

Package ‘trqwe’

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Description Performance oriented statistical metrics and utility functions.

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allDups	<i>All duplicates.</i>
---------	------------------------

Description

Finds all duplicates in a vector including first instances of duplicates.

Usage

```
allDups(vec)
```

Arguments

vec A vector.

Value

A boolean vector of the same length as vec. TRUE if the element is duplicated, FALSE if not.

Examples

```
allDups(sample(1:100, size=100, replace=T))
```

append<-	<i>Append to a vector.</i>
----------	----------------------------

Description

Appends the 2nd argument to the 1st.

Usage

```
append(x) <- value
```

Arguments

x	A vector.
value	The element to append.

Examples

```
x <- 1:5
append(x) <- 6
print(x)

[1] 1 2 3 4 5 6
```

bioc	<i>Install Bioconductor package.</i>
------	--------------------------------------

Description

Utility function for installing packages from bioconductor easily.

Usage

```
bioc(package)
```

Arguments

package	unquoted package name.
---------	------------------------

Examples

```
\code{bioc(DESeq2)}
```

`chop`*Cleans leading and trailing whitespace.*

Description

Removes leading and trailing whitespace in a vector of strings.

Usage

```
chop(x)
```

Arguments

`x` A character vector.

Value

A vector with leading and trailing whitespace removed.

Examples

```
chop(c("  hello ", " 123 \t"))
```

```
[1] "hello" "123"
```

`cindex`*Concordance Index.*

Description

Calculates the concordance index from the results of a censored survival model. Very fast compared to other packages.

Usage

```
cindex(probs, time, event)
```

Arguments

`probs` The prognostic score of each patient.

`time` The time of each patient.

`event` The death of each patient: 1 for a patient death, 0 for censored.

Value

The concordance index.

cosineDist	<i>Cosine distance.</i>
------------	-------------------------

Description

Calculates the cosine distance of rows of a matrix.

Usage

```
cosineDist(x)
```

Arguments

x	A matrix.
---	-----------

Value

Cosine distance as a dist object.

See Also

<http://stackoverflow.com/questions/2535234/find-cosine-similarity-in-r>

Examples

```
rbind(c(1,1,1,0,0,0), c(0,0,0,1,1,1), c(1,1,1,0,0,1)) -> x
cosineDist(x)
      1      2
2 1.0000000
3 0.1339746 0.7113249
```

f1score	<i>F1 score.</i>
---------	------------------

Description

Calculates F1 score from the results of a classification model.

Usage

```
f1score(probs, class)
```

Arguments

probs	A numeric vector where 1 is predicted positive and 0 is predicted negative.
class	A numeric vector where 1 is positive and 0 is negative.

Value

The F1 score.

See Also

https://en.wikipedia.org/wiki/F1_score

fastAUC	<i>Fast AUC</i>
---------	-----------------

Description

This function calculates the Area Under the Receiver-Operator Curve from the results of a classification model.

Usage

```
fastAUC(probs, class, method = "trqwe")
```

Arguments

probs	A numeric vector of probabilities or likelihoods for each data point.
class	A numeric vector where 1 is positive and 0 is negative.

Value

The AUC.

See Also

Reference <https://stat.ethz.ch/pipermail/r-help/2005-September/079872.html>

Examples

```
library(AppliedPredictiveModeling)
data(abalone)
library(glmnet)
class <- factor(as.numeric(abalone$Type == "M"))
data <- data.matrix(abalone[, -1])
fit <- cv.glmnet(data, class, family="binomial")
probs <- predict(fit, newx=data)[,1]
fastAUC(probs, class)
```

fastPR	<i>Precision-Recall curve</i>
--------	-------------------------------

Description

Calculates the points in a Precision-Recall from the results of a classification model.

Usage

```
fastPR(probs, class)
```

Arguments

probs	A numeric vector of probabilities or likelihoods for each data point.
class	A numeric vector where 1 is positive and 0 is negative.

Value

a list containing the ROC curve.

Examples

```
library(AppliedPredictiveModeling)
data(abalone)
library(glmnet)
class <- factor(as.numeric(abalone$Type == "M"))
data <- data.matrix(abalone[, -1])
fit <- cv.glmnet(data, class, family="binomial")
probs <- predict(fit, newx=data)[,1]
with(fastPR(probs, class), plot(recall, precision, type="l"))
```

fastReadLines	<i>Fast readLines.</i>
---------------	------------------------

Description

Replacement for readLines, faster.

Usage

```
fastReadLines(fname, newlinechar = "\n", nchars = NULL)
```

Arguments

fname	Filename to read.
newlinechar	The new line character in the file.

Value

A vector containing all the lines in the file.

Examples

```
library(microbenchmark)
writeLines(replicate(100000, sample(letters, size=100, replace=T)), con="/tmp/temp.txt")
microbenchmark(fastReadLines("/tmp/temp.txt"),
               readLines("/tmp/temp.txt"), times=3)[,c("expr", "mean"), drop=F]

Unit: milliseconds
      expr      min       lq     mean  median      uq
fastReadLines("/tmp/temp.txt") 335.3874 335.9188 336.5900 336.4502 337.1912
  readLines("/tmp/temp.txt") 724.9136 725.4523 727.2444 725.9911 728.4098
```

fastROC	<i>Fast ROC</i>
---------	-----------------

Description

Calculates the points in a Receiver-Operator Curve from the results of a classification model.

Usage

```
fastROC(probs, class)
```

Arguments

probs	A numeric vector of probabilities or likelihoods for each data point.
class	A numeric vector where 1 is positive and 0 is negative.

Value

a list containing the ROC curve.

Examples

```
library(AppliedPredictiveModeling)
data(abalone)
library(glmnet)
class <- factor(as.numeric(abalone$Type == "M"))
data <- data.matrix(abalone[, -1])
fit <- cv.glmnet(data, class, family="binomial")
probs <- predict(fit, newx=data)[,1]
with(fastROC(probs, class), plot(fpr, tpr, type="l"))
```

fpkmFromCounts	<i>Calculate FPKM.</i>
----------------	------------------------

Description

Calculates FPKM from a count matrix.

Usage

```
fpkmFromCounts(mat, gene_lengths, uq_norm = T)
```

Arguments

mat	An integer matrix representing the counts of a RNA-Seq dataset.
gene_lengths	A named vector with the length of each gene.
uq_norm	If TRUE, will perform upper-quartile normalization.

Value

A matrix containing FPKM with the same dimensions as mat.

See Also

FPKM-UQ - A normalized read count in which gene expression values, in FPKM, are divided by the 75th percentile value.

https://gdc-docs.nci.nih.gov/Data/Bioinformatics_Pipelines/Expression_mRNA_Pipeline/
<http://vinaykmittal.blogspot.com/2013/10/fpkmrpk-normalization-caveat-and-upper.html>

head2	<i>Upper-left corner of matrix.</i>
-------	-------------------------------------

Description

Shortcut function for previewing a matrix or data.frame by displaying the upper-left corner. Similar to head.

Usage

```
head2(x, n = 10, ncols = 10)
```

Arguments

x	A wide matrix or data.frame.
n	Number of lines to display. Default 10.
ncols	Number of columns to display. Default 10.

Value

n by ncols subset of the matrix taken from the upper-left corner.

install	<i>Install CRAN package.</i>
---------	------------------------------

Description

Utility function for installing packages from CRAN easily.

Usage

```
install(package, repos = "http://cran.us.r-project.org")
```

Arguments

package	unquoted package name.
---------	------------------------

Examples

```
\code{install(Rcpp)}
```

logit	<i>Logit Transformation.</i>
-------	------------------------------

Description

Calculates the results of the Logit Transformation.

Usage

```
logit(x)
```

Arguments

x	Input to the function (e.g., probabilities from 0 to 1).
---	--

Value

Logit(x)

make_percent	<i>Make percentage from number</i>
--------------	------------------------------------

Description

Make percentage from number

Usage

```
make_percent(x, digits = 2)
```

Arguments

x	A number.
digits	Number of decimal places, default 2.

Value

A percentage.

Examples

```
make_percent(0.424, 2)
```

matrixFactor	<i>Matrix Factor Design.</i>
--------------	------------------------------

Description

From a factor, returns a design matrix with a column for each level.

Usage

```
matrixFactor(x, names = NULL)
```

Arguments

x	A factor.
names	The name of each instance in the resulting matrix (i.e., the rownames).

Value

The design matrix.

Examples

```
matrixFactor(factor(letters))
```

mccscore	<i>Matthew's correlation coefficient</i>
----------	--

Description

Calculates Matthew's correlation coefficient. Requires the Rmpfr package.

Usage

```
mccscore(probs, class)
```

Arguments

probs	A numeric vector where 1 is predicted positive and 0 is predicted negative.
class	A numeric vector where 1 is positive and 0 is negative.

Value

The MCC score.

See Also

https://en.wikipedia.org/wiki/Matthews_correlation_coefficient

mcreadRDS	<i>Multi-threaded readRDS</i>
-----------	-------------------------------

Description

Uses the pigz utility to improve loading large R objects. This is compatible with saveRDS and readRDS functions. Requires pigz (sudo apt-get install pigz on Ubuntu).

Uses the pigz utility to improve loading large R objects. This is compatible with saveRDS and readRDS functions. Requires pigz (sudo apt-get install pigz on Ubuntu).

Usage

```
mcreadRDS(file, mc.cores = min(parallel::detectCores(), 4))
```

```
mcreadRDS(file, mc.cores = min(parallel::detectCores(), 4))
```

Arguments

file	The filename of the rds object.
mc.cores	How many cores to use in pigz. The program does not seem to benefit after more than about 4 cores.
file	The filename of the rds object.
mc.cores	How many cores to use in pigz. The program does not seem to benefit after more than about 4 cores.

Value

The R object.

The R object.

See Also

<http://stackoverflow.com/questions/28927750/>

<http://stackoverflow.com/questions/28927750/>

Examples

```
x <- sample(1e4, 1e7, replace=T)
saveRDS(x, file="temp.Rds")
xmc <- mcreadRDS("temp.Rds")
x <- sample(1e4, 1e7, replace=T)
saveRDS(x, file="temp.Rds")
xmc <- mcreadRDS("temp.Rds")
```

mcsaveRDS	<i>Multi-threaded saveRDS</i>
-----------	-------------------------------

Description

Uses the pigz utility to improve saving large R objects. This is compatible with saveRDS and readRDS functions. Requires pigz (sudo apt-get install pigz on Ubuntu).

Usage

```
mcsaveRDS(object, file, mc.cores = min(parallel::detectCores(), 4))
```

Arguments

object	An r object to save.
file	The filename to save to.
mc.cores	How many cores to use in pigz. The program does not seem to benefit after more than about 4 cores.

See Also

<http://stackoverflow.com/questions/28927750/>

Examples

```
x <- sample(1e4, 1e7, replace=T)
y <- sample(1e4, 1e7, replace=T)
microbenchmark(mcsaveRDS(x, file="temp.Rds"), saveRDS(y, file="temp2.Rds"))
```

Unit: seconds

	expr	min	lq	mean	median	uq
mcsaveRDS(x, file = "temp.Rds")		1.908310	1.908310	1.908310	1.908310	1.908310
saveRDS(y, file = "temp2.Rds")		6.271499	6.271499	6.271499	6.271499	6.271499

mcsplitapply	<i>Parallel split-matrix or dataframe apply loop</i>
--------------	--

Description

Splits a matrix or data.frame into subsets based on a factor, and applies a function to each subset. Typical use case: sum exon count data to gene count data. This is similar to the dplyr idiom: `df %>% group_by(f) %>% do(...)`, but has several advantages. 1) mcsplitapply can be used on matrices (and is therefore much faster), 2) inherently parallelized, 3) can return results other than dataframes, 4) you can specify how the data are combined (default is rbind).

Usage

```
mcsplitapply(mat, f, func, mc.cores = 4, .combine = rbind, ...)
```

Arguments

mat	The matrix.
f	A factor of length equal to nrow(mat). The levels of this factor will split the matrix into subsets.
func	The function to apply to each subset.
mc.cores	The number of cores to use.
.combine	The function to combine the results with. Default is rbind. Use NA to return a list.

Value

A list or a combined object depending on the .combine parameter.

Examples

```
library(pasilla)
library(DEXSeq)
library(trqwe)
data(pasillaDEXSeqDataSet)
exon_counts <- counts(dxd)
f <- rowData(dxd)$groupID
gene_counts <- mcsplitapply(exon_counts, f, colSums)
```

mgrepl	<i>Multiple grepl.</i>
--------	------------------------

Description

Takes in a list of regex patterns and returns true if any pattern matches.

Usage

```
mgrepl(patterns, x, ...)
```

Arguments

`x` A vector of strings to search.
`pattern` A vector of regex patterns.

Value

A vector of the same length as `x`, TRUE if any pattern matches.

Examples

```
x <- fastReadLines("http://textfiles.com/ufo/ufobooks.ufo", newlinechar="\r\n", 1e5)
head(x[mgrepl(c("ALIENS", "UFO"), x)])
```

```
[1] "                                UFOLOGY BOOKS (REVISION 2.1 343 books)"
[2] "    H. S. Stewart on the  subject of UFO's. The list is alphabetic"
[3] "    Tom Mickus's most excellent board UFONET I.  (416-237-1204)"
[4] "    Bill Adler                *  LETTERS TO THE AIR FORCE ON UFOS  1967"
[5] "    Gordon W. Allen           OVERLORDS OLYMPIANS AND THE UFO  1974"
[6] "    Robert B. Beard           FLYING SAUCERS, UFO'S AND EXTRA"
```

<code>nelson_aelen_surv</code>	<i>Nelson Aalen estimator</i>
--------------------------------	-------------------------------

Description

Nelson–Aalen estimator is an estimator of the cumulative hazard function in survival data. It can be used to compare the overall risks of two groups or used to estimate the number of deaths before a certain time.

Usage

```
nelson_aelen_surv(time, event)
```

Arguments

`time` The time of each patient.
`event` The death of each patient: 1 for a patient death, 0 for censored.

Value

A list containing the cumulative hazard function.

See Also

[#https://en.wikipedia.org/wiki/Nelson-Aalen_estimator](https://en.wikipedia.org/wiki/Nelson-Aalen_estimator)

Examples

```
library(survival)
data(veteran)
with(nelson_aelen_surv(veteran$time, veteran$status), plot(ti, Hi, type="b"))
```

posteriorBalance	<i>Posterior probability adjustment.</i>
------------------	--

Description

Adjusts the posterior probability of a classifier based on unbalanced datasets. In classification model where the negative data is randomly under-sampled and all the positive data is used, the adjustment factor (beta) is $p(s=1|-) = p(+)/p(-)$. I.e., the probability that a negative datapoint is selected in the classifier. $\text{beta} \sim N+/N-$.

Usage

```
posteriorBalance(probs, beta = NULL, Nplus, Nminus)
```

Arguments

probs	The original posterior probability.
beta	The adjustment factor.
Nplus	The number of positive examples in the real data. Only used if beta is NULL.
Nminus	The number of negative examples in the real data. Only used if beta is NULL.

Value

The adjusted posterior probability.

See Also

Dal Pozzolo, Andrea, et al. "Calibrating probability with undersampling for unbalanced classification." Computational Intelligence, 2015 IEEE Symposium Series on. IEEE, 2015.

prepend<-	<i>Prepend to a vector.</i>
-----------	-----------------------------

Description

Prepends the 2nd argument to the 1st.

Usage

```
prepend(x) <- value
```

Arguments

x	A vector.
value	The element to append.

Examples

```
x <- 1:5
prepend(x) <- 6
print(x)

[1] 6 1 2 3 4 5
```

readRz

*readRz***Description**

faster serialization + compression using fst package

Usage

```
readRz(file)
```

reload

*Reload a package.***Description**

Unload and reload a package and sets the namespace search order.

Usage

```
reload(package, pos = 2)
```

Arguments

package	Unquoted package name.
pos	Namespace search position.

Examples

```
\code{reload(trqwe)}
```

reloadtrqwe

*Unload and reload trqwe.***Description**

Unload and reload trqwe. Shortcut for `reload(trqwe)`

Usage

```
reloadtrqwe()
```

saveRz	<i>saveRz</i>
--------	---------------

Description

faster serialization + compression using fst package

Usage

```
saveRz(object, file, compressor = "ZSTD", compression = 0)
```

se	<i>Standard error.</i>
----	------------------------

Description

Calculates the standard error of a sampling distribution.

Usage

```
se(x)
```

Arguments

x	A vector.
---	-----------

Value

The standard error of x.

Examples

```
x <- rnorm(1e3)
se(x)
[1] 0.03192027
```

set_colnames	<i>Set colnames of data.frame or matrix</i>
--------------	---

Description

Set colnames of data.frame or matrix and return it, for use with pipes

Usage

```
set_colnames(df, colnames)
```

Arguments

df	data.frame or matrix
colnames	colnames to add

Value

df with colnames added

set_rownames	<i>Set rownames of data.frame or matrix</i>
--------------	---

Description

Set rownames of data.frame or matrix and return it, for use with pipes

Usage

```
set_rownames(df, rownames)
```

Arguments

df	data.frame or matrix
rownames	rownames to add

Value

df with rownames added

sigmoid	<i>Sigmoid Function.</i>
---------	--------------------------

Description

Calculates the results of the sigmoid function.

Usage

```
sigmoid(x)
```

Arguments

probs x Input to the function.

Value

Sigmoid(x)

statsCallback	<i>Variable information.</i>
---------------	------------------------------

Description

Automatically stores basic information of variables in the previous command. This function adds a callback which reports information on previous variables and stores this information in `.stats`.

Usage

```
statsCallback()
```

Examples

```
statsCallback()
my_data <- VADeaths
.stats
> [1] "dim: 5 4, length: 20, class: matrix, typeof: double"
```

tablec	<i>Fast C++ tabulation.</i>
--------	-----------------------------

Description

Takes in a character, integer or factor vector and tabulates the number of times each element appears.

Usage

```
tablec(x, sort = F)
```

Arguments

x A character, integer or factor vector. NAs are allowed.
sort TRUE if the result names should be sorted alphanumerically.

Value

A integer vector of counts of each element.

Examples

```
x <- factor(sample(1e5, 1e8, replace=T))
microbenchmark(table(x), tablec(x), times=3)
```

Unit: milliseconds

expr	min	lq	mean	median	uq	max
table(x)	9777.6457	10479.0949	10717.3382	11180.544	11187.1844	11193.8246
tablec(x)	678.0364	685.9467	713.1181	693.857	730.6589	767.4608

```
x <- sample(letters, 1e8, replace=T)
microbenchmark(table(x), tablec(x), tablec(x,sort=T), times=3)
```

Unit: seconds

expr	min	lq	mean	median	uq	max
table(x)	5.778514	5.829125	5.855560	5.879737	5.894083	5.908430
tablec(x)	1.589360	1.589381	1.589516	1.589402	1.589594	1.589786
tablec(x, sort = T)	1.589386	1.590520	1.591824	1.591655	1.593044	1.594432

tail2	<i>Lower-left corner of matrix.</i>
-------	-------------------------------------

Description

Shortcut function for previewing the bottom of a matrix or data.frame by displaying the lower-left corner. Similar to tail.

Usage

```
tail2(x, n = 10, ncol = 10)
```

Arguments

x	A wide matrix or data.frame.
n	Number of lines to display. Default 10.
ncols	Number of columns to display. Default 10.

Value

n by ncols subset of the matrix taken from the lower-left corner.

TCGA_barcode	<i>Parse TCGA barcode.</i>
--------------	----------------------------

Description

Taking in a full TCGA sample barcode, or any subset of the barcode, and return extracted values.

Usage

```
TCGA_barcode(x, what = "patient")
```

Arguments

x	TCGA barcode, or a vector of barcodes.
what	Which information to return.

Value

The specified information contained in the barcode of the same length as x.

Examples

```
TCGA_barcode(c("TCGA-02-0001-01C-01D-0182-01", "TCGA-02-0001-11C-01D-0182-01"), what="tissue")

[1] "01" "11"
```

timePrompt	<i>Time profiling.</i>
------------	------------------------

Description

Reports time in seconds to the R prompt of the previous command. This function adds a callback which saves the running of individual commands and reports the time in seconds on the next line.

Usage

```
timePrompt()
```

Examples

```
> timePrompt()
0.000s> x <- sample(1:10, size=1e8, replace=T)
1.240s>
Note - this time is not accurate if child processes or multithreading is involved.
```

topn	<i>Highest elements in a vector.</i>
------	--------------------------------------

Description

Finds the top elements in a vector very quickly. Equivalent of `-sort(-x, partial=1:n)`

Usage

```
topn(x, n = 100, value = F, lowest = F)
```

Arguments

x	A numeric vector.
n	The number of top elements to return.
value	If TRUE, returns the values of the top elements. If FALSE, returns the indices.
lowest	If TRUE, returns the lowest elements instead of the highest.

Value

A vector containing the indices or the values of the top elements.

See Also

<http://stackoverflow.com/questions/18450778/>

Examples

```
naive_top <- function(x, n) {
  -sort(-x, partial=1:n)
}
x <- runif(1e7)
microbenchmark(naive_top(x,100), topn(x,100,value=T), times=10)
```

Unit: milliseconds

	expr	min	lq	mean	median	uq
	naive_top(x, 100)	1070.0180	1071.5951	1075.964	1072.3520	1073.9989
	topn(x, 100, value = T)	433.6682	433.8882	434.771	434.4986	435.6029

TOSTtwo	<i>#' Multiple return assignment #' @description Python style multiple return assignment. #' @examples #' mreturn[x,y,z] <- list("hello", c(1,2,3), sqrt(2)) #' print(x) #' [1] "hello" #' print(y) #' [1] 1 2 3 #' print(z) #' [1] 1.414214 #' @rdname mreturn mreturn <- structure(NA, class = "mreturn")</i>
---------	---

Description

```
'[<-.mreturn' <- function(mreturn, ..., value) vars <- sapply(substitute(list(...)), deparse)[-1] stopifnot(all(make.names(vars) == vars)) # invalid variable names stopifnot(length(value) == length(vars)) # incorrect number of return values for(i in 1:length(vars)) assign(vars[i], value=value[[i]], envir = parent.frame()) return(mreturn)
```

Usage

```
TOSTtwo(m1, m2, sd1, sd2, n1, n2, low_eqbound_d, high_eqbound_d, alpha, var.equal)
```

tost_power	<i>Calculates power/sample size of a TOST test</i>
------------	--

Description

Calculates the sample size required for a TOST test, given a sample size through exact statistics or simulation

Usage

```
tost_power(mu_A, mu_B, sd_A, sd_B, delta = 0.3, kappa = 1, alpha = 0.05, power = 0.8, method = "equivalence", n_iterations = 1000, paired = F, sample_var = 0)
```

Arguments

mu_A	mean of group A
mu_B	mean of group B
sd_A	SD of group A
sd_B	SD of group B
delta	maximum tolerated difference (i.e., $\text{abs}(\mu_A - \mu_B)$)
kappa	sample size of group A / sample size of group B (n_A / n_B)
alpha	significance threshold (0.05 default)
power	Desired power level (0.8 default)
method	Whether to use the exact statistics or simulation. "Exact" - exact statistics, "equivalence" - simulate using equivalence package tost function, "TOSTER" - simulate using TOSTER functions
n_iterations	Number of simulation iterations
paired	If the TOST is paired (kappa should be 1)
paired_var	Sample variance for paired TOST

Value

For exact statistics, returns a list of two values: minimum sample size to achieve desired power and the power at that sample size. For simulation, returns a series of sample sizes and achieved power up to the desired power.

See Also

<http://powerandsamplesize.com/Calculators/Compare-2-Means/2-Sample-Equality> Chow S, Shao J, Wang H. 2008. Sample Size Calculations in Clinical Research. 2nd Ed. Chapman & Hall/CRC Biostatistics Series. page 58.

Examples

```
res_equivalence <- tost_power(mu_A=1, mu_B=1, sd_A=0.2, sd_B=0.2, delta=0.2, method="equivalence")
res_toster <- tost_power(mu_A=1, mu_B=1, sd_A=0.2, sd_B=0.2, delta=0.2, method="TOSTER")
res_exact <- tost_power(mu_A=1, mu_B=1, sd_A=0.2, sd_B=0.2, delta=0.2, method="exact")

plot(res_equivalence$nB, res_equivalence$Power, xlim=c(1,20), ylim=c(0,1), type="s", main="TOST power analysis")
lines(res_toster$nB, res_toster$Power, col="green", type="s")
abline(h=0.8, col="blue", lty=2)
points(res_exact$nB, res_exact$Power, col="red", pch=18, cex=2)
legend("topleft", legend=c("equivalence", "TOSTER", "exact"), col=c("black", "green", "red"), pch=1)

res_equivalence <- tost_power(mu_A=1, mu_B=1, sd_A=0.2, sd_B=0.2, delta=0.1, method="equivalence")
res_exact <- tost_power(mu_A=1, mu_B=1, sd_A=0.2, sd_B=0.2, delta=0.1, method="exact")
plot(res_equivalence$nB, res_equivalence$Power, xlim=c(1,80), ylim=c(0,1), type="s", main="TOST power analysis")
abline(h=0.8, col="blue", lty=2)
points(res_exact$nB, res_exact$Power, col="red", pch=18, cex=2)
legend("topleft", legend=c("equivalence", "exact"), col=c("black", "red"), pch=1)
```

trqwe_KNN

*Fast binary KNN classifier***Description**

Fast binary KNN classifier

Usage

```
trqwe_KNN(distmat, train_idx, test_idx, classes, K, mc.cores = 1)
```

Arguments

distmat	A NxN pre-computed distance matrix
train_idx	Train indices
test_idx	Test indices
classes	vector length N, 1 or 0
K	Number of nearest neighbors parameter
mc.cores	Number of threads to use

Value

A prediction vector for the test set based on the class labels of the train set.

varSizes	<i>Size of R objects.</i>
----------	---------------------------

Description

Prints out the size of all R objects in the environment.

Usage

```
varSizes(env = globalenv(), units = "KB")
```

Arguments

env	The environment to search (default global environment).
units	Units to print out for each variable.

Value

A data.frame containing the size of each object.

ww_test	<i>Wald-Wolfowitz Runs Tests for Randomness</i>
---------	---

Description

This is the k-category asymptotic Z Test with continuity correction. Imagine rolling a die multiple times to obtain a sequence of rolls. This statistic tests whether there exists a "run" within the sequence where one particular number comes up more times in a row than expected randomly. If the test is significant, it can be concluded that the die rolls are not independent.

Usage

```
ww_test(x)
```

Arguments

x	A vector of items, coerced into a factor.
---	---

Value

A p-value for the statistical test.

See Also

Reference https://ncss-wpengine.netdna-ssl.com/wp-content/themes/ncss/pdf/Procedures/NCSS/Analysis_of_Runs.pdf

Examples

```
set.seed(1)
ww_test(sample(2, 100, replace=T))
ww_test(c(sample(6, 90, replace=T), rep(1,10)))
```

%Q%	<i>Concatenate strings.</i>
-----	-----------------------------

Description

Concatenates two strings.

Usage

a %Q% b

Arguments

a	First string.
b	Second string.

Value

The concatenated string.

Examples

```
'Hello ' %Q% 'World'
```

```
[1] "Hello World"
```

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