

Article Title

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Abstract

There's nothing here yet

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1 Introduction

The apicomplexan parasite *Plasmodium falciparum* is the most virulent causative agent of malaria, and responsible for over 600,000 deaths annually [1]. Along with other members of the *Plasmodium* family, *P. falciparum* has a complex life-cycle, moving between several different tissues in both mammalian and arthropod hosts. Symptomatic disease in humans occurs when *P. falciparum* undergoes rounds of asexual reproduction inside human red blood cells (RBCs) [2].

In some respects, the intracellular environment of a red blood cell is an ideal location for parasite proliferation. The cells' lack of an MHC (Major Histocompatibility Complex) system, which would otherwise be used to identify intracellular pathogens to the host immune system, renders parasites immunologically invisible [3], whilst the vascular system allows the parasite to travel throughout the body. The highly specialised nature of RBCs, however, means that the intracellular environment also presents significant challenges to parasite survival.

Mature red blood cells lack protein production and export machinery, and are a nutritionally poor environment, with a proteome dominated by haemoglobin, which typically accounts for around 98% of the protein content of the cell [4]. *P. falciparum* is able to digest RBC proteins, however haemoglobin lacks several amino acids required for protein production. Red blood cells are also subject to regular 'quality control' in the spleen, where damaged or infected cells are killed and recycled [5].

In order to survive and proliferate inside RBCs, *P. falciparum* exports a range of proteins which radically transform the red blood cell, collectively termed the **exportome**. Many of these proteins are involved in setting up a parasite-derived protein export system, capable of directing exported *P. falciparum* proteins to sites both inside and outside the RBC. *P. falciparum* resides inside a parasitophorous vacuole, and exports proteins via structures termed Maurer's Clefts [6], which bear some similarity to golgi apparatus [7].

In order to avoid detection in the spleen, many exported proteins are associated with the formation of knobs, specialised structures which form at the RBC membrane and promote cytoadherence to epithelial cells, platelets and other red blood cells [8]. Severe forms of malaria, including cerebral malaria, are believed to be caused by sequestration of infected red blood cells in deep tissues, as well as overinduction of inflammatory cytokines [2]. Other exported components have been associated with increasing RBC membrane permeability to facilitate nutrient and waste exchange and strengthening the RBC cytoskeleton (Reviewed in [5]).

To date, at least 10% of the protein products of the *P. falciparum* genome have been shown to be exported to the host cell [9]. Within this exportome, there exist 360 distinct proteins once close duplicates are excluded.

Whilst the *P. falciparum* genome has been available since 2002 [10], comparatively few genes have been studied in depth, and many remain of unknown function. The discovery of a motif termed the PEXEL (Plasmodium Export Element) shared between many exportome components has made it possible to reliably predict the *P. falciparum* exportome in the absence of other information [11].

The PEXEL motif is pentameric, located near the N-terminal of the protein, and can be generalised as the amino acid sequence RxLxE/Q/D [12], where x is any non-charged amino acid [13] although the non-canonical PEXEL motif KxLxE/Q/D and relaxed PEXEL motif RxxLxE/Q/D are also seen occasionally [5]. It is known that the amino acid sequence cleaved after the leucine residue in the parasite ER [12] although how the PEXEL motif targets the protein for export

remains unclear. The *P. falciparum* protein Plasmeprin V has been shown to cleave a subset of PEXEL-carrying proteins [14].

In addition to exported PEXEL proteins, several PEXEL Negative exported proteins have been identified using transcription profiling [15]. Based on experimental evidence, a cryptic signal is thought to exist near the N-terminal of both mature (cleaved) PEXEL proteins and PNEPs [16], although the nature of this sequence remains unclear.

Understanding the protein-protein interactions responsible for the transformation of the infected red blood cell is both an interesting scientific challenge and an important step towards a better understanding of malaria in humans. Whilst an interactome for *P. falciparum* proteins has been produced using a yeast-two-hybrid [17] it is of low quality and contains many false positives.

1.1 Protein Structure Prediction

The design of constructs for experimental use is hampered somewhat a lack of structural information about *P. falciparum* exportome components. To date, only 4 components of the *P. falciparum* exportome have solved structures in the PDB (own research, <https://gist.github.com/tomnewport/a04868602d3482d33921>), and so knowledge based modelling using a tool such as MODELLER [18] is not applicable. There exist several tools, however, which are able to predict features and domains of proteins using a variety of approaches.

Secondary Structure and Disorder Prediction

Parts of a protein such as helices and strands will have a fixed 3D structure and pattern of amino acids and are said to be ordered. Other parts of the protein, especially loops, may not possess such an ordered or fixed structure and so are said to be disordered. Disordered parts of a protein often connect domains or features of the protein secondary structure and so disorder prediction can be used to find domain boundaries. Several algorithms exist to predict both secondary structure and disorder from amino acid sequence, and the tool metaPrDOS [19] can be used to obtain a consensus from a selection of disorder prediction algorithms.

Coiled Coil Prediction

Coiled coils are a common structural motif whereby two or more alpha helices are wound together, often important in the formation of oligomers and complexes. The COILS software tool uses a database of known coiled coil motifs to predict the likelihood of a coiled coil at particular sites on in an amino acid sequence [20].

Transmembrane Prediction

Proteins may include several domains which cross or interact with the hydrophobic environment of the lipid membrane of a cell or compartments within the

cell. The software tool TMHMM uses a hidden Markov model based approach to predict these domains based on the protein’s amino acid sequence alone [21].

Combined Approaches

Some software tools combine several different approaches to structure prediction. InterPro [22] performs coiled coil prediction and transmembrane prediction, and searches for known domains with similar sequences. Results are presented in a single graphic showing predicted domains along the amino acid sequence.

Phyre2 [23] performs disorder prediction, secondary structure prediction and transmembrane prediction before comparing against a fold database and attempting to build a model of the tertiary structure based on the amino acid sequence. Results are presented in a series of figures as well as 3D models based on different templates in a PDB format.

2 Software implementation

2.1 Aims

This project aims to build software tools to automate bioinformatic analysis of *P. falciparum* exported proteins and red blood cell proteins and present the results in a web-based interface. This will be used to aid in the design of protein constructs used to produce a high quality interactome for exported *P. falciparum* proteins.

The *P. falciparum* genome and other associated data are already available from several online databases, including the dedicated PlasmoDB (<http://plasmodb.org/>) [24] which provides functional genomic data for genes found in several *Plasmodium* species and includes annotations such as gene location, polymorphisms and expression data, as well as some limited structural annotations (Figure 1). No database presently provides a broad range of structural annotations required for construct design, or allows the user to look only at proteins known to be exported to the red blood cell.

Protein structure prediction is a fast evolving field. As existing tools are updated and new tools are published, both the inputs and outputs of such a tool may change. It is therefore vital to consider modularity and flexibility at an early stage to ensure future development is not compromised by design decisions made earlier in development.

2.2 Overview

The finished software will consist of three distinct parts. A local client implemented in Python, will request analyses of newly added sequences from remote servers and then parse and store the response, whilst a remote client implemented in JavaScript will load and display protein files to the end user. A simple HTTP server will serve and cache JavaScript files necessary for the remote client and data files generated by the Python client.



Figure 1: **PlasmoDB Structural information for PF3D7_1149000** — This includes, amongst others, InterPro domains, secondary structure prediction and export domains. Note that the protein product of PF3D7_1149000 has been shown to be exported.

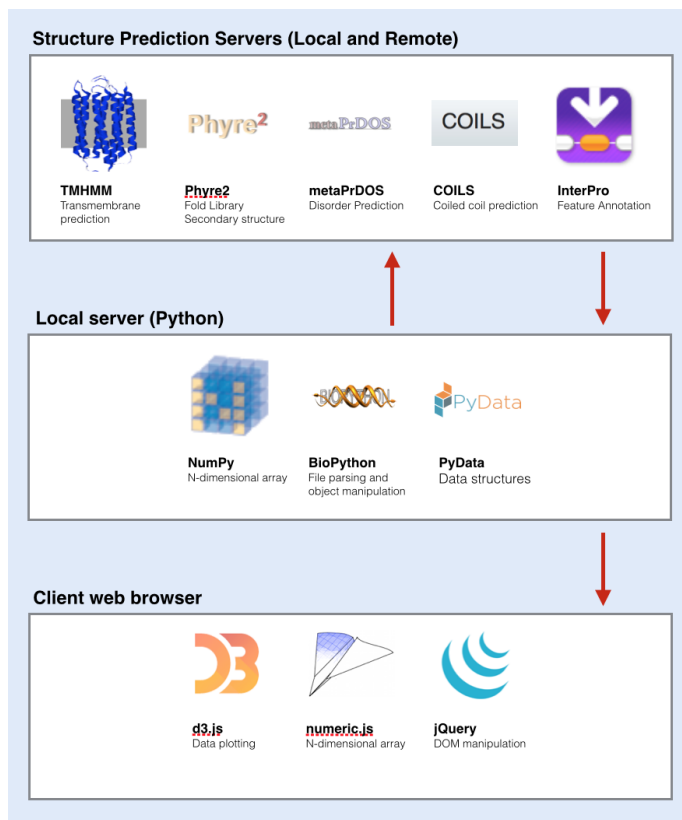


Figure 2: **High-level overview of planned software** — Sequence data is submitted to the servers TMHMM, Phyre2, metaPrDOS, COILS and InterPro using a python script running on a local server. The data is then retrieved and processed. Once processed and stored, data may be requested by the client and displayed using client-side scripts for visualisation.

2.2.1 Python server

The local python client, which is responsible for requesting, retrieving and parsing structure predictions, may be provided with a list of gene names which correspond to genes available through PlasmoDB (<http://plasmodb.org/>) [24]. These will then be retrieved in FASTA format from PlasmoDB and submitted several servers: TMHMM for transmembrane prediction; Phyre2 for fold recognition and secondary structure predication; metaPrDOS for disorder prediction; COILS for coiled coil prediction and InterPro for feature annotation. Submission to web-based servers will be performed using HTTP requests to CGI (Common Gateway Interface) scripts on respective remote servers using the Python Requests library (<http://python-requests.org>) and similarly retrieved once completed.

Parsing of results will be performed primarily using the Pandas dataframe provided by PyData (<http://pandas.pydata.org>), assisted by BioPython [25], used to open files in biological formats such as PDB and FASTA, and Numpy [26], which implements a computationally efficient n-dimensional array for the Python language. Results from many servers will be saved to two files in comma-separated values format, one storing series with a defined value for each amino acid (a prediction of the likelihood of each amino acid forming part of a coiled coil, for example), and one (annotations) storing annotations identifying features stretching across several amino acids (domains identified based on similarity to common motifs in a database, for example).

| Position | Amino Acid | Series 1 | Series 2 | ... | Series N |
|----------|------------|----------|----------|-----|----------|
| 1 | S | 0.2 | 20 | ... | 0.9 |
| 2 | N | 0.3 | 14 | ... | 0.1 |
| 3 | I | 0.2 | 12 | ... | 0.3 |

Table 1: **Series Table Example** — Each series has a defined value for each amino acid. Each column represents a single series, and the table possesses an unlimited number of columns. Each row represents a single amino acid.

| Start Position | End Position | Source | Description |
|----------------|--------------|----------|----------------------------|
| 10 | 143 | InterPro | Transmembrane Region |
| 53 | 197 | Phyre2 | Duffy receptor, alpha form |

Table 2: **Annotation Table Example** — Each annotation consists of a start position and an end position, a source (the server which identified it) and a description (the feature which was identified by the server). Each row is an annotation, and the table may contain an unlimited number of rows.

2.2.2 Public facing server

Since the public-facing HTTP server only retrieves files (and is not able to trigger any additional server-side processing), any expertly built HTTP server may be used without modification, greatly simplifying the problems of installing, maintaining and securing a publicly accessible server on the web.

Unlike most existing web-based tools for viewing genomic data where visualisations are rendered on the server and transmitted as images, visualisations will be rendered client-side based on raw data. Users will therefore require a recently updated web browser in order to view visualisations of data using the remote client. The client-side approach removes the need for the server to render images, reducing load on the server, and also removes the need to transmit images, reducing the load on the network. Additionally, advanced users are able to tweak visualisations as required simply by changing rules in the CSS (Cascading StyleSheet) files supplied with the remote client.

2.2.3 Remote JavaScript client

An HTML (HyperText Markup Language) document provided by the server will download the remote client and all dependencies (including default style sheets) to the user's web browser. The remote client will consist of two main classes, a `data_manager` class which will load data from the server, perform some client side processing and then provide an object which can be used to retrieve data for plotting, and a `ui_bindings` class, which will provide an interface which may be used to create, update and link different UI elements.

2.2.4 The `data_manager` class

The `data_manager` class will first download a schema from the server in JSON format which will describe where data required for visualisations may be found.

2.2.5 The `ui_bindings` class

[?]

2.3 Implementation

3 Discussion

4 Limitations and Further Work

Acknowledgments

I am grateful to John Vakonakis for his insight and encouragement whilst supervising this project.

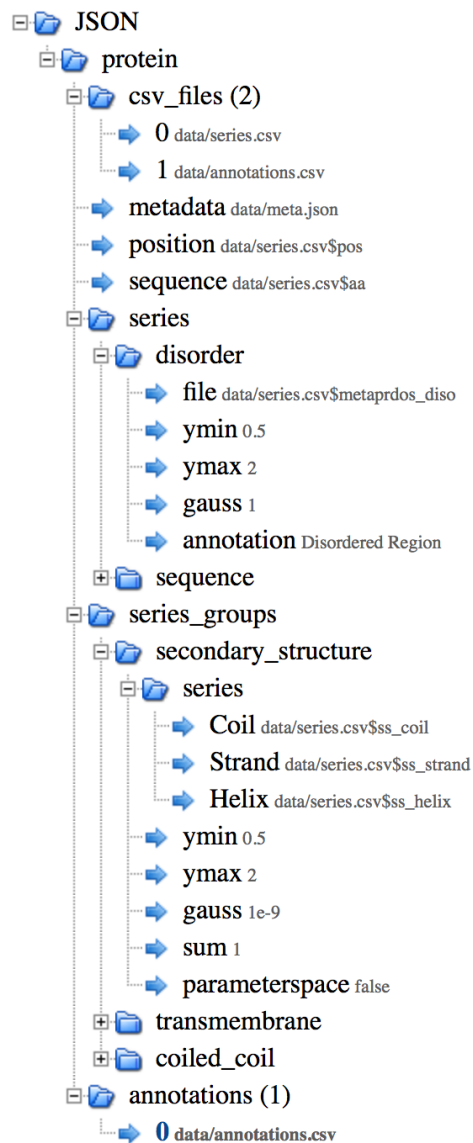


Figure 3: **Tree visualisation of JSON Schema** — Each protein object contains a list of required CSV files and a link to a protein metadata file. Position identifies a spreadsheet column which contains the amino acid position, and sequence identifies a spreadsheet column which contains a single-letter amino acid code for each position. Series are then defined, followed by series groups and finally annotations. Only one object of each type is shown expanded. Visualised using <http://www.jsontree.com>.

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