

Detection of cell assemblies with extracellular multi-electrode recordings

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Outline

Background

- Cell assemblies
- Spatiotemporal spike patterns in monkey motor cortex
- Cell-assembly structure and detectability

Model

- Model of the measurement setup
- Minimal assembly model
- Pattern statistics

Fitting procedure and results

Summary

Ressources

Cell assemblies

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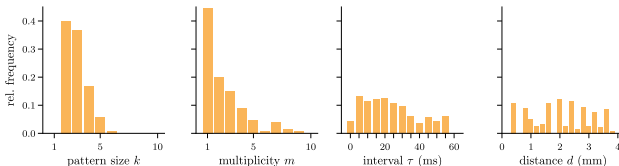
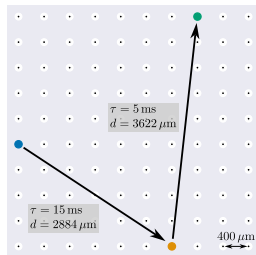
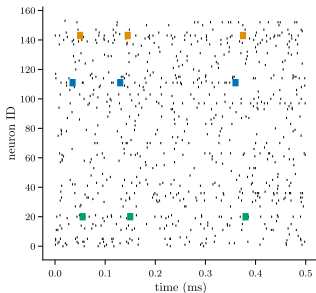
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 - neurons that reliably and recurrently generate spatio-temporal spike patterns with high temporal precision, such as neurons in a synfire chain (Abeles 1991) or in a braid network (polychronous patterns) (Bienenstock 1995; Izhikevich 2006)

Spatiotemporal spike patterns in monkey motor cortex

- single-unit spiking activity from reach-to-grasp experiment (Riehle et al. 2013)
- extracellular recordings with 10×10 Utah array, $400\mu\text{m}$ spacing
- identification of spatio-temporal patterns with millisecond precision by SPADE analysis

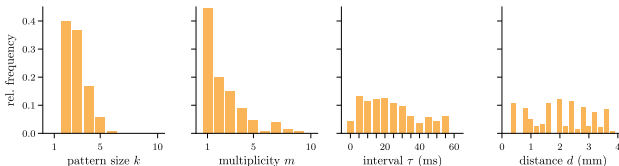
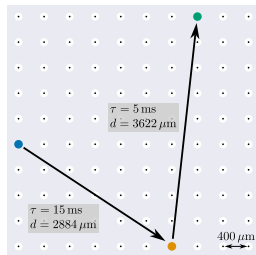
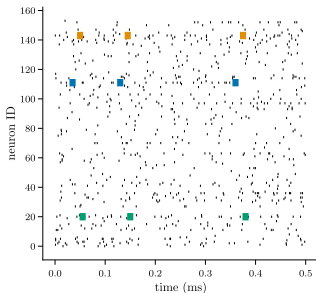
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Neuronal substrate generating such patterns? Spatiotemporal structure of these assemblies?

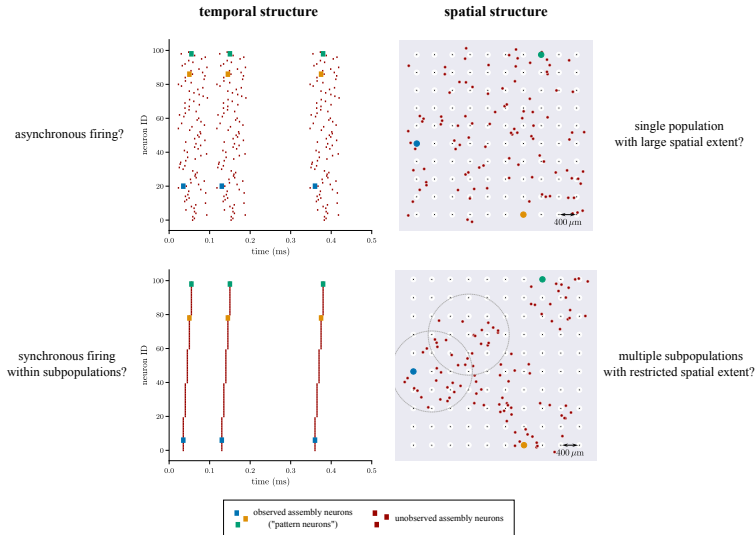
Spatiotemporal cell-assembly structure

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What does the rest of the iceberg look like?



Questions

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 - the recording constraints, and
 - the statistics of observed patterns?
- given a certain recording configuration (e.g., type/number of/distance between electrodes):
How likely is it to observe cell assemblies with a specific structure?
(not discussed in this talk)

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Model of the measurement setup

- total number of electrodes K
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- example: $K = 96$, $V = 4 \times 4 \times 1.5 \text{ mm}^3$, $U = 1.1$

$$q = \begin{cases} 0.0001 & \text{if } \rho = 35000 / \text{mm}^3 \\ 0.002 & \text{if } \rho = 2100 / \text{mm}^3 \end{cases}$$

Model

Minimal assembly model

- minimal model of spatial arrangement, size and number of assemblies
- no assumptions on network connectivity and dynamics

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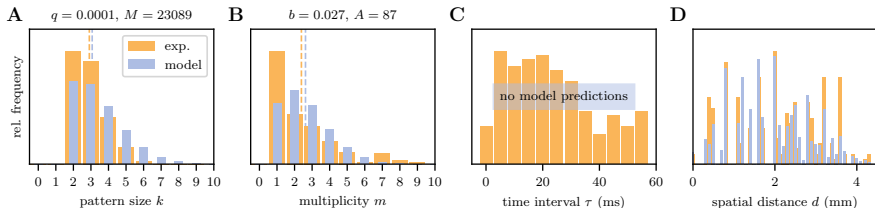
- minimal model of spatial arrangement, size and number of assemblies
- no assumptions on network connectivity and dynamics
- assumptions:
 - probed volume V contains A cell assemblies
 - each cell assembly composed of M neurons
 - assembly neurons are uniformly and independently distributed across V

Pattern statistics

- **pattern size k** : probability of detecting k neurons in a given assembly

$$p(k; q, M) = \binom{M}{k} q^k (1 - q)^{M-k}$$

with neuron-detection probability $q = KU/\rho V$ and assembly size M



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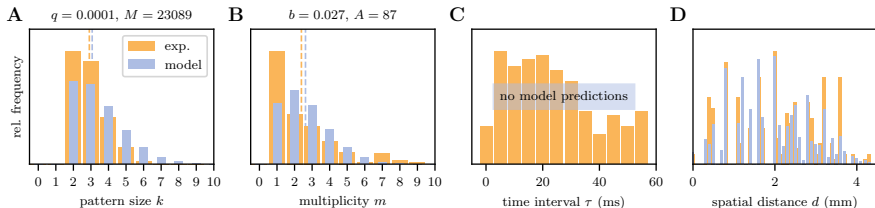
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- **membership multiplicity m** : probability of some neuron participating in m different assemblies

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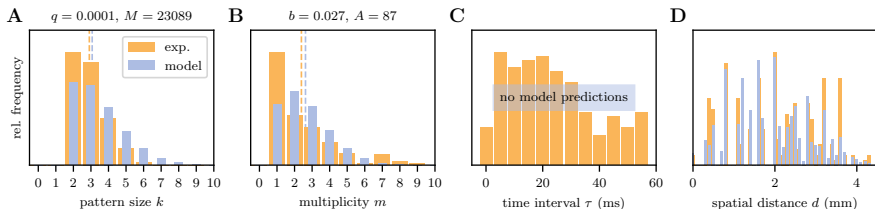
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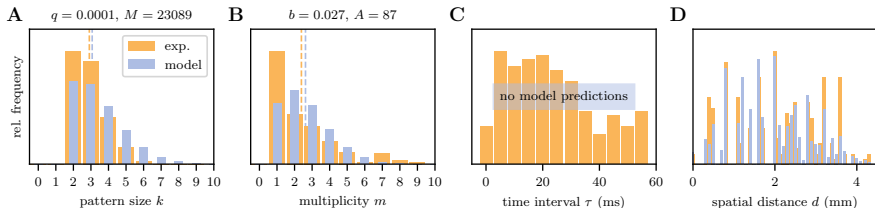
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- **pattern spike interval τ** : probability of observing time interval τ between consecutive pattern spikes
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- **pattern neuron distance d** : probability of Euclidean distance d between two pattern neurons
 - = frequency of inter-electrode distance d (independent + uniform neuron positions within observed volume)



Fitting procedure and results

- fix $q = KU/\rho V$ with $K = 96$, $U = 1.1$, $V = 4 \times 4 \times 1.5 \text{ mm}^3$, $\rho = 2100, \dots, 35000 \text{ mm}^{-3}$
- adjust model parameters M , $b = M/\rho V$ and A by maximizing sum of normalized model likelihoods, i.e., by minimizing cost function

$$E = -S_k^{-1} \sum_{i=1}^{S_k} \log [p(k_i; q, M)] - S_m^{-1} \sum_{j=1}^{S_m} \log [u(m_j; b, A)]$$

with model distributions $p(\cdot)$ and $u(\cdot)$, empirical pattern sizes and multiplicities k_i and m_j , and sample sizes S_k and S_m

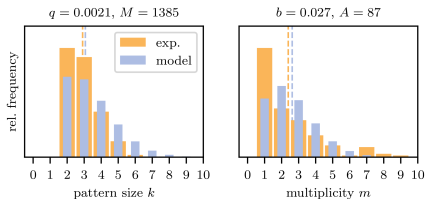
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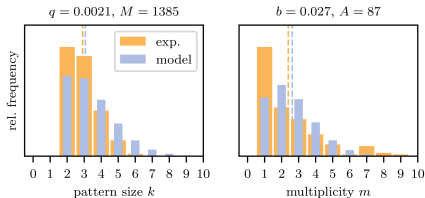
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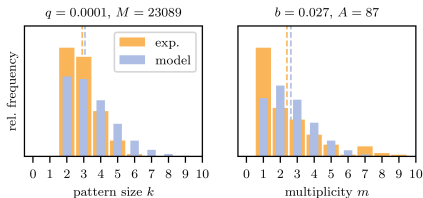
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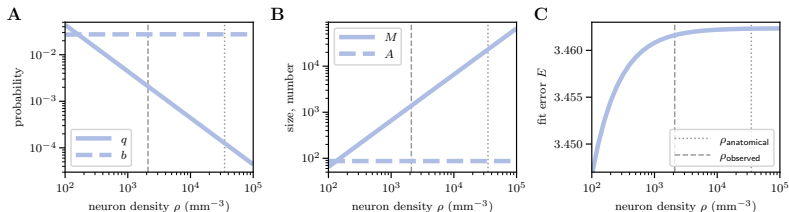
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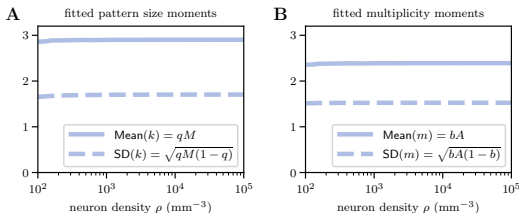
Fitting procedure and results

- best-fit assembly sizes M proportional to ρ , with little effect on fit error (same for V)
- best-fit assembly participation probability $b = 0.027$ and number of assemblies $A = 87$ independent of ρ

best-fit parameters and fit error:



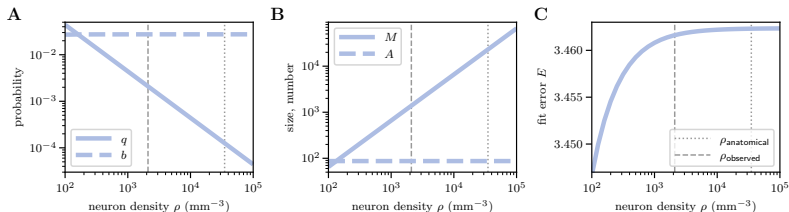
best-fit moments:



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best-fit parameters and fit error:



explanation: Poisson theorem

$$p(k; q, M) = \binom{M}{k} q^k (1 - q)^{M-k} \xrightarrow{q \rightarrow 0, Mq = \text{const.}} \frac{\lambda^k}{k!} e^{-\lambda} \quad \text{with} \quad \lambda = Mq$$

$$q = \frac{KU}{\rho V} \quad \curvearrowright \quad \lambda = Mq = \frac{MKU}{\rho V} \quad \curvearrowright \quad M = \frac{\rho V \lambda}{KU} \quad \curvearrowright \quad b = \frac{M}{\rho V} = \frac{\lambda}{KU}$$

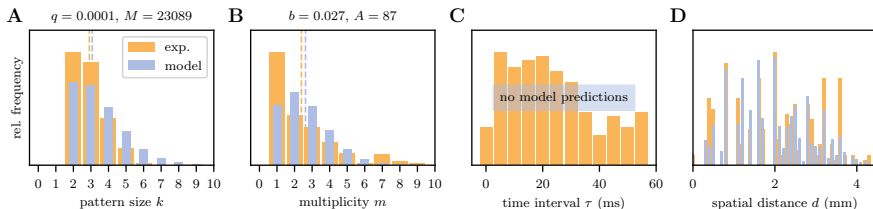
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 - many (~ 100) and
 - large cell assemblies containing $10^3 \dots 10^4$ neurons

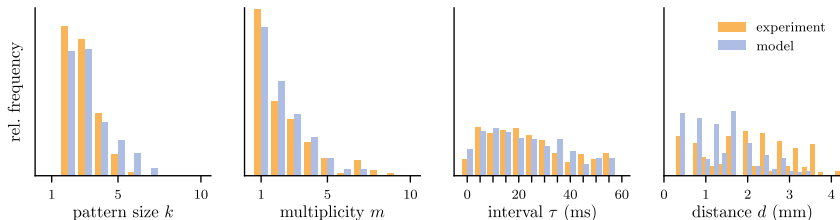
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 - many (~ 100) and
 - large cell assemblies containing $10^3 \dots 10^4$ neurons
- minimal assembly model and more complex synfire-chain model make similar predictions

minimal assembly model:



synfire-chain model:



Outlook

- include minimal model of spike timing (asynchronous firing of assembly neurons) to predict pattern spike interval distributions
- quantitative comparison between minimal assembly model and synfire-chain model (use same metrics for fit performance)

Resources

- **scientific tools:**

python, numpy, scipy, matplotlib

- **workflow tools:**

snakemake

- **project locations:**

https://github.com/INM-6/simulate_patterns_from_synfire_chains

https://github.com/INM-6/synfire_manuscript

- **data sources:**

pattern characteristics (pattern sizes, multiplicities, pattern spike intervals, pattern neuron distances)

https://github.com/INM-6/simulate_patterns_from_synfire_chains/blob/master/minimal_assembly_model/py/experimental_results.npy

obtained from reach-to-grasp data (Riehle et al. 2013)

data set: <https://doi.gin.g-node.org/10.12751/g-node.f83565>

metadata: https://github.com/INM-6/DataGrasp_Metadata

- **computing:**

laptop

Thanks

References I

- Abeles, Moshe (1991). **Corticonics: Neural Circuits of the Cerebral Cortex**. 1st edition. Cambridge: Cambridge University Press. DOI: 10.1017/CB09780511574566.
- (2011). “Cell assemblies”. In: **Scholarpedia** 6.7, p. 1505. DOI: 10.4249/scholarpedia.1505.
- Beul, Sarah F, Helen Barbas, and Claus C Hilgetag (2017). “A predictive structural model of the primate connectome”. In: **Scientific Reports** 7.43176, pp. 1–12. DOI: 10.1038/srep43176.
- Bienenstock, Elie (1995). “A model of neocortex”. In: **Network: Computation in neural systems** 6.2, pp. 179–224. DOI: 10.1088/0954-898X\6\2\004.
- Gallego, Juan A et al. (2017). “Neural manifolds for the control of movement”. In: **Neuron** 94.5, pp. 978–984.
- Hebb, D. O. (1949). **The organization of behavior: A neuropsychological theory**. New York: John Wiley & Sons. DOI: 10.1002/sce.37303405110.
- Henze, D.A. et al. (2000). “Intracellular features predicted by extracellular recordings in the hippocampus in vivo”. In: **Journal of Neurophysiology** 1.84, pp. 390–400. DOI: 10.1152/jn.2000.84.1.390.
- Hopfield, J. J. (1982). “Neural networks and physical systems with emergent collective computational abilities”. In: **Proceedings of the National Academy of Sciences of the United States of America** 79, pp. 2554–2558.
- Hubel, D. H. and T. N. Wiesel (1959). “Receptive fields of single neurones in the cat’s striate cortex”. In: **Journal of Physiology** 148, pp. 574–591.
- Izhikevich, Eugene M (2006). “Polychronization: computation with spikes”. In: **Neural Computation** 18.2, pp. 245–282. DOI: 10.1162/089976606775093882.
- Jaeger, H. (2001). **The “echo state” approach to analysing and training recurrent neural networks**. Tech. rep. GMD Report 148. St. Augustin, Germany: German National Research Center for Information Technology.

References II

- Jaeger, Herbert and Harald Haas (2004). "Harnessing nonlinearity: Predicting chaotic systems and saving energy in wireless communication". In: **Science** 304.5667, pp. 78–80.
- Maass, Wolfgang, Thomas Natschläger, and Henry Markram (2002). "Real-time computing without stable states: a new framework for neural computation based on perturbations". In: **Neural Computation** 14.11, pp. 2531–2560.
- Pettersen, Klas H. and Gaute T. Einevoll (2008). "Amplitude variability and extracellular low-pass filtering of neuronal spikes." In: **Biophysical Journal** 94.3, pp. 784–802. DOI: 10.1529/biophysj.107.111179.
- Riehle, Alexa et al. (2013). "Mapping the spatio-temporal structure of motor cortical LFP and spiking activities during reach-to-grasp movements". In: **Frontiers in Neural Circuits** 7, p. 48. DOI: 10.3389/fncir.2013.00048.
- Shoham, Shy, Daniel H O'Connor, and Ronen Segev (2006). "How silent is the brain: is there a "dark matter" problem in neuroscience?" In: **Journal of Comparative Physiology A** 192, pp. 777–784. DOI: 10.1007/s00359-006-0117-6.
- Stella, Alessandra et al. (2019). "3d-SPADE: Significance Evaluation of Spatio-Temporal Patterns of Various Temporal Extents". In: **Biosystems** 185, p. 104022. DOI: 10.1016/j.biosystems.2019.104022.
- Torre, Emiliano et al. (2013). "Statistical evaluation of synchronous spike patterns extracted by frequent item set mining". In: **Frontiers in computational neuroscience** 7, p. 132. DOI: 10.3389/fncom.2013.00132.