

# SPECTROSCOPY

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*Every honest researcher I know admits he's just a professional amateur.  
He's doing whatever he's doing for the first time. That makes him an  
amateur. He has sense enough to know that he's going to have a lot of  
trouble, so that makes him a professional.*

— Charles F. Kettering (1876-1958) (Holder of 186 patents)

## ACKNOWLEDGEMENTS

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I express my sincere gratitude to our instructors, Dr. K. S. Viswanathan and Dr. K. R. Shamasundar, for bringing the subject to life and helping us discover, in depth, the science behind the procedures.

I also thank Vivek Sagar (MS11017) for his contribution to this report as my lab-partner, who made the task of performing experiments immensely comfortable and productive at the same time.



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## Part I

### EXPERIMENTS





## INTRODUCTION TO INFRA RED AND ULTRAVIOLET - VISIBLE SPECTROSCOPY

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August 9, 2012

### 1.1 OBJECTIVE

To

1. understand the method of Fourier Transformed Infrared and Ultraviolet spectroscopy
2. understand the preparation substances for analysis
3. handle and operate the Spectroscopes provided

### 1.2 THEORY

#### 1.2.1 Basic Concept

To find the presence of elements and/or compounds within a given substance, we can use spectroscopy techniques, specially when their concentrations are small and they satisfy certain requirements. The essential idea behind this measurement comes from the fact that elements/compounds absorb lights of certain frequencies to get to a higher energy state. These frequencies are mostly discrete as they correspond to quantized energy levels. This energy could be absorbed for, say, changing the vibrational energy (IR Spectroscopy) or for exciting an electron in the substance to a higher energy level (UV-vis Spectroscopy). We note here that these quantized energy levels are properties of individual substances and are, for most practical purposes, unique.

For the analysis to be possible, the first condition is that the substance must *absorb* light incident to it. Granted this, we can obtain an absorption spectrum for the given substance, which behaves like a fingerprint of the substance. This can thus be used to not only identify the compound, but also to quantify it. For identification, in the simplest case, we simply need to observe the frequency corresponding to the peaks in the absorption spectrum and match it with the known/expected substance(s). Quantification harnesses a rather “obvious” law, termed *Beer-Lambert’s Law*. In the simplest form, the law



How much absorption, well, the limit comes from the sensitivity of the experimental setup and concentration of substance given.

quantifies the intuitive notion; higher the concentration of the analyte, higher is the absorption. The relation is given as

$$T = \frac{I}{I_0} = 10^{-\alpha l} = 10^{-\epsilon lc} \quad (1)$$

where  $I$  is intensity of incident light,  $I_0$  is intensity of transmitted light,  $\epsilon$  is molar absorptivity,  $l$  is the optical path length, and  $c$  is molar concentration.

### 1.2.2 Infra Red Spectroscopy

Infra Red spectroscopy usually deals with energies of the level that cause change in vibrational energies. The wavelength ranges from  $2.5\mu\text{m}$  to  $25\mu\text{m}$  ( $4000\text{cm}^{-1}$  to  $400\text{cm}^{-1}$ ). These energies are characteristic for different bonds which is how, using the spectrum, we can identify (and quantify) the bonds present and thus the compound.

The way the spectroscope works for Infra Red, is rather interesting and ingenious. The setup uses a Michelson interferometer.

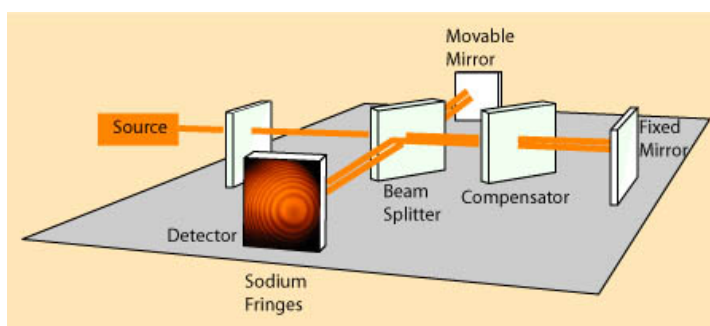




Figure 1: Michelson Interferometer [? ]

 Coherent means phase locked and Monochromatic means single frequency

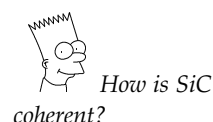
Before complicating things, let us assume that the source of light is coherent and monochromatic. Now in the [Figure 1](#), we assume the source of light to be the light transmitted through the sample to be analysed. Say the interference at the detector is, at the given configuration, constructive. If we move the movable mirror by  $\lambda/4$ , where  $\lambda$  is the wavelength of the light, then the detector will receive a dark, destructive interference. If we plot the intensity at the detector as a function of displacement of the moveable mirror, we will, in this case, receive a sine wave.

 Note the fact that here, the measurement is simultaneous!

Now let us crank it up a notch. Let us consider the light to still be coherent, but not monochromatic. Let the source of light contain all the transmitted frequencies. Now if the intensity is plotted against the displacement of the moveable mirror, we will get a superposition of sine waves, and we already know how to decompose them to find individual frequencies using *Fourier Transformation*. This essentially

gives us the spectrum, with wavenumber (dimensions of one over distance) on one axis, and intensity on the other.

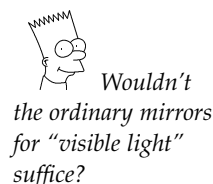
The source of infra-red light is a Silicon Carbide rod, that is heated to produce the desired radiations. The detector is capacitive, or so we were told.



### 1.2.3 Ultraviolet-Visible Spectroscopy

Here the wavelengths range from 200nm to 900nm. This range is often in the range of differences of electronic energy levels. The light source is a Tungsten-Neon lamp. In this technique, usually movable diffraction grating is used to split the transmitted light into its component frequencies, "one" of which goes to an intensity detector. Which component is detected depends on the orientation of the diffraction grating. Using this information, an intensity vs. frequency plot is generated which is the desired spectrum.

It must be noted here that the process of moving the orientation of the grating, requires time, which means that the measurement for various frequencies' intensities is not simultaneous, which in certain cases might not be suitable. One solution to such a problem is the use of multiple (arrayed) detectors. Yet it doesn't explain why a Michelson interferometer can't be used in this case. The answer to this comes from the fact that the wavelength in this case is much smaller, thereby complicating the task for preparing reflectors.



## 1.3 EXPERIMENTAL DETAILS

For this session, both IR and UV-visible spectroscopy techniques were demonstrated to us in groups of two. For IR spectroscopy, we used KBr as the base and Benzoic Acid as the analyte. For UV-vis, we used water as the base and  $\text{KMnO}_4$  as the analyte.

### 1.3.1 Infrared

#### *Procedure*

The general procedure is as follows:

- A. Weighed 100mg of KBr and ground it well.
- B. Transferred it the dye (palletizing module).
- C. Converted it into a pellet using the Hydraulic Press (about 8 tons of pressure).
- D. Did IR spectroscopy.
- E. Weighed 100mg of KBr and with it, 2mg of sample (in this case Benzoid Acid) and repeated from [item B](#).

Details about the IR spectroscopy are as follows:

- A. Ran the background to get rid of noise.
- B. Did the accumulation 16 times (set the option).
- C. Subtracted the first spectroscopy from the second

*Observation*

Detected the carboxylic group (peaks for OH and C = O)

1.3.2 UV-Vis

*Procedure*

The general procedure is as follows:

- A. Took a suitable amount of water.
- B. Did UV-Vis spectroscopy.
- C. Again took water and added a suitable amount of  $\text{KMnO}_4$  and mixed it well.
- D. Did UV-Vis spectroscopy.

Details about the UV-vis spectroscopy is corresponding to the previous section.

*Observation*

Peaks were detected in the range 520nm-550nm, complimentary to Purple as expected.

1.4 ACKNOWLEDGEMENTS

I thank the PhD students, Ms. Shruti and Ms. Shilpa, who helped us with the session and performed the experiment. They also walked us through the procedure for using the apparatus.

## EXPERIMENT 1A: UV-VISIBLE SPECTROSCOPY

August 16, 2012

## 2.1 OBJECTIVE

To

1. prepare 10, 25, 50 and 75 ppm solutions of Benzoic Acid in water
2. use one of these for obtaining a spectrogram
3. use three of these for calibration and find the concentration of the third experimentally

## 2.2 THEORY

This section is the same as and has been covered in the previous experiment.

A few simple relations must be stated however, for clarity. Transmissivity or Transmission (T) is related to observables by Equation 2, where I is intensity of the light transmitted,  $I_0$  is intensity of incident light,  $\epsilon$  is extinction coefficient, l is the path length and c is concentration of the solution.

Relation between Absorbance (A) and intensity of light is as given by Equation 3. Clearly, A has a linear relation with  $\epsilon$  as given by Equation 4 and we'll use this very relation to find the concentration of the 'unknown' sample.

$$T = \frac{I}{I_0} = 10^{-\epsilon lc} \quad (2)$$

$$A = -\log_{10}\left(\frac{I}{I_0}\right) \quad (3)$$

$$A = \epsilon lc \quad (4)$$

## 2.3 PROCEDURE

1. First calculated the mass of Benzoic Acid required for making a 25mL a 250 ppm solution in water, to be 6.25 mg.



Just in case,  
1ppm = 1μL/mL

Mass of salt	0.0066	g
Mass of paper	0.0001	g
Mass of Benzoic Acid	6.5	mg
Concentration of Benzoid Acid Used	$\frac{6500}{25} = 260$	ppm

Table 1: For the Stock Solution of Benzoic Acid

10 ppm	$\frac{50}{260} = 0.192$	mL
25 ppm	$\frac{125}{260} = 0.480$	mL
10 ppm	$\frac{250}{260} = 0.961$	mL
10 ppm	$\frac{50}{375} = 1.442$	mL

Table 2: For varying concentrations of Benzoic Acid Solution



Used the  
upper meniscus  
consistently for  
volume  
measurements



But does  
that mean it  
wouldn't affect the  
result?

2. Weighed roughly the calculated amount of Benzoic Acid and transferred it to a volumetric flask of 25mL, and made the volume 25mL. This solution is henceforth referred to as the stock solution.
3. Calculated the volume of the stock solution required to prepare 10, 25, 50 and 75 ppm, 5 mL Benzoic Acid solutions.
4. Using a graduated pipette, measured the precise volume in accordance with the calculations and transferred it to a 5mL volumetric flask, and filled it to 5mL of water. Repeated this step for all four concentrations and labelled the flasks accordingly.
5. Calibrated the spectroscope with water first to suppress noise, if any.
6. Used one of the concentrations to generate a spectrogram.
7. Identified the maximum wavelength for use in Beer-Lambert law.
8. Used three of the concentrations to calibrate and fourth as an unknown, using the spectroscope.
9. Printed the spectrogram and noted the observations off of the screen for the known and unknown concentrations.

## 2.4 CALCULATIONS AND MEASUREMENTS

Refer to [Table 1](#) for the stock solution and to [Table 2](#) for the variants.

WAVELENGTH ( $\lambda$ )	ABSORBANCE
544.00nm	1.710
524.00nm	1.915

Table 3: Wavelengths Absorbed

CONCENTRATION (c)	ABSORBANCE
75ppm	1.902
50ppm	0.971
10ppm	0.237

Table 4: Absorption for various Concentrations

## 2.5 SPECTROSCOPIC OBSERVATIONS AND ANALYSIS

The maximum wavelength ( $\lambda_{\max}$ ) was found to be 524.00nm. Refer to [Table 3](#). The Absorbance (A) for various concentration is given in [Table 4](#)

Absorbance of 'unknown' sample (25 ppm): 0.489

In accordance with the application, the best fit for the three points was not linear and the concentration of the unknown sample was calculated to be 30.0720 ppm which amounts to a 20% error.

In accordance with least square straight line fit, we get the slope as  $0.0249395 \pm 20.34\%$  but the intercept as  $-0.0856124 \pm 310.2\%$ . In accordance with these numbers, the concentration works out to be 16.174 ppm, which is 35% off. Refer to [Figure 2](#) for details.



Expected  
accuracy is 2 – 3%

## 2.6 DISCUSSION

The accuracy of this experiment, as told by our instructor, is expected to be less than 3%. Since we clearly failed at achieving that number, we analyse in this section, what could've caused the enormous error.

The error could be caused by making errors in any of the following processes

1. Weighing
2. Volume measurement
  - a) Pipetting
  - b) Filling the Volumetric Flask

Despite taking sufficient care, even if one of the steps mentioned above go wrong, for even one of the samples, since the number of samples for calibration is just three, the entire experiment can, and in this case, did go wrong.

Thus for ensuring such errors do *not* crop up in the future, we can attempt the following

## 1. Weighing:

- a) Put the paper on the weighing scale and tare it
- b) Add the required amount of substance ensuring nothing gets wet
- c) Keep the container, viz. Volumetric Flask, with accessories, viz. Funnel etc. near the setup
- d) After transferring the contents, weigh the paper again to ensure everything got transferred

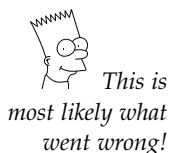
## 2. Volume Measurement

## a) Pipetting

- i. Read the upper meniscus consistently (for this experiment and in general for opaque solutions)
- ii. Use graduated pipettes that can provide sufficient precision
- iii. *Ensure the liquid doesn't stick on the walls*

## b) Filling Volumetric Flask

- i. Again, use the upper meniscus
- ii. Use a dropper to add towards the end to avoid accidentally adding extra volume

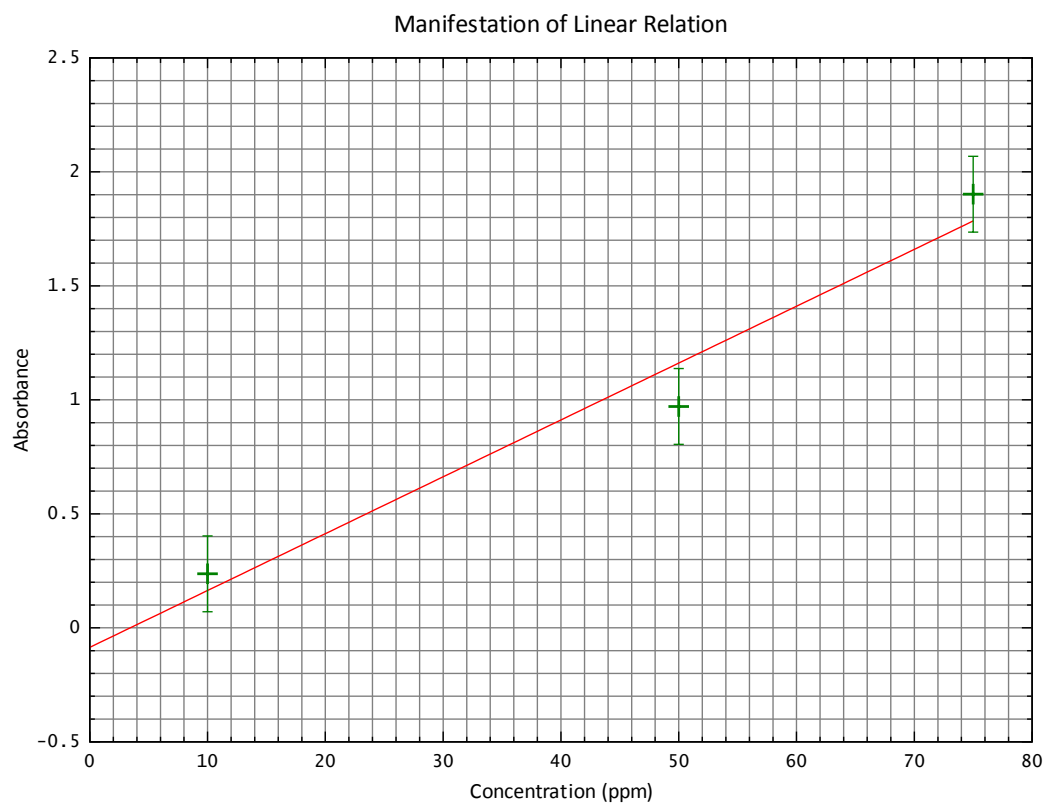


## 2.7 ACKNOWLEDGEMENTS

I thank Ms. Sumyra, the PhD student, who helped us with the use of the spectroscope. I also acknowledge the contribution of Ms. Athira J. Nair and Mr. Arpit Porwal for the performance of the experiment as team members.



## Experiment: UV-visible Spectroscopy



Slope of Best Fit Line : +0.1663  
Intercept of Best Fit Line : +0.0249

Performed on: August 16, 2012  
Performed by: Athira, Arpit and Atul

Figure 2: Least Square Fit



## Part II

### THE SHOWCASE

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## COLOPHON

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