511](https://doi.org/10.1021/jacs.7b08511).
- (29) De Vleeschouwer, F.; Denayer, M.; Pinter, B.; Geerlings, P.; De Proft, F. *Journal of Computational Chemistry* **2017**, 557–572,
DOI: [10.1002/jcc.25099](https://doi.org/10.1002/jcc.25099).
- (30) Kolář, M. H.; Hobza, P. *Chemical Reviews* **2016**, *116*, 5155–5187,
DOI: [10.1021/acs.chemrev.5b00560](https://doi.org/10.1021/acs.chemrev.5b00560).
- (31) Gleiter, R.; Haberhauer, G.; Werz, D. B.; Rominger, F.; Bleiholder, C. *Chemical Reviews* **2018**, *118*, 2010–2041,
DOI: [10.1021/acs.chemrev.7b00449](https://doi.org/10.1021/acs.chemrev.7b00449).
- (32) Präsang, C.; Whitwood, A. C.; Bruce, D. W. *Crystal Growth and Design* **2009**, *9*, 5319–5326,
DOI: [10.1021/cg900823d](https://doi.org/10.1021/cg900823d).

- (33) Mulliken, R. S. *Journal of the American Chemical Society* **1950**, *72*, 600–608,
DOI: [10.1021/ja01157a151](https://doi.org/10.1021/ja01157a151).
- (34) Balkrishna, S. J.; Bhakuni, B. S.; Chopra, D.; Kumar, S. *Organic Letters* **2010**,
12, 5394–5397,
DOI: [10.1021/ol102027j](https://doi.org/10.1021/ol102027j).
- (35) Batsanov, S. S. *Inorganic Materials* **2001**, *37*, 871–885,
DOI: [10.1023/A:1011625728803](https://doi.org/10.1023/A:1011625728803).
- (36) Berthelot, M.; Laurence, C.; Safar, M.; Besseau, F. *Journal of the Chemical Society, Perkin Transactions 2* **1998**, 283–290,
DOI: [10.1039/a706696a](https://doi.org/10.1039/a706696a).
- (37) Graton, J.; Besseau, F; Berthelot, M.; Raczynska, E. D.; Laurence, C. *Canadian Journal of Chemistry* **2002**, *80*, 1375–1385,
DOI: [10.1139/v02-176](https://doi.org/10.1139/v02-176).
- (38) Cowieson, N. P.; Aragao, D.; Clift, M.; Ericsson, D. J.; Gee, C.; Harrop, S. J.;
Mudie, N.; Panjikar, S.; Price, J. R.; Riboldi-Tunnicliffe, A.; Williamson, R.;
Caradoc-Davies, T. *Journal of Synchrotron Radiation* **2015**, *22*, 187–190,
DOI: [10.1107/S1600577514021717](https://doi.org/10.1107/S1600577514021717).
- (39) Sheldrick, G. M. *Acta Crystallographica Section C: Structural Chemistry* **2015**,
71, 3–8,
DOI: [10.1107/S2053229614024218](https://doi.org/10.1107/S2053229614024218).
- (40) Farrugia, L. J. *Journal of Applied Crystallography* **1997**, *30*, 565,
DOI: [10.1107/S0021889897003117](https://doi.org/10.1107/S0021889897003117).
- (41) Farrugia, L. J. *Journal of Applied Crystallography* **1999**, *32*, 837–838,
DOI: [10.1107/S0021889899006020](https://doi.org/10.1107/S0021889899006020).

Chapter 4

Further investigations into Ch-bonded complexes

4.1 Introduction

In the previous chapter, we established that Ch-bonding is not only present in ebselen derivatives, but dominates packing in crystals of the pure compound (through Se \cdots O interactions) and in co-crystals with a variety of Lewis bases. In this chapter, we extend our investigation to a wider range of derivatives with systematically varied electronic properties.

Linear free energy relationships (LFERs) relate the rate of a chemical reaction with some electronic property of the substrate molecule(s). Although the single crystal structures presented here are *static* snapshots of reality, we can think of them as representative of the transition state of the breaking of the endocyclic Se–N bond (fig. 4.1). The lower the transition state energy, the shorter will be the Se \dots N Ch-bond distance, and this is clearly influenced by the electronic properties of both the Ch-bond donor and acceptor. They have been used previously to explain trends in X-bonding.^{1,2}

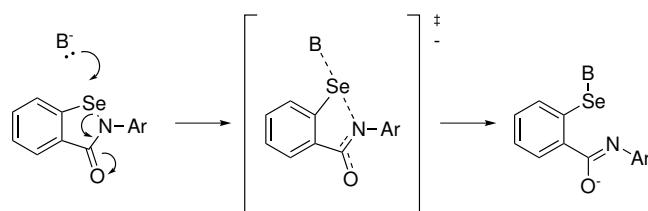
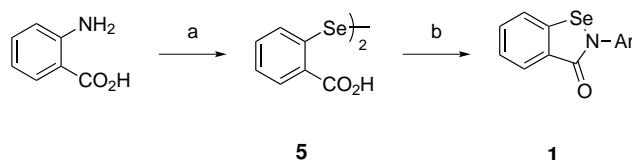


FIGURE 4.1: Depiction of the bond breaking/forming process of which Ch-bonding is a ground state manifestation.

4.2 Results and discussion

Our previous work had shown that electron-rich pyridines (specifically DMAP) formed the strongest Ch-bonds out of all the bases trialled (chapter 3). This is roughly consistent with the hydrogen bond basicity (pK_{HB}) of the bases (table 3.2). The planar geometry and aromatic character of DMAP may also facilitate crystallisation, as opposed to the relatively bulky and flexible aliphatic bases which may not pack as efficiently. For these reasons, we restricted the bases used in this study to other electron-rich pyridines. Although DMAP is already a very strong base, the basicity can be increased by incorporating the aniline nitrogen in another ring. This reduces the energetic penalty associated with the delocalisation of the lone pair into the pyridine ring, by forcing a more planar geometry upon the nitrogen.^{3,4} We therefore prepared co-crystals with **4a** and **4b**, which are slightly stronger and weaker bases than DMAP respectively.



SCHEME 4.1: Synthesis of benzisoselenazolinone derivatives. (a) 1. NaNO_2 , HCl , 0°C ; 2. Na_2Se_2 , NaOH , rt, 1 h, 37%; (b) 1. SOCl_2 , cat. DMF , reflux, 1.5 h; 2. ArNH_2 , TEA , MeCN , rt, 2 h, 14–67%.

The benzisoselenazolinone derivatives **1** and **1NO₂,CN,CF₃,Br,CO₂Et,Me,OMe,OEt** were prepared by the reaction of substituted anilines with a selenenyl chloride

intermediate **5**, ultimately derived from anthranilic acid (scheme 4.1). The one-pot selenocyclisation reaction used in chapter 3 often did not tolerate the various functional groups on the aryl ring, affording reduction, cross-coupling, or protode-halogenation byproducts. All derivatives except **1NO₂** were isolated in acceptable yield at room temperature using acetonitrile as a solvent, and in the presence of a weak base (triethylamine). Due to the strongly electron-withdrawing nature of the nitro substituent, the parent aniline of **1NO₂** was not sufficiently nucleophilic to react under the same conditions. We therefore first deprotonated the aniline using NaH (60% suspension in mineral oil) in anhydrous THF before adding the selenyl chloride, which afforded the benzisoselenazolinone in excellent yield.

We also note the solubility trends of the products as the electron demand of the substituent increases. The electron rich derivatives **1OMe**, **1OEt** and **1Me** were highly soluble even in relatively poor solvents (Et₂O, EtOAc), while the electron poor derivatives **1NO₂** and **1CN** were insoluble in all but the strongest solvents (DMSO). This likely reflects the varying strength of the crystal packing due to the Se···O=C Ch-bond.

Co-crystals of the benzisoselenazolinones and pyridines were grown by vapour diffusion from an equimolar solution in dichloromethane/pentane. Relevant structural parameters are given in table 4.1. In addition to the structural parameters, we characterised the various Ch-bonds using Bader's QTAIM framework. Depending on the quality of the data we were able to obtain for each crystal, we used:

- experimental electron density from a multipole refinement with atomic coordinates determined from high angle refinement ($d < 0.8\text{\AA}$),
- experimental electron density from a multipole refinement with atomic coordinates determined by refinement using aspherical scattering factors derived from the DFT electron density,
- density derived from a DFT calculation on the crystal geometry.

The second “hybrid” strategy has the advantage of being able to capture electron density effects not fully described by DFT, while not requiring a large amount of weak high-angle (or neutron) data for the reliable determination of atomic coordinates. The low-angle data, however, does need to be free from faults, so this strategy may still not be appropriate for all data sets.

For the second and third strategies, the densities and scattering factors were calculated using NoSphereA2 incorporated in Olex2.⁵

4.2.1 Hammett plots of crystallographic data.

Evident in this data is a trend of increasing Ch-bond length with increasing electron donating character of the substituent. There are a number of methods to quantify this property of the substituent, and we will explore a couple of the most common. The Hammett *para* substituent parameter σ_p for a given substituent at the *para* position is determined from the ionization equilibrium of the parent carboxylic acid. Although originally used to explain reaction kinetics with an associated reaction constant ρ , the substituent constant provides a convenient measure of electron donating character for ground state phenomena as well. Within the context of Ch-bonding, this can be rationalised by considering the approach of the Lewis base to form the Ch-bond as an incipient nucleophilic substitution at the selenium, and invoking the Hammond postulate to relate the transition state geometry to the ground state (fig. 4.1).

Closely linked to the substituent constant σ_p are the values of σ_p^+ and σ_p^- , which can be used when mesomeric effects have a strong influence on the property being studied. Hammett substituent parameters are particularly convenient as they have been determined for a wide variety of substituents. Parameters for the following analysis were obtained from reference [6].

TABLE 4.1: Selected structural and electron density parameters of Ch-bonded complexes.

Complex	r(N···Se)	r(Se–N ₁)	r(Se–C ₁)	r(N ₁ –C(O))	∠(N···Se–N ₁)	∠(C _{para} ···N···Se)	$\rho_{BCP}(Se\cdots N)$	$\nabla^2(\rho_{BCP})(Se\cdots N)$
	Å	Å	Å	Å	°	°	e/Å ³	e/Å ⁵
1	—	1.896(3)	1.892(4)	1.359(5)	—	—	—	—
1NO₂	—	—	—	—	—	—	—	—
1CN	—	1.894(2)	1.877(2)	1.372(2)	—	—	—	—
1CF₃^a	—	1.880(9)	1.889(9)	1.38(1)	—	—	—	—
1CF₃^b	—	1.898(8)	1.901(9)	1.37(1)	—	—	—	—
1Br	—	1.898(2)	1.889(2)	1.371(3)	—	—	—	—
1CO₂Et	—	1.902(2)	1.879(2)	1.371(3)	—	—	—	—
1Me	—	1.904(3)	1.890(3)	1.365(4)	—	—	—	—
1OMe	—	1.8741(9)	1.887(1)	1.356(1)	—	—	—	—
1OEt	—	1.901(3)	1.885(4)	1.357(4)	—	—	—	—
1-DMAP	2.371(1)	1.968(1)	1.896(1)	1.358(2)	174.18(4)	173.51(6)	0.3511	2.6960 ^c
1NO₂-DMAP	2.2424(5)	2.0200(4)	1.9086(4)	1.3592(4)	173.57(2)	175.47(2)	0.5372	3.8680 ^c
1CN-DMAP	2.301(1)	1.997(1)	1.899(2)	1.368(2)	174.79(5)	167.57(6)	0.4130	2.5210 ^c
1CN-DMAP^{d,a}	2.254(2)	2.019(2)	1.902(1)	1.366(2)	174.46(6)	176.33(7)	0.4780	2.4816 ^e
1CN-DMAP^{d,b}	2.308(2)	1.993(1)	1.901(1)	1.372(2)	174.93(5)	167.85(7)	0.4284	2.4558 ^e
1CF₃-DMAP	2.3347(9)	1.9855(9)	1.899(1)	1.372(1)	175.24(4)	162.65(5)	0.4048	2.4112 ^e
1Br-DMAP	2.3215(7)	1.9840(7)	1.9021(8)	1.362(1)	173.85(3)	173.56(4)	0.4058	3.1160 ^c
1CO₂Et-DMAP^d	2.322(1)	1.982(1)	1.902(1)	1.367(2)	174.96(4)	172.86(6)	0.4143	2.4585 ^e
1Me-DMAP	2.4301(4)	1.9341(4)	1.8918(4)	1.3650(6)	175.33(1)	158.64(2)	0.2843	3.2570 ^c
1OMe-DMAP^{f,a}	2.270(1)	1.9689(9)	1.899(1)	1.350(1)	174.17(3)	159.44(4)	0.4545	2.5544 ^e
1OMe-DMAP^{f,b}	2.4496(9)	1.9267(9)	1.895(1)	1.357(1)	174.57(3)	160.53(4)	0.3207	2.1731 ^e
1OMe-DMAP^{g,a}	2.334(1)	1.965(1)	1.894(1)	1.363(1)	175.95(5)	155.71(6)	0.4421	3.1590 ^c
1OMe-DMAP^{g,b}	2.407(1)	1.941(1)	1.894(1)	1.358(2)	176.12(5)	156.60(6)	0.3806	2.9170 ^c
1OEt-DMAP^a	2.517(2)	1.921(1)	1.895(2)	1.356(3)	172.30(6)	158.37(8)	0.3641	2.9344 ^c
1OEt-DMAP^b	2.327(5)	1.931(2)	1.894(2)	1.353(3)	171.4(1)	151.2(3)	0.2565	2.3493 ^c
1-4a	2.350(1)	1.9830(9)	1.8985(7)	1.3616(8)	174.20(3)	176.46(4)	0.2419	5.4650 ^c
1NO₂-4a	—	—	—	—	—	—	—	—
1CN-4a	2.289(1)	2.000(1)	1.902(1)	1.363(2)	174.77(5)	175.06(7)	0.4914	3.4270 ^b
1CF₃-4a	—	—	—	—	—	—	—	—
1Br-4a	2.319(2)	1.981(2)	1.895(1)	1.362(2)	174.57(6)	174.60(8)	0.3617	3.7020 ^c
1CO₂Et-4a	2.337(3)	1.982(3)	1.906(4)	1.374(5)	173.5(1)	170.1(2)	0.4014	2.4107 ^e
1Me-4a	2.272(1)	1.986(1)	1.905(1)	1.351(2)	172.66(5)	173.51(6)	0.4545	2.5279 ^e
1OMe-4a	—	—	—	—	—	—	—	—
1OEt-4a^a	2.356(3)	1.972(3)	1.898(3)	1.355(4)	174.5(1)	170.2(1)	0.3878	2.4124 ^e
1OEt-4a^b	2.371(3)	1.964(3)	1.898(3)	1.360(4)	174.9(1)	170.9(1)	0.3774	2.3640 ^e
1-4b^f	2.414(2)	1.960(2)	1.902(2)	1.366(2)	175.46(6)	166.23(8)	0.2584	3.4620 ^c
1-4b^g	2.420(2)	1.967(2)	1.903(2)	1.363(2)	176.7(1)	174.23(8)	0.3416	2.2739 ^e
1NO₂-4b	—	—	—	—	—	—	—	—
1CN-4b^d	2.301(1)	1.993(1)	1.907(1)	1.368(2)	173.08(5)	164.75(6)	0.4306	2.4546 ^h
1CF₃-4b	—	—	—	—	—	—	—	—
1Br-4b	2.381(2)	1.975(2)	1.905(2)	1.367(2)	173.84(6)	174.97(8)	0.3536	3.4890 ^c
1CO₂Et-4b	2.337(3)	1.982(3)	1.906(4)	1.374(5)	173.5(1)	170.1(2)	0.4286	2.4674 ^e
1Me-4b	2.412(5)	1.977(5)	1.908(5)	1.360(7)	174.4(2)	178.9(2)	0.3456	2.2774 ^e
1OMe-4b	—	—	—	—	—	—	—	—
1OEt-4b^a	2.398(4)	1.958(4)	1.894(6)	1.367(6)	174.6(2)	174.1(2)	0.3553	2.3309 ^e
1OEt-4b^b	2.448(6)	1.951(4)	1.901(5)	1.367(5)	175.5(2)	170.5(3)	0.3239	2.2095 ^e

^aEnvironment *a*^bEnvironment *b*^cFully experimental density used^dDCM solvate^eDFT density used^fPolymorph 1^gPolymorph 2^hHybrid density used

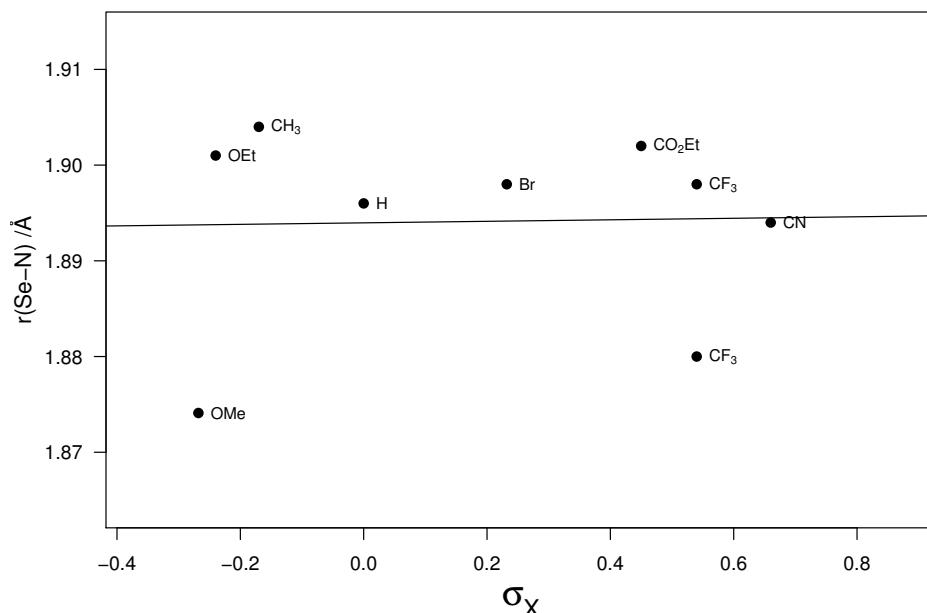


FIGURE 4.2: Hammett plot of endocyclic Se–N bond length of uncomplexed ebselen derivatives.

The structural effects most relevant to Ch-bonding will manifest themselves in the vicinity of the selenium atom. Of particular interest is the endocyclic Se–N bond length, which serves as a measure of $\sigma^*(\text{Se}–\text{N})$ orbital occupancy, thus the degree of hyperconjugation and character of the Ch-bond.

As can be seen in fig. 4.2, there is practically no correlation between the electronic properties of the aryl ring and the endocyclic Se–N bond length in crystals of the unbound ebselen derivatives. This is not unexpected, as in all cases the crystals consist of one dimensional chains of ebselen molecules Ch-bonded to the carbonyl oxygen of the next (fig. 4.3). As the magnitude of the σ -hole (Ch-bond donor ability) is *increased* by an electron withdrawing substituent, the Ch-bond acceptor ability of the carbonyl is *decreased*. These opposing effects appear to be approximately equal in magnitude, so cancel each other out and give a very flat and featureless Hammett plot.

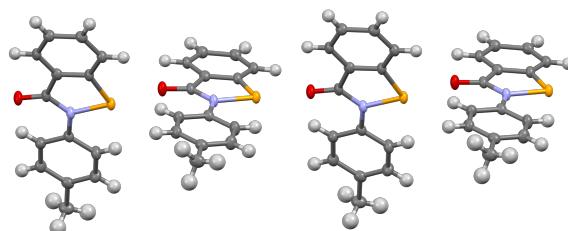


FIGURE 4.3: One dimensional chains formed by Ch-bonding between the selenium and carbonyl oxygen in **1Me**. All other ebselen derivatives display a similar packing motif.

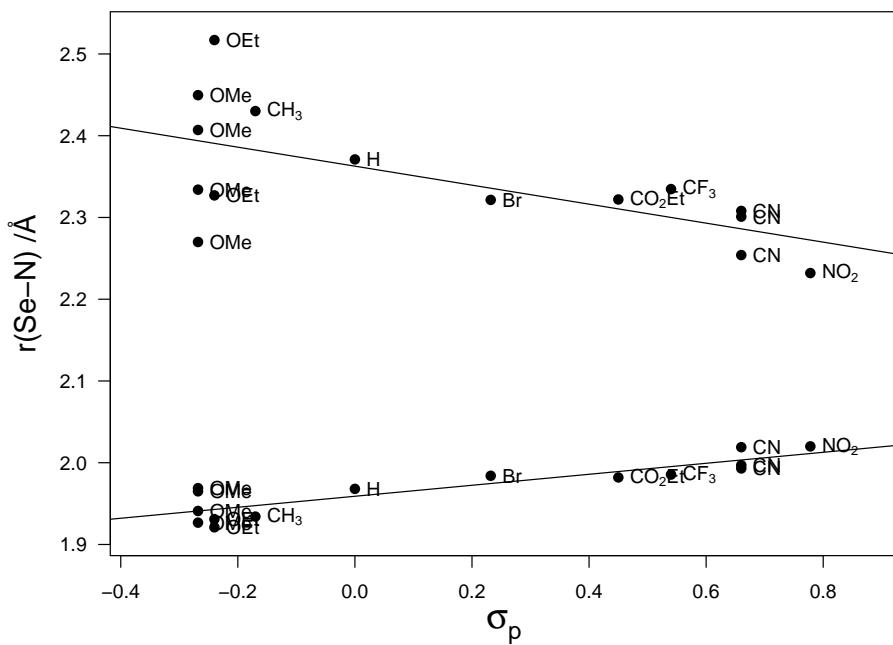


FIGURE 4.4: Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with DMAP. The lines are described by the equations $r(\text{Se}-\text{N}) = (1.959(5) + 0.067(10) \times \sigma_p) \text{\AA}$ ($R^2 = 0.7857$) and $r(\text{Se}\cdots\text{N}) = (2.363(17) - 0.12(4) \times \sigma_p) \text{\AA}$ ($R^2 = 0.4104$).

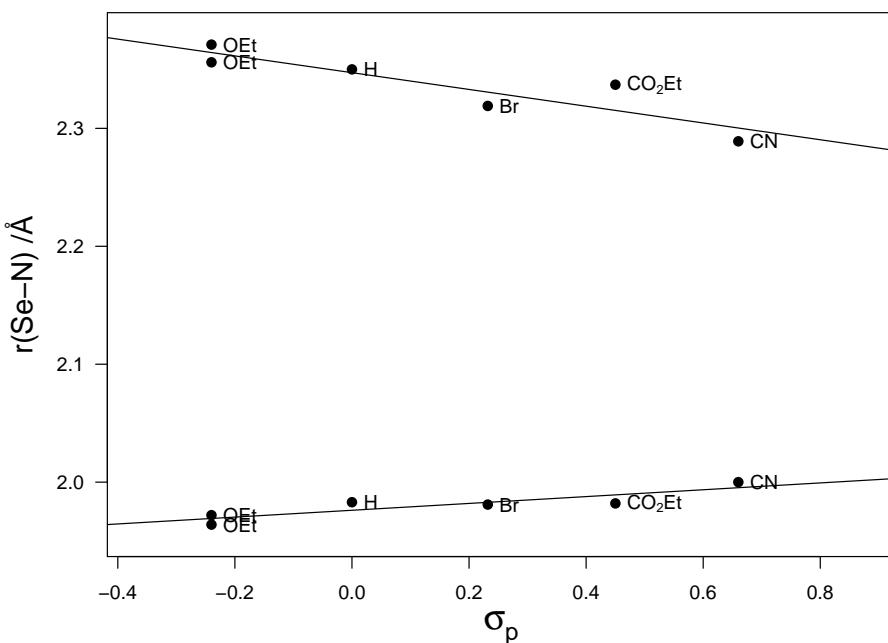


FIGURE 4.5: Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with **4a**. The lines are described by the equations $r(\text{Se}-\text{N}) = (1.9761(28) + 0.029(8) \times \sigma_p) \text{\AA}$ ($R^2 = 0.7828$) and $r(\text{Se}\cdots\text{N}) = (2.347(7) - 0.071(17) \times \sigma_p) \text{\AA}$ ($R^2 = 0.8005$)

TABLE 4.2: Linear regression results for Hammett plots in figs. 4.4 to 4.6.

Base	pK_{HB}^3	$r(\text{Se}\cdots\text{N})$			$r(\text{Se}-\text{N})$		
		gradient	intercept	R^2	gradient	intercept	R^2
4b	2.68	-0.13(2)	2.400(7)	0.8522	0.036(9)	1.967(3)	0.7286
DMAP	2.80	-0.12(4)	2.363(17)	0.4104	0.067(10)	1.959(5)	0.7857
4a	2.93	-0.071(17)	2.347(7)	0.8005	0.029(8)	1.9761(28)	0.7828

Fortunately, inspecting the same bond length in co-crystals of ebselen derivatives and a Lewis base gives us a much clearer dependence, as can be seen in figs. 4.4 to 4.6. This is because the Ch-bond acceptor ability is now independent of the electronic properties of the Ch-bond donor, so the Se–N bond length is determined solely by the latter. An inverse correlation can be seen in the Se···N Ch-bond length. The results are tabulated in table 4.2. Moderately good correlation is obtained, as measured by the R^2 values, although there is clearly some deviation on the left hand

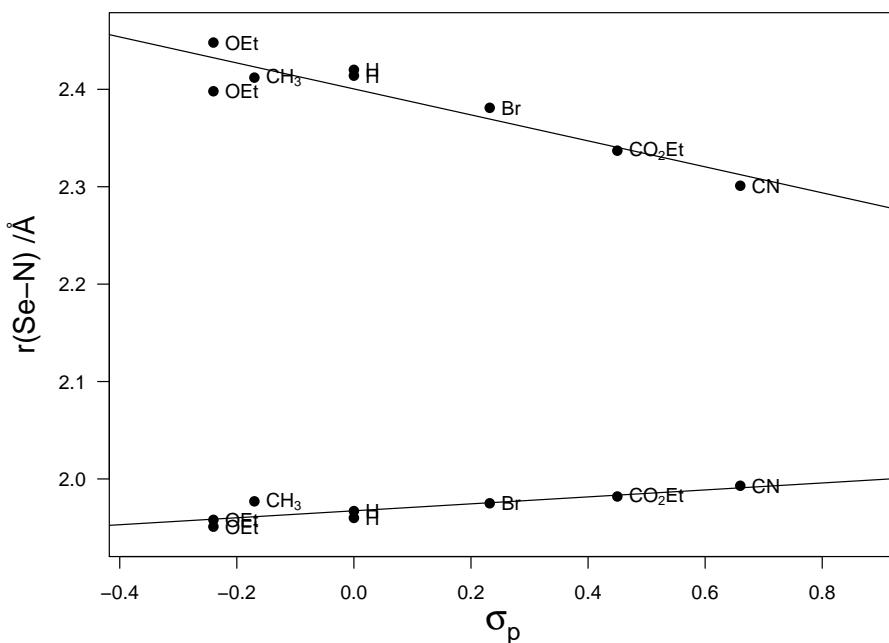


FIGURE 4.6: Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with **4b**. The line is described by the equation $r(\text{Se}-\text{N}) = (1.967(3) + 0.036(9) \times \sigma_p) \text{ \AA}$ ($R^2 = 0.7386$) and $r(\text{Se}\cdots\text{N}) = (2.400(7) - 0.13(2) \times \sigma_p) \text{ \AA}$ ($R^2 = 0.8522$).

side of the plots. While the more strongly Ch-bonded systems (with electron withdrawing substituents) are generally very well described by the model, the electron rich derivatives **1OMe** and **1OEt** vary significantly in their bond lengths.* Indeed, in the case of DMAP complexes, omitting these data points improves the correlation coefficient to 0.8148 while the gradient and intercept are almost unchanged at $-0.156(28) \text{ \AA}$ and $2.386(15) \text{ \AA}$ respectively, suggesting that the model is appropriate, and that there is some other effect occurring in electron rich systems. For a further discussion of this phenomenon, see section 4.2.1.1.

It has been found that phenomena that are dominated by inductive or field effects (as opposed to mesomeric effects) are better described by the *meta* substituent constant σ_m .^{2,7} We therefore constructed Hammett plots using this parameter as well, which are shown in figs. 4.7 to 4.9, and the results tabulated in table 4.3.

*This is also visible, though less apparent, in the plot of endocyclic bond lengths (fig. 4.4).

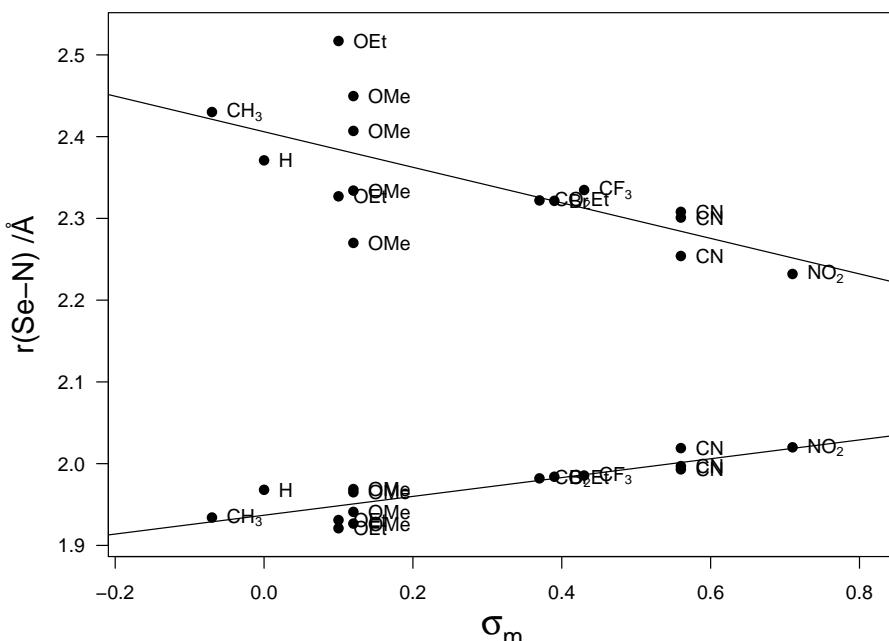


FIGURE 4.7: Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with DMAP. The lines are described by the equations $r(\text{Se}-\text{N}) = (1.937(7) + 0.011(2) \times \sigma_m) \text{\AA}$ ($R^2 = 0.7501$) and $r(\text{Se}\cdots\text{N}) = (2.41(2) - 0.22(6) \times \sigma_m) \text{\AA}$ ($R^2 = 0.4680$).

TABLE 4.3: Linear regression results for Hammett plots in figs. 4.7 to 4.9.

Base	pK_{HB}^3	$r(\text{Se}\cdots\text{N})$			$r(\text{Se}-\text{N})$		
		gradient	intercept	R^2	gradient	intercept	R^2
4b	2.68	-0.18(4)	2.42(1)	0.7438	0.04(2)	1.963(5)	0.4620
DMAP	2.80	-0.22(6)	2.41(2)	0.4680	0.11(2)	1.937(7)	0.7501
4a	2.93	-0.12(3)	2.368(10)	0.7892	0.04(2)	1.970(6)	0.5098

We obtain a slightly poorer fit when using σ_m compared to σ_p , which suggests that mesomeric effects may play a significant role. We note, however, that the correlation is well below what would be accepted as “good” for a Hammett plot ($R^2 < 0.95$),⁸ which we attribute to scatter caused by crystal packing forces. The difference between the σ_m and σ_p correlations are therefore probably not significant.

Nonetheless, in both cases we can see a qualitative link between the strength of the base used and the gradient of the line of best fit. As the base gets stronger,

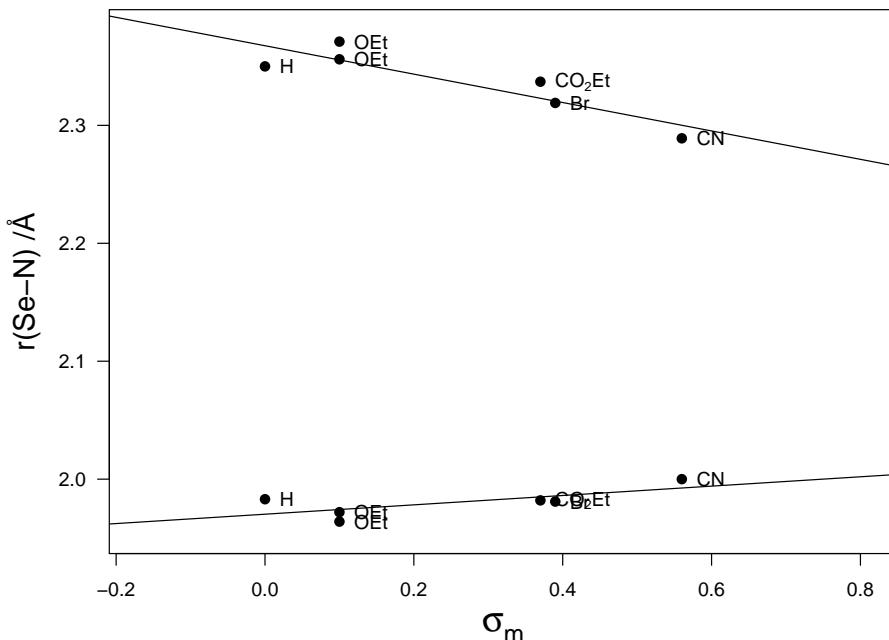


FIGURE 4.8: Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with **4a**. The lines are described by the equations $r(\text{Se}-\text{N}) = (1.970(6) + 0.04(2) \times \sigma_m) \text{ \AA}$ ($R^2 = 0.5098$) and $r(\text{Se}\cdots\text{N}) = (2.368(10) - 0.12(3) \times \sigma_m) \text{ \AA}$ ($R^2 = 0.7892$)

the Ch-bond length varies less with the electron demand of the Ch-bond donor. This can be rationalised as the potential energy surface for the Ch-bond stretching becomes steeper as the base gets stronger, meaning the distance is less affected by the electronics of the donor. Incidentally, we believe this is also why the more electron poor systems do not display the same variation in bond lengths that we see in the electron rich systems (see section 4.2.1.1 for further details).

Hammett plots can also be constructed using the electron density parameters at the bond critical point (ρ_{BCP} and $\nabla^2\rho_{\text{BCP}}$) of the Ch bond (figs. 4.10 to 4.12). The electron density ρ_{BCP} shows a similar trend to the bond distance ($\rho_{\text{BCP}} = (0.379(16) + 0.105(37) \times \sigma_p) \text{ e}/\text{\AA}^3$ ($R^2 = 0.3783$)), while the Laplacian $\nabla^2\rho_{\text{BCP}}$ is more or less independent of the Hammett constant ($\nabla^2\rho_{\text{BCP}} = (2.74(13) + 0.05(29) \times \sigma_p) \text{ e}/\text{\AA}^5$ ($R^2 = 0.002$)). This reinforces the significance of the Laplacian not as an indicator of the

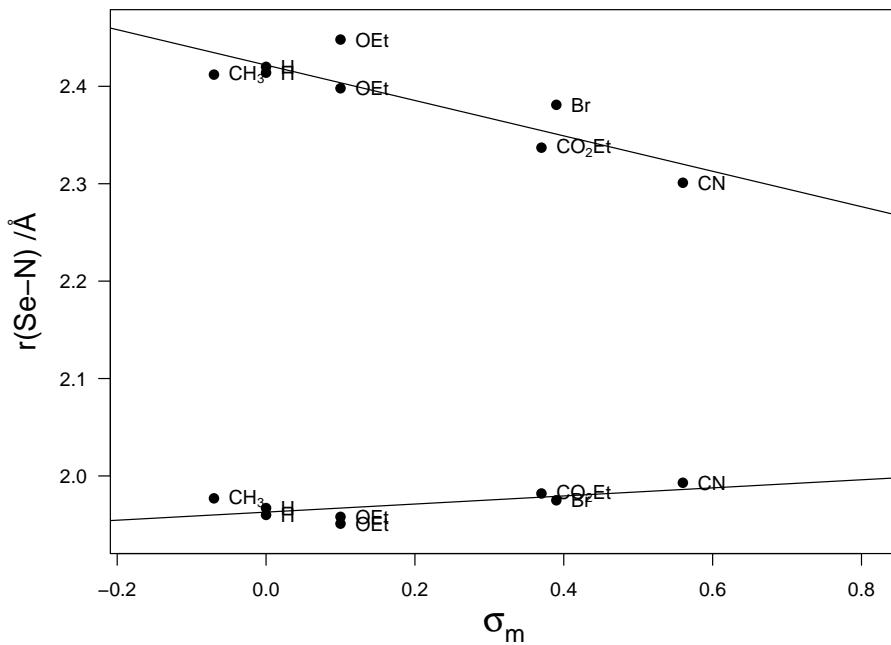


FIGURE 4.9: Hammett plots of endocyclic Se–N bond length and Se···N Ch-bond length of ebselen derivatives complexed with **4b**. The line is described by the equation $r(\text{Se}-\text{N}) = (1.963(5) + 0.04(2) \times \sigma_m) \text{\AA}$ ($R^2 = 0.4620$) and $r(\text{Se}\cdots\text{N}) = (2.42(1) - 0.18(4) \times \sigma_m) \text{\AA}$ ($R^2 = 0.7438$).

strength of an interaction, but as a measure of its ionic or covalent character, which does not appear to change significantly across the range of electron demand.

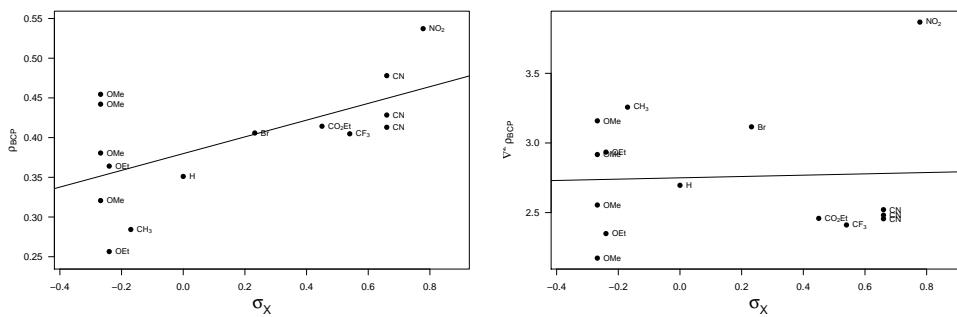


FIGURE 4.10: Hammett plots of QTAIM parameters ρ_{BCP} and $\nabla^2 \rho_{\text{BCP}}$ of ebselen derivatives complexed with DMAP. The line is described by the equation $\rho_{\text{BCP}} = (0.379(16) + 0.105(37) \times \sigma_p) \text{ e}/\text{\AA}^3$ ($R^2 = 0.3783$) and $\nabla^2 \rho_{\text{BCP}} = (2.74(13) + 0.05(29) \times \sigma_p) \text{ e}/\text{\AA}^5$ ($R^2 = 0.002$).

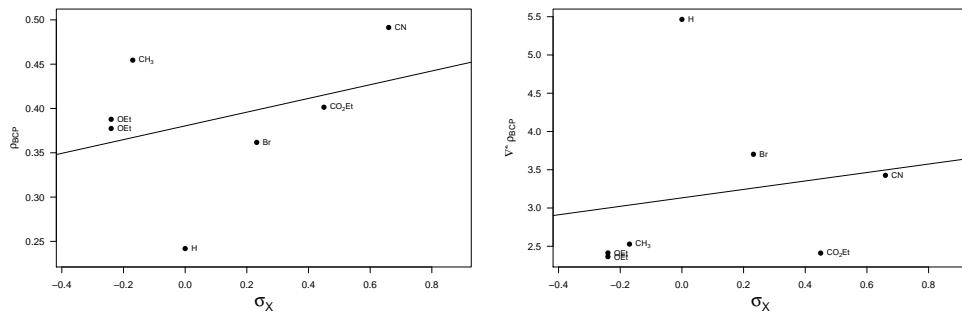


FIGURE 4.11: Hammett plots of QTAIM parameters ρ_{BCP} and $\nabla^2 \rho_{BCP}$ of ebselen derivatives complexed with **4a**. The line is described by the equation $\rho_{BCP} = (0.380(32) + 0.08(9) \times \sigma_p) \text{ e}/\text{\AA}^3$ ($R^2 = 0.123$) and $\nabla^2 \rho_{BCP} = (3.13(49) + 0.6(1.4) \times \sigma_p) \text{ e}/\text{\AA}^5$ ($R^2 = 0.0299$)

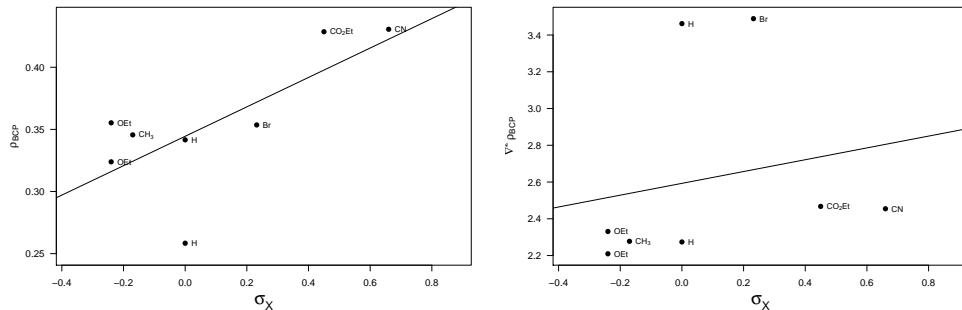


FIGURE 4.12: Hammett plots of QTAIM parameters ρ_{BCP} and $\nabla^2 \rho_{BCP}$ of ebselen derivatives complexed with **4b**. The line is described by the equation $\rho_{BCP} = (0.344(16) + 0.12(5) \times \sigma_p) \text{ e}/\text{\AA}^3$ ($R^2 = 0.5013$) and $\nabla^2 \rho_{BCP} = (2.59(21) + 0.3(6) \times \sigma_p) \text{ e}/\text{\AA}^5$ ($R^2 = 0.0400$)

4.2.1.1 Co-crystals where $Z' = 2$

DFT calculations show that the vibrational mode associated with Ch-bond stretching is found at very low energy. The force constant is 5–10 $\mu\text{dyne}\cdot\text{\AA}^{-1}$, which means that the energetic penalty associated with a 0.18 Å deformation (the difference between the shortest and longest Ch-bond length in **10Me**·DMAP) is only 0.02–0.03 kcal·mol⁻¹. Crystal packing forces (the sum of weak interactions such as C–H/ π , π/π and C–H/O interactions) are commonly accepted to be in the range of 1–2 kcal·mol⁻¹, so it is perhaps not surprising that the Ch-bond is deformed by the crystal environment.⁹

That said, there are no obvious differences between the two Ch-bond environments in any of the crystals that display this effect. We performed a solid state IR experiment to probe the crystalline environment surrounding the carbonyl, which is dependent on the Ch-bond environment due to conjugation through the amidic nitrogen (fig. 4.13).

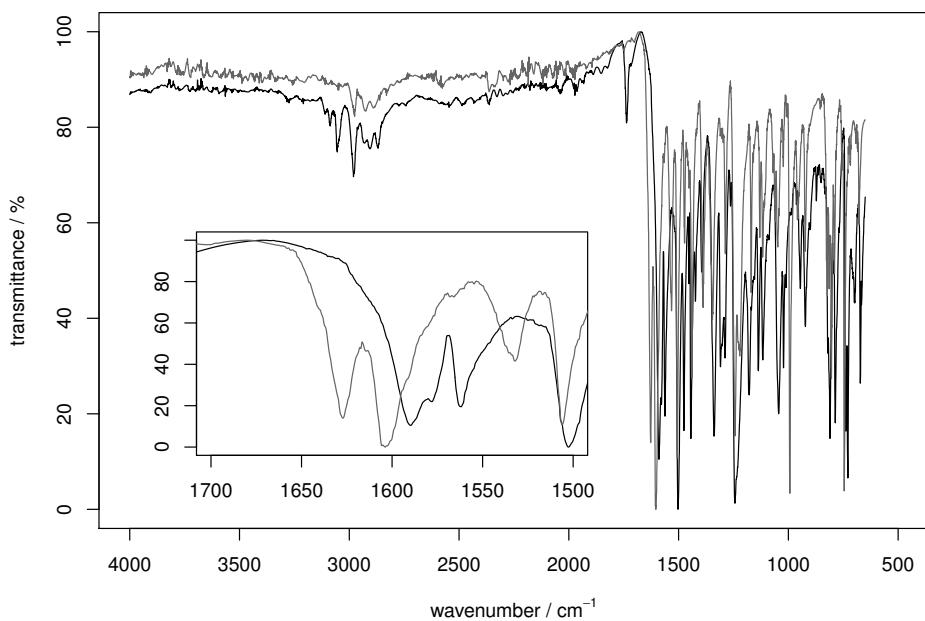


FIGURE 4.13: Solid state IR spectrum of **10Et·DMAP**. The spectrum of the pure **10Et** is shown in gray, which contains only one molecule in the asymmetric unit.

In the pure compound the carbonyl peak is found at approximately 1590 cm^{-1} and is relatively sharp and well defined. In the co-crystal, we observe the carbonyl peak at higher wave number (1610 cm^{-1}), due to the increased double bond character, as the π system and oxygen lone pair are no longer involved in the Ch-bond. This effect ostensibly outweighs the *decreased* double bond character caused by the shortened Ch-bond formed between the pyridyl nitrogen and selenium (see fig. 4.1 for a depiction of the decreased carbonyl double bond character).

If there are truly two Ch-bonded environments, we would expect to see splitting or at least broadening of the carbonyl signal in the IR spectrum of the co-crystal. This

is not the case, meaning that either the difference is too small to be seen in the spectrum, or the two environments are actually the same, and the measured differences are a crystallographic artefact, either a manifestation of missed symmetry, disorder, or a doubled cell.

However, we do not believe this is likely, for two reasons. Firstly, the data was of extremely high quality, and no alerts were raised in the ADDSYM routine of PLATON. Secondly, a refinement was conducted with a tight SADI restraint on the Ch-bonds ($\sigma = 0.0001$), which forced them to adopt the same length of approximately 1.949 Å. This increased the R-factor by 1.2%, and significant residual density was visible where the pyridyl nitrogen had been displaced. Furthermore, removal of the restraint recovered the original model, ruling out the possibility of a false minimum.

Solid state NMR of the co-crystal was also used to investigate the crystalline environment. A sample of **1OMe**·DMAP was first characterised by single crystal x-ray diffraction, which confirmed the polymorph and structural parameters (fig. 4.14).

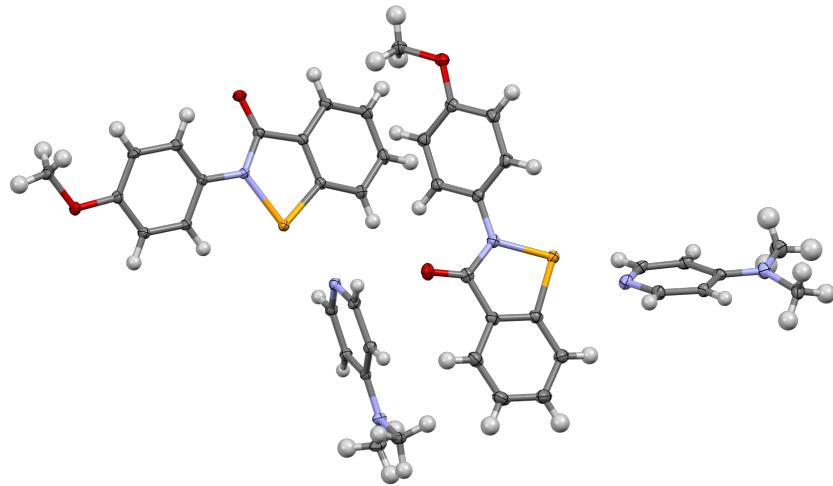


FIGURE 4.14: Single crystal x-ray structure of **1OMe**·DMAP, displaying the two systems in the asymmetric unit.

The bulk material was then crushed and homogenised, and characterised by powder x-ray diffraction. The measured powder pattern was in excellent agreement with

the pattern calculated from the single crystal data (fig. 4.15), indicating good phase purity.

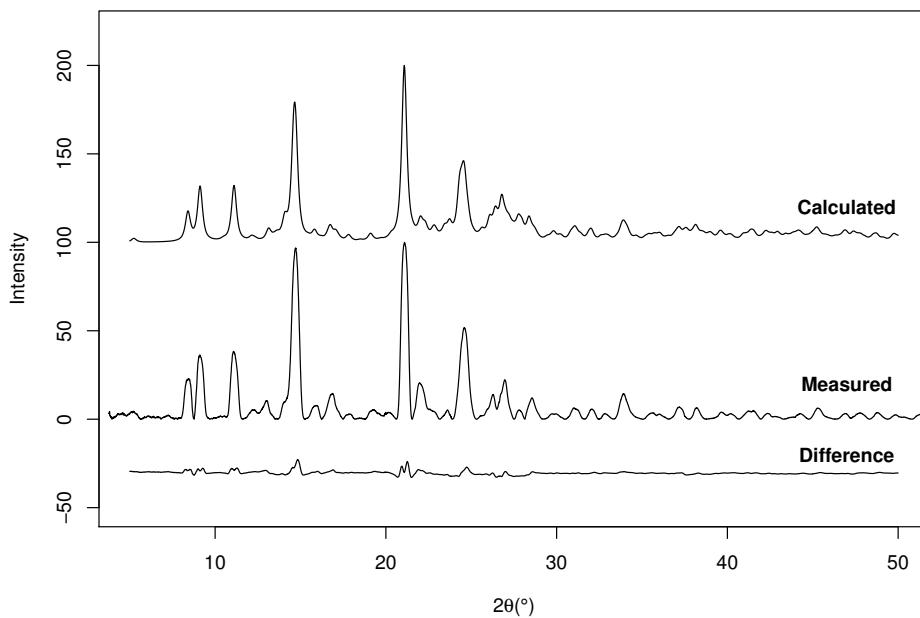


FIGURE 4.15: Calculated vs measured powder diffraction pattern for **10Me**·DMAP.

Spectra were then acquired using a 400 MHz instrument and triple resonance MAS room temperature probe tuned to ^1H , ^{13}C and ^{77}Se . ^1H - ^{13}C or ^1H - ^{77}Se cross polarisation was used for signal enhancement, and a spin frequency of 10 kHz was used in most cases.

CDCl_3 solution spectra of the complex were also obtained on a 500 MHz instrument. The ^1H and ^1H - ^{13}C HSQC spectra (fig. 4.16) were used to unambiguously assign the ^{13}C spectrum.

The aromatic region of the solid state ^{13}C spectrum is shown in fig. 4.17, overlaid with the corresponding solution spectrum*. Good agreement is observed for all signals, with some interesting phenomena visible in the solid state spectrum. Firstly, the signal at 138.62 ppm corresponding to C8 is split into a 1:1:1 triplet, possibly

*The ^{13}C spectrum was referenced to adamantine at $\delta\text{CH} = 29.45$ ppm and $\delta\text{CH}_2 = 38.48$ ppm.¹⁰

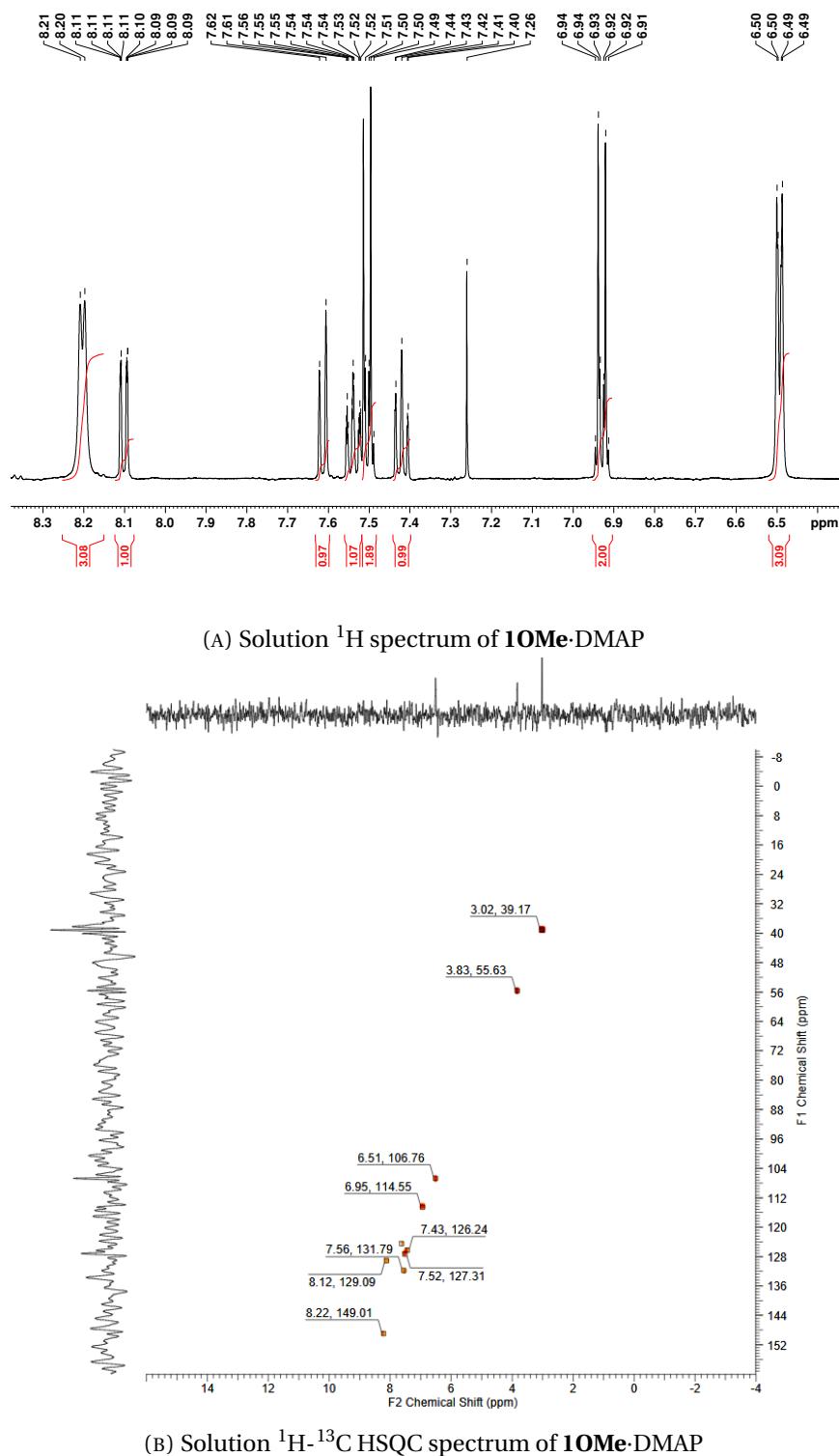


FIGURE 4.16

due to coupling to the spin 1 ^{14}N nucleus adjacent. However this is not observed for the C7 signal, nor any signals in the pyridyl ring. Secondly, shoulders can be seen on the C15/C19 and C16/C18 signals. This indicates that the crystalline environment surrounding each DMAP is different, which provides further evidence that there are indeed two systems in the asymmetric unit. The relatively poor resolution of the solid state ^{13}C NMR spectrum limits further analysis, particularly of the C1 signal which is obscured by several other signals.

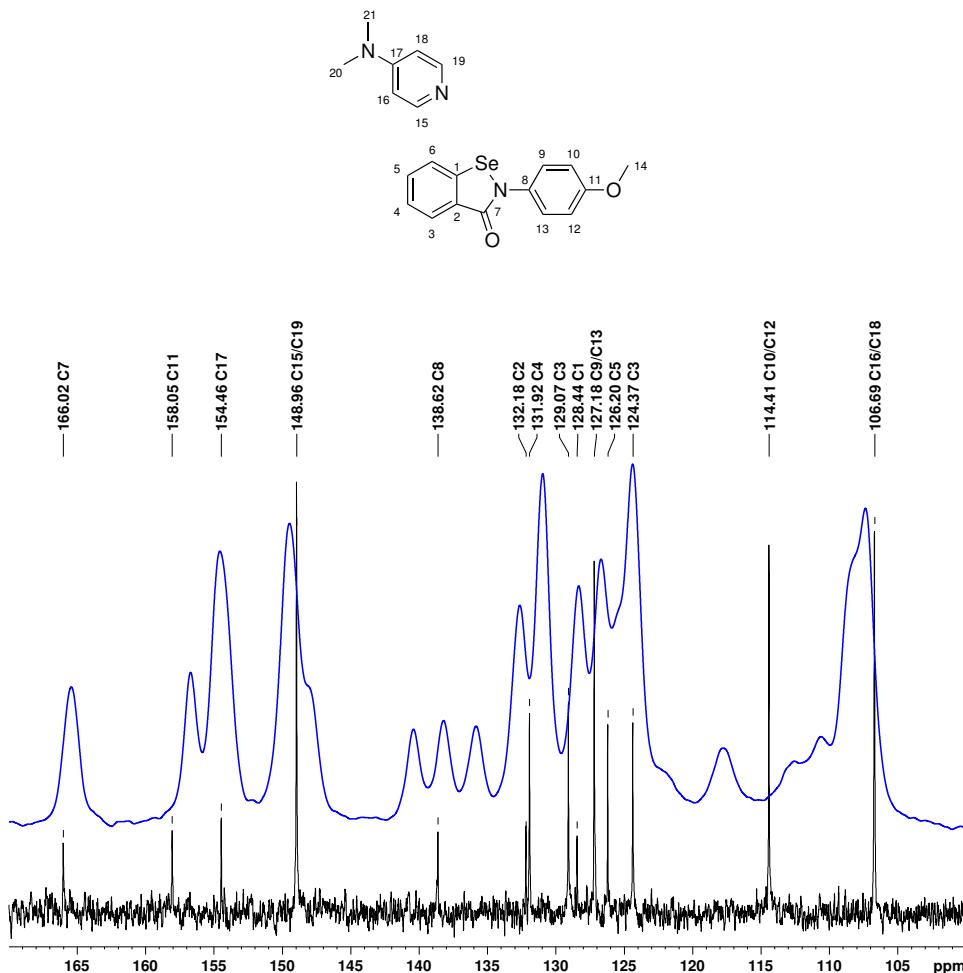


FIGURE 4.17: Solid state ^{13}C -NMR spectrum of **10Me-DMAP** (blue) overlaid on solution spectrum of the same (black). Signals are assigned according to the numbering scheme above.

To conclusively demonstrate that there are two systems in the asymmetric unit

we conducted a solid state ^{77}Se NMR experiment, which is shown in fig. 4.18.* In solution, the ^{77}Se resonance is found around 900 ppm relative to dimethylselenide ($\delta = 0$ ppm), and appears as one singlet due to the averaging of all environments. In the solid phase, the spectrum is significantly more complex, primarily due to chemical shift anisotropy which manifests as spinning sidebands. The true anisotropic chemical shifts can only be discerned by varying the MAS spinning speed, which changes the spacing of the sidebands while leaving the parent signals in the same place. Spinning at 12 kHz instead of 10 kHz showed that the signals at 834.69 and 867.02 ppm are the true isotropic chemical shifts, and the fact that there are two signals show that there are indeed two Ch-bonded systems in the asymmetric unit.

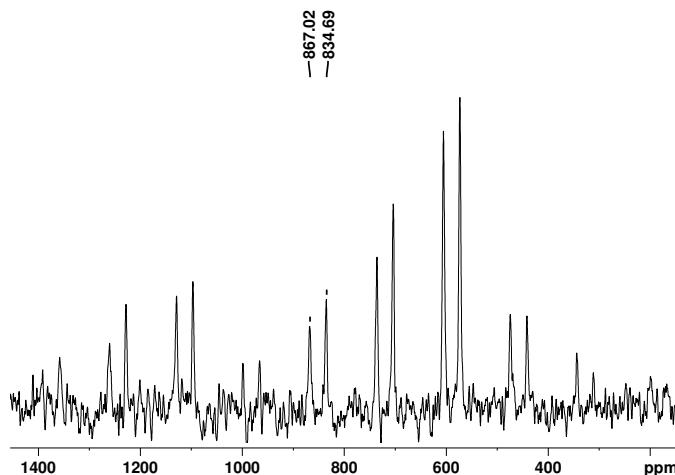


FIGURE 4.18: Solid state ^{77}Se -NMR spectrum of **1OMe**·DMAP. The primary resonances are visible at 834.69 and 867.02 ppm. The remaining peaks are spinning sidebands, and are separated from the parent signals by multiples of 10 kHz (the magic angle spinning frequency).

4.2.2 Measurement of chemical shift anisotropy

We acquired solid state spectra of three complexes spanning the range of electron demand, **1OMe**·DMAP, **1**·DMAP, and **1NO₂**·DMAP. The resulting spectra are presented in fig. 4.19, and the extracted principal values of the chemical shift tensor

*The ^{77}Se spectrum was referenced to diphenyl diselenide at $\delta = 463$ ppm.

are presented in table 4.4.

1OMe·DMAP differs from the other structures, in that it crystallises in the space group $P2_1/c$, whereas **1**·DMAP and **1NO₂**·DMAP both crystallise in $P\bar{1}$. The 2_1 screw axis in the former crystal is oriented such that it generates a symmetry equivalent molecule which is rotated by an angle of about 45°. This means that the observed chemical shift tensor is the average of these two symmetry related orientations, further complicating the analysis. Fundamentally this is due to the fact that an ellipsoid does not have twofold rotational symmetry, except about its principal axes. The triclinic complexes **1**·DMAP and **1NO₂**·DMAP do not suffer from this issue, as the inversion symmetry operation preserves the shape of the tensor. This can be seen in the latter two principal values of the chemical shift tensor in table 4.4, which have the same value (within experimental error), describing a cigar-shaped tensor.

TABLE 4.4: Principal values of the chemical shift tensor extracted from powder spectra in fig. 4.19.

Complex	δ_{iso}	IUPAC			Haeberlen		Herzfeld-Berger	
		δ_{11}	δ_{22}	δ_{33}	Δ_{CSA}	η_{CSA}	ω	κ
1OMe ·DMAP*	835.2	1572.58	466.52	466.50	724.69	0.003	1088.11	-1.0
1OMe ·DMAP†	866.9	1628.91	485.95	485.92	759.35	-0.003	1140.06	-1.0
1 ·DMAP	864.5	1616.74	596.21	380.64	752.21	-0.287	1236.09	-0.65
1NO₂ ·DMAP	860.8	1596.92	547.82	437.66	736.12	-0.15	1159.27	-0.81

4.2.3 Calculation of chemical shift anisotropy

In order to verify our results, we also *calculated* the chemical shielding tensors in order to compare the principal values, and also determine the likely orientation of the tensors with respect to the rest of the molecule (fig. 4.20a). These calculations were conducted at the ω B97X-D/def2-TZVP level, using the GIAO method of calculating shielding tensors.^{11,12} The Gauge Independent Atomic Orbital (GIAO)

*Site a

†Site b

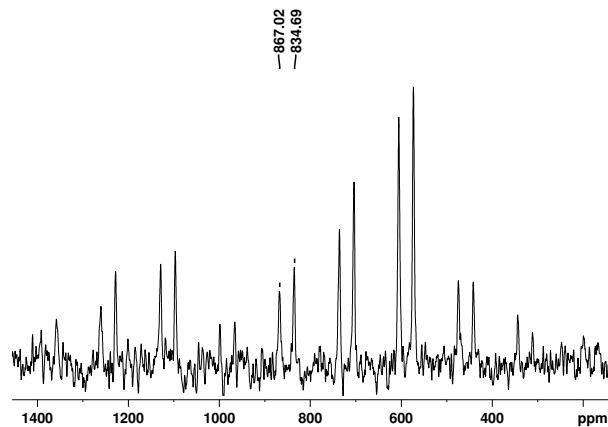
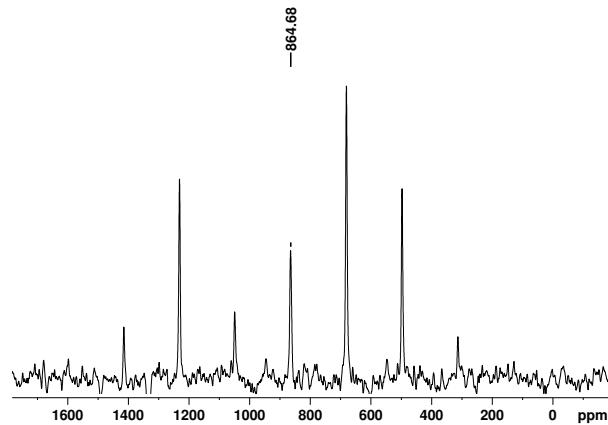
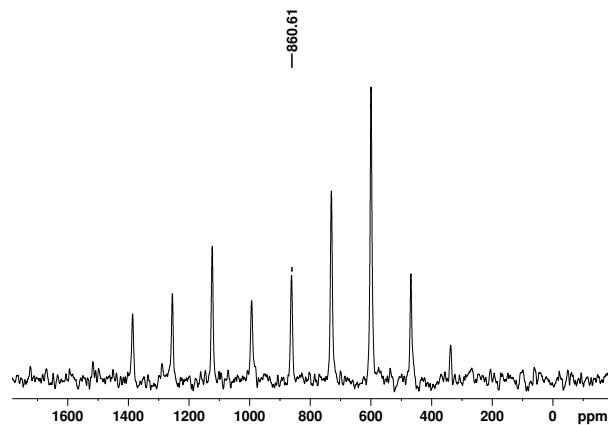
(A) ⁷⁷Se CPMAS NMR spectrum of **1OMe**·DMAP.(B) ⁷⁷Se CPMAS NMR spectrum of **1**·DMAP.(C) ⁷⁷Se CPMAS NMR spectrum of **1NO₂**·DMAP.

FIGURE 4.19

overcomes the so-called gauge problem in evaluating perturbations in the wave function due to a magnetic field. For an atomic orbital described by a finite basis set, the perturbation is proportional to $B_z \hat{l}_z$, where \hat{l}_z is the z component of the angular momentum operator which is dependent on the distance to the origin of the coordinate system. This is obviously unphysical, and typical basis sets are not large enough to accommodate these non-atom centred perturbations. The gauge problem is overcome in GIAO by transforming the atomic orbitals using a complex gauge factor which is dependent on the magnetic field and the location of the basis function relative to the origin. This affords chemical shielding tensors using reasonably sized basis sets. In order to convert these chemical shielding tensors σ into chemical shifts δ , we must reference them to a standard σ_{ref} .

$$\delta = \sigma_{\text{ref}} - \sigma_{\text{sample}} \quad (4.1)$$

Convention dictates that for ^{77}Se , dimethylselenide is assigned a chemical shift of 0 ppm. The structure of dimethylselenide was therefore optimised and shielding tensors calculated at the same level.

The tensors (in the computation reference frame) were found to be

$$\sigma_{\text{Me}_2\text{Se}} = \begin{bmatrix} 2276.3834 & -0.0073 & -0.0175 \\ -0.0159 & 1579.1094 & 0.0349 \\ -0.0622 & -0.0041 & 1583.4669 \end{bmatrix}$$

$$\sigma_1 = \begin{bmatrix} 1086.4346 & 929.1822 & 929.1822 \\ 862.9168 & 514.6826 & -87.328 \\ 105.4202 & -92.4104 & 1101.0194 \end{bmatrix}$$

$$\sigma_{\mathbf{1}\text{OMe}\cdot\text{DMAP}} = \begin{bmatrix} 1686.2267 & 105.2547 & -3.2274 \\ 31.1129 & 187.9487 & 48.4737 \\ -19.1247 & 50.3984 & 1081.5651 \end{bmatrix}$$

$$\sigma_{\mathbf{1}\cdot\text{DMAP}} = \begin{bmatrix} 1595.7999 & 367.4957 & 6.7322 \\ 322.6389 & 291.3611 & 10.8588 \\ -19.9165 & -35.8017 & 1069.2648 \end{bmatrix}$$

$$\sigma_{\mathbf{1}\text{NO}_2\cdot\text{DMAP}} = \begin{bmatrix} 1671.4068 & -76.2797 & 20.6203 \\ -100.0242 & 221.8577 & -17.9295 \\ -44.1025 & -120.0346 & 1055.0806 \end{bmatrix}$$

$$\sigma_{\mathbf{1}\cdot\text{DMF}} = \begin{bmatrix} 1642.3856 & -138.5717 & -175.547 \\ -111.063 & 175.617 & -253.364 \\ -215.991 & -275.178 & 1028.493 \end{bmatrix}$$

affording reference frame independent principal values in table 4.5.

These were converted to chemical shifts using eq. (4.1), which are presented in table 4.6.

These values are encouragingly close to those derived from the fitting of the experimental MAS line shape, with the exception of **1OMe**·DMAP, for reasons explained above.

TABLE 4.5: Principal values of the chemical shielding tensor calculated from optimised structures.

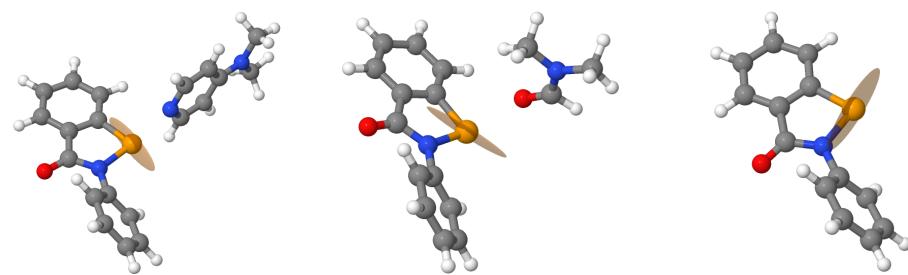
Complex	σ_{iso}	σ_{11}	σ_{22}	σ_{33}
Me ₂ Se	1812.9866	1579.1093	1583.4670	2276.3834
1	900.7122	-156.9566	1114.4138	1744.6794
1OMe ·DMAP	985.2468	182.0846	1084.2014	1689.4545
1 ·DMAP	985.4753	205.5761	1069.2485	1681.6013
1NO₂ ·DMAP	982.7817	210.7844	1060.7206	1676.8400
1 ·DMF	948.8319	80.7668	1064.7403	1700.9887

TABLE 4.6: Principal values of the chemical shift tensor calculated from optimised structures.

Complex	δ_{iso}	δ_{11}	δ_{22}	δ_{33}
1	912.2744	1969.9432	698.5728	68.3072
1OMe ·DMAP	827.7398	1630.9020	728.7852	123.5321
1 ·DMAP	827.5113	1607.4105	743.7381	131.3853
1NO₂ ·DMAP	830.2049	1602.2022	752.2660	136.1466
1 ·DMF	864.1547	1732.2198	748.2463	111.9979

Plotting the resulting tensors as ellipsoids reveals aspects of the electron density and hence shielding around the selenium (fig. 4.20a). A major contributor to the tensor is of course the aromatic ring current, which leads to deshielding perpendicular to the plane of the ring. There is also a ring current in the pyridine ring, which deshields the selenium roughly in the direction of the Se–C bond (perpendicular to the plane of the pyridine ring). This accounts for the large δ_{11} and δ_{22} components of the chemical shift. The remainder is presumably due to diamagnetic shielding from the electron density. In this we are able to see deformation of the electron cloud due to the lone pairs and the Ch-bond. The δ_{33} component is substantially smaller than δ_{11} and δ_{22} , and is aligned with the Ch-bond, reflecting a very strong shielding in this direction. This clearly shows the contribution of the lone pair of the pyridyl nitrogen to the electron density around the selenium, and hence the strength of the Ch-bond.

This can be contrasted to the chemical shift of the non-complexed selenium, which



(A) Calculated chemical shift tensor for **1**·DMAP. (B) Calculated chemical shift tensor for **1**·DMF. (C) Calculated chemical shift tensor for **1**.

FIGURE 4.20

is shown in fig. 4.20c. In the absence of the lone pair of the pyridyl nitrogen, the σ -hole strongly deshields the selenium, therefore the large δ_{11} component is roughly aligned with it. There is also no deshielding ring current from the pyridine, so the δ_{33} (which is aligned with the Se–C bond) is very small. The δ_{22} component perpendicular to the plane of the benzisoselenazolinone ring is roughly the same, as there is no significant change in either electron density or ring currents in this direction.

To demonstrate that the δ_{11} component in the **1**·DMAP complex is not entirely due to the pyridine ring current, a **1**·DMF complex was optimised and the chemical shift tensor calculated. This is shown in fig. 4.20b. The largest component of the tensor is still approximately aligned with the Se–C bond, even in the absence of the aromatic base.

These results must be taken with a grain of salt, as the chemical shifts are calculated in the gas phase (i.e. in the absence of crystal packing). This is particularly important for the chemical shift of the non-complexed heterocycle, as such a “free” σ -hole would not be observed in any condensed phase (it is always filled by solvent in solution, or the carbonyl of an adjacent molecule in the solid). Nonetheless we may draw comparisons with this caveat in mind.

4.2.3.1 Measurement of CSA in a single crystal

The calculated chemical shift tensors were useful in the absence of experimental data, and the fact that the principal values of the tensors matched the powder data so well was encouraging. However, we wished to verify the orientation of the tensor by an experimental method.

We were fortunate to obtain a single crystal of **1**·DMAP of sufficient size (approx $10 \times 3 \times 1$ mm, fig. 4.21) for a single crystal SS-NMR experiment, allowing us to measure the orientation of the tensor in that crystal for comparison with our computational results.*

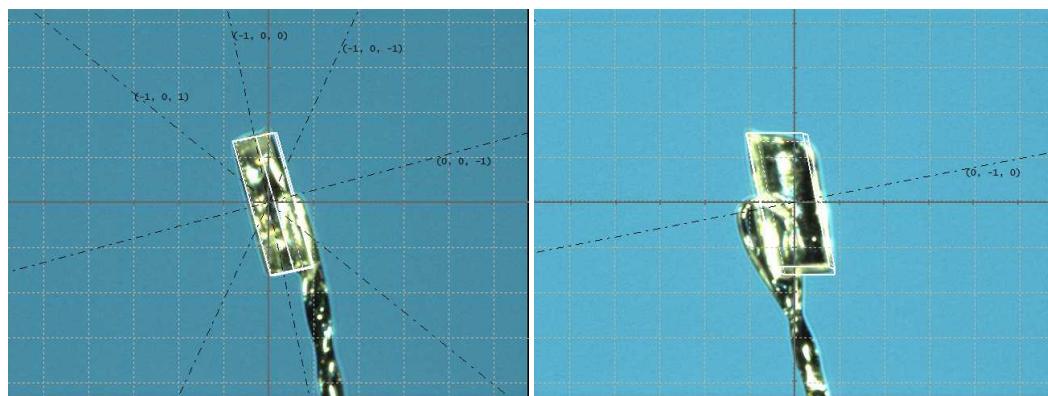


FIGURE 4.21: Single crystal of **1**·DMAP used for SS-NMR.

In order to do this, the faces of the crystal had to be indexed to the internal structure, which was done by x-ray diffraction. As the crystal was far too big to mount on the diffractometer, two faces were marked with different coloured pens to preserve the relation to the large crystal, then a small fragment was removed from the crystal. This was mounted, and a short data collection afforded an indexable pattern. The resulting planes are shown in fig. 4.22.

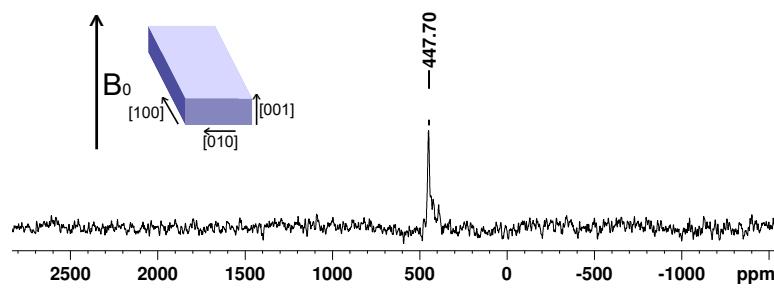
A number of these large crystals were visually indexed based on their morphology, and packed in a rectangular sample holder. Spectra were acquired on a 600 MHz instrument using a Hahn echo pulse sequence. This was necessary to preserve the

*Unfortunately no other derivatives formed crystals of sufficient size or morphology for the SS-NMR experiment.

FIGURE 4.22: Indexed faces of **1**·DMAP.

extremely weak signal by isolating it from the excitation pulse in the time domain. The spectra were referenced to a saturated solution of diphenyl diselenide in CDCl_3 at 463 ppm, which was contained in a zirconia MAS rotor inside the sample holder.¹³

A spectrum was collected with the crystals lying flat relative to the magnetic field, and the resulting spectrum is shown in fig. 4.23. The selenium nucleus in this orientation resonates at 448 ppm.

FIGURE 4.23: ^{77}Se NMR spectrum of a single crystal of **1**·DMAP in the orientation depicted in the inset.

The sample holder was then re-packed with the crystals now rotated 90° to their original orientation, and a spectrum was again acquired. This is shown in fig. 4.24, where the selenium now resonates at 567 ppm.

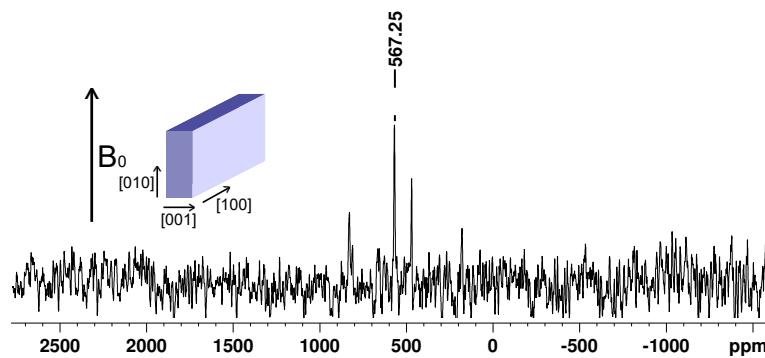


FIGURE 4.24: ^{77}Se NMR spectrum of a single crystal of **1**·DMAP in the orientation depicted in the inset. The smaller satellite signals are due to slightly misaligned crystals in the sample holder. This also explains the poorer signal to noise ratio of the main peak.

In order to orient the tensor in the laboratory reference frame, we must determine the declination and azimuth angles θ and ϕ . This is done by writing an expression which relates these angles, the principal values (determined from the powder experiment), and the observed chemical shift. The rotation of the tensor into the lab reference frame is modelled by a simple rotation matrix of the two polar angles:

$$\mathbf{Q} = \begin{bmatrix} \cos\theta & 0 & \sin\theta \\ \sin\theta \sin\phi & \cos\phi & -\cos\theta \sin\phi \\ -\sin\theta \cos\phi & \sin\phi & \cos\theta \cos\phi \end{bmatrix} \quad (4.2)$$

The 90° rotation about the a axis is modelled by a similar rotation matrix which, for generality, is derived from the Rodrigues rotation formula describing a rotation of

angle ψ about a unit vector $\hat{\omega} = (\omega_x, \omega_y, \omega_z)$:

$$\mathbf{R}_{\hat{\omega}}(\psi) = \mathbf{I} + \tilde{\omega} \sin \psi + \tilde{\omega}^2 (1 - \cos \psi) \quad (4.3)$$

where $\tilde{\omega}$ is the antisymmetric matrix:

$$\tilde{\omega} = \begin{bmatrix} 0 & -\hat{\omega}_z & \hat{\omega}_y \\ \hat{\omega}_z & 0 & -\hat{\omega}_x \\ -\hat{\omega}_y & \hat{\omega}_x & 0 \end{bmatrix} \quad (4.4)$$

and \mathbf{I} denotes the 3×3 identity matrix. This notation is particularly convenient for crystallography, as the unit vector for a rotation is easily generated from the Miller indices, which generally correspond to a macroscopic face of a crystal. In this case the rotation axis was simply $\hat{\omega} = (1, 0, 0)$ i.e. the a axis.

Sequentially applying these two rotations allows us to represent the tensor in any orientation. The projection of the tensor onto the Z axis (the magnetic field B_0), which corresponds to the observed chemical shift in any given orientation, is given by the δ_{ZZ} component of the tensor. We therefore simply optimise the function

$$\delta = \mathbf{RQ} \begin{bmatrix} 1616.74 & 0 & 0 \\ 0 & 596.21 & 0 \\ 0 & 0 & 380.64 \end{bmatrix} \mathbf{Q}^{-1} \mathbf{R}^{-1} \quad (4.5)$$

to reproduce the observed δ_{ZZ} components for $\psi = 0^\circ$ and $\psi = 90^\circ$. The polar angles thus calculated are equal to $\theta = 79.9^\circ$ and $\phi = 23.8^\circ$.

We can calculate the chemical shift tensor using the diagonal components from the powder experiment and the rotation matrix from the polar angles which affords

$$\delta = \begin{bmatrix} 1578.72 & -86.12 & 195.26 \\ -86.12 & 567.29 & 65.55 \\ 195.26 & 65.55 & 447.56 \end{bmatrix} \quad (4.6)$$

This can be plotted within the unit cell of the crystal, where we observe an excellent qualitative agreement with the calculated tensor (fig. 4.25). The most deshielded axis is aligned with the Se–C bond, and the most shielded axis is aligned with the Ch-bond, as in the calculated tensor. This verifies that the tensor is oriented correctly in the crystal, and that the observed shielding is due to the approach of the lone pair into the σ -hole.

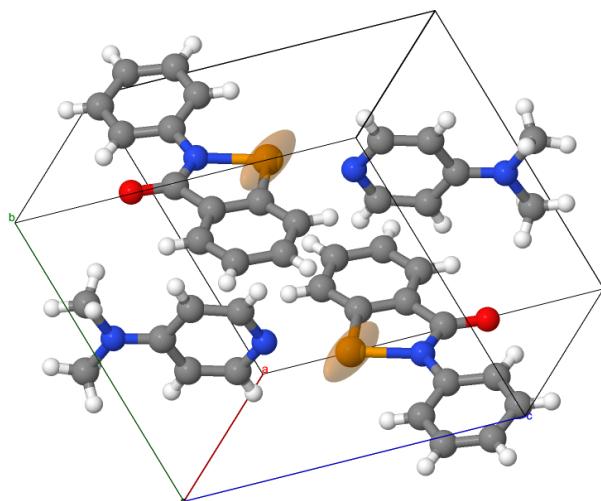


FIGURE 4.25: Experimentally determined chemical shift tensor for **1**·DMAP. Compare with the calculated tensor in fig. 4.20a.

To briefly summarise, Ch-bonding induces enormous chemical shift anisotropy at the selenium, which is highly sensitive to the approach of the Lewis base and can be measured using both powder and single crystal SS-NMR. While the technique is perhaps not as sensitive as diffraction based methods, the great advantage lies in the fact that SS-NMR does not require crystallinity, even at the nano scale. This means that amorphous species such as polymers may be studies, as long as they contain an NMR active nucleus.

4.2.4 Solution-phase studies

While the above crystallographic analysis gives a useful qualitative understanding of the strength of the Ch-bond, we sought an experimental method that could accurately determine bond energies, so that we could easily compare our Ch-bonded complexes with those dominated by other interactions.

4.2.4.1 NMR titration experiments

NMR titrations are a useful tool for the determination of binding affinities in supramolecular and host-guest chemistry.^{2,14–16} ^1H and ^{19}F NMR have both been used to study halogen bonded systems, however we decided to take advantage of the unique NMR characteristics of the ^{77}Se nucleus to probe the Ch-bond in our systems for the following reasons:

- the nucleus has an extremely wide chemical shift range ($-1000\text{--}2000$ ppm),
- the chemical shift is very sensitive to the electronic environment around the selenium,
- the selenium is at the heart of the interaction, so any electronic changes should manifest clearly,
- the spectrum (for our compounds) is extremely simple, featuring only one singlet,
- the experiment is moderately sensitive (slightly more sensitive than ^{13}C).

We devised a titration experiment, where a solution of a Lewis base is gradually added to a solution of the Ch-bond donor, and the chemical shift of the selenium is measured at the various concentrations of base. As the concentration of base increases, a greater proportion of the selenium species will be in a Ch-bonded

environment, with an associated increase in electron density at the selenium due to the coordinated base. This will lead to an upfield shift of the selenium resonance.

It is important to note that even in the absence of a Lewis base, the organoselenium species will still likely feature a Ch-bond to the carbonyl oxygen of another molecule. This is substantially weaker than a Se...N interaction, but non-negligible. We must therefore view these as *competition* experiments, rather than an absolute measure of binding energy.

For single site binding (a valid approximation for these systems, as the single σ -hole is likely to out-compete all other interactions), the dissociation constant can be expressed as

$$K_d = \frac{[\text{ebs}][\text{base}]}{[\text{ebs} \cdot \text{base}]} \quad (4.7)$$

If the Ch-bond formation/breaking is slow on the NMR timescale, two distinct resonances will be observed that correspond to the “free” (Ch-bonded to a carbonyl oxygen) and “bound” (Ch-bonded to the Lewis base) selenium species, and their relative concentrations can be determined by integration of the signals. As it happens, this is not the case, and the process is fast relative to the NMR timescale. The observed chemical shift is therefore the mole fraction ($f_{\text{ebs}} = [\text{ebs}] / [\text{ebs}]_0$ and $f_{\text{ebs}\cdot\text{base}} = [\text{ebs} \cdot \text{base}] / [\text{ebs}]_0$) weighted average of the chemical shifts of the two species:

$$\delta_{\text{observed}} = \delta_{\text{ebs}} f_{\text{ebs}} + \delta_{\text{ebs}\cdot\text{base}} f_{\text{ebs}\cdot\text{base}} \quad (4.8)$$

If we consider only the *change* in chemical shift from the free species, this becomes simply:

$$\Delta(\delta_{\text{observed}}) = \Delta(\delta_{\text{ebs}\cdot\text{base}}) f_{\text{ebs}\cdot\text{base}} \quad (4.9)$$

$$= \Delta(\delta_{\text{ebs}\cdot\text{base}}) \left(\frac{[\text{ebs} \cdot \text{base}]}{[\text{ebs}]_0} \right) \quad (4.10)$$

From here we can rearrange the equilibrium expression (eq. 4.7) and mass balance equation ($[ebs]_0 = [ebs] + [ebs \cdot base]$), and substitute them in to arrive at the generic binding isotherm equation:

$$\Delta(\delta) = \frac{\Delta(\delta_{ebs \cdot base}) \times [base]}{K_d + [base]} \quad (4.11)$$

This assumes that there is insignificant depletion of the base concentration due to complexation.¹⁷ Such an assumption may not be entirely valid for this situation, which may explain the imperfect fitting. However, the analysis is considerably simplified, and adequate standard deviations are obtained using this method.

The resulting K_d values can be converted to free energies:

$$\Delta G = -RT \ln K_d \quad (4.12)$$

A saturated solution of the organoselenium derivative in chloroform was used, due to the high solubility (to reduce acquisition time) and non-coordinating nature (to minimise Ch-bonding to the solvent). This was spiked with a small amount of deuteriochloroform for the lock signal. Spectra were acquired on a 500 MHz instrument, using a 60° pulse and 1 s relaxation delay, until an unambiguous ^{77}Se resonance was observed. The resulting chemical shifts were then tabulated and plotted against the concentration of the Lewis base. These are shown in fig. 4.26.

Non-linear regression analysis was performed with the `nls` function in the R software package using the relationship derived above, and the calculated values are given in table 4.7.¹⁸

Although there is insufficient data to derive clear trends from a Hammett relationship, we can nevertheless visualise the data in a plot, which shows that the Ch-bond gets appreciably stronger with more electron withdrawing substituents (fig. 4.27).

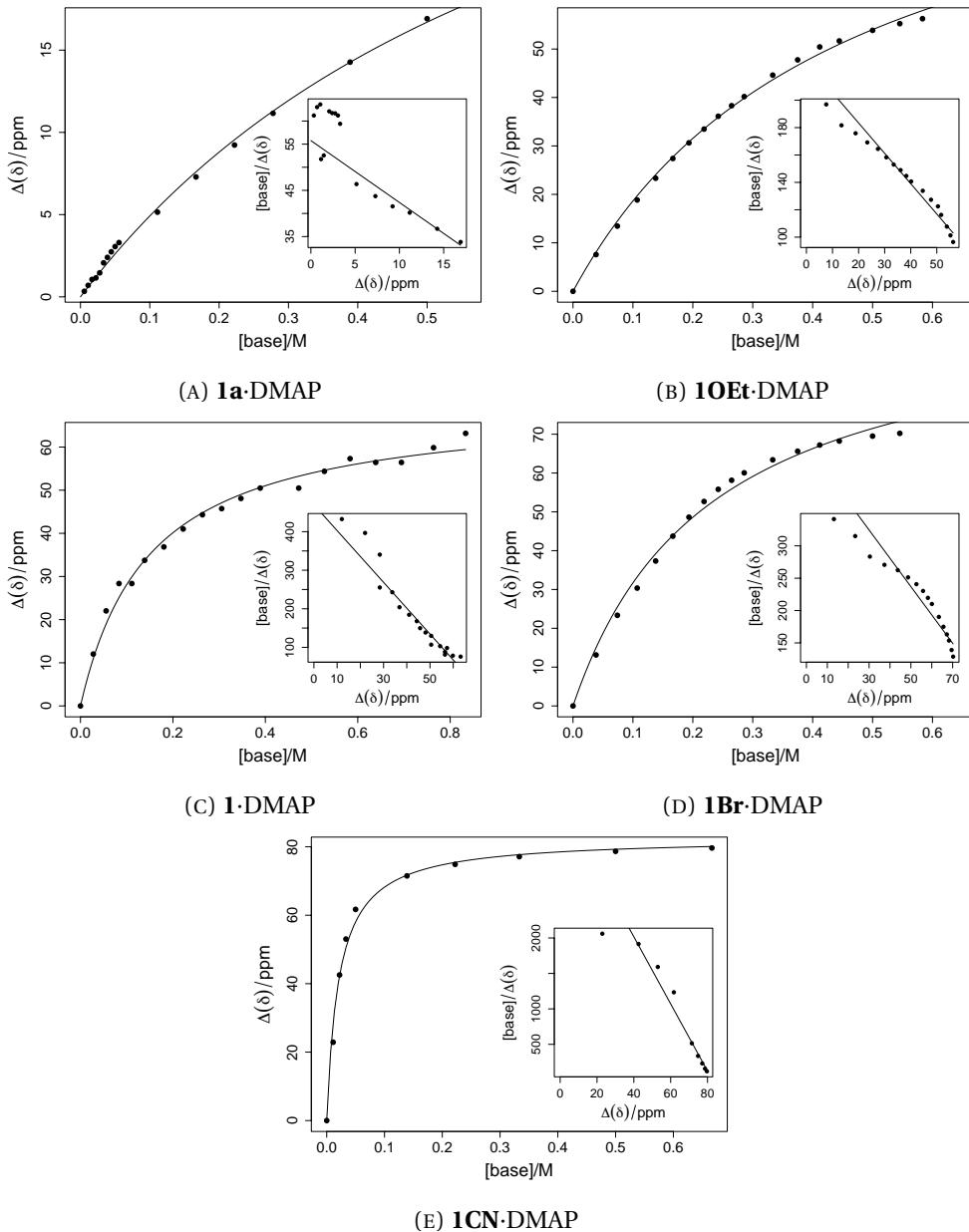


FIGURE 4.26: Binding isotherms for ebselen derivatives **1a**, **1OEt**, **1**, **1Br** and **1CN** with DMAP. Scatchard plots are inset.

TABLE 4.7: Binding energies for complexes of **1a**, **1OEt**, **1Br**, **1** and **1CN** with DMAP, derived from ^{77}Se -NMR titration experiments.

Complex	Binding energy (kcal/mol)	$\Delta(\delta_{\text{ebs-base}})$ (ppm)
1a ·DMAP	0.17(6)	41.6
1OEt ·DMAP	0.47(3)	102.8
1Br ·DMAP	0.87(5)	104.2
1 ·DMAP	1.12(4)	70.05
1CN ·DMAP	2.28(5)	82.62

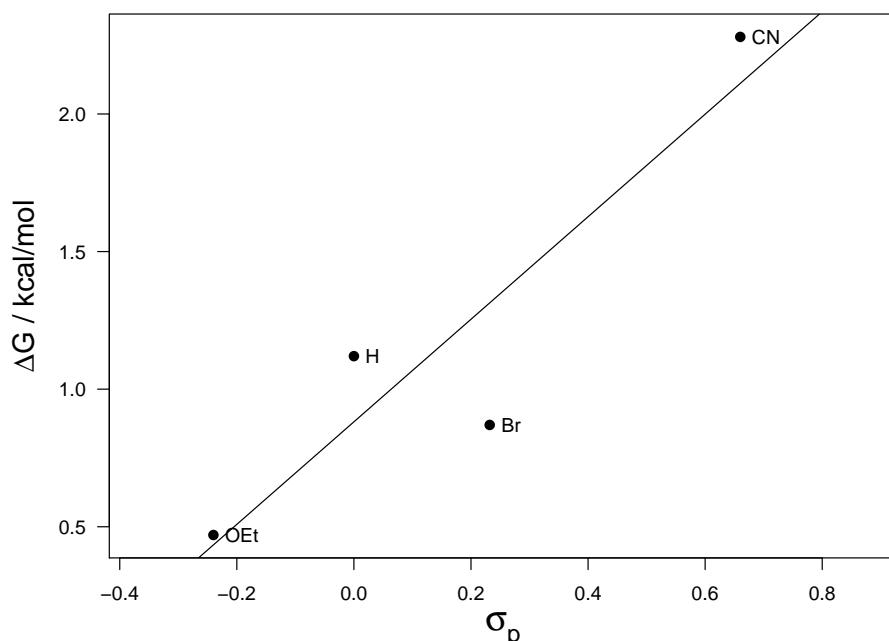


FIGURE 4.27: Hammett plot of binding energies between ebselen derivatives and DMAP, determined by ^{77}Se -NMR titration. The line is described by the equation $\Delta G = (0.88(21) + 1.86(57) \times \sigma_p) \text{ kcal} \cdot \text{mol}^{-1}$ ($R^2 = 0.8437$).

4.2.4.2 UV-Vis titration experiment

Although extremely useful, the NMR titration technique had a number of drawbacks. Firstly, relatively large amounts (~ 100 mg) of the Ch-bond donor were required. For simple systems such as these, this was not an issue, however we hoped to apply the technique to less synthetically accessible molecules, which may not be available

in such quantities. Secondly, the experiment is quite time consuming, with tens of minutes required per spectrum.

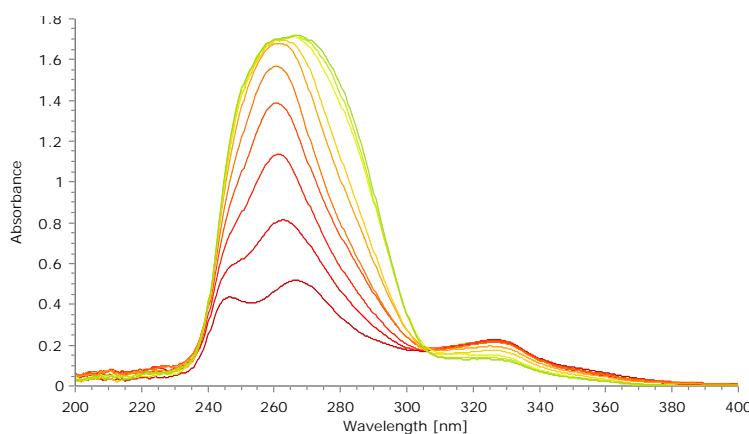


FIGURE 4.28: UV-Vis titration of **1** with DMAP. The pure solution of **1** is shown in red, with increasing concentrations of DMAP in green.

UV-Vis spectroscopy presented a possible solution to both of these issues, with spectra collected in seconds even with extremely dilute solutions. Interactions which have a charge-transfer component (such as Ch-bonding) can give rise to strong absorbances attributable to the $n \rightarrow \sigma^*$ transition.¹⁹ However, we were unable to observe any evidence of such absorbances in our systems (fig. 4.28). This may be due to the fact that these Ch-bonds are primarily electrostatic in origin, or because the aromatic systems are already strongly absorbing, thus obscuring the charge-transfer band.

4.2.5 Ch-bonding in the gas phase

To complete our trifecta of experiments, we sought evidence that supports Ch-bonding in the gas phase. Using an ESI source, LTQ ion trap and TOF detector, we intended to isolate an ion corresponding to the complex, and subsequently fragment it by collision induced dissociation (CID) to regain the original species. CID is a process by which ions are fragmented by accelerating them into inert gas particles (noble gases such as He and Ar are commonly used). The energy of

the collision can be controlled by varying the potential applied to accelerate the ions. A collision with a noble gas particle causes the kinetic energy of the ion to be converted into vibrational potential energy among the bonds of the ion. If sufficient energy is deposited in the ion, the weakest bonds will break, and product ions will be observed. Mass can also be lost as neutral molecules or radicals, which are not detected in the mass spectrometer due to the lack of charge.

We sought to break the Ch-bond of a Ch-bonded complex, and observe a fragment ion. Naturally, this requires that one (or both) of the components of the complex have a charge, so that it can be detected by the mass spectrometer. We initially injected an equimolar mixture of DMAP and ebselen **1** in methanol, and isolated an ion of m/z 398.08 a.m.u. However, this could be assigned to either the desired Ch-bonded complex, protonated at the carbonyl oxygen, or to a H-bonded complex.* CID of this ion exclusively gave an ion of m/z 123.09 a.m.u., corresponding to protonated DMAP (fig. 4.29). We interpreted this as evidence that we had formed the H-bonded complex, as migration of the proton on the timescale of a collision is unlikely, and the proton is most likely to remain with the more basic species in the H-bond. Furthermore, $\text{N}^+ - \text{H} \cdots \text{O}$ hydrogen bonds are very strong (in excess of 15 kcal/mol),²⁰ which is likely to out compete Ch-bonding, with estimated bond enthalpies of < 10 kcal/mol in the gas phase.

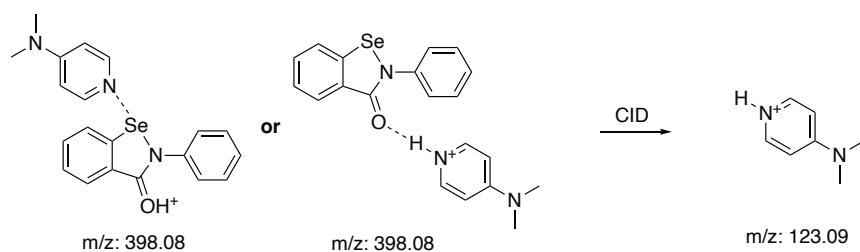


FIGURE 4.29: Dissociation of the complex **1**·DMAP·H⁺ under CID.

To circumvent this preferential formation of an H-bonded complex, we devised a system with no possibility of H-bonding. The isonicotinate ion was used as the Lewis base, and the complex isolated in the negative ion mode with m/z 397.01

*Source/detector conditions here

a.m.u. CID of this ion exclusively gave a species of m/z 122.02 a.m.u., corresponding to isonicotinate (fig. 4.30). This provides strong evidence that a Ch-bonded complex is able to be formed in the gas phase.

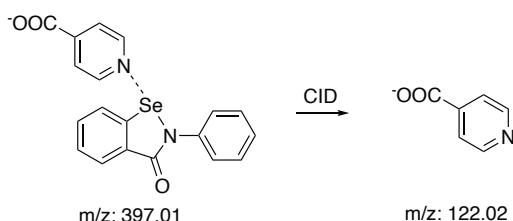


FIGURE 4.30: Dissociation of the complex **1**·isonicotinate under CID.

Using the technique of CID, one could imagine an experiment where such a Ch-bonded complex is fragmented with gradually increasing energy, and the yield of the product ion measured. This could theoretically be correlated with the gas phase bond energy. Indeed, this is the basis of the technique of Threshold CID (TCID) developed by Armentrout.^{21–23} Although the experiment is simple, the data analysis is not. This is due to the often poorly characterised energy distribution of the reactant ions, and vibrational energy redistribution in the products. These barriers are not insurmountable for simple systems (the technique has been successfully applied to metal complexes, and small non-covalent complexes), but in our moderately complex molecules with many vibrational modes, errors are likely to dominate any values extracted from the experiment. However, we present here a proof of concept experiment, which shows the decrease in reactant ion intensity and increase in product ion as CID energy is increased (fig. 4.31 and fig. 4.32).

The ion corresponding to the **1**·isonicotinate/**1a**·isonicotinate was first isolated directly from the ESI source in the negative ion mode, then this was subjected to increasing CID energies, recording spectra when the product isonicotinate ion at m/z 122 was first detected. Qualitatively, the **1**·isonicotinate Ch-bond can be seen to be stronger than the **1a**·isonicotinate Ch-bond, as a higher CID energy is needed to begin breaking the bond (7 arbitrary units *versus* 6 units), and the parent ion persists until 12 units as opposed to 10 units.

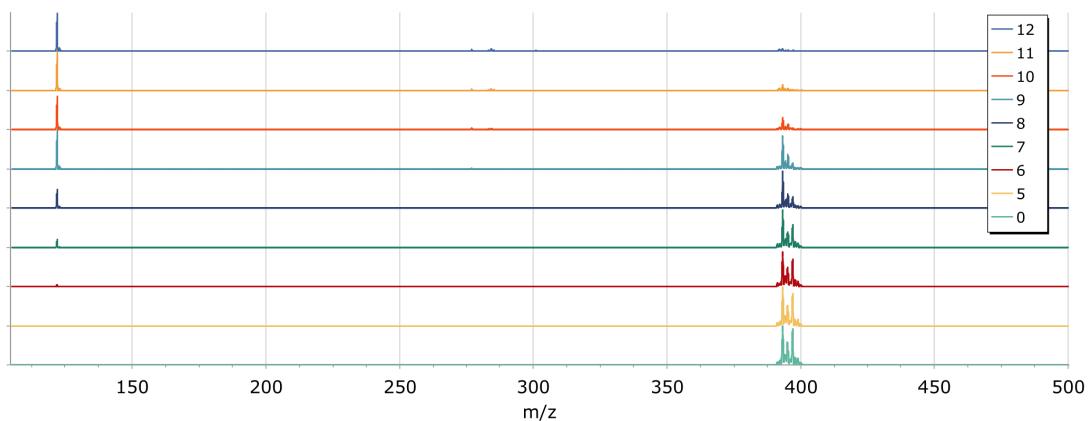


FIGURE 4.31: Rudimentary TCID experiment of **1**·isonicotinate. The CID energies are given in arbitrary units.

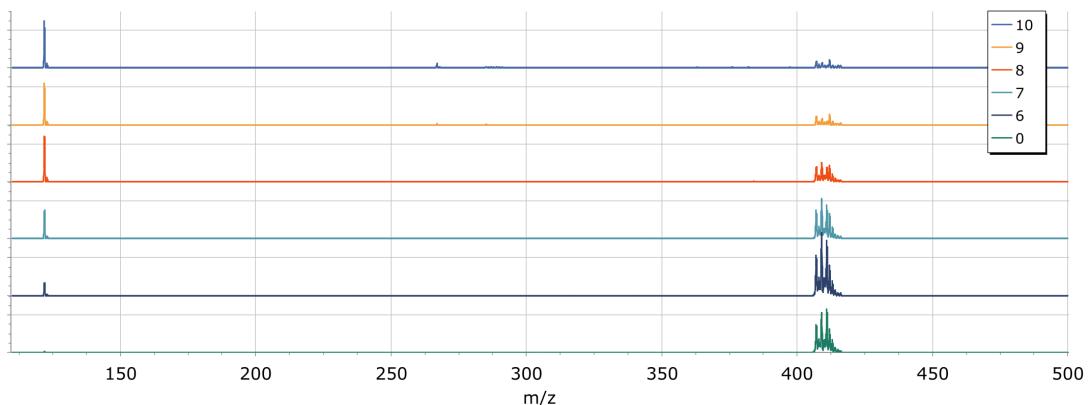


FIGURE 4.32: Rudimentary TCID experiment of **1a**·isonicotinate. The CID energies are given in arbitrary units.

4.3 Conclusion

To conclude, we have shown that ebselen derivatives reliably form Ch-bonded complexes in the solid, solution, and gas phases, and have introduced a number of techniques by which the complexes can be studied. Ch-bonding dominates the crystal packing of these derivatives, and geometric and electron density parameters vary in a manner consistent with the electron demand of the various substituents. Using this information, we constructed Hammett plots for a number of Lewis bases, which may be used to extrapolate to stronger or weaker systems. We also studied perturbations in the electron density by solid state NMR, which revealed large

chemical shift anisotropy of the selenium which was sensitive to the approach of a Lewis base. We exploited this sensitivity in the solution phase to evaluate dissociation constants by titration experiments, and showed that the Ch-bond comparable in energy to H-bonding. Finally, we attempted to isolate a Ch-bonded complex in the gas phase, which we were able to do using a negatively charged Lewis base. Low energy CID of this complex recovered the ion of the Lewis base, providing strong structural evidence that the complex was bound by a Ch-bond.

4.4 Supplementary materials

4.4.1 Synthetic procedures

4.4.1.1 Preparation of bis(2-carboxyphenyl)diselenide 5

Anthranilic acid (3.460 g, 25.33 mmol) was dissolved in 1 M hydrochloric acid (80 mL) and cooled to 0°C. Sodium nitrite (1.751 g, 25.37 mmol) in water (40 mL) was then added drop-wise with stirring. The mixture was kept at 0°C.

Selenium (2.080 g, 26.34 mmol) was suspended in 1 M sodium hydroxide (120 mL) and degassed by repeatedly evacuating the flask until the solvent began to boil, then backfilling with N₂. One grain of sodium borohydride was added under N₂ counter flow, followed by hydrazine hydrate (1.3 mL, 26 mmol). The mixture was heated until a red colour appeared, then cooled to 0°C.

The diazonium solution was neutralised with a solution of sodium hydroxide (4.320 g, 108.0 mmol) in water (20 mL), then added slowly to the cooled diselenide solution. Upon completion, the mixture was warmed to room temperature and stirred for 1 h, then neutralised with concentrated hydrochloric acid. The dense precipitate was separated by centrifuging, the supernatant decanted off, and the residue re-suspended in hot methanol (300 mL). The methanol solution was subjected to hot

filtration, and the orange filtrate concentrated to a volume of ~100 mL. This was cooled to -20°C, and the resulting crystals filtered off and dried. Recrystallisation from methanol gave **5** as a pale brown powder (1.8718 g, 37%).

¹H NMR (400 MHz, *d*₆-DMSO) δ ppm 7.29–7.38 (m, 1 H), 7.47 (t, *J*=7.43 Hz, 1 H), 7.66 (d, *J*=8.22 Hz, 1 H), 8.02 (d, *J*=7.43 Hz, 1 H) 13.70 (br. s., 1 H).

MS (ESI +ve) m/z 424.8806 (MNa⁺) C₁₄H₁₁O₄Se₂⁺ requires 424.8802 (Δ=0.941 ppm).

4.4.1.2 Preparation of 2-(chlorocarbonyl)phenylselenyl chloride **6**

Diselenide **5** (1.4758 g, 3.6881 mmol) was dissolved in thionyl chloride (10 ml) with 2 drops DMF, and heated to reflux for 90 minutes. The excess thionyl chloride was distilled off under reduced pressure, and the solid residue extracted into dry hexane (30 mL). Evaporation of the solvent afforded **6** as yellow crystals (1.870 g, 100%), which were used without further purification or characterisation.

4.4.1.3 General procedure for the preparation of benzisoselenazolinone derivatives **1CN**, **1CF₃**, **1Br**, **1CO₂Et**, **1OMe** and **1OEt** (procedure A)

The appropriate amine (2.5 mmol) was dissolved in anhydrous acetonitrile (5 mL) and cooled to 0°C. To this was added triethylamine (1 mL, distilled from CaH₂), followed by **6** (2.5 mmol) in a further 5 mL anhydrous acetonitrile. The mixture was stirred at room temperature for 2 h, then the solvent was removed under vacuum. The solid residue was triturated with water (5 mL) and 1 M hydrochloric acid to afford a friable solid, which was purified by recrystallisation from ethyl acetate at -20° C to afford the pure benzisoselenazolinone derivative.

4.4.1.4 Procedure for the preparation of benzisoselenazolinone derivative $\mathbf{1NO_2}$ (procedure B).

Sodium hydride (60% suspension in mineral oil, 125.0 mg, 3.125 mmol) was suspended in anhydrous THF (5 mL) and cooled to 0°C. 4-Nitroaniline (treated with charcoal and crystallised from aqueous ethanol, 277 mg, 2.01 mmol) was then added and the resulting dark red suspension stirred for 5 minutes at 0°C. DMAP (16.3 mg, 0.133 mmol) was then added, followed by a solution of the dichloride **6** (2.00 mmol) in a further 5 mL anhydrous THF. The mixture was then warmed to room temperature, and stirred for 18 h to give a light yellow suspension, which was quenched with methanol (5 mL), then suspended in water (50 mL). The resulting solid was filtered off, washed with 1 M HCl and water, then dried to give a light yellow powder (336.0 mg, 52%).

4.4.1.5 Procedure for the preparation of benzisoselenazolinone derivative **1 (procedure C).**

Copper iodide (98.4 mg, 0.517 mmol) and 1,10-phenanthroline (83.1 mg, 0.461 mmol) were stirred in anhydrous DMF (3 mL) for 15 mins at r.t., then 2-iodo-N-phenylbenzamide (653.2 mg, 2.021 mmol), selenium (196.9 mg, 2.495 mmol) and potassium carbonate (627.3 mg, 4.539 mmol) were added sequentially under a flow of argon. The mixture was heated at 110°C for 8 h, when TLC showed consumption of starting material. The mixture was then tipped into brine (30 mL) and stirred to form a brown precipitate, which was extracted into DCM (40 mL) and washed with water (2 × 20 mL). The DCM solution was filtered through a silica plug, then evaporated, and the residue applied to a SNAP 25 g silica cartridge, eluting with a petroleum ether/ethyl acetate gradient. The major peak was evaporated to afford colourless crystals of **1** (378.8 mg, 69%).

4.4.1.6 General procedure for the preparation of substituted pyridines **4a** and **4b**.

???

4.4.2 Crystallisation methods

Crystals of sufficient quality for x-ray diffraction analysis were grown by vapour diffusion of pentane into a dichloromethane solution (~50 mg/mL) of either the pure Ch-bond donor, or a 1:1 mixture of the donor and the appropriate Lewis base, with the following exceptions:

- the non-solvate of **1CN**·DMAP was grown by slow evaporation from THF,
- second polymorphs of **1OEt**·DMAP and **1OMe**·DMAP were grown from diffusion of pentane into diethyl ether,
- the γ polymorph of **1** was grown by vapour diffusion of pentane into a dichloromethane solution in the presence of triethylamine.

4.4.3 Characterisation data

4.4.3.1 Ebselen 1

Colourless crystals, m.p. 182.1–182.3°C (α polymorph), 173.4–174.6°C (β and γ polymorph), yield 69%.

^1H NMR (499 MHz, d_6 -DMSO) δ ppm 8.07 (1H, d, J = 8.02 Hz), 7.90 (1H, d, J = 7.45 Hz), 7.68 (1H, t, J = 7.25 Hz), 7.61 (2H, d, J = 7.68 Hz), 7.52–7.42 (3H, m), 7.26 (1H, t, J = 7.39 Hz).

4.4.3.2 4-Nitro ebselen **1NO₂**

Pale yellow crystals, yield 52%.

¹H NMR (499 MHz, *d*₆-DMSO) δ ppm 8.30 (2H, d, *J* = 9.15 Hz), 8.18 (1H, d, *J* = 8.08 Hz), 8.05 (2H, d, *J* = 9.16 Hz), 7.94 (1H, d, *J* = 7.72 Hz), 7.71 (1H, t, *J* = 7.02 Hz), 7.5 (1H, t, *J* = 7.45 Hz).

MS (ESI +ve) m/z 401.0005 ($M+Na^+$ + acetone) C₁₆H₁₄N₂NaO₄Se⁺ requires 401.00110 (Δ=1.52 ppm).

4.4.3.3 4-Cyano ebselen **1CN**

Colourless crystals, m.p. 191°C (dec.), yield 61%.

¹H NMR (600 MHz, *d*₆-DMSO) δ ppm 8.20 (1H, d, *J* = 8.08 Hz), 7.94–7.83 (5H, m), 7.65 (1H, t, *J* = 7.61 Hz), 7.45 (1H, t, *J* = 7.42 Hz).

¹³C NMR (151 MHz, *d*₆-DMSO) δ ppm 166.03, 145.18, 139.4, 133.88, 133.81, 133.03, 129.37, 128.47, 126.78, 126.74, 124.28, 119.20, 114.08, 107.18.

⁷⁷Se NMR (95 MHz, *d*₆-DMSO) δ ppm 897.93.

1CN·DMAP m.p. 157.6–160.2°C (DCM solvate, P2₁/c).

1CN·DMAP m.p. 176.0–178.8°C (non-solvate, Pbca).

4.4.3.4 4-Trifluoromethyl ebselen **1CF₃**

Colourless crystals, m.p. 184.6–186.4°C, yield 14%.

¹H NMR (499 MHz, *d*₆-DMSO) δ ppm 8.27 (1H, d, *J* = 7.41 Hz), 8.01–7.96 (1H, m), 7.95–7.89 (1H, m), 7.88–7.81 (3H, m), 7.76 (2H, d, *J* = 8.41 Hz).

1CF₃·DMAP m.p. 178.3–179.6°C.

4.4.3.5 4-Bromo ebselen 1Br

^1H NMR (499 MHz, d_6 -DMSO) δ ppm 8.09 (1H, d, J = 8.07 Hz), 7.89 (1H, d, J = 7.6 Hz), 7.71 (1H, s), 7.62 (4H, s), 7.48 (1H, t, J = 7.4 Hz).

Colourless crystals, m.p. 189.7–190.7°C, yield 18%.

1Br·DMAP m.p. 162.3–164.4°C.

4.4.3.6 4-Carboxyethyl ebselen 1CO₂Et

Colourless crystals, m.p. 173.1–174.5°C, yield 13%.

^1H NMR (500 MHz, d_6 -DMSO) δ 8.08 (d, J = 7.8 Hz, 1H), 8.02 (d, J = 8.7 Hz, 2H), 7.92 (dd, J = 0.4, 8.0 Hz, 1H), 7.88 (d, J = 8.6 Hz, 2H), 7.71 (t, J = 7.0 Hz, 1H), 7.50 (t, J = 7.3 Hz, 1H), 4.32 (q, J = 7.2 Hz, 2H), 1.33 (t, J = 7.1 Hz, 3H).

1CO₂Et·DMAP m.p. 136.0–138.7°C.

4.4.3.7 4-Methyl ebselen 1Me

Light brown crystals, dec.* , yield 58%.

^1H NMR (400 MHz, d_6 -DMSO) δ 8.06 (d, J = 8.0 Hz, 1H), 7.87 (d, J = 7.3 Hz, 1H), 7.66 (dt, J = 0.3, 7.4 Hz, 1H), 7.48 (d, J = 8.3 Hz, 2H), 7.46 (t, J = 7.3 Hz, 1H), 7.24 (d, J = 8.2 Hz, 2H), 2.31 (s, 3H).

^{13}C NMR (100 MHz, d_6 -DMSO) δ ppm 165.38, 139.3, 137.53, 135.73, 132.61, 130.06, 128.92, 128.35, 126.69, 126.24, 125.09, 21.01.

* The electron rich benziselenazolinones decomposed slightly above room temperature, so a melting point was not able to be obtained.

4.4.3.8 4-Methoxy ebselen 1OMe

Light brown crystals, dec.^{*}, yield 67%.

¹H NMR (500 MHz, *d*₆-DMSO) δ 8.07 (d, *J* = 7.9 Hz, 1H), 7.88 (d, *J* = 7.6 Hz, 1H), 7.67 (t, *J* = 7.1 Hz, 1H), 7.49 (d, *J* = 8.4 Hz, 2H), 7.47 (t, *J* = 6.9 Hz, 1H), 7.01 (d, *J* = 8.6 Hz, 2H), 3.78 (s, 3H).

4.4.3.9 4-Ethoxy ebselen 1OEt

Light brown crystals, dec.^{*}, yield 67%.

¹H NMR (500 MHz, *d*₆-DMSO) δ 8.06 (d, *J* = 8.0 Hz, 1H), 7.87 (d, *J* = 7.5 Hz, 1H), 7.67 (t, *J* = 7.2 Hz, 1H), 7.48 (t, *J* = 3.2 Hz, 1H), 7.47 (d, *J* = 8.8 Hz, 2H), 6.99 (d, *J* = 8.8 Hz, 2H), 4.05 (q, *J* = 7.0 Hz, 2H), 1.34 (t, *J* = 6.9 Hz, 3H).

4.4.4 Crystallographic Data

Intensity data was collected on a Rigaku XtalLAB Synergy at the specified temperature. The temperature was maintained using an Oxford Cryostream cooling device. The structures were solved by direct methods and difference Fourier synthesis.²⁴

4.4.4.1 Crystal data for 1

α and β polymorphs have previously been reported (CSD codes **SENGOH** and **SENGOH1**, respectively).²⁵

$C_{13}H_9NOSe$, γ polymorph, $M = 274.17$, $T = 101(2)$ K, $\lambda = 1.54184$ Å, monoclinic, space group $P2_1/c$ (no. 14), $a = 14.9405(5)$ Å, $b = 5.9905(2)$ Å, $c = 12.1716(4)$ Å, $\beta = 104.862(3)$ °, $V = 1052.93(6)$ Å³, $Z = 4$. $D_c = 1.730$ mg M⁻³, $\mu(\text{Cu K}\alpha) = 4.616$ mm⁻¹, $F(000) = 544$, crystal size $0.022 \times 0.057 \times 0.138$ mm. 4087 reflections measured,

$2\theta_{\max} = 154.39^\circ$, 4087 independent reflections, $R_{\text{int}} = 0.0626$, the final R was 0.0401 ($I > 2\theta(I)$, 3784 reflections) and $wR(F^2)$ was 0.1427 (all data), GOF 1.153.

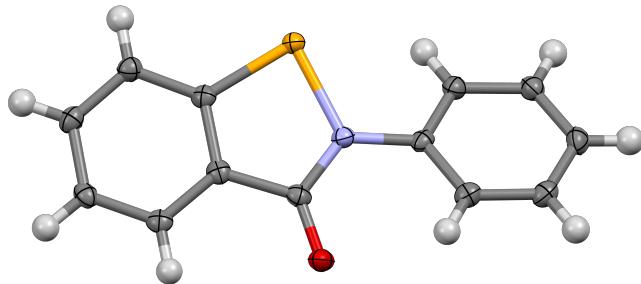


FIGURE 4.33: X-ray crystal structure of **1**.

4.4.4.2 Crystal data for **1·DMAP**

$C_{20}H_{19}N_3OSe$, $M = 396.34$, $T = 249.97(10)$ K, $\lambda = 1.54184$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 8.40490(10)$ Å, $b = 9.9751(3)$ Å, $c = 10.7243(3)$ Å, $\alpha = 93.231(2)^\circ$, $\beta = 93.031(2)^\circ$, $\gamma = 100.288(2)^\circ$, $V = 881.45(4)$ Å³, $Z = 2$. $D_c = 1.493$ mg M⁻³, $\mu(\text{Cu K}\alpha) = 2.980$ mm⁻¹, $F(000) = 404$, crystal size $0.096 \times 0.164 \times 0.331$ mm. 10762 reflections measured, $2\theta_{\max} = 154.198^\circ$, 3629 independent reflections, $R_{\text{int}} = 0.0375$, the final R was 0.0291 ($I > 2\theta(I)$, 3464 reflections) and $wR(F^2)$ was 0.0771 (all data), GOF 1.066.

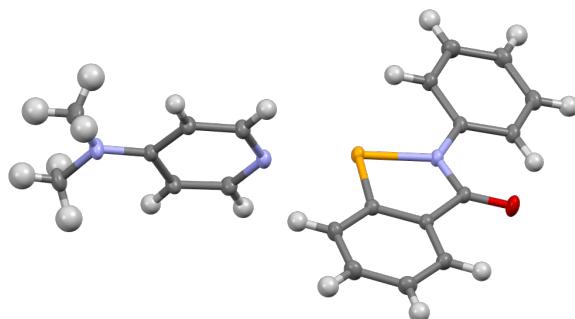


FIGURE 4.34: X-ray crystal structure of **1·DMAP**.

4.4.4.3 Crystal data for 1·4a

$C_{22}H_{21}N_3OSe$, $M = 422.38$, $T = 100.00(10)$ K, $\lambda = 0.71073$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 9.64890(10)$ Å, $b = 9.7748(2)$ Å, $c = 11.1296(2)$ Å, $\alpha = 112.887(2)^\circ$, $\beta = 108.4340(10)^\circ$, $\gamma = 90.8000(10)^\circ$, $V = 906.11(3)$ Å³, $Z = 2$. $D_c = 1.548$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 2.090$ mm⁻¹, $F(000) = 432$, crystal size $0.287 \times 0.361 \times 0.543$ mm. 112871 reflections measured, $2\theta_{\max} = 102.666^\circ$, 19957 independent reflections, $R_{\text{int}} = 0.0658$, the final R was 0.0423 ($I > 2\sigma(I)$, 13231 reflections) and $wR(F^2)$ was 0.1000 (all data), GOF 1.017.

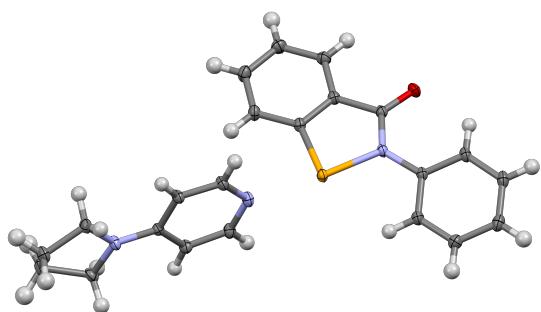
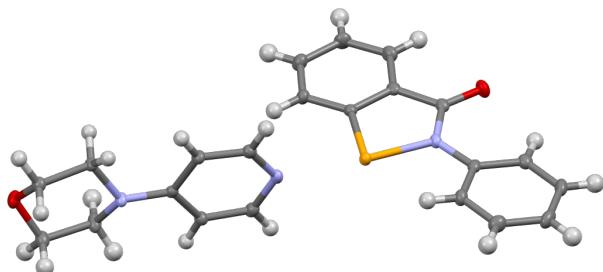


FIGURE 4.35: X-ray crystal structure of 1·4a.

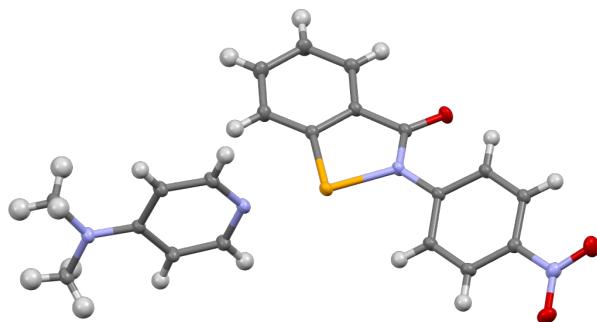
4.4.4.4 Crystal data for 1·4b

$C_{22}H_{21}N_3O_2Se$, $M = 438.38$, $T = 100.00(10)$ K, $\lambda = 0.71073$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 9.6802(4)$ Å, $b = 10.0031(3)$ Å, $c = 11.0601(5)$ Å, $\alpha = 72.555(3)^\circ$, $\beta = 65.653(4)^\circ$, $\gamma = 89.402(3)^\circ$, $V = 922.93(7)$ Å³, $Z = 2$. $D_c = 1.577$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 2.059$ mm⁻¹, $F(000) = 448$, crystal size $0.228 \times 0.379 \times 0.406$ mm. 74878 reflections measured, $2\theta_{\max} = 102.822^\circ$, 20252 independent reflections, $R_{\text{int}} = 0.0372$, the final R was 0.0372 ($I > 2\sigma(I)$, 15377 reflections) and $wR(F^2)$ was 0.0953 (all data), GOF 1.031.

FIGURE 4.36: X-ray crystal structure of **1·4b**.

4.4.4.5 Crystal data for $\text{1NO}_2\cdot\text{DMAP}$

$\text{C}_{20}\text{H}_{18}\text{N}_4\text{O}_3\text{Se}$, $M = 441.34$, $T = 99.99(10)$ K, $\lambda = 0.71073$ Å, triclinic, space group $\text{P}\bar{1}$ (no. 2), $a = 9.33330(10)$ Å, $b = 9.5828(2)$ Å, $c = 12.0004(2)$ Å, $\alpha = 109.266(2)^\circ$, $\beta = 108.2410(10)^\circ$, $\gamma = 98.5910(10)^\circ$, $V = 923.28(3)$ Å³, $Z = 2$. $D_c = 1.588$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 2.064$ mm⁻¹, $F(000) = 448$, crystal size $0.109 \times 0.182 \times 0.607$ mm. 73764 reflections measured, $2\theta_{\max} = 108.038^\circ$, 22302 independent reflections, $R_{\text{int}} = 0.0309$, the final R was 0.0338 ($I > 2\sigma(I)$, 15459 reflections) and $wR(F^2)$ was 0.0813 (all data), GOF 1.030.

FIGURE 4.37: X-ray crystal structure of $\text{1NO}_2\cdot\text{DMAP}$.

4.4.4.6 Crystal data for $\text{1NO}_2\cdot\text{DMF}$

$\text{C}_{16}\text{H}_{15}\text{N}_3\text{O}_4\text{Se}$, $M = 392.27$, $T = 100.00(10)$ K, $\lambda = 1.54184$ Å, triclinic, space group $\text{P}\bar{1}$ (no. 2), $a = 7.2085(2)$ Å, $b = 8.1572(2)$ Å, $c = 13.6748(3)$ Å, $\alpha = 74.985(2)^\circ$, $\beta =$

$82.896(2)^\circ$, $\gamma = 84.509(2)^\circ$, $V = 768.98(3)$ Å 3 , $Z = 2$. $D_c = 1.694$ mg M $^{-3}$, $\mu(\text{Cu K}\alpha) = 3.559$ mm $^{-1}$, $F(000) = 396$, crystal size $0.148 \times 0.183 \times 0.662$ mm. 8506 reflections measured, $2\theta_{\max} = 154.322^\circ$, 3155 independent reflections, $R_{\text{int}} = 0.0304$, the final R was 0.0269 ($I > 2\theta(I)$, 3127 reflections) and $wR(F^2)$ was 0.0721 (all data), GOF 1.088.

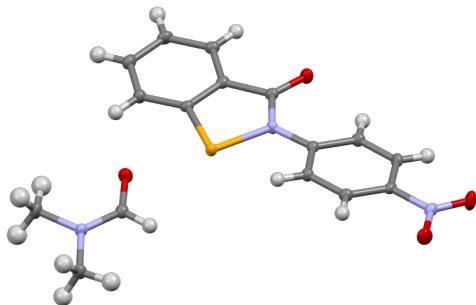


FIGURE 4.38: X-ray crystal structure of **1NO₂**·DMF.

4.4.4.7 Crystal data for **1CN**

$\text{C}_{14}\text{H}_8\text{N}_2\text{OSe}$, $M = 299.18$, $T = 123(30)$ K, $\lambda = 1.54184$ Å, monoclinic, space group C2/c (no. 15), $a = 28.4741(7)$ Å, $b = 11.5814(3)$ Å, $c = 7.4663(2)$ Å, $\beta = 91.134(2)^\circ$, $V = 2461.68(11)$ Å 3 , $Z = 8$. $D_c = 1.615$ mg M $^{-3}$, $\mu(\text{Cu K}\alpha) = 4.034$ mm $^{-1}$, $F(000) = 1184$, crystal size $0.057 \times 0.093 \times 0.198$ mm. 8242 reflections measured, $2\theta_{\max} = 154.516^\circ$, 2539 independent reflections, $R_{\text{int}} = 0.0304$, the final R was 0.0314 ($I > 2\theta(I)$, 2412 reflections) and $wR(F^2)$ was 0.0915 (all data), GOF 1.117.

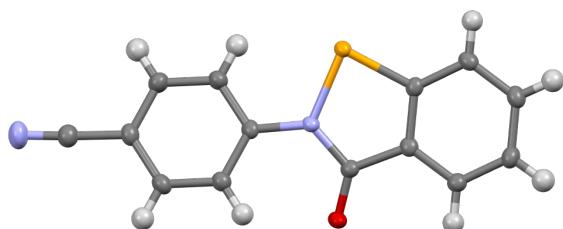


FIGURE 4.39: X-ray crystal structure of **1CN**.

4.4.4.8 Crystal data for **1CN·DMAP**

$C_{21}H_{18}N_4OSe$, $M = 421.35$, $T = 200.00(10)$ K, $\lambda = 0.71073$ Å, monoclinic, space group $P2_1/c$ (no. 14), $a = 10.1440(3)$ Å, $b = 9.1899(2)$ Å, $c = 20.6385(7)$ Å, $\beta = 95.836(3)^\circ$, $V = 1914.00(10)$ Å³, $Z = 4$. $D_c = 1.462$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 1.980$ mm⁻¹, $F(000) = 856$, crystal size $0.237 \times 0.485 \times 0.782$ mm. 47246 reflections measured, $2\theta_{\max} = 82.326^\circ$, 12358 independent reflections, $R_{\text{int}} = 0.0466$, the final R was 0.0432 ($I > 2\theta(I)$, 6716 reflections) and $wR(F^2)$ was 0.1046 (all data), GOF 0.999.

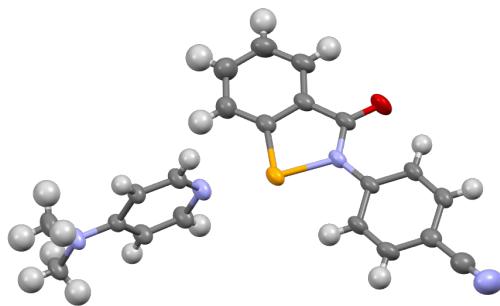
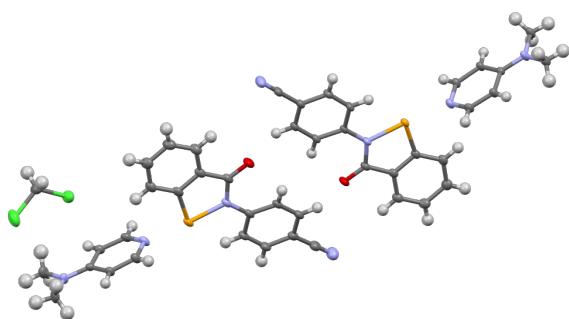


FIGURE 4.40: X-ray crystal structure of **1CN·DMAP**.

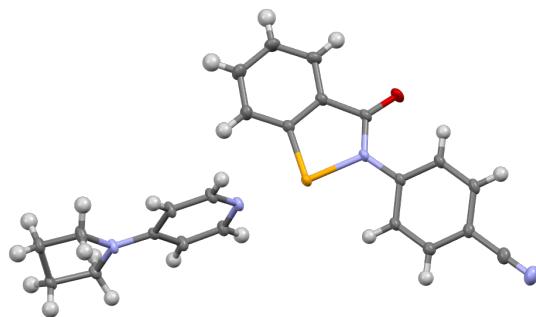
4.4.4.9 Crystal data for **1CN·DMAP·DCM**

$C_{21} \cdot 50H_{19}ClN_4OSe$, $M = 463.82$, $T = 99.99(10)$ K, $\lambda = 0.71073$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 12.7673(3)$ Å, $b = 13.5668(2)$ Å, $c = 13.5764(2)$ Å, $\alpha = 85.4790(10)^\circ$, $\beta = 76.0730(10)^\circ$, $\gamma = 62.304(2)^\circ$, $V = 2019.45(7)$ Å³, $Z = 4$. $D_c = 1.526$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 2.012$ mm⁻¹, $F(000) = 940$, crystal size $0.205 \times 0.279 \times 0.391$ mm. 76018 reflections measured, $2\theta_{\max} = 81.524^\circ$, 25014 independent reflections, $R_{\text{int}} = 0.0670$, the final R was 0.0687 ($I > 2\theta(I)$, 15296 reflections) and $wR(F^2)$ was 0.1307 (all data), GOF 1.040.

FIGURE 4.41: X-ray crystal structure of **1CN·DMAP·DCM**.

4.4.4.10 Crystal data for **1CN·4a**

$C_{23}H_{20}N_4OSe$, $M = 447.39$, $T = 100.0(2)$ K, $\lambda = 0.71073$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 9.7432(4)$ Å, $b = 9.7525(4)$ Å, $c = 12.2261(4)$ Å, $\alpha = 110.327(3)^\circ$, $\beta = 112.660(3)^\circ$, $\gamma = 90.219(3)^\circ$, $V = 992.49(7)$ Å³, $Z = 4$. $D_c = 2.994$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 3.828$ mm⁻¹, $F(000) = 912$, crystal size $0.152 \times 0.339 \times 0.549$ mm. 22193 reflections measured, $2\theta_{\max} = 74.602^\circ$, 9250 independent reflections, $R_{\text{int}} = 0.0341$, the final R was 0.0369 ($I > 2\sigma(I)$, 7367 reflections) and $wR(F^2)$ was 0.0899 (all data), GOF 1.037.

FIGURE 4.42: X-ray crystal structure of **1CN·4a**.

4.4.4.11 Crystal data for **1CN·4b·DCM**

$C_{24}H_{22}Cl_2N_4O_2Se$, $M = 548.31$, $T = 100.00(10)$ K, $\lambda = 0.71073$ Å, monoclinic, space group $P2_1/c$ (no. 14), $a = 9.8472(2)$ Å, $b = 9.1428(2)$ Å, $c = 25.8520(5)$ Å, $\beta = 96.395(2)^\circ$, $V = 2313.00(8)$ Å³, $Z = 4$. $D_c = 1.575$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 1.885$ mm⁻¹,

$F(000) = 1112$, crystal size $0.179 \times 0.511 \times 0.673$ mm. 49263 reflections measured, $2\theta_{\max} = 74.33^\circ$, 11127 independent reflections, $R_{\text{int}} = 0.0462$, the final R was 0.0378 ($I > 2\theta(I)$, 8288 reflections) and $wR(F^2)$ was 0.0920 (all data), GOF 1.031.

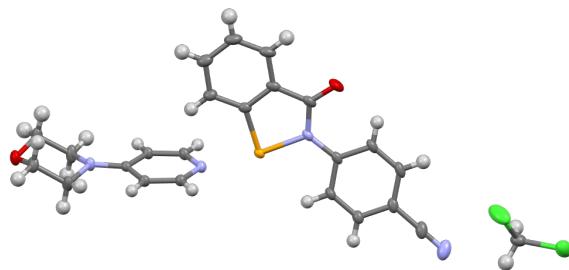


FIGURE 4.43: X-ray crystal structure of **1CN·4b·DCM**.

4.4.4.12 Crystal data for **1CF₃**

$C_{14}H_8F_3NOSe$, $M = 342.17$, $T = 100.00(10)$ K, $\lambda = 1.54184$ Å, monoclinic, space group P2₁/c (no. 14), $a = 7.54860(10)$ Å, $b = 11.7195(2)$ Å, $c = 27.6207(5)$ Å, $\beta = 90.998(2)^\circ$, $V = 2443.12(7)$ Å³, $Z = 8$. $D_c = 1.861$ mg M⁻³, $\mu(\text{Cu K}\alpha) = 4.497$ mm⁻¹, $F(000) = 1344$, crystal size $0.022 \times 0.1 \times 0.135$ mm. 15455 reflections measured, $2\theta_{\max} = 154.602^\circ$, 5042 independent reflections, $R_{\text{int}} = 0.0408$, the final R was 0.0957 ($I > 2\theta(I)$, 4332 reflections) and $wR(F^2)$ was 0.2514 (all data), GOF 1.148.

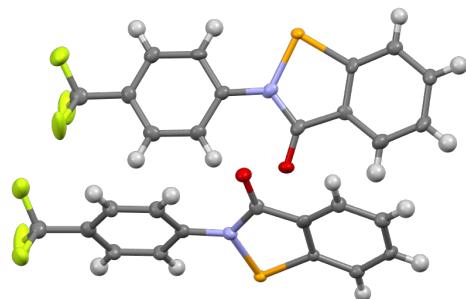


FIGURE 4.44: X-ray crystal structure of **1CF₃**.

4.4.4.13 Crystal data for $\mathbf{1CF_3\cdot DMAP}$

$C_{21}H_{18}F_3N_3OSe$, $M = 464.34$, $T = 100.00(10)$ K, $\lambda = 0.71073$ Å, monoclinic, space group $P2_1/n$ (no. 14), $a = 10.1115(3)$ Å, $b = 9.1437(2)$ Å, $c = 20.9441(6)$ Å, $\beta = 96.060(2)^\circ$, $V = 1925.60(9)$ Å³, $Z = 4$. $D_c = 1.602$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 1.996$ mm⁻¹, $F(000) = 936$, crystal size $0.032 \times 0.123 \times 0.259$ mm. 98727 reflections measured, $2\theta_{\max} = 116.164^\circ$, 26657 independent reflections, $R_{\text{int}} = 0.0669$, the final R was 0.0509 ($I > 2\theta(I)$, 12525 reflections) and $wR(F^2)$ was 0.1406 (all data), GOF 0.979.

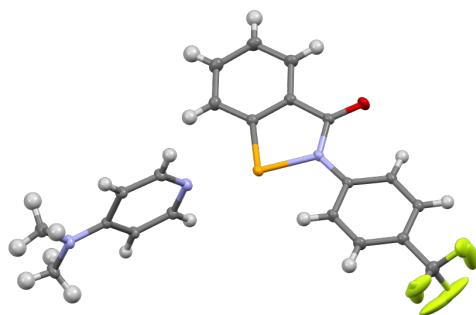


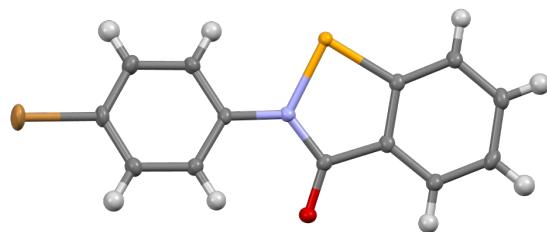
FIGURE 4.45: X-ray crystal structure of $\mathbf{1CF_3\cdot DMAP}$.

4.4.4.14 Crystal data for $\mathbf{1Br}$

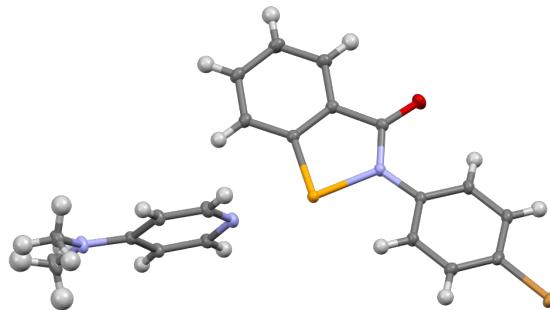
$C_{13}H_8BrNOSe$, $M = 201.90$, $T = 101(1)$ K, $\lambda = 1.54184$ Å, monoclinic, space group $P2_1/c$ (no. 14), $a = 13.9453(2)$ Å, $b = 11.7592(2)$ Å, $c = 7.47100(10)$ Å, $\beta = 102.4360(10)^\circ$, $V = 1196.39(3)$ Å³, $Z = 12$. $D_c = 3.363$ mg M⁻³, $\mu(\text{Cu K}\alpha) = 22.937$ mm⁻¹, $F(000) = 1092$, crystal size $0.022 \times 0.046 \times 0.124$ mm. 7959 reflections measured, $2\theta_{\max} = 153.406^\circ$, 2454 independent reflections, $R_{\text{int}} = 0.0227$, the final R was 0.0239 ($I > 2\theta(I)$, 2358 reflections) and $wR(F^2)$ was 0.0658 (all data), GOF 1.097.

4.4.4.15 Crystal data for $\mathbf{1Br\cdot DMAP}$

$C_{20}H_{18}BrN_3OSe$, $M = 475.24$, $T = 100.00(10)$ K, $\lambda = 0.71073$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 8.5472(2)$ Å, $b = 10.0870(2)$ Å, $c = 12.2848(2)$ Å, $\alpha = 68.572(2)^\circ$,

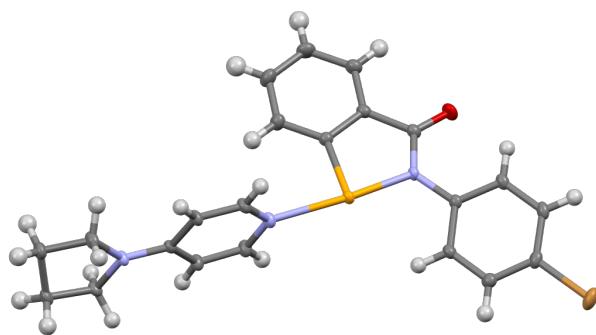
FIGURE 4.46: X-ray crystal structure of **1Br**.

$\beta = 70.720(2)^\circ$, $\gamma = 89.997(2)^\circ$, $V = 921.83(4) \text{ \AA}^3$, $Z = 2$. $D_c = 1.712 \text{ mg M}^{-3}$, $\mu(\text{Mo K}\alpha) = 4.218 \text{ mm}^{-1}$, $F(000) = 472$, crystal size $0.105 \times 0.195 \times 0.425 \text{ mm}$. 67535 reflections measured, $2\theta_{\max} = 102.658^\circ$, 20158 independent reflections, $R_{\text{int}} = 0.0377$, the final R was 0.0332 ($I > 2\sigma(I)$, 13604 reflections) and $wR(F^2)$ was 0.0729 (all data), GOF 1.020.

FIGURE 4.47: X-ray crystal structure of **1Br·DMAP**.

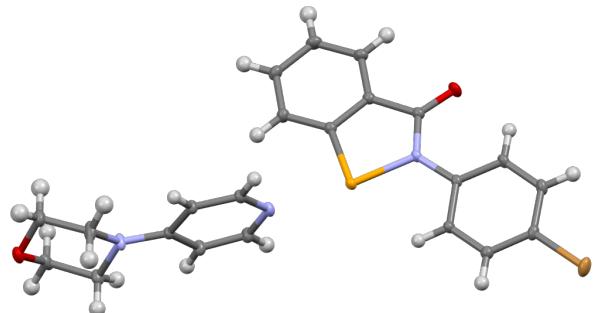
4.4.4.16 Crystal data for **1Br·4a**

$C_{22}H_{20}BrN_2OSe$, $M = 487.27$, $T = 100.00(10) \text{ K}$, $\lambda = 0.71073 \text{ \AA}$, triclinic, space group $P\bar{1}$ (no. 2), $a = 9.5724(3) \text{ \AA}$, $b = 9.8138(3) \text{ \AA}$, $c = 12.5411(3) \text{ \AA}$, $\alpha = 67.273(2)^\circ$, $\beta = 67.640(3)^\circ$, $\gamma = 89.385(2)^\circ$, $V = 992.05(5) \text{ \AA}^3$, $Z = 4$. $D_c = 3.262 \text{ mg M}^{-3}$, $\mu(\text{Mo K}\alpha) = 7.841 \text{ mm}^{-1}$, $F(000) = 972$, crystal size $0.047 \times 0.309 \times 0.369 \text{ mm}$. 51194 reflections measured, $2\theta_{\max} = 74.618^\circ$, 9698 independent reflections, $R_{\text{int}} = 0.0748$, the final R was 0.0428 ($I > 2\sigma(I)$, 7012 reflections) and $wR(F^2)$ was 0.0969 (all data), GOF 1.030.

FIGURE 4.48: X-ray crystal structure of **1Br·4a**.

4.4.4.17 Crystal data for **1Br·4b**

$C_{22}H_{20}BrN_3O_2Se$, $M = 517.28$, $T = 100.00(10)$ K, $\lambda = 0.71073$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 9.1265(2)$ Å, $b = 10.2233(2)$ Å, $c = 11.9313(2)$ Å, $\alpha = 111.203(2)^\circ$, $\beta = 105.914(2)^\circ$, $\gamma = 93.809(2)^\circ$, $V = 980.92(4)$ Å³, $Z = 2$. $D_c = 1.751$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 3.976$ mm⁻¹, $F(000) = 516$, crystal size $0.172 \times 0.234 \times 0.408$ mm. 57051 reflections measured, $2\theta_{\max} = 74.526^\circ$, 9605 independent reflections, $R_{\text{int}} = 0.0731$, the final R was 0.0403 ($I > 2\sigma(I)$, 7636 reflections) and $wR(F^2)$ was 0.0957 (all data), GOF 1.025.

FIGURE 4.49: X-ray crystal structure of **1Br·4b**.

4.4.4.18 Crystal data for **1CO₂Et**

$C_{16}H_{13}NO_3Se$, $M = 346.23$, $T = 100.5(9)$ K, $\lambda = 1.54184$ Å, monoclinic, space group $P2_1/c$ (no. 14), $a = 24.0629(3)$ Å, $b = 4.75510(10)$ Å, $c = 11.8795(2)$ Å, $\beta = 95.6310(10)^\circ$,

$V = 1352.71(4) \text{ \AA}^3$, $Z = 4$. $D_c = 1.700 \text{ mg M}^{-3}$, $\mu(\text{Cu K}\alpha) = 3.854 \text{ mm}^{-1}$, $F(000) = 696$, crystal size $0.022 \times 0.045 \times 0.222 \text{ mm}$. 8636 reflections measured, $2\theta_{\max} = 154.516^\circ$, 2767 independent reflections, $R_{\text{int}} = 0.0365$, the final R was $0.0317 (I > 2\theta(I), 2564 \text{ reflections})$ and $wR(F^2)$ was 0.0878 (all data), GOF 1.085.

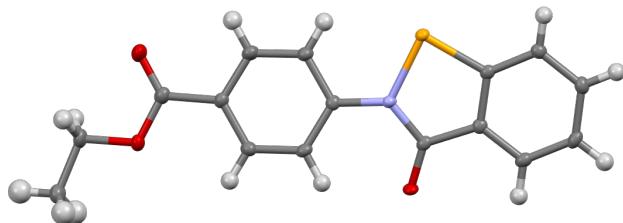


FIGURE 4.50: X-ray crystal structure of **1CO₂Et**.

4.4.4.19 Crystal data for **1CO₂Et·DMAP·DCM**

$C_{24}H_{25}ClN_3O_3Se$, $M = 517.88$, $T = 100.6(10) \text{ K}$, $\lambda = 0.71073 \text{ \AA}$, triclinic, space group $P\bar{1}$ (no. 2), $a = 8.9533(2) \text{ \AA}$, $b = 10.2462(2) \text{ \AA}$, $c = 12.2540(2) \text{ \AA}$, $\alpha = 81.391(2)^\circ$, $\beta = 82.583(2)^\circ$, $\gamma = 84.484(2)^\circ$, $V = 1098.80(4) \text{ \AA}^3$, $Z = 2$. $D_c = 1.565 \text{ mg M}^{-3}$, $\mu(\text{Mo K}\alpha) = 1.863 \text{ mm}^{-1}$, $F(000) = 530$, crystal size $0.065 \times 0.226 \times 0.601 \text{ mm}$. 78896 reflections measured, $2\theta_{\max} = 102.748^\circ$, 24099 independent reflections, $R_{\text{int}} = 0.0411$, the final R was $0.0588 (I > 2\theta(I), 14613 \text{ reflections})$ and $wR(F^2)$ was 0.1711 (all data), GOF 1.036.

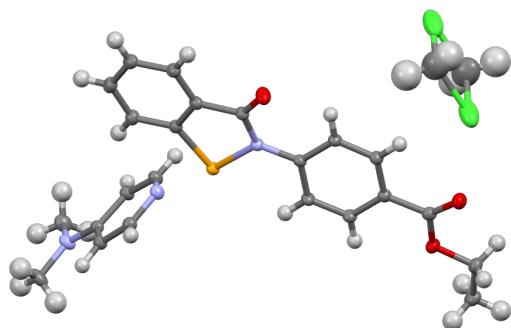


FIGURE 4.51: X-ray crystal structure of **1CO₂Et·DMAP·DCM**.

4.4.4.20 Crystal data for $\mathbf{1CO_2Et \cdot 4a}$

$C_{25}H_{25}N_3O_3Se$, $M = 494.44$, $T = 100.00(10)$ K, $\lambda = 1.54184$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 9.8257(6)$ Å, $b = 9.8381(3)$ Å, $c = 11.6093(7)$ Å, $\alpha = 97.830(4)^\circ$, $\beta = 100.220(5)^\circ$, $\gamma = 92.729(4)^\circ$, $V = 1091.15(10)$ Å³, $Z = 2$. $D_c = 1.505$ mg M⁻³, $\mu(\text{Cu K}\alpha) = 2.602$ mm⁻¹, $F(000) = 508$, crystal size $0.092 \times 0.171 \times 0.35$ mm. 12186 reflections measured, $2\theta_{\max} = 154.082^\circ$, 4458 independent reflections, $R_{\text{int}} = 0.0471$, the final R was 0.0554 ($I > 2\theta(I)$, 4151 reflections) and $wR(F^2)$ was 0.1539 (all data), GOF 1.051.

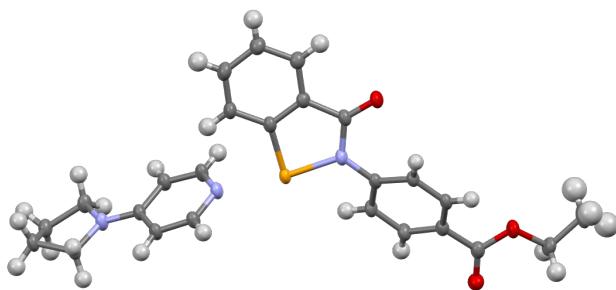


FIGURE 4.52: X-ray crystal structure of $\mathbf{1CO_2Et \cdot 4a}$.

4.4.4.21 Crystal data for $\mathbf{1CO_2Et \cdot 4b}$

$C_{25}H_{25}N_3O_4Se$, $M = 510.44$, $T = 100.00(10)$ K, $\lambda = 0.71073$ Å, monoclinic, space group $P2_1/n$ (no. 14), $a = 9.12020(10)$ Å, $b = 10.7781(2)$ Å, $c = 22.8685(4)$ Å, $\beta = 96.436(2)^\circ$, $V = 2233.77(6)$ Å³, $Z = 4$. $D_c = 1.518$ mg M⁻³, $\mu(\text{MoK}\alpha) = 1.719$ mm⁻¹, $F(000) = 1048$, crystal size $0.051 \times 0.087 \times 0.329$ mm. 50769 reflections measured, $2\theta_{\max} = 78.776^\circ$, 13312 independent reflections, $R_{\text{int}} = 0.0344$, the final R was 0.0354 ($I > 2\theta(I)$, 9910 reflections) and $wR(F^2)$ was 0.0846 (all data), GOF 1.027.

4.4.4.22 Crystal data for $\mathbf{1Me}$

$C_{14}H_{11}NOSe$, $M = 288.20$, $T = 100(1)$ K, $\lambda = 1.54184$ Å, monoclinic, space group $P2_1/c$ (no. 14), $a = 16.2262(3)$ Å, $b = 6.03740(10)$ Å, $c = 12.3400(2)$ Å, $\beta = 106.925(2)^\circ$,

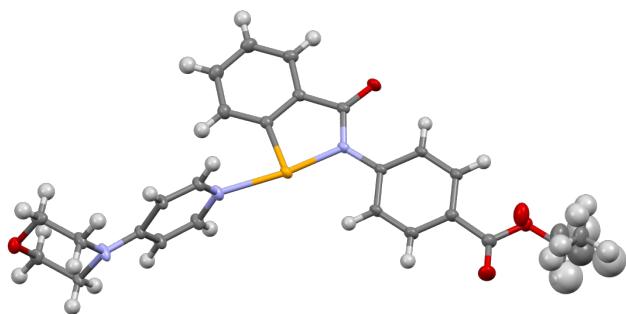


FIGURE 4.53: X-ray crystal structure of **1CO₂Et·4b**. The ethyl group is disordered over two sites.

$V = 1156.52(4) \text{ \AA}^3$, $Z = 4$. $D_c = 1.655 \text{ mg M}^{-3}$, $\mu(\text{Cu K}\alpha) = 4.234 \text{ mm}^{-1}$, $F(000) = 576$, crystal size $0.017 \times 0.066 \times 0.098 \text{ mm}$. 11405 reflections measured, $2\theta_{\max} = 153.926^\circ$, 2412 independent reflections, $R_{\text{int}} = 0.0442$, the final R was 0.0433 ($I > 2\sigma(I)$, 2186 reflections) and $wR(F^2)$ was 0.1206 (all data), GOF 1.065.

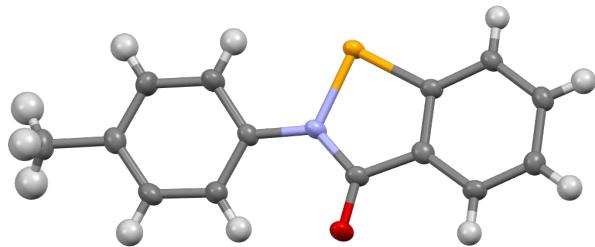


FIGURE 4.54: X-ray crystal structure of **1Me**.

4.4.4.23 Crystal data for **1Me·DMAP**

$C_{21}H_{21}N_3OSe$, $M = 410.380$, $T = 100.00(10) \text{ K}$, $\lambda = 0.71073 \text{ \AA}$, monoclinic, space group $P2_1/c$ (no. 14), $a = 12.9280(2) \text{ \AA}$, $b = 8.6159(1) \text{ \AA}$, $c = 17.4656(3) \text{ \AA}$, $\beta = 109.549(2)^\circ$, $V = 1833.29(5) \text{ \AA}^3$, $Z = 4$. $D_c = 1.487 \text{ mg M}^{-3}$, $\mu(\text{Mo K}\alpha) = 2.063 \text{ mm}^{-1}$, $F(000) = 840.219$, crystal size $0.099 \times 0.202 \times 0.321 \text{ mm}$. 118568 reflections measured, $2\theta_{\max} = 102.6^\circ$, 20042 independent reflections, $R_{\text{int}} = 0.0464$, the final R was 0.0298 ($I > 2\sigma(I)$, 11677 reflections) and $wR(F^2)$ was 0.0365 (all data), GOF 0.9855.

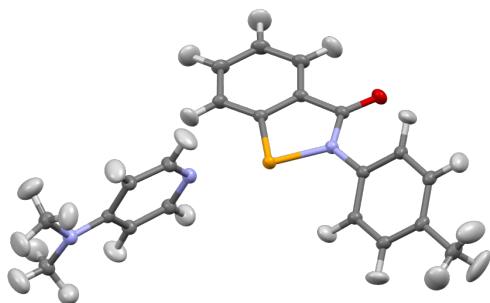


FIGURE 4.55: X-ray crystal structure of **1Me·DMAP**.

4.4.4.24 Crystal data for **1Me·4a**

$C_{23}H_{23}N_3OSe$, $M = 436.40$, $T = 169.99(10)$ K, $\lambda = 0.71073$ Å, monoclinic, space group $P2_1/c$ (no. 14), $a = 9.7501(2)$ Å, $b = 13.2335(3)$ Å, $c = 16.0268(4)$ Å, $\beta = 99.956(2)^\circ$, $V = 2036.76(8)$ Å³, $Z = 4$. $D_c = 1.423$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 1.862$ mm⁻¹, $F(000) = 896$, crystal size $0.072 \times 0.257 \times 0.343$ mm. 94736 reflections measured, $2\theta_{\max} = 89.802^\circ$, 16669 independent reflections, $R_{\text{int}} = 0.0938$, the final R was 0.0491 ($I > 2\sigma(I)$, 6838 reflections) and $wR(F^2)$ was 0.1154 (all data), GOF 0.976.

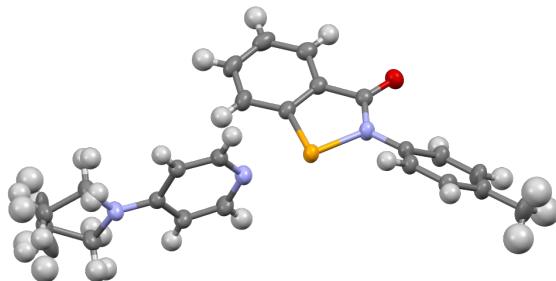


FIGURE 4.56: X-ray crystal structure of **1Me·4a**. The pyrrol group is disordered over two sites.

4.4.4.25 Crystal data for **1Me·4b**

$C_{23}H_{23}N_3O_2Se$, $M = 452.40$, $T = 100.01(10)$ K, $\lambda = 1.54184$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 9.2634(3)$ Å, $b = 10.1601(3)$ Å, $c = 11.9076(4)$ Å, $\alpha = 110.439(3)^\circ$, $\beta = 108.347(3)^\circ$, $\gamma = 93.315(3)^\circ$, $V = 978.88(6)$ Å³, $Z = 2$. $D_c = 1.535$ mg M⁻³, $\mu(\text{Cu K}\alpha) =$

2.801 mm^{-1} , $F(000) = 464$, crystal size $0.035 \times 0.085 \times 0.156\text{ mm}$. 11592 reflections measured, $2\theta_{\max} = 154.304^\circ$, 4003 independent reflections, $R_{\text{int}} = 0.0474$, the final R was 0.0814 ($I > 2\theta(I)$, 3783 reflections) and $wR(F^2)$ was 0.1934 (all data), GOF 1.042.

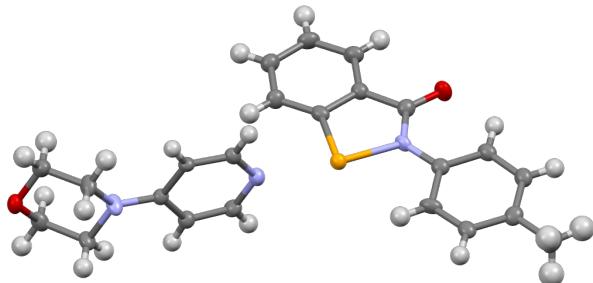


FIGURE 4.57: X-ray crystal structure of **1Me·4b**.

4.4.4.26 Crystal data for **1OMe**

$\text{C}_{14}\text{H}_{11}\text{NO}_2\text{Se}$, $M = 304.20$, $T = 100.01(10)\text{ K}$, $\lambda = 0.71073\text{ \AA}$, monoclinic, space group $\text{P}2_1/\text{c}$ (no. 14), $a = 6.85470(10)\text{ \AA}$, $b = 12.0394(2)\text{ \AA}$, $c = 15.1698(3)\text{ \AA}$, $\beta = 102.659(2)^\circ$, $V = 1221.48(4)\text{ \AA}^3$, $Z = 4$. $D_c = 1.654\text{ mg M}^{-3}$, $\mu(\text{Mo K}\alpha) = 3.066\text{ mm}^{-1}$, $F(000) = 608$, crystal size $0.077 \times 0.184 \times 0.380\text{ mm}$. 52768 reflections measured, $2\theta_{\max} = 89.722^\circ$, 9983 independent reflections, $R_{\text{int}} = 0.0591$, the final R was 0.0342 ($I > 2\theta(I)$, 7115 reflections) and $wR(F^2)$ was 0.0827 (all data), GOF 1.013.

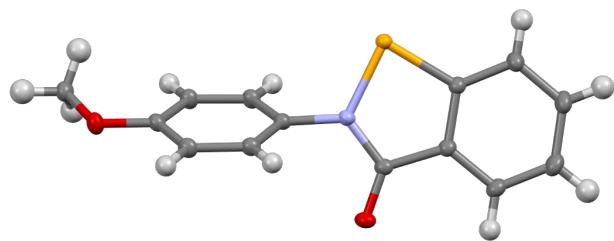


FIGURE 4.58: X-ray crystal structure of **1OMe**.

4.4.4.27 Crystal data for **1OMe·DMAP**

$C_{21}H_{21}N_3O_2Se$, polymorph A, $M = 426.37$, $T = 100.00(10)$ K, $\lambda = 0.71073$ Å, monoclinic, space group $P2_1/n$ (no. 14), $a = 12.0928(2)$ Å, $b = 17.5615(2)$ Å, $c = 18.6689(2)$ Å, $\beta = 106.4180(10)^\circ$, $V = 3803.01(9)$ Å³, $Z = 8$. $D_c = 1.489$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 1.996$ mm⁻¹, $F(000) = 1744$, crystal size $0.173 \times 0.271 \times 0.381$ mm. 371431 reflections measured, $2\theta_{\max} = 116.262^\circ$, 53275 independent reflections, $R_{\text{int}} = 0.0888$, the final R was 0.0518 ($I > 2\theta(I)$, 21468 reflections) and $wR(F^2)$ was 0.1103 (all data), GOF 0.985.

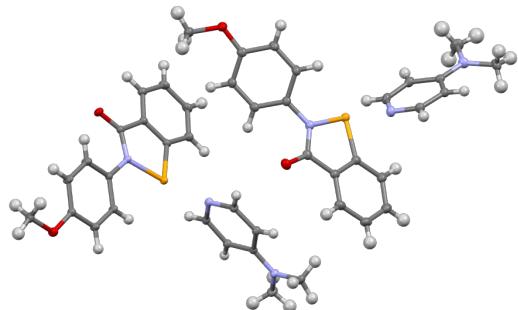
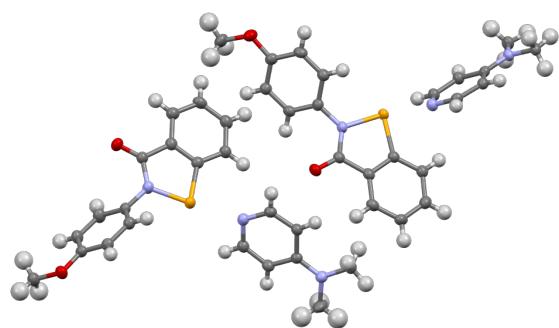


FIGURE 4.59: X-ray crystal structure of **1OMe·DMAP**, polymorph A.

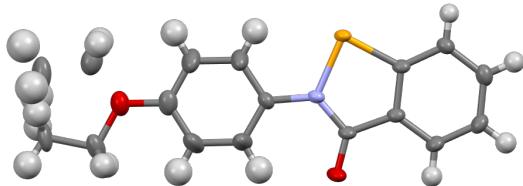
$C_{21}H_{21}N_3O_2Se$, polymorph B, $M = 426.37$, $T = 99.99(10)$ K, $\lambda = 0.71073$ Å, monoclinic, space group $P2_1/c$ (no. 14), $a = 20.2207(2)$ Å, $b = 8.72690(10)$ Å, $c = 21.9128(3)$ Å, $\beta = 106.5990(10)^\circ$, $V = 3705.68(8)$ Å³, $Z = 8$. $D_c = 1.528$ mg M⁻³, $\mu(\text{Mo K}\alpha) = 2.048$ mm⁻¹, $F(000) = 1744$, crystal size $0.234 \times 0.439 \times 1.346$ mm. 166933 reflections measured, $2\theta_{\max} = 102.63^\circ$, 40553 independent reflections, $R_{\text{int}} = 0.0373$, the final R was 0.0370 ($I > 2\theta(I)$, 26403 reflections) and $wR(F^2)$ was 0.0855 (all data), GOF 1.016.

4.4.4.28 Crystal data for **1OEt**

$C_{15}H_{13}NO_2Se$, $M = 318.22$, $T = 112(18)$ K, $\lambda = 1.54184$ Å, monoclinic, space group $P2_1/c$ (no. 14), $a = 18.5506(5)$ Å, $b = 6.1578(2)$ Å, $c = 12.3151(3)$ Å, $\beta = 101.778(3)^\circ$, $V = 1377.15(7)$ Å³, $Z = 4$. $D_c = 1.535$ mg M⁻³, $\mu(\text{Cu K}\alpha) = 3.671$ mm⁻¹, $F(000) = 640$,

FIGURE 4.60: X-ray crystal structure of **10Me·DMAP**, polymorph B.

crystal size $0.03 \times 0.115 \times 0.205$ mm. 9007 reflections measured, $2\theta_{\max} = 153.694^\circ$, 2836 independent reflections, $R_{\text{int}} = 0.0445$, the final R was 0.0564 ($I > 2\sigma(I)$, 2580 reflections) and $wR(F^2)$ was 0.1617 (all data), GOF 1.111.

FIGURE 4.61: X-ray crystal structure of **10Et**. The ethyl group is disordered over two sites.

4.4.4.29 Crystal data for **10Et·DMAP**

$C_{22}H_{23}N_3O_2Se$, $M = 440.39$, $T = 100.00(10)$ K, $\lambda = 1.54184$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 14.3280(3)$ Å, $b = 14.6892(3)$ Å, $c = 20.0386(3)$ Å, $\alpha = 83.051(2)^\circ$, $\beta = 72.057(2)^\circ$, $\gamma = 82.557(2)^\circ$, $V = 3963.81(14)$ Å 3 , $Z = 8$. $D_c = 1.476$ mg M $^{-3}$, $\mu(\text{Cu } K\alpha) = 2.749$ mm $^{-1}$, $F(000) = 1808$, crystal size $0.196 \times 0.239 \times 0.513$ mm. 54565 reflections measured, $2\theta_{\max} = 155.022^\circ$, 16338 independent reflections, $R_{\text{int}} = 0.0392$, the final R was 0.0375 ($I > 2\sigma(I)$, 14382 reflections) and $wR(F^2)$ was 0.1121 (all data), GOF 1.097.

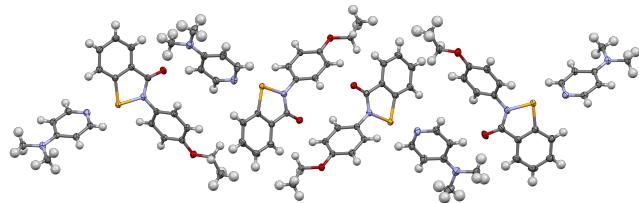


FIGURE 4.62: X-ray crystal structure of **1OEt·DMAP**.

4.4.4.30 Crystal data for **1OEt·4a**

$C_{24}H_{25}N_3O_2Se$, $M = 466.43$, $T = 101(1)$ K, $\lambda = 1.54184$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 9.7799(3)$ Å, $b = 12.7158(5)$ Å, $c = 18.9949(6)$ Å, $\alpha = 75.672(3)^\circ$, $\beta = 77.214(3)^\circ$, $\gamma = 68.133(3)^\circ$, $V = 2102.03(14)$ Å³, $Z = 4$. $D_c = 1.474$ mg M⁻³, $\mu(\text{Cu K}\alpha) = 2.626$ mm⁻¹, $F(000) = 960$, crystal size $0.049 \times 0.347 \times 0.451$ mm. 27336 reflections measured, $2\theta_{\max} = 154.508^\circ$, 8651 independent reflections, $R_{\text{int}} = 0.0487$, the final R was 0.0484 ($I > 2\sigma(I)$, 7620 reflections) and $wR(F^2)$ was 0.1388 (all data), GOF 1.131.

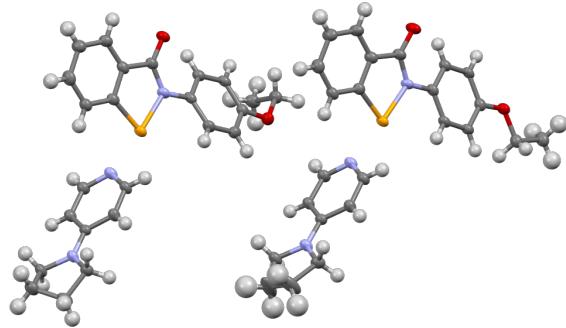


FIGURE 4.63: X-ray crystal structure of **1OEt·4a**.

4.4.4.31 Crystal data for **1OEt·4b**

$C_{24}H_{25}N_3O_3Se$, $M = 482.43$, $T = 101(1)$ K, $\lambda = 1.54184$ Å, triclinic, space group $P\bar{1}$ (no. 2), $a = 10.1213(3)$ Å, $b = 12.3068(3)$ Å, $c = 18.9297(6)$ Å, $\alpha = 75.580(2)^\circ$, $\beta = 77.135(2)^\circ$, $\gamma = 71.395(3)^\circ$, $V = 2137.77(11)$ Å³, $Z = 4$. $D_c = 1.499$ mg M⁻³, $\mu(\text{Cu K}\alpha) = 2.639$ mm⁻¹, $F(000) = 992$, crystal size $0.079 \times 0.142 \times 0.216$ mm. 27505

reflections measured, $2\theta_{\max} = 154.896^\circ$, 8820 independent reflections, $R_{\text{int}} = 0.0439$, the final R was 0.0923 ($I > 2\theta(I)$, 7842 reflections) and $wR(F^2)$ was 0.2668 (all data), GOF 1.145.

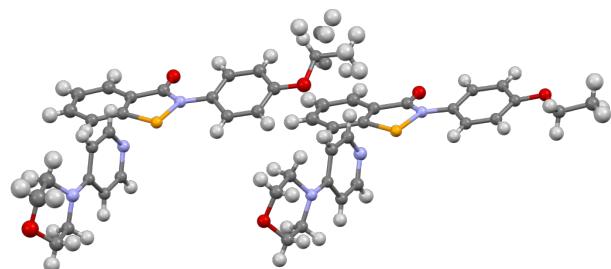


FIGURE 4.64: X-ray crystal structure of **1OEt·4b**. One of the ethyl groups is disordered over two sites.

References

- (1) Laurence, C.; Queignec-Cabanetos, M.; Wojtkowiak, B. *Canadian Journal of Chemistry* **1983**, *61*, 135–138,
DOI: [10.1139/v83-024](https://doi.org/10.1139/v83-024).
- (2) Sarwar, M. G.; Dragisic, B.; Salsberg, L. J.; Gouliaras, C.; Taylor, M. S. *Journal of the American Chemical Society* **2010**, *132*, 1646–1653,
DOI: [10.1021/ja9086352](https://doi.org/10.1021/ja9086352).
- (3) Berthelot, M.; Laurence, C.; Safar, M.; Besseau, F. *Journal of the Chemical Society, Perkin Transactions 2* **1998**, 283–290,
DOI: [10.1039/a706696a](https://doi.org/10.1039/a706696a).
- (4) Heinrich, M. R.; Klisa, H. S.; Mayr, H.; Steglich, W.; Zipse, H. *Angewandte Chemie International Edition* **2003**, *42*, 4826–4828,
DOI: [10.1002/anie.200352289](https://doi.org/10.1002/anie.200352289).
- (5) Kleemiss, F. et al. *Chem. Sci.* **2021**, *12*, 1675–1692,
DOI: [10.1039/D0SC05526C](https://doi.org/10.1039/D0SC05526C).

- (6) Hansch, C.; Leo, A.; Taft, R. W. *Chemical Reviews* **1991**, *91*, 165–195,
DOI: [10.1021/cr00002a004](https://doi.org/10.1021/cr00002a004).
- (7) Reynolds, W. F. *Journal of the Chemical Society, Perkin Transactions 2* **1980**,
985–991,
DOI: [10.1039/P29800000985](https://doi.org/10.1039/P29800000985).
- (8) Jaffé, H. H. *Chemical Reviews* **1953**, *53*, 191–261,
DOI: [10.1021/cr60165a003](https://doi.org/10.1021/cr60165a003).
- (9) Dunitz, J. D.; Maverick, E. F.; Trueblood, K. N. *Atomic Motions in Molecular Crystals from Diffraction Measurements*, 1988,
DOI: [10.1002/anie.198808801](https://doi.org/10.1002/anie.198808801).
- (10) Morcombe, C. R.; Zilm, K. W. *J. Magn. Reson.* **2003**, *162*, 479–486,
DOI: [10.1016/S1090-7807\(03\)00082-X](https://doi.org/10.1016/S1090-7807(03)00082-X).
- (11) Schreckenbach, G.; Ziegler, T. *The Journal of Physical Chemistry* **1995**, *99*,
606–611,
DOI: [10.1021/j100002a024](https://doi.org/10.1021/j100002a024).
- (12) Schreckenbach, G.; Ziegler, T. *International Journal of Quantum Chemistry* **1996**, *60*, 753–766,
DOI: [10.1002/\(SICI\)1097-461X\(1996\)60:3<753::AID-QUA4>3.0.CO;2-W](https://doi.org/10.1002/(SICI)1097-461X(1996)60:3<753::AID-QUA4>3.0.CO;2-W).
- (13) Duddeck, H. *Prog. Nucl. Magn. Reson. Spectrosc.* **1995**, *27*, 1–323,
DOI: [10.1016/0079-6565\(94\)00005-F](https://doi.org/10.1016/0079-6565(94)00005-F).
- (14) Foyle, É. M.; White, N. G. *CrystEngComm* **2020**, *22*, 2526–2536,
DOI: [10.1039/d0ce00241k](https://doi.org/10.1039/d0ce00241k).
- (15) Gilday, L. C.; White, N. G.; Beer, P. D. *Dalton Transactions* **2013**, *42*, 15766–
15773,
DOI: [10.1039/c3dt52093e](https://doi.org/10.1039/c3dt52093e).
- (16) Garrett, G. E.; Gibson, G. L.; Straus, R. N.; Seferos, D. S.; Taylor, M. S. *Journal of the American Chemical Society* **2015**, *137*, 4126–4133,
DOI: [10.1021/ja512183e](https://doi.org/10.1021/ja512183e).

- (17) Thordarson, P. *Chem. Soc. Rev.* **2011**, *40*, 1305–1323,
DOI: [10.1039/C0CS00062K](https://doi.org/10.1039/C0CS00062K).
- (18) R Core Team R: A Language and Environment for Statistical Computing, Vienna, Austria, 2019.
- (19) Blackstock, S. C.; Lorand, J. P.; Kochi, J. K. *Journal of Organic Chemistry* **1987**, *52*, 1451–1460,
DOI: [10.1021/jo00384a013](https://doi.org/10.1021/jo00384a013).
- (20) Emsley, J. *Chemical Society Reviews* **1980**, *9*, 91–124,
DOI: [10.1039/CS9800900091](https://doi.org/10.1039/CS9800900091).
- (21) Armentrout, P. B. In *Modern Mass Spectrometry*; Springer: Berlin, Heidelberg, 2003, pp 233–262,
DOI: [10.1007/b10468](https://doi.org/10.1007/b10468).
- (22) Rodgers, M. T.; Armentrout, P. B. *Mass Spectrometry Reviews* **2000**, *19*, 215–247,
DOI: [10.1002/1098-2787\(200007\)19:4<215::AID-MAS2>3.0.CO;2-X](https://doi.org/10.1002/1098-2787(200007)19:4<215::AID-MAS2>3.0.CO;2-X).
- (23) Narancic, S.; Bach, A.; Chen, P. *Journal of Physical Chemistry A* **2007**, *111*, 7006–7013,
DOI: [10.1021/jp0720921](https://doi.org/10.1021/jp0720921).
- (24) Sheldrick, G. M. *Acta Crystallographica Section C: Structural Chemistry* **2015**, *71*, 3–8,
DOI: [10.1107/S2053229614024218](https://doi.org/10.1107/S2053229614024218).
- (25) Dupont, L.; Dideberg, O.; Jacquemin, P. *Acta Crystallographica Section C Crystal Structure Communications* **1990**, *46*, 484–486,
DOI: [10.1107/s0108270189007894](https://doi.org/10.1107/s0108270189007894).

Part III

Chalcogen bonding at oxygen

Chapter 5

Experimental evidence of Chalcogen bonding at oxygen.

This chapter was published in Chem. Commun., 13 Feb 2020.^{1†}

5.1 Abstract

An *o*-nitro-O-aryl oxime was observed to exhibit a short O ··· O contact, which exhibited characteristics consistent with a chalcogen bond. The O–N bond length of the oxime was appreciably longer than the expected value, and NBO calculations indicated the presence of a $n(O) \rightarrow \sigma^*(O-N)$ orbital delocalisation. Topological analysis of the experimental electron density of two analogues shows the presence of a bond path between the two oxygen atoms, with $\rho(r)$ and $\nabla^2\rho(r)$ values consistent with an electrostatic interaction. Finally, electrostatic potential calculations indicate the presence of a σ -hole, the “smoking gun” indicating a Ch-bond. These results are unusual as oxygen is not typically considered to be a Ch-bond donor, especially in unactivated systems such as oximes.

[†]Compound numbering, section titles, and terminology have been updated to fit this thesis.

5.2 Introduction

Chalcogen bonding (Ch-bonding) is a weak interaction of the broader family of σ -hole interactions, in which a chalcogen atom adopts some electrophilic character, allowing it to form a stabilised complex with a Lewis base. Much work has been directed towards applying Ch-bonding principles to fields such as supramolecular engineering, catalysis, and anion sensing.^{2–7} Typically only the heavier chalcogen atoms form Ch-bonds (tellurium, selenium or sulfur, the latter having particular relevance to protein folding).⁸ The reason for this is that the electron cloud of the chalcogen must be distorted by an electron withdrawing substituent in order to form the electrophilic σ -hole. The larger and more polarisable the chalcogen atom, the more electrophilic the σ -hole, and the stronger the Ch-bond.⁹

Oxygen, as the second most electronegative element with its extremely low polarisability, is not considered to be a strong Ch-bond donor, although recent theoretical studies indicate that it can form a σ -hole in species such as OF_2 and FNO_3 .^{10,11} We report here a compound which appears to contain an intramolecular $\text{O}\cdots\text{O}$ Ch-bond, which was synthesised and crystallised as part of an investigation into the Beckmann rearrangement.¹²

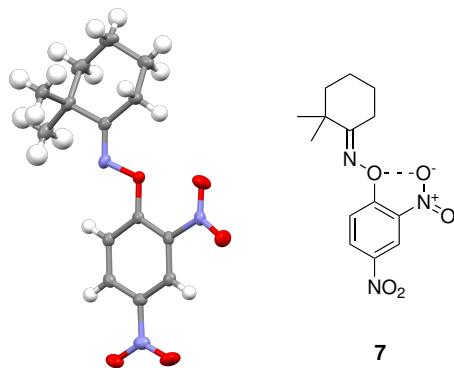


FIGURE 5.1: Oxime 7 displays a close $\text{O}\cdots\text{O}$ contact, which has characteristics consistent with a Ch-bond.

5.3 Results and Discussion

The compound **7** is an *o*-nitro-O-aryl oxime, in which the nitro group constitutes the Ch-bond acceptor, and the oxime is the Ch-bond donor. Although the coplanar geometry of the oxime and nitro groups is expected on the grounds of favourable conjugation of these groups into the aromatic ring, there are a number of signs that suggest that there is an additional stabilising interaction between the groups, namely, the Ch-bond.

5.3.1 Structural effects attributable to the Ch-bond

TABLE 5.1: Selected geometric parameters of oximes **7**, **8** and **9**. The two geometries in the asymmetric unit of **8** are indicated as **8a** and **8b**.

Compound	$r(O_1 \cdots O_2)$ Å	$r(N_1O_1)$ Å	$r(N_2O_2)$ Å	$\angle(O_2 \cdots O_1N_1)$ °	$\angle(C_1C_2N_2O_2)$ °
7	2.619(2)	1.489(2)	1.208(2)	161.80(8)	-3.5(2)
8a	2.5423(8)	1.4556(7)	1.2264(9)	160.56(5)	-0.7(1)
8b	2.5522(8)	1.4522(7)	1.223(1)	160.29(4)	-0.2(1)
9	2.6390(8)	1.4480(7)	1.2266(8)	148.60(4)	-30.94(8)

Firstly, bond lengths measured from the crystal structure deviate somewhat from expected values. The O ··· O Ch-bond distance was measured to be 2.619(2) Å, which is well within the van der Waals radii of two oxygen atoms (3.04 Å).¹³ The O–N bond of the oxime is also lengthened appreciably. Comparisons with other O-aryl oximes are convoluted by the significant effect of the electronic properties of the phenyl ring on the oxime bond length. However, an expected value can be calculated using the acidity of the corresponding 2,4-dinitro phenol ($pK_a = 5.91$) and the structural correlation given in eq. (5.1).^{12,14}

$$r_{N-OR}(\text{\AA}) = 1.467 - (3.20 \times 10^{-3}) \times pK_a(\text{ROH}) \quad (5.1)$$

This gives an expected bond length of 1.448 Å versus the measured bond length 1.489(2) Å ($\Delta = 0.041$ Å = 13.7σ).

This is consistent with the charge transfer model of Ch-bonding, as the lone pair of the nitro oxygen donor is delocalised into the $\sigma^*(O-N)$ anti bonding orbital, leading to a weakening and lengthening of this bond.¹⁵

5.3.2 NBO calculations

Secondly, DFT calculations were carried out on the solid phase geometry of 7 in order to ascertain the nature of the O ··· O close contact within the NBO (Natural Bond Orbital) and QTAIM (Quantum Theory of Atoms In Molecules) frameworks.^{16,17} The two frameworks are complementary descriptions of bonding within molecules. The calculations were performed at the M06-2X/aug-cc-pVQZ level as implemented in the Gaussian16 suite.^{18–20} This has been shown to give accurate densities for Ch-bonded complexes.²¹

It is customary to optimise structures so they are at a gas-phase minimum, ensuring that any derived energies, vibrational modes, and other quantities are reliable. In this case, however, we were unable to locate a minimum corresponding to the crystal geometry; the nitro group was forced out of the plane of the aryl ring, disrupting the requisite Ch-bond geometry. We note that the potential energy surface of this torsion is quite shallow, so the lack of a satisfactory minimum is likely due to the absence of crystal packing. Regardless, a thorough investigation was considered to be outside of the scope of this preliminary study, therefore solid phase geometries were used instead, with the caveat that certain quantities are likely to be unreliable.

The NBO calculations revealed the presence of a $n(O) \rightarrow \sigma^*(O-N)$ orbital delocalisation, which is one criterion by which a Ch-bond can be defined.²² As the geometry at which the NBO analysis was performed was not a gas phase minimum, orbital energies are inaccurate, and so the usual estimate of the strength of a delocalisation

(E^2) should not be used. We instead measure the magnitude of the delocalisation using the off-diagonal element of the Fock matrix $F(i, j)$, from which the energy E^2 is calculated using eq. (5.2).

$$E^2 = q_i \frac{F(i, j)^2}{\epsilon_j - \epsilon_i} \quad (5.2)$$

The value of $F(i, j)$ for the $n(O) \rightarrow \sigma^*(O-N)$ orbital delocalisation in **7** is 0.023 a.u., indicating a small delocalisation.

5.3.3 QTAIM analysis of electron density

Bader's Quantum Theory of Atoms In Molecules (QTAIM) describes bonding in terms of the observable quantity of electron density and its topology. We analysed the DFT density of oxime **7**, which revealed the presence of a bond path between the two oxygen atoms. Bond paths are associated with bond critical points (bcps), where the gradient vector of the electron density is zero.

The values of both the electron density $\rho(r)$ and the Laplacian of the electron density $\nabla^2\rho(r)$ at the bcp are diagnostic of certain types of bonds. Bcps with a small $\rho(r)$ and positive $\nabla^2\rho(r)$ (indicating local depletion of electron density at the bcp) are characteristic of ionic bonds or purely electrostatic interactions. The values of $\rho(r)$ and $\nabla^2\rho(r)$ respectively are 0.0161 and +0.0790 (in a.u.), suggesting this is primarily a closed shell electrostatic interaction, and that the NBO delocalisation, although present, is secondary.

These observations piqued our interest in the potential for Ch-bonding at oxygen, so we decided to extend our investigation to verify that this interaction was not simply a quirk of the initial oxime.

5.3.4 CSD search for similar structures

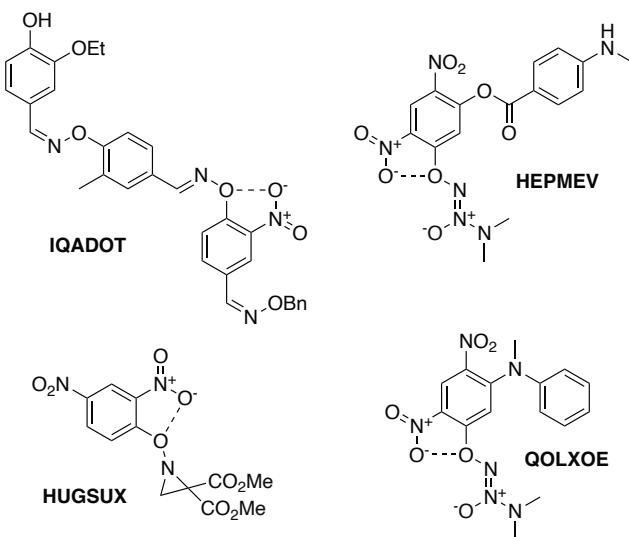


FIGURE 5.2: Structures of CSD matches for the *o*-nitro-O-aryl oxime motif.

TABLE 5.2: Selected geometric parameters of structures **HEPMEV**, **HUGSUX**, **QOLXOE**, and **IQADOT**.

Compound	r(O ₁ ⋯O ₂) Å	r(N ₁ –O ₁) Å	r(N ₂ –O ₂) Å	∠(O ₂ ⋯O ₁ –N ₁) °	∠(C ₁ –C ₂ –N ₂ –O ₂) °
HEPMEV	2.554(5)	1.426(4)	1.222(4)	161.2(2)	-15.4(6)
HUGSUX	2.563(2)	1.461(2)	1.220(2)	158.4(1)	20.8(2)
QOLXOE	2.557(4)	1.419(4)	1.216(5)	158.7(2)	-10.9(6)
IQADOT^a	2.532(2)	1.445(2)	1.207(3)	154.4(1)	-9.2(3)
IQADOT^b	—	1.433(2)	—	—	—

A search of the Cambridge Structural Database for *o*-nitro-O-aryl oximes afforded several results.²³ Structures HEPMEV, HUGSUX, and QOLXOE all contained similar motifs to 7, with a nitro group approximately coplanar with an *ortho* oxime or oxime-like group.^{24–26} Structure IQADOT was particularly interesting, as it contains two electronically similar oximes, only one of which has an *ortho* nitro group.²⁷ The difference in bond lengths between them is significant (1.445(2) Å versus 1.433(2) Å, $\Delta = 0.012$ Å = 6σ), and the same trend is observed as in 7. Relevant structural parameters are given in table 5.2.

5.3.5 Analysis of analogues and determination of experimental electron density

Two analogues of oxime **7** (**8** and **9**, fig. 5.3) were prepared, forming crystals which gave data of sufficient quality for multipole refinement, allowing us to analyse the experimental electron density as opposed to the calculated DFT density.²⁸ This refinement was performed using the MoProSuite software package.²⁹ **8** crystallised in a similar coplanar geometry to **7**, exhibiting a Ch-bond*, while **9** displayed a torsion of the nitro group, disrupting the requisite geometry. This torsion can be attributed to crystal packing forces (taken to be worth 0.5–2 kcal/mol) disrupting this extremely weak interaction.³⁰

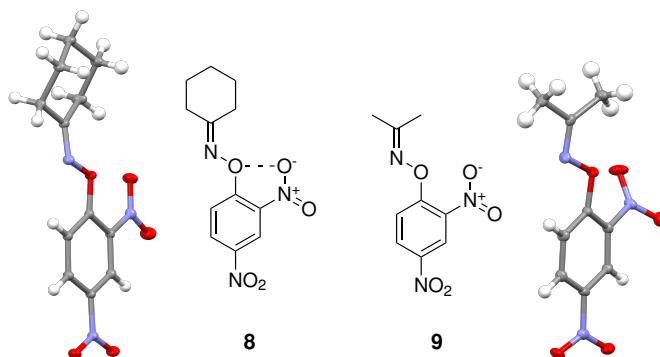


FIGURE 5.3: Oxime **8** adopts a similar conformation to **7**, while the nitro group of **9** is twisted, leading to poor alignment with the oxime.

The oxime bond lengths differ significantly between **8** and **9** (1.4556(7)–1.4522(7) versus 1.4480(7) Å, $\Delta = 0.0042$ Å = 6σ), supporting the suggestion that the n(O) \rightarrow $\sigma^*(O-N)$ orbital delocalisation plays a role in these Ch-bonds. NBO analysis of these geometries confirmed this, with $F(i,j)$ values of 0.026 a.u. for **8**, and an absence of any delocalisation in **9**.

QTAIM analysis of the experimental density revealed the presence of a bond path and bcp in **8** but not in **9**, showing that there is indeed an interaction present when the geometry allows. In **8**, the values of $\rho(r)$ and $\nabla^2\rho(r)$ respectively are 0.018 and

*The asymmetric unit contained two molecules, however both had similar geometries, and in the interests of simplicity values for only one molecule are presented.

+0.094 (in a.u.), which agree well with the DFT values calculated for **7**, and are consistent with a closed-shell origin for the interaction (fig. 5.4).

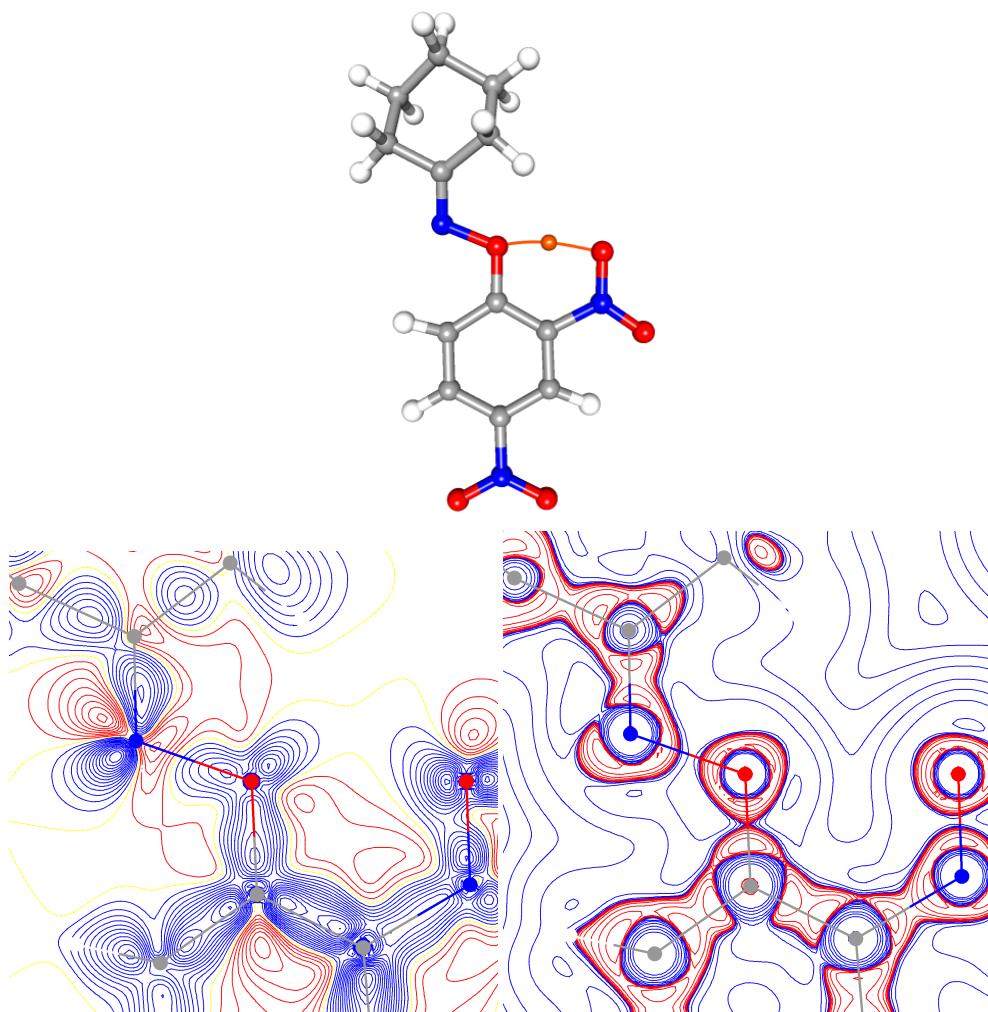


FIGURE 5.4: Deformation and Laplacian maps of the experimental electron density for **8**. Negative values depicted by red contours, and positive by blue. Above is shown the bond path and bcp in orange.

The NCI (non-covalent interaction) index is an extension of QTAIM, which characterises interactions based on the reduced density gradient and the second eigenvalue of the electron density Hessian.³¹ This can be used to construct informative maps, showing surfaces where non-covalent interactions appear to be occurring. NCI maps were computed for **8** and **9** based on the experimental electron density, and these are shown in fig. 5.5.

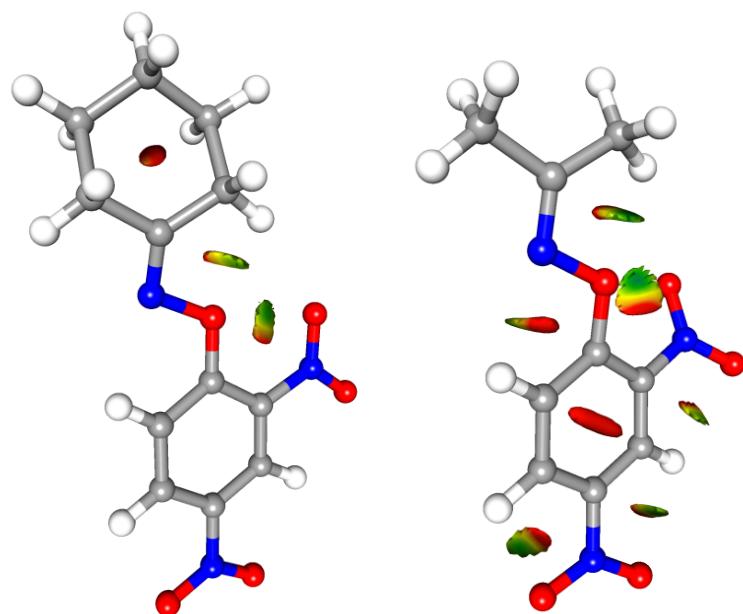


FIGURE 5.5: NCI maps for **8** and **9**. Positive values (non-bonding) are shown in red, and negative values (attractive) are shown in blue.

To provide further evidence for the interaction, we took advantage of the fact that derivative **9** does not display a Ch-bond, and searched for a σ -hole in the experimental electron density. Due to the crowded surroundings of the oxime group, we were unable to visualise the 0.001 a.u. isosurface as would be typical for ESP analysis. The extension of the oxime bond (therefore, the σ -hole) was simply not visible. We instead mapped the electrostatic potential onto intersecting planes through the oxime bond, which clearly showed an positive region along the extension of this bond consistent with a σ -hole (fig. 5.6).

When investigating such weak interactions as these, it is important to consider the limitations of each method of analysis. For example, the presence of a topological bond path within the QTAIM framework does not necessarily imply the presence of an attractive bonding interaction. It may simply represent the inevitable consequence of placing two electron clouds in close proximity.^{32–36} However, in this case, the NBO calculations, QTAIM and electrostatic potential analysis of the experimental electron densities are all consistent with the statistically significant lengthening of the oxime N–O bond, and all signs point toward a true σ -hole interaction.

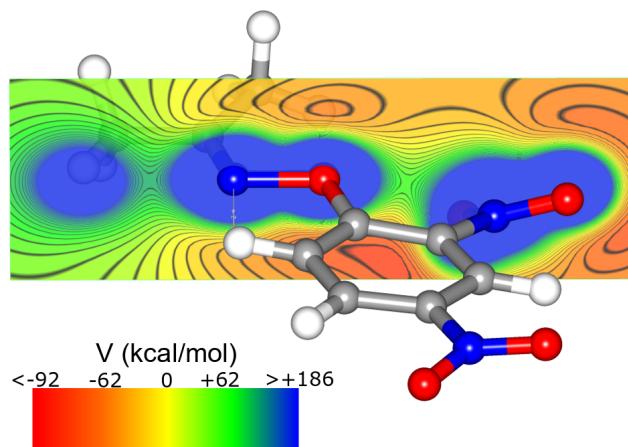


FIGURE 5.6: Electrostatic potential mapped onto a plane through the oxime bond. The σ -hole is visible as a green (positive ESP) elongation of the nuclear potential towards the nitro group to the right. Note that the oxime nitrogen also displays a σ -hole to the left.

We believe this is the first documented experimental evidence of a σ -hole interaction at oxygen, and it is remarkable that it occurs in such air- and water-stable molecules as these oximes. We therefore challenge the consensus that Ch-bonding can only occur at highly activated oxygen atoms, and suggest that there may be implications when considering the reactivity of trace molecules such as peroxides, and nitric and nitrous oxides in biological systems.

5.4 Supplementary material

5.4.1 Synthesis

5.4.1.1 Preparation of *O*-(2,4-dinitrophenyl) oximes **8** and **9**

Cyclohexanone oxime (556.8 mg, 4.920 mmol) and sodium hydride (60% in mineral oil, 249.5 mg, 6.237 mmol) were dissolved in anhydrous THF (10 mL) and stirred for 5 minutes. This was then added to a solution of 2,4-dinitro-1-fluorobenzene (950.6 mg, 5.108 mmol) in a further 10 mL anhydrous THF. This was stirred for 1 h

at room temperature, then diluted with water (100 mL) and extracted with ethyl acetate (2×30 mL). The combined organic phases were washed with brine (30 mL), dried (MgSO_4) and evaporated to afford **8** as a waxy pale yellow solid (895.1 mg, 65%, m.p. 100.2–100.9° C).

Acetone oxime **9** was isolated as a pale yellow powder, yield 72%, m.p. 88.5–88.9° C.

Crystals suitable for x-ray diffraction analysis were grown by slow diffusion of pentane into a saturated solution of the compound in dichloromethane.

5.4.2 Crystallographic data

5.4.2.1 Crystal data for **7**

$\text{C}_{14}\text{H}_{17}\text{N}_3\text{O}_5$ ($M = 307.30$ g/mol): monoclinic, space group $\text{P}2_1/c$ (no. 14), $a = 7.17630(10)$ Å, $b = 28.9373(4)$ Å, $c = 7.06700(10)$ Å, $\beta = 92.1860(10)$ °, $V = 1466.48(4)$ Å³, $Z = 4$, $T = 130.15$ K, $\mu(\text{Cu K}\alpha) = 0.902$ mm⁻¹, $D_{\text{calc}} = 1.392$ g/cm³, 10358 reflections measured ($12.234^\circ \leq 2\theta \leq 158.922^\circ$), 2902 unique ($R_{\text{int}} = 0.0231$, $R_{\text{sigma}} = 0.0149$) which were used in all calculations. The final R_1 was 0.0472 ($I > 2\sigma(I)$) and wR_2 was 0.1242 (all data).

5.4.2.2 Crystal data for **8**

$\text{C}_{12}\text{H}_{13}\text{N}_3\text{O}_5$ ($M = 279.25$ g/mol): monoclinic, space group $\text{P}2_1$ (no. 4), $a = 6.66740(10)$ Å, $b = 16.3925(3)$ Å, $c = 11.6108(2)$ Å, $\beta = 101.0310(10)$ °, $V = 1245.56(4)$ Å³, $Z = 4.00$, $Z' = 2.00$, $T = 100(1)$ K, $\mu(\text{Mo K}\alpha) = 0.118$ mm⁻¹, $D_{\text{calc}} = 1.489$ g/cm³, 51580 reflections measured ($7.016^\circ \leq 2\theta \leq 102.638^\circ$), 25111 unique ($R_{\text{int}} = 0.0214$, $R_{\text{sigma}} = 0.0317$) which were used in all calculations. The final R_1 was 0.0409 ($I > 2\sigma(I)$) and wR_2 was 0.1153 (all data). Flack parameter = 0.32(14).

5.4.2.3 Crystal data for 9

$C_9H_9N_3O_5$ ($M = 239.175$ g/mol): orthorhombic, space group $P2_12_12_1$ (no. 19), $a = 5.49110(10)$ Å, $b = 10.5173(2)$ Å, $c = 18.2619(3)$ Å, $V = 1054.65(3)$ Å 3 , $Z = 4$, $T = 100.00(10)$ K, $\mu(\text{Mo K}\alpha) = 0.125$ mm $^{-1}$, $D_{\text{calc}} = 1.506$ g/cm 3 , 24292 reflections measured ($5.908^\circ \leq 2\theta \leq 102.41^\circ$), 10696 unique ($R_{\text{int}} = 0.0224$, $R_{\text{sigma}} = 0.0326$) which were used in all calculations. The final R_1 was 0.0378 ($I > 2\sigma(I)$) and wR_2 was 0.1032 (all data). Flack parameter = -0.02(19).

References

- (1) Fellowes, T.; Harris, B.; White, J. *Chemical Communications* **2020**, 56, 3313–3316,
DOI: [10.1039/c9cc09896h](https://doi.org/10.1039/c9cc09896h).
- (2) Riwar, L. J.; Trapp, N.; Root, K.; Zenobi, R.; Diederich, F. *Angewandte Chemie - International Edition* **2018**, 57, 17259–17264,
DOI: [10.1002/anie.201812095](https://doi.org/10.1002/anie.201812095).
- (3) Mahmudov, K. T.; Kopylovich, M. N.; Guedes Da Silva, M. F. C.; Pombeiro, A. J. *Dalton Transactions* **2017**, 46, 10121–10138,
DOI: [10.1039/c7dt01685a](https://doi.org/10.1039/c7dt01685a).
- (4) Wonner, P.; Dreger, A.; Vogel, L.; Engelage, E.; Huber, S. M. *Angewandte Chemie - International Edition* **2019**, 58, 17082–17082,
DOI: [10.1002/anie.201913472](https://doi.org/10.1002/anie.201913472).
- (5) Ho, P. C.; Szydlowski, P.; Sinclair, J.; Elder, P. J. W.; Kübel, J.; Gendy, C.; Lee, L. M.; Jenkins, H.; Britten, J. F.; Morim, D. R.; Vargas-Baca, I. *Nature Communications* **2016**, 7, 11299,
DOI: [10.1038/ncomms11299](https://doi.org/10.1038/ncomms11299).

- (6) Kremer, A.; Fermi, A.; Biot, N.; Wouters, J.; Bonifazi, D. *Chemistry (Weinheim an der Bergstrasse, Germany)* **2016**, *22*, 5665–5675,
DOI: [10.1002/chem.201504328](https://doi.org/10.1002/chem.201504328).
- (7) Benz, S.; Mareda, J.; Besnard, C.; Sakai, N.; Matile, S. *Chemical Science* **2017**, *8*, 8164–8169,
DOI: [10.1039/C7SC03866F](https://doi.org/10.1039/C7SC03866F).
- (8) Iwaoka, M.; Isozumi, N. *Molecules* **2012**, *17*, 7266–7283,
DOI: [10.3390/molecules17067266](https://doi.org/10.3390/molecules17067266).
- (9) Murray, J. S.; Lane, P.; Politzer, P. *International Journal of Quantum Chemistry* **2008**, *108*, 2770–2781,
DOI: [10.1002/qua.21753](https://doi.org/10.1002/qua.21753).
- (10) Varadwaj, P. R.; Varadwaj, A.; Marques, H. M.; MacDougall, P. J. *Physical Chemistry Chemical Physics* **2019**, *21*, 19969–19986,
DOI: [10.1039/c9cp03783g](https://doi.org/10.1039/c9cp03783g).
- (11) Varadwaj, P. R. *Molecules* **2019**, *24*, 3166,
DOI: [10.3390/molecules24173166](https://doi.org/10.3390/molecules24173166).
- (12) Yeoh, S. D.; Harris, B. L.; Simons, T. J.; White, J. M. *Australian Journal of Chemistry* **2012**, *65*, 905–917,
DOI: [10.1071/CH12079](https://doi.org/10.1071/CH12079).
- (13) Bondi, A. *Journal of Physical Chemistry* **1964**, *68*, 441–451,
DOI: [10.1021/j100785a001](https://doi.org/10.1021/j100785a001).
- (14) Socrates, G. *Transactions of the Faraday Society* **1970**, *66*, 1052–1057,
DOI: [10.1039/TF9706601052](https://doi.org/10.1039/TF9706601052).
- (15) Reed, A. E.; Curtiss, L. A.; Weinhold, F. *Chemical Reviews* **1988**, *88*, 899–926,
DOI: [10.1021/cr00088a005](https://doi.org/10.1021/cr00088a005).
- (16) Bader, R. F. *Chemical Reviews* **1991**, *91*, 893–928,
DOI: [10.1021/cr00005a013](https://doi.org/10.1021/cr00005a013).

- (17) Glendening, E. D.; Badenhoop, J. K.; Reed, A. E.; Carpenter, J. E.; Bohmann, J. A.; Morales, C. M.; Karafiloglou, P.; Landis, C. R.; Weinhold, F. NBO 7.0, Madison, 2018.
- (18) Frisch, M. J. et al. Gaussian16 Revision C.01, 2016.
- (19) Zhao, Y.; Truhlar, D. G. *Theoretical Chemistry Accounts* **2008**, *120*, 215–241, DOI: [10.1007/s00214-007-0310-x](https://doi.org/10.1007/s00214-007-0310-x).
- (20) Woon, D. E.; Dunning, T. H. *The Journal of Chemical Physics* **1995**, *103*, 4572–4585, DOI: [10.1063/1.470645](https://doi.org/10.1063/1.470645).
- (21) Kim, Y.; Song, S.; Sim, E.; Burke, K. *Journal of Physical Chemistry Letters* **2019**, *10*, 295–301, DOI: [10.1021/acs.jpclett.8b03745](https://doi.org/10.1021/acs.jpclett.8b03745).
- (22) Pascoe, D. J.; Ling, K. B.; Cockroft, S. L. *Journal of the American Chemical Society* **2017**, *139*, 15160–15167, DOI: [10.1021/jacs.7b08511](https://doi.org/10.1021/jacs.7b08511).
- (23) Groom, C. R.; Bruno, I. J.; Lightfoot, M. P.; Ward, S. C. *Acta Crystallographica Section B: Structural Science, Crystal Engineering and Materials* **2016**, *72*, 171–179, DOI: [10.1107/S2052520616003954](https://doi.org/10.1107/S2052520616003954).
- (24) Saavedra, J. E.; Srinivasan, A.; Bonifant, C. L.; Chu, J.; Shanklin, A. P.; Flippenn-Anderson, J. L.; Rice, W. G.; Turpin, J. A.; Davies, K. M.; Keefer, L. K. *Journal of Organic Chemistry* **2001**, *66*, 3090–3098, DOI: [10.1021/jo0016529](https://doi.org/10.1021/jo0016529).
- (25) Kostyanovsky, R. G.; Schurig, V.; Trapp, O.; Lyssenko, K. A.; Averkiev, B. B.; Kadorkina, G. K.; Prosyanyik, A. V.; Kostyanovsky, V. R. *Mendeleev Communications* **2002**, *12*, 137–140, DOI: [10.1070/MC2002v012n04ABEH001612](https://doi.org/10.1070/MC2002v012n04ABEH001612).

- (26) Saavedra, J. E. et al. *Journal of Medicinal Chemistry* **2006**, *49*, 1157–1164,
DOI: [10.1021/jm050700k](https://doi.org/10.1021/jm050700k).
- (27) Renaudet, O.; Reymond, J. L. *Organic Letters* **2003**, *5*, 4693–4696,
DOI: [10.1021/o1035867r](https://doi.org/10.1021/o1035867r).
- (28) Hansen, N. K.; Coppens, P. *Acta Crystallographica Section A* **1978**, *34*, 909–921,
DOI: [10.1107/S0567739478001886](https://doi.org/10.1107/S0567739478001886).
- (29) Jelsch, C.; Guillot, B.; Lagoutte, A.; Lecomte, C. *Journal of Applied Crystallography* **2005**, *38*, 38–54,
DOI: [10.1107/S0021889804025518](https://doi.org/10.1107/S0021889804025518).
- (30) Dunitz, J. D.; Gavezzotti, A. *Chemical Society Reviews* **2009**, *38*, 2622–2633,
DOI: [10.1039/b822963p](https://doi.org/10.1039/b822963p).
- (31) Johnson, E. R.; Keinan, S.; Mori-Sánchez, P.; Contreras-García, J.; Cohen, A. J.;
Yang, W. *Journal of the American Chemical Society* **2010**, *132*, 6498–6506,
DOI: [10.1021/ja100936w](https://doi.org/10.1021/ja100936w).
- (32) Spackman, M. A. *Chemical Physics Letters* **1999**, *301*, 425–429,
DOI: [10.1016/S0009-2614\(99\)00071-8](https://doi.org/10.1016/S0009-2614(99)00071-8).
- (33) Gatti, C. *Zeitschrift fur Kristallographie* **2005**, *220*, 399–457,
DOI: [10.1524/zkri.220.5.399.65073](https://doi.org/10.1524/zkri.220.5.399.65073).
- (34) Bader, R. F. *Journal of Physical Chemistry A* **2009**, *113*, 10391–10396,
DOI: [10.1021/jp906341r](https://doi.org/10.1021/jp906341r).
- (35) Cerpa, E.; Krapp, A.; Vela, A.; Merino, G. *Chemistry - A European Journal* **2008**,
14, 10232–10234,
DOI: [10.1002/chem.200800934](https://doi.org/10.1002/chem.200800934).
- (36) Cerpa, E.; Krapp, A.; Flores-Moreno, R.; Donald, K. J.; Merino, G. *Chemistry - A European Journal* **2009**, *15*, 1985–1990,
DOI: [10.1002/chem.200801399](https://doi.org/10.1002/chem.200801399).

Part IV

Applications of Ch-bonding

Chapter 6

Thermal rearrangement of a Ch-bonded solvate

This chapter was published in Cryst. Eng. Comm., 26 May 2020, and was originally titled *Thermal conversion of a pyridine solvate to a de-solvate facilitated by rearrangement of chalcogen bonds. The solvate and non-solvate structures of N-(2-nitro-4-(3-oxobenzo[d][1,2]selenazol-2(3H)-yl)phenyl)picolinamide.*^{1†}

6.1 Abstract

The pyridine solvate of benzisoselenazolinone **13**·pyridine is characterised by planar sheets of the benzisoselenazolinone **13** pierced by channels of pyridine molecules at an angle of 133°, the pyridine molecules are held in place by N···Se chalcogen bonding to the isoselenazolinone moiety. These channels, which extend through the structure to the surface of the crystal, provide a means for escape of pyridine from the lattice when the crystal is heated to ca. 100°C. Upon loss of the pyridine from these channels the remaining molecules undergo rearrangement to fill the

[†]Compound numbering, section titles, and terminology have been updated to fit this thesis.

space and in doing so the N···Se chalcogen bond in **13**·pyridine is replaced by a C=O···Se chalcogen bond to give the non solvate **13**(ex.DMF). The geometry of the chalcogen bond requires that the two benzisoselenazolinone ring systems which are essentially coplanar in **13**·pyridine twist by an angle of 138° resulting in the formation of highly corrugated sheets in the non solvate.

6.2 Introduction

Chalcogen bonding (Ch-bonding) is an attractive non-covalent interaction between a Lewis base and a chalcogen atom bearing an electron withdrawing substituent (X) (fig. 6.1);^{2–4} it is directional and has a strength similar to hydrogen bonding. Chalcogen bonding has applications in fields as diverse as medicinal chemistry,^{5–9} anion sensing,^{10–12} materials chemistry,¹³ supramolecular chemistry,^{3,14–18} and catalysis.¹⁹ In medicinal chemistry the chalcogen bond is considered as an isostere to N–H···A hydrogen bonding (fig. 6.1), this property is currently being exploited in the development of new pharmaceuticals.

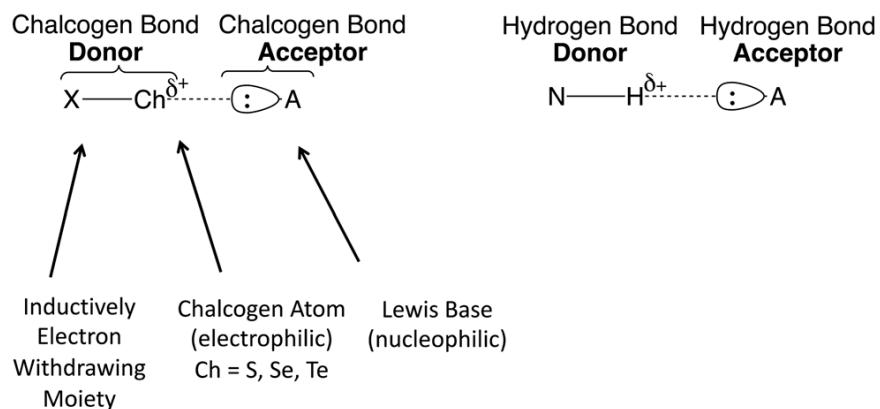


FIGURE 6.1: Chalcogen bonding model, and similarity to H-bonding.

The mechanism of chalcogen bonding is believed to involve both electrostatic and orbital interaction components with dispersion being a lesser contributor. The electrostatic component involves attraction between the Lewis base and a positively

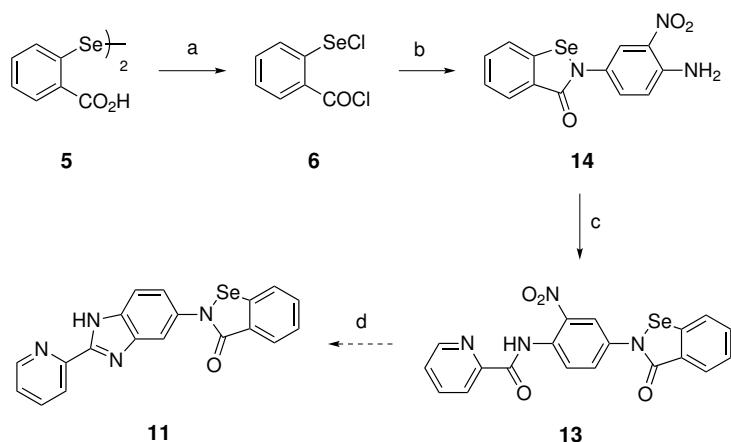
charged σ -hole which is generated along the extension of the Ch–X bond due to polarisation of the bonding orbital, while the orbital interaction component involves mixing between the occupied lone pair orbital of the Lewis base and the vacant $\sigma_{\text{Ch}-\text{X}}^*$ bond on the chalcogen, both these interactions account for the directionality of this interaction to different extents. Whereas the electrostatic σ -hole interaction is believed to be the main contributor to the closely related halogen bonding interactions,^{20–25} significant lengthening of the Ch–X bond observed in the crystal structures of a number of chalcogen bonded systems is suggestive of a significant charge-transfer component to this interaction.^{26,27}

6.3 Results and discussion

As part of our efforts towards a chalcogen-bonding based DNA minor-groove binding agent we required the benzisoselenazolinone **13**, which we proposed to cyclise to the benzimidazole-substituted benzisoselenazolinone **11**, an analog to the Hoechst-type DNA-binding bis-benzimidazoles.^{28–30}

6.3.1 Synthesis

The benzisoselenazolinone **13** was prepared from the diselenide **5** by conversion to the bis-electrophilic selenium reagent **6**,³¹ followed by condensation with 2-nitro-1,4-benzenediamine to give the benzisoselenazolinone **14**, which was then coupled to picolinic acid via a Yamaguchi intermediate (scheme 6.1).



SCHEME 6.1: Synthesis of precursor **13**. (a) SOCl_2 , cat. DMF, reflux, 30 min; (b) 2-nitro-1,4-benzenediamine, Et_3N , THF, rt, 18 h, 61%; (c) Picolinic acid, TCBC, Et_3N , rt, 24 h, 20%; (d) [H], H^+ .

6.3.2 Structural characterisation

Benzisoselenazolinone **13** was found to be of low solubility in most organic solvents, but very small orange needles were obtained from slow evaporation from dimethylformamide. The crystal structure obtained using data collected at the Australian Synchrotron was that of a non-solvate form, referred to herein as **13(ex.DMF)** in the orthorhombic space group *Pbca*. A thermal ellipsoid plot for **13(ex.DMF)** is presented in fig. 6.2.

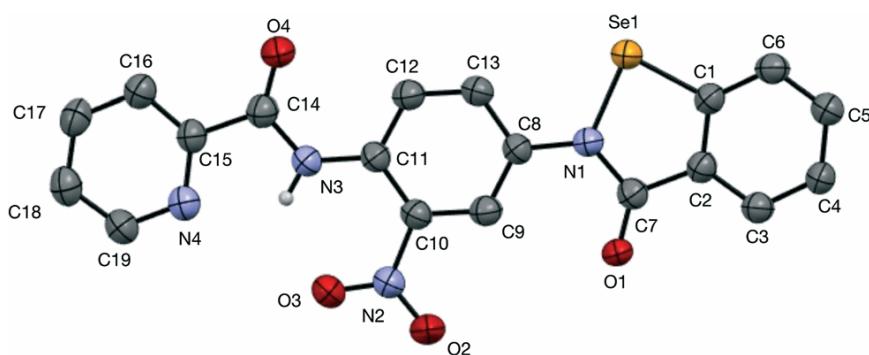


FIGURE 6.2: Thermal ellipsoid plot of **13(ex.DMF)**. Ellipsoids are at the 50% probability level.

The crystal packing in **13(ex.DMF)** is characterised by a number of non-covalent interactions including π -stacking which forms columns extending down the *b* axis

between molecules of **13** related by the *b* glide plane (fig. 6.3). The planes defined by the central aromatic ring C8-C13 are inclined at an angle 10.3(2) $^{\circ}$ to the adjacent π -stacked molecule, with a centroid (C8-C13) plane distance of 3.414(4) Å and a centroid–centroid distance of 3.786(4) Å representing a slip distance of 1.636 Å.

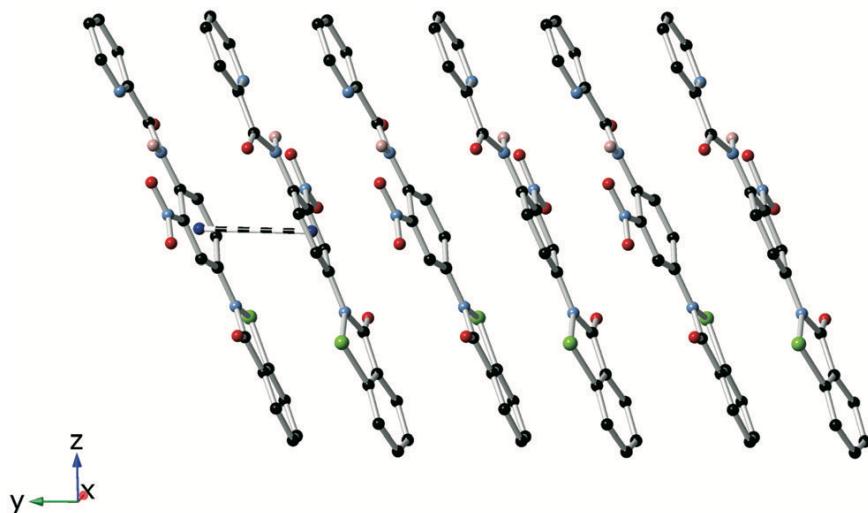


FIGURE 6.3: Offset π -stacking of **13**(ex.DMF) extending down the *b*-axis. The centroid-centroid distance is 3.786(4) Å.

Adjacent π -stacked columns are held together by chalcogen bonding interactions between the isoselenazolinone amide carbonyl oxygen and the selenium atom in the heterocyclic ring forming chains generated by the *a*-glide plane extending down the *a*-axis. The O \cdots Se distance is 2.659(3) Å and the O \cdots Se–N angle is 173.6(2) $^{\circ}$. The angle between the planes defined by the benzisoselenazolinone rings in the chalcogen bonded pairs is 138.8 $^{\circ}$ a coplanar arrangement is presumably disfavoured as this would result in severe steric clashes with the C5 of the benzisoselenazolinone ring. Similar O \cdots Se chalcogen bonding interactions have been observed in other benzisoselenazolinone derivatives related to the drug ebselen with comparable geometries.^{26,32–34} The N3–H3 group which is flanked by the pyridine nitrogen and the nitro group engages in intramolecular N–H \cdots N and N–H \cdots O hydrogen bonds, but is not involved in any significant intermolecular interactions (fig. 6.4, fig. 6.5, fig. 6.6).

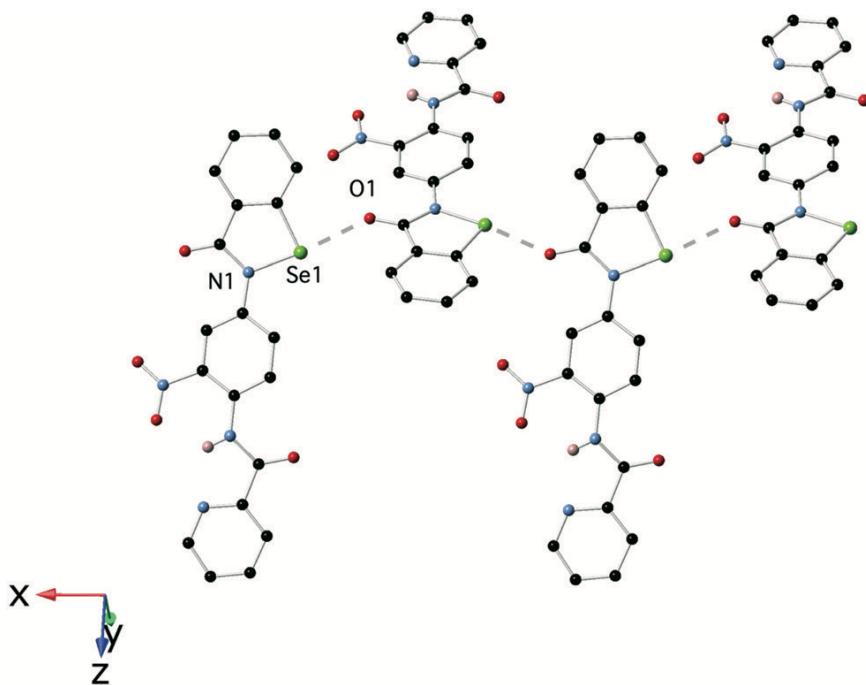


FIGURE 6.4: O \cdots Se chalcogen bonding interactions in **13**(ex.DMF).

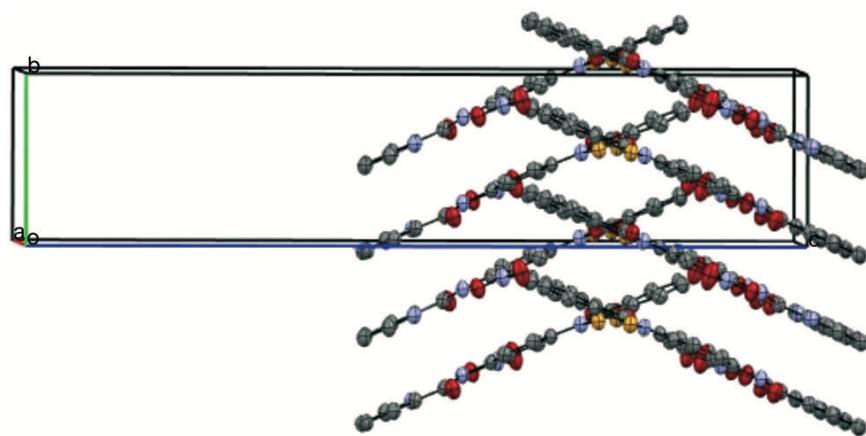


FIGURE 6.5: 2-D layers of **13**(ex.DMF) π -stacking extends along the *b*-axis while the Se \cdots O chalcogen bond interactions extend down the *a*-axis.

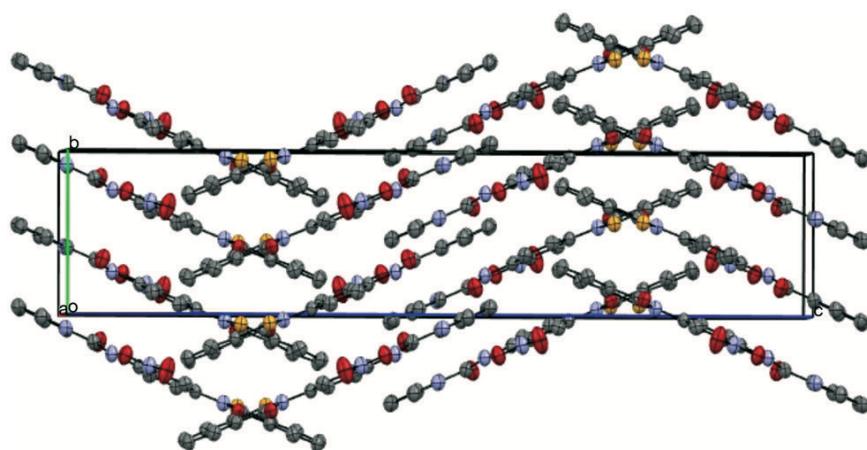


FIGURE 6.6: Extension of the 2-D layers of **13**(ex.DMF) into 3-D by van der Waals interactions.

The combination of the π -stacking and chalcogen bonding generates a 2-dimensional network lying parallel to the *ab* plane. These layers are extended into 3-dimensions by weaker Van-der Waals contacts between molecules related by the *c* glide.

Crystallisation of benzisoselenazolinone **13** from pyridine gave rise to orange needles which were found to diffract well on the home-source diffractometer. The structure solved in the monoclinic space group $P\bar{c}$ and was found to be a pyridine solvate, herein labelled **13**·pyridine. The structure of **13**·pyridine is presented in fig. 6.7 and shows that the pyridine solvate forms a $N \cdots Se$ chalcogen bond between the pyridine solvent molecule and the isoselenazolinone moiety. The $N5 \cdots Se1$ distance is $2.466(5)$ Å, the $N5 \cdots Se1 - N1$ angle is $173.2(2)^\circ$, and the angle between the plane of the coordinated pyridine ring and the benzisoselenazolinone ring is $89.2(1)^\circ$. This geometry compares with previously reported chalcogen bonded adducts involving dimethylaminopyridine with simple benzisoselenazolinones.²⁶ It is interesting to note that upon formation of the chalcogen bond to pyridine, there is a significant lengthening of the $Se1 - N1$ bond distance from $1.910(3)$ Å in **13**(ex.DMF) to $1.945(5)$ Å in **13**·pyridine, which is consistent with the expected structural effects arising from the charge-transfer component of chalcogen-bonding.^{26,27}

Molecules of **13** assemble into planar sheets lying parallel to the $(\bar{1}04)$ plane, the

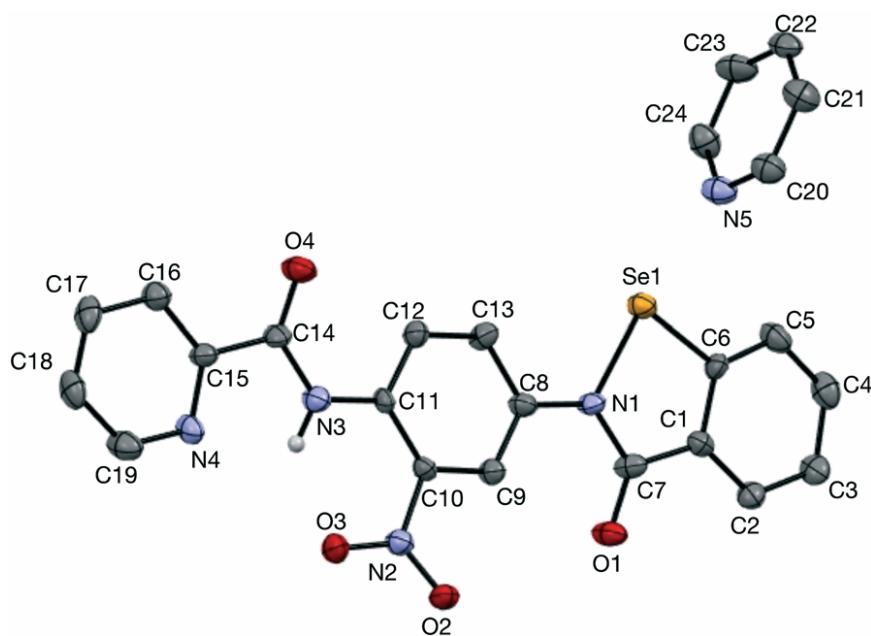


FIGURE 6.7: Thermal ellipsoid plot of **13**·pyridine. Ellipsoids are at the 50% probability level.

distance between these sheets as defined by the distance between the centroid of the atoms C8–C13 and the adjacent plane is 3.336 Å while the centroid-centroid distance is 4.973 Å representing a slippage of 3.688 Å (fig. 6.8, fig. 6.9). The parallel sheets of molecules of **13** are pierced by channels of chalcogen bonded pyridine molecules which run parallel to the *a*-axis at an angle of approximately 133° to the plane.

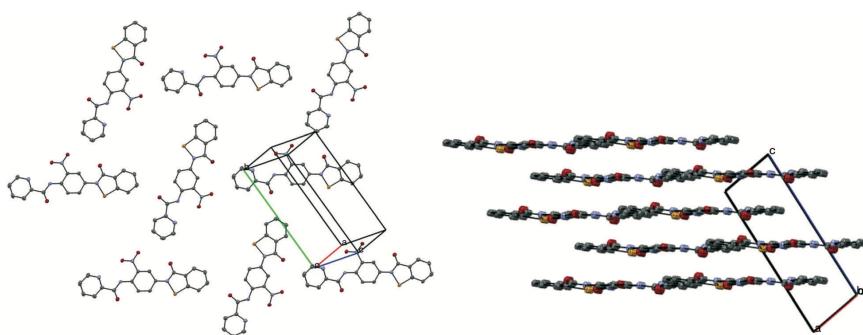


FIGURE 6.8: Sheets of compound **13** viewed from orthogonal and parallel directions. Pyridine solvate has been excluded.

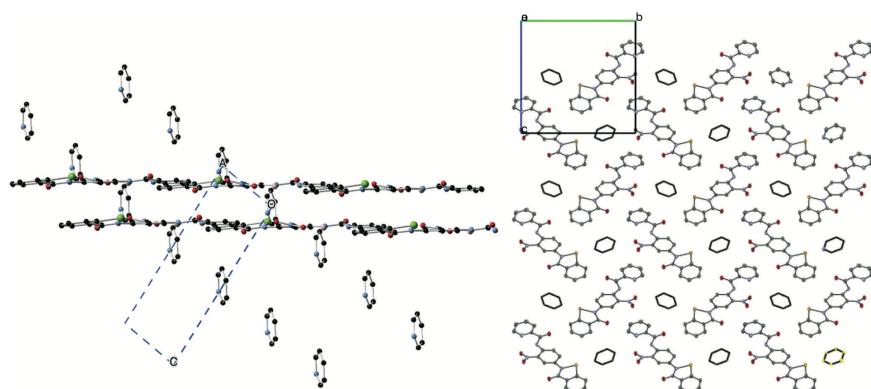


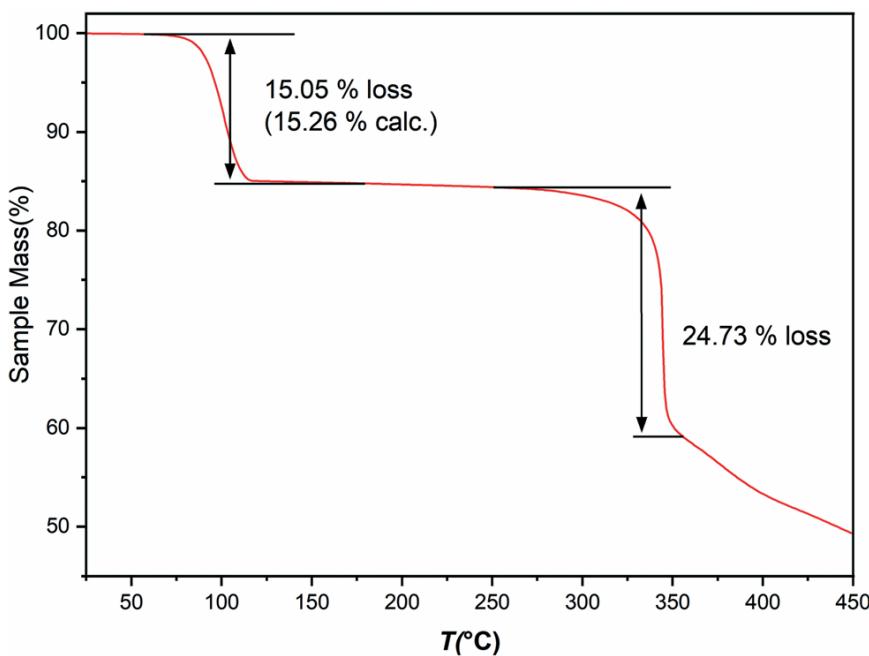
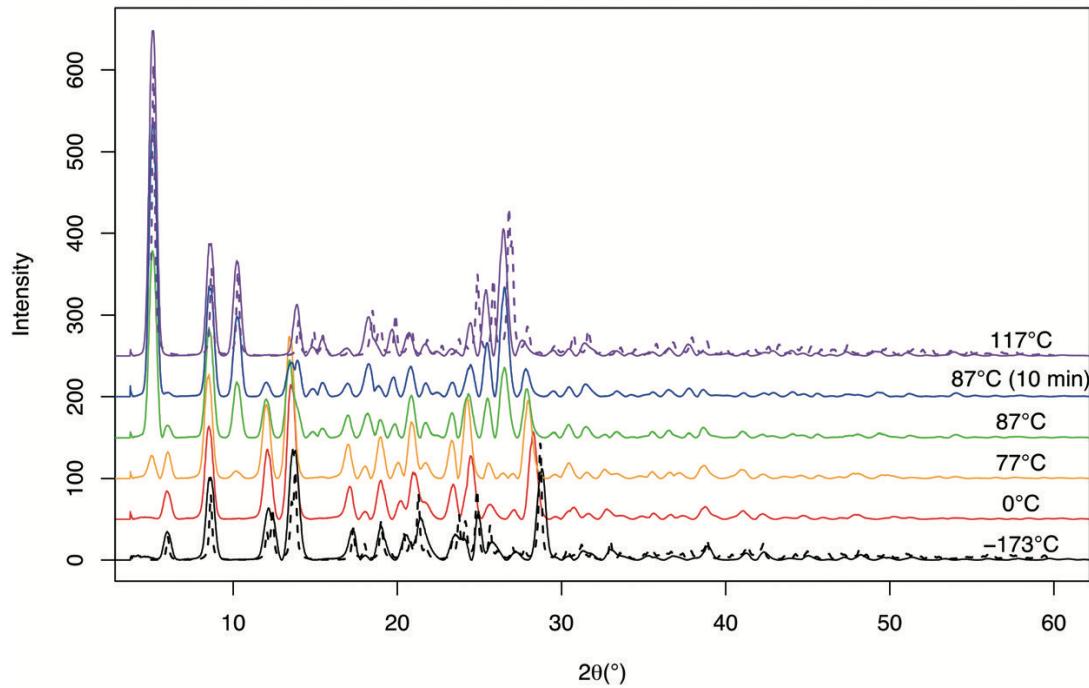
FIGURE 6.9: Two orthogonal views of parallel sheets of compound **13**, pierced by channels of pyridine molecules running parallel to the a -axis.

6.3.3 Variable temperature studies

Thermal gravimetric analysis of the pyridine solvate **13**·pyridine was conducted on a Mettler TGA/SDTA851 apparatus in 40 μL aluminium crucibles. A mass loss of 15.04% occurred between 90–110°C corresponding to the loss of the pyridine solvate (calc. 15.26%) followed by a second loss of 24.73% between 300–360°C which is consistent with the loss of 121 a.m.u. very likely associated with the Pyr-C(O)NH moiety (fig. 6.10).

The observation that both the non-solvate **13**(ex.DMF) and **13**·pyridine had the same melting point (experimental section below) intrigued us to establish whether desolvation of **13**·pyridine which occurs between 90–120°C results in conversion to non-solvate **13**(ex.DMF). Thus we carried out variable temperature powder X-ray diffraction measurements on **13**·pyridine from -173°C to 117°C. The resulting diffractograms are shown in fig. 6.11.

From 87°C to 117°C when pyridine is known to be lost from the sample from the TGA analysis, there is a smooth change from one crystalline phase to another. Furthermore comparison of the final diffractogram obtained after being kept at 117°C for 10 minutes very closely matches the calculated powder pattern for the non-solvate **13**(ex.DMF) suggesting the transformation from **13**·pyridine into **13**(ex.DMF). The same experiment was applied to a single crystal of **13**·pyridine glued on a glass fibre

FIGURE 6.10: TGA analysis of **13**·pyridine.FIGURE 6.11: Variable temperature powder XRD patterns from **13**·pyridine, calculated powder pattern for **13**·pyridine and **13**(ex.DMF) are the dotted lines in the top and bottom traces respectively.

to establish whether this transformation occurs from a single crystal of **13**·pyridine to a single crystal of **13**(ex.DMF). The crystal was heated to 90°C and heated at a rate of 0.5°C per minute to 120°C. While there was a steady decrease in the intensity of the reflections for **13**·pyridine, individual reflections consistent with a single crystal of **13**(ex.DMF) were not observed, but rather, there was the development of the powder pattern for **13**(ex.DMF). Interestingly, throughout this transformation the crystal morphology did not appear to change significantly, but the final diffraction pattern was clearly that of a powder. It is likely that the transformation (which must begin at the surface of the crystal) results in fragmentation of daughter crystals of the non-solvate, thus eroding away at the mother crystal. We believe that a single crystal to single crystal transformation is very unlikely as collapsing of the channels containing the pyridine solvate would prevent complete desolvation. A plausible mechanism for this interconversion likely involves replacement of the N···Se chalcogen bond in **13**·pyridine, with a O···Se chalcogen bond involving the isoselenazolinone carbonyl group from a molecule in an adjacent layer which is at a distance of 12.051 Å (fig. 6.12).

6.4 Conclusion

The structure of **13**·pyridine is characterised by planar sheets of the benzisoselenazolinone **13** pierced by channels of pyridine molecules at an angle 133°, the pyridine molecules are held in place by N···Se chalcogen bonding to the isoselenazolinone moiety. These channels which extend through the structure to the surface of the crystal provide a means for escape of pyridine from the lattice when the crystal is heated to ca. 100°C. Upon loss of the pyridine from these channels the remaining molecules undergo rearrangement to fill the space and in doing so the N···Se chalcogen bond in **13**·pyridine is replaced by a C=O···Se chalcogen bond to give the non solvate **13**(ex.DMF). The geometry of the chalcogen bond requires that the two benzisoselenazolinone ring systems which are essentially coplanar in **13**·pyridine

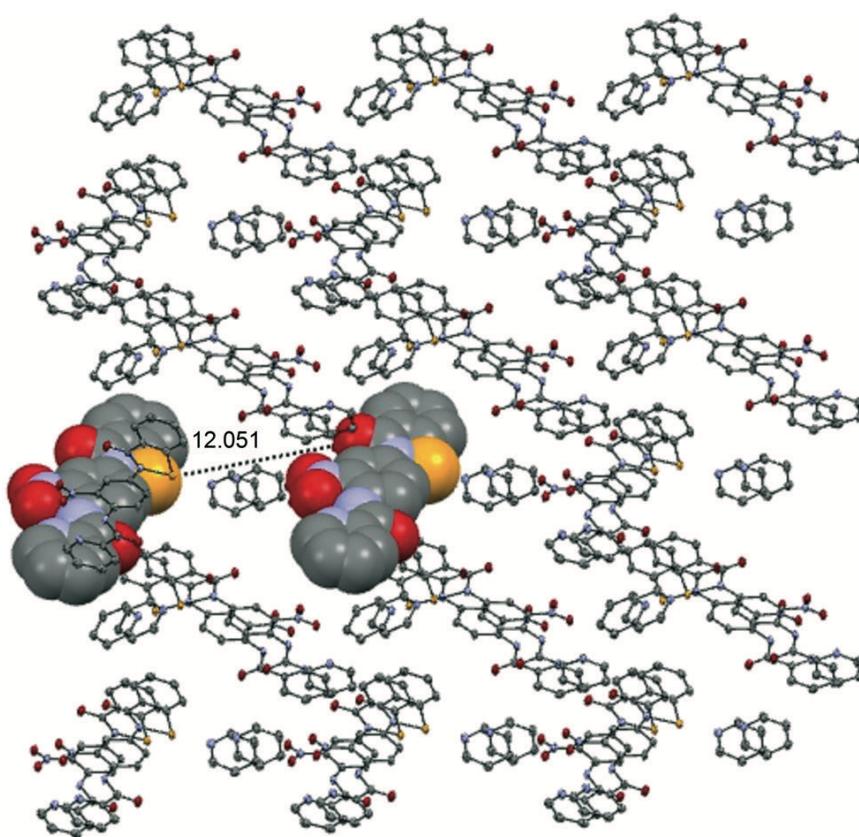


FIGURE 6.12: Interlayer benzisoselenazolinones **13** believed to form a O⋯Se chalcogen bond upon desolvation of **13**·pyridine.

twist by an angle of 138° resulting in the formation of highly corrugated sheets in the non solvate.

6.5 Experimental

6.5.1 Synthesis

6.5.1.1 Preparation of 2-(4-amino-3-nitrophenyl)benzo[*d*][1,2]-selenazol-3(2*H*)-one **14**.

Diselenide **5** (1.2025 g, 3.005 mmol) was dissolved in thionyl chloride (20 mL) and refluxed for 30 min, and the colour changed from purple to light yellow. Excess

thionyl chloride was removed *in vacuo*, and the residue dissolved in anhydrous THF (50 mL). To this was added a solution of 2-nitro-1,4-benzenediamine (991.9 mg, 6.477 mmol) and triethylamine (dist. from CaH₂, 3 mL) in anhydrous THF (50 mL). This mixture was stirred at room temperature for 18 h, then filtered, washing the precipitate with water, to afford 2-(4-amino-3-nitro-phenyl)benzo[*d*][1,2]-selenazol-3(2*H*)-one **14** as a brick red solid (940 mg, 61%, m.p. 300.8–302.0°C (DMF)).

¹H NMR (400 MHz, *d*₆-DMSO) δ ppm 7.08 (d, *J* = 9.00 Hz, 1 H), 7.46 (t, *J* = 7.43 Hz, 1 H), 7.55 (s, 2 H), 7.59–7.70 (m, 2 H), 7.86 (d, *J* = 7.43 Hz, 1 H), 8.06 (d, *J* = 7.83 Hz, 1 H), 8.12 (d, *J* = 1.96 Hz, 1 H).

¹³C NMR (100 MHz, *d*₆-DMSO) δ ppm 120.29, 121.49, 126.32, 126.71, 127.77, 128.31, 128.45, 129.68, 132.66, 134.13, 139.33, 145.03, 165.70.

MS (ESI +ve) m/z 335.9882 (MH⁺) C₁₃H₁₀N₃O₃Se⁺ requires 335.9881 (Δ=0.3 ppm).

6.5.1.2 Preparation of N-(2-nitro-4-(3-oxobenzo[*d*][1,2]selenazol-2(3*H*)-yl)phenyl) picolinamide **13**.

Picolinic acid (256.6 mg, 2.084 mmol) was dissolved in anhydrous THF (5 mL) and triethylamine (dist. from CaH₂, 1 mL), then trichlorobenzoyl chloride (325 μL) was added and the mixture stirred for 10 min. The above nitroaniline **14** (242.8 mg, 0.956 mmol) was then added, and the mixture stirred under argon for 24 h, during which time the dark red colour faded to give a yellow solution. The mixture was tipped into water (100 mL) and filtered to afford a yellow precipitate, which was recrystallised from pyridine (50 mL) to give N-(2-nitro-4-(3-oxobenzo[*d*][1,2]selenazol-2(3*H*)-yl)phenyl)picolinamide **13** as yellow needles (93.2 mg, 20%), m.p. 342.7–343.7°C (*d*). While recrystallization by slow evaporation from DMF gave small yellow needles m.p. 342–344°C (*d*).

¹H NMR (500 MHz, *d*₆-DMSO) δ ppm 7.51 (t, *J* = 7.40 Hz, 1 H) 7.66–7.83 (m, 1 H) 7.94 (d, *J* = 7.63 Hz, 1 H) 8.04 (dd, *J* = 7.8, 2.4 Hz, 1 H) 8.09–8.17 (m, 2 H) 8.23 (d, *J* = 7.78 Hz, 1 H) 8.68 (d, *J* = 2.44 Hz, 1 H) 8.73 (d, *J* = 9.00 Hz, 1 H) 8.81 (d, *J* = 4.43 Hz, 1 H) 12.22 (s, 1 H).

MS (ESI +ve) m/z 441.010 (MH⁺) C₁₉H₁₃N₄O₄Se⁺ requires 441.0096 (Δ=0.9 ppm).

6.5.2 Crystallography

Intensity data for **13**·pyridine was collected on a Rigaku XtalLAB Synergy at 100.0(1) K. Data for **13**(ex.DMF) was collected on the MX1 beam line³⁵ at the Australian Synchrotron. The temperature was maintained using an Oxford Cryostream cooling device. The structures were solved by direct methods and difference Fourier synthesis.³⁶ Thermal ellipsoid plots were generated using the program Mercury³⁷ integrated within the WINGX³⁸ suite of programs. Other figures were obtained with both Mercury software and Crystal Maker software.

6.5.2.1 Crystal data for **13**(ex.DMF)

$C_{19}H_{12}N_4O_4Se$, $M = 439.29$, $T = 100.0$ K, $\lambda = 0.82656$ Å, orthorhombic, space group Pbc_a, $a = 12.652(3)$, $b = 7.5250(15)$, $c = 34.367(7)$ Å, $V = 3272.0(11)$ Å³, $Z = 8$, $D_c = 1.784$ mg M⁻³, $\mu = 3.383$ mm⁻¹, $F(000) = 1760$, crystal size $0.05 \times 0.005 \times 0.005$ mm³, 35758 reflections measured, $\theta_{\max} = 32.28^\circ$, 3282 independent reflections [R(int) = 0.0803], the final R was 0.0601 [$I > 2\sigma(I)$, 2513 reflections] and $wR(F^2)$ was 0.1802 (all data), GOF 1.037.

6.5.2.2 Crystal data for **13**·pyridine

$C_{19}H_{12}N_4O_4Se \cdot C_5H_5N$, $M = 518.39$, $T = 100.0$ K, $\lambda = 1.54184$ Å, monoclinic, space group P_c, $a = 4.9726(2)$, $b = 14.6836(5)$, $c = 14.4255(5)$ Å, $\beta = 98.154(4)^\circ$, $V = 1042.64(7)$ Å³, $Z = 2$, $D_c = 1.651$ mg M⁻³, $\mu(\text{Cu}-K\alpha) = 2.829$ mm⁻¹, $F(000) = 524$, crystal size $0.12 \times 0.033 \times 0.026$ mm³, 6794 reflections measured, $\theta_{\max} = 76.79^\circ$, 3132 independent reflections [R(int) = 0.0558], the final R was 0.0422 [$I > 2\sigma(I)$, 2977 data] and $wR(F^2)$ was 0.1107 (all data), GOF 1.098.

6.6 Acknowledgements

We gratefully acknowledge Sirtex Medical for funding and the Australian Synchrotron for beam time via the Collaborative Access Program (proposal 13618b). The Australian Research Council for Post Graduate Scholarships (TF and MPVK).

References

- (1) Fellowes, T.; Van Koeverden, M. M. P.; White, J. J. M. *CrystEngComm* **2020**, *22*, 4023–4029,
DOI: [10.1039/d0ce00662a](https://doi.org/10.1039/d0ce00662a).
- (2) Vogel, L.; Wonner, P.; Huber, S. M. *Angewandte Chemie International Edition* **2019**, *58*, 1880–1891,
DOI: [10.1002/anie.201809432](https://doi.org/10.1002/anie.201809432).
- (3) Bleiholder, C.; Werz, D. B.; Köppel, H.; Gleiter, R. *Journal of the American Chemical Society* **2006**, *128*, 2666–2674,
DOI: [10.1021/ja056827g](https://doi.org/10.1021/ja056827g).
- (4) Garrett, G. E.; Gibson, G. L.; Straus, R. N.; Seferos, D. S.; Taylor, M. S. *Journal of the American Chemical Society* **2015**, *137*, 4126–4133,
DOI: [10.1021/ja512183e](https://doi.org/10.1021/ja512183e).
- (5) Beno, B. R.; Yeung, K. S.; Bartberger, M. D.; Pennington, L. D.; Meanwell, N. A. *Journal of Medicinal Chemistry* **2015**, *58*, 4383–4438,
DOI: [10.1021/jm501853m](https://doi.org/10.1021/jm501853m).
- (6) Clark, T.; Hennemann, M.; Murray, J. S.; Politzer, P. *Journal of Molecular Modeling* **2007**, *13*, 291–296,
DOI: [10.1007/s00894-006-0130-2](https://doi.org/10.1007/s00894-006-0130-2).

- (7) Hudson, B. M.; Nguyen, E.; Tantillo, D. J. *Organic & Biomolecular Chemistry* **2016**, *14*, 3975–3980,
DOI: [10.1039/C6OB00254D](https://doi.org/10.1039/C6OB00254D).
- (8) Iwaoka, M.; Takemoto, S.; Tomoda, S. *Journal of the American Chemical Society* **2002**, *124*, 10613–10620,
DOI: [10.1021/ja026472q](https://doi.org/10.1021/ja026472q).
- (9) Reid, R. C.; Yau, M. K.; Singh, R.; Lim, J.; Fairlie, D. P. *Journal of the American Chemical Society* **2014**, *136*, 11914–11917,
DOI: [10.1021/ja506518t](https://doi.org/10.1021/ja506518t).
- (10) Lim, J. Y. C.; Marques, I.; Thompson, A. L.; Christensen, K. E.; Félix, V.; Beer, P. D. *Journal of the American Chemical Society* **2017**, *139*, 3122–3133,
DOI: [10.1021/jacs.6b12745](https://doi.org/10.1021/jacs.6b12745).
- (11) Garrett, G. E.; Carrera, E. I.; Seferos, D. S.; Taylor, M. S. *Chem. Commun.* **2016**, *52*, 9881–9884,
DOI: [10.1039/C6CC04818H](https://doi.org/10.1039/C6CC04818H).
- (12) Borissov, A.; Marques, I.; Lim, J. Y. C.; Félix, V.; Smith, M. D.; Beer, P. D. *Journal of the American Chemical Society* **2019**, jacs.9b00148,
DOI: [10.1021/jacs.9b00148](https://doi.org/10.1021/jacs.9b00148).
- (13) Biot, N.; Bonifazi, D. *Chemistry - A European Journal* **2018**, *24*, 5439–5443,
DOI: [10.1002/chem.201705428](https://doi.org/10.1002/chem.201705428).
- (14) Chen, L.; Xiang, J.; Zhao, Y.; Yan, Q. *Journal of the American Chemical Society* **2018**, *140*, 7079–7082,
DOI: [10.1021/jacs.8b04569](https://doi.org/10.1021/jacs.8b04569).
- (15) Ho, P. C.; Szydlowski, P.; Sinclair, J.; Elder, P. J. W.; Kübel, J.; Gendy, C.; Lee, L. M.; Jenkins, H.; Britten, J. F.; Morim, D. R.; Vargas-Baca, I. *Nature Communications* **2016**, *7*, 11299,
DOI: [10.1038/ncomms11299](https://doi.org/10.1038/ncomms11299).

- (16) Gleiter, R.; Haberhauer, G.; Werz, D. B.; Rominger, F; Bleiholder, C. *Chemical Reviews* **2018**, *118*, 2010–2041,
DOI: [10.1021/acs.chemrev.7b00449](https://doi.org/10.1021/acs.chemrev.7b00449).
- (17) Huynh, H.-T.; Jeannin, O.; Fourmigué, M. *Chem. Commun.* **2017**, 8467–8469,
DOI: [10.1039/C7CC04833E](https://doi.org/10.1039/C7CC04833E).
- (18) Gleiter, R.; Werz, D. B.; Rausch, B. J. *Chemistry - A European Journal* **2003**, *9*,
2676–2683,
DOI: [10.1002/chem.200204684](https://doi.org/10.1002/chem.200204684).
- (19) Wang, W.; Zhu, H.; Feng, L.; Yu, Q.; Hao, J.; Zhu, R.; Wang, Y. *Journal of the American Chemical Society* **2020**, *142*, 3117–3124,
DOI: [10.1021/jacs.9b12610](https://doi.org/10.1021/jacs.9b12610).
- (20) Präsang, C.; Whitwood, A. C.; Bruce, D. W. *Crystal Growth and Design* **2009**, *9*,
5319–5326,
DOI: [10.1021/cg900823d](https://doi.org/10.1021/cg900823d).
- (21) Sarwar, M. G.; Dragisic, B.; Salsberg, L. J.; Gouliaras, C.; Taylor, M. S. *Journal of the American Chemical Society* **2010**, *132*, 1646–1653,
DOI: [10.1021/ja9086352](https://doi.org/10.1021/ja9086352).
- (22) Beale, T. M.; Chudzinski, M. G.; Sarwar, M. G.; Taylor, M. S. *Chemical Society Reviews* **2013**, *42*, 1667–1680,
DOI: [10.1039/c2cs35213c](https://doi.org/10.1039/c2cs35213c).
- (23) Aakeröy, C. B.; Chopade, P. D.; Desper, J. *Crystal Growth and Design* **2013**, *13*,
4145–4150,
DOI: [10.1021/cg400988m](https://doi.org/10.1021/cg400988m).
- (24) Goud, N. R.; Bolton, O.; Burgess, E. C.; Matzger, A. J. *Crystal Growth and Design* **2016**, *16*, 1765–1771,
DOI: [10.1021/acs.cgd.6b00074](https://doi.org/10.1021/acs.cgd.6b00074).
- (25) Stone, A. J. *Journal of the American Chemical Society* **2013**, *135*, 7005–7009,
DOI: [10.1021/ja401420w](https://doi.org/10.1021/ja401420w).

- (26) Fellowes, T.; White, J. M. *CrystEngComm* **2019**, *21*, 1539–1542,
DOI: [10.1039/C8CE01853G](https://doi.org/10.1039/C8CE01853G).
- (27) Pascoe, D. J.; Ling, K. B.; Cockroft, S. L. *Journal of the American Chemical Society* **2017**, *139*, 15160–15167,
DOI: [10.1021/jacs.7b08511](https://doi.org/10.1021/jacs.7b08511).
- (28) Loewe, H.; Urbanietz, J. *Arzneimittel-Forschung* **1974**, *24*, 1927–1933.
- (29) Pjura, P. E.; Grzeskowiak, K.; Dickerson, R. E. *Journal of Molecular Biology* **1987**, *197*, 257–271,
DOI: [10.1016/0022-2836\(87\)90123-9](https://doi.org/10.1016/0022-2836(87)90123-9).
- (30) Martin, R. F.; Broadhurst, S.; Reum, M. E.; Squire, C. J.; Clark, G. R.; Lobachevsky, P. N.; White, J. M.; Clark, C.; Sy, D.; Spotheim-Maurizot, M.; Kelly, D. P. *Cancer Research* **2004**, *64*, 1067–1070,
DOI: [10.1158/0008-5472.CAN-03-2423](https://doi.org/10.1158/0008-5472.CAN-03-2423).
- (31) Lesser, R.; Weiβ, R. *Berichte der deutschen chemischen Gesellschaft (A and B Series)* **1924**, *57*, 1077–1082,
DOI: [10.1002/cber.19240570703](https://doi.org/10.1002/cber.19240570703).
- (32) Thomas, S. P.; Satheeshkumar, K.; Mugesh, G.; Guruow, T. N. *Chemistry - A European Journal* **2015**, *21*, 6793–6800,
DOI: [10.1002/chem.201405998](https://doi.org/10.1002/chem.201405998).
- (33) Bhabak, K. P.; Mugesh, G. *Chemistry - A European Journal* **2007**, *13*, 4594–4601,
DOI: [10.1002/chem.200601584](https://doi.org/10.1002/chem.200601584).
- (34) Piatek, M.; Oleksyn, B.; Sliwinski, J. *Acta Crystallographica Section C Crystal Structure Communications* **1995**, *51*, 298–301,
DOI: [10.1107/s0108270193012983](https://doi.org/10.1107/s0108270193012983).

- (35) Cowieson, N. P.; Aragao, D.; Clift, M.; Ericsson, D. J.; Gee, C.; Harrop, S. J.; Mudie, N.; Panjikar, S.; Price, J. R.; Riboldi-Tunnicliffe, A.; Williamson, R.; Caradoc-Davies, T. *Journal of Synchrotron Radiation* **2015**, *22*, 187–190, DOI: [10.1107/S1600577514021717](https://doi.org/10.1107/S1600577514021717).
- (36) Sheldrick, G. M. *Acta Crystallographica Section C: Structural Chemistry* **2015**, *71*, 3–8, DOI: [10.1107/S2053229614024218](https://doi.org/10.1107/S2053229614024218).
- (37) Macrae, C. F.; Bruno, I. J.; Chisholm, J. A.; Edgington, P. R.; McCabe, P.; Pidcock, E.; Rodriguez-Monge, L.; Taylor, R.; Van De Streek, J.; Wood, P. A. *Journal of Applied Crystallography* **2008**, *41*, 466–470, DOI: [10.1107/S0021889807067908](https://doi.org/10.1107/S0021889807067908).
- (38) Farrugia, L. J. *Journal of Applied Crystallography* **1999**, *32*, 837–838, DOI: [10.1107/S0021889899006020](https://doi.org/10.1107/S0021889899006020).

Chapter 7

Development of a Ch-bonding DNA binder

7.1 Introduction

7.1.1 Cancer and treatment

In Australia, cancer consistently ranks as one of the leading causes of death among all age groups. In 2015, lung cancer alone accounted for 8,466 deaths, which ranked only behind ischaemic heart disease (19,777), dementia (12,625), and cerebrovascular diseases (10,869). If other cancers are included, the total number of deaths attributable to cancer is 1.5 times those caused by heart disease.¹

It has clear and serious consequences among families and communities, and also a wider economic price, costing Australia \$2 billion each year, of which the majority is spent on treatment.² Developing treatments for cancer has therefore attracted significant interest from the pharmaceutical industry, government, and the general public.³

Cancer is not a single disease, but an umbrella term which encompasses many different types of malignancies. The uniting theme among them is the uncontrolled and damaging division of cells. Because of the diversity of diseases broadly termed “cancer”, treatments are often difficult to develop and highly specific to one particular class.⁴

Some more general treatments for cancer exist. These include surgery and radiation therapy. While these treatments do have limitations (e.g. surgery is only viable for solid tumours), they are usually very effective, and are often combined with more specific therapies where appropriate.

7.1.2 Radiation therapy

Radiation therapy is one such general treatment for cancer. A course of radiation therapy consists of a series of exposures to radiation (x-rays, generated electrically; γ -rays, generated by radioactive decay; or, less commonly, fermionic particles such as protons, neutrons or electrons), with the intention of damaging the DNA of the cancer cells beyond the cells’ ability to repair, resulting in death of the cells and remission of the cancer.⁵ The mechanism of this damage is either direct, where a photon ejects an electron from the DNA polymer itself (via Compton scattering); or indirect, where other molecules (usually water) are ionised by the radiation, generating reactive oxygen species (ROS) such as $\cdot\text{OH}$ and O_2^- .⁶

If generated in the vicinity of the nucleus, these ROS may diffuse short distances and abstract electrons from DNA, causing damage to the DNA.⁷ Due to the minuscule volume occupied by DNA within the cell, indirect damage is the dominant mechanism in external beam radiotherapy.⁶

Damage to DNA can occur at the sugar-phosphate backbone or within the bases themselves, resulting in a range of fragmentation products or cross-linked adducts.⁷

Such products are not effective substrates for cellular processes such as DNA replication, so cell death usually occurs at the time of mitosis, if not before.⁸

Radiation therapy is targeted to inflict the greatest damage on cancer cells, while leaving healthy cells untouched. In other words, the aim is to maximise therapeutic value while minimising side effects. This is achieved by a number of means, including physically shaping the beam, using orthogonal beams focussed on the tumour, and fractionating the dosage. Compounds which can differentially alter the sensitivities of cancer cells and healthy cells (*radiomodifiers*) represent another way of achieving this. This project is concerned with the subset of radiomodifiers which reduce the damage done to healthy tissue (*radioprotectors*).

7.1.3 Radioprotection

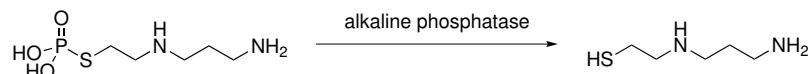


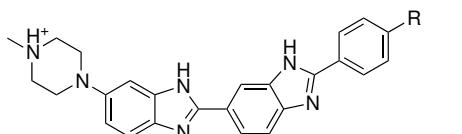
FIGURE 7.1: Amifostine, and its active metabolite.

Despite the clear and serious side effects of radiotherapy, there is currently only one radioprotective pharmaceutical licensed by the FDA for use as such. Amifostine (fig. 7.1) is administered as a prodrug, which is phosphorylated *in vivo* by alkaline phosphatase to form the free thiol which is able to scavenge radicals formed by irradiation. Selectivity for healthy cells ultimately stems from the hypoxic nature of cancer cells, which inhibits dephosphorylation to the active compound.⁹ However, due to the need for it to be present in relatively high concentration in the cytosol to effectively scavenge radicals, large doses are required, and this magnifies issues of cytotoxicity. A more targeted approach is therefore required for effective radioprotection and a larger therapeutic window.

7.1.4 Minor groove binding

DNA is known to exist in many different structures (A-, B-, C-DNA etc.), depending on factors such as solvation and the presence of metal ions. B-DNA, however, is by far the most prevalent in cells.¹⁰ The familiar double helix of B-DNA forms a major and a minor groove on either side of the sugar-phosphate backbone, where the nitrogenous bases are exposed. There exists a class of small molecules which bind to this minor groove, and impart some property to the bound DNA.

The Hoechst stains, first synthesised in the early 1970s by the company *Hoechst Aktiengesellschaft* as antihelmintic agents, are prototypical minor groove binders. As can be seen in scheme fig. 7.2, the gentle curvature of the molecule allows it to follow the DNA helix. The benzimidazole moieties provide H-bond donors, which interact with acceptors in adenine and thymine. At physiological pH, the molecule is positively charged, allowing further stabilisation of the DNA-Hoechst complex via electrostatic interactions with the electron rich sugar-phosphate backbone.¹¹ The conjugated aromatic system imparts a strong UV absorbance at around 350 nm. Fluorescence in the visible spectrum is also observed, giving Hoechst compounds utility as DNA stains.



Hoechst 33258 R: OH
Hoechst 33342 R: OCH₂CH₃
Hoechst 34580 R: N(CH₃)₂

FIGURE 7.2: Hoechst stains, featuring the characteristic bis-benzimidazole motif.

7.1.5 Bis-benzimidazole derived radioprotectors

In collaboration with the Peter MacCallum Cancer Centre, the White group has investigated a range of minor groove binders inspired by the Hoechst stains. Methylproamine is a lead compound evaluated for radioprotection. It was found to effectively protect cells from γ and x-ray radiation, based on a clonogenic survival assay. Pulse radiolysis studies suggested that the mechanism (for directly targeted cells) is based on direct repair of oxidative lesions, as opposed to scavenging ROS like amifostine. When a DNA base (usually guanine) is ionised, a “hole” in the DNA is created. This can be effectively transported along the length of the DNA molecule.¹² The bound ligand (methylproamine) can then act as a reducing agent and neutralise the hole.

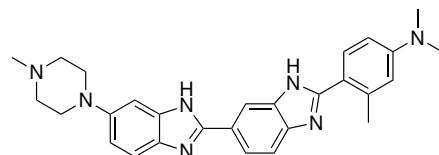


FIGURE 7.3: Methylproamine, a lead compound identified for radioprotection.

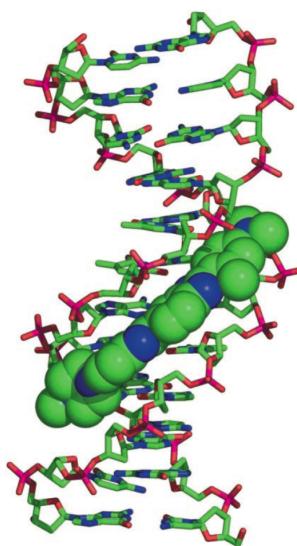


FIGURE 7.4: Single crystal structure of methylproamine complexed with an AT rich DNA oligomer.

7.1.6 Proton-coupled electron transfer (PCET)

The reduction mechanism of methylproamine and other bis-benzimidazole compounds is thought to be a proton-coupled electron transfer, in which an electron and proton are transferred from the ligand to the DNA in a stepwise process. This was based on QSAR studies of a wide range of derivatives.¹³ It was found that the radioprotective potential was proportional to the acidity of the first benzimidazole proton. This observation led to a new generation of radioprotectors with a 2-pyridyl group capping the first benzimidazole. The lone pair of the pyridine nitrogen is able to donate into the $\sigma^* \text{N}-\text{H}$ anti-bonding orbital and weaken the bond, allowing the proton to be more easily abstracted.

Substituting the N-methyl piperazinyl group for a N-morpholino group further improved the radioprotective ability of the bis-benzimidazole. *Ab initio* modelling showed the HOMO was centred around the lone pair of the morpholine nitrogen, and x-ray crystallography indicated that, when complexed with DNA, this lone pair pointed into the minor groove in an acceptable geometry for direct electron transfer. The lead compound M2PB **10** was therefore synthesised, and showed very promising radioprotective ability, albeit with a relatively small therapeutic window.

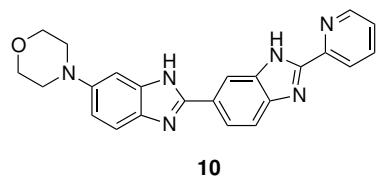


FIGURE 7.5: M2PB **10**, an optimised lead compound for radioprotection.

7.1.7 Chalcogens as nitrogen bioisosteres

Heavier group VI elements such as sulfur and selenium have been observed to behave similarly to nitrogen in biological systems. A range of drugs have been developed that display this bioisosteric behaviour.¹⁴ The similarity between these

elements is postulated to be due to the σ -hole that heavy main group elements exhibit, and their resulting propensity for Ch-bonding.

The fundamental aspects of Ch-bonding have been thoroughly discussed elsewhere in this thesis, suffice it to say that derivatives of the drug ebselen **1** were identified early on as likely bioisosteres for a benzimidazole moiety.

7.1.8 Ebselen as an antioxidant

Ebselen itself, of course, displays antioxidant properties. These are believed to be the basis for its neuroprotective, anti-helminthic, and anti-inflammatory behaviour. The mechanism, however, appears to be completely different. Ebselen behaves as a glutathione peroxidase mimic, that is to say, a catalyst that increases the rate at which glutathione reduces reactive oxygen species.

For this reason, we did not constrain ourselves to 2-pyridyl analogues (as the mechanism may not involve PCET), nor did we incorporate the morpholino substituent (as the HOMO is likely to be centred on the selenium).

7.2 Synthesis of analogues

Initial targets **11** and **12** were identified as likely minor groove binders, based on their similarity to the bis-benzimidazoles previously studied in the group. Due to the difficulty of functionalising the benzisoselenazolinone system and the rationalisation in section 7.1.8, we simplified our targets to exclude the morpholino group. Among our targets, bis-benzisoselenazolinone **12** was particularly interesting, as it has the potential to be a completely H-bond free minor groove binder. However, we first focussed on the synthesis of benzisoselenazolinone-benzimidazole **11**, as this appeared to be the most synthetically accessible using the chemistry developed in the group.

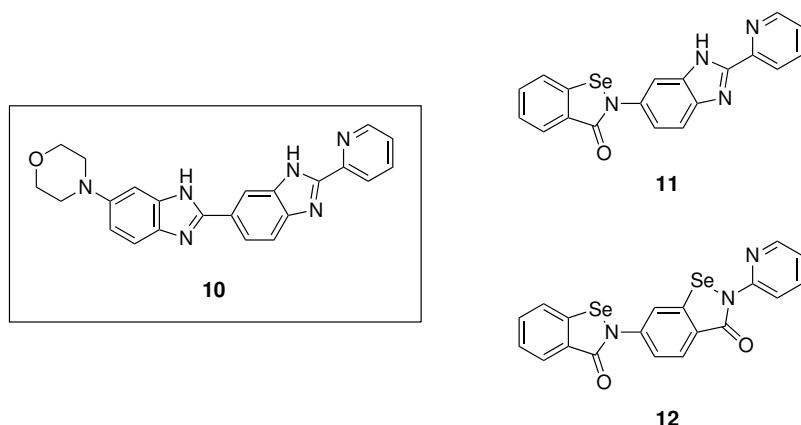


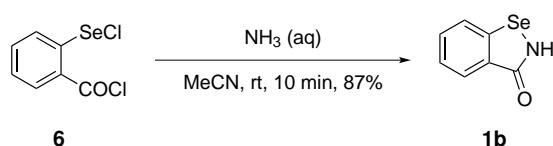
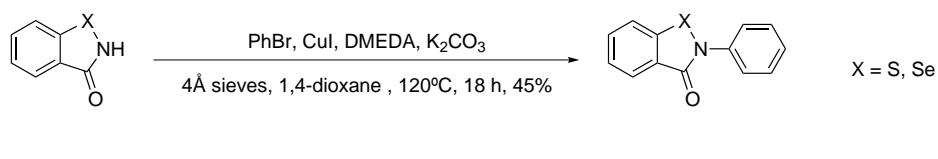
FIGURE 7.6: Lead bis-benzimidazole **10** and initial target compounds **11** and **12**.

7.2.1 Preparation of benziselenazolinone-benzimidazole **11**.

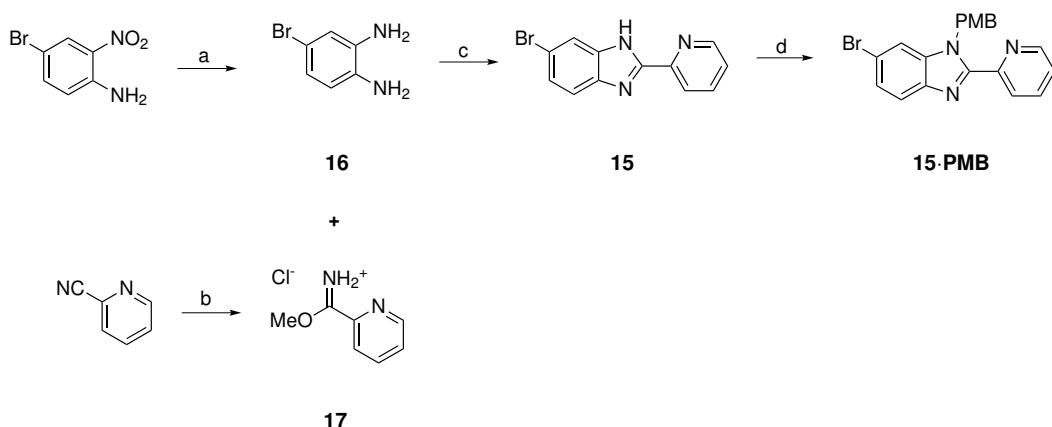
7.2.1.1 Ullmann coupling

A number of routes to this compound were envisaged, via various intermediates. Firstly, we were inspired by the Ullmann-type chemistry exploited by Balkrishna et al. in their selenocyclisation, and by the observation that the amidic proton in **1b** goes on to react further to furnish the tetracyclic compound **3** (see chapter 3).^{15,16} We supposed that the intermediate **1b**, which can be prepared by reaction of the dichloride **6** with aqueous ammonia (scheme 7.1) would be a suitable substrate for an Ullmann coupling with an aryl halide. A number of test reactions were performed using the benzisothiazolinone **1b·S**, which is commercially available and inexpensive. Typical Ullmann conditions were used (high temperature, DMF solvent, high CuI catalyst loading) to initially couple **1b·S** with bromobenzene to afford **1·S** in moderate yield (scheme 7.2). We were also able to prepare ebselen **1** via this route in similar yield. Optimisation of the reaction was considered to be outside the scope of this work, so we proceeded with the synthesis of the bromobenzimidazole coupling partner **15**.

4-bromo-2-nitroaniline was reduced to the diamine **16** using tin (II) chloride, then reacted with 2-pyridyl carboximidate hydrochloride **17** (prepared from 2-pyridyl

SCHEME 7.1: Synthesis of **1b**.SCHEME 7.2: Synthesis of **1**.

carbonitrile) to afford the benzimidazole **15** in excellent yield. Unfortunately this gave no detectable product in the coupling reaction. We suspected that the benzimidazole proton may be competing with the amidic proton in the benzisothiazolinone ring, as it is substantially more acidic (pK_a 16.4 vs > 20), so we protected it using *p*-methoxy benzyl chloride to give **15·PMB** (scheme 7.3). This afforded two distinct isomers from the two freely interconverting tautomers of **15**. Although these were resolved by chromatography for subsequent reactions, this was likely not necessary, as deprotection would afford both tautomers. Both isomers are therefore treated as a single compound here.

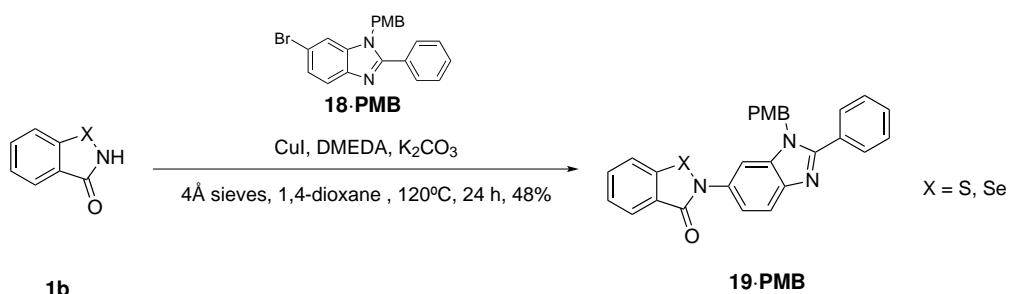
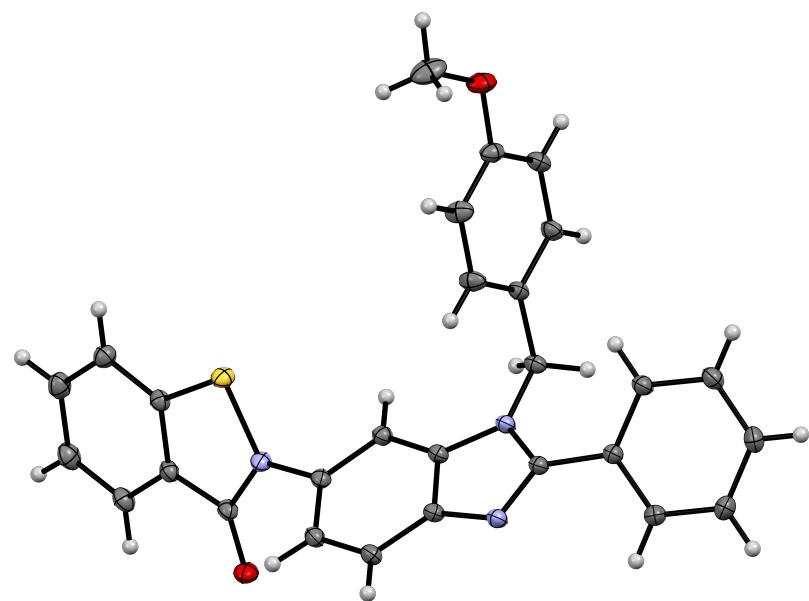
SCHEME 7.3: Synthesis of **15** and protection to form **15·PMB**. (a) $\text{SnCl}_2 \cdot 2\text{H}_2\text{O}$, EtOH, reflux, 8 h, 92%; (b) NaOMe , MeOH, 80°C, 1 h; (c) MeOH, AcOH, reflux, 18 h, 91%; (d) NaH , PMBCl , DMF, 0°C → rt, 18 h, 82%.

The PMB-protected benzimidazoles also gave no detectable coupling product, so we turned our attention to the catalyst/ligand system. Ullmann couplings require a bidentate nitrogen-based ligand to coordinate the copper (I) species. We used dimethylethylenediamine (DMEDA), although salicylates, 1,10-phenanthroline, 2,2'-bipyridyl, tetramethylethylenediamine, and proline have also been used successfully in similar reactions.^{17–19} The 2-(2-pyridyl)benzimidazole, even protected, resembles these ligands, and could reasonably be anticipated to coordinate the copper in competition with the ligand. Although this does not necessarily preclude the coupling reaction, it does remove our ability to modify the coordination environment of the copper, to which the Ullmann reaction is very sensitive. We attempted a ligand-free reaction to no avail, and thus concluded that the coordination environment of the copper is inappropriate for the reaction, and we would have to use another substrate.

We therefore prepared the phenyl derivative **18** via a metabisulfite coupling with benzaldehyde, and protected it similarly to give both isomers of **18·PMB**. This gave the desired coupling product **19·PMB·S** as a mixture of isomers, from one of which we obtained a single crystal structure (scheme 7.4 and fig. 7.7). We were also able to prepare the benzisoselenazolinone derivative **19·PMB** as a pair of isomers. Unfortunately, all efforts to deprotect the benzimidazole nitrogen failed. A number of other protecting groups were trialled, including trityl, triphenylsilyl, *t*-butyl and ethyl carbamates, however all either failed to give any protected product, or were too labile to survive the Ullmann reaction conditions. We therefore abandoned this synthetic route.

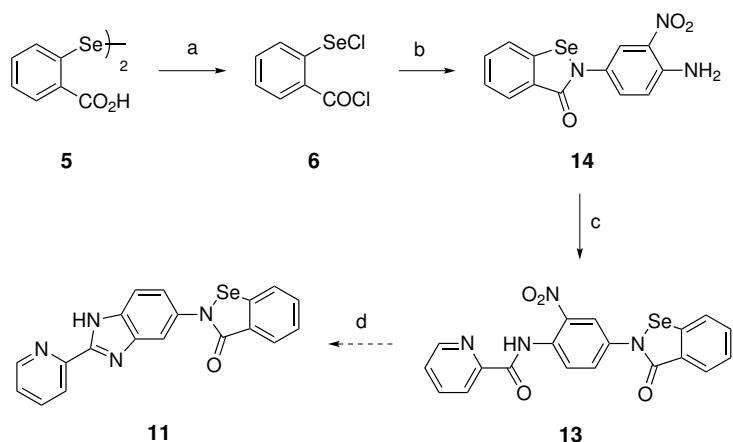
7.2.1.2 Benzisoselenazolinone-first route

A revised route to this molecule is shown in scheme 7.5, which involved first synthesising the benzisoselenazolinone moiety, then constructing the benzimidazole in a condensation and reductive dehydrocyclisation. The results of this, specifically

SCHEME 7.4: Synthesis of **19·PMB**.FIGURE 7.7: X-ray crystal structure of one isomer of **19·PMB·S**.

the interesting crystal packing of the late stage intermediate **13** spurred the investigation detailed in chapter 6. As this pathway did not involve copper catalysis, we reintroduced the 2-pyridyl group on the benzimidazole. However, this route was not able to furnish the desired product, as the benzoiselenazolinone moiety did not tolerate the reduction conditions trialled. Among the conditions trialled were:

- palladium catalysed hydrogenation, from which we only recovered starting material presumably due to the catalyst being poisoned by the divalent selenium,



SCHEME 7.5: Proposed synthesis of **11**. (a) SOCl_2 , cat. DMF, reflux, 30 min; (b) 2-nitro-1,4-benzenediamine, Et_3N , THF, rt, 18 h, 61%; (c) Picolinic acid, TCBC, Et_3N , rt, 24 h, 20%; (d) [H], H^+ .

- tin (II) chloride/HCl reduction, which afforded a diselenide **20** (fig. 7.8),
- dithionite reduction, which afforded a complex mixture of products.

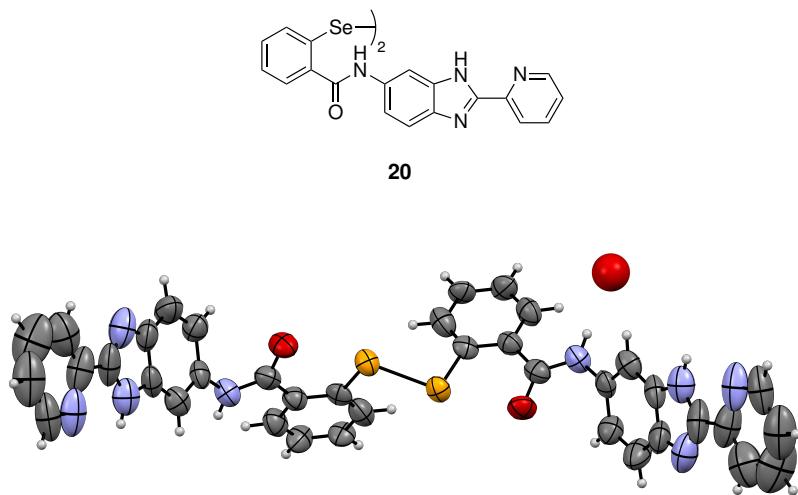
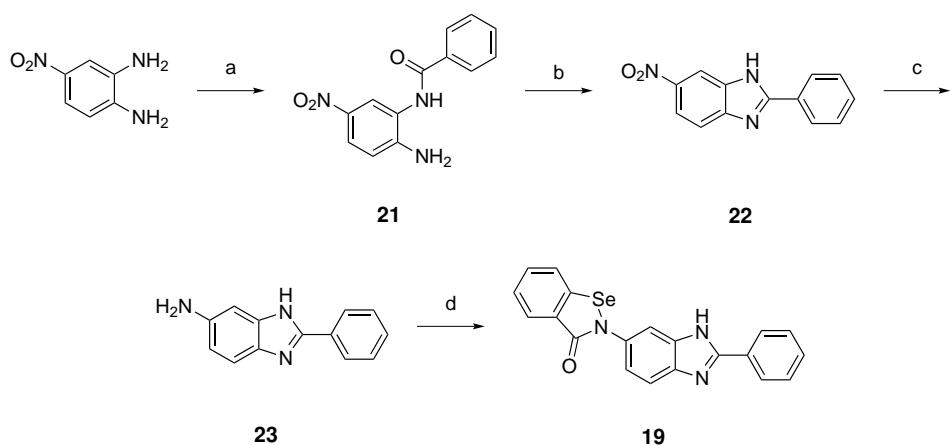


FIGURE 7.8: X-ray crystal structure of **20** with a disordered water molecule.

7.2.1.3 Benzoiselenazoline-last route

We therefore devised another scheme in which the benzoiselenazolinone ring is formed last, as the delicate nature of this group appeared to be hindering our



SCHEME 7.6: Synthesis of benzisoselenazolinone-benzimidazole Hoechst analogue **19**. (a) BzCl, NEt₃, THF, rt, 18 h, 75%; (b) BF₃ · OEt₂, 1,4-dioxane, reflux, 3.5 h, 93%; (c) SnCl₂, EtOH, reflux, 2 h, 87%; (d) **6**, MeCN, NEt₃, rt, 18 h, 8%.

efforts (scheme 7.6). This initially involved formation of an amide **21** by treating 3,4-diamino-1-nitrobenzene with an acid chloride. It was found that the acid chloride of picolinic acid (which would ultimately afford the 2-(2-pyridyl) benzimidazole) was not stable, decomposing almost as soon as it was formed. We investigated using the carboximide as in section 7.2.1.1, however the nitro group on the diamine proved to be too deactivating; the aniline nitrogens were not sufficiently nucleophilic to react with the carboximide. Rather than investigate other pathways to this compound (such as the use of coupling agents), we again decided to simplify the system to a phenyl ring (to ultimately give **19**). We therefore formed **21** using benzoyl chloride in excellent yield, which underwent a dehydrocyclisation in the presence of the strong Lewis acid BF₃ · OEt₂ to form **22**. Tin (II) chloride reduction of the nitroarene **22** afforded the aniline **23** as the hexachlorostannate salt. This was coupled to the bis-electrophile **6** to afford the desired benzisoselenazolinone-benzimidazole **19**, which formed single crystals suitable for x-ray diffraction by slow evaporation from DMSO (fig. 7.9).

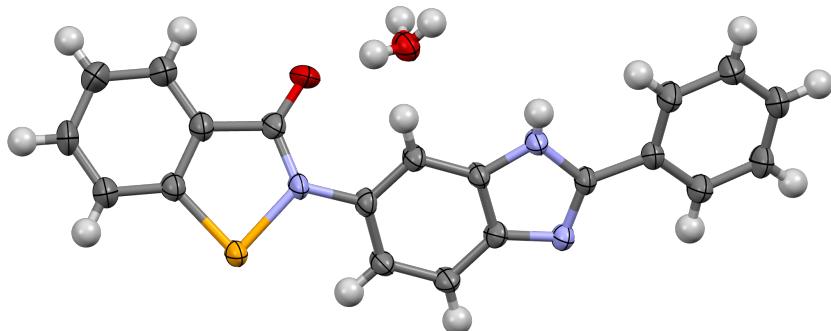


FIGURE 7.9: X-ray crystal structure of **19**. The water is disordered over 3 sites, with each proton having an occupancy of 2/3.

7.3 DNA binding studies

Previous studies of Hoechst derivatives have shown that radioprotective ability is strongly correlated with a high affinity for the minor groove of DNA. Although it is conceivable that ROS may be reduced in the cytoplasm by radioprotector molecules, this does not appear to be the major method of radioprotection for these compounds.

7.3.1 Co-crystallisation results

The main technique used is co-crystallisation of the ligand with DNA oligomers. Unlike the co-crystallisation experiments used earlier in this thesis, this experiment bears more similarity to the techniques employed in macromolecular crystallisation, simply due to the delicate nature of the DNA oligomers. Vapour diffusion from hanging drops was selected as the method for growing the crystals. Hanging drop

crystallisation relies on the equilibration of water between a reservoir of certain osmotic potential, and a droplet just above it containing the components of the crystal. The main benefit is the slow rate of diffusion of water between the droplet and the reservoir, which leads to the formation of high quality crystals. The technique is also quite efficient, in that only small amounts of compound are used.

The components of the drops were as follows:

- DNA oligomer, the A2T2 self-complementary 16mer was used for all experiments (CGCGCGAATTCTCGCGCG);
- ligand molecule **19**;
- spermine hydrochloride, a polycationic tetramine which helps to stabilise the negative charge on the phosphate backbone of the DNA;
- MgCl_2 , a source of Mg^{2+} cations to further stabilise the negative DNA;
- sodium cacodylate, a buffer to ensure constant pH, and to inhibit microbial growth;
- water, the solvent;
- 2-methyl-2,4-pentanediol, the anti-solvent.

The solvent reservoir contained only water and 2-methyl-2,4-pentanediol.

Unfortunately, we were not able to obtain co-crystals of **19** with the DNA oligomer, as precipitation of an amorphous solid occurred upon addition of the ligand solution to the drop. This suggested that solubility of the compound may be an issue, which is fairly common among bis-benzimidazoles.

In order to circumvent this, we instead grew crystals of the DNA oligomer in the absence of the ligand. This afforded large crystals suitable for x-ray diffraction. To introduce the ligand into the minor groove, we soaked the DNA crystals in a solution

of the ligand in crystallisation buffer. The DNA crystal structure contains substantial amounts of disordered water, forming channels through which the ligand may diffuse (fig. 7.10). However, upon exposure to the ligand, the DNA crystals appeared to disintegrate to give a similar amorphous precipitate.

We therefore abandoned this method of characterising the binding of the ligand.

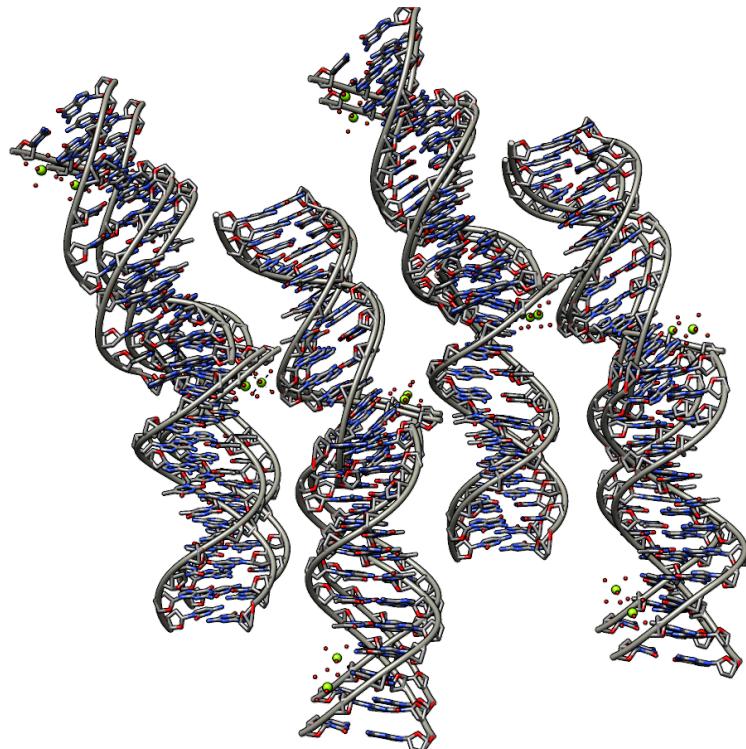


FIGURE 7.10: Crystal packing of the A2T2 self-complementary DNA 16mer, showing the open channels between the DNA molecules.

7.3.2 UV-Vis titration

In the absence of a co-crystal structure, we sought to measure the binding of the ligand by Uv-Vis titration. Briefly, this works by measuring the degree of conjugation between the two benzimidazole (or benzimidazole-like) systems. Free bis-benzimidazoles have a significant torsion about the central bond, partially due to the push-pull character of these compounds. This leads to relatively poor orbital

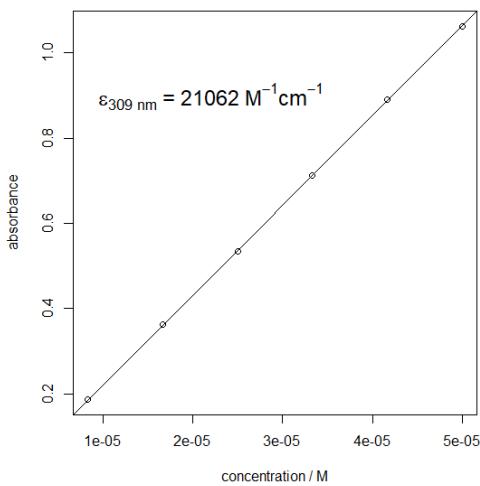
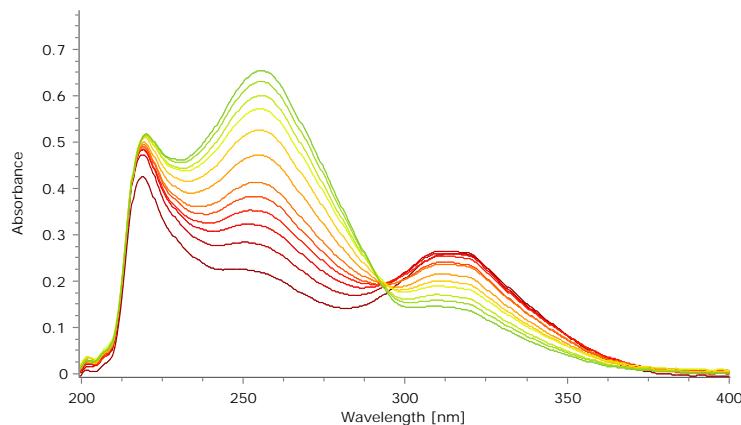
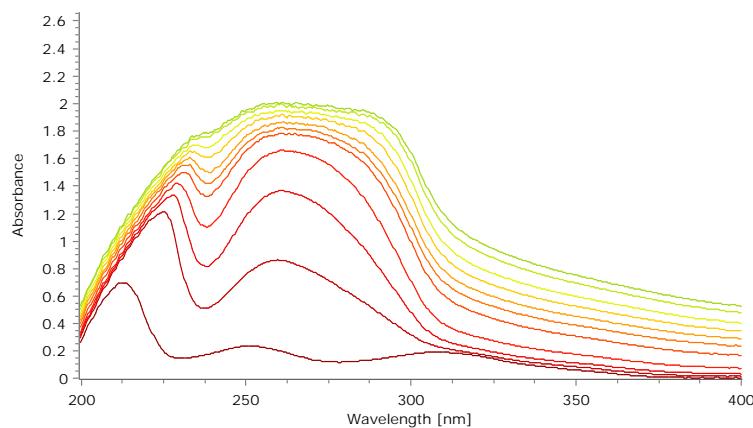
overlap and conjugation between the benzimidazoles, and an associated blue shift in the absorbance spectrum as compared to a completely planar molecule of the same size. As the bis-benzimidazole binds to the minor groove of a DNA molecule, a more planar geometry is imposed, so the orbital overlap is improved and the absorbance peak moves towards the red. By the application of a simple binding model, a dissociation constant can be derived.

Also visible in these titration experiments are spectroscopic signatures of different types of binding, including intercalation and major groove binding. Both of these are referred to as non-specific binding, as they do not rely on the presence of four consecutive A-T pairs to expose H-bond acceptors in the minor groove. These types of binding are not associated with a bathochromic shift in the absorbance spectrum, but instead with a quenching of the signal. It is important to note that non-specific binding occurs simultaneously with specific minor groove binding, but the latter, when present, dominates the changes in the absorbance spectrum.

In order to assess the binding constant, the absorption spectrum and extinction coefficient for the free ligand must be measured. The extinction coefficient at $\lambda_{\text{max}} = 309 \text{ nm}$ for **19** in 0.1% TFA/45% MeOH was determined to be $21062 \text{ M}^{-1}\text{cm}^{-1}$ (fig. 7.11).

The titration was then carried out using a solution of the ligand in 0.1% TFA/45% MeOH ($15 \mu\text{M}$) and diluted DNA stock (0.025 mMbp), and the resulting spectra are shown in fig. 7.12. A shift in absorbance is not readily apparent at such low DNA concentrations, so the experiment was repeated using 2 mMbp DNA stock, and the resulting spectra are shown in fig. 7.13. At this concentration, we observed precipitation of the ligand and DNA, which can be seen in the broadly absorbing tail in the spectra. This is highly undesirable, as the resulting data cannot be used to calculate the binding affinity.

A further sample of the precipitate was prepared using the same conditions, and

FIGURE 7.11: Calibration curve for **19**.FIGURE 7.12: UV-vis titration of **19** with ct-DNA at low DNA stock concentration.FIGURE 7.13: UV-vis titration of **19** with ct-DNA at high DNA stock concentration.

the solid isolated by centrifuging. The supernatant was pipetted off, and the UV-vis spectrum revealed no absorbance at 309 nm, so we concluded that all of the ligand was contained in the precipitate. The solid was washed with T/E/N buffer (section 7.5.1) to remove excess DNA, and 0.1% TFA/45% MeOH to remove excess ligand, dried in vacuo, then dissolved in d_6 -DMSO. ^1H NMR revealed that the solid contained nucleic acid material (in the downfield region) as well as aromatic signals consistent with the ligand (fig. 7.14). We suspected that this was the same amorphous material that formed in the co-crystallisation experiment, and attribute its formation to a reaction between the ligand and the DNA.

Unfortunately this makes **19** useless for the radioprotectors project, as a core requirement is the formation of a *non*-covalent complex.

7.4 Conclusions

7.5 Experimental procedures

7.5.1 DNA titration procedure

The following solutions were prepared:

- Tris stock – 1 M, pH 7.2
 - 1.214 g tris(hydroxymethyl)aminomethane freebase
 - 9 mL 1 M HCl
 - dilute to 10 mL with water
- EDTA·2Na⁺ stock – 0.1 M
 - 372 mg EDTA·2Na⁺·2H₂O

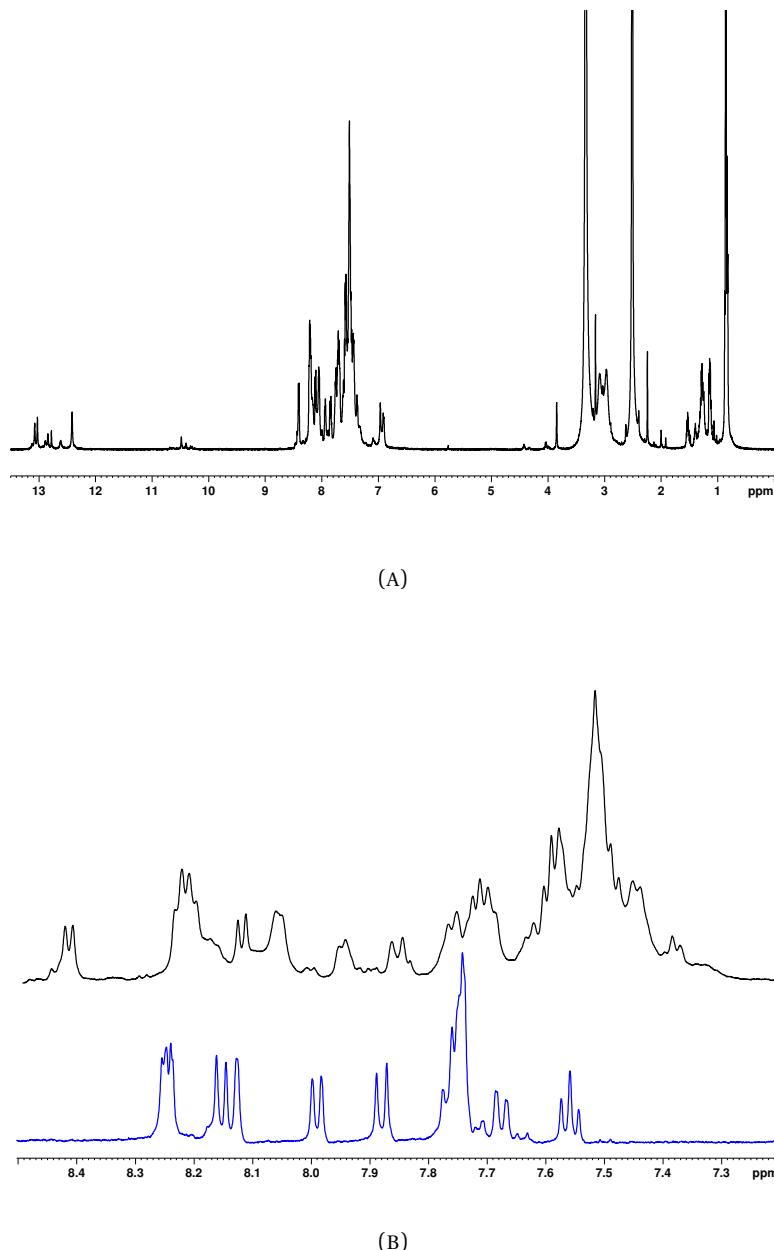


FIGURE 7.14: ¹H NMR spectra of the adduct of **19** with ct-DNA, and the pure ligand (blue) for comparison.

- dilute to 10 mL with water
- NaCl stock – 1 M
 - 584 mg NaCl
 - dilute to 10 mL with water
- T/E/N \times 2 buffer
 - 400 μ L Tris stock – final concentration 40 mM, pH 7.2
 - 200 μ L EDTA stock – final concentration 2 mM
 - 2 mL NaCl stock – final concentration 200 mM
 - 2 mL DMSO – final concentration 20% v/v
 - dilute to 10 mL with water
- T/E buffer
 - 500 μ L Tris stock – final concentration 10 mM, pH 7.2
 - 500 μ L EDTA stock – final concentration 2 mM
 - dilute to 50 mL with water
- TFA/M/W
 - 10 μ L TFA
 - 4.5 mL methanol
 - dilute to 10 mL with water
- ct-DNA stock
 - 30 mg ct-DNA
 - 10 mL T/E buffer*

*The DNA can be cut into pieces using clean scissors, then magnetically stirred overnight to dissolve in the buffer. MW and viscosity can be reduced by drawing up rapidly through a 22G needle, then sonicating for up to 30 minutes. It is critical that the temperature remains below 30°C. The final concentration can be checked by diluting the stock solution fiftyfold and measuring the absorbance at 260 nm. An absorbance of 1.0 corresponds to a concentration of 75 μ Mbp.

- Ligand stock

7.5.2 Synthetic methods

Selenium was purified by refluxing in 32% hydrochloric acid for 2 h, then washing with deionised water, methanol, then ether.

7.5.2.1 Preparation of **1b**.

Aqueous ammonia (30% w/w, 637 μ L, 10 mmol) was dissolved in anhydrous acetonitrile (5 mL), and to this was added a solution of dichloride **6** in acetonitrile (10 mL, 0.5 mmol/mL). The solution was stirred at room temperature for 10 min, then the solvent was evaporated to give a cream powder which was used without further purification (858.4 mg, 87%).

^1H NMR (499 MHz, d_6 -DMSO) δ ppm 9.16 (1H, s), 8.04 (1H, d, J = 8.03 Hz), 7.79 (1H, d, J = 7.76 Hz), 7.59 (1H, t, J = 6.95 Hz), 7.41 (1H, t, J = 7.02 Hz).

^{13}C NMR (151 MHz, d_6 -DMSO) δ ppm 169.12, 141.85, 131.91, 128.04, 127.65, 126.72, 125.99.

^{77}Se NMR (95 MHz, d_6 -DMSO) δ ppm 795.01.

MS (ESI +ve) m/z 199.9609 (MH^+) $\text{C}_7\text{H}_6\text{NOSe}^+$ requires 199.9609 ($\Delta=0$ ppm).

7.5.2.2 Preparation of **1·S**.

To a sealed tube containing 4 Å molecular sieves (740.7 mg) was added copper (I) iodide (34.3 mg, 0.180 mmol), DMEDA (27.6 mg, 31.3 mmol), potassium carbonate (305.3 mg, 2.209 mmol), benzisothiazolinone **1·S**, bromobenzene (190.6 mg, 1.213 mmol), and dioxane (3 mL). The tube was capped and heated to 120°C for

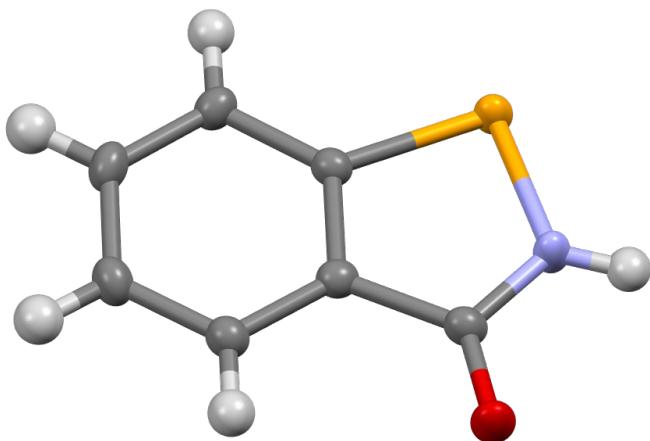


FIGURE 7.15: X-ray crystal structure of **1b**.

18 h, then cooled and the mixture tipped into water (30 mL). The resulting blue suspension was centrifuged, and the blue supernatant discarded. The residue was dissolved in 1:1 DCM/methanol, and decanted off from the sieves. The solvent was evaporated to afford a greenish solid, which was chromatographed to give the product as colourless needles (105.0 mg, 45%).

^1H NMR (400 MHz, d_6 -DMSO) δ ppm 8.05 (1H, d, J = 8.2 Hz), 7.94 (1H, d, J = 7.75 Hz), 7.75 (1H, t, J = 7.69 Hz), 7.69 (2H, d, J = 8.13 Hz), 7.56–7.46 (3H, m), 7.36 (1H, t, J = 7.39 Hz).

MS (ESI +ve) m/z 228.0476 (MH^+) $\text{C}_{13}\text{H}_{10}\text{NOS}^+$ requires 228.0478 ($\Delta=0.88$ ppm).

7.5.2.3 Preparation of 16.

The nitrobenzene (4.9982 g, 23.031 mmol) was dissolved in ethanol (35 mL), and to this was added tin (II) chloride dihydrate (26.022 g, 115.25 mmol, 5 eq). The mixture was refluxed for 8 h, then cooled, concentrated, and diluted with water to a volume of 300 mL. The thick white suspension was basified to pH 12, cooled again, and extracted with diethyl ether (2×200 mL). The combined organic layers were washed with brine (150 mL), dried (MgSO_4), and evaporated to give a brownish oil,

which was triturated with petroleum ether to give a light yellow solid which was used without further purification (3.9759 g, 92%).

^1H NMR (400 MHz, d_6 -DMSO) δ ppm 6.6 (1H, d, J = 2.17 Hz), 6.47–6.42 (1H, dd, J = 2.17, 8.2 Hz), 6.39 (1H, d, J = 8.2 Hz), 4.61 (4H, br. s.).

7.5.2.4 Preparation of 15.

A 0.1 M solution of sodium methoxide was prepared by dissolving sodium metal (137 mg) in anhydrous methanol (50 mL). Of this, 6.8 mL was added to 2-pyridine carbonitrile (714.0 mg, 6.858 mmol) and the mixture heated to 80°C under an argon atmosphere for 1 h to afford the carboximide **17**. To this was then added a solution of the diamine **16** (1.0226 g, 5.467 mmol) in a further 20 mL anhydrous methanol, followed by glacial acetic acid (650 μ L, 11.5 mmol), and the mixture gently refluxed for 18 h. Upon cooling, the mixture was concentrated to give a yellow oil which was triturated with 1 M aqueous ammonia to give a friable pale yellow solid, which was washed with water to give the product (1.3595 g, 91%).

^1H NMR (400 MHz, d_6 -DMSO + d -TFA vapour) δ ppm 8.72 (1H, d, J = 4.21 Hz), 8.31 (1H, d, J = 7.85 Hz), 8.01 (1H, t, J = 7.7 Hz), 7.79 (1H, s), 7.63–7.49 (2H, m), 7.37 (1H, d, J = 8.53 Hz).

MS (ESI +ve) m/z 273.9975 (MH^+) $\text{C}_{12}\text{H}_9\text{BrN}_3^+$ requires 273.9974 (Δ =0.36 ppm).

7.5.2.5 Preparation of 15·PMB.

The benzimidazole **15** (1034.1 mg, 3.797 mmol) and sodium hydride (60% in oil, 298.2 mg, 7.450 mmol, 2.5 eq) were dissolved in anhydrous DMF (6 mL) at 0°C, and to this was added *p*-methoxybenzyl chloride (600 μ L, 4.42 mmol, 1.2 eq). The mixture was warmed to room temperature and stirred for 18 h, then diluted with water (50 mL) and extracted with DCM (2 \times 80 mL). The combined organic phases

were washed with brine, dried (MgSO_4) and evaporated to give a yellow oil, which was chromatographed to afford two major fractions:

1. 633 mg

2. 591 mg

which when combined gave a 82% overall yield.

^1H NMR (499 MHz, CDCl_3) δ ppm 8.67 (1H, d, $J = 4.12$ Hz), 8.4 (1H, d, $J = 7.95$ Hz), 7.98 (1H, d, $J = 1.52$ Hz), 7.86 (1H, dt, $J_1 = 1.65$ Hz, $J_2 = 7.78$ Hz), 7.38–7.34 (2H, m), 7.23 (1H, d, $J = 8.63$ Hz), 7.11 (2H, d, $J = 8.65$ Hz), 6.78 (2H, d, $J = 8.65$ Hz), 6.08 (2H, s), 3.73 (3H, s).

MS (ESI +ve) m/z 394.0555 (MH^+) $\text{C}_{20}\text{H}_{17}\text{BrN}_3\text{O}^+$ requires 394.0550 ($\Delta=1.27$ ppm).

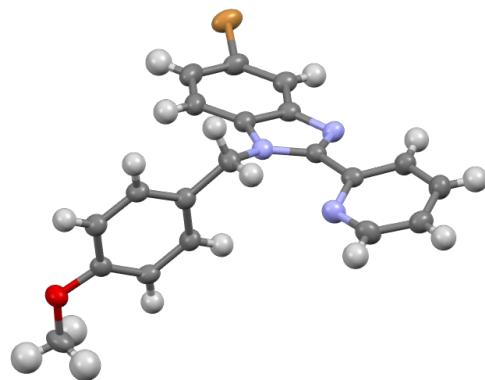


FIGURE 7.16: X-ray crystal structure of **15·PMB**.

7.5.2.6 Preparation of **18**.

Sodium metabisulfite (9.670 g, 50.85 mmol) was dissolved in 50 mL water and added to 200 mL ethanol, then benzaldehyde (5.444 g, 51.30 mmol) was added. This was stirred at room temperature for 1 h, then a solution of the diamine **16** (4.0437 g, 21.6 mmol) in ethanol (40 mL) was added, and the mixture refluxed under nitrogen

for 24 h, then cooled and left to stand for 60 h. The solvent was removed by rotary evaporator, and the residue triturated with 1 M aqueous ammonia (50 mL). The resulting precipitate was filtered off, washed with water (2×25 mL), cold diethyl ether (5 mL), and dried affording the benzimidazole **18** (5.1646 g, 79%).

^1H NMR (499 MHz, d_6 -DMSO + d -TFA) δ ppm 8.18 (2H, d, $J = 6.89$ Hz), 7.83 (1H, s), 7.64–7.52 (4H, m), 7.41 (1H, dd, $J_1 = 1.51$ Hz, $J_2 = 8.52$ Hz).

7.5.2.7 Preparation of **19·PMB·S**.

Benzisothiazolinone **1b·S** (173.1 mg, 1.142 mmol), aryl bromide **18·PMB** (401.9 mg, 1.022 mmol), 4 Å molecular sieves (607.1 mg), DMEDA (32.7 mg, 0.371 mmol), potassium carbonate (283.8 mg, 2.053 mmol), and dioxane (3 mL) were combined in a sealed tube under a nitrogen atmosphere, and heated at 120°C for 24 h. The mixture was then cooled to room temperature and tipped into water (40 mL). The suspension was centrifuged, and the supernatant discarded, then the residue redissolved in methanol (2×40 mL). This was filtered, and the filtrate evaporated to give a light brown oil, which was applied to a SNAP 25 g silica cartridge and eluted with an ethyl acetate/petroleum ether gradient to give two fractions:

1. 160.9 mg, aryl bromide **18·PMB**,
2. 231.2 mg, 48%, benzisothiazolinone-benzimidazole **19·PMB·S**.

19·PMB·S crystallised in two distinct polymorphs:

1. P $\bar{1}$, m.p. 101.0–104.2°C
2. P $2_1/n$, m.p. 134.1–138.4°C

¹H NMR (600 MHz, *d*₆-DMSO) δ ppm 8.03 (1H, d, *J* = 8.08 Hz), 7.92 (1H, d, *J* = 7.67 Hz), 7.86–7.79 (2H, m), 7.73 (3H, d, *J* = 4.68 Hz), 7.55 (3H, d, *J* = 3.42 Hz), 7.51–7.44 (2H, m), 6.93 (2H, d, *J* = 8.38 Hz), 6.82 (2H, d, *J* = 8.46 Hz), 5.52 (2H, s), 3.66 (3H, s).

MS (ESI +ve) m/z 464.1431 (MH⁺) C₂₈H₂₂N₃O₂S⁺ requires 464.1427 (Δ=0.86 ppm).

7.5.2.8 Preparation of N-(2-amino-5-nitrophenyl)benzamide 21

4-Nitro-1,2-benzenediamine (3.0797 g, 20.110 mmol) and TEA (3 mL, distilled from CaH₂) were dissolved in anhydrous THF (150 mL). Benzoyl chloride (1.5 mL, 1.8 g, 13 mmol) was then added at –10°C and the mixture slowly warmed to room temperature while stirring for 18 h. The mixture was then tipped into water (400 mL), and extracted with ethyl acetate (3 × 100 mL). The combined organic layers were washed with water (2 × 200 mL), brine (200 mL), then dried (MgSO₄) and evaporated to give a reddish solid. This was recrystallised from ethyl acetate to give the amide **21** as yellow crystals (2.5027 g, 75%).

¹H NMR (400 MHz, DMSO-*d*6) δ 9.66 (s, 1H), 8.04 (s, 1H), 7.92 (d, *J* = 7.4 Hz, 2H), 7.83 (d, *J* = 9.2 Hz, 1H), 7.50 (d, *J* = 6.8 Hz, 1H), 7.44 (t, *J* = 7.3 Hz, 2H), 6.71 (d, *J* = 9.0 Hz, 1H), 6.52 (s, 2H).

7.5.2.9 Preparation of 6-nitro-2-phenyl-1*H*-benzo[*d*]imidazole 22

Amide **21** (1.2283 g, 4.7749 mmol) was refluxed in a solution of BF₃ · OEt₂ (0.75 mL) and dioxane (75 mL) for 1.5 h. The mixture was then cooled to room temperature, stirred for 18 h, then heated to reflux again for 2 h. The yellow solution was diluted with water (300 mL) and extracted with ethyl acetate (3 × 75 mL). The combined organic layers were washed with water and brine (100 mL each), then dried (Na₂SO₄), and evaporated to give a brown oil. Trituration with petroleum spirit/dichloromethane afforded **22** as a light brown friable solid (1.0606 g, 93%).

¹H NMR (400 MHz, DMSO-*d*6 + *d*-TFA) δ 8.47 (s, 1H), 8.21 (q, *J* = 3.2 Hz, 2H), 8.13 (q, *J* = 3.7 Hz, 1H), 7.76 (d, *J* = 8.9 Hz, 1H), 7.54–7.62 (m, 3H).

MS (ESI +ve) m/z 240.0768 (MH⁺) C₁₃H₁₀N₃O₂⁺ requires 240.0768 ($\Delta=0$ ppm).

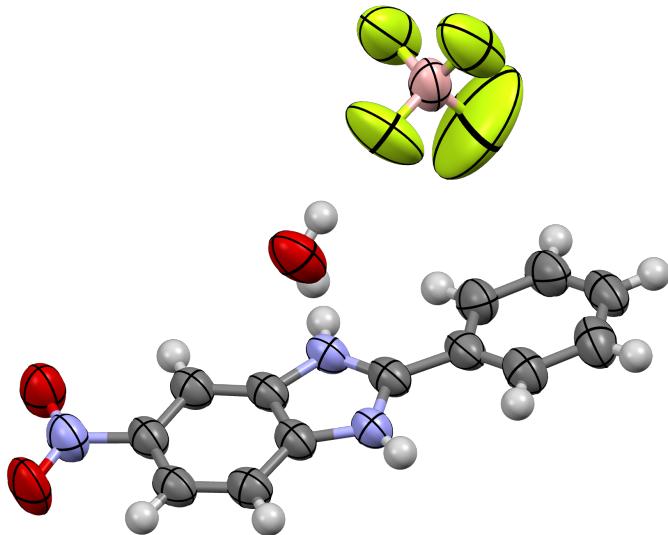


FIGURE 7.17: X-ray crystal structure of **22** as the tetrafluoroborate salt.

7.5.2.10 Preparation of 2-phenyl-1*H*-benzo[*d*]imidazol-6-amine **23**

A solution of the nitrobenzene **22** (893.6 mg, 3.735 mmol) and tin (II) chloride dihydrate (4.410 g, 19.54 mmol) in concentrated hydrochloric acid (20 mL) was refluxed for 2 h. The solution was then cooled, and basified to pH 10 using 5 M aqueous sodium hydroxide. This was then extracted with ethyl acetate (2 × 30 mL), washing with water and brine (30 mL each). The combined organic layers were dried (Na₂SO₄) then evaporated to give **23** as a brownish solid (677.2 mg, 87%).

¹H NMR (400 MHz, *d*₆-DMSO + *d*-TFA) δ ppm 8.2–8.01 (1H, m), 7.64 (1H, d, *J* = 6.54 Hz), 7.53 (1H, d, *J* = 8.66 Hz), 7.07 (0H, s), 6.94 (0H, d, *J* = 8.59 Hz).

MS (ESI +ve) m/z 210.1026 (MH⁺) C₁₃H₁₂N₃⁺ requires 210.1026 ($\Delta=0$ ppm).

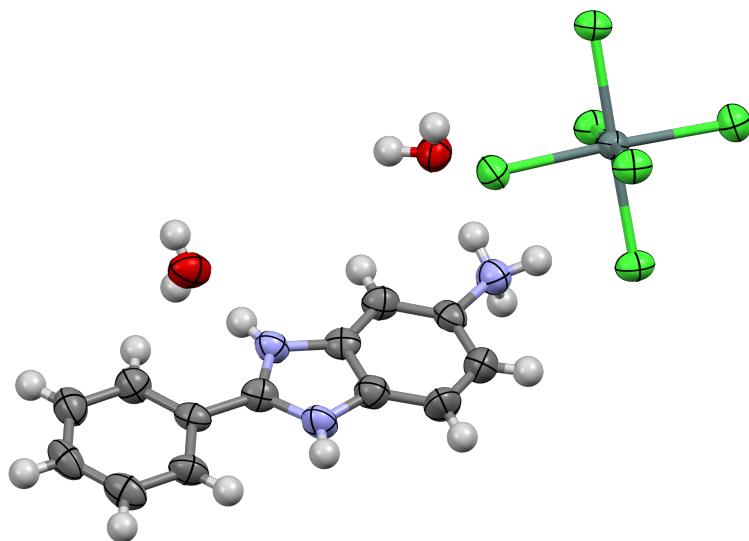


FIGURE 7.18: X-ray crystal structure of **23** as the hexachlorostannate salt.

7.5.2.11 Preparation of 2-(2-phenyl-1*H*-benzo[*d*]imidazol-6-yl)benzo[*d*][1,2]-selenazol-3(2*H*)-one **19**

Diselenide **5** (440.1 mg, 1.100 mmol) was refluxed in thionyl chloride with 2 drops DMF for 2 h. The excess reagent was removed by vacuum distillation, and the residue was dissolved in anhydrous acetonitrile (10 mL). This was added drop-wise to a solution of the aniline **23** (440.4 mg, 2.105 mmol) and TEA (2 mL) in a further 10 mL anhydrous acetonitrile. The mixture was stirred at room temperature for 18 h, then briefly brought to reflux. It was then cooled, diluted with water (30 mL) and extracted into ethyl acetate (3×50 mL). The combined organic layers were washed with water and brine (50 mL each), then dried (MgSO_4), then evaporated onto celite. This was chromatographed on a SNAP 25 g silica cartridge using an ethyl acetate/methanol gradient, affording 2 major fractions:

1. orange solid, 67.7 mg, 8%, **19**, m.p. 170°C (dec.)
2. yellow solid, 166.0 mg, 38%, starting material **23**

¹H NMR (499 MHz, *d*₆-DMSO) δ ppm 8.23–8.17 (2H, m), 8.12 (1H, d, *J* = 8.08 Hz), 8.07 (1H, s), 7.94 (1H, d, *J* = 7.64 Hz), 7.83 (1H, d, *J* = 8.66 Hz), 7.76–7.67 (4H, m), 7.63 (1H, d, *J* = 8.63 Hz), 7.51 (1H, t, *J* = 7.34 Hz).

MS (ESI +ve) m/z 392.0298 (MH⁺) C₂₀H₁₄N₃OSe⁺ requires 392.0297 (Δ=0.26 ppm).

7.5.2.12 Preparation of 2-(4-amino-3-nitrophenyl)benzo[*d*][1,2]selenazol-3(2*H*)-one **14**

Diselenide **5** (1.2025 g, 3.005 mmol) was dissolved in thionyl chloride (20 mL) and refluxed for 30 minutes, and the colour changed from purple to light yellow. Excess thionyl chloride was removed *in vacuo*, and the residue dissolved in anhydrous THF (50 mL). To this was added a solution of 2-nitro-1,4-benzenediamine (991.9 mg, 6.477 mmol) and triethylamine (3 mL, dist. from CaH₂) in anhydrous THF (50 mL). This mixture was stirred at room temperature for 18 h, then filtered, washing the precipitate with water, to afford 2-(4-amino-3-nitrophenyl)benzo[*d*][1,2]selenazol-3(2*H*)-one **14** as a brick red solid (940 mg, 61%, m.p. (DMF) 300.8–302.0 °C).

¹H NMR (400 MHz, *d*₆-DMSO) δ ppm 7.08 (d, *J*=9.00 Hz, 1 H), 7.46 (t, *J*=7.43 Hz, 1 H), 7.55 (s, 2 H), 7.59–7.70 (m, 2 H), 7.86 (d, *J*=7.43 Hz, 1 H), 8.06 (d, *J*=7.83 Hz, 1 H), 8.12 (d, *J*=1.96 Hz, 1 H).

¹³C NMR (100 MHz, *d*₆-DMSO) δ ppm 120.29 (s), 121.49 (s), 126.32 (s), 126.71 (s), 127.77 (s), 128.31 (s), 128.45 (s), 129.68 (s), 132.66 (s), 134.13 (s), 139.33 (s), 145.03 (s), 165.70 (s).

MS (ESI +ve) m/z 335.9882 (MH⁺) C₁₃H₁₀N₃O₃Se⁺ requires 335.9881 (Δ=0.298 ppm).

7.5.2.13 Preparation of *N*-(2-nitro-4-(3-oxobenzo[*d*][1,2]selenazol-2(3*H*)-yl)-phenyl) picolinamide **13**

Picolinic acid (256.6 mg, 2.084 mmol) was dissolved in anhydrous THF (5 mL) and triethylamine (1 mL, dist. from CaH₂), then trichlorobenzoyl chloride (325 uL) was added and the mixture stirred for 10 minutes. The nitroaniline **14** (242.8 mg, 0.956 mmol) was then added, and the mixture stirred under argon for 24 h, during which time the dark red colour faded to give a yellow solution. The mixture was tipped into water (100 mL) and filtered to afford a yellow precipitate, which was recrystallised from pyridine (50 mL) to give 2-(4-amino-3-nitrophenyl)benzo[*d*][1,2]-selenazol-3(2*H*)-one **13** as a yellow solid (93.2 mg, 20%, m.p. 342.7–343.7 °C).

¹H NMR (500 MHz, *d*₆-DMSO) δ ppm 7.51 (t, *J*=7.40 Hz, 1 H) 7.66–7.83 (m, 1 H) 7.94 (d, *J*=7.63 Hz, 1 H) 8.04 (dd, *J*=9.00, 2.40 Hz, 1 H) 8.09–8.17 (m, 2 H) 8.23 (d, *J*=7.78 Hz, 1 H) 8.68 (d, *J*=2.44 Hz, 1 H) 8.73 (d, *J*=9.00 Hz, 1 H) 8.81 (d, *J*=4.43 Hz, 1 H) 12.22 (s, 1 H)

MS (ESI +ve) m/z 441.0100 (MH⁺) C₁₉H₁₃N₄O₄Se⁺ requires 441.0096 (Δ=0.907 ppm).

References

- (1) Australian Bureau of Statistics 3303.0 - Causes of Death, Australia, 2015, 2016.
- (2) Mathers, C. New report analyses costs of cancer in Australia, 1998.
- (3) Rankin, J. Cancer breakthroughs trigger big pharma interest in drugs and deals, 2015.
- (4) Hanahan, D.; Weinberg, R. A. *Cell* **2011**, *144*, 646–674,
DOI: [10.1016/j.cell.2011.02.013](https://doi.org/10.1016/j.cell.2011.02.013).
- (5) Ward, J. F. *International Journal of Radiation Biology* **1994**, *66*, 427–32,
DOI: [10.1017/CBO9781107415324.004](https://doi.org/10.1017/CBO9781107415324.004).

- (6) Ward, J. In *Progress in Nucleic Acid Research and Molecular Biology*, 1988, pp 95–125,
DOI: [10.1016/S0079-6603\(08\)60611-X](https://doi.org/10.1016/S0079-6603(08)60611-X).
- (7) Evans, M. D.; Cooke, M. S. *BioEssays* **2004**, *26*, 533–542,
DOI: [10.1002/bies.20027](https://doi.org/10.1002/bies.20027).
- (8) Rodney Withers, H. *The Lancet* **1992**, *339*, 156–159,
DOI: [10.1016/0140-6736\(92\)90218-R](https://doi.org/10.1016/0140-6736(92)90218-R).
- (9) Kouvaris, J. R.; Kouloulias, V. E.; Vlahos, L. J. *The Oncologist* **2007**, *12*, 738–747,
DOI: [10.1634/theoncologist.12-6-738](https://doi.org/10.1634/theoncologist.12-6-738).
- (10) Miyahara, T.; Nakatsuji, H.; Sugiyama, H. *Journal of Physical Chemistry A* **2012**, *117*, 42–55,
DOI: [10.1021/jp3085556](https://doi.org/10.1021/jp3085556).
- (11) Teng, M.; Usman, N.; Frederick, C. A.; Wang, A. H. *J. Nucleic Acids Research* **1988**, *16*, 2671–2690,
DOI: [10.1093/nar/16.6.2671](https://doi.org/10.1093/nar/16.6.2671).
- (12) Giese, B. *Annual review of biochemistry* **2002**, *71*, 51–70,
DOI: [10.1146/annurev.biochem.71.083101.134037](https://doi.org/10.1146/annurev.biochem.71.083101.134037).
- (13) Kakkar, R.; Suruchi; Pathak, M. *Journal of Molecular Structure: THEOCHEM* **2005**, *714*, 35–42,
DOI: [10.1016/j.theochem.2004.09.045](https://doi.org/10.1016/j.theochem.2004.09.045).
- (14) Beno, B. R.; Yeung, K. S.; Bartberger, M. D.; Pennington, L. D.; Meanwell, N. A. *Journal of Medicinal Chemistry* **2015**, *58*, 4383–4438,
DOI: [10.1021/jm501853m](https://doi.org/10.1021/jm501853m).
- (15) Balkrishna, S. J.; Bhakuni, B. S.; Chopra, D.; Kumar, S. *Organic Letters* **2010**, *12*, 5394–5397,
DOI: [10.1021/ol102027j](https://doi.org/10.1021/ol102027j).
- (16) Fellowes, T.; White, J. M. *CrystEngComm* **2019**, *21*, 1539–1542,
DOI: [10.1039/C8CE01853G](https://doi.org/10.1039/C8CE01853G).

- (17) Klapars, A.; Huang, X.; Buchwald, S. L. *Journal of the American Chemical Society* **2002**, *124*, 7421–7428,
DOI: [10.1021/ja0260465](https://doi.org/10.1021/ja0260465).
- (18) Altman, R. A.; Buchwald, S. L. *Nature Protocols* **2007**, *2*, 2474–2478,
DOI: [10.1038/nprot.2007.364](https://doi.org/10.1038/nprot.2007.364).
- (19) Sherborne, G. J.; Adomeit, S.; Menzel, R.; Rabehah, J.; Brückner, A.; Fielding, M. R.; Willans, C. E.; Nguyen, B. N. *Chem. Sci.* **2017**, 7203–7210,
DOI: [10.1039/C7SC02859H](https://doi.org/10.1039/C7SC02859H).

Chapter 8

Concluding remarks

IM DONE YAY

