

Graduate School of Sciences and Technology for Innovation,  
Yamaguchi University

Division of Fundamental Sciences

# A Computational Model of Cell Migration of Keratocytes

Yu Tokunaga

January 9, 2019

# Contents

<b>1</b>	<b>Introduction</b>	<b>1</b>
1.1	Introduction . . . . .	1
<b>2</b>	<b>Keratocytes</b>	<b>3</b>
2.1	Characteristics of Cell Migration of Keratocytes . . . . .	3
2.2	Molecular Mechanism of Cell Migration . . . . .	3
<b>3</b>	<b>Simulation Methods</b>	<b>5</b>
3.1	Simulation Methods of Cell Membrane Molecules . . . . .	5
3.2	Simulation Methods of Actin Molecules . . . . .	5
3.2.1	Actin Porymerazation . . . . .	5
3.2.2	Actin Retrograde Flow . . . . .	6
3.3	Initial Condition and Relocation Condition . . . . .	6
<b>4</b>	<b>Results</b>	<b>9</b>
4.1	Simulation Results . . . . .	9
4.2	Discussion . . . . .	9
<b>5</b>	<b>Conclusion</b>	<b>11</b>
5.1	Morphology of Keratosite and Its Motor Function . . . . .	11
5.2	Future Prospects . . . . .	11



# Chapter 1

## Introduction

### 1.1 Introduction

Fish epidermal cells called keratocytes have the characteristic of keeping a half-moon shape when migrating. In order to explain how strange this is, we introduce the migration of *Amoeba proteus*, which is a common amoeba cell. *Amoeba proteus* migrates irregularly while repeating complicated morphological changes when migrating. The migration of *Amoeba proteus* is completed by the pseudopodium made by stretching from the cell membrane landing away from itself. In other words, in general amoeba cells, migration does not occur without causing shape change. However, after the shape changes to a half-moon shape, keratocytes can advance forward without changing its shape. It is not clear why keratocytes form a half moon shape, why keratocytes can migrate without changing its shape.

Keratocytes are about  $70\text{ }\mu\text{m}$  in size and is a columnar cell present on the back side of the fish scales. When a fish bleeds due to trauma, it gathers to cover the wound. At this point, Keratocytes is doing the amoeba movement. The most interesting feature of keratocytes is this amoeba movement. Amoeba Movement of keratocytes contrasts with the typical Amoeba movement and moves straight forward while maintaining a shape like a half-moon shape during migrating.

Humans walk by bending and floating the legs when moving from the spot. As with other organisms, if the form of the organism and its motor function are closely related, what is the effect of the half-moon shape formed by the migration of keratocytes on the motor function of keratocytes? Elucidating this problem is interesting to

know the relationship between the form of organism and motor function.

In this study, the purpose is to clarify the relationship between keratocytes morphology and motor function in cell migration. Specifically, we will elucidate the intracellular mechanism that makes it possible to go straight while keeping the cell shape half-moon.

The structure of this paper is as follows. In Chapter 2, we explain the specific features of keratinocytes and the outline of cell migration mechanism. In Chapter 3, details of simulation experiment method will be described. Section 4 presents the results of simulation experiments and discusses the considerations. In the last chapter, we explain the improvements and future prospects of the facts and methods obtained from this survey.



# Chapter 2

## Keratocytes

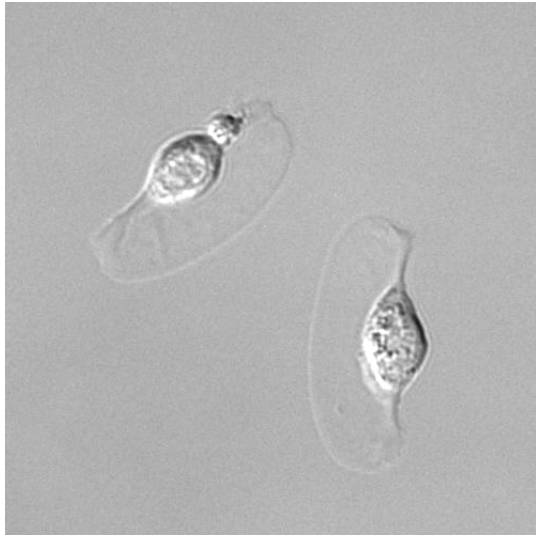


Figure 2.1: Keratocytes during cell migration. (Source: Takako Tanaka, Iwate Lab).

### 2.1 Characteristics of Cell Migration of Keratocytes

A photograph of the actual keratocytes are shown in Figure 2.1. Keratocytes are migratory fish epidermal cells. Migrating cells are cells that can freely move around in the same way as lymphocytes and the like. Keratocytes advances by about one body length in one minute. The physiological findings of cell migration will be introduced in the next section.

Studies using keratocytes in the field of biophysics have often revealed facts in cell migration mechanisms. Svitkina et al. showed that the forward translocation of the cell body and retrograde flow

in the lamellipodia are both driven by contraction of an actin – myosin network in the lamellipodial/cell body transition zone [1]. The importance of both AP and ARF was suggested. Nakata et al. Focused on pulling force against temperament and flow rate gradient of actin retrograde flow (ARF), and reported that stress fiber (SF) plays an important role in morphological control of keratinocytes [2]. Asano et al. Showed that movement similar to keratocytes was observed when AmiB gene was deleted from ameba cells of cellular slime mold *Dictyostelium discoideum* which performs typical ameba movement [3]. Even in other ameboid cells, if some condition is satisfied, there is a possibility of maintaining a half-moon shape, which means that morphogenesis of keratocyte is not unique to keratocyte and is a physical mechanism causing cell migration. Okimura et al. showed that the rotation of these stress fibers plays the role of a “wheel” in crawling migration of keratocytes [4]. Removal of the stress fibers decreased migration velocity and induced the collapse of the left-right balance of crawling migration. From the previous studies above, it is known that both AP and ARF play an important role in cell migration of keratocytes.

### 2.2 Molecular Mechanism of Cell Migration

A cell has a structure called a cytoskeleton. The cytoskeleton is like bones for humans, but in contrast to static human bones, the cytoskeleton is dynamic and is an

important organelle that maintains the shape of the cell. The main component of the cytoskeleton is the actin molecule. As the actin molecule polymerizes and depolymerizes, the shape of the cell membrane changes. The actin molecule is elongated by polymerization in the cell membrane but retreats in the direction opposite to the elongation direction by the bundle of actomyosin called stress fiber. This phenomenon is called actin retrograde flow, and it is not clear what kind of role it plays in cell migration. The cell membrane extruded by polymerization of the actin molecule becomes the pseudopodia and adheres to the substrate at the advancing position. Thereafter, the adhesion between the substrates of the rear cell membrane is released and dragged forward. By repeating this cycle, cell migration mechanism is formed.

# Chapter 3

## Simulation Methods

### 3.1 Simulation Methods of Cell Membrane Molecules

In the cell membrane model, each cell membrane molecule moves under the resultant of force and velocity from the actin molecule and resistance force proportional to force between the cell membrane molecules. The equation of motion of the cell membrane molecule is as follows.

$$m \frac{d^2 \mathbf{x}_i}{dt^2} = \mathbf{F}_i^m + \mathbf{F}_i^a - \eta \frac{d\mathbf{x}_i}{dt} \quad (3.1)$$

where  $\mathbf{F}^m$  is the elastic force received from the membrane molecule,  $\mathbf{F}^a$  is the repulsive force received from the actin molecule,  $\eta$  is the viscous drag coefficient, which is  $8.9 \times 10^{-6} \text{kg s}^{-1}$ , and  $\mathbf{x}$  is the position vector of the membrane molecule. As shown in the formula, the membrane molecule has the force  $\mathbf{F}^m$  received from the membrane molecule and the force  $\mathbf{F}^a$  received from the actin molecule. That is the force as follows.

$$\mathbf{F}^m = \sum_{\{\forall j \mid \|\mathbf{x}_j - \mathbf{x}_i\| < D_1\}} -k((\mathbf{x}_j - \mathbf{x}_i) - \mathbf{l}_{ij}) \quad (3.2)$$

where  $k$  is a constant,  $D_1$  is a distance range to receive a force, and  $\mathbf{l}_{ij}$  is a vector representing the difference between the two points (membrane molecules  $i$  and  $j$ ) in the initial state.

$$\mathbf{F}^a = \sum_{\{\forall i \mid \|\mathbf{x}_j - \mathbf{B}_i\| < D_2\}} \frac{s}{\|\mathbf{x}_j - \mathbf{B}_i\|} \frac{\mathbf{x}_j - \mathbf{B}_i}{\|\mathbf{x}_j - \mathbf{B}_i\|} \quad (3.3)$$

where  $s$  is a constant, and  $D_2$  is a distance range to receive a force.

### 3.2 Simulation Methods of Actin Molecules

#### 3.2.1 Actin Polymerization

The polymerization of actin has directionality, the end where polymerization is carried out is called barbed-end, and the end which is not done is called pointed-end. Since polymerization is carried out well in a region where many actin molecules are present, the polymerization rate is proportional to the actin concentration in the vicinity. The filament formed by actin polymerization is called F-actin. F-actin on the simulation is treated as a rod. F-actin is defined as the position vector at both ends of the rod. Here, the position vector of pointed-end is  $\mathbf{P}$ , and the position vector of barbed-end is  $\mathbf{B}$ .  $\mathbf{P}$  and  $\mathbf{B}$  are updated step by step to control polymerization and depolymerization.

As described above, the polymerization direction  $\mathbf{L}$  is determined at the initial stage of cell migration. Therefore, the formula for updating the polymerization is

$$\mathbf{B} \leftarrow \mathbf{P} + \mathbf{L}$$

. However, considering the surrounding actin concentration and time step, the formula can be rewritten as follows.

$$\mathbf{B} \leftarrow \mathbf{P} + f^p(c) \mathbf{L} \cdot dt \quad (3.4)$$

where  $f^p(c)$  is a function that proportional to actin concentration  $c$  and  $dt$  is the time step.

Regarding depolymerization, the update formula is as follows.

$$\mathbf{P} \leftarrow \mathbf{P} + f^d(c) \mathbf{L} \cdot dt \quad (3.5)$$



where  $f^d(c)$  is a function inversely proportional to  $c$ . However, these formulas are not done every time, but are done with probability

$$p_i(c) = c_i = \frac{a_i}{N}$$

where  $p_i(c)$  is the probability that depends on the concentration  $c_i$  in region  $i$ ,  $a_i$  is the number of actin in region  $i$ , and  $N$  is the total number of actin. In this simulation, the original space was divided into 16020 spaces ( $i = 16020$ ).

### 3.2.2 Actin Retrograde Flow

Because ARF is often unknown, we simulate with some assumptions. Although the element that causes ARF is a stress fiber, since the cell model of this paper does not introduce a stress fiber, it substitutes the membrane molecule at the end of the cell membrane. Since it is not clear how much the effect on actin molecule also works, it was assumed that the closer to the membrane molecule the stronger the effect.

The cell membrane molecule with the smallest x coordinate is the reference point  $E$ . The calculation formula for implementing ARF is shown below.

$$\begin{cases} B \leftarrow B - \alpha \frac{B - E}{\|B - E\|} & (3.6) \\ P \leftarrow P - \beta \frac{P - E}{\|P - E\|} & (3.7) \end{cases}$$

where the vector  $B$  is barbed-end, the vector  $P$  is pointed-end, and  $\alpha$  and  $\beta$  are constants.

## 3.3 Initial Condition and Relocation Condition

Let us decide the initial condition of actin. The total number of actin is 2000. First, the direction of polymerization is randomly determined. Some actins invade areas

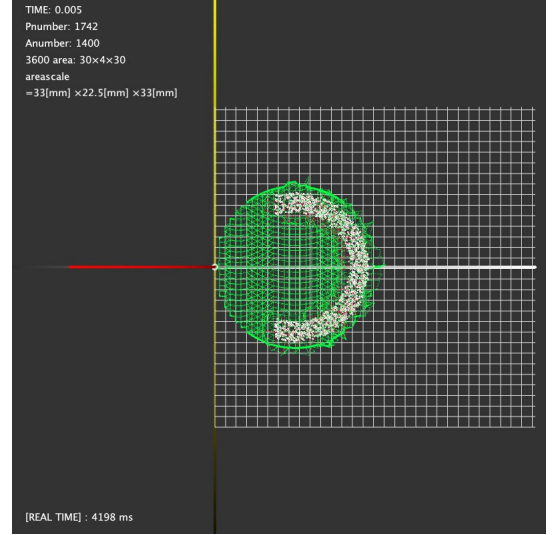


Figure 3.1: Top view of keratocyte model at time  $t = 0$ .

where they can not normally invade due to the progress of polymerization. For example, it is outside the cell membrane and in the nucleus. In such a case, relocation processing is applied to the actin. F-actin escaping from the cell membrane disappears and moves to a random place close to the cell membrane. The region to be repositioned is outside of the cell membrane and regions where the concentration of actin molecule is low.

The initial state in the simulation experiment is shown in Fig. 3.1 and Fig. 3.2.

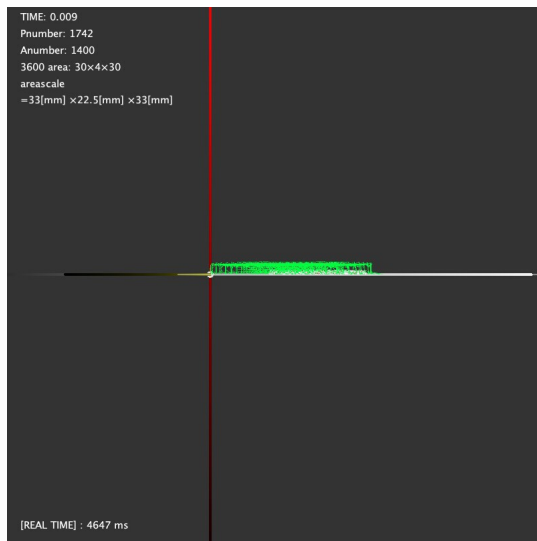


Figure 3.2: Side view of keratocyte model at time  $t = 0$ .



# Chapter 4

## Results

### 4.1 Simulation Results

### 4.2 Discussion



# Chapter 5

## Conclusion

### 5.1 Morphology of Ker- atosite and Its Mo- tor Function

### 5.2 Future Prospects



# Acknowledgements

We gratefully acknowledge the work of past and present members of our laboratory.





# Bibliography

- [1] Tatyana M Svitkina, Alexander B Verkhovsky, Kyle M McQuade, and Gary G Borisy. Analysis of the actin–myosin ii system in fish epidermal keratocytes: mechanism of cell body translocation. *The Journal of cell biology*, Vol. 139, No. 2, pp. 397–415, 1997.
- [2] Takako Nakata, Chika Okimura, Takafumi Mizuno, and Yoshiaki Iwadate. The role of stress fibers in the shape determination mechanism of fish keratocytes. *Biophysical journal*, Vol. 110, No. 2, pp. 481–492, 2016.
- [3] Yukako Asano, Takafumi Mizuno, Takahide Kon, Akira Nagasaki, Kazuo Sutoh, and Taro QP Uyeda. Keratocyte-like locomotion in amib-null dictyostelium cells. *Cell motility and the cytoskeleton*, Vol. 59, No. 1, pp. 17–27, 2004.
- [4] Chika Okimura, Atsushi Taniguchi, Shigenori Nonaka, and Yoshiaki Iwadate. Rotation of stress fibers as a single wheel in migrating fish keratocytes. *Scientific reports*, Vol. 8, No. 1, p. 10615, 2018.