Analyses of the high frequency time series on example of Circadian rhythms in the Long-Tail Pocket mouse.

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Abstract

The present study aimed to examine the forecasting performance of various univariate approaches to forecast high-frequency time series which contains more than one seasonal pattern that must be taken into consideration.

To achieve the purpose of the study, we performed the following procedures with major findings. First, we investigated different frequencies and produced traditional forecasts for it using ts object. The limitation of ts objects for only low frequency data and also for using multiple seasonalities led us to make additional sets of forecasts by applying non-benchmark approaches with double seasonality using msts object with Fourier terms for each seasonal pattern. This indicates that forecast accuracy was able to be improved by incorporating two different types of seasonal patterns simultaneously.

Seasonality is the main component of time series, and the consideration of seasonality has become more important with the increasing frequency of time series produced in industry. Circadian rhythms in the Long-Tail Pocket mouse time series is a representative example of business time series data that has been collected every two minutes for a long period of time. The high frequency data, with time series containing closely spaced time intervals creates a necessity of creating accurate modeling approaches for modeling seasonal patterns of time series.

As each seasonal pattern has distinct periods and effects, it is not trivial to design a model that can capture multiple seasonal patterns at once. We found that fourier regression gives more accurate results and can detect multiple seasonalities better then etc and thats and sarima.

Introduction

The investigating dataset is a temperature recording made at a 2 minutes interval for 83 days on a nocturnal mammal. The Long-tailed pocket mouse live in the South West of the United States and Northern Mexico. The data represents interest because it does contain periodicities in the behaviour of the animals in regards to circadian rhythm. The Circadian is the reaction to light and darkness of their environment. The experiments used separate equipment for each mammal to monitor the environment and so data for each animal could be varied. The data given here are the telemeter frequency temperature recordings for one Long-tailed pocket mouse from the first experiment, with higher counts indicating higher temperature. Since the interest was in periodicity, no effort was made to relate the telemeter frequency to actual temperature. The file that was collected had 59616 observations that correspond to the 83 days. The proportion of outliers in the original data has been 8% and outliers were removed.

Typically, multiple seasonality patterns are more likely to occur whenever the series has data with high frequency (for example, daily, hourly, half-hourly, and so on), as there are more options to aggregate the series to a lower frequency. This data is a typical example of multiple seasonality, which could have multiple seasonal patterns, as the observations taken every two minutes of the hours of the day, the day of the week. On the other hand, as the frequency of the series is lower (for example, monthly, quarterly, and so on), it is more likely to have only one dominant seasonal pattern as opposed to a high frequency series, as there are fewer aggregation options for another type of frequencies. Seven last days of data for investigation were selected.

Model Specification

7days of observations = 30(in one hour) * 24(hours) * 7(days) = 5040 observations

Having more observations per cycle unit, that is, high-frequency time series data, could potentially provide more insightful information about the series behavior as opposed

to a lower frequency time series data. However, this comes with the price of additional complexity, which therefore requires more effort in the analysis process.

Potentially, as mentioned previously, the series can have three different seasonal patterns.

```
1 week = 30*24 *7 =5040

Daily 30 * 24 = 720

Hourly = 30

2 hourly = 30*2=60

6 hourly = 30*6 = 180

10 hourly = 30 *10 = 330
```

Time series objects allow maximum frequency = 350 and therefore we will use hourly seasonality with frequency = 30 and 10 hourly with frequency = 300. For multiple seasonalities we have to use ms object instead of ts object.

Original data

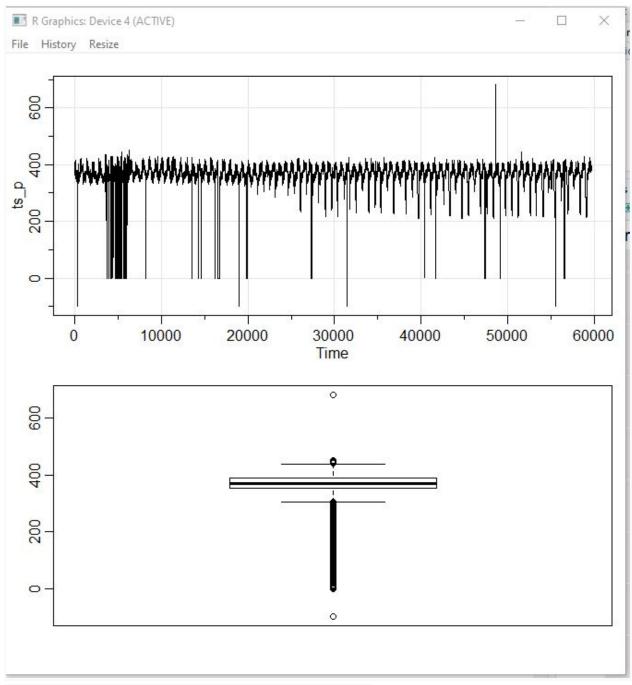
```
library(astsa)
library("forecast")
library("lubridate")
examine.mod <- function(mod.fit.obj, p, d, q, P=0, D=0, Q=0, S=-1, lag.max=24) {
dev.new(width=6, height=6)
par(mfrow=c(2,1))
pacf(mod.fit.obj$fit$residuals, main="PACF of Residuals", lag.max)
if ((P==0)&(D==0)&(Q==0)) {
 title(paste("Model: (", p, ",", d, ",", q, ")", sep=""), adj=0, cex.main=0.75)
}
else {
 title(paste("Model: (", p, ",", d, ",", q, ") (", P, ",", D, ",", Q, ") [", S, "]", sep=""), adj=0, cex.main=0.75)
}
std.resid <- mod.fit.obj$fit$residuals/sqrt(mod.fit.obj$fit$sigma2)
hist(std.resid, main="Histogram of Standardized Residuals", xlab="Standardized Residuals",
freq=FALSE)
curve(expr=dnorm(x, mean=mean(std.resid), sd=sd(std.resid)), col="red", add=TRUE)
```

pformosu<-read.table(file = "C:/Users/inna/Desktop/DepaulClasses/ApplyMathClasses/Time_Series/Final Project/pformosu.txt") row =3726 col= 16 pformosu_dat = numeric(col*row) count = 1 for(i in 1:row) { for(j in 1:col) { pformosu_dat[count]=pformosu[i,j] count = count + 1 } } ts_p<-ts(pformosu_dat) ength(ts_p)

dev.new()

tsplot(ts_p)
boxplot(ts_p)\$out

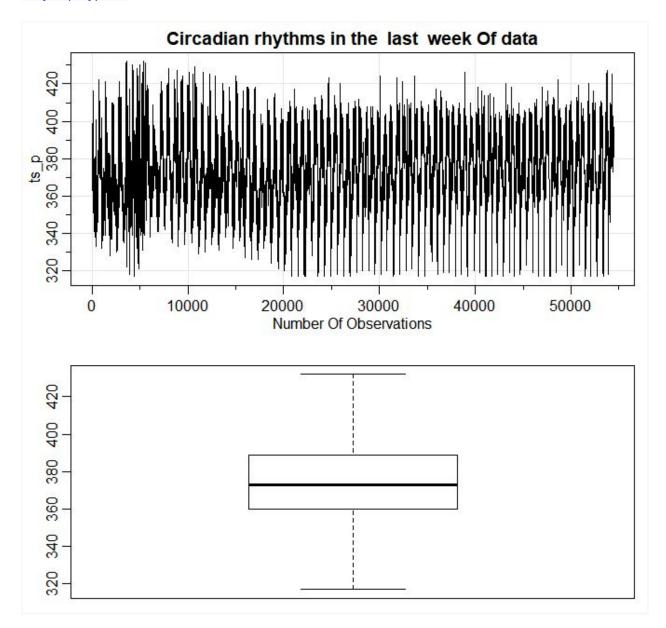
par(mfrow=c(2,1))



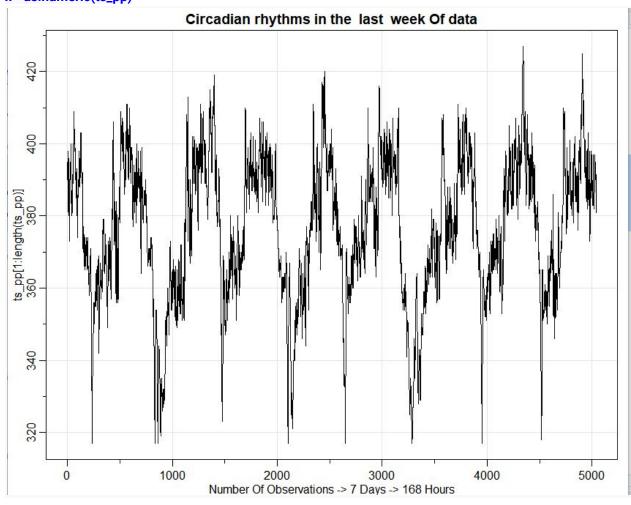
Removing outliers.

```
num_of_outliers = 0
repeat
{
  outliers <- boxplot(ts_p, plot=FALSE)$out
  v<-(which(ts_p %in% outliers))
  if(length(v)== 0)
  break</pre>
```

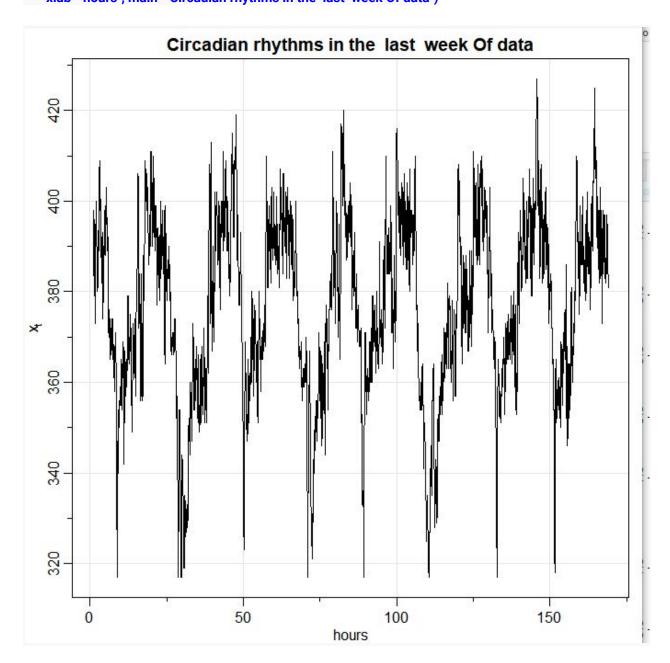
```
else
{
    ts_p<-ts_p[-v]
    num_of_outliers = num_of_outliers + length(v)
}
print(paste("Number Of Outliers removed = ", num_of_outliers))
print(paste("Length Of Time Series",length(ts_p)))
dev.new()
par(mfrow=c(2,1))
tsplot(ts_p,xlab="Number Of Observations",main="Circadian rhythms in the last week Of data")
boxplot(ts_p)$out</pre>
```



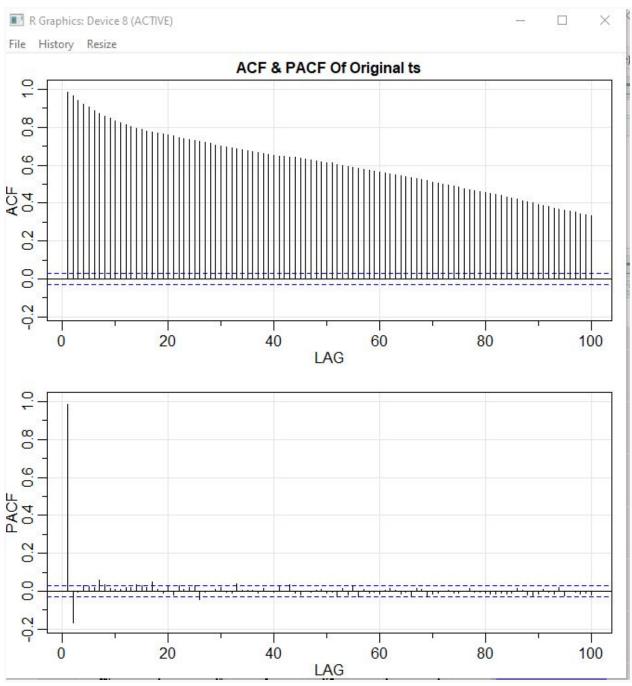
Getting one week of data and plotting it.



dev.new()
tsplot(ts_pp, ylab=expression(x[t]),
 xlab="hours", main="Circadian rhythms in the last week Of data")



dev.new()
acf2(x, max.lag = length(ts_pp) - 1,main = "ACF & PACF Of Original ts")

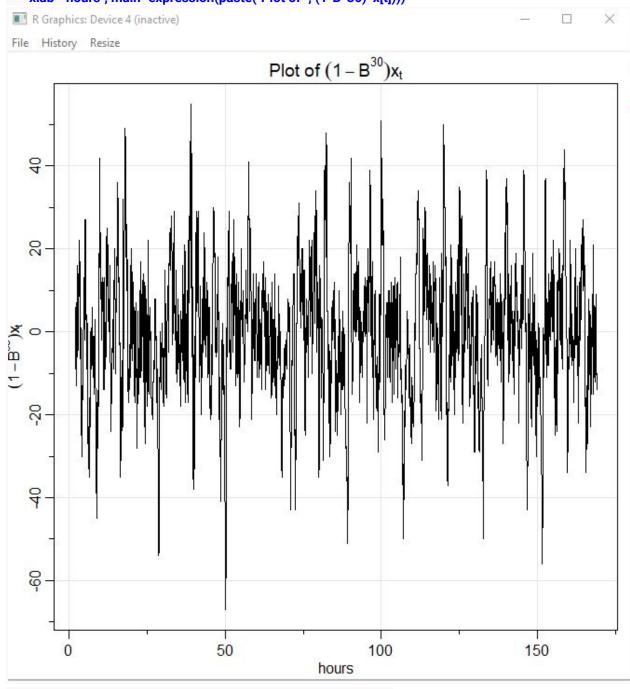


We can see from the plot that ACF is decaying exponentially and PACF has 2 lags but we have to differentiate to get rid of seasonalities (1- B^30)

Plot of (1-B^30)*x_t

dev.new()

tsplot(diff(ts_pp, lag=frequency, differences=1), ylab=expression((1-B^30)*x[t]), xlab="hours", main=expression(paste("Plot of ", (1-B^30)*x[t])))

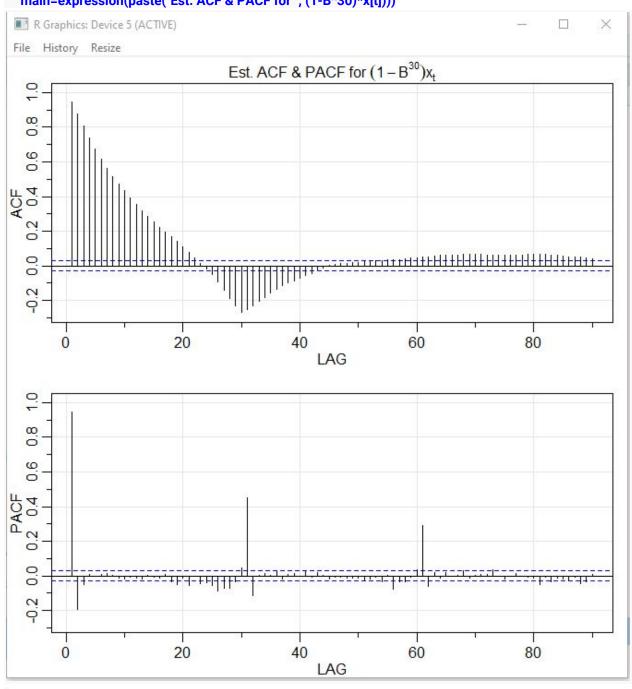


ACF indicated cut of after 30 lag that suggests that ARIMA Q=1,

PACF exponentially decay that suggests ARIMA P=0

Diff =1 S=18 (1-B^30)*x_t

dev.new()
acf2(diff(x, lag=frequency, differences=1), max.lag=frequency*3,
 main=expression(paste("Est. ACF & PACF for ", (1-B^30)*x[t])))

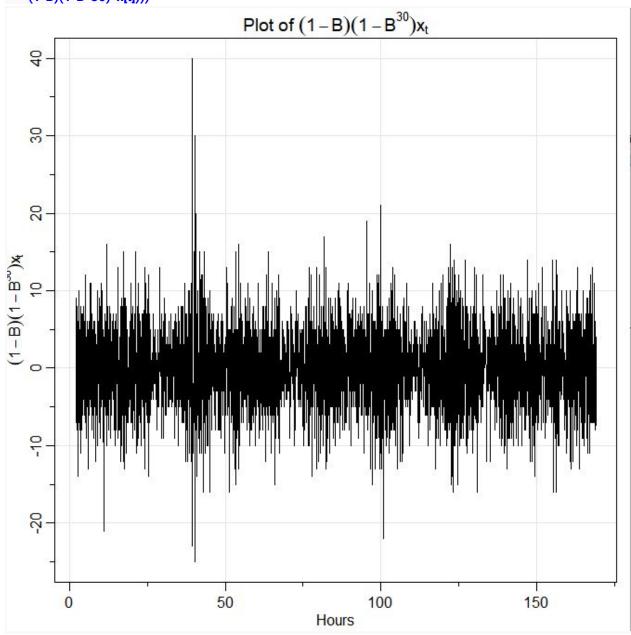


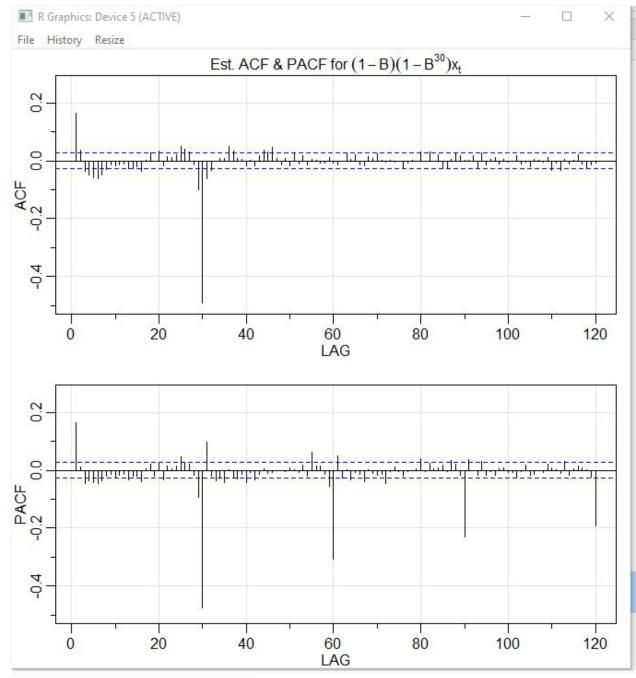
PACF exponentially decaying at lags 30,60 indicating that P = 0

ACF seasonal pick at Lag = 30 and no other seasonal picks at lags 60, 90 Indication that Q = 1, giving us the initial estimation for ARIMA(),(0,1,1)30. But graph has some little trend so we have to remove the trend using non seasonal difference (1-B)

```
ev.new()
```

```
tsplot(diff(diff(ts_pp, lag=frequency, differences=1)),
ylab=expression((1-B)(1-B^30)*x[t]),
xlab="Hours", main=expression(paste("Plot of ",
(1-B)(1-B^30)*x[t])))
```



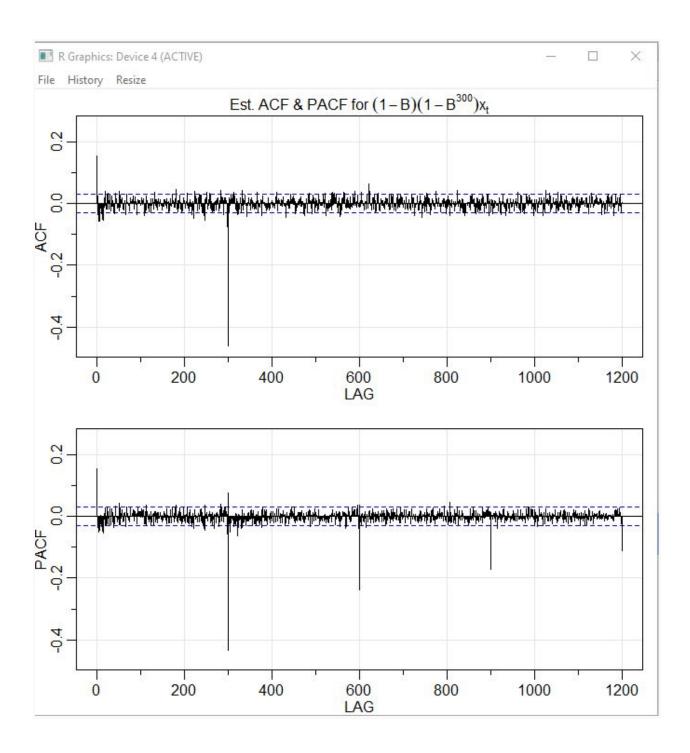


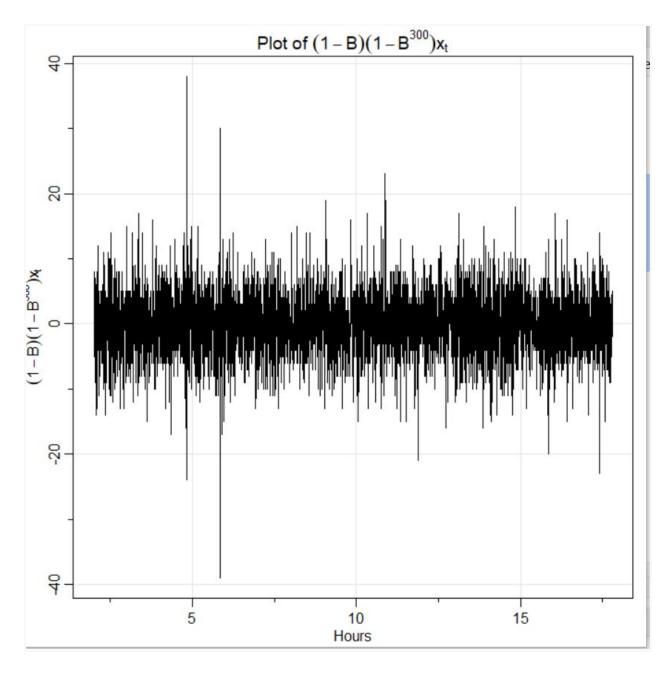
ACF seasonal Q = 1, non-seasonal q = 1-5

PACF decaying at lags 30,60,90, and clusters around each lag indication that seasonal P = 0, nonseasonal p = 1-5

ARIMA(p=(0-5),d=1,q=(0-5)),(P=0,D=1,Q=1)[30]

Trying ACF frequency = 300





ACF shows seasonal spike at lag 300 and non-seasonal at some cluster around leg 1 PACF shows seasonal exponential decay at 300,600 .. and also cluster around lag one Also indicating ARIMA(p=(0-5),d=1,q=(0-5)),(P=0,D=1,Q=1)[300] ARIMA(p=(0-5),d=1,q=(0-5),P=0,D=1,Q=1,S=30)

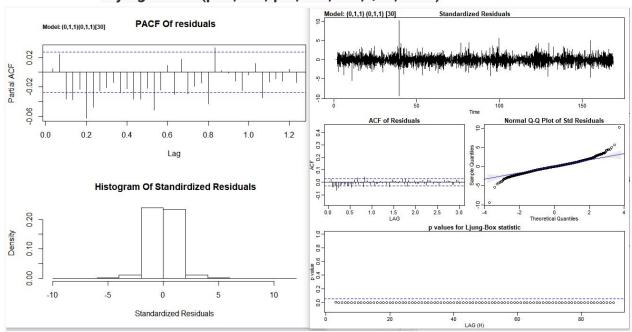
Fitting and Diagnostics

Trying ARIMA(p=(0),d=1,q=(0),P=0,D=1,Q=1,S=30)

```
dev.new()
mod.fit1<- sarima(ts_pp,p=0,d=1,q=0,P=0,D=1,Q=1,S=30)
mod.fit1
dev.new()
par(mfrow=c(2,1))
pacf(mod.fit1$fit$residuals,main = "PACF Of residuals")
title(paste("Model: (",
        p = 0, ",",
        d = 1, ",",
        q = 0, ")(",
        P = 0, ",",
       D = 1, ",",
        Q = 1, ")[30]",
        sep=""),adj=0,cex.main=0.75)
std.resid1 <- mod.fit1$fit$residuals / sqrt(mod.fit1$fit$sigma2)
hist(std.resid1,main = "Histogram Of Standardized Residuals",
   xlab='Standardized Residuals',
   freq = FALSE
                                                                                            Standardized Residuals
                                                                     Model: (0,1,0) (0,1,1) [30]
                          PACF Of residuals
       Model: (0,1,0)(0,1,1)[30]
Partial ACF
    0.05
    -0.05
                                                                                                         Normal Q-Q Plot of Std Residuals
       0.0
               02
                        0.4
                                06
                                        0.8
                                                1.0
                                                        1.2
                                                                  4.
                                 Lag
                                                                  O.2
                  Histogram Of Standirdized Residuals
    0.3
    0.2
                                                                  0.8
    0.1
                                                                  p value
0.4 0.6
    0.0
                 -5
                              0
                                                        10
                          Standardized Residuals
```

```
$'fit'
call:
stats::arima(x = xdata, order = c(p, d, q), seasonal = list(order = c(p, D,
    Q), period = S), include.mean = !no.constant, transform.pars = trans, fixed = fixed,
    optim.control = list(trace = trc, REPORT = 1, reltol = tol))
Coefficients:
            sma1
        -1.0000
s.e. 0.0152
sigma^2 estimated as 12.15: log likelihood = -13438.07, aic = 26880.13
$degrees_of_freedom
[1] 5008
$ttable
     Estimate SE t.value p.value
L -1 0.0152 -65.6435 0
$AIC
[1] 5.335477
$AICC
[1] 5.335477
$BIC
[1] 5.338065
```

Trying ARIMA(p=0,d=1,q=1,P=0,D=1,Q=1,S=30)

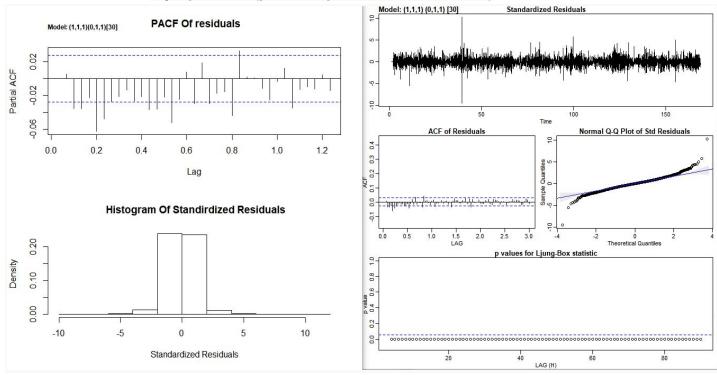


```
> mod.fit1
$`fit'
call:
coefficients:
       ma1
             sma1
    0.1538
           -1.0000
    0.0135
            0.0126
sigma^2 estimated as 11.85: log likelihood = -13376.12, aic = 26758.24
$degrees_of_freedom
[1] 5007
$ttable
              SE t.value p.value
    Estimate
     0.1538 0.0135 11.3692
ma1
    -1.0000 0.0126 -79.6657
sma1
$AIC
[1] 5.311282
$AICC
[1] 5.311282
$BIC
[1] 5.315164
```

ARIMA(p=0,d=1,q=1,P=0,D=1,Q=1,S=30)

model seems like better that the previous because BIC = 5.315164 compare to ARIMA(p=0,d=1,q=0,P=0,D=1,Q=1,S=30) BIC = 5.338065

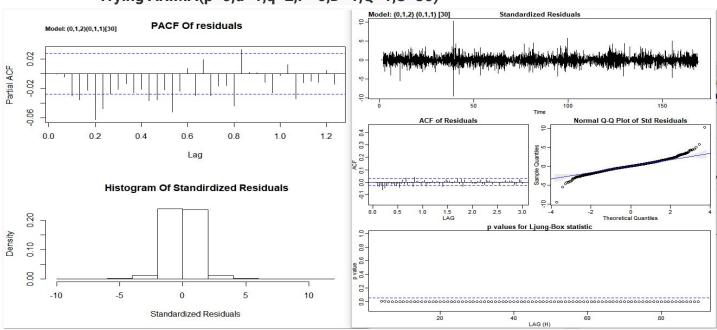
Trying ARIMA(p=1,d=1,q=1,P=0,D=1,Q=1,S=30)



```
> mod.fit1
$`fit`
call:
stats::arima(x = xdata, order = c(p, d, q), seasonal = list(order = c(P, D,
Q), period = S), include.mean = !no.constant, transform.pars = trans, fixed = fixed,
optim.control = list(trace = trc, REPORT = 1, reltol = tol))
Coefficients:
                      ma1
                                sma1
            ar1
        0.1370
                 0.0219
                             -1.0000
                             0.0129
       0.0727 0.0730
sigma^2 estimated as 11.84: log likelihood = -13374.49, aic = 26756.98
$degrees_of_freedom
[1] 5006
$ttable
      Estimate
                            t.value p.value
                        SE
ar1
         0.1370 0.0727
                              1.8833
ma1
         0.0219 0.0730
                              0.2995
                                         0.7646
sma1
       -1.0000 0.0129 -77.7387
$AIC
[1] 5.311032
$AICC
[1] 5.311033
[1] 5.316208
```

The ARIMA(p=1,d=1,q=1,P=0,D=1,Q=1,S=30) model has p-value for ma1 > 0.05 and this makes ARIMA(p=0,d=1,q=1,P=0,D=1,Q=1,S=30) is better out of all previous models

Trying ARIMA(p=0,d=1,q=2,P=0,D=1,Q=1,S=30)

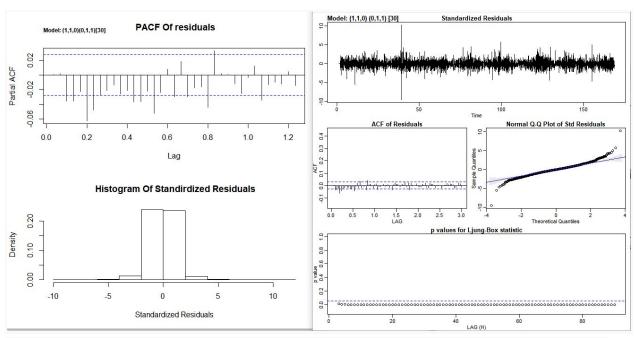


```
Coefficients:
         ma1
                 ma2
                        sma1
      0.1599
              0.0320
                      -1.000
s.e. 0.0142
              0.0147
                       0.013
sigma^2 estimated as 11.84: log likelihood = -13373.75, aic = 26755.5
$degrees_of_freedom
[1] 5006
$ttable
                  SE t.value p.value
     Estimate
       0.1599 0.0142
                      11.2785
ma1
                               0.0000
ma2
       0.0320 0.0147
                       2.1813
                               0.0292
      -1.0000 0.0130 -76.7035
                               0.0000
$AIC
[1] 5.310738
$AICc
[1] 5.310739
$BIC
[1] 5.315914
```

The ARIMA(p=0,d=1,q=2,P=0,D=1,Q=1,S=30) model has aal p-value for ma1 < 0.05 and this makes And BIC=5.315914 $ARIMA(p=0,d=1,q=1,P=0,D=1,Q=1,S=30) \ \, BIC=5.315164 < 5.315914 = BIC \\ ARIMA(p=0,d=1,q=2,P=0,D=1,Q=1,S=30) \ \, which makes the best out of all previous models \\ ARIMA(p=0,d=1,q=1,P=0,D=1,Q=1,S=30)$

When Trying ARIMA(p=0,d=1,q=(3 or 4),P=0,D=1,Q=1,S=30) values of BIC is increasing and therefore the best model now is still ARIMA(p=0,d=1,q=1,P=0,D=1,Q=1,S=30)

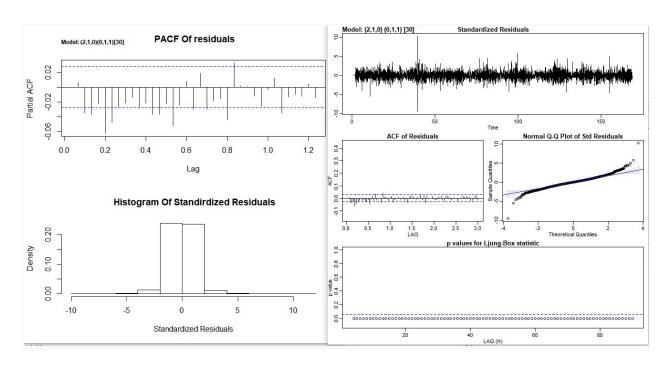
Trying ARIMA(p=1,d=1,q=0,P=0,D=1,Q=1,S=30)



```
Coefficients:
         ar1
              -1.0000
      0.1583
               0.0129
      0.0139
s.e.
sigma^2 estimated as 11.84: log likelihood = -13374.53, aic = 26755.07
$degrees_of_freedom
[1] 5007
$ttable
                  SE t.value p.value
     Estimate
       0.1583 0.0139 11.3460
ar1
                                     0
      -1.0000 0.0129 -77.3007
                                     0
sma1
$AIC
[1] 5.310653
$AICC
[1] 5.310653
$BIC
[1] 5.314535
```

The ARIMA(p=1,d=1,q=0,P=0,D=1,Q=1,S=30) model has all p-value for ma1 < 0.05 and And BIC=5.314535 < BIC = 5.315164 of ARIMA(p=0,d=1,q=1,P=0,D=1,Q=1,S=30) This makes ARIMA(p=1,d=1,q=0,P=0,D=1,Q=1,S=30) better model

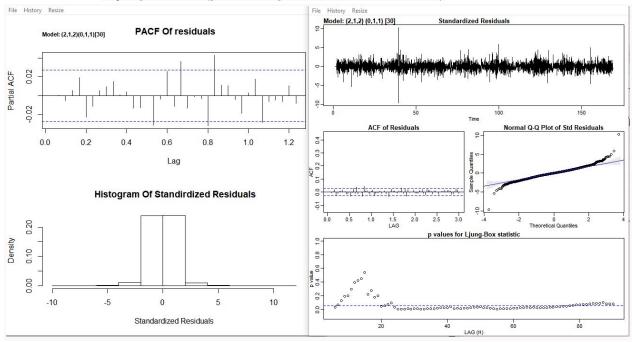
Trying ARIMA(p=2,d=1,q=0,P=0,D=1,Q=1,S=30)



```
$ttable
     Estimate
                  SE t.value p.value
ar1
       0.1591 0.0141
                     11.2615
                               0.0000
ar2
      -0.0052 0.0141
                     -0.3683
                               0.7127
sma1
     -1.0000 0.0128 -77.9575
                              0.0000
$AIC
[1] 5.311023
$AICC
[1] 5.311024
$BIC
[1] 5.316199
> dev.new()
```

Ar2 pvalue >0.05 which makes the ARIMA(p=1,d=1,q=0,P=0,D=1,Q=1,S=30) better model

Trying ARIMA(p=2,d=1,q=2,P=0,D=1,Q=1,S=30)



```
coefficients:
                    ar2
                                       ma2
                                                sma1
                              ma1
         ar1
      1.3022
               -0.3833
                          -1.1594
                                   0.2056
                                             -1.0000
      0.0682
                0.0654
                          0.0722
                                   0.0707
sigma^2 estimated as 11.64: log likelihood = -13332.36, aic = 26676.72
$degrees_of_freedom
[1] 5004
$ttable
     Estimate
                    SE
                        t.value p.value
       1.3022 0.0682
ar1
                        19.0957
                                   0.0000
       -0.3833 0.0654
                         -5.8623
                                   0.0000
      -1.1594 0.0722 -16.0541
                                   0.0000
ma2
      0.2056 0.0707 2.9097
-1.0000 0.0102 -97.6950
                                   0.0036
                                  0.0000
sma1
[1] 5.295102
$AICc
[1] 5.295104
[1] 5.302866
```

p values <0.05 which makes the ARIMA(p=2,d=1,q=2,P=0,D=1,Q=1,S=30) BIC =5.302866 < BIC=5.314535 ARIMA(p=1,d=1,q=0,P=0,D=1,Q=1,S=30) which makes ARIMA(p=2,d=1,q=2,P=0,D=1,Q=1,S=30) better than all above models

```
Trying ARIMA(p=3,d=1,q=3,P=0,D=1,Q=1,S=30)

Trying ARIMA(p=3,d=1,q=2,P=0,D=1,Q=1,S=30)

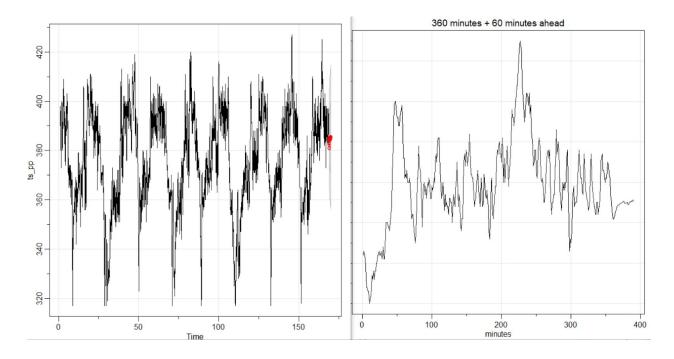
ARIMA(p=2,d=1,q=3,P=0,D=1,Q=1,S=30)

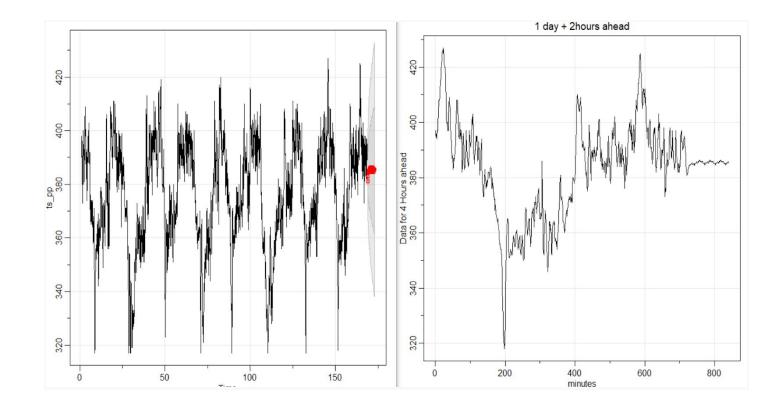
P-values >0.05 and therefore the best selected model is

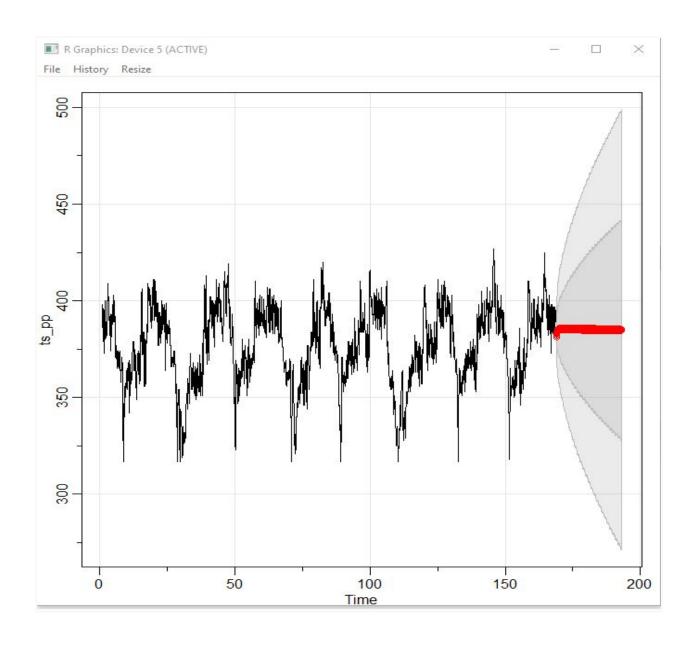
ARIMA(p=2,d=1,q=2,P=0,D=1,Q=1,S=30)
```

Forecasting

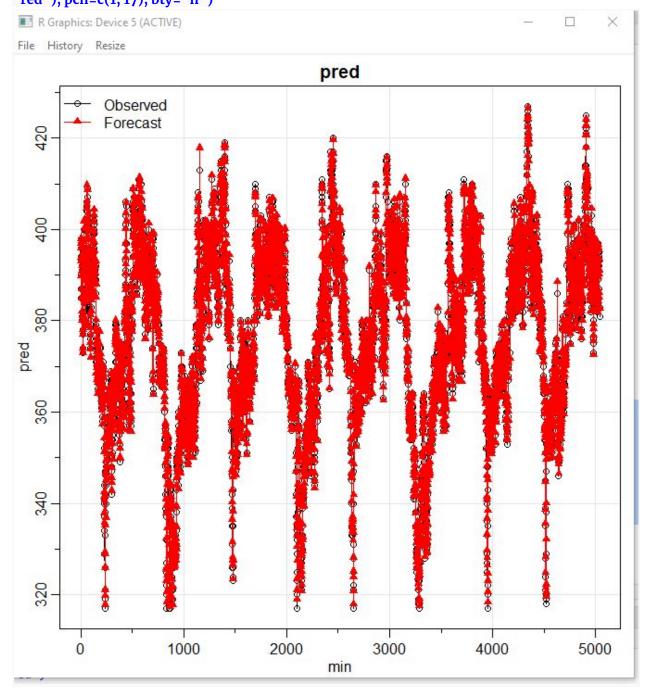
ahead_1_hours <-30 dev.new() fore.mod <- sarima.for(ts_pp, n.ahead=ahead_1_hours, p=2, d=1, q=2, P=0, D=1, Q=1, S=30, plot.all=TRUE)





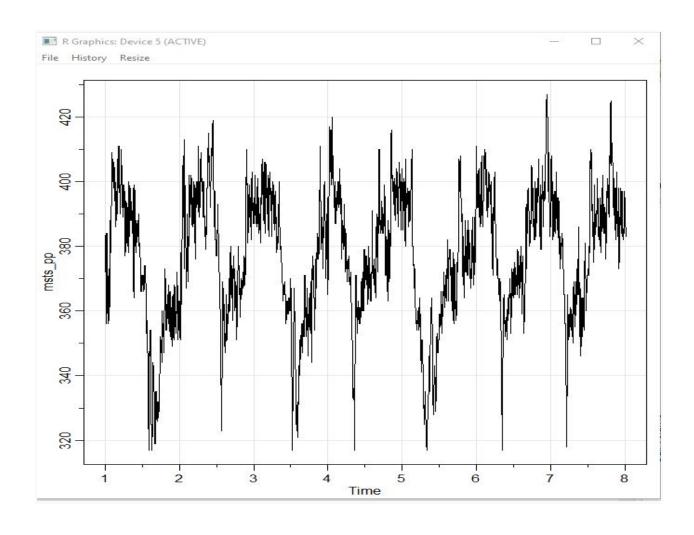


```
pred.mod <- ts(x - mod.fit.110.011$fit$residuals)
dev.new()
tsplot(ts_pp[1:length(ts_pp)], ylab="pred", xlab="min", type="o", main="pred")
lines(pred.mod, col="red", type="o", pch=17)
legend("topleft", legend=c("Observed", "Forecast"), lty=c("solid", "solid"), col=c("black", "red"), pch=c(1, 17), bty="n")</pre>
```



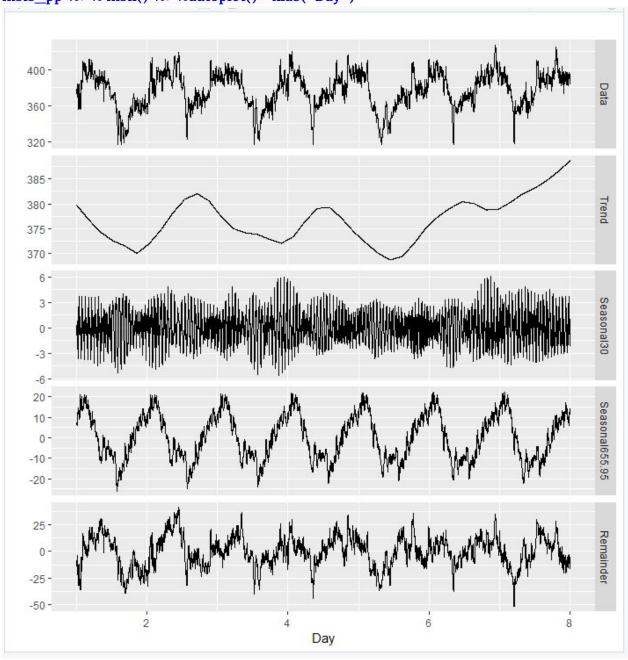
Discussion

```
On_hour <- 30
one_day <- 30*24
one_week <- 30*24*7
frequency <- c(30,one_day)
msts_pp <- msts(ts_p[(length(ts_p)-(one_week) + 1):(length(ts_p))],seasonal.periods = frequency)
dev.new()tsplot(msts_pp)
```



The $_{\tt mstl}$ () function is a variation on $_{\tt stl}$ () designed to deal with multiple seasonality. It will return multiple seasonal components, as well as a trend and remainder component.

msts_pp %>% mstl() %>%autoplot() + xlab("Day")



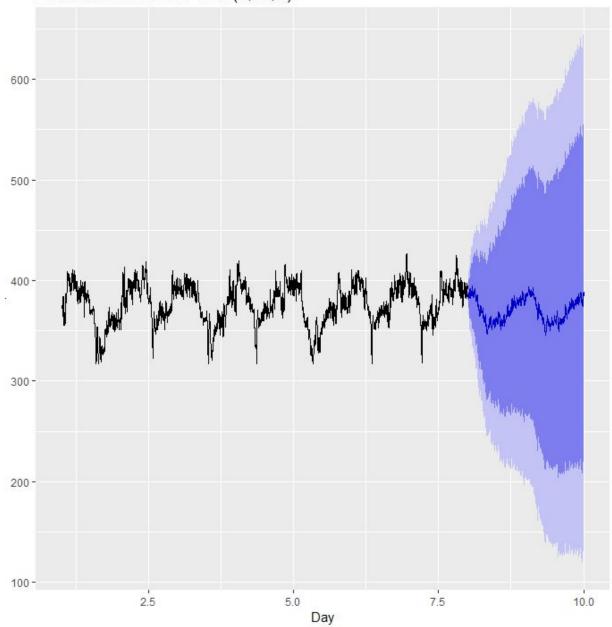
There are two seasonal patterns shown, one for the time of minutes in hour (the third panel), and one for the time of day (the fourth panel). To properly interpret this graph, it is important to notice the vertical scales. In this case,

the trend and the daily seasonality have relatively narrow ranges compared to the other components, because there is little trend seen in the data, and the daily seasonality is weak.

The decomposition can also be used in forecasting, with each of the seasonal components forecast using a seasonal naïve method, and the seasonally adjusted data forecasting using ETS (or some other user-specified method). The stlf() function will do this automatically.

msts_pp %>% stlf() %>%
autoplot() + xlab("Day")

Forecasts from STL + ETS(A,Ad,N)



Dynamic harmonic regression with multiple seasonal periods

Because there are multiple seasonalities, we need to add Fourier terms for each seasonal period. In this case, the seasonal periods are 30 and 656, so the Fourier terms are of the form

```
\sin(2\pi kt/30), \cos(2\pi kt/30), \sin(2\pi kt/656), \cos(2\pi kt/656)
```

For k = 1,2, ...

The fourier() function can generate these for you.

We will fit a dynamic harmonic regression model with an ARMA error structure. The total number of Fourier terms for each seasonal period have been chosen to minimise the AICc. We will use a log transformation (lambda=0) to ensure the forecasts and prediction intervals remain positive.

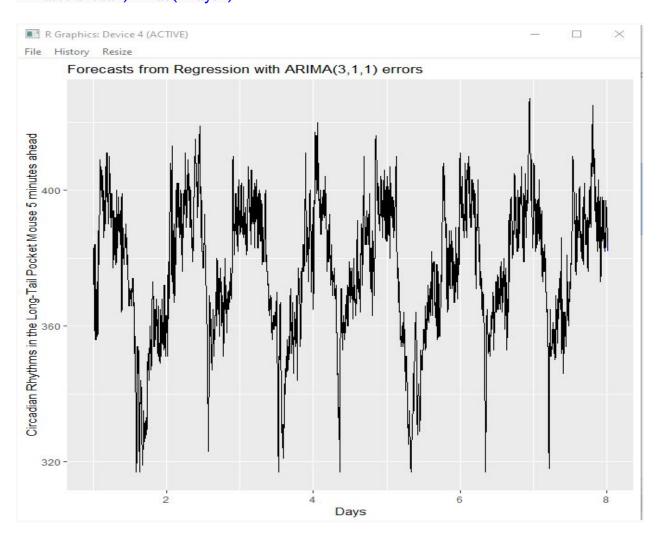
fit <- auto.arima(msts_pp, seasonal=FALSE, lambda=0,xreg=fourier(msts_pp, K=c(14,65))) fit

```
Coefficients:
                         ar1 ar2 ar3 mal s1-30 c1-30 s2-30 c2-30 s3-30 c3-30 s4-30 c4-30 s5-30 c5-30 s6-30 s6-30 s1-30 s1
                                                                                                                                                                                                                                                                                                                                                                                                                                                             2e-04 1e-04 0e+00 2e-04 -3e-04
s.e. 0.0187
                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                     -2e-04 0.0069 0.0343 0.0034
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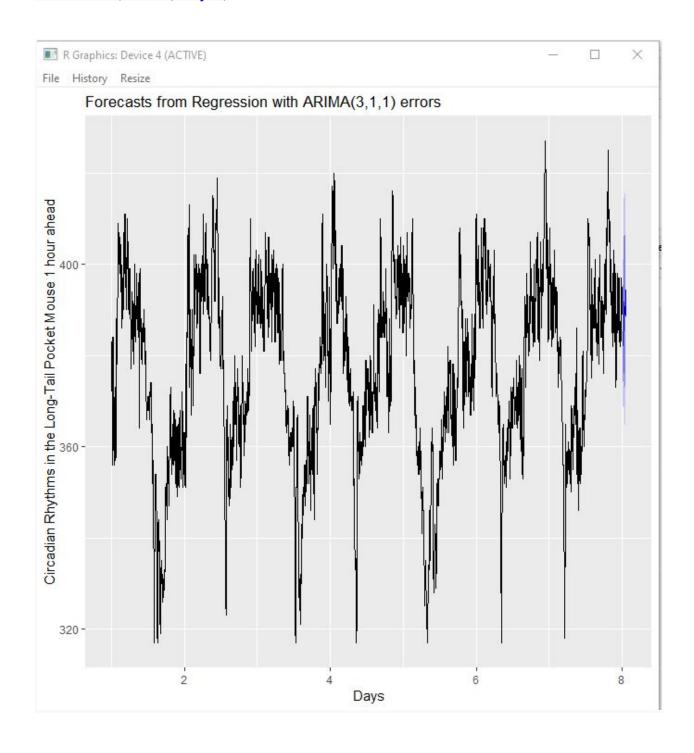
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                              1e-04
                                                                             -1e-04
                                                          log likelihood=15190.22
BIC=-29006.08
sigma^2 estimated as 8.103e-05:
AIC=-30054.44 AICC=-30042.36
```

dev.new() minutes_10<- 5 fit %>%

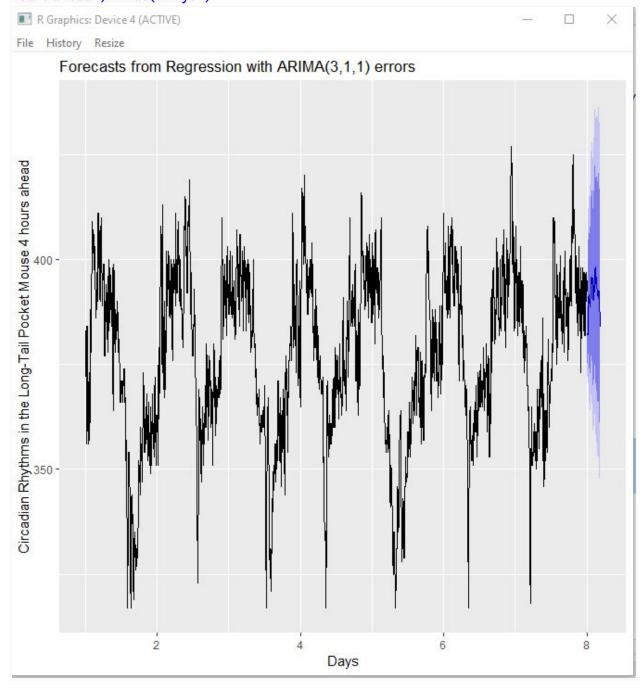
forecast(xreg=fourier(msts_pp, K=c(14,65), minutes_10)) %>% autoplot(include=one_week) + ylab("Circadian Rhythms in the Long-Tail Pocket Mouse 10 minutes ahead") + xlab("Days")



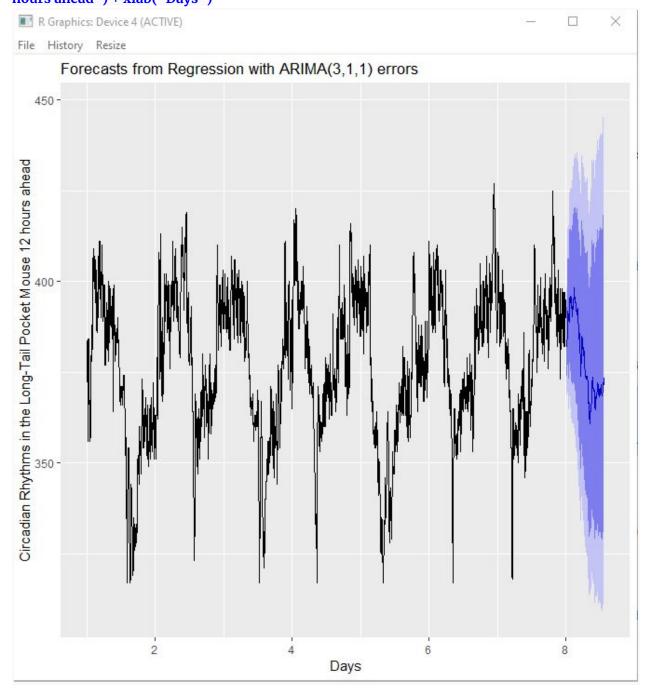
```
dev.new()
hour_1<- 30
fit %>%
forecast(xreg=fourier(msts_pp, K=c(14,65), h=hour_1)) %>%
  autoplot(include=one_week) + ylab("Circadian Rhythms in the Long-Tail Pocket Mouse 1 hour ahead") + xlab("Days")
```



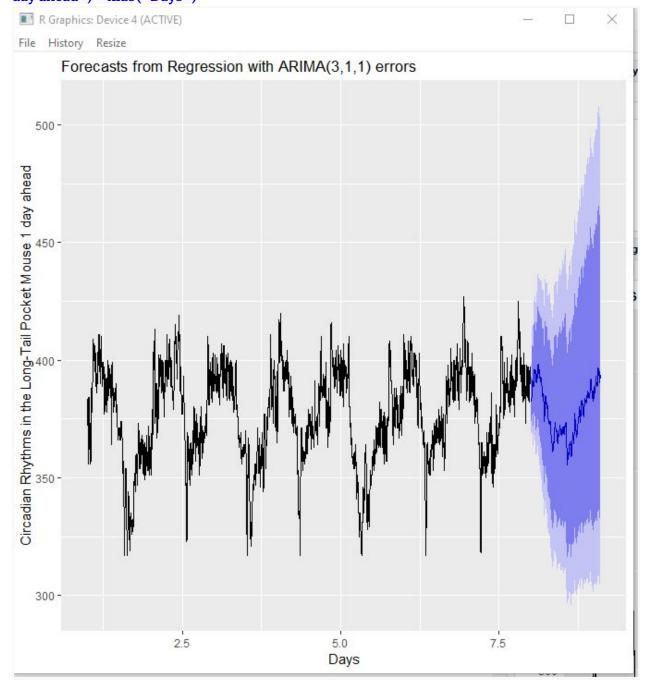
```
dev.new()
hours_4<- 30 *4
fit %>%
forecast(xreg=fourier(msts_pp, K=c(14,65), h=hours_4)) %>%
autoplot(include=one_week) + ylab("Circadian Rhythms in the Long-Tail Pocket Mouse 4
hours ahead") + xlab("Days")
```



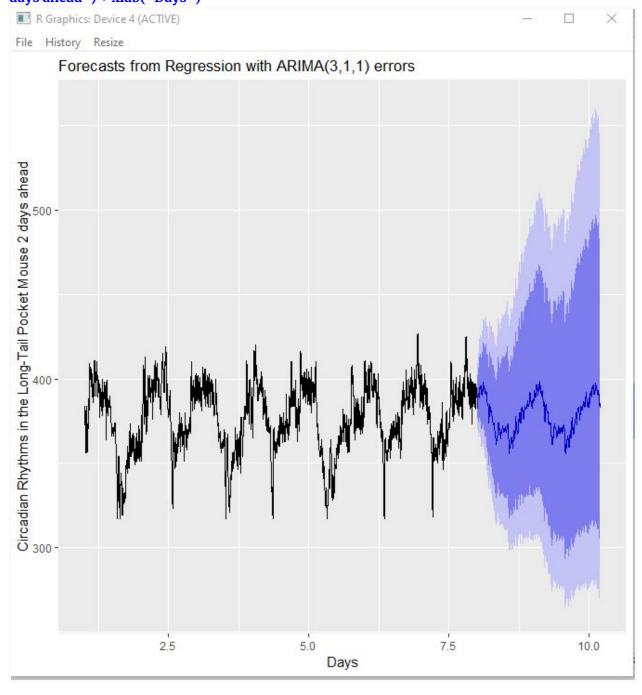
dev.new()
hours_12<- 30 * 12
fit %>%
forecast(xreg=fourier(msts_pp, K=c(14,65), h=hours_12)) %>%
autoplot(include=one_week) + ylab("Circadian Rhythms in the Long-Tail Pocket Mouse 12 hours ahead") + xlab("Days")



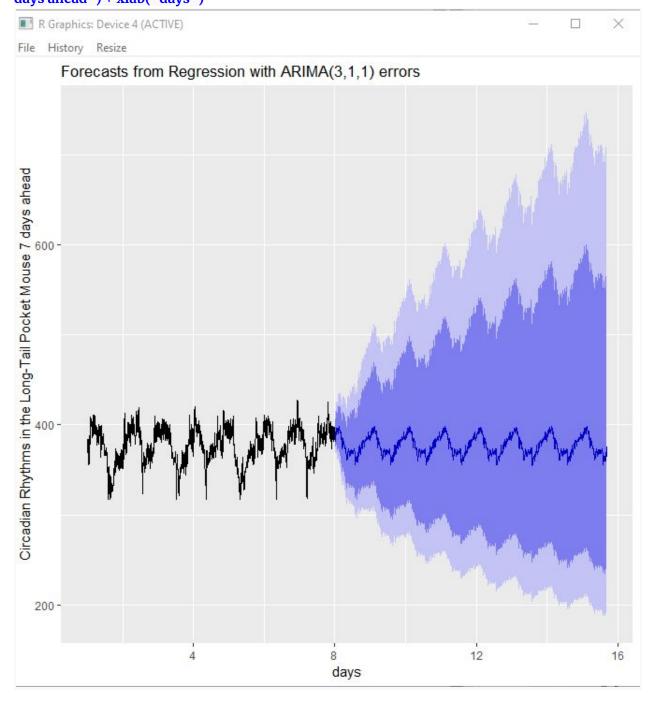
dev.new()
day_1<- 30 * 24
fit %>%
 forecast(xreg=fourier(msts_pp, K=c(14,65), h=day_1)) %>%
 autoplot(include=one_week) + ylab("Circadian Rhythms in the Long-Tail Pocket Mouse 1
day ahead") + xlab("Days")



```
dev.new()
day_2<- 30 * 24 * 2
fit %>%
forecast(xreg=fourier(msts_pp, K=c(14,65), h=day_2)) %>%
autoplot(include=one_week) + ylab("Circadian Rhythms in the Long-Tail Pocket Mouse 2 days ahead") + xlab("Days")
```



```
dev.new()
week_1<- 30 * 24 * 7
fit %>%
forecast(xreg=fourier(msts_pp, K=c(14,65), h=week_1)) %>%
autoplot(include=one_week) + ylab("Circadian Rhythms in the Long-Tail Pocket Mouse 7 days ahead") + xlab("days")
```



Bibliography

David Benson, Forecasting Daily Data with Multiple Seasonality in R http://www.dbenson.co.uk/Rparts/subpages/forecastR/

E. E. Holmes, M.D. Scheuerell, and E. J. Ward 2020-02-03, Analysis for Fisheries and Environmental Sciences

Rob J Hyndman, George Athanasopoulos, Forecasting Principles And Practice

DataSet http://www.statsci.org/data/general/pformos5.html

Appendix

```
library(astsa)
library("forecast")
library("lubridate")
examine.mod <- function(mod.fit.obj, p, d, q, P=0, D=0, Q=0, S=-1, lag.max=24) {
dev.new(width=6, height=6)
par(mfrow=c(2,1))
pacf(mod.fit.obj$fit$residuals, main="PACF of Residuals", lag.max)
if ((P==0)&(D==0)&(Q==0)) {
 title(paste("Model: (", p, ",", d, ",", q, ")", sep=""), adj=0, cex.main=0.75)
}
else {
 title(paste("Model: (", p, ", ", d, ", ", q, ") (", P, ", ", D, ", ", Q, ") [", S, "]", sep=""), adj=0,
cex.main=0.75)
}
std.resid <- mod.fit.obj$fit$residuals/sqrt(mod.fit.obj$fit$sigma2)</pre>
hist(std.resid, main="Histogram of Standardized Residuals", xlab="Standardized
Residuals", freq=FALSE)
curve(expr=dnorm(x, mean=mean(std.resid), sd=sd(std.resid)), col="red", add=TRUE)
}
pformosu<-read.table(file =
"C:/Users/inna/Desktop/DepaulClasses/ApplyMathClasses/Time Series/Final
Project/pformosu.txt")
row = 3726
col= 16
pformosu dat = numeric(col*row)
count = 1
for(i in 1:row)
{
for(j in 1:col)
 pformosu dat[count]=pformosu[i,j]
 count = count + 1
}
}
```

```
ts p<-ts(pformosu dat)
dev.new()
par(mfrow=c(2,1))
tsplot(ts p)
boxplot(ts p)$out
num of outliers = 0
repeat
{
outliers <- boxplot(ts p, plot=FALSE)$out
v<-(which(ts p %in% outliers))</pre>
if(length(v)==0)
 break
else
 ts_p<-ts_p[-v]
 num of outliers = num of outliers + length(v)
}
}
print(paste("Number Of Outliers removed = ", num_of_outliers))
print(paste("Length Of Time Series",length(ts p)))
dev.new()
par(mfrow=c(2,1))
tsplot(ts_p,xlab="Number Of Observations",main="Circadian rhythms in the last week Of
data")
boxplot(ts p)$out
frequency <- 30
one day<- 30*24
one week<- 24*30*7
ts pp <- ts(ts p[(length(ts p)-(one week) + 1):(length(ts p))],frequency = frequency)
dev.new()
tsplot(ts pp,xlab="Number Of Observations -> 7 Days -> 168 Hours",
  main="Circadian rhythms in the last week Of data")
x<-as.numeric(ts pp)
dev.new()
tsplot(ts_pp, ylab=expression(x[t]),
  xlab="hours", main="Circadian rhythms in the last week Of data")
dev.new()
acf2(x, max.lag = length(ts_pp) - 1,main = "ACF & PACF Of Original ts")
dev.new()
acf2(x, max.lag = 100, main = "ACF & PACF Of Original ts")
```

```
# Plot of (1-B^30)*x t
dev.new()
tsplot(diff(ts pp, lag=frequency, differences=1), ylab=expression((1-B^30)*x[t]),
  xlab="hours", main=expression(paste("Plot of ", (1-B^30)*x[t])))
# ACF indicated cut of after 330 lag that sugests that ARIMA Q=1,
# PACF exponencially decay that sugests ARIMA P=0
# Diff = 1 S = 18 (1 - B^30) \times t
dev.new()
acf2(diff(x, lag=frequency, differences=1), max.lag=frequency*5,
main=expression(paste("Est. ACF & PACF for ", (1-B^30)*x[t])))
# Plot of (1-B)(1-B^30)*x t
dev.new()
tsplot(diff(diff(ts_pp, lag=frequency, differences=1)),
  vlab=expression((1-B)(1-B^30)*x[t]),
  xlab="Hours", main=expression(paste("Plot of ",
                (1-B)(1-B^{30})*x[t]))
# ACF and PACF of (1-B)(1-B^30)*x t
dev.new()
acf2(diff( diff(x[1:length(x)], lag=frequency, differences=1)),
 max.lag=frequency,
 main=expression(paste("Est. ACF & PACF for ", (1-B)(1-B^30)*x[t])))
#########
# First Estimate ARIMA(p=0,d=1,q=0,P=0,D=1,Q=1,S=30) #######
dev.new()
mod.fit1 < - sarima(ts_pp,p=0,d=1,q=0,P=0,D=1,Q=1,S=30)
mod.fit1
dev.new()
par(mfrow=c(2,1))
pacf(mod.fit1$fit$residuals,main = "PACF Of residuals")
title(paste("Model: (",
   p = 0, ",",
d = 1, ",",
q = 0, ")(",
P = 0, ",",
D = 1, ",",
    Q = 1, ")[30]",
    sep=""),adj=0,cex.main=0.75)
std.resid1 <- mod.fit1$fit$residuals / sqrt(mod.fit1$fit$sigma2)</pre>
hist(std.resid1,main = "Histogram Of Standirdized Residuals",
```

```
xlab='Standardized Residuals',
freq = FALSE
### # First Estimate ARIMA(p=0,d=1,q=1,P=0,D=1,Q=1,S=30) #######
mod.fit1<- sarima(ts_pp,p=0,d=1,q=1,P=0,D=1,Q=1,S=30)
mod.fit1
dev.new()
par(mfrow=c(2,1))
pacf(mod.fit1$fit$residuals,main = "PACF Of residuals")
title(paste("Model: (",
    p = 0, ", ",
d = 1, ",",
q = 1, ")(",
P = 0, ", ",
D = 1, ",",
Q = 1, ")[30]",
     sep=""),adj=0,cex.main=0.75)
std.resid1 <- mod.fit1$fit$residuals / sqrt(mod.fit1$fit$sigma2)</pre>
hist(std.resid1,main = "Histogram Of Standardized Residuals",
 xlab='Standardized Residuals',
 freq = FALSE
### ARIMA(p=1,d=1,q=1,P=0,D=1,Q=1,S=30)
dev.new()
mod.fit1 < - sarima(ts pp,p=1,d=1,q=1,P=0,D=1,Q=1,S=30)
mod.fit1
dev.new()
par(mfrow=c(2,1))
pacf(mod.fit1$fit$residuals,main = "PACF Of residuals")
title(paste("Model: (",
p = 1, ",",
d = 1, ",",
q = 1, ")(",
P = 0, ", ",
D = 1, ",",
    Q=1,")[30]",
     sep=""),adj=0,cex.main=0.75)
std.resid1 <- mod.fit1$fit$residuals / sqrt(mod.fit1$fit$sigma2)</pre>
hist(std.resid1,main = "Histogram Of Standirdized Residuals",
 xlab='Standardized Residuals',
 freq = FALSE
#### ARIMA(p=0,d=1,q=2,P=0,D=1,Q=1,S=30)
dev.new()
```

```
mod.fit1 < - sarima(ts pp,p=0,d=1,q=2,P=0,D=1,Q=1,S=30)
mod.fit1
dev.new()
par(mfrow=c(2,1))
pacf(mod.fit1$fit$residuals,main = "PACF Of residuals")
title(paste("Model: (",
    p = 0, ", ",
d = 1, ",",
q = 2, ")(",
P = 0, ",",
D = 1, ",",
Q = 1, ")[30]",
     sep=""),adj=0,cex.main=0.75)
std.resid1 <- mod.fit1$fit$residuals / sqrt(mod.fit1$fit$sigma2)</pre>
hist(std.resid1,main = "Histogram Of Standardized Residuals",
  xlab='Standardized Residuals',
  freq = FALSE
#### ARIMA(p=0,d=1,q=3,P=0,D=1,Q=1,S=30)
dev.new()
mod.fit1 < - sarima(ts_pp,p=0,d=1,q=3,P=0,D=1,Q=1,S=30)
mod.fit1
dev.new()
par(mfrow=c(2,1))
pacf(mod.fit1$fit$residuals,main = "PACF Of residuals")
title(paste("Model: (",
p = 0, ",",
d = 1, ",",
q = 3, ")(",
P = 0, ", ", "
D = 1, ",",
    Q = 1, ")[30]",
     sep=""),adj=0,cex.main=0.75)
std.resid1 <- mod.fit1$fit$residuals / sqrt(mod.fit1$fit$sigma2)</pre>
hist(std.resid1,main = "Histogram Of Standirdized Residuals",
  xlab='Standardized Residuals',
  freq = FALSE
#### ARIMA(p=1,d=1,q=0,P=0,D=1,Q=1,S=30)
dev.new()
mod.fit1<- sarima(ts_pp,p=1,d=1,q=0,P=0,D=1,Q=1,S=30)
mod.fit1
dev.new()
par(mfrow=c(2,1))
pacf(mod.fit1$fit$residuals,main = "PACF Of residuals")
```

```
title(paste("Model: (",
p = 1, ",",
d = 1, ",",
q = 0, ")(",
P = 0, ",",
D = 1, ",",
Q=1,")[30]",
     sep=""),adj=0,cex.main=0.75)
std.resid1 <- mod.fit1$fit$residuals / sqrt(mod.fit1$fit$sigma2)</pre>
hist(std.resid1,main = "Histogram Of Standirdized Residuals",
  xlab='Standardized Residuals'.
  frea = FALSE
#### ARIMA(p=2,d=1,q=0,P=0,D=1,Q=1,S=30)
mod.fit1 < - sarima(ts_pp,p=2,d=1,q=0,P=0,D=1,Q=1,S=30)
mod.fit1
dev.new()
par(mfrow=c(2,1))
pacf(mod.fit1$fit$residuals,main = "PACF Of residuals")
title(paste("Model: (",
    p = 2, ",",
d = 1, ",",
q = 0, ")(",
P = 0, ",",
D = 1, ",",
    Q = 1, ")[30]",
     sep=""),adj=0,cex.main=0.75)
std.resid1 <- mod.fit1$fit$residuals / sqrt(mod.fit1$fit$sigma2)</pre>
hist(std.resid1,main = "Histogram Of Standardized Residuals",
  xlab='Standardized Residuals'.
freq = FALSE
#### ARIMA(p=2,d=1,q=2,P=0,D=1,Q=1,S=30)
dev.new()
mod.fit1<- sarima(ts_pp,p=2,d=1,q=2,P=0,D=1,Q=1,S=30)
mod.fit1
dev.new()
par(mfrow=c(2,1))
pacf(mod.fit1$fit$residuals,main = "PACF Of residuals")
title(paste("Model: (",
p = 2, ",",
d = 1, ",",
q = 2, ")(",
```

```
P = 0, ", ",
D = 1, ",",
Q = 1, ")[30]",
    sep=""),adj=0,cex.main=0.75)
std.resid1 <- mod.fit1$fit$residuals / sqrt(mod.fit1$fit$sigma2)
hist(std.resid1,main = "Histogram Of Standardized Residuals",
 xlab='Standardized Residuals',
freq = FALSE
#### ARIMA(p=2,d=1,q=3,P=0,D=1,Q=1,S=30)
dev.new()
mod.fit1<- sarima(ts_pp,p=2,d=1,q=3,P=0,D=1,Q=1,S=30)
mod.fit1
dev.new()
par(mfrow=c(2,1))
pacf(mod.fit1$fit$residuals,main = "PACF Of residuals")
title(paste("Model: (",
p = 3, ",",
d = 1, ",",
q = 2, ")(",
P = 0, ",",
D = 1, ",",
    Q = 1, ")[30]",
    sep=""),adj=0,cex.main=0.75)
std.resid1 <- mod.fit1$fit$residuals / sqrt(mod.fit1$fit$sigma2)</pre>
hist(std.resid1,main = "Histogram Of Standardized Residuals",
 xlab='Standardized Residuals',
 freq = FALSE
dev.new()
mod.fit.110.011 < - sarima(x, p=2,d=1,q=2,P=0,D=1,Q=1,S=30)
mod.fit.110.011
examine.mod(mod.fit.110.011, 2,1,2, 0,1,1, 30)
ahead 1 hours <-30
dev.new()
fore.mod <- sarima.for(ts_pp, n.ahead=ahead_1_hours,</pre>
   p=2, d=1, q=2, P=0, D=1, Q=1, S=30, plot.all=TRUE)
dev.new()
ts pp 1 hour < c(ts pp[(length(ts pp)-12*ahead 1 hours + 1):(length(ts pp))],
       fore.mod$pred[1:length(fore.mod$pred)])
tsplot(ts pp 1 hour,
   ylab="Data for 1 Hour ahead",
```

```
xlab="minutes", main=expression("360 minutes + 60 minutes ahead"))
ahead 4 hours <- 4*30
dev.new()
fore.mod <- sarima.for(ts_pp, n.ahead=ahead_4_hours,
  p=2, d=1, q=2, P=0, D=1, Q=1, S=30, plot.all=TRUE)
dev.new()
ts_pp_4_hour <- c(ts_pp[(length(ts_pp)-6*ahead_4_hours + 1):(length(ts_pp))],
      fore.mod$pred[1:length(fore.mod$pred)])
tsplot(ts_pp_4_hour,
  ylab="Data for 4 Hours ahead",
  xlab="minutes", main=expression("1 day + 2hours ahead"))
ahead 24 hours <- 24*30
dev.new()
fore.mod <- sarima.for(ts pp, n.ahead=ahead 24 hours,
        p=2, d=1, q=2, P=0, D=1, Q=1, S=30, plot.all=TRUE)
dev.new()
ts_pp_24_hour <- c(ts_pp[(length(ts_pp)-2*ahead_24_hours+1):(length(ts_pp))],
      fore.mod$pred[1:length(fore.mod$pred)])
tsplot(ts pp 24 hour,
  ylab="Data for 24 Hours ahead",
  xlab="minutes", main=expression("4 day + 24 hours ahead"))
ahead 24 hours <- 24*30
dev.new()
fore.mod <- sarima.for(ts_pp, n.ahead=ahead_24_hours, p=2, d=1, q=2, P=0, D=1, Q=1,
S=30, plot.all=TRUE)
pred.mod <- ts(x - mod.fit.110.011$fit$residuals)</pre>
dev.new()
tsplot(ts pp, ylab="pred", xlab="min", type="o", main="Predicted Model fitting with
Data")
lines(pred.mod, col="red", type="o", pch=17)
legend("topleft", legend=c("Observed", "Forecast"), lty=c("solid", "solid"), col=c("black",
"red"),
pch=c(1, 17), bty="n")
```