

Trafficking synaptic cargo involves tradeoffs in speed and precision

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ABSTRACT

Nervous system function depends on the appropriate development, maintenance, and plasticity of synaptic connectivity. In order to form and modify synaptic connections, organelles and molecular cargo need to be trafficked from the site of synthesis to the vicinity of the synapse. An open question is how cargo can be delivered to appropriate synapses in spite of the complex dendritic arbors and thousands of postsynaptic sites typical of many neurons. We develop a biophysically-rooted yet general model of microtubule transport that captures known mechanistic properties of intracellular transport. In spite of the assumption that molecular transport responds only to local chemical cues, we show that arbitrarily complex spatial distributions of cargo can be achieved. However, the model reveals important tradeoffs that can be tested experimentally. We find that precise and flexible delivery of synaptic cargo requires the capture of synaptic cargo to occur on a slower timescale than the movement of cargo across the microtubule network. Tuning the transport parameters reveals that optimal solutions exist for specific distributions of synapses, but these solutions sensitively depend on the location as well as the number of synapses, suggesting that biological neurons face a compromise between flexibility of connectivity and efficacy in regulating synaptic function.

Keywords: Synaptic tagging hypothesis, LTP, Transport, Morphology

INTRODUCTION

Neurons often have complex dendritic trees, reflecting the dense and structured synaptic connectivity that supports nervous system function. In order to maintain appropriate connectivity, or to alter it in response to changes in circuit activity, numerous enzymes and macromolecules must be continually replenished and distributed among a large number of synaptic contacts. Although the molecular machinery that solves this logistical problem is characterized in increasing detail (Doyle and Kiebler, 2011; Buxbaum et al., 2014a; Hancock, 2014), there are few attempts to formalize how subcellular transport processes might constrain a neuron's ability to regulate its connectivity and synaptic function.

Not all aspects of neurite metabolism require delivery of cargo from the soma. For example, some forms of long-term potentiation (LTP) depend on local protein synthesis and can function in isolated dendrites (Kang and Schuman, 1996; Aakalu et al., 2001; Vickers et al., 2005; Sutton and Schuman, 2006). Nonetheless, components originating in the soma need to find their way to synapses in the first place, and in the long run, the mRNAs and the machinery that supports local biosynthesis needs to be replenished. Moreover, many long-lasting forms of synaptic plasticity are known to depend on anterograde transport of mRNAs (Kandel, 2001; Puthanveetil et al., 2008) and specific mRNAs are known to be selectively transported to regions of heightened synaptic activity (Steward et al., 1998; Steward and Worley, 2001; Moga et al., 2004) or to developing synaptic contacts (Lyles et al., 2006). These observations fit with the well-known synaptic tagging hypothesis (Frey and Morris, 1997), which proposes that synapses produce a biochemical 'tag' that signals a requirement for synaptic building blocks as part of the plasticity process.

Kinesin and dyenin motor proteins mediate transport along microtubules at rates that far exceed passive diffusion (Block et al., 1990; Smith and Simmons, 2001; Hirokawa et al., 2010; Gagnon and Mowry, 2011; Park et al., 2014). In many cases, the movement of individual cargoes is stochastic and bidirectional (Hancock, 2014), leading to the hypothesis that motor proteins "search" the dendritic tree for dropoff locations, instead of following a pre-determined path (Welte, 2004; Bressloff and Newby, 2009; Doyle and Kiebler, 2011). Thus, the local rate of transport and the decision to release cargo are thought to be controlled by localized signaling pathways, rather than a centralized addressing system. These localized mechanisms

are not fully understood, but are believed to involve transient elevations in second-messengers like $[Ca^{2+}]$ and ADP (Mironov, 2007; Wang and Schwarz, 2009; Buxbaum et al., 2014b), and changes in microtubule-associate proteins (Soundararajan and Bullock, 2014). These pathways are influenced by the spatial pattern of synaptic activity, spontaneous release of calcium from internal stores, and locally released factors from other cells (Wong and Ghosh, 2002; Parrish et al., 2007; Park and Poo, 2013; Zagrebelsky and Korte, 2014). The composition and expression of molecular cargo is also influenced by global factors, such as the cell's average activity level (Spitzer, 2006; Puthanveettil et al., 2008; Turrigiano, 2011) and localization sequences within 3'-untranslated regions of mRNA, which act as a coarse addressing system (Andreassi and Riccio, 2009).

Here, we aim to express and examine existing conceptual models of molecular transport (Doyle and Kiebler, 2011) in a mathematical framework. We develop a minimal biophysical model that can account for activity-dependent accumulation of molecular cargo at synaptic sites in realistic neuronal morphologies, and show how local biochemical signals can modify the movement and/or release of cargo to produce complex spatial patterns. However, we find that decentralized transport strategies face unavoidable tradeoffs between the speed and precision of cargo delivery. Tuning the transport parameters reveals that while optimal solutions exist to circumvent this tradeoff, these solutions sensitively depend on the location as well as the number of synapses, suggesting that biological neurons may face a compromise between flexibility in connectivity and their ability to precisely regulate synaptic function.

RESULTS

Model description

Transport along microtubules is mediated by kinesin and dynein motors, which mediate anterograde and retrograde transport, respectively (Hirokawa et al., 2010; Gagnon and Mowry, 2011). Cargo is often simultaneously bound to both forms of motor protein, resulting in stochastic back-and-forth movements with a net direction determined by the balance of opposing movements (Welte, 2004; Hancock, 2014; Buxbaum et al., 2014a) (Fig. 1A).

To obtain a general model that can accommodate variations in the biophysical details, we consider microtubule-based transport as a biased random walk along a one-dimensional cable

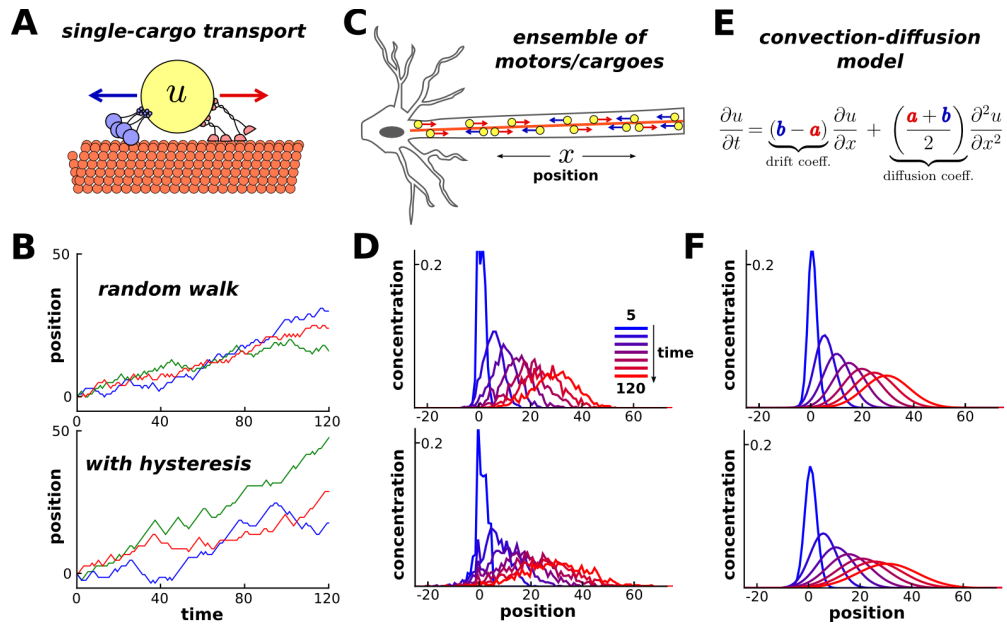


Figure 1. Model of microtubular transport. **(A)** Molecular cargo, u , undergoes stochastic back-and-forth movements driven by opposing motor proteins. **(B)** Three example random walks on a cable representing the movement of individual cargoes. In a simple random walk, each movement is independent of previous movements (top panel); longer run lengths result from adding history-dependence to the model, such that each movement is likely to continue in the same direction as the previous step (bottom panel). **(C)** An ensemble of synaptic cargoes transported along the length of a neurite. **(D)** The concentration profile of transported cargo along a cable over time, simulated as 1,000 independent random walks. Simulations with (bottom) and without (top) history-dependence. **(E)** In the limit of many individual particles, the concentration of u is described by a mass-action model (eqn 1). The parameters a and b respectively scale the anterograde and retrograde rate of transport. **(F)** The mass-action model provides a good fit for the simulations in panel D.

(Bressloff, 2006; Bressloff and Newby, 2009; Newby and Bressloff, 2010; Bressloff and Newby, 2013). For each time step, the cargo moves forwards, backwards, or remains in the same place with different probabilities that together sum to 1. In the simplest model, the probabilities associated with each movement are fixed and independent across each time step, with the forward jump more probable than a reverse jump, leading to a biased random walk (Fig. 1B, top panel). A potentially more realistic model (see, e.g., Muller et al. (2008)) incorporates hysteresis into the biased random walk (see Methods). In this case, the cargo is more likely to continue in the same direction as it moved during the previous time step, leading to extended runs in one direction along the microtubule (Fig. 1B, bottom panel).

While the position of individual cargoes is highly stochastic, the net movement of a population of cargoes (Fig. 1C) is predictable. Figure 1D shows the distribution of 1000 molecules

over time with (top panel) and without (bottom panel) hysteresis. Thus, the bulk trafficking of cargo can be modeled as a deterministic process, which we refer to as the “mass-action model” of transport. The model discretizes the dendritic tree into small compartments, and describes the transfer of cargo between neighboring compartments as reactions with first order kinetics. In a cable with N compartments, the mass-action model is (Fig. 1E):



where u_i is the amount of cargo in each compartment, and a_i and b_i respectively denote local rate constants for anterograde and retrograde transport. The mass-action model with spatially uniform transport rates (Fig. 1F) reproduces the dynamics of the ensemble simulations (Fig. 1D). In this case, the mass-action model maps onto the well-known drift-diffusion partial differential equation (see *Methods*).

The mass-action model makes two important assumptions. It first assumes that the net movement of molecules between neighboring compartments is a stochastic and memoryless process. This assumption is reasonable for cargoes that change direction often; specifically, on a length scale comparable to the size of the compartments. This appears to be the case under many (Muller et al., 2008; Verbrugge et al., 2009), though not necessarily all (Dynes and Steward, 2007; Soundararajan and Bullock, 2014) circumstances. However, we observed that the mass-action model could still be well-fit to simulated data where this assumption was violated (Fig. 1F, lower panel). We also provide a theoretical argument in the *Methods* section showing that the mass-action model only breaks down for very large run lengths (i.e. unidirectional transport).

The second assumption of the mass-action model is that there are many transported particles distributed throughout the dendritic tree. Many types of dendritic cargo are present in high numbers (CITE), and a deterministic model can provide a good approximation in this regime (Fig. 1D).

The model also provides insight into the stochastic dynamics of transport for cargoes with fewer copy numbers. Instead of interpreting u as the amount of cargo in each compartment, this variable (when appropriately normalized) can be interpreted as the probability that a single particle lies within a particular compartment (see *Methods*). Thus, in the stochastic regime, the mass-action model captures average, or expected, behavior of the ensemble.

A simple transport mechanism distributes cargo according to demand

We used the mass-action model for our simulations because it is easy to extend it to branched morphologies (Fig. 2A), and to cases where the trafficking rate constants (a_i and b_i) are spatially non-uniform. This corresponds to the kinetics of motor proteins depending on local biochemical signals (Mironov, 2007; Wang and Schwarz, 2009; Soundararajan and Bullock, 2014).

In the model, the exchange of cargo between compartments approaches a equilibrium, or steady-state (ss), distribution over time (see *Methods*). The equilibrium occurs when no net movement of cargo occurs between connected compartments. Mathematically, this occurs when:

$$\left. \frac{u_p}{u_c} \right|_{ss} = \frac{b}{a} \quad (2)$$

where u_p is the level in the “parent” compartment (closer to soma) and u_c is the level in the “child” compartment (closer to periphery); b and a refer to the trafficking rate constants between this pair of compartments.

Intuitively, in the mass-action model, the rate of cargo transfer is proportional to the concentration of cargo and the trafficking rate constant. Thus, the flow of cargo is equal and opposite when the ratio of cargo matches the reciprocal ratio of the rate constants between a pair of connected compartments.

The above result means that the trafficking rates can be locally tuned to produce a desired spatial distribution of cargo. Denote the desired steady-state level of cargo in each compartment by \tilde{u} (we will soon show how this can be encoded by a localized biochemical signal). To match this desired distribution, the ratio of local trafficking rates should satisfy:

$$\frac{b}{a} = \frac{\tilde{u}_p}{\tilde{u}_c} \quad (3)$$

For example, we can produce a linear expression gradient (Magee, 1998; Hoffman et al., 1997) by setting \tilde{u} directly proportional to the distance from the soma. This produces trafficking rate constants that satisfy $b_i/a_i = i/i + 1$ (where i indexes on increasing distance to the soma). Figure 2B-C shows that this rule produces the expected profile, with the slope of the linear gradient controlled by tuning the total amount of cargo in the cable (Fig. 2C).

Together, this analysis shows that by manipulating local trafficking rates, arbitrary distribu-

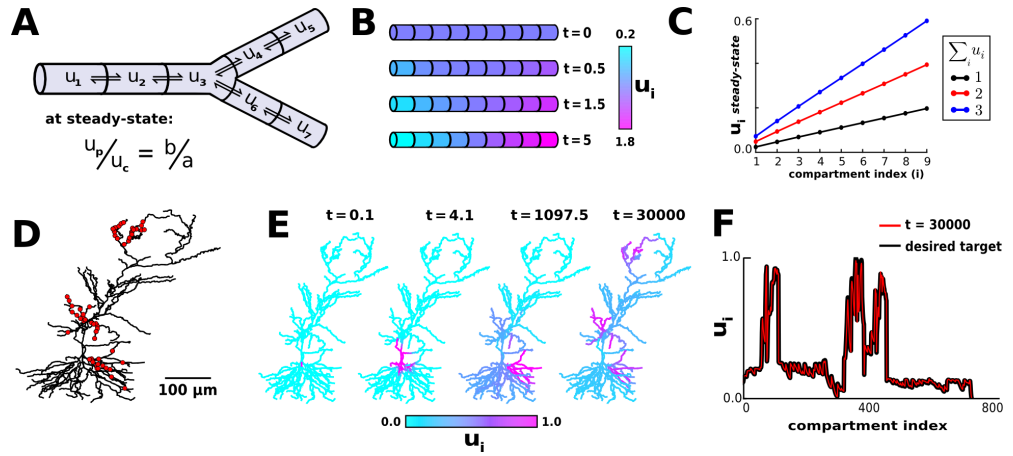


Figure 2. Local trafficking parameters determine the spatial distribution of biomolecules by a simple rule. **(A)** Diagram of the mass action transport model for a simple branched morphology. **(B)** A simulation of a nine compartment cable, with trafficking rate constants set to produce a linear gradient using the steady-state relation shown in panel A. **(C)** The slope of the linear gradient shown in panel B is determined by the total amount of u present in the model; the slope increases as more of u is present, but the linear profile is preserved. **(D)** A model of a CA1 pyramidal cell with 742 compartments based on Migliore and Migliore (2012); excitatory synapses were added at the locations marked by red dots. **(E)** The average membrane potential in each compartment of the CA1 model cell was used to define a desired spatial distribution, and transport rate constants were chosen according to equation (4). The desired spatial distribution of cargo emerges over time, after starting in the soma. **(F)** The desired and steady-state profile of u for each compartment in the simulation shown in panel E.

tions of cargo can be achieved over time. Local trafficking rates, in turn, can be modulated by a simple biochemical reaction resulting from synaptic activation or local release of a growth factor. We can consider the local cytoplasmic calcium concentration, $[ca]_i$, as an candidate second messenger. Increases in $[ca]_i$ simultaneously arrest anterograde and retrograde microtubular transport for certain cargoes (Wang and Schwarz, 2009). Thus, for any pair of compartments, the anterograde rate constant is determined by calcium in the parent compartment, $a = f([ca]_p)$, and the retrograde rate constant is determined by calcium in the child compartment, $b = f([ca]_c)$, where f is a function that describes how calcium alters the transport rates. This results in:

$$\frac{b}{a} = \frac{f([ca]_c)}{f([ca]_p)} = \frac{\tilde{u}_p}{\tilde{u}_c} \quad (4)$$

where $\tilde{u}_i = 1/f([ca]_i)$.

Thus, in principle, local calcium transients that serve to arrest transport could provide a mechanism for distributing cargo to an arbitrary target profile. To demonstrate the plausibility

of this model in a complex morphology, we extended an existing multi-compartment model of a CA1 pyramidal neuron (Migliore and Migliore, 2012). Excitatory synaptic input was delivered to 120 locations within three dendritic regions (red dots, Fig. 2D), and the average membrane potential in each electrical compartment determined the target level (\tilde{u}_i) in each compartment (Methods). This models how molecular cargo could be selectively trafficked to active synaptic sites. Figures 2E and 2F confirm that the spatial distribution of u approaches the desired steady-state.

Convergence rate

Biochemical processes are time-sensitive. For example, newly synthesized proteins must be delivered to synapses within ~ 1 hour to support long-term potentiation in CA1 pyramidal cells (Frey and Morris, 1997, 1998). We therefore examined how quickly the spatial pattern of cargo converged to its target distribution.

In equations (3) and (4), we implicitly required each $\tilde{u}_i > 0$ in order to avoid division by zero. Intuitively, if $\tilde{u}_i = 0$ for some compartment, then no cargo can flow through that compartment, cutting off more peripheral compartments from the transport system. Similarly, if certain \tilde{u}_i are nearly equal to zero, then transport through these compartments will act as a bottleneck for transport, and convergence to the desired distribution will be slow.

Figure 3A-C illustrates and analyzes a bottleneck in a simple three compartment model. The two compartments on the end of the cable have the same desired level, $\tilde{u}_1 = \tilde{u}_3$; the middle compartment acts as a bottleneck when \tilde{u}_2 is very small (Fig. 3A). This results in the mass-action model:

$$u_1 \xrightleftharpoons[x]{\varepsilon} u_2 \xrightleftharpoons[\varepsilon]{x} u_3 \quad (5)$$

We assume that u begins at one end of the cable, and examine the how taking ε to zero affects the convergence rate. Figure 3B shows that the convergence rate slows dramatically as ε decreases. The convergence rate is determined by the smallest magnitude, non-zero eigenvalue of the system (see Methods), which can be thought of as a rate-limiting step or process for the system. Simulations confirmed this analysis and showed that the convergence rate diverges to infinity as ε approaches zero (Fig. 3C).

We then asked whether the intuition from this three-compartment model extended to a cell

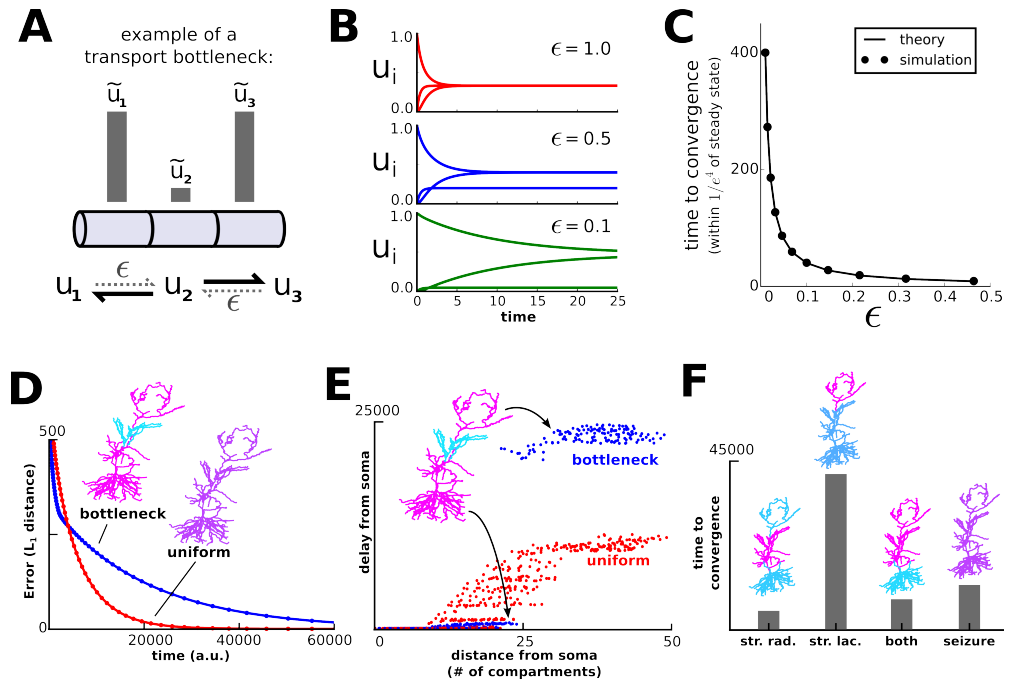


Figure 3. Convergence to steady-state is slow when molecules must be transported across bottlenecks —compartments with low target levels. **(A)** A three-compartment transport model, with the middle compartment acting as a bottleneck. The vertical bars represent the desired steady-state level of u in each compartment. The rate of transport into the middle compartment is small (ϵ , dashed harpoons) relative to transport out of the middle compartment. **(B)** As ϵ decreases, the model converges more slowly and the steady-state level decreases in the middle compartment. **(C)** Simulations (black dots) confirm that the time to convergence is given by the smallest non-zero eigenvalue of the system (analytically calculated line). This eigenvalue can be thought of as the rate-limiting step of the system. **(D)** Convergence (L_1 distance) to a uniform target distribution (red line) is faster than a target distribution containing a bottleneck (blue line) in the CA1 model. **(E)** For all compartments that reach a threshold level ($u_i = 0.001$), the simulated time it takes to reach threshold is plotted against the distance of that compartment to the soma. **(F)** Predicted convergence times for various target distributions (*str. rad.*, stratum radiatum; *str. lac.*, stratum lacunosum/moleculare).

with realistic morphology. We obtained qualitatively similar results. The CA1 model converged to a uniform target distribution more quickly than to a “bottleneck” target distribution, in which the middle third of the apical dendrite had low steady-state levels of cargo (Fig. 3D). Each pair of anterograde and retrograde rate constants were normalized to sum to one; thus, differences in convergence were not due to the scale of the trafficking rate constants.

In addition to this global view of convergence (Fig. 3D), we considered how the transport bottleneck affected transport to individual dendritic compartments. Consider a scenario where transported cargo produces a local chemical reaction after a certain quantity of cargo accumulates;

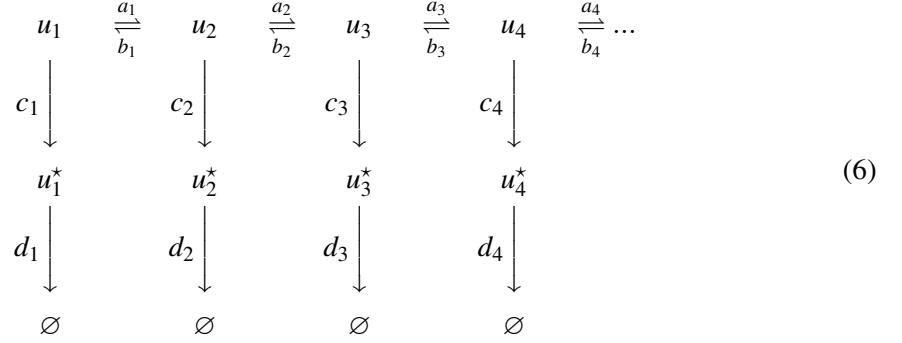
for example, a recently potentiated synapse might be stabilized after enough plasticity-related factors are delivered from the soma. Figure 3E plots the duration of time it took for u_i to reach a pre-defined threshold for each compartment as a measure of the local transport delay. As expected, introducing a bottleneck caused much longer delays to compartments distal to that bottleneck (Fig. 3E, upper right portion of plot). The presence of a bottleneck *shortens* the transport delay to proximal compartments, compared to the uniform target distribution (Fig. 3E, lower left portion of plot). This occurs because cargo delayed by the bottleneck spreads throughout the proximal compartments, reaching higher levels earlier in the simulation. We observed qualitatively similar results for different local threshold values (data not shown).

The model predicts that transport to distal compartments is quickened by increasing the steady-state level of cargo in the proximal compartments (Fig. 3E). This might be experimentally tested by measuring the convergence time of a retrogradely transported molecule that aggregates at recently activated synapses, such as *Arc* mRNA (Steward et al. (1998), see discussion). To illustrate this in the model CA1 cell, we characterized the time course of transport to the distal apical dendrite (stimulating stratum lacunosum/moleculare), proximal apical dendrite (stimulating stratum radiatum), entire apical dendrite (stimulating both layers), and entire cell (seizure). Notably, the model converges more slowly to distal input alone, than to paired distal and proximal input, or to an entirely uniform input distribution (Fig. 3F).

Microtubular trafficking, detachment and degradation on separated time scales

We have so far considered how a target distribution of cargo can be generated by modulating the local rates of molecular motors. However, while certain types of molecular cargo stay on the microtubule network (e.g. mitochondria), many kinds of cargo must detach from the microtubules in order to be used at their final location. For example, dendritic mRNAs are transported along microtubules within densely-packed granules, and are released following granule disassembly (Krichevsky and Kosik, 2001; Buxbaum et al., 2014b). In this case, and in other cases where the cargo is disassembled or sequestered, it is reasonable to model detachment from the microtubule as an irreversible process, followed by an eventual degradation. We thus incorporated these

features into the mass-action model of transport:



As before, a molecule u is transported along a network of microtubules (top row, in equation 7). In each compartment, molecules can irreversibly detach from the microtubules in a reaction $u_i \xrightarrow{c_i} u_i^*$, where u^* denotes the biochemically active or released form of cargo. The final reaction, $u_i^* \xrightarrow{d_i} \emptyset$, models degradation in each compartment. Note that only u^* is subject to degradation; the molecule is assumed to be protected from degradation during transport. This model can be extended to branched morphologies (Fig. 3A).

One way to analyze this system is to assume that these three processes — trafficking, detachment, and degradation — occur on separate timescales. If trafficking is sufficiently faster than detachment ($a, b \gg c$), then u approaches a quasi-steady state distribution defined by our previous analysis (equation 4). We then choose detachment rate constants that transform the microtubular distribution into our desired distribution for u^* :

$$c_i \propto \frac{\tilde{u}_i^*}{\tilde{u}_i} \tag{7}$$

Here, \tilde{u} and \tilde{u}^* respectively denote the quasi-steady state distributions for u and u^* , respectively. As long as degradation is sufficiently slow ($c \gg d$) the desired distribution is transiently achieved.

The addition of spatially varied detachment rates (c_i) produces new strategies for achieving a desired distribution of cargo (Fig. 4B). First, the strategy introduced in figure 2 can be reproduced in this expanded model: the transport rates (a_i, b_i) are chosen according to equation (4) and the detachment rate constants are spatially uniform ($c_i = \text{constant}$, with $c \ll a, b$). In this case, the desired spatial distribution is first achieved both along the microtubules (red line, Fig. 4C) and then for the detached/activated cargo (blue line, Fig. 4C).

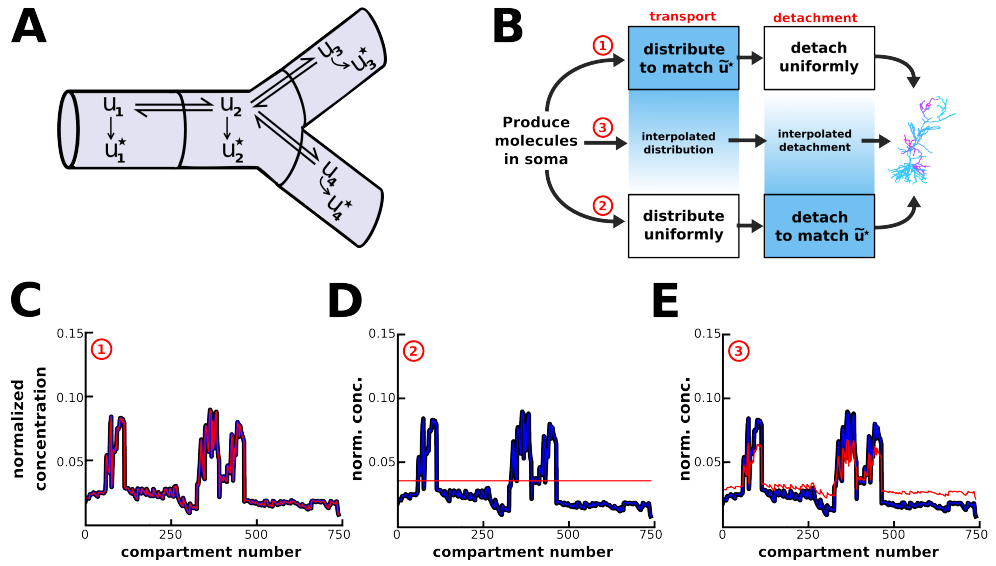


Figure 4. Multiple strategies for transport in a model including nonuniform microtubular detachment/activation. **(A)** Schematic of microtubular transport model with irreversible detachment in a branched morphology. The localized degradation reactions ($u_i^* \xrightarrow{d_i} \emptyset$) is omitted. **(B)** Multiple strategies for producing a desired distribution of activated biomolecules (\tilde{u}^*). When the timescale of detachment/delivery is sufficiently slow, the distribution of cargo on the microtubules approaches a quasi-steady-state (transport step). This known distribution can then be transformed into the desired distribution for \tilde{u} (detachment step). As long as these two steps are appropriately matched (blends of blue), then the desired distribution will be achieved (CA1 cell, right). **(C-E)** Normalized distributions of u (red), u^* (blue), and \tilde{u}^* (black) as $t \rightarrow \infty$ for various strategies diagrammed in panel B (see circled red numbers).

A second strategy begins by choosing uniform transport rates ($a_i = b_i$). This evenly distributes cargo throughout the dendritic tree. The desired distribution is then achieved by locally delivering cargo at a rate proportional to the desired level ($c_i \propto \tilde{u}_i^*$; Fig 4D). Unlike the first solution, this strategy avoids the transport bottlenecks examined in Figure 3, and can achieve target patterns with \tilde{u}^* equal to zero in certain compartments by setting $c_i = 0$.

We refer to the first strategy (Fig. 4C) as the “specific” trafficking model, because cargo is selectively transported to target sites. We refer to the second strategy (Fig. 4D) as the “uniform” trafficking model, because cargo is uniformly distributed throughout the dendrite. These two strategies represent extremes on a spectrum of possible models (Fig. 4B). Figure 4E shows the behavior of an intermediate model, whose parameters are a linear interpolation between the extreme strategies shown in Figure 4C and 4D. Thus, effective trafficking systems can be described as a spectrum of strategies that may be suited to different situations and purposes (see Discussion).

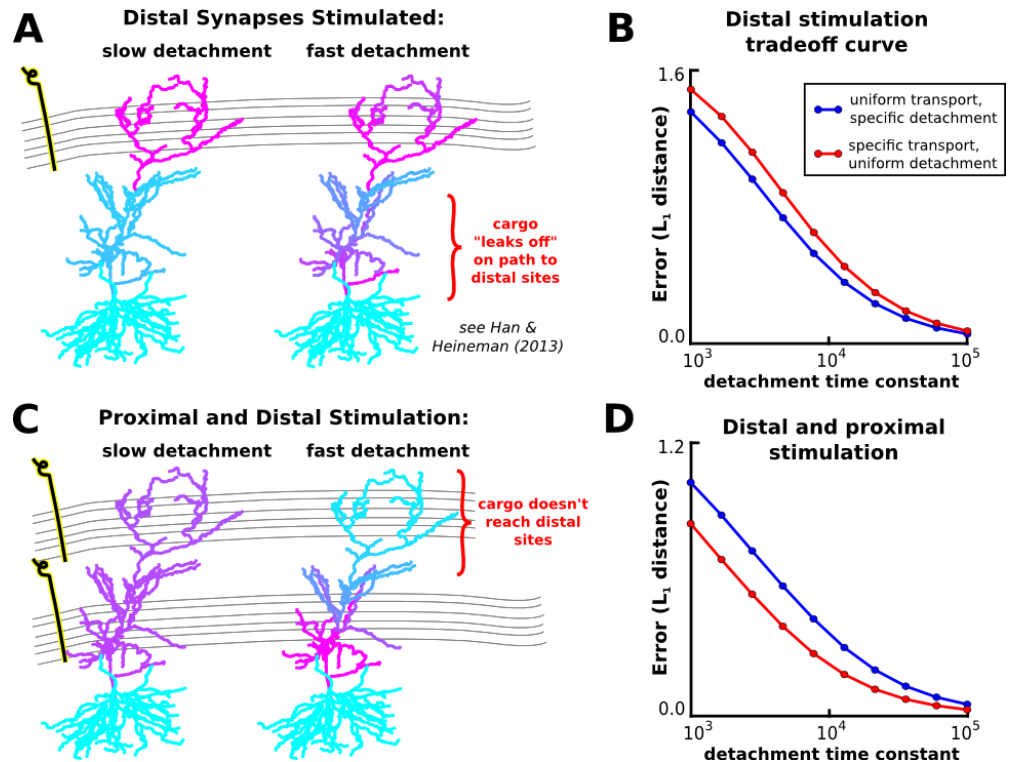


Figure 5. Proximal synapses capture more cargo at the expense of distal synapses when detachment rates are naïvely increased. (A) Delivery of cargo to the distal apical zone with slow (left) and fast detachment rates (right). The steady-state cargo distribution does not match the stimulated pattern when detachment is fast. (B) Tradeoff curves between non-specificity and convergence rate for two trafficking strategies (blue line, see Fig 4D; red line, see Fig 4C). (C-D) Same as (A-B) but for cargo delivery to the entire apical tuft.

Non-specific cargo delivery occurs when trafficking and detachment occur on similar timescales

We have demonstrated possible strategies for neurons to achieve precise and flexible transport of cargo by assuming that cargo detachment is sufficiently slow relative to trafficking. However, biological neurons are unlikely to require perfect matching between demand and distribution of cargo and may therefore tolerate loss of precision in order to speed up delivery globally. We therefore examined the consequences of relaxing the separation of time scales between transport and detachment.

Returning to the model of Figure 4, we consider a scenario where distal synaptic inputs on the apical tuft are stimulated (Fig. 5A). If the average detachment rate constants are sufficiently slow, then, as before, cargo is delivered selectively to the stimulated region (Fig. 5A, left). If we increase the detachment rates by a uniformly scaling, some cargo “leaks off” the microtubule

path on its way to the distal synapses (Fig. 5A, right). We refer to this as a “non-specific” delivery of cargo, since cargo is not selectively delivered to the stimulated sites. Thus, speeding up detachment relative to transport improves the overall rate at which cargoes are delivered to synapses, but this comes at the cost of decreased accuracy of delivery.

Tradeoff curves between the average detachment rate constant and the non-specificity of transport for this stimulation pattern are shown in figure 5B. Importantly, this tradeoff exists for both trafficking strategies we examined in figure 4 — the selective transport strategy (see Fig 4D) and uniform transport strategy (see Fig. 4C). For this stimulation pattern, the uniform trafficking strategy (Fig. 5B, blue line) outperforms the specific trafficking strategy (Fig. 5B, red line) since the latter suffers from bottleneck in the proximal apical zone.

The pattern of non-specific delivery is stimulation-dependent. When the entire apical tree is stimulated, fast detachment can result in a complete occlusion of cargo delivery to distal synaptic sites (Fig. 5C). As before, a tradeoff between specificity and delivery speed is present for both transport strategies (Fig. 5D). Interestingly, the specific trafficking strategy outperforms the uniform trafficking strategy in this case (in contrast to Fig. 5A-B). This is due to the lack of a bottleneck, and the fact that the uniform trafficking strategy initially sends cargo to the basal dendrites where it is not released.

Together, these results show that an increase in the efficiency of synaptic cargo delivery comes at the cost of loss of specificity and that the final destination of mis-trafficked cargo depends on the pattern of stimulation. We next asked whether we can tune the kinetic parameters of the transport mechanism to find an ideal compromise between precision and trafficking efficiency.

Transport can be optimized for both speed and precision for a given stimulation pattern

In figure 5, we showed that scaling the detachment rates (c_i), while leaving the transport rates (a_i , b_i) fixed produces a proximal bias in cargo delivery. That is, synapses closer to the soma are more likely to capture cargo at the expense of distal synapses. We reasoned that increasing the anterograde transport rate of cargo could correct for this bias, producing transport rules that are both fast and precise.

We examined this possibility in a simpler model — an unbranched cable — so that we could

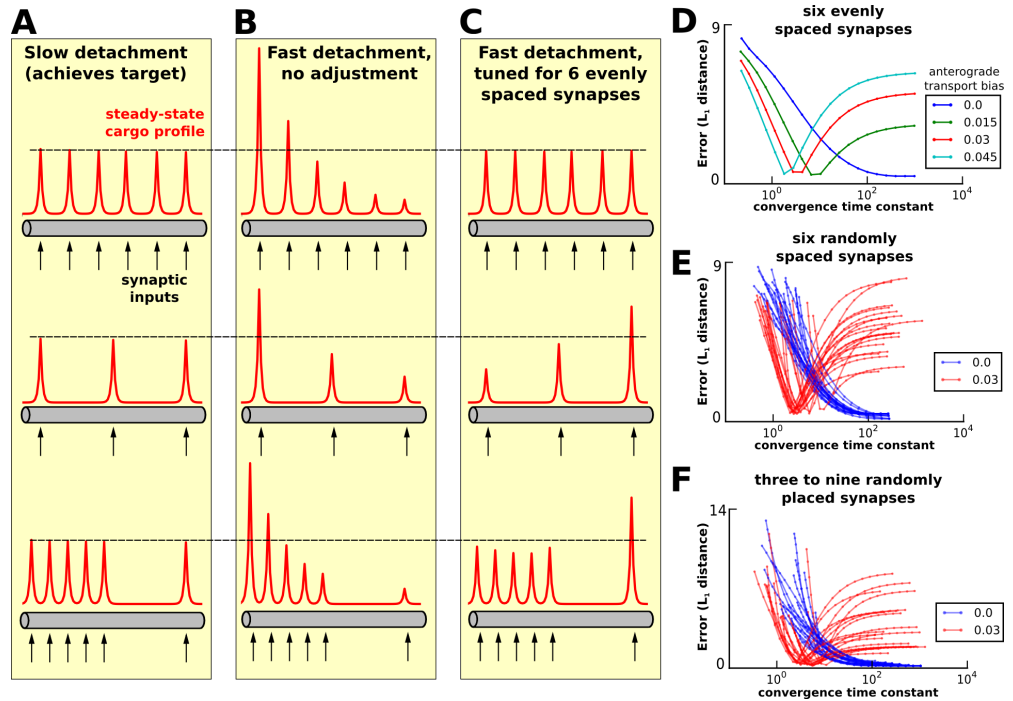


Figure 6. Producing speed and specificity for particular stimulation patterns does not extend to other patterns. Cargo begins on the left end and is transported to the right. **(A-C)** Steady-state cargo profiles (red) for three stimulation patterns (black arrows) along an unbranched cable. The dotted black line corresponds to the ‘target’ steady-state level at each synapse. **(A)** When the timescale of detachment is sufficiently slow, cargo can be evenly distributed to the synapses regardless of their number and position. Transport parameters were set according to the procedure shown in figure 4D. **(B)** When detachment is naïvely increased (all rates multiplicatively scaled) a proximal bias in the steady-state distribution of cargo across all stimulation patterns. **(C)** Transport rate constants, a_i and b_i , were tuned to optimize the distribution of cargo to six equally spaced synapses (top row); detachment rate constants were the same as in panel B. Changing the number of synapses (middle row) or the position of the synapses (bottom row) causes the unequal distribution of cargo to synapses. **(D)** Tradeoff curves between non-specificity and convergence rate for six evenly spaced synapses (top row of A-C). The curve shifts to the left and becomes non-monotonic as the anterograde bias (β) increases. **(E)** Tradeoff curves for two levels of anterograde bias on multiple simulations; for each simulation six synapse positions were drawn from random uniform distribution over the length of the cable. **(F)** Same as panel E, except each the number of synapses was randomly chosen between 3 to 9 for each simulation.

develop simple heuristics that precisely achieve a desired distribution of cargo while minimizing the convergence time. In the reduced model, cargo begins on the left end of the cable and is transported to a number of synaptic sites, each of which is modeled as a double-exponential curve.

As before, cargo can be precisely delivered to a variety of stimulation patterns when the

detachment rate is sufficiently slow (Fig. 6A); when the detachment rate is naïvely increased to speed up the rate of delivery, a proximal bias develops for all stimulation patterns (Fig. 6B). This bias is qualitatively exponential.

We then hand-tuned the transport rate constants to deliver equal cargo to six evenly spaced synaptic sites (top row of Fig. 6C). Specifically, we increased the anterograde rate constants (a_i) and decreased the retrograde rate constants (b_i) by a decreasing linear function of position so that $a_{N-1} = b_{N-1}$ at the right side of the cable. On the left end of the cable, we set $a_1 = 0.5 + \beta$ and $b_1 = 0.5 - \beta$, where β is the parameter controlling anterograde bias. Intuitively, this pushes cargo past the proximal synaptic sites more quickly, canceling out the exponential delivery bias induced by fast detachment.

However, this tuned transport model does not precisely deliver cargo for other stimulation patterns. When the number of synapses on the cable is decreased, a distal delivery bias is observed because too little cargo is released on the proximal portion of the cable (middle row, Fig. 6C). Even when the number of synapses is held constant, changing the position of the synapses can disrupt equitable delivery of cargo to synapses. This is shown in the bottom row of figure 6C, where a distal bias again develops when the majority of activated synapses are positioned proximally. Thus, within the simple framework we've developed, the delivery of cargo can be tuned to achieve both precision and speed for a specified target distribution. However, non-specific cargo delivery occurs when different stimulation patterns are applied (assuming the transport parameters are not re-tuned).

To examine these observations over a larger range of stimulation patterns and transport parameters, we plotted tradeoff curves between delivery precision and speed. We first examined a cable with six evenly spaced delivery sites (same as top row of Fig. 6A-C). As before, a hard tradeoff between specificity and delivery speed exists for the untuned model (blue line in Fig. 6D, same transport rate constants as Fig. 6A and 6B). As the anterograde transport bias is introduced and increased, the optimal points along the tradeoff curve move to the left, representing faster transport times. The tradeoff curves also become nonmonotonic: the error (non-specificity) initially decreases as the detachment rate decreases, but begins to increase after a certain well-matched point. Points on the descending left branch of the curve represent cargo distributions with proximal bias (detachment is too fast); points on the ascending right branch correspond to

distal bias (detachment is too slow).

We observed qualitatively similar tradeoff curves for randomly positioned synapses (Fig. 6E), and for randomizing the number and position of synapses (Fig. 6F). There is substantial variability in the tradeoff curves in both cases, but particularly when randomizing the number of synaptic inputs (Fig. 6F). Notably, the untuned transport model (blue curves) always converge to zero error as the detachment rate decreases. Thus, for this model, it appears that the only way to achieve very precise, reliable and flexible transport is to have a slow detachment rate.

DISCUSSION

A microscope image of a typical neuronal dendritic tree hints at the complex logistical task that the neuron must solve to maintain synapses and alter them during development and learning. Synapses, such as the extensively-studied excitatory synapses of the mammalian nervous system, comprise hundreds of different proteins, including receptors, structural and anchoring proteins, and cytosolic signaling enzymes (Liu et al., 2014; Laßek et al., 2015). How do these components find their way to the synapse, and how do synapses that have varying demands for these components receive an appropriate supply?

A large experimental literature addresses the molecular mechanisms underlying this question, and several reviews have outlined a two-step model in which molecular cargo first samples the dendritic arbor by a noisy walk, before its eventual release or capture (Welte, 2004; Doyle and Kiebler, 2011; Buxbaum et al., 2014a, 2015). Both stages of transport are thought to be influenced by localized signaling pathways, rather than being pre-determined by a precise molecular address. The signaling pathways affecting transport require further characterization and discovery; candidate second-messengers include calcium (Wang and Schwarz, 2009), ADP (Mironov, 2007), and microtubule-associated proteins (Soundararajan and Bullock, 2014).

Our goal was to formulate conceptual accounts of activity-dependent transport as mathematical models to test their feasibility and examine relationships between the speed, flexibility and specificity of cargo delivery. A primary question we asked was whether a transport system based on purely local rules was feasible, or whether a complex global addressing system was required. We showed that local rules can produce arbitrarily precise and flexible cargo delivery provided that the release of cargo occurs on a slow enough timescale. Intuitively, this requirement means that the cargo has time to sample the dendritic tree for potential delivery sites (Welte,

2004). Similar decentralized strategies are utilized by other biological systems, such as ant colonies, which converge onto a common path during exploration based on localized trail-laying and -following decisions made by a distributed population of agents (Deneubourg et al., 1990). Such self-organizing systems often enjoy a number of advantageous properties – including scalability, flexibility, and fault-tolerance – and are therefore a current topic of substantial interest to engineers (Werfel et al., 2014; Rubenstein et al., 2014).

Cargo does not need to search the dendritic tree for target sites in a uniform or unbiased manner. We showed that the same distribution of cargo could be achieved by (a) specific microtubular transport followed by uniform release, (b) from uniform transport followed by specific release, or (c) a combination of these strategies (Fig. 4). Experimental findings are variable across systems and span these possibilities. For example, Kim and Martin (2015) identified three mRNAs that were uniformly distributed throughout cultured *Aplysia* sensory neurons, but were targeted to synapses at the level of protein expression by localized translation. In contrast, the nonuniform expression of *Arc* mRNA is closely matched to the pattern of Arc protein in granule cells of the dentate gyrus (Steward et al., 1998; Farris et al., 2014; Steward et al., 2015). Even the same type of molecular cargo can show diverse movement statistics in single-particle tracking experiments (Dynes and Steward, 2007) Others?.

Given that different strategies are feasible and appear to be represented in biological neurons, it is important to know the pros and cons of a given strategy, and the general tradeoffs that exist in a transport mechanism that uses local signals. One experimentally testable phenomenon seen in the model is the emergence of bottlenecks that arise when cargo is trafficked nonuniformly; for example, when distal regions require high levels of cargo and proximal regions require low levels (Fig. 3). Other forms of trafficking bottlenecks can be experimentally induced by expressing kinesin at high densities to induce molecular crowding Leduc et al. (2012). An important open question is whether trafficking bottlenecks and crowding events occur as part of normal physiology, and whether severe activity-induced bottlenecks might underlie pathophysiological states.

We showed that having sufficiently slow capture of cargo can produce a well-behaved transport system that matches demand using local mechanisms and is not sensitive to the complex geometry of the neuron. Mathematically, this regime occurs when the system contains

two separated timescales, so that the distribution of cargo along the microtubules approaches a quasi-steady-state that matches the long-term average of local demand. However, because the transport of certain cargo is time-sensitive (Frey and Morris, 1997, 1998), we asked whether the model could maintain precise delivery while relaxing the restriction of slow detachment. We found that increasing the rate of detachment produces a proximal bias in the delivery of cargo due to spontaneous leakage, leading to a mismatch between supply and demand across a population of synapses. Thus, the family of models that corresponds to a large separation of timescales between transport and detachment face a tradeoff between the speed and specificity of cargo delivery.

We then asked whether a different family of models might circumvent this tradeoff by globally tuning the anterograde and retrograde movement rates. Studies show that a directional bias in transport can be induced by changing the complement of motor proteins (Kanai et al., 2004; Amrute-Nayak and Bullock, 2012), and that such a change might be induced at the soma in response to synaptic activity (Puthanveettil et al., 2008). In a simple case involving an unbranched cable we showed that increasing the anterograde transport rate can precisely overcome the proximal bias in cargo delivery, and therefore provide both fast and precise transport for a particular target distribution of cargo (Fig. 6).

However, tuning trafficking bias to counterbalance leakage does not provide a completely general solution to the speed/specificity tradeoff in a biologically realistic situation. First, anterograde rate constants need to be tuned non-uniformly over the cable. In the cases we studied, we used a linearly decreasing profile (see *Methods*). How might such a non-uniform transport bias arise at the molecular level? Simply changing the composition of motor proteins (Amrute-Nayak and Bullock, 2012) is unlikely induce a spatial profile in transport bias. On the other hand, non-uniform modulation of the microtubule network or expression of microtubule-associated proteins provide potential mechanisms (Kwan et al., 2008; Soundararajan and Bullock, 2014). It would therefore be intriguing to experimentally test the existence of spatial gradients in anterograde movement bias, for example, using single-particle tracking in living neurons.

Secondly, tuning transport bias can provide fast and rapid cargo delivery for a specific density and arrangement of synapses along a dendrite, but this tuned solution will not be optimal for different densities and spatial distributions of synapses, as seen in Figure 6. The overall

morphology of the neurite may also need to be taken into account, as introducing an anterograde bias can cause the accumulation of cargo at the tips of short branches, proximal to the soma (data not shown). However, other mechanisms, not modeled in this study, might prevent this accumulation (Soundararajan and Bullock, 2014). Certain neuron types in specific brain regions may nonetheless have morphology and synaptic connectivity that is sufficiently stereotyped to permit some degree of tuning. In such cases the qualitative prediction that anterograde bias should decrease as a function of distance to the soma can be tested. Such tuning, while providing improved efficiency in trafficking intracellular cargo, would also constitute a site of vulnerability to perturbations that alter synapse density and reflect an additional constraint on the possibilities for wiring and re-wiring neural circuits.

METHODS

All simulation code is available on: <https://github.com/ahwillia/Williams-et-al-Synaptic-Transport>

Model of single-particle transport

Let x_n denote the position of a particle along a 1-dimensional cable at timestep n . Let v_n denote the velocity of the particle at timestep n ; for simplicity, we assume the velocity can take on three discrete values, $v_n = \{-1, 0, 1\}$, corresponding to a discrete retrograde movement, pause, or anterograde movement. As a result, x_n is constrained to take on integer values. In the memoryless transport model (top plots in Fig. 1B, 1D, and 1F), we assume that v_n is drawn with fixed probabilities on each step. The update rule for position is:

$$x_{n+1} = x_n + v_n$$

$$v_{n+1} = \begin{cases} -1 & \text{with probability } p_- \\ 0 & \text{with probability } p_0 \\ 1 & \text{with probability } p_+ \end{cases}$$

We chose $p_- = 0.2$, $p_0 = 0.35$ and $p_+ = 0.45$ for the illustration shown in Figure 1. For the model with history-dependence (bottom plots in Fig. 1B, 1D, and 1F), the movement probabilities at each step depend on the previous movement. For example, if the motor was moving in an anterograde direction on the previous timestep, then it is more likely to continue to moving in

that direction in the next time step. In this case the update rule utilizes *conditional* probabilities:

$$v_{n+1} = \begin{cases} -1 & \text{with probability } p(-|v_n) \\ 0 & \text{with probability } p(0|v_n) \\ 1 & \text{with probability } p(+|v_n) \end{cases}$$

In the extreme case of history-dependence, the particle always steps in the same direction as the previous time step.

	$v_n = -1$	$v_n = 0$	$v_n = 1$
$p(v_{n+1} = -1)$	1	0	0
$p(v_{n+1} = 0)$	0	1	0
$p(v_{n+1} = 1)$	0	0	1

We introduce a parameter $k \in [0, 1]$ to linearly interpolate between this extreme case and the memoryless model. The rest of the panels in the figure show individual runs, 5000 molecule simulations, and mass action approximations for $k = 0.5$.

Relationship of single-particle transport to the mass-action model

The mass-action model (equation 1, in the *Results*) simulates the bulk movement of cargo u across discrete compartments. The rate of cargo transfer is modeled as elementary chemical reactions (Keener and Sneyd, 1998). For an unbranched cable, the change in cargo in compartment i is given by:

$$\dot{u}_i = au_{i-1} + bu_{i+1} - (a+b)u_i \quad (8)$$

For now, we assume that the anterograde (a) and retrograde (b) trafficking rate constants are spatially uniform.

The mass-action model approximates the drift-diffusion partial differential equation (Fig. 1E) by discretizing u into spatial compartments. This can be seen by Taylor expanding around some position $u(x)$:

$$\begin{aligned} \dot{u}(x) &\approx a \left[u(x) - \Delta \frac{\partial u}{\partial x} + \frac{\Delta^2}{2} \frac{\partial^2 u}{\partial x^2} \right] + b \left[u(x) + \Delta \frac{\partial u}{\partial x} + \frac{\Delta^2}{2} \frac{\partial^2 u}{\partial x^2} \right] - (a+b) u(x) \\ &= a \left[-\Delta \frac{\partial u}{\partial x} + \frac{\Delta^2}{2} \frac{\partial^2 u}{\partial x^2} \right] + b \left[\Delta \frac{\partial u}{\partial x} + \frac{\Delta^2}{2} \frac{\partial^2 u}{\partial x^2} \right] \end{aligned}$$

where Δ is the compartment size. In the limit $\Delta \rightarrow 0$, we arrive precisely at the drift-diffusion equation.

$$\dot{u}(x) = \frac{\partial u}{\partial t} = \underbrace{(b-a)}_{\text{drift coefficient}} \frac{\partial u}{\partial x} + \underbrace{\left(\frac{a+b}{2}\right)}_{\text{diffusion coefficient}} \frac{\partial^2 u}{\partial x^2}$$

When all cargo starts in one location (delta function), then the distribution after some elapsed time will be a Gaussian function. The mean of this Gaussian function changes in direct proportion to the drift coefficient over time, while the variance is inversely proportional to the diffusion coefficient (CITE?).

How does this equation relate to the model of single-particle transport (Fig. 1A-B)? For a memoryless biased random walk, the expected position of a particle after n time steps is $E[x_n] = n(p_+ - p_-)$ and the variance in position after n steps is $n(p_+ + p_- - (p_+ - p_-)^2)$. For large numbers of independently moving particles (assuming *ergodicity*), the ensemble of cargo will approach a Gaussian distribution by the Central Limit Theorem. This means that the mean and variance calculations for a single particle can be directly related to the ensemble statistics outlined in the previous paragraph. We find:

$$a = \frac{2p_+ - (p_+ - p_-)^2}{2}$$

$$b = \frac{2p_- - (p_+ - p_-)^2}{2}$$

Can we say the following: For large numbers of particles and long times, the central limit theorem still holds when there are long runs in single-particle trajectories. $E[x_n] = n(p_+ - p_-)$ is still the expected value for mean, but the variance is different. However, the mass-action family of models still captures the ensemble behavior. This gives some intuition as to why we aren't too sensitive to the assumption of short run lengths, though I it breaks down when run lengths are very long (because then the system isn't very stochastic).

Model of bulk/mass-action transport

Many of our results take advantage of the fact that the steady-state ratio of cargo in two neighboring compartments equals the ratio of trafficking rate constants (equation 2). We prove this result for a cable with non-uniform anterograde (a_i) and retrograde (b_i) rate constants.

Examining the left end of the cable at steady-state ($\dot{u}_i = 0$), we immediately verify our result:

$$-a_1 u_1 + b_1 u_2 = 0 \Rightarrow \left. \frac{u_1}{u_2} \right|_{ss} = \frac{b_1}{a_1}$$

Successively applying the same logic down the cable confirms the condition in equation 2 holds globally. A similar procedure (starting at the tips and moving in) can be applied to branched morphologies to prove the more general condition (equation 4).

It is helpful to re-express the model as a matrix differential equation, $\dot{\mathbf{u}} = A\mathbf{u}$, where $\mathbf{u} = [u_1, u_2, \dots, u_N]^T$ is the state vector, and A is the state-transition matrix. For a general branched morphology, A will be nearly tridiagonal, with off-diagonal elements corresponding to branch points; matrices in this form are called Hines matrices (Hines, 1984). For the simpler case of an unbranched cable, A is tridiagonal:

$$A = \begin{bmatrix} -a_1 & b_1 & 0 & & \dots & 0 \\ a_1 & -b_1 - a_2 & b_2 & 0 & & \\ 0 & a_2 & -b_2 - a_3 & b_3 & \ddots & \vdots \\ \vdots & 0 & a_3 & \ddots & & 0 \\ & & \ddots & & -b_{N-2} - a_{N-1} & b_{N-1} \\ 0 & & \dots & 0 & a_{N-1} & -b_{N-1} \end{bmatrix}$$

For both branched and unbranched morphologies, each column of A sums to zero, which reflects conservation of mass within the system. The rank of A is exactly $N - 1$ (this can be seen by taking the sum of the first $N - 1$ rows, which results in -1 times the final row). Thus, the nullspace of A is one-dimensional (red lines in Supp. Fig. 1).

The desired steady-state distribution, $\tilde{\mathbf{u}}$, is an eigenvector that spans the nullspace of A . It is simple to show that all other eigenvalues A are negative using the Gershgorin circle theorem; thus, the fixed point described by equations 2 and 4 is stable. The convergence rate is determined by the non-zero eigenvalue with smallest magnitude of A .

Stochastic Interpretation of Mass-Action Model

The mass-action model described in figure 2 is a deterministic dynamical system. However, this determinism only holds in the limit of having a very large number of transported particles.

Cargoes with small copy numbers (e.g. mitochondria) will operate in a stochastic regime. We evoke a standard assumption that the system is *ergodic*, meaning that position of one particle averaged over very long time intervals is the same as the ensemble steady-state distribution.

Thus, in addition to modeling how the spatial distribution of cargo changes over time, the mass-action model equivalently models the spatial probability distribution of a single cargo. That is, imagine we track a single cargo and ask its position after a long period of transport. The probability ratio between of finding this particle in any parent-child pair of compartments converges to:

$$\left. \frac{p_p}{p_c} \right|_{ss} = \frac{b}{a}$$

which mirrors our analysis of the deterministic model.

At steady-state, the number of molecules in any single compartment follows a binomial distribution; the coefficient of variation in each compartment is given by:

$$\sqrt{\frac{1 - p_i^{(ss)}}{n p_i^{(ss)}}}$$

This suggests that there are two ways of decreasing noise in the system. First, increasing the number of transported molecules, n , decreases the noise by a factor of $1/\sqrt{n}$. Additionally, transport is more reliable to compartments with a high steady-state occupation probability.

Detachment and Degradation Reactions and Quasi-Steady-State Approximation

Introducing detachment and degradation reactions into the transport model is straightforward. For an arbitrary compartment in a cable, the differential equations become:

$$\begin{aligned}\dot{u}_i &= a_{i-1}u_{i-1} - (a_i + b_{i-1} + c_i)u_i + b_i u_{i+1} \\ \dot{u}_i^* &= c_i u_i - d_i u_i^*\end{aligned}$$

When $a_i, b_i \gg c_i \gg d_i$, then the variables u_i and u_i^* approach a quasi-steady-state, which we denote \tilde{u}_i and \tilde{u}_i^* . For simplicity we assume $d_i = 0$ in our simulations. We present two strategies for achieving a desired distribution for \tilde{u}_i^* in figure 4C and 4D. To interpolate between these strategies, let F be a scalar between 0 and 1, and let \tilde{u}^* be normalized to sum to one. We choose

a_i and b_i to achieve:

$$\tilde{u}_i = F \tilde{u}_i^* + (1 - F)/N$$

along the microtubular network and choose c_i to satisfy

$$c_i \propto \frac{\tilde{u}_i^*}{F \tilde{u}_i^* + (1 - F)/N}$$

Here, N is the number of compartments in the model. Setting $F = 1$ results in the simulation in figure 4C, while setting $F = 0$ results in the simulation in figure 4D. An interpolated strategy is shown in figure 4E ($F = 0.3$).

Globally tuning transport rates to circumvent the speed-specificity tradeoff

to be written soon.

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REFERENCES

- Aakalu, G., Smith, W. B., Nguyen, N., Jiang, C., and Schuman, E. M. (2001). Dynamic visualization of local protein synthesis in hippocampal neurons. *Neuron*, 30(2):489–502.
- Amrute-Nayak, M. and Bullock, S. L. (2012). Single-molecule assays reveal that rna localization signals regulate dynein-dynactin copy number on individual transcript cargoes. *Nat. Cell Biol.*, 14(4):416–423.
- Andreassi, C. and Riccio, A. (2009). To localize or not to localize: mrna fate is in 3 utr ends. *Trends Cell Biol.*, 19(9):465–474.
- Block, S. M., Goldstein, L. S. B., and Schnapp, B. J. (1990). Bead movement by single kinesin molecules studied with optical tweezers. *Nature*, 348(6299):348–352.
- Bressloff, P. and Newby, J. (2009). Directed intermittent search for hidden targets. *New J. Phys.*, 11(2):023033.
- Bressloff, P. C. (2006). Stochastic model of protein receptor trafficking prior to synaptogenesis. *Phys. Rev. E Stat. Nonlin. Soft. Matter. Phys.*, 74(3).
- Bressloff, P. C. and Newby, J. M. (2013). Stochastic models of intracellular transport. *Rev. Mod. Phys.*, 85(1):135–196.

- Buxbaum, A. R., Haimovich, G., and Singer, R. H. (2014a). In the right place at the right time: visualizing and understanding mRNA localization. *Nat. Rev. Mol. Cell Biol.*, 16(2):95–109.
- Buxbaum, A. R., Wu, B., and Singer, R. H. (2014b). Single-Actin mRNA Detection in Neurons Reveals a Mechanism for Regulating Its Translatability. *Science*, 343(6169):419–422.
- Buxbaum, A. R., Yoon, Y. J., Singer, R. H., and Park, H. Y. (2015). Single-molecule insights into mrna dynamics in neurons. *Trends Cell Biol.*, 25(8):468–475.
- Deneubourg, J.-L., Aron, S., Goss, S., and Pasteels, J. M. (1990). The self-organizing exploratory pattern of the argentine ant. *J. Insect Behav.*, 3(2):159–168.
- Doyle, M. and Kiebler, M. A. (2011). Mechanisms of dendritic mRNA transport and its role in synaptic tagging. *EMBO J.*, 30(17):3540–3552.
- Dynes, J. L. and Steward, O. (2007). Dynamics of bidirectional transport of *Arc* mRNA in neuronal dendrites. *J. Comp. Neurol.*, 500(3):433–447.
- Farris, S., Lewandowski, G., Cox, C. D., and Steward, O. (2014). Selective Localization of *Arc* mRNA in Dendrites Involves Activity- and Translation-Dependent mRNA Degradation. *J. Neurosci.*, 34(13):4481–4493.
- Frey, U. and Morris, R. (1998). Weak before strong: dissociating synaptic tagging and plasticity-factor accounts of late-LTP. *Neuropharmacol.*, 37(4-5):545–552.
- Frey, U. and Morris, R. G. M. (1997). Synaptic tagging and long-term potentiation. *Nature*, 385(6616):533–536.
- Gagnon, J. and Mowry, K. (2011). Molecular motors: directing traffic during RNA localization. *Crit Rev Biochem Mol Biol*, 46:229–39.
- Hancock, W. O. (2014). Bidirectional cargo transport: moving beyond tug of war. *Nat. Rev. Mol. Cell Biol.*, 15(9):615–628.
- Hines, M. (1984). Efficient computation of branched nerve equations. *Int. J. Biomed. Comput.*, 15(1):69–76.
- Hirokawa, N., Niwa, S., and Tanaka, Y. (2010). Molecular Motors in Neurons: Transport Mechanisms and Roles in Brain Function Development, and Disease. *Neuron*, 68(4):610–638.
- Hoffman, D., Magee, J., Colbert, C., and Johnston, D. (1997). K⁺ channel regulation of signal propagation in dendrites of hippocampal pyramidal neurons. *Nature*, 387:869–75.
- Kanai, Y., Dohmae, N., and Hirokawa, N. (2004). Kinesin transports rna: isolation and charac-

- terization of an rna-transporting granule. *Neuron*, 43(4):513–525.
- Kandel, E. R. (2001). The Molecular Biology of Memory Storage: A Dialogue Between Genes and Synapses. *Science*, 294(5544):1030–1038.
- Kang, H. and Schuman, E. M. (1996). A requirement for local protein synthesis in neurotrophin-induced hippocampal synaptic plasticity. *Science*, 273(5280):1402–1406.
- Keener, J. P. and Sneyd, J. (1998). *Mathematical Physiology*. Springer, New York.
- Kim, S. and Martin, K. C. (2015). Neuron-wide rna transport combines with netrin-mediated local translation to spatially regulate the synaptic proteome. *eLife*, 4:e04158.
- Krichevsky, A. M. and Kosik, K. S. (2001). Neuronal RNA Granules A Link between RNA Localization and Stimulation-Dependent Translation. *Neuron*, 32(4):683–696.
- Kwan, A. C., Dombeck, D. A., and Webb, W. W. (2008). Polarized microtubule arrays in apical dendrites and axons. *Proc. Natl. Acad. Sci. U.S.A.*, 105(32):11370–11375.
- Laßek, M., Weingarten, J., and Volkandt, W. (2015). The synaptic proteome. *Cell Tissue Res.*, 359(1):255–265.
- Leduc, C., Padberg-Gehle, K., Varga, V., Helbing, D., Diez, S., and Howard, J. (2012). Molecular crowding creates traffic jams of kinesin motors on microtubules. *Proc. Natl. Acad. Sci. U.S.A.*, 109(16):6100–6105.
- Liu, X.-A., Kadakkuzha, B., Pascal, B., Steckler, C., Akhmedov, K., Yan, L., Chalmers, M., and Puthanveetil, S. V. (2014). New approach to capture and characterize synaptic proteome. *Proc. Natl. Acad. Sci. U.S.A.*, 111(45):16154–16159.
- Lyles, V., Zhao, Y., and Martin, K. C. (2006). Synapse Formation and mRNA Localization in Cultured Aplysia Neurons. *Neuron*, 49(3):349–356.
- Magee, J. (1998). Dendritic hyperpolarization-activated currents modify the integrative properties of hippocampal CA1 pyramidal neurons. *J Neurosci*, 18:7613–24.
- Migliore, M. and Migliore, R. (2012). Know your current i_h : Interaction with a shunting current explains the puzzling effects of its pharmacological or pathological modulations. *PLoS ONE*, 7(5):e36867.
- Mironov, S. L. (2007). ADP Regulates Movements of Mitochondria in Neurons. *Biophys. J.*, 92(8):2944–2952.
- Moga, D., Calhoun, M., Chowdhury, A., Worley, P., Morrison, J., and Shapiro, M. (2004).

- Activity-regulated cytoskeletal-associated protein is localized to recently activated excitatory synapses. *Neurosci.*, 125(1):7–11.
- Muller, M. J. I., Klumpp, S., and Lipowsky, R. (2008). Tug-of-war as a cooperative mechanism for bidirectional cargo transport by molecular motors. *Proc. Natl. Acad. Sci. U.S.A.*, 105(12):4609–4614.
- Newby, J. and Bressloff, P. C. (2010). Local synaptic signaling enhances the stochastic transport of motor-driven cargo in neurons. *Phys. Biol.*, 7(3):036004.
- Park, H. and Poo, M.-m. (2013). Neurotrophin regulation of neural circuit development and function. *Nat. Rev. Neurosci.*, 14(1):7–23.
- Park, H. Y., Lim, H., Yoon, Y. J., Follenzi, A., Nwokafor, C., Lopez-Jones, M., Meng, X., and Singer, R. H. (2014). Visualization of dynamics of single endogenous mrna labeled in live mouse. *Science*, 343(6169):422–424.
- Parrish, J. Z., Emoto, K., Kim, M. D., and Jan, Y. N. (2007). Mechanisms that regulate establishment, maintenance, and remodeling of dendritic fields. *Annu. Rev. Neurosci.*, 30:399–423.
- Puthanveetil, S. V., Monje, F. J., Miniaci, M. C., Choi, Y.-B., Karl, K. A., Khandros, E., Gawinowicz, M. A., Sheetz, M. P., and Kandel, E. R. (2008). A New Component in Synaptic Plasticity: Upregulation of Kinesin in the Neurons of the Gill-Withdrawal Reflex. *Cell*, 135(5):960–973.
- Rubenstein, M., Cornejo, A., and Nagpal, R. (2014). Programmable self-assembly in a thousand-robot swarm. *Science*, 345(6198):795–799.
- Smith, D. and Simmons, R. (2001). Models of Motor-Assisted Transport of Intracellular Particles. *Biophys. J.*, 80(1):45–68.
- Soundararajan, H. C. and Bullock, S. L. (2014). The influence of dynein processivity control, maps, and microtubule ends on directional movement of a localising mrna. *eLife*, 3:e01596.
- Spitzer, N. C. (2006). Electrical activity in early neuronal development. *Nature*, 444(7120):707–712.
- Steward, O., Farris, S., Pirbhoy, P. S., Darnell, J., and Driesche, S. J. V. (2015). Localization and local translation of Arc/Arg3.1 mRNA at synapses: some observations and paradoxes. *Front. Mol. Neurosci.*, 7.

- Steward, O., Wallace, C., Lyford, G., and Worley, P. (1998). Synaptic Activation Causes the mRNA for the IEG *Arc* to Localize Selectively near Activated Postsynaptic Sites on Dendrites. *Neuron*, 21(4):741–751.
- Steward, O. and Worley, P. F. (2001). Selective Targeting of Newly Synthesized *Arc* mRNA to Active Synapses Requires NMDA Receptor Activation. *Neuron*, 30(1):227–240.
- Sutton, M. A. and Schuman, E. M. (2006). Dendritic protein synthesis, synaptic plasticity, and memory. *Cell*, 127(1):49–58
- Turrigiano, G. (2011). Too many cooks? intrinsic and synaptic homeostatic mechanisms in cortical circuit refinement. *Annual review of neuroscience*, 34:89–103.
- Verbrugge, S., van den Wildenberg, S. M., and Peterman, E. J. (2009). Novel Ways to Determine Kinesin-1's Run Length and Randomness Using Fluorescence Microscopy. *Biophys. J.*, 97(8):2287–2294.
- Vickers, C. A., Dickson, K. S., and Wyllie, D. J. A. (2005). Induction and maintenance of late-phase long-term potentiation in isolated dendrites of rat hippocampal cal pyramidal neurones. *J. Physiol.*, 568(3):803–813.
- Wang, X. and Schwarz, T. L. (2009). The Mechanism of Ca^{2+} -Dependent Regulation of Kinesin-Mediated Mitochondrial Motility. *Cell*, 136(1):163–174.
- Welte, M. A. (2004). Bidirectional transport along microtubules. *Curr. Biol.*, 14(13):R525–R537.
- Werfel, J., Petersen, K., and Nagpal, R. (2014). Designing collective behavior in a termite-inspired robot construction team. *Science*, 343(6172):754–758.
- Wong, R. O. and Ghosh, A. (2002). Activity-dependent regulation of dendritic growth and patterning. *Nat. Rev. Neurosci.*, 3(10):803–812.
- Zagrebelsky, M. and Korte, M. (2014). Form follows function: Bdnf and its involvement in sculpting the function and structure of synapses. *Neuropharmacol.*, 76:628–638.