

Package ‘myFun’

September 21, 2023

Type Package

Title myFun is a collection of my favorite R functions, packaged for simplicity

Version 1.0.2

Date 2023-09-21

Author Tom Lesluyes [aut, cre]

Maintainer Tom Lesluyes <lesluyes.tom@iuct-oncopole.fr>

Description My utility functions for R.

URL <https://github.com/tlesluyes/myFun>

BugReports <https://github.com/tlesluyes/myFun/issues>

License GPL-3 + file LICENSE

Encoding UTF-8

LazyData true

Depends R (>= 3.4.0)

Imports GenomicRanges,
IRanges,
doParallel,
foreach

Roxygen list(markdown = TRUE)

RoxygenNote 7.2.3

R topics documented:

adjustPositions	2
checkGRlist	3
computeISA	3
computeMD	4
generate_cytoband_and_CHRsize	5
harmonizeGRanges	6
load_CHRsize	7
load_cytoband	8
occurrenceGRanges	8

Index**10**

adjustPositions	<i>adjustPositions</i>
-----------------	------------------------

Description

Adjust genomic positions

Usage

```
adjustPositions(  
  DF,  
  CHRsize,  
  chr_column = "chr",  
  start_column = "start",  
  end_column = "end",  
  suffix = "_adj"  
)
```

Arguments

DF	a data.frame
CHRsize	a data.frame from the load_CHRsize function
chr_column	a column name with chromosome information (default: "chr")
start_column	a column name with start position (default: "start")
end_column	a column name with end position (default: "end")
suffix	a suffix for the adjusted positions (default: "_adj")

Details

This function adjusts genomic positions according to the chromosome sizes. The first nucleotide of chromosome 2 corresponds to the size of the chromosome 1 + 1bp and so on.

Value

A data.frame with adjusted genomic positions

Author(s)

tlesluyes

Examples

```
DF=data.frame(chr=c(1:3), start=rep(1e6, 3), end=rep(125e6, 3))  
load_CHRsize("hg19")  
adjustPositions(DF, CHRsize)
```

`checkGRlist`*checkGRlist*

Description

Check that the given object is a list of GRanges objects

Usage

```
checkGRlist(myGRList)
```

Arguments

`myGRList` a list of GRanges objects

Details

This function checks that the given object is a list of GRanges objects.

Value

TRUE if the input is a list of GRanges objects

Author(s)

tlesluyes

Examples

```
require("GenomicRanges")
GR1=GRanges(seqnames="1", ranges=IRanges(start=1, end=1000))
GR2=GRanges(seqnames="1", ranges=IRanges(start=10, end=2000))
checkGRlist(list(GR1, GR2))
```

`computeISA`*computeISA*

Description

Compute the inter-sample agreement (ISA)

Usage

```
computeISA(GR1, GR2, CNstatus = "CNstatus")
```

Arguments

GR1	a GRanges object corresponding to a single CNA profile
GR2	a GRanges object corresponding to a single CNA profile
CNstatus	a metadata column name for the copy-number status (default: "CNstatus"). Can be total (e.g. "3") or allele-specific (e.g. "2+1")

Details

This function computes the inter-sample agreement (ISA) between two profiles (as GRanges objects). This corresponds to the fraction of the genome (%) with the same CN status.

Value

A percentage representing the ISA

Author(s)

tlesluyes

Examples

```
require("GenomicRanges")
GR1=GRanges(seqnames=rep("1", 3),
             ranges=IRanges(start=c(1, 1001, 10001),end=c(1000, 10000, 20000)),
             CNstatus=c("1+1", "2+1", "1+1"))
GR2=GRanges(seqnames=rep("1", 2),
             ranges=IRanges(start=c(500, 10001),end=c(10000, 25000)),
             CNstatus=c("2+1", "1+1"))

# in this example:
#   Region 500-1000 (size=501) is 1+1 for GR1 and 2+1 for GR2
#   Region 1001-20000 (size=19000) is identical between GR1 and GR2 (both 2+1 and 1+1)
#   ISA is: 19000/19501 = 97.43%
computeISA(GR1, GR2)
```

computeMD

computeMD

Description

Compute the Manhattan distance (MD)

Usage

```
computeMD(GR1, GR2, nMajor = "nMajor", nMinor = "nMinor", convertMb = FALSE)
```

Arguments

GR1	a GRanges object corresponding to a single CNA profile
GR2	a GRanges object corresponding to a single CNA profile
nMajor	a metadata column name for the major allele (default: "nMajor")
nMinor	a metadata column name for the minor allele (default: "nMinor")
convertMb	a boolean, the MD will be converted to megabases if set to TRUE (default: FALSE)

Details

This function computes the Manhattan distance (MD) between two profiles (as GRanges objects).

Value

A numeric value representing the MD

Author(s)

tlesluyes

Examples

```
require("GenomicRanges")
GR1=GRanges(seqnames=rep("1", 3),
             ranges=IRanges(start=c(1, 1001, 10001),end=c(1000, 10000, 20000)),
             nMajor=c(1, 2, 1),
             nMinor=c(1, 1, 1))
GR2=GRanges(seqnames=rep("1", 2),
             ranges=IRanges(start=c(500, 10001),end=c(10000, 25000)),
             nMajor=c(2, 1),
             nMinor=c(1, 1))

# in this example:
#   Region 500-1000 (size=501) is 1+1 for GR1 and 2+1 for GR2
#   Region 1001-20000 (size=19000) is identical between GR1 and GR2 (both 2+1 and 1+1)
#   MD is: (abs(2-1)+abs(1-1))*501 = 501
computeMD(GR1, GR2)
```

generate_cytoband_and_CHRsize

generate_cytoband_and_CHRsize

Description

Generate cytoband and CHRsize information

Usage

```
generate_cytoband_and_CHRsize(cytoband_file)
```

Arguments

cytoband_file a cytoband file

Details

This function generates cytoband and CHRsize information from a cytoband file. This can be obtained from the UCSC table browser -> select a genome/assembly -> "Mapping and Sequencing" -> "Chromosome Band" (not the ideogram version!) -> "get output" -> Remove the first "#" character (keep the header!).

Value

A list with both the cytoband and CHRsize information

Author(s)

tlesluyes

See Also

load_CHRsize("hg38"); load_cytoband("hg38")

harmonizeGRanges

harmonizeGRanges

Description

Harmonize GRanges objects

Usage

```
harmonizeGRanges(myGRList, cores = 1)
```

Arguments

myGRList a list of GRanges objects, each object should correspond to one CNA profile
cores a numeric, the number of cores to use (default: 1)

Details

This function harmonizes GRanges objects by keeping only regions covered by all samples.

Value

A list of harmonized GRanges objects

Author(s)

tlesluyes

Examples

```
require("GenomicRanges")
GR1=GRanges(seqnames="1", ranges=IRanges(start=1, end=1000), nMajor=1, nMinor=1)
GR2=GRanges(seqnames="1", ranges=IRanges(start=10, end=2000), nMajor=2, nMinor=1)
harmonizeGRanges(list(GR1, GR2))
```

load_CHRsize	<i>load_CHRsize</i>
--------------	---------------------

Description

Load CHRsize information

Usage

```
load_CHRsize(assembly)
```

Arguments

assembly	an assembly (hg19 or hg38)
----------	----------------------------

Details

This function loads CHRsize information for a given assembly. It is then available as a data.frame called CHRsize in the environment.

Value

A data.frame with the CHRsize information

Author(s)

tesluyes

Examples

```
load_CHRsize("hg38"); head(CHRsize)
```

load_cytoband	<i>load_cytoband</i>
---------------	----------------------

Description

Load cytoband information

Usage

load_cytoband(assembly)

Arguments

assembly an assembly (hg19 or hg38)

Details

This function loads cytoband information for a given assembly. It is then available as a data.frame called cytoband in the environment.

Value

A data.frame with the cytoband information

Author(s)

tlesluyes

Examples

load_cytoband("hg38"); head(cytoband)

occurrenceGRanges	<i>occurrenceGRanges</i>
-------------------	--------------------------

Description

Get the occurrence of events

Usage

occurrenceGRanges(myGRList, myMetadata)

Arguments

myGRList a list of GRanges objects, each object should correspond to one CNA profile
myMetadata a vector of metadata to consider

Details

This function gets the occurrence of events in a list of GRanges objects. All objects must have the same metadata columns and metadata must be TRUE/FALSE.

Value

A GRanges object with nSamples as the total number of samples and metadata columns with the occurrence of events

Author(s)

tlesluyes

Examples

```
require("GenomicRanges")
GR1=GRanges(seqnames="1", ranges=IRanges(start=1, end=1000), Gain=TRUE, Loss=FALSE)
GR2=GRanges(seqnames="1", ranges=IRanges(start=10, end=2000), Gain=FALSE, Loss=TRUE)
occurrenceGRanges(list(GR1, GR2),c("Gain", "Loss"))
```

Index

`adjustPositions`, [2](#)

`checkGRlist`, [3](#)

`computeISA`, [3](#)

`computeMD`, [4](#)

`generate_cytoband_and_CHRsize`, [5](#)

`harmonizeGRanges`, [6](#)

`load_CHRsize`, [7](#)

`load_cytoband`, [8](#)

`occurrenceGRanges`, [8](#)