Practical work:

Small RNA sizes and differential expression



Task 1

Size distribution

Login - reminder

ssh ec-username@fox.educloud.no

- One-time password and password
- https://uio-in-biosx000.readthedocs.io/en/latest/Educloud/index.html
- https://www.uio.no/english/services/it/research/platforms/edu-research/help/fox/index.md
- Work on the interactive nodes.
- ssh int-<N> choose the one with the least load
- Files are availeble here:
 - /projects/ec34/in-biosx000/smallRNA

P.1 Size distribution

AIM: Visualize the size distribution (lengths) of RNAs from the small RNAseq data

- Data for this lecture are here:
 - /projects/ec34/in-biosx000/smallRNA/fastq
- Files
 - Sample10_clipped_single.fq
 - Sample11_clipped_single.fq
 - Sample12_clipped_single.fq
- The files are trimmed and uncompressed
- Choose one!



• The fastq format:

We have the sequence in every fourth line, starting with the second

- We can extract every fourth line using for example AWK
 - programming language designed for text processing
 - Awk Built-in Variables
 - Awk NR gives you the total number of records being processed or line number
 - awk 'NR%2==1' filename.fq prints every second line starting with the first in file.txt
- awk 'NR%4==2'

- We need the length of the sequence
 - awk length(string) calculates the length of a string
 - Length of what ?
 - AWK treats tab or whitespace for file separator by default
 - \$0 is the whole line, \$1 for first field...\$n for nth field
- awk '{if(NR%4==2) print length(\$0)}' Sample10_clipped_single.fq

Sort the length

• awk '{if(NR%4==2) print length(\$0)}' Sample10_clipped_single.fq|sort

Count the lengths

• awk '{if(NR%4==2) print length(\$0)}' Sample10_clipped_single.fq|sort |uniq -c

• Print to file

awk '{if(NR%4==2) print length(\$0)}' Sample10_clipped_single.fq|sort |uniq -c > home/Sample10_len.txt

Plot the size distributions



R

- Programming language
- Free software environment for statistical computing

- Long video introduction
 - https://www.youtube.com/watch?v=_V8eKsto3Ug&ab_channel=freeCodeCamp.org
- Short video introduction:
 - https://www.youtube.com/watch?v=SWxoJqTqo08&list=PLjgj6kdf_snYBkIsW QYcYtUZiDpam7ygg&ab_channel=DataCamp

R on Educloud

- https://www.uio.no/english/services/it/research/platforms/eduresearch/help/fox/installing-software-r.md
- module spider Bioconductor
- module load R-bundle-Bioconductor/3.9-foss-2019a-R-3.6.0
- R

Size distributions of small RNA seq data

- R
- setwd("PATH")
 - setwd("/fp/homes01/u01/ec-trinro/smallRNA/test1") # example
- ## Read the size distribution files

```
len<-read.table("filename.txt")
len</pre>
```

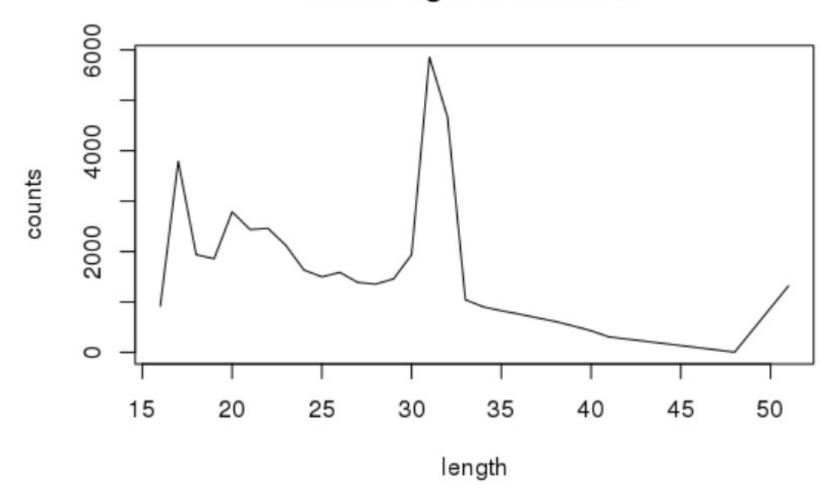
colnames(len)<-c("counts","lengths")

```
## In R - plot the distribution
plot(len$lengths, len$counts)
## with lines
plot(len$lengths, len$counts, type="l")
## make it nice
plot(len$lengths, len$counts, type="l", main="RNA length distribution",
xlab="length", ylab="counts")
```



```
## In R
# Open a pdf file
pdf("rplot.pdf")
# plot
plot(len$lengths, len$counts, type="l", main="RNA length distribution", xlab="length",
ylab="counts")
# Close the pdf file
dev.off()
```

RNA length distribution





Task 2

Differential expression



Task

AIM: Identify differential expressed circulating RNAs between serum samples from lung cancer and healthy individuals.

- The data for this task is here:
 - /work/IN-BIOSx/data/smallrna/de
- Select RNA class
 - miRNA
 - piRNA
 - tRF
- We will be using DEseq2 for differential expression analyses

Need help – new to R?

Take a look at the script is you are stuck:

 https://drive.google.com/file/d/18BqumBzN6BHjKWuh2yMTBXddK35 l2fEb/view?usp=sharing

R package

• Install packages you need (This takes time – so lets skip this)

```
# manually install Hmisc - old version due to issues
#install.packages("https://cran.r-project.org/src/contrib/Archive/Hmisc/Hmisc_3.9-3.tar.gz", repos = NULL,
type="source",dependencies=TRUE)
# https://bioconductor.org/packages/release/bioc/html/DESeq2.html
#if (!requireNamespace("BiocManager", quietly = TRUE)) install.packages("BiocManager")
```

- You will use libraries I already installed for you to avoid problems
- .libPaths("/work/IN-BIOSx/data/smallrna/Rlibs")
- Load the pachage you need
 - Use library() or require()
 - Load DEseq2



Manual and tutorial

DESeq2 vignett:

https://www.bioconductor.org/packages/devel/bioc/vignettes/DESeq 2/inst/doc/DESeq2.html

 DESeq2 manual: https://bioconductor.org/packages/release/bioc/manuals/DESeq2/man/DESeq2.pdf

Files

- Locate the small RNA files
- Set working directory
 - Use getwd() and setwd()
- Read in the count table
 - Use read.csv()
 - Be aware of separators and headers
- Read in the metadata table with case vs control
 - Use read.csv(), remember separators and headers

Check your data

• Use the str() function to check your data

 The differential expression analyses will only axcept int in the count tables

- Change rowname to RNA names and remove the RNA name column
 - Use rownames()
 - And remove column dataframe[-1]

Check your data

What is the average number of counts per RNA?

- rowMeans()
- plot(rowMeans())
- ## What is the average number of counts per sample?
- colMeans()
- plot(colMeans())

Filter your data

- To remove very low count RNAs
 - Reduce multiple testing
 - rowMeans
- Remove RNAs with mean counts less than 100
 - df[which(rowMeans(df)>100),]

Design the DE analyses

- Carefully set up your design variable
 - Use DESeqDataSetFromMatrix
 - Add
 - countData your filtered count data frame
 - colData the dataframe with the contrast groups
 - design ~design will contrast lung cancer cases vs controls

DE analyses

- Normalise and analyse the count file using DESeq2
 - DESeq()
 - This will take a bit of time
- Extract the results
 - results()
 - summary()

Identify RNAs that are DE

- # Extract the results with alpha (q value) less than 0.05 as a criteria for significance
 - res_05 <- results()
 - summary()

- # Extract significantly DE list and write them to a file
 - subset()
 - write.table()

Visulize the result

- Refer to DESeq2 manual for plot description
 - plotDispEsts(dds_process)
 - plotPCA(DESeqTransform(dds_process))
 - plotMA(dds_process)
 - sizeFactors(dds_process)

Extra: Evaluate your results

- Select one DE RNA
- What is the strenght of the result
- What is the biological meaning of the result
 - Use internett resourses such as
 - UCSC genome Browser, TargetScan, +++