

Tin Sinh học Bioinformatics

Bài thực hành BioPython 1

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Định dạng FastA, GenBank
3. Các thao tác xử lý chuỗi trình tự



1. Cài đặt môi trường Window Subsystem Linux và Ubuntu

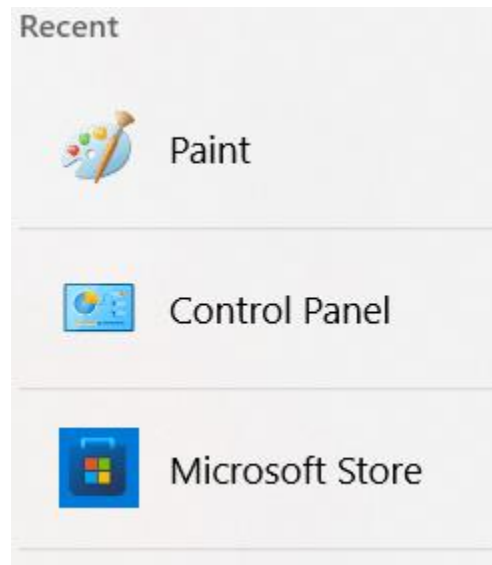


Báo cáo bài thực hành

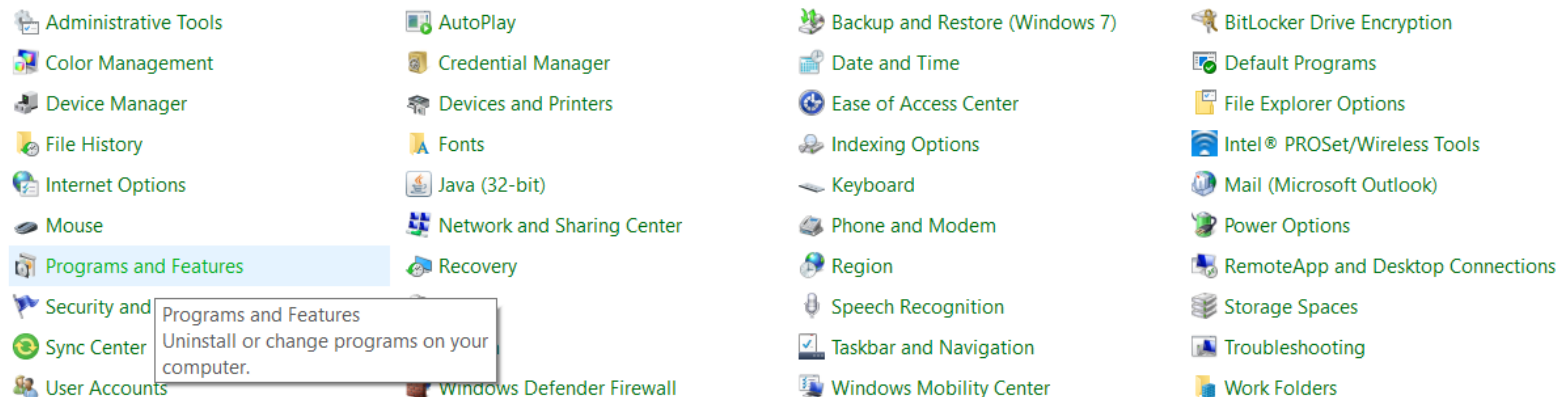
- Thực hiện các thao tác trong bài thực hành
- Ghi lại kết quả vào báo cáo

I. Cài đặt Windows Subsystem Linux

- Vào Control Panel > Program and Features > Turn Windows feature on or off



Adjust your computer's settings

View by: [Small icons](#)

Programs and Features

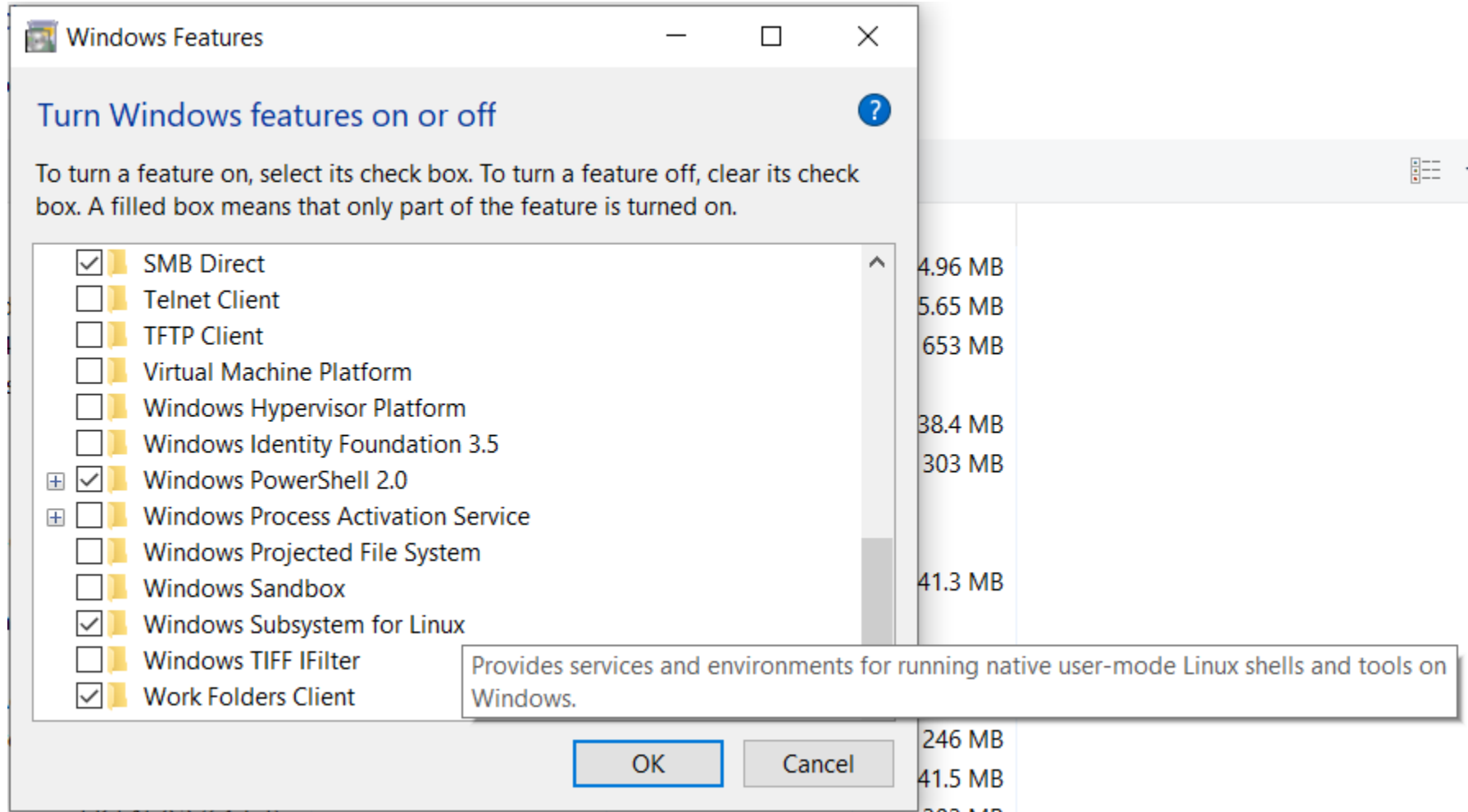
[Control Panel Home](#)[View installed updates](#)[Turn Windows features on or off](#)

Uninstall or change a program

To uninstall a program, select it from the list and then

Organize ▾

Cài đặt Windows Subsystem Linux



Cài đặt Ubuntu 22.04 LTS Vào Microsoft Store

Microsoft Store

ubuntu

Home

Apps

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Ubuntu

Apps | Developer tools

Canonical Group Limited

4.2 ★ Average

397 Ratings

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Get

Install a complete Ubuntu terminal environment in minutes with Windows Subsystem for Linux (WSL). Develop cross-platform applications, improve your data science or web development workflows and manage IT...

Ubuntu 22.04.2 LTS Free

4.3 ★ Apps | Developer t...

Install a complete Ubuntu terminal environment in minutes with Windows Subsystem for Linux (WSL)...

Ubuntu 20.04.5 LTS Installed

4.0 ★ Apps | Develo...

Install a complete Ubuntu terminal environment in minutes with Windows Subsystem for Linux (WSL)...

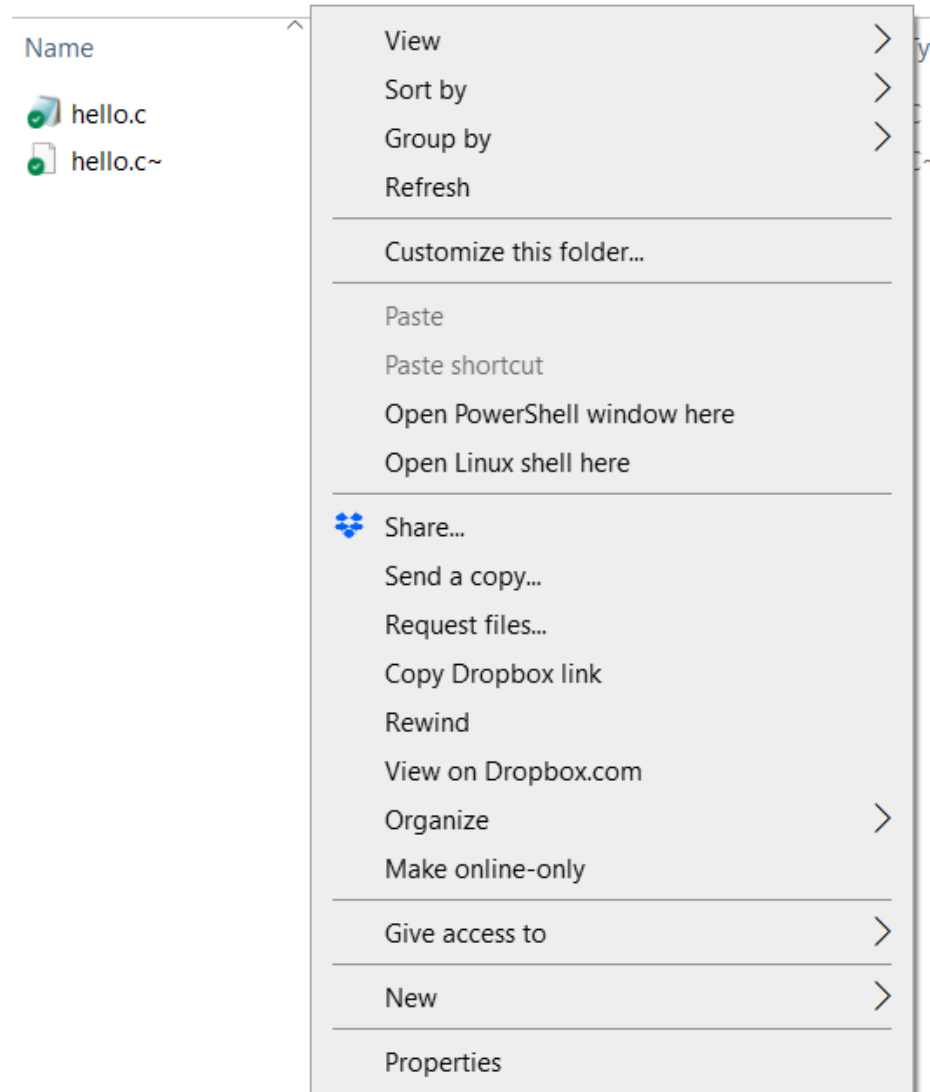
Ubuntu 18.04.5 LTS Free

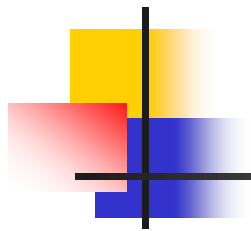
4.5 ★ Apps | Developer t...

Install a complete Ubuntu terminal environment in minutes with Windows Subsystem for Linux (WSL)...

Đổ vào môi trường WSL Linux

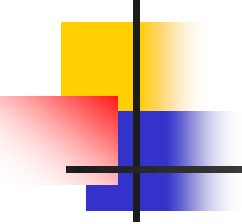
- Vào thư mục hiện tại
- Giữ nút Shift và ấn chuột phải
- Chọn “Open Linux shell here”





2. Quick Start

What can you do with Biopython?



```
import Bio
from Bio.Seq import Seq
from Bio import SeqIO
from Bio.SeqUtils import GC
print("biopython version: ", Bio.__version__)
my_seq = Seq("AGTACACTGGT")
print(my_seq)
print(my_seq.complement())
print("Reverse complement: ",
my_seq.reverse_complement())
```



Đọc các định dạng file phổ biến: FASTA, GENBANK

FASTA parsing example

```
for seq_record in SeqIO.parse("ls_orchid.fasta", "fasta"):
    print("Id của chuỗi: ", seq_record.id)
    print(repr(seq_record.seq))
    print(seq_record.seq, " => ", len(seq_record.seq), "
nucleotides")
```

```
print("\n", seq_record)
break
```

**Báo cáo: Mô tả chi tiết
định dạng FastA**



GenBank file format

```
for seq_record in SeqIO.parse("ls_orchid.gb", "genbank"):
    print(seq_record.id)
    print(repr(seq_record.seq))
    print(seq_record.seq, " => ", len(seq_record.seq))

print("\n### Thông tin bản ghi theo định dạng GENBANK")
print(seq_record)
break
```

**Báo cáo: Mô tả chi tiết
định dạng GenBank**



3. Các thao tác xử lý chuỗi trình tự



3.1 Sequences act like strings

```
my_seq = Seq("GATCG")
for index, letter in enumerate(my_seq):
    print("%i %s" % (index, letter))
    #print(index, letter)
    #print(my_seq[index])

print("Len sequence: ", len(my_seq))
print(my_seq[0])
print(my_seq[2])
print(my_seq[-1])
```



Non-overlapping count

```
print("AAAA".count("AA"))
```

```
my_seq = Seq("GATCGATGGGCCTATATAGGATCGAAAATCGC")
print("Len my_seq: ", len(my_seq))
print("Tổng số G: ", my_seq.count("G"))
print("Tổng số C: ", my_seq.count("C"))
gc = 100 * float(my_seq.count("G") + my_seq.count("C")) / len(my_seq)
print("Tỷ lệ GC: ", gc)
```

```
my_seq = Seq("GATCGATGGGCCTATATAGGATCGAAAATCGC")
print("Tỷ lệ GC: ", GC(my_seq))
```




3.2. Slicing a sequence

```
my_seq = Seq("GATCGATGGGCCTATATAGGATCGAAAATCGC")
```

```
my_seq[4:12]
```

```
# get the first, second and third codon positions of this DNA  
sequence:
```

```
# với 3 ORF (Open Reading Frame)
```

```
print(my_seq[0::3])
```

```
print(my_seq[1::3])
```

```
print(my_seq[2::3])
```

```
# Reverse the string
```

```
print(my_seq[::-1])
```



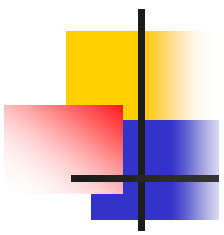
3.3. Turning Seq objects into strings

```
print(str(my_seq))
```

```
# Convert to FASTA format
```

```
fasta_format_string = ">Name\n%s\n" %  
my_seq
```

```
print("\nFASTA:\n", fasta_format_string)
```



3.4. Concatenating or adding sequences

```
dna_seq_1 = Seq("ACGT")
dna_seq_2 = Seq("CGTATG")
dna_seq = dna_seq_1 + dna_seq_2
print("dna_seq: ", dna_seq)
```

```
list_of_seqs = [Seq("ACGT"), Seq("AACC"), Seq("GGTT")]
concatenated = Seq("")
for s in list_of_seqs:
    concatenated += s
print("concatenated: ", concatenated)
```



join method

```
contigs = [Seq("ATG"), Seq("ATCCCG"),  
Seq("TTGCA")]
```

```
spacer = Seq("N"*10)
```

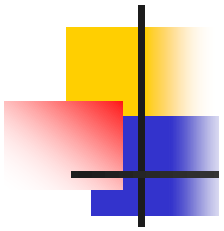
```
new_seq = spacer.join(contigs)
```

```
print("new_seq: ", new_seq)
```



Changing case

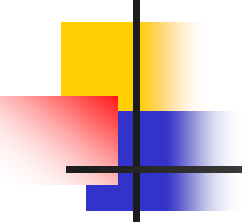
```
dna_seq = Seq("acgtACGT")  
dna_seq_upper = dna_seq.upper()  
dna_seq_lower = dna_seq.lower()  
print("dna_seq_upper: ", dna_seq_upper)  
print("dna_seq_lower: ", dna_seq_lower)
```

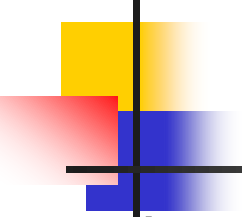


Nucleotide sequences and (reverse) complements

```
my_seq = Seq("ACGGTA")  
print("\n Complement: ",  
my_seq.complement())  
print("Reverse complement: ",  
my_seq.reverse_complement())
```

- 1000000

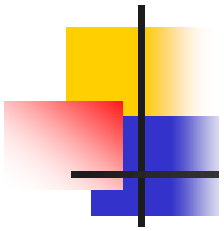
- 
- The actual biological transcription process works from the template strand, doing a reverse complement (TCAG \rightarrow CUGA) to give the mRNA. However, in Biopython and bioinformatics in general, we typically work directly with the coding strand because this means we can get the mRNA sequence just by switching T \rightarrow U.
 - These should match the figure above - remember by convention nucleotide sequences are normally read from the 5' to 3' direction, while in the figure the template strand is shown reversed.



```
coding_dna = Seq("ATGGCC")
print("coding_dna:  ", coding_dna)
template_dna = coding_dna.reverse_complement()
print("template_dna: ", template_dna)
```

```
messenger_rna = coding_dna.transcribe()
print("messenger_rna: ", messenger_rna)
```

```
# The Seq object also includes a back-transcription method for
# going from the mRNA to the coding strand of the DNA.
# Again, this is a simple U → T substitution:
print("back_transcribe: ", messenger_rna.back_transcribe())
```



3.8. Bảng dịch mã Translation Table

```
from Bio.Data import CodonTable
standard_table = CodonTable.unambiguous_dna_by_name["Standard"]
#standard_table = CodonTable.unambiguous_dna_by_id[1]

# Vertebrate Mitochondrial: ty thể của động vật có xương sống
mito_table = CodonTable.unambiguous_dna_by_name["Vertebrate
Mitochondrial"]
mito_table = CodonTable.unambiguous_dna_by_id[2]

print(standard_table)
#print(mito_table)

print("Stop codons: ", standard_table.stop_codons)
print("Start codons: ", standard_table.start_codons)
```



3.9. Quá trình dịch mã: Translation

```
messenger_rna =  
Seq("AUGGCCAUUGUAAUGGGCCGCUGAAAGGGUGCCCCGAUAG")  
protein_seq = messenger_rna.translate()  
print("protein_seq: ", protein_seq)  
print(messenger_rna.translate(to_stop=True))
```

You can also translate directly from the coding strand DNA sequence:

```
coding_dna =  
Seq("ATGGCCATTGTAATGGGCCGCTGAAAGGGTGCCCGATAG")  
protein_seq = coding_dna.translate(to_stop=True)  
print("protein_seq: ", protein_seq)
```



3.10. Comparing Seq objects

```
# Comparing Seq objects  
seq1 = Seq("ACGT")  
print("ACGT" == seq1)  
print(seq1 == "ACGT")
```



3.11. MutableSeq objects

```
from Bio.Seq import Seq
```

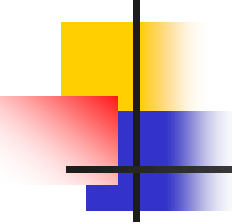
```
my_seq =
```

```
Seq("GCCATTGTAATGGGCCGCTGAAAGGGTGCCCGA")
```

```
my_seq[5] = "G"
```

```
-----  
TypeError                                Traceback (most recent call last)  
<ipython-input-26-50e848a55f5c> in <module>  
----> 1 my_seq[5] = "G"
```

```
TypeError: 'Seq' object does not support item assignment
```

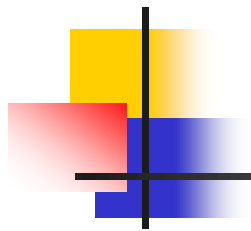


```
from Bio.Seq import MutableSeq
mutable_seq =
MutableSeq("GCCATTGTAATGGGCCGCTGAAAGGGTGCCCCGA")
mutable_seq[5] = "C"
mutable_seq
mutable_seq.remove("T")
mutable_seq
mutable_seq.reverse()
mutable_seq
# get back to a read-only Seq object
new_seq = mutable_seq.toseq()
new_seq
```



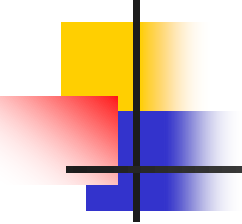
3.12. UnknownSeq objects

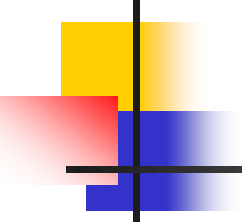
- The UnknownSeq object is a subclass of the basic Seq object and its purpose is to represent a sequence where we know the length, but not the actual letters making it up.
- You could of course use a normal Seq object in this situation, but it wastes rather a lot of memory to hold a string of a million "N" characters when you could just store a single letter "N" and the desired length as an integer.



```
from Bio.Seq import UnknownSeq
unk = UnknownSeq(20)
unk
print(unk)
len(unk)
```

```
????????????????????
20
```


- 
-
- For DNA or RNA sequences, unknown nucleotides are commonly denoted by the letter “N”, while for proteins “X” is commonly used for unknown amino acids.
 - When creating an `UnknownSeq`, you can specify the character to be used instead of “?” to represent unknown letters.



```
from Bio.Seq import UnknownSeq
unk_dna = UnknownSeq(20, character="N")
unk_dna
print(unk_dna)
```

NNNNNNNNNNNNNNNNNNNNNNNNNNNN



Tài liệu tham khảo

- Biopython Tutorial and Cookbook

- Link:

<http://biopython.org/DIST/docs/tutorial/Tutorial.html>