Monday, Feb 28

Modeling Counts

Example: Consider the following data.

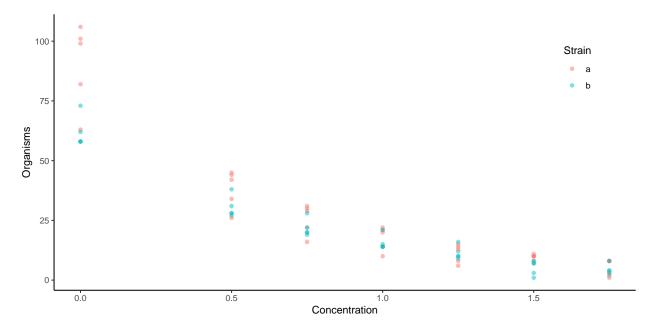
```
library(trtools) # for ceriodaphniastrain data
ceriodaphniastrain$strainf <- factor(ceriodaphniastrain$strain,
    levels = c(1,2), labels = c("a","b"))
head(ceriodaphniastrain)</pre>
```

```
count concentration strain strainf
     82
1
                     0
                             1
2
     58
                     0
                             2
                                      b
3
    106
                     0
                             1
4
     58
                     0
                             2
                                     b
5
     63
                     0
                             2
6
     62
                                     b
```

tail(ceriodaphniastrain)

```
count concentration strain strainf
65
                   1.75
       3
                             1
66
       2
                   1.75
67
       8
                   1.75
                              1
                                      a
68
       8
                   1.75
                              2
                                      b
69
       1
                   1.75
                              1
70
       4
                   1.75
                                      b
```

```
p <- ggplot(ceriodaphniastrain, aes(x = concentration, y = count, color = strainf)) +
   geom_point(alpha = 0.5) + theme_classic() +
   theme(legend.position = c(0.9, 0.8)) +
   labs(x = "Concentration", y = "Organisms", color = "Strain")
plot(p)</pre>
```



What are the complications when the response variable is a count?

- 1. Nonlinear models may be necessary because $E(Y_i) > 0$.
- 2. Heteroscedasticity because $Var(Y_i)$ tends to increase with $E(Y_i)$.
- 3. Non-normal discrete distribution.

One solution would be to use a *nonlinear* regression model combined with some method to account for the heteroscedasticity, and we will revisit this approach, but for now we will consider instead a specialized model that assumes a Poisson rather than a normal distribution of Y_i .

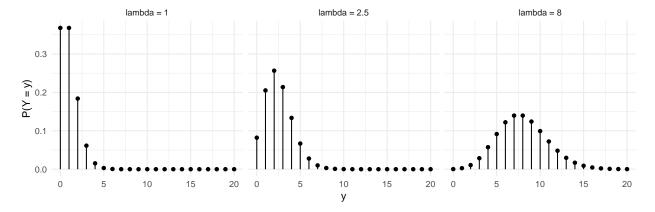
Poisson Regression

A random variable Y has a $Poisson\ distribution$ if

$$P(Y = y) = \frac{\lambda^y e^{-\lambda}}{y!},$$

where y is a non-negative integer (i.e., y = 0, 1, 2, ...) and $\lambda > 0$ is the parameter of the distribution. Also note that y! is the factorial of y, defined as $y! = y \times (y - 1) \times (y - 2) \times ... \times 2 \times 1$ and 0! = 1.

It can be shown that if Y has a Poisson distribution then $E(Y) = \lambda$ and $Var(Y) = \lambda$. The parameter λ is sometimes called a "rate" parameter.



A regression model can be specified for a response variable with a Poisson distribution by assuming that

$$P(Y_i = y) = \frac{\lambda_i^y e^{-\lambda_i}}{y!}$$

where λ_i is a function of $x_{i1}, x_{i2}, \dots, x_{ik}$. Since $\lambda_i > 0$ we might use

$$\lambda_i = \exp(\beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \dots + \beta_k x_{ik}).$$

This implies the *nonlinear* regression model

$$E(Y_i) = \exp(\beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \dots + \beta_k x_{ik}),$$

which can also be written as

$$\log E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \dots + \beta_k x_{ik}$$

This kind of model is sometimes called a *log-linear* model. And because $Var(Y_i) = E(Y_i)$ the model assumes a certain pattern of heteroscedasticity. This kind of regression model is called a *Poisson regression* model.

Generalized Linear Models

Poisson regression is a member of a family of models known as *generalized linear models* (GLM). A generalized linear model has the form

$$g[E(Y_i)] = \underbrace{\beta_0 + \beta_1 x_{i1} + \beta_1 x_{i2} + \dots + \beta_k x_{ik}}_{n_i}$$

where g is the link function and η_i is the linear predictor or systematic component. The link function is invertable so that we can also write

$$E(Y_i) = g^{-1}(\underbrace{\beta_0 + \beta_1 x_{i1} + \beta_1 x_{i2} + \dots + \beta_k x_{ik}}_{\eta_i}) = g^{-1}(\eta_i).$$

Example: A linear regression model can be written as

$$E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_1 x_{i2} + \dots + \beta_k x_{ik}$$

which implies that the link function is the "identity" function g(u) = u, and thus so is the inverse link function $g^{-1}(v) = v$.

Example: A Poisson regression model can be written as

$$\log E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_1 x_{i2} + \dots + \beta_k x_{ik},$$

so the link function is $g(u) = \log(u)$, and the inverse link function is the exponential function $g^{-1}(v) = \exp(v)$, also written as e^v .

In a GLM the variance of Y_i is

$$Var(Y_i) = \phi V[E(Y_i)]$$

where ϕ is a dispersion parameter and V is the variance function. That is, the variance of Y_i is proportional to some function of $E(Y_i)$.

Example: Linear models typically assume homoscedasticity meaning that $Var(Y_i) = \sigma^2$ is a constant. Here the dispersion parameter is $\phi = \sigma^2$, and the variance function is just $V[E(Y_i)] = 1$.

Example: In Poisson regression we have that $Var(Y_i) = E(Y_i)$. Here the dispersion parameter is $\phi = 1$, and the variance function is the identity function $V[E(Y_i)] = E(Y_i)$.

We can write a GLM concisely as

$$E(Y_i) = g^{-1}(\eta_i), \tag{1}$$

$$Var(Y_i) = \phi V[g^{-1}(\eta_i)], \tag{2}$$

where again

$$\eta_i = \beta_0 + \beta_1 x_{i1} + \dots + \beta_k x_{ik},$$

to define the mean structure and a variance structure for Y_i , respectively. In general, specification of a GLM involves specifying three component parts:

- 1. The linear predictor $\eta_i = \beta_0 + \beta_1 x_{i1} + \cdots + \beta_k x_{ik}$.
- 2. The link function g to relate $E(Y_i)$ to η_i as $g[E(Y_i)] = \eta_i$.
- 3. The distribution of Y_i , or the variance structure $\phi V[g^{-1}(\eta_i)]$.

The choice of distribution implies a certain variance structure when using glm. The distribution comes from a family of distributions known as the *exponential family* (not to be confused with what is called an *exponential distribution*).

Example: Normal linear regression is a GLM where g(u) = u, $g^{-1}(v) = v$, $\phi = \sigma^2$, and V(u) = 1 so that

$$E(Y_i) = \eta_i, \tag{3}$$

$$Var(Y_i) = \sigma^2. (4)$$

Example: Poisson regression is a GLM where $g(u) = \log(u)$, $g^{-1}(v) = e^v$, $\phi = 1$, and V(u) = u so that

$$\log[E(Y_i)] = \eta_i,\tag{5}$$

$$Var(Y_i) = \exp(\eta_i). \tag{6}$$

In R the function glm can be used to specify a generalized linear model.

Example: Recall the model for the whiteside data.

```
library(MASS)
m <- lm(Gas ~ Insul + Temp + Insul:Temp, data = whiteside)
summary(m)$coefficients</pre>
```

```
Estimate Std. Error t value Pr(>|t|)
(Intercept) 6.8538 0.13596 50.409 7.997e-46
InsulAfter -2.1300 0.18009 -11.827 2.316e-16
Temp -0.3932 0.02249 -17.487 1.976e-23
InsulAfter:Temp 0.1153 0.03211 3.591 7.307e-04
```

```
m <- glm(Gas ~ Insul + Temp + Insul:Temp, data = whiteside,
  family = gaussian(link = identity))
summary(m)$coefficients</pre>
```

```
Estimate Std. Error t value Pr(>|t|)
(Intercept) 6.8538 0.13596 50.409 7.997e-46
InsulAfter -2.1300 0.18009 -11.827 2.316e-16
Temp -0.3932 0.02249 -17.487 1.976e-23
InsulAfter:Temp 0.1153 0.03211 3.591 7.307e-04
```

Note that we do not explicitly state the variance structure (although we could — more on that later). Here the variance structure is implied by the choice of distribution.

Example: Now consider the following Poisson regression model for the ceriodaphniastrain data.

```
m <- glm(count ~ concentration + strainf, data = ceriodaphniastrain,
  family = poisson(link = log))
summary(m)$coefficients</pre>
```

```
Estimate Std. Error z value Pr(>|z|) (Intercept) 4.455 0.03914 113.819 0.000e+00 concentration -1.543 0.04660 -33.111 2.057e-240 strainfb -0.275 0.04837 -5.684 1.313e-08
```

This model can be written as

$$E(Y_i) = \exp(\beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2}),$$

or

$$\log E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2}$$

where x_{i1} is the concentration for the *i*-th observation, and x_{i2} is an indicator variable for strain such that

$$x_{i2} = \begin{cases} 1, & \text{if the strain is b,} \\ 0, & \text{otherwise,} \end{cases}$$

so that the model can be written case-wise as

$$\log E(Y_i) = \begin{cases} \beta_0 + \beta_1 c_i, & \text{if the strain is a,} \\ \beta_0 + \beta_2 + \beta_1 c_i, & \text{if the strain is b,} \end{cases}$$

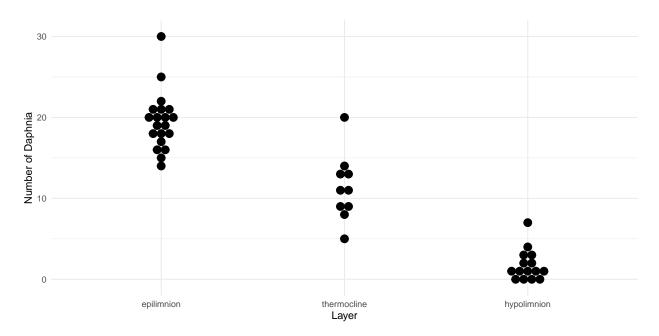
or

$$E(Y_i) = \begin{cases} \exp(\beta_0 + \beta_1 c_i), & \text{if the strain is a,} \\ \exp(\beta_0 + \beta_2 + \beta_1 c_i), & \text{if the strain is b,} \end{cases}$$

if we let $c_i = x_{i1}$ denote the concentration for the *i*-th observation. Also note that because Y_i is assumed to have a Poisson distribution, $Var(Y_i) = E(Y_i)$.

Example: Recall the daphnia survey.

```
library(trtools) # for daphniastrat
library(ggplot2)
p <- ggplot(daphniastrat, aes(x = layer, y = count)) +
  geom_dotplot(binaxis = "y", binwidth = 1, stackdir = "center") +
  labs(x = "Layer", y = "Number of Daphnia") + theme_minimal()
plot(p)</pre>
```



A Poisson regression model for these data might be specified as follows.

```
m <- glm(count ~ layer, family = poisson(link = log), data = daphniastrat)
summary(m)$coefficients</pre>
```

```
Estimate Std. Error z value Pr(>|z|)
(Intercept) 2.9704 0.05064 58.661 0.000e+00
layerthermocline -0.5456 0.10683 -5.107 3.272e-07
layerhypolimnion -2.4204 0.20255 -11.950 6.519e-33
```

So the model is $\log E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2}$, where Y_i is the *i*-th count, and x_{i1} and x_{i2} are defined as

$$x_{i1} = \begin{cases} 1, & \text{if the } i\text{-th observation is from the thermocline layer,} \\ 0, & \text{otherwise,} \end{cases}$$

and

$$x_{i2} = \begin{cases} 1, & \text{if the } i\text{-th observation is from the hypolimnion layer,} \\ 0, & \text{otherwise.} \end{cases}$$

So the model can be written case-wise as

$$\log E(Y_i) = \begin{cases} \beta_0, & \text{if the i-th observation is from the epilimnion layer,} \\ \beta_0 + \beta_1, & \text{if the i-th observation is from the thermocline layer,} \\ \beta_0 + \beta_2, & \text{if the i-th observation is from the hypolimnion layer,} \end{cases}$$

or

$$E(Y_i) = \begin{cases} \exp(\beta_0), & \text{if the } i\text{-th observation is from the epilimnion layer,} \\ \exp(\beta_0 + \beta_1), & \text{if the } i\text{-th observation is from the thermocline layer,} \\ \exp(\beta_0 + \beta_2), & \text{if the } i\text{-th observation is from the hypolimnion layer.} \end{cases}$$

And of course we have that $Var(Y_i) = E(Y_i)$.

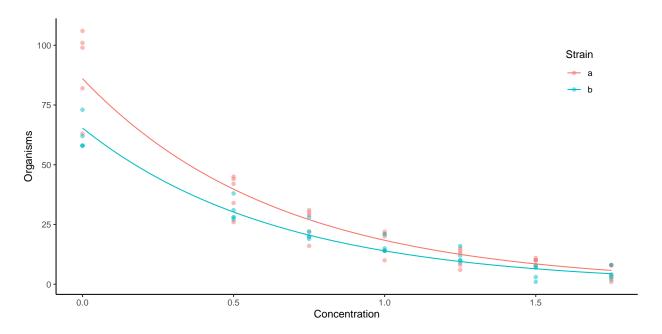
Visualization of a GLM

Visualization of a GLM be done in the usual way provided we use the type = response option when using predict. The default, which is type = link, returns $\hat{\eta}_i = \hat{\beta}_0 + \hat{\beta}_1 x_{i1} + \cdots + \hat{\beta}_k x_{ik}$ which is the estimate of $\log E(Y_i)$.

```
m <- glm(count ~ concentration + strainf, data = ceriodaphniastrain,
    family = poisson(link = log))

d <- expand.grid(concentration = seq(0, 1.75, length = 100), strainf = c("a","b"))
d$yhat <- predict(m, newdata = d, type = "response")

p <- ggplot(ceriodaphniastrain, aes(x = concentration, y = count, color = strainf)) +
    geom_point(alpha = 0.5) + theme_classic() + geom_line(aes(y = yhat), data = d) +
    theme(legend.position = c(0.9, 0.8)) +
    labs(x = "Concentration", y = "Organisms", color = "Strain")
plot(p)</pre>
```



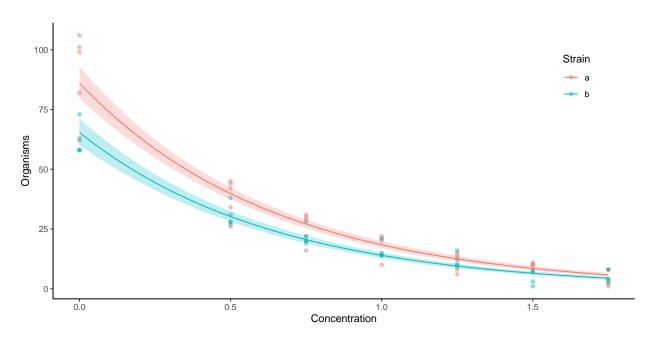
To compute confidence intervals for the expected response you can use the glmint function from the **trtools** package.

```
d <- expand.grid(concentration = seq(0, 1.75, length = 100), strainf = c("a","b"))
d <- cbind(d, glmint(m, newdata = d))
head(d)</pre>
```

```
concentration strainf
                          fit
                                 low
        0.00000
                      a 86.03 79.67 92.88
1
2
        0.01768
                      a 83.71 77.60 90.30
3
        0.03535
                      a 81.46 75.58 87.79
4
        0.05303
                      a 79.27 73.61 85.35
5
                      a 77.13 71.69 82.99
        0.07071
                      a 75.06 69.82 80.68
        0.08838
p \leftarrow ggplot(ceriodaphniastrain, aes(x = concentration, y = count, color = strainf)) +
  geom point(alpha = 0.5) + theme classic() +
  geom_line(aes(y = fit), data = d) +
  geom_ribbon(aes(ymin = low, ymax = upp, y = NULL, fill = strainf),
    color = NA, alpha = 0.25, data = d) +
  theme(legend.position = c(0.9, 0.8)) + guides(fill = FALSE) +
  labs(x = "Concentration", y = "Organisms", color = "Strain")
```

Warning: `guides(<scale> = FALSE)` is deprecated. Please use `guides(<scale> = "none")`
instead.

plot(p)

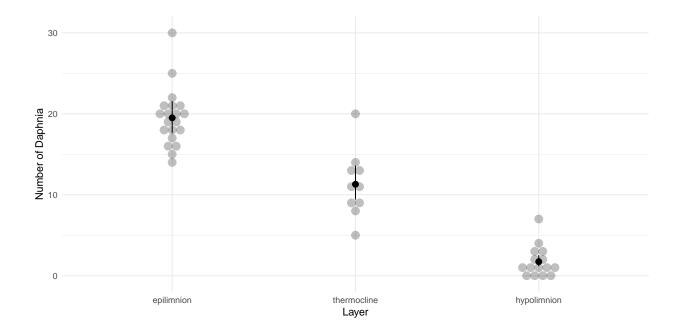


We might visualize the model for the daphniastrat data as follows.

```
m <- glm(count ~ layer, family = poisson(link = log), data = daphniastrat)

d <- data.frame(layer = unique(daphniastrat$layer))
d <- cbind(d, glmint(m, newdata = d))

p <- ggplot(daphniastrat, aes(x = layer, y = count)) +
    geom_dotplot(binaxis = "y", binwidth = 1, stackdir = "center", alpha = 0.25) +
    labs(x = "Layer", y = "Number of Daphnia") +
    theme_minimal() + geom_pointrange(aes(y = fit, ymin = low, ymax = upp), data = d)
plot(p)</pre>
```



GLMs Versus Nonlinear Regression

There is a very close relationship between a GLM and a nonlinear regression model. We can try to estimate the model above using nlm as follows, using the estimates as starting values for convenience.

```
m <- nls(count ~ exp(b0 + b1 * concentration + b2 * (strainf == "b")),
    data = ceriodaphniastrain, start = list(b0 = 7, b1 = -2, b2 = 0.12))
summary(m)$coefficients</pre>
```

```
Estimate Std. Error t value Pr(>|t|)

b0 4.4821 0.02894 154.86 2.486e-87

b1 -1.5679 0.06232 -25.16 7.441e-36

b2 -0.3267 0.04506 -7.25 5.402e-10
```

The estimates are not the same. But consider that the GLM assumes that

$$Var(Y_i) = E(Y_i),$$

so our weights should be $w_i = 1/E(Y_i)$. Consider then an iteratively weighted least squares algorithm with $w_i = 1/\hat{y}_i$.

```
ceriodaphniastrain$w <- 1
for (i in 1:10) {
  m <- nls(count ~ exp(b0 + b1 * concentration + b2 * (strainf == "b")),
    data = ceriodaphniastrain, start = list(b0 = 7, b1 = -2, b2 = 0.12),
    weights = w)
  ceriodaphniastrain$w <- 1 / predict(m, type = "response")
}
summary(m)$coefficients</pre>
```

```
Estimate Std. Error t value Pr(>|t|)

b0 4.455 0.04272 104.273 7.167e-76

b1 -1.543 0.05087 -30.334 7.309e-41

b2 -0.275 0.05280 -5.208 1.988e-06
```

Compare that with what we obtained using glm.

```
m <- glm(count ~ concentration + strainf, data = ceriodaphniastrain,
  family = poisson(link = log))
summary(m)$coefficients</pre>
```

The parameter estimates are the same, but the standard errors are not. Why? The GLM assumes $Var(Y_i) = E(Y_i)$ whereas the iteratively weighted least squares approach assumes $Var(Y_i) \propto E(Y_i)$.

GLMs Versus Response Variable Transformations

It is important to note that, for example, the GLM

$$\log E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \dots + \beta_k x_{ik}.$$

is not equivalent to a linear model with a transformed response,

$$E[\log(Y_i)] = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \dots + \beta_k x_{ik},$$

because $\log[E(Y_i)] \neq E[\log(Y_i)]$ (although in practice they can produce similar results).