Spontaneous generation of persistent activity in diffusively coupled cellular assemblies

Ria Ghosh^{1,2} and Shakti N. Menon¹

¹ The Institute of Mathematical Sciences, CIT Campus, Taramani, Chennai 600113, India ² Homi Bhabha National Institute, Anushaktinagar, Mumbai 400 094, India (Dated: September 27, 2021)

The spontaneous generation of electrical activity underpins a number of essential physiological processes, and is observed even in tissues where specialized pacemaker cells have not been identified. The emergence of periodic oscillations in diffusively coupled assemblies of excitable and electrically passive cells (which are individually incapable of sustaining autonomous activity) has been suggested as a possible mechanism underlying such phenomena. In this paper we investigate the dynamics of such assemblies in more detail by considering simple motifs of coupled electrically active and passive cells. The resulting behavior encompasses a wide range of dynamical phenomena, including chaos. However, embedding such assemblies in a lattice yields spatio-temporal patterns that either correspond to a quiescent state or partial/globally synchronized oscillations. The resulting reduction in dynamical complexity suggests an emergent simplicity in the collective dynamics of such large, spatially extended systems. Furthermore, we show that such patterns can be reproduced by a reduced model comprising only excitatory and oscillatory elements. Our results suggest a generalization of the mechanism by which periodic activity can emerge in a heterogeneous system comprising non-oscillatory elements by coupling them diffusively, provided their steady states in isolation are sufficiently dissimilar.

Spontaneously recurring electrical activity is of crucial significance in a number of physiological contexts [1–3]. This is typically driven by pacemaker cells [4–6], such as the sinoatrial node in the heart which comprises specialized cells that periodically generate signals initiating excitatory activity, leading to mechanical contraction [7]. However, such cells have not been observed in other contractile tissue, such as the myometrium of the gravid uterus [8]. It has been hypothesized that spontaneous activity in the latter contexts arise through interactions between electrically active and passive cells, local assemblies of which are capable of generating periodic waves of activation in the tissue through diffusive coupling [9, 10]. These waves traveling through an organ are capable of sustaining spatio-temporally coherent contractions [11]. Indeed it has been demonstrated that one of the simplest ways to achieve this is by having an excitable cell coupled by gap-junctions to one (or more) electrically passive cells characterized by a resting state membrane potential that is much higher than that of the excitable cell [12]. The coupling between these heterogeneous cell types causes the membrane potential of the excitable cell to be driven beyond its threshold, resulting in the generation of an action potential. Subsequently, the excitable cell attempts to return to its resting state, but after a period of recovery is again driven to exceed its threshold by the passive cells coupled to it, thereby resulting in a periodically recurring series of action potentials. Thus, although neither excitable nor passive cells are individually capable of spontaneous sustained activation, an assembly of these two cell types can generate periodic oscillations [13].

The emergence of periodic activity in a heterogeneous assembly of excitable and passive cells makes such a mechanism a viable candidate for self-organized systemwide coherent oscillations in physiological contexts where no pacemakers have been reported [14, 15]. Indeed, it has been demonstrated that a lattice of excitable cells that are each coupled to a variable number of passive cells, can exhibit a range of spatio-temporal phenomena consistent with those observed in the uterus during the transition to coherent activity seen prior to parturition [11, 16, 17]. However, noting that each local cellular assembly are either in an excitable or an oscillatory dynamical regime in isolation, raises an important question; can the observed collective behavior be reproduced in an even simpler setting, viz., where each lattice site is occupied by either an oscillatory or an excitable element, a situation reminiscent of percolation [18]. In this paper, we consider the dynamics of two classes of systems, each capable of exhibiting spontaneous collective dynamics, one comprising electrically active and passive cells (EP) and the other comprising oscillatory and excitable cells (OE). We observe that simple motifs of cells described using the EP model are capable of exhibiting a wide range of complex collective dynamical patterns. However, several of these are no longer observed when such cells are embedded in a spatially extended system, suggesting an emergent simplicity of the collective dynamics. More importantly, we observe that when cells described by the OE model are placed on a lattice, the resulting dynamics are qualitatively very similar to that obtained using the EP model. This points towards a more fundamental mechanism that could explain the emergence of spontaneous recurrent activity in physiologically relevant contexts.

To investigate in detail the range of complex behavior that emerges upon diffusively coupling excitable cells, each of which are in turn coupled to one or more passive cells, we consider the simplest possible assemblies of these cells capable of exhibiting spontaneous oscillatory activity. Following Ref. [11], we simulate the electrical

activity of an excitable cell using the FitzHugh-Nagumo (FHN) model [19], which is capable of both excitable and oscillatory dynamics. The model describes the temporal evolution of an activation variable V_e (the membrane potential), and an inactivation variable g (an effective trans-membrane conductance) as $V_e = \mathcal{F}(V_e, g)$, $\dot{g} = \mathcal{G}(V_e, g)$. Here, $\mathcal{F}(V_e, g) = A V_e (V_e - \alpha)(1 - V_e) - g$ and $\mathcal{G}(V_e, g) = \epsilon(k_e V_e - g - b)$, where A(=3) and $k_e(=1)$ govern the kinetics, $\alpha (=0.2)$ is the excitation threshold and $\epsilon (= 0.08)$ is the recovery rate, while b provides a measure of the asymmetry of the limit cycle. We note that for $b_{c1} (= 0.127) < b < b_{c2} (= 0.343)$ the cell exhibits periodic oscillations, while outside this range it is excitable with a stable resting state. The value of V_e in the resting state is close to 0 for $b < b_{c1}$ and consequently this regime is characterized as a "low" stable state. For $b > b_{c2}$, the resting state value of V_e is relatively large, with the regime being referred to as a "high" stable state. The temporal evolution of the passive cell is described in terms of its membrane potential V_p as $\dot{V}_p = K(V_p^R - V_p)$, where V_p^R (= 1.5) is the resting state and K (= 0.25) is the relaxation rate [20]. Each excitable cell is electrically coupled to $n_p (= 0, 1, 2...)$ passive cells, where the conductance of the gap junctions between the two cell types is C_r . Thus, the set of equations used to describe the dynamics of an excitable cell i coupled to n_p^i passive cells, as well as to an excitable cell j, is:

$$\frac{dV_e^i}{dt} = \mathcal{F}(V_e^i, g^i) + n_p^i C_r (V_p^i - V_e^i) + D(V_e^j - V_e^i),
\frac{dg^i}{dt} = \mathcal{G}(V_e^i, g^i),
\frac{dV_p^i}{dt} = K(V_p^R - V_p^i) - C_r (V_p^i - V_e^i).$$
(1)

The dynamics of an isolated excitable cell coupled to one or more passive cells depends on n_p and C_r . As V_p^R is much higher than the excitation threshold α , for a range of n_p and C_r the coupled excitable-passive system can exhibit oscillations. To demonstrate that such emergent oscillations result exclusively from the coupling we have chosen b=0, so that in isolation the FHN dynamics converges to the low stable state. It is important to note in the context of the results reported here that when $C_r > 0.5$, oscillations are seen only for $n_p = 1$ while those excitable cells coupled to $n_p > 1$ passive cells converge to high stable states. We first consider the simplest nontrivial assembly of dissimilar excitable-passive units, viz., a pair of excitable cells diffusively coupled with strength D, each interacting with a different number n_p of passive cells with strength C_r (Fig. 1). The heterogeneity in n_p implies that the intrinsic behavior of the two units are dissimilar, and we observe a range of distinct collective dynamics upon varying D and C_r , as shown in Fig. 1 (a-d) for four distinct connection topologies of the assemblies [illustrated in the top right corners of the corresponding panels. The dynamical regimes obtained can

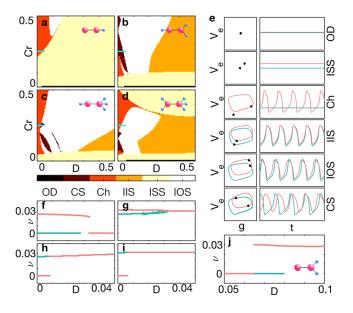


FIG. 1. Emergent dynamics obtained with different motifs of diffusively coupled excitable and passive cells. (a-d) Collective dynamical patterns observed over a range of values of diffusive coupling strengths between two excitable cells (D) and between an excitable and a passive cell (C_r) , obtained using different motifs (shown as insets in the corresponding panels, where the larger and smaller circles represent excitable and passive cells, respectively). The regimes are classified according to the dominant attractor obtained for the given parameter set, and include oscillation death (OD), inhomogeneous steady state (ISS), chimera (Ch), inhomogeneous in-phase synchronization (IIS), inhomogeneous out-of-phase synchronization (IOS) and cluster synchronization (CS). (e) Phase plane trajectories and time series of the excitable cells for the different collective dynamical regimes displayed in (a-d). The black dots represent the instantaneous position of the two cells on the corresponding limit cycle. (f-i) Variation of the frequency ν of the excitable cells on the left (green) and right (maroon) in each of the four motifs in (a-d) for $C_r = 0.25$, over the range of D indicated by horizontal cyan bars in (a-d). As D increases, frequencies of the two cells merge and for sufficiently strong coupling the system can either stop oscillating (f), or display a frequency that is between (g), greater than (h) or equal to (i) the maximum of the intrinsic frequencies. (j) Variation of ν with Dfor $C_r = 0.6$, obtained using the motif shown as an inset. Although both cells are quiescent for low coupling strength, they exhibit oscillations for sufficiently large D.

be classified on the basis of the V_e time-series of the excitable cells, using a set of order parameters with specified threshold values (see SI for details): (i) oscillation death (OD), where both cells are in the same temporally invariant non-zero steady state; (ii) inhomogeneous steady state (ISS), where both cells are in different temporally invariant steady states; (iii) chimera (Ch), where only one of the two cells oscillate; (iv) inhomogeneous in-phase synchronization (IIS), where both cells oscillate in-phase; (v) inhomogeneous out-of-phase synchroniza-

tion (IOS), where both cells have the same frequency but are out-of-phase with each other, and, (vi) cluster synchronization (CS), where the two cells have different oscillation frequencies [Fig. 1 (e)]. At low values of D, the two units can behave very differently, and we observe collective states such as Ch or CS. As D increases, the cells either become frequency locked or cease oscillating altogether. Note that the intrinsic heterogeneity of the two units prevents exact synchronization between them even for large D. For a given value of D, as C_r is decreased, eventually the cells stop oscillating (in isolation, neither an excitable nor a passive cell is capable of spontaneous activity).

In Fig. 1 (f-i), we display the variation of the frequency ν of the periodic activity exhibited by the excitable cells in the low D regime in each of the different assemblies $(C_r \text{ is fixed at } 0.25 \text{ in each case})$. We observe that an increase in D either gives rise to the cessation of oscillations [Fig. 1(f)] or a synchronized state in which the two units oscillate at a common frequency that is either lower [Fig. 1(g)], higher [Fig. 1(h)] or equal to [Fig. 1(i)] the higher of the pair of intrinsic frequencies (i.e., the frequencies of each unit at D = 0). Just as coupling an excitable cell to one or more passive cells can, under appropriate conditions, give rise to oscillatory dynamics, we observe that spontaneous activity can arise upon coupling a pair of dissimilar units that do not oscillate in isolation. Fig. 1 (j) shows a pair of excitable cells, having $n_p = 0$ and $n_p = 2$ respectively, such that neither can independently oscillate for $C_r = 0.6$. However, upon increasing D sufficiently, we observe a transition to a Ch state and eventually to a frequency synchronized state of the two units.

Upon increasing the number of units in an assembly, we observe that the system becomes capable of exhibiting more complex collective behavior including chaotic activity. However, a particularly intriguing collective state of coexisting chaotic and non-chaotic activity is observed in an assembly of three excitable cells, having $n_p = 1, 2, 3$ respectively, that are coupled in a chain [see top right corner of Fig. 2 (a)]. For a large range of values of C_r and D, the system exhibits IOS [Fig. 2 (a)]. However, in the CS regime, we observe a collective dynamical state that is characterized by chaotic behavior in the excitable cell with $n_p = 1$ with non-chaotic, periodic oscillations in the other two cells [Fig. 2 (b-d)] The qualitative difference in the dynamics of the three excitable cells is evident upon comparing their time series [Fig. 2(b)], phase plane portraits [Fig. 2(c)] and power spectral densities [Fig. 2(d)]. A more rigorous comparison, considering the response of each cell to small perturbations, shows rapid divergence of the resulting trajectory from the unperturbed one for the chaotic unit, with no such deviation observed for the other two units (see SI). We note that permutations of the connection topology of this assembly, i.e. changing the order in which the cells with different values of n_p are

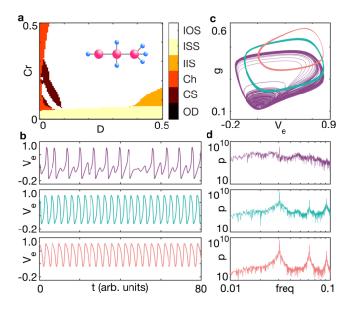


FIG. 2. Coexistence of chaotic and non-chaotic dynamical activity in a system of three diffusively coupled excitable cells, each coupled to a different number (n_n) of passive cells. (a) Collective dynamical patterns observed over a range of values of diffusive coupling strengths D between excitable cells and the coupling C_r between excitable and passive cells for the system shown as an inset. The dynamical regimes are classified according to the dominant attractor obtained for the given parameter set, and are the same as those detailed in Fig. 1 (a-d). (b) Time series of membrane potential V_e of the excitable cells coupled to (top) $n_p = 1$, (middle) $n_p = 2$ and (bottom) $n_p = 3$ passive cells, for D = 0.02 and $C_r = 0.19$. (c-d) Phase plane trajectories and power spectral densities of the three excitable cells, colored as in the corresponding panels of (b).

placed in the chain, yields similar qualitative behavior, with chaotic dynamics consistently observed in the unit with the lowest n_p .

It may appear that increasing the size of the assemblies further, by adding more coupled excitable-passive units, can only lead to a further increase in the complexity of the collective dynamics. However, surprisingly, we observe an emergent simplicity in the behavior of large lattices of such units, with neighboring elements coupled diffusively to each other. Indeed, such an example is provided by a spatially extended model of uterine tissue, which is heterogeneous by nature, comprising electrically active myocytes that are excitable (thereby facilitating muscle contractions), as well as electrically passive cells such as interstitial Cajal-like cells (ICLC) [21] and fibroblasts [see top panel of Fig. 3 (a)]. The system exhibits spontaneous oscillations for a range of values of C_r even though, in isolation, none of the individual cells are capable of autonomous periodic activity, as has been experimentally observed in uterine tissue [14, 15]. More important from the perspective of the dynamical transition

to periodic coordinated contraction of the myometrium, it is seen that increasing D results in the self-organized emergence of global synchronization, and eventually coherence [11, 17]. It is striking that such coordination is achieved exclusively through local interactions between cells and does not require a centralized pacemaker such as that present in the heart (viz., the sino-atrial node).

The relative simplicity of the collective behavior of such a lattice of heterogeneous coupled cells can be shown by demonstrating that it can be captured by a reduced description of the system in terms of interacting dynamical elements, each of which are either in an oscillating or a steady state. In particular, we can replace excitable-passive cell assemblies that are capable of spontaneous periodic activation by a single FHN unit with $b_{c1} < b < b_{c2}$ (for concreteness, we choose $b_{osc} = 0.192$ for the simulations whose results are shown here), and FHN units with $b < b_{c1}$ (> b_{c2}) for cell assemblies exhibiting a low (high) stable state (we choose $b_{exc}^{low}=0$ and $b_{exc}^{high}=0.394$ for the simulations shown here). The resulting equivalent lattice now comprises only FHN units, a fraction f of which are in an oscillatory regime with the remaining being excitable by virtue of having different values of b [see Fig. 3 (a), bottom panel]. Nevertheless, the system exhibits qualitatively identical behavior to that seen in models of uterine tissue simulated by coupling assemblies of excitable and passive elements, e.g., the occurrence of cluster synchronization at relatively low inter-cellular coupling that gives way to global synchronization of periodic activity (coordinated by propagating waves of excitation that traverse the lattice) for stronger coupling [Fig. 3 (b)].

The similarity of the emergent properties of the simpler model can be established further by comparing the different dynamical regimes of the f-D parameter space with that observed in the uterine model having heterogeneous cell types [11]. Indeed all the qualitatively distinct types of behavior reported in the latter can be seen in Fig. 3 (c), including No Oscillation (NO, with all cells in steady states), Cluster Synchronization (CS, marked by coexistence of multiple groups of cells, each characterized by a different frequency), Local Synchronization (LS, coexistence of quiescent cells with cells oscillating at a common frequency), Global Synchronization (GS, all cells have the same oscillation frequency) and Coherence (COH, all cells exhibit phase synchrony). In the limit of large D, the dynamics of lattice can be further simplified and an implicit analytical equation can be obtained for f_c , the fraction of FHN units that should be oscillatory for the system to exhibit persistent periodic excitations. It marks the boundary between the NO and COH regimes and is given as $b_{c1} = (1 - f_c) b_{exc} + f_c b_{osc}$. For the situation shown in Fig. 3 (c), $b_{exc} = b_{exc}^{low}$, which yields $f_c \sim 0.7$ upon inserting the corresponding numerical values.

We can investigate the collective dynamics around this

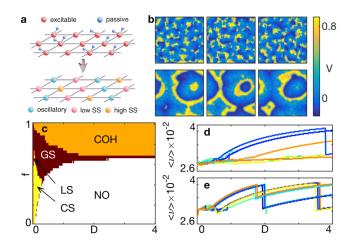


FIG. 3. Collective dynamics of a lattice of diffusively coupled elements that can be either excitable or oscillatory. (a) Schematic diagram of uterine tissue, modeled as a two-dimensional lattice where each site comprises an excitable cell coupled to a variable number of passive cells (top, the "EP" model). The dynamics at each site can equivalently be described through cells that are either oscillatory or excitable (bottom, the "OE" model). The latter cell type could be in one of two possible steady states, characterized by low and high values of the state variable V. Note that the number of passive cells coupled to each excitable cell in the top panel is chosen such that the uncoupled dynamics at that site is qualitatively the same as that of the corresponding site in the bottom panel. (b) Snapshots of the activity V in a planar simulation domain comprising an equal mixture of excitable and oscillatory elements (f = 0.5) diffusively coupled with strength D to nearest neighbors, showing (top row, D = 0.1) cluster synchronization (CS) and (bottom row, D=0.3) global synchronization (GS). (c) Varying the diffusion coefficient D and the fraction of oscillatory cells f in the lattice for the case of the OE model, several distinct dynamical regimes are observed. In addition to CS and GS, these include local synchronization (LS), coherence (COH) and no oscillations (NO). The regions are identified according to the collective dynamics observed for the majority (> 50%)of initial conditions. (d-e) Variation of the mean oscillatory frequency $\langle \nu \rangle$ with D. Each curve is obtained by starting from different random initial states at low D and then gradually increasing D over time. In panel (d), the cell at each site can be either oscillatory (with probability f = 0.7) or excitable (with probability 1-f). In panel (e), we first associate with each lattice site a random number n drawn from a Poisson distribution (with mean $\lambda = 0.7$), and then place oscillatory cells in sites with n=1 and excitable cells having a low or high V steady states at sites with n = 0 and n > 1, respectively. All simulations are performed on square lattices comprising 64×64 cells, with periodic boundary conditions.

asymptotic boundary between persistent oscillatory activity and a quiescent steady state by considering the case f=0.7. In particular, we focus on the variation of the overall activation rate, measured by the mean frequency of periodic activation $\langle \nu \rangle$ (averaged over all oscil-

lating elements in the lattice), as D is increased. In order to be consistent with the physiological setting, where the coupling between cells increases over the gestation period (as a result of hormone induced increased expression of gap junctions that electrically couple the cells [22]), D is increased adiabatically over the course of a single realization, from a random initial condition at a low value of D. For the situation when $b_{exc} = b_{exc}^{low}$, shown in Fig. 3 (d), $\langle \nu \rangle$ increases with D until it reaches a maximum value related to the reciprocal of the refractory period (set by the parameters of the FHN model). Increasing D further results in an abrupt drop in $\langle \nu \rangle$ as the number of propagating wavefronts in the system changes. A subsequent increase in D results in an increase in $\langle \nu \rangle$ generated by the new spatio-temporal pattern. This is qualitatively the same as the phenomenon observed for the model of uterine tissue involving assemblies of excitable and passive cells. An even closer match between the two classes of models can be obtained if we replace each of the excitable elements with FHN elements having either $b_{exc}=b_{exc}^{low}$ or $=b_{exc}^{high}$ according to the following procedure: first, each lattice site is assigned a value n chosen from a Poisson distribution with mean $\lambda = f$. Note that this is identical to the process by which the number of passive cells (given by n) coupled to an excitable cell are determined in modeling uterine tissue with excitable-passive cell assemblies [11]. Next, FHN units in the oscillatory regime ($b = b_{osc}$) are placed at sites having n=1, while FHN units with $b=b_{exc}^{low}$ (i.e., excitable element with a low stable state) are placed at sites with n = 0. At sites having n > 1, corresponding to excitable-passive cell assemblies whose activity is arrested at a high stable state, FHN units with $b=b_{exc}^{high}$ are placed. The resulting oscillatory-excitable (OE) model can accurately reproduce dynamical behaviors reported for the model comprising excitable-passive (EP) cell assemblies [11]. These include the emergence of clusters characterized by a common oscillation frequency, propagating wavefronts, as well as self-sustained spiral waves in the GS regime [see SI for details]. Note that persistent periodic activity can arise upon coupling two FHN units that cannot independently oscillate, provided one of them is in the low and the other in the high stable state - a phenomenon analogous to the appearance of oscillations in assemblies of excitable and passive cells which cannot sustain autonomous activity. Fig. 3 (e) shows the evolution of the mean frequency with D when the cellular coupling is increased adiabatically starting from a random initial condition over the course of a single realization, displaying an even closer agreement to the behavior seen in the EP model of uterine tissue [11].

The qualitative equivalence of the collective behavior in large lattices for the two classes of models is all the more surprising as the dynamics of network motifs comprising excitable-passive cell assemblies (discussed above, see Figs. 1 and 2) is much more complex than that observed upon replacing each assembly by a FHN unit in the oscillatory or excitable regime. For instance, coupling a pair of EP cell assemblies, each of which oscillate at different frequencies, cannot give rise to exact synchronization even at large D. However, two FHN oscillators characterized by distinct b values (and hence, frequencies) can exhibit exact synchronization when coupled with sufficient strength. Furthermore, while we have reported motifs of connected EP cell assemblies that exhibit chimera (Ch) regimes over a range of coupling strengths, such behavior cannot be seen in two coupled FHN oscillators with distinct intrinsic frequencies. We would also like to point out that nothing equivalent to the chaotic behavior observed in a motif comprising coupled EP cell assemblies (Fig. 2) is seen in systems of coupled FHN oscillators arranged in a similar topology (viz., a chain comprising two oscillators having different intrinsic frequencies and an excitable element). Thus, even though the OE model reproduces the collective behavior of a large system of coupled EP cell assemblies, the dynamics at the microscopic level (i.e., motifs comprising only a few elements) can be extremely different for the two classes of models (see SI).

To conclude, in this paper we have shown that while coupled excitable-passive cell assemblies are capable of exhibiting a wide range of dynamical behaviors including chaos, a macroscopic system comprising a large number of such elements diffusively coupled to their nearest neighbors on a lattice shows relatively simpler spatiotemporal phenomena. In particular, this resulting collective dynamics can be reproduced by a model comprising many elements, each described by a generic model for an excitable cell that is either in a steady state or in an oscillatory regime. Indeed, it suggests that the behavior associated with physiologically detailed models of uterine tissue activity [16, 17, 23] can be understood in terms of a reduced model involving a heterogeneous assembly of coupled oscillatory and excitable elements. More importantly, our results point towards a generalization of the mechanism proposed in Ref. [12] for the emergence of periodic activity in systems where none of the individual elements are intrinsically capable of oscillating. While it was shown there that persistent oscillations arise upon coupling excitable and electrically passive cells under certain circumstances, here we have shown that an oscillating system may emerge upon coupling elements, each of which are in isolation at time-invariant steady states provided these states are dissimilar (i.e., the state variables associated with them have sufficiently distinct numerical values, corresponding to "low" and "high"). Furthermore, our demonstration of a large variety of dynamical attractors in small assemblies of excitable and passive elements can provide an understanding of the complex dynamics seen in electrically coupled heterogeneous sub-cellular compartments in neurons [24, 25] and small networks of neurons interacting via gap-junctions [26].

We would like to thank Sitabhra Sinha and K. A. Chandrashekar for helpful discussions. SNM has been supported by the IMSc Complex Systems Project (12th Plan), and the Center of Excellence in Complex Systems and Data Science, both funded by the Department of Atomic Energy, Government of India. The simulations and computations required for this work were supported by the Institute of Mathematical Sciences High Performance Computing facility (hpc.imsc.res.in) [Nandadevi and Satpura clusters].

- [1] L. Glass, Nature (London) **410** 277 (2001). doi: 10.1038/35065745
- [2] M. I. Rabinovich, P. Varona, A. I. Selverston, and H. D. Abarbanel, Rev. Mod. Phys. 78, 1213 (2006). doi: 10.1103/RevModPhys.78.1213
- [3] S. Sinha and S. Sridhar, Patterns in Excitable Media: Genesis, Dynamics, and Control, (CRC Press, Boca Raton, FL, 2015).
- [4] J. D. Huizinga, L. Thuneberg, M. Klüppel, J. Malysz,
 H. B. Mikkelsen, and A. Bernstein, Nature (Lond.) 373,
 347 (1995). doi:10.1038/373347a0
- [5] L. Thomson, T. L. Robinson, J. C. F. Lee, L. A. Farraway, M. J. G. Hughes, D. W. Andrews, and J. D. Huizinga, Nat. Med. 4, 848 (1998). doi: 10.1038/nm0798-848
- [6] J. D. Huizinga et. al., Nat. Commun. 5, 3326 (2014). doi:10.1038/ncomms4326
- [7] M. R. Boyett, H. Honjo, and I. Kodama, Cardiovasc. Res. 47 658 (2000). doi:10.1016/S0008-6363(00)00135-8
- [8] R. Smith, M. Imtiaz, D. Banney, J. W. Paul, and R. C. Young, Am. J. Obstet. Gynecol. 213, 181 (2015). doi:10.1016/j.ajog.2015.06.040
- [9] J. H. E. Cartwright, Phys. Rev. E 62, 1149 (2000). doi: 10.1103/PhysRevE.62.1149
- [10] C. Degli Esposti Boschi, E. Louis, and G. Ortega, Phys. Rev. E 65, 012901 (2001). doi: 10.1103/PhysRevE.65.012901
- [11] R. Singh, J. Xu, N. B. Garnier, A. Pumir, and S. Sinha, Phys. Rev. Lett. 108, 068102 (2012). doi: 10.1103/PhysRevLett.108.068102
- [12] V. Jacquemet, Phys. Rev. E 74, 011908 (2006). doi: 10.1103/PhysRevE.74.011908
- [13] T. A. Quinn, P. Camelliti, E. A. Rog-Zielinska, U. Siedlecka, T. Poggioli, E. T. O'Toole, T. Knöpfel, and P. Kohl, Proc. Natl. Acad. Sci. USA 113, 14852 (2016). doi:10.1073/pnas.1611184114
- [14] S. Wray, S. Kupittayanant, A. Shmygol, R. D. Smith, and, T. Burdyga, Exp. Physiol. 86, 239 (2001). doi: 10.1113/eph8602114
- [15] R. C. Young, Best Pract. Res. Clin. Obstet. Gynaecol. 52, 68 (2018). doi:10.1016/j.bpobgyn.2018.04.002
- [16] J. Xu, S. N. Menon, R. Singh, N. B. Garnier, S. Sinha, and A. Pumir, PLoS ONE 10, e0118443 (2015). doi: 10.1371/journal.pone.0118443
- [17] J. Xu, R. Singh, N. B. Garnier, S. Sinha, and A. Pumir, New J. Phys. 15, 093046 (2013). doi:10.1088/1367-2630/15/9/093046
- [18] D. Stauffer and A. Aharony, Introduction to Percolation

- Theory, (CRC Press, Boca Raton, FL, 1994).
- [19] J. Keener and J. Sneyd, Mathematical Physiology, (Springer, New York, 1998).
- [20] P. Kohl, A. G. Kamkin, I. S. Kiseleva, and D. Noble, Exp. Physiol. 79, 943 (1994). doi: 10.1113/expphysiol.1994.sp003819
- [21] R. A. Duquette, A. Shmygol, C. Vaillant, A. Mobasheri, M. Pope, T. Burdyga, and S. Wray, Biol. Reprod. 72, 276 (2005). doi:10.1095/biolreprod.104.033506
- [22] E. Jahn, I. Classen-Linke, M. Kusche, H. M. Beier, O. Traub, R. Grummer, and E. Winterhager, Hum. Reprod. 10, 2666 (1995). doi: 10.1093/oxfordjournals.humrep.a135764
- [23] W. C. Tong, C. Y. Choi, S. Karche, A. V. Holden, H. Zhang, and M. J. Taggart, PLoS ONE 6, e18685 (2011). doi:10.1371/journal.pone.0018685
- [24] B. V. Safronov, M. Wolff, and W. Vogel, Biophys. J. 78, 2998 (2000). doi:10.1016/S0006-3495(00)76838-X
- [25] J. M. Bekkers and M. Häusser, Proc. Natl. Acad. Sci. USA 104, 11447 (2007). doi:10.1073/pnas.0701586104
- [26] G. J. Gutierrez, T. O'Leary, and E. Marder, Neuron 77, 845 (2013). doi:10.1016/j.neuron.2013.01.016