

# The Use of Lattice Upstream Targeting for the Analysis of mRNA Expression for Cancers

LUST 2019

Tristan Holmes   J.B. Nation et al

University of Hawaii at Manoa  
*tristanh314@gmail.com*

PSU Systems Science Seminar  
February 14, 2023

# Presentation Overview

- 1 Introduction
- 2 Data Setup
- 3 The Lattice Upstream Targeting Algorithm
- 4 Conclusions and Future Research

# Abstract

In 2019 the UH Cancer Center hosted a project to identify genetic factors of interest in various types of cancer.

# Abstract

In 2019 the UH Cancer Center hosted a project to identify genetic factors of interest in various types of cancer.

One of the results of this effort was the use of the Lattice Upstream Targeting (LUST) Algorithm to analyze mRNA expression data for 33 different types of cancer in the TCGA database. This effort will be the topic of this presentation.

# Abstract

In 2019 the UH Cancer Center hosted a project to identify genetic factors of interest in various types of cancer.

One of the results of this effort was the use of the Lattice Upstream Targeting (LUST) Algorithm to analyze mRNA expression data for 33 different types of cancer in the TCGA database. This effort will be the topic of this presentation.

The full results of this effort can be found on GitHub.

# Abstract

In 2019 the UH Cancer Center hosted a project to identify genetic factors of interest in various types of cancer.

One of the results of this effort was the use of the Lattice Upstream Targeting (LUST) Algorithm to analyze mRNA expression data for 33 different types of cancer in the TCGA database. This effort will be the topic of this presentation.

The full results of this effort can be found on GitHub.

Results of a similar project conducted using data proprietary to the UH cancer center led to studies seeking to identify new chemical treatments.

# Overview of Procedure

- The LUST algorithm is a discrete mathematical method for analyzing continuous data, e.g., mRNA expression.

# Overview of Procedure

- The LUST algorithm is a discrete mathematical method for analyzing continuous data, e.g., mRNA expression.
- For a given array of expression data, the algorithm is applied twice.



# Overview of Procedure

- The LUST algorithm is a discrete mathematical method for analyzing continuous data, e.g., mRNA expression.
- For a given array of expression data, the algorithm is applied twice.
- ① The first run is on the entire expression matrix and uses a graph theoretic objective function to rank the groups obtained. This pass identifies and ranks a small set of *metagenes* associated with the given cancer.

# Overview of Procedure

- The LUST algorithm is a discrete mathematical method for analyzing continuous data, e.g., mRNA expression.
- For a given array of expression data, the algorithm is applied twice.
- - 1 The first run is on the entire expression matrix and uses a graph theoretic objective function to rank the groups obtained. This pass identifies and ranks a small set of *metagenes* associated with the given cancer.
  - 2 The second run is on the expression matrix for each metagene and supervised by survival time as the objective function using the Fisher score to rank the results. This pass identifies small predictive *signiatures* for each metagene.

# Overview of Procedure

- The LUST algorithm is a discrete mathematical method for analyzing continuous data, e.g., mRNA expression.
- For a given array of expression data, the algorithm is applied twice.
- - 1 The first run is on the entire expression matrix and uses a graph theoretic objective function to rank the groups obtained. This pass identifies and ranks a small set of *metagenes* associated with the given cancer.
  - 2 The second run is on the expression matrix for each metagene and supervised by survival time as the objective function using the Fisher score to rank the results. This pass identifies small predictive *signiatures* for each metagene.
- In some cases, certain signiatures would seem appropriate to use as guides for treatment.

# Data Aquisition and Cleaning

- TCGA mRNA expression and clinical data are downloaded from the Broad Institute via the Firehose GDAC portal.

# Data Aquisition and Cleaning

- TCGA mRNA expression and clinical data are downloaded from the Broad Institute via the Firehose GDAC portal.
- Normalized gene expression files sequenced by Illumina HiSeq are used, reporting expression levels for 20,531 genes.

# Data Acquisition and Cleaning

- TCGA mRNA expression and clinical data are downloaded from the Broad Institute via the Firehose GDAC portal.
- Normalized gene expression files sequenced by Illumina HiSeq are used, reporting expression levels for 20,531 genes.
- Samples from tissue surrounding tumors are removed so that each patient has a single record representing tumor tissue.

# Data Acquisition and Cleaning

- TCGA mRNA expression and clinical data are downloaded from the Broad Institute via the Firehose GDAC portal.
- Normalized gene expression files sequenced by Illumina HiSeq are used, reporting expression levels for 20,531 genes.
- Samples from tissue surrounding tumors are removed so that each patient has a single record representing tumor tissue.
- The expression data is log transformed, quantile normalized, and row centered.

# Data Acquisition and Cleaning

- TCGA mRNA expression and clinical data are downloaded from the Broad Institute via the Firehose GDAC portal.
- Normalized gene expression files sequenced by Illumina HiSeq are used, reporting expression levels for 20,531 genes.
- Samples from tissue surrounding tumors are removed so that each patient has a single record representing tumor tissue.
- The expression data is log transformed, quantile normalized, and row centered.
- Survival times and censoring information for each patient are contained in the clinical data and used later in the process.



# Data Discretization

- The expression data is represented by a  $20531 \times N$  real valued matrix **E**, where  $N$  is the number of samples.

# Data Discretization

- The expression data is represented by a  $20531 \times N$  real valued matrix **E**, where  $N$  is the number of samples.
- The matrix **E** is discretized into a  $20531 \times N$  matrix **M** with entries in  $\{-1, 0, 1\}$ .

# Data Discretization

- The expression data is represented by a  $20531 \times N$  real valued matrix  $\mathbf{E}$ , where  $N$  is the number of samples.
- The matrix  $\mathbf{E}$  is discretized into a  $20531 \times N$  matrix  $\mathbf{M}$  with entries in  $\{-1, 0, 1\}$ .
- The desired density  $D$  of non-zero entries in  $\mathbf{M}$  is obtained by adjusting a threshold variable  $\phi$  using the matrix secant method.

# Data Discretization

- The expression data is represented by a  $20531 \times N$  real valued matrix **E**, where  $N$  is the number of samples.
- The matrix **E** is discretized into a  $20531 \times N$  matrix **M** with entries in  $\{-1, 0, 1\}$ .
- The desired density  $D$  of non-zero entries in **M** is obtained by adjusting a threshold variable  $\phi$  using the matrix secant method.
- For this study  $D = 0.5$  for all cancers. In any particular study, one may seek to vary  $D$  to optimize the results.

# Specifications

The LUST algorithm is used to find metagenes (*Part I*), or signiatures (*Part II*).

# Specifications

The LUST algorithm is used to find metagenes (*Part I*), or signatures (*Part II*).

## Input

- Discretized expression matrix **M**.
- Parameters *density*, *confol*, *overlap* and *noregs*.
- For Part II only, clinical data such as survival.

# Specifications

The LUST algorithm is used to find metagenes (*Part I*), or signiatures (*Part II*).

## Input

- Discretized expression matrix **M**.
- Parameters *density*, *confol*, *overlap* and *noregs*.
- For Part II only, clinical data such as survival.

## Output

- Metagenes (Part I) or signiatures (Part II) ranked by an objective function.
- For Part II only, a score placing patients into high and low risk groups.
- For Part II only, Kaplan-Meyer survival curves.

# Regulation and Equivalence

Assume the density  $D$  has been fixed (0.5 in this study). We use *conftol* (in this study 0.75 for Part I and either 0.66, 0.7, or 0.74 for Part II) to adjust sensitivity.



# Regulation and Equivalence

Assume the density  $D$  has been fixed (0.5 in this study). We use *conftol* (in this study 0.75 for Part I and either 0.66, 0.7, or 0.74 for Part II) to adjust sensitivity.

## Definition

For a gene  $X$ , let  $X^+$  denote the number of columns marked with 1 and  $x^-$  the number of columns marked with -1. We say  $X$  *regulates*  $Y$ , denoted  $X \rightarrow Y$ , if

$$① \frac{|X^+ \cap Y^+|}{|X^+|} \geq \text{conftol}, \text{ and}$$

$$② \frac{|X^- \cap Y^-|}{|X^-|} \geq \text{conftol}.$$

# Regulation and Equivalence

Assume the density  $D$  has been fixed (0.5 in this study). We use *conftol* (in this study 0.75 for Part I and either 0.66, 0.7, or 0.74 for Part II) to adjust sensitivity.

## Definition

For a gene  $X$ , let  $X^+$  denote the number of columns marked with 1 and  $x^-$  the number of columns marked with -1. We say  $X$  *regulates*  $Y$ , denoted  $X \rightarrow Y$ , if

$$① \frac{|X^+ \cap Y^+|}{|X^+|} \geq \text{conftol}, \text{ and}$$

$$② \frac{|X^- \cap Y^-|}{|X^-|} \geq \text{conftol}.$$

## Definition

We say gene  $X$  is *equivalent* to gene  $Y$  and write  $X \approx Y$  if  $X \rightarrow Y$  and  $Y \rightarrow X$ .

# Forming Groups

The algorithm begins by computing, for each gene  $X$

$$F_X := \{Y : Y \approx X\}$$

# Forming Groups

The algorithm begins by computing, for each gene  $X$

$$F_X := \{Y : Y \approx X\}$$

Note:  $F_X$  is not necessarily an equivalence class as  $\approx$  is not transitive. Different groups are merged if

$$\frac{|F_X \cap F_Y|}{\min(|F_X|, |F_Y|)} \geq \text{overlap}$$

# Forming Groups

The algorithm begins by computing, for each gene  $X$

$$F_X := \{Y : Y \approx X\}$$

Note:  $F_X$  is not necessarily an equivalence class as  $\approx$  is not transitive. Different groups are merged if

$$\frac{|F_X \cap F_Y|}{\min(|F_X|, |F_Y|)} \geq \textit{overlap}$$

In this study, default values for *overlap* were 0.5 for Part I and 0.6 for Part II. Merging was performed only once.

# Objective Functions

## Part I

For a given metagene  $M$  with  $n$  genes, we consider  $M$  as a directed graph with edges determined by  $X \rightarrow Y$ , and let  $E$  be the set of edges of this graph.

## Part I

For a given metagene  $M$  with  $n$  genes, we consider  $M$  as a directed graph with edges determined by  $X \rightarrow Y$ , and let  $E$  be the set of edges of this graph.

We use a measure of the probability of obtaining a set of vertices of size  $n$  with  $|E|$  edges.

$$f(M) = \frac{|E|}{n-1}$$

# Refinement Using Upstream Regulators

Score every gene  $X$  to measure it's effectiveness regulating the entire set of genes.

$$s_X = \frac{1}{N} \cdot \sum_{X \rightarrow Y} \frac{(|X^+ \cap Y^+| + |X^- \cap Y^-|)^2}{|X^+| + |X^-|}$$



# Refinement Using Upstream Regulators

Score every gene  $X$  to measure it's effectiveness regulating the entire set of genes.

$$s_X = \frac{1}{N} \cdot \sum_{X \rightarrow Y} \frac{(|X^+ \cap Y^+| + |X^- \cap Y^-|)^2}{|X^+| + |X^-|}$$

Let  $G$  be a group from the previous step. For each  $X \notin G$ , consider

$$G_X = \{X\} \cup \{Y \in G : X \rightarrow Y\},$$

# Refinement Using Upstream Regulators

Score every gene  $X$  to measure it's effectiveness regulating the entire set of genes.

$$s_X = \frac{1}{N} \cdot \sum_{X \rightarrow Y} \frac{(|X^+ \cap Y^+| + |X^- \cap Y^-|)^2}{|X^+| + |X^-|}$$

Let  $G$  be a group from the previous step. For each  $X \notin G$ , consider

$$G_X = \{X\} \cup \{Y \in G : X \rightarrow Y\},$$

and assign a score

$$p_{X,G} = \frac{|G_X|}{|G|}(1 + s_X)$$

# Refinement Using Upstream Regulators

Score every gene  $X$  to measure it's effectiveness regulating the entire set of genes.

$$s_X = \frac{1}{N} \cdot \sum_{X \rightarrow Y} \frac{(|X^+ \cap Y^+| + |X^- \cap Y^-|)^2}{|X^+| + |X^-|}$$

Let  $G$  be a group from the previous step. For each  $X \notin G$ , consider

$$G_X = \{X\} \cup \{Y \in G : X \rightarrow Y\},$$

and assign a score

$$p_{X,G} = \frac{|G_X|}{|G|}(1 + s_X)$$

For  $noregs = k$  (default 5), keep  $G_{X_1}, \dots, G_{X_k}$  with the  $k$  highest scores  $p_{X,G}$  for further analysis.

# Objective Functions

## Part II

For each  $G_X$  form a submatrix  $\mathbf{E}_{G_X}$  from the undiscretized expression data.

# Objective Functions

## Part II

For each  $G_X$  form a submatrix  $\mathbf{E}_{G_X}$  from the undiscretized expression data.

Use eigen-survival analysis to produce a predictive score for each patient that is a linear combination of their expression values for  $G_X$ .

# Objective Functions

## Part II

For each  $G_X$  form a submatrix  $\mathbf{E}_{G_X}$  from the undiscretized expression data.

Use eigen-survival analysis to produce a predictive score for each patient that is a linear combination of their expression values for  $G_X$ .

The top and bottom quartiles of the predictive scores are identified and used to calculate Kaplan-Meier expected survival curves.

# Objective Functions

## Part II

For each  $G_X$  form a submatrix  $\mathbf{E}_{G_X}$  from the undiscretized expression data.

Use eigen-survival analysis to produce a predictive score for each patient that is a linear combination of their expression values for  $G_X$ .

The top and bottom quartiles of the predictive scores are identified and used to calculate Kaplan-Meier expected survival curves.

Use the logrank and Cox tests to measure the separation of these two curves. Each test produces a  $p$ -value ( $p_1$  and  $p_2$ , respectively). The *Fisher score* combines these measures to rank how well the signature separates the survival curves.

$$F(G_X) = -\ln(p_1) - \ln(p_2)$$

# Fase Discovery Rates - Notation

Fix a density  $D$ , let  $p = \frac{D}{2}$ , and let  $\gamma = \text{conf tol}$ . Consider an  $m \times n$  matrix with entries from  $\{-1, 0, 1\}$  assigned from uniform probability distributions with densities  $p, p, 1 - 2p$ .



# Fase Discovery Rates - Notation

Fix a density  $D$ , let  $p = \frac{D}{2}$ , and let  $\gamma = \text{conf tol}$ . Consider an  $m \times n$  matrix with entries from  $\{-1, 0, 1\}$  assigned from uniform probability distributions with densities  $p, p, 1 - 2p$ .

Probability row  $X$  has  $a$  entries 1 and  $b$  entries  $-1$

$$g(n, a, b, p) = \binom{n}{a+b} p^{a+b} (1 - 2p)^{n-a-b}$$

Probability row  $Y$  has  $c$  entries 1 in  $a$  columns

$$h(a, c, p) = \binom{n}{c} p^c (1 - p)^{n-c}$$

# Fase Discovery Rates - Derivation

Fix a density  $D$ , let  $p = \frac{D}{2}$ , and let  $\gamma = \text{confol}$ . Consider an  $m \times n$  matrix with entries from  $\{-1, 0, 1\}$  assigned from uniform probability distributions with densities  $p, p, 1 - 2p$ .

# Fase Discovery Rates - Derivation

Fix a density  $D$ , let  $p = \frac{D}{2}$ , and let  $\gamma = \text{confol}$ . Consider an  $m \times n$  matrix with entries from  $\{-1, 0, 1\}$  assigned from uniform probability distributions with densities  $p, p, 1 - 2p$ .

Probability  $X \rightarrow Y$

$$\sum_{1 \leq a, b \leq n} g(n, a, b, p) \left( \sum_{a \geq c \geq \gamma a} h(a, c, p) \right) \left( \sum_{b \geq d \geq \gamma d} h(b, d, p) \right)$$

# Fase Discovery Rates - Derivation

Fix a density  $D$ , let  $p = \frac{D}{2}$ , and let  $\gamma = \text{conf tol}$ . Consider an  $m \times n$  matrix with entries from  $\{-1, 0, 1\}$  assigned from uniform probability distributions with densities  $p, p, 1 - 2p$ .

Probability  $X \rightarrow Y$

$$\sum_{1 \leq a, b \leq n} g(n, a, b, p) \left( \sum_{a \geq c \geq \gamma a} h(a, c, p) \right) \left( \sum_{b \geq d \geq \gamma d} h(b, d, p) \right)$$

Expected number of relations  $X \rightarrow Y$

$$E = m(m-1) \cdot \text{prob}(X \rightarrow Y)$$

# False Discovery Rates - In Practice

Testing on permuted data matrices shows these estimates are quite accurate for values of *conf<sub>tol</sub>* used in this study.

# False Discovery Rates - In Practice

Testing on permuted data matrices shows these estimates are quite accurate for values of *confitol* used in this study.

The probability of random edges or used *confitol* is very low, on the order of  $10^{-5}$  at most.

# False Discovery Rates - In Practice

Testing on permuted data matrices shows these estimates are quite accurate for values of *confitol* used in this study.

The probability of random edges or used *confitol* is very low, on the order of  $10^{-5}$  at most.

The worst-case scenario in this study was cholangiocarcinoma, with only 36 patients. Here  $E$  is about 9,220, but the analysis found 830,000 arrows.

# False Discovery Rates - In Practice

Testing on permuted data matrices shows these estimates are quite accurate for values of *confitol* used in this study.

The probability of random edges or used *confitol* is very low, on the order of  $10^{-5}$  at most.

The worst-case scenario in this study was cholangiocarcinoma, with only 36 patients. Here  $E$  is about 9,220, but the analysis found 830,000 arrows.

For Part II, there are even fewer random arrows expected.



# Sensitivity - Simulations

To test the sensitivity of LUST, simulations were run  $5,000 \times 120$  signal matrix **S** with a step signal in the first 200 rows consisting of 30 entries of 1, 30 entries of  $-1$ , and 60 zeros.

# Sensitivity - Simulations

To test the sensitivity of LUST, simulations were run  $5,000 \times 120$  signal matrix **S** with a step signal in the first 200 rows consisting of 30 entries of 1, 30 entries of  $-1$ , and 60 zeros. A Gaussian noise matrix was made to create  $\mathbf{M} = \mathbf{S} + a\mathbf{N}$ , using  $a$  to adjust signal-to-noise ratio.

# Sensitivity - Simulations

To test the sensitivity of LUST, simulations were run  $5,000 \times 120$  signal matrix  $\mathbf{S}$  with a step signal in the first 200 rows consisting of 30 entries of 1, 30 entries of  $-1$ , and 60 zeros. A Gaussian noise matrix was made to create  $\mathbf{M} = \mathbf{S} + a\mathbf{N}$ , using  $a$  to adjust signal-to-noise ratio. Repeated tests were run at various levels of *conf*tol.

# Sensitivity - Simulations

To test the sensitivity of LUST, simulations were run  $5,000 \times 120$  signal matrix **S** with a step signal in the first 200 rows consisting of 30 entries of 1, 30 entries of  $-1$ , and 60 zeros. A Gaussian noise matrix was made to create  $\mathbf{M} = \mathbf{S} + a\mathbf{N}$ , using  $a$  to adjust signal-to-noise ratio. Repeated tests were run at various levels of *confitol*. The conclusion was that the signals detected by Part I are quite strong.

# Sensitivity - Results

Subtitle

SNR	Rows Found	False Positives
$-10db$	188	0
$-12.5db$	4	0
$-15db$	0	0

Table: *conf*tol = 0.7

# Sensitivity - Results

Subtitle

SNR	Rows Found	False Positives
-10db	188	0
-12.5db	4	0
-15db	0	0

Table: *conf*tol = 0.7

SNR	Rows Found	False Positives
-10db	200	0
-12.5db	196	0
-15db	50	0

Table: *conf*tol = 0.6

# Sensitivity - More Results

Subtitle

SNR	Rows Found	False Positives
$-10db$	200	0
$-12.5db$	200	0
$-15db$	199	4

Table: *conf*tol = 0.5

# Conclusions

- Several metagenes appear to be of interest across multiple types of tumors with several variations. Other metagenes are prominent for only a single kind of tumor.



# Conclusions

- Several metagenes appear to be of interest across multiple types of tumors with several variations. Other metagenes are prominent for only a single kind of tumor.
- Metagenes with signatures that result in the separation of Kaplan-Meier survival curves indicate biological processes of interest.

# Conclusions

- Several metagenes appear to be of interest across multiple types of tumors with several variations. Other metagenes are prominent for only a single kind of tumor.
- Metagenes with signatures that result in the separation of Kaplan-Meier survival curves indicate biological processes of interest.
- Separating tumors by stage results in different metagenes of interest, seeming to indicate that different biological processes become more prominent as the disease progresses.

# Future Investigations

- Using signatures to determine a patient's risk and aggressiveness of treatment (Nation, 2019).

# Future Investigations

- Using signatures to determine a patient's risk and aggressiveness of treatment (Nation, 2019).
- Include methylation and microRNA expression in the analysis.

# Future Investigations

- Using signatures to determine a patient's risk and aggressiveness of treatment (Nation, 2019).
- Include methylation and microRNA expression in the analysis.
- Modify the  $X \rightarrow Y$  relationship to include negative correlation.

# Future Investigations

- Using signiatures to determine a patient's risk and aggresiveness of treatment (Nation, 2019).
- Include methylation and microRNA expression in the analysis.
- Modify the  $X \rightarrow Y$  relationship to include negative correlation.
- Use the algorithm to study continuous data related to other diseases, specifically where the diseased tissue can be isolated and sampled.

## The Last Word

*"LUST is good..."*

# Future Investigations

- Using signatures to determine a patient's risk and aggressiveness of treatment (Nation, 2019).
- Include methylation and microRNA expression in the analysis.
- Modify the  $X \rightarrow Y$  relationship to include negative correlation.
- Use the algorithm to study continuous data related to other diseases, specifically where the diseased tissue can be isolated and sampled.

## The Last Word

*"LUST is good...*

*...and so is the algorithm."* - J.B. Nation

# Future Investigations

- Using signiatures to determine a patient's risk and aggressiveness of treatment (Nation, 2019).
- Include methylation and microRNA expression in the analysis.
- Modify the  $X \rightarrow Y$  relationship to include negative correlation.
- Use the algorithm to study continuous data related to other diseases, specifically where the diseased tissue can be isolated and sampled.

## The Last Word

*"LUST is good...*

*...and so is the algorithm."* - J.B. Nation

Thank you!