Introduction

The microsporidia

Microsporidia is a group of obligate intracellular parasites that infect a broad range of species from vertebrates to invertebrates. They are now classified as sister group of fungi. Microsporidia species have genome sizes from 2,3MB to 19,5MB (Heinz *et al.*, 2012). Most of these parasites have a very small number of genes, from 2000 to 4000 genes, which are thought to be very essential that cannot be lost or they are specific genes of microsporidia. The understanding of the evolution of microsporidia is still unclear and there are many question about the reducing process in their genomes, such as when they lost their genes, if the losing process happened once or many times, if those genes lost forever or just temporarily...

In order to find the answer for those questions, the proteins sets of the microsporidian LCA (LCA) will be inferred using hierarchical orthology inference approach. The distribution of these proteins will be then analyzed to find their origins and to get an overview how common they are in the living organisms. Moreover, we try to investigate the functionality of these microsporidian LCA proteins and identify their roles to understand how they can work together with their hosts' metabolic pathways.

within a few hours of in- fection, a PV forms within the host cell. It is a membrane- bounded organelle that houses developing microsporidial stages (meronts and sporonts) and mature spores in a highly organized manner. Characteristically, developing stages are found near the membrane of the vacuole; the mature spores, which are fully differentiated, lie more centrally. Infected cells house one or more PVs. As the developing stages of the parasite divide and mature within the PV, the host cell enlarges several-fold to accommodate the proliferating parasites. The PVs frequently occupy most of the host cell cytoplasm, squeezing the host cell nucleus, organelles and cytoskeleton to the periphery (Fig. 1B-D). Infection can last for several days before rupture of the membranes of both the host cell and PV is triggered. (Scanlon *et al.*, 2000)

Outside of their host, microsporidia exist as highly durable but largely dormant spores that contain an extremely elabo- rate and highly specialised suite of structures that medi- ate infection [1]. When a spore is triggered, the most dramatic event is the eversion of a tube (the polar tube) that otherwise lays coiled within the spore. This tube can be many times the length of the spore, and when everted in the presence of a potential host, it can adhere to or penetrate the membrane of the host cell, which in turn allows the infectious sporoplasm to pass though the tube and into the host [2]. (Fast and Keeling, 2001)

Note that the host ± parasite relationships are variable among micro- sporidian genera. For example, Enterocytozoon and Nosema are in direct contact with the host cytoplasm while Nucleospora develops within the nuclei of fish lymphoblasts. (Vivarès and Méténier, 2001)

Thus, the haploid genome size was estimated to be 2.5±3.0 Mbp in E. hellem and 2.4 Mbp in E. intestinalis. (Vivarès and Méténier, 2001)

Today, the microsporidia are known to be an extremely diverse group of par- asites. There are currently approximately 150 described genera of microsporidia with over 1200 individual species (78, 79). By far, most microsporidia infect ani- mals, where they have been characterized in all vertebrate orders as well as most invertebrates, including the parasitic myxosporidia (95). / Although widespread among animals, microsporidia are apparently most preva- lent in arthropods and fish. They are used as biological control agents against insect pests and, under natural circumstances, are found to be destructive to apiculture, fish, and some crustacea important to aquaculture (4, 75). The first microsporidian infection described in mammals was Encephalitozoon cuniculi, originally found in rabbits in 1922 (106). / In 1959 the first clear case of a human microsporidian in- fection was recorded (60), but such cases were relatively rare until the mid-1970s, when a dramatic increase in recorded infections accompanied the increased preva- lence of immunosuppressed individuals, either resulting from infection with HIV or due to the use of immunosuppressing drugs (98). The most common of these opportunistic human microsporidia is Enterocytozoon bieneusi, which was first described as a gastrointestinal parasite causing “wasting syndrome” (a potentially lethal diarrhea) in 1985 (20) / Presently, 13 species of microsporidia have been found to infect humans (30, 92, 99) leading to a long list of human diseases, including chronic diarrhea and wasting syndrome, keratoconjunctivitis, pneumonia, bronchitis, nephritis, urethritis, prostatitis, hepatitis, encephalitis, myostitis, and peritonitis (30, 98, 99, 105). (Keeling and Fast, 2002)

Presently, the sizes of 13 microsporidian genomes are known and they fall between 19.5 Mbp and only 2.3 Mbp [(6, 70); for a summary table see (63)]. Apart from their small size, microsporidian genomes are in all characteristics eukaryotic: They have multiple linear chromosomes, telomeres, and segregate by closed mitosis. / The general characteristics of the 2.9-Mbp Encephalitozoon genome mirror what has been observed in other highly reduced eukaryotic genomes (23): Genes are typically flanked by short intergenic regions (although there are no overlapping genes), there are few repeat sequences, little evidence of selfish elements, and few introns. (Keeling and Fast, 2002)

One of the surprising characteristics of the Encephalitozoon genome that demonstrates the extreme degree of reduction is the finding that Encephalito- zoon genes themselves are actually shorter on average than their homologs from other organisms (Katinka *et al.*, 2001)

In essence, selection acts to eliminate any redundancy caused by overlapping functions between the two organisms, leaving only genes that serve some essential function. In the case of parasitic relationships, these genes will include those responsible for ensuring the parasite’s transmission among its hosts. / e.g., N. locustae 5.4 Mb, N. pyrausta 10.5 Mb, and N. bombycis 15.3 Mb (data taken from Méténier and Vivarès, 2001) (Agnew *et al.*, 2003)

Microsporidia are obligate intracellular parasites related to unicellular fungi. These microorganisms are ubiquitous in the animal kingdom, with more than 1,200 species invading a wide range of invertebrates and vertebrates. Among these species is A. algerae, which was originally isolated from the larvae of the Anopheles stephensis mosquito2 and has one of the broadest known host ranges3. / In the microsporidia family, A. algerae has one of the larg- est genome sizes, estimated at 23Mbp7 (Peyretaillade *et al.*, 2012)

Microsporidia are an abundant group of obligate intracellular parasites of other eukaryotes, including immunocompromised humans, but the molecular basis of their intracellular lifestyle and pathobiology are poorly understood. / Core microsporidian genes shared with other eukaryotes are enriched in orthologs that, in yeast, are highly expressed, highly connected, and often essential, consistent with strong negative selection against further reduction of the conserved gene set since the LCMA. Our study reveals that microsporidian genome evolution is a highly dynamic process that has balanced constraint, reductive evolution, and genome expansion during adaptation to an extraordinarily successful obligate intracellular lifestyle. / The number of predicted protein-coding genes varies over a much smaller range of approximately 1,750–3,266 genes, depending on the species and method of analysis (Cornman et al. 2009; Cuomo et al. 2012; Heinz et al. 2012; Peyretaillade et al. 2012). / Although some of these gene differences are of known functional significance, for example, the presence or absence of glycolytic enzymes (Keeling et al. 2010; Peyretaillade et al. 2012), or variation in copy number of the functionally important surface-located nucleotide transport proteins (Tsaousis et al. 2008; Cuomo et al. 2012), most dif- ferences involve hypothetical genes of unknown function (Cuomo et al. 2012; Heinz et al. 2012; Peyretaillade et al. 2012). (Nakjang *et al.*, 2013)

Microsporidia are strict obligate intracellular parasites that infect a wide range of eukaryotes including humans and economically important fish and insects. Surviving and flourishing inside another eukaryotic cell is a very specialised lifestyle that requires evolutionary inno- vation. Microsporidia are fungi-related eukaryotic parasites with over 1,400 reported species that infect a wide range of hosts including humans, mammals, and insects / The parasite cell (meront) grows and multiplies within the host cell cytoplasm through several rounds of division and differentiates into a spore, which exits the host cell, typically through host cell lysis, to complete the cycle (Dean *et al.*, 2016)

Microsporidia represent a large phylum of obligate intracellular pathogens related to fungi, which can infect a diverse array of hosts from protists to humans. They have features consistent with having adapted to proliferate exclusively within the host cellular environment, including greatly reduced genome sizes and the loss of true mitochondria. (Luallen *et al.*, 2016)

Symbiosis life-style

A symbiotic relationship is one in which two dissimilar organisms live in close association with one another. This does not imply that they necessarily live together in harmony; symbiosis also applies to host–parasite relationships. The relationships formed by the microsporidia are extreme forms of symbiosis: these unicellular eukaryotes have become obligate intracellular parasites that cannot live independently outside of their host-cell environment. (Agnew *et al.*, 2003)

Microsporidia are highly specialized eukaryotic unicells, living only as obligate intracellular parasites of other eukaryotes (1). They lack the mitochondria and peroxisomes typical of most eukaryotes. (Hirt *et al.*, 1999)

The microsporidia are a large group of obligate intracellular parasites. Their hosts are especially common among Arthropods. Long evolution of parasites inside of host cell suggests the strong dependence of microsporidia on host metabolism. (Dolgikh, 2000)

are obligate, intracellular protozoan par- asites that infect many vertebrate and invertebrate hosts (Canning and Lom 1986). Until recently, microsporidia were primarily of interest because of their ability to infect insects and fish, in the former case for their potential use in insect control, and in the latter, because of their impact on the fishing industry (Wittner 1999). Prior to the AIDS epidemic, a few isolated reports of human microsporidiosis were described in the literature (Shadduck 1989; Wittner 1999)

The microsporidia are a group of obligately intracellular parasites that infect a broad variety of animal hosts, as well as a small number of protists. (Fast and Keeling, 2001)

The microsporidia are a group of unusual, obligately parasitic protists that infect a great variety of other eukaryotes, including vertebrates, arthropods, molluscs, annelids, nematodes, cnidaria and even various ciliates, myxosporidia and gregarines1. (Vossbrinck *et al.*, 1987)

Microsporidia are small spore-forming unicellular eukaryotes with an obligate intracellular parasitic lifestyle and a worldwide distribution. About 1000 microsporidian species are known; their hosts mainly include various members of almost all animal phyla. In the mid-nineteenth century, the silk worm disease (``peÂ brine'') was caused by a microsporidian Nosema bombycis. In 1922, Wright and Craighead identified a new species (Nosema cuniculi) in brain, spinal chord and kidneys of a paralysed rabbit. This species was assigned in 1923 to a new genus (Encephalito- zoon) by Levatidi et al., and Encephalitozoon cuniculi was the first microsporidian to be propagated in cell culture.(1) Its successful transmission to mice(2,3) facilitated the analysis of host±parasite relationships. Illustrating the propensity of microsporidia for infecting immunocompromised humans, two other Encephalitozoon species (E. hellem, E. intestinalis) have been identified in AIDS patients. (Vivarès and Méténier, 2001)

Nosema bombycis (Naegeli, 1857), was from an insect. This species was later shown by Pasteur (1870) to be the causative agent of pébrine, a disease responsible for serious economic harm to the commercial silk industry at the time; a related species, N. apis, still causes similar problems for commercial honey producers today. / No microsporidia develop outside of their host cells. This pattern strongly suggests that these parasites require their host environment for the uptake of energy and nutrients. (Agnew *et al.*, 2003)

In the most extreme cases, genes are tightly packed with intergenic spaces averaging just over 100 bp, and several pro- tein-coding sequences physically overlap with their neighbours (Corradi *et al.*, 2010)

Origin of microsporidia (check Agnew 2003, corradi 2009)

They lack the mitochondria and peroxisomes typical of most eukaryotes. Thus, in 1983 Cavalier-Smith (2) included the Microsporidia with other amitochondriate protists, Parabasa- lia (e.g., Trichomonas), Metamonada (e.g., Giardia), and Ar- chamoebae (e.g., Entamoeba) in the kingdom Archezoa. These protists were presumed to have diverged from other eu- karyotes before the acquisition of mitochondria and were suggested as the earliest eukaryotic lineages. Phylogenetic trees based initially on small-subunit ribosomal RNA (SSUrRNA) (3) and then on protein translation elon- gation factor (EF-1 and EF-2) (4, 5) sequences showed that Microsporidia indeed diverged early, along with the Parabasa- lia and Metamonada. Thus, these data apparently confirmed the archezoal hypothesis in general and what we call the ‘‘Microsporidia-early’’ hypothesis in particular—Archamoe- bae were eliminated from the Archezoa when their SSUrRNAs neither placed them together nor as early branches (6). However, more recent trees constructed from tubulin (7, 8) and Hsp70 data (9, 10) placed Microsporidia in the eukaryotic ‘‘crown,’’ favoring a position within, or as the sister group to, Fungi. We have determined complete gene se- quences encoding the largest subunit of the RNA polymerase II (RBP1) from two Microsporidia, Vairimorpha necatrix and Nosema locustae. Phylogenetic analyses of these and other RPB1 sequences strongly support the notion that Microspo- ridia are not early-diverging eukaryotes but instead are specifically related to Fungi. (Hirt *et al.*, 1999)

The evolutionary relationship between the micros- poridia (Microspora) and other eukaryotes has been a long-standing and difficult issue to resolve. Based on structural characters, microsporidia have historically been classified with various combinations of other in- tracellular parasites such as myxosporidia, actinomyxi- dia, haplosporidia, and sporozoa (see, e.g., Lom and Va ́- vra 1962; Kudo 1966; Desportes and Nashed 1983). However, many of the characters uniting these groups are associated with their highly specialized modes of parasitism or are present in many other groups of eu- karyotes, and this raises the possibility of convergence. In addition, many of the other characters frequently used for eukaryotic classification (e.g., mitochondria, and 9 2 microtubule structures) are missing or unrecogniz- able in microsporidia, further obscuring their origins. / Eventually, this absence of characters, particularly of the mitochondrion, led to the inclusion of the mi- crosporidia in the Archezoa, a group proposed to de- scend from eukaryotes that lived prior to the acquisition of the mitochondrial endosymbiont (Cavalier-Smith 1983). Initial molecular evidence provided support for this putatively ancient origin of microsporidia by show- ing that they were among the deepest eukaryotic branch- es in trees based on ribosomal RNA (Vossbrinck et al. 1987) and translation elongation factors (Kamaishi et al. 1996a, 1996b). However, these molecular data were not without complications. Like the cells in which they re- side, microsporidian genes tend to be very odd—typi- cally characterized by unique insertions or deletions,strong base-composition bias, and highly accelerated rates of substitution. Genes with accelerated rates are often erroneously placed in phylogenies because of at- traction to other accelerated or divergent sequences (Fel- senstein 1978; Philippe and Laurent 1998). Thus, even considering the apparent agreement between morpho- logical and molecular data, the position of the micro- sporidia was never beyond skepticism (Cavalier-Smith 1993). / as sam- pling of more microsporidian genes soon yielded a strong alternative to their ancient position, namely that microsporidia are somehow related to fungi. Similarities between microsporidian and fungal mitosis and meiosis have been noted over many years (Desportes and The ́o- doride`s 1979; Desportes and Nashed 1983; Flegel and Pasharawipas 1995), but these similarities failed to gen- erate much taxonomic enthusiasm because the shared characters were not unique to these two groups. The first molecular phylogenies to argue for a relationship be- tween microsporidia and fungi were those of alpha- and beta-tubulins, both of which placed the microsporidia within the fungi (Edlind et al. 1996; Keeling and Doo- little 1996). Since then, only one additional gene (again involving the translation apparatus) has been found to support an ancient origin of microsporidia (Keeling, Fast, and McFadden 1998), whereas several more sup- port a fungal relationship. / Altogether, there is a fairly strong body of evidence that microsporidia are related to fungi, but very little evidence that the microsporidia actually evolved from fungi, which is a very significant distinction. Molecular phylogenies that support the relationship between the two are generally only scantily sampled; almost all are restricted to sequences from only one or two micro- sporidia and a few ascomycete fungi (Germot, Philippe, and Le Guyader 1997; Hirt et al. 1997, 1999; Peyre- taillade et al. 1998b; Fast, Logsdon, and Doolittle 1999). In these analyses, the microsporidia are a sister group to fungi but do not actually arise from within the group. Similarly, molecular features uniting microsporidia with fungi really unite them with both animals and fungi (Ka- maishi et al. 1996a, 1996b; Vivare`s et al. 1996). These phylogenies and features are not sufficiently resolved or sampled to address the issue of whether microsporidia are highly derived fungi or only close relatives of fungi. / he resulting beta-tubulin phylogeny is in general agreement with what is believed to be the organismal phylogeny of the two groups and shows that the micro- sporidian beta-tubulins emerge from within the fungal clade. These results provide the first clear demonstration that microsporidia evolved from a fungus. (Keeling *et al.*, 2000)

Their cellular simplicity, in particular the lack of mitochondria, led to the proposal that microsporidia could be one of the earliest eukaryotic lineages, per- haps descended from primitive nucleated cells that lived before typical eukaryotic characters arose [5]. This conclusion was supported by early molecular phylogenies including microsporidian data [6–8], how- ever, it is now clear that this is not the case, and that microsporidia are instead highly derived and extremely divergent fungi. This fungal origin of microsporidia was first proposed based on the relationship between microsporidia and fungi in tubulin phylogenies [9,10], and is now supported by the majority of phylogenetic analyses using a variety of genes (e.g. Refs. [11–15]). (Fast and Keeling, 2001)

The uniqueness of the microsporidia may reflect their phylogenetic position, because comparative sequence analysis shows that the small subunit rRNA of the microsporidium Vairimorpha necatrix7,8 is more unlike those of other eukaryotes than any known eukaryote 18S rRNA sequence. We conclude that the lineage leading to microsporidia branched very early from that leading to other eukaryotes. (Vossbrinck *et al.*, 1987)

Altogether, there are now a number of gene phylogenies that provide robust sup- port for some relationship between microsporidia and other fungi, but what exactly is this relationship? Most genes that have been used to test this have only included ascomycetes and occasionally basidiomycetes. With such poor sampling of fungi, it is not clear from these studies if microsporidia actually are fungi, or if they are merely a closely related sister group of fungi. Unfortunately, only two genes have currently been sampled from diverse fungi to better define this relationship, and these are alpha- and beta-tubulins. In the case of beta-tubulin there is strong support for microsporidia actually evolving from within the fungi, but phylogenies fail to distinguish whether microsporidia are specifically related to ascomycetes or zy- gomycetes (48). Alpha-tubulin also strongly supports microsporidia evolving from within the fungi, but in this case, and in analyses combining both genes, the mi- crosporidia show a specific relationship to zygomycetes. While the exact relationship between microsporidia and fungi remains to be clar- ified, nearly all current evidence does support one major conclusion: Microsporidia are not ancient eukaryotes, but are instead highly evolved fungi. This conclusion colors nearly all other aspects of microsporidia in a new light: No longer are they primitive in lacking mitochondria, flagella, or peroxisomes—these features result from reductive evolution, probably in response to their growing adaptation to intracellular parasitism / In addition to questions of biology, our current interpretations of microsporidian origins are still clouded by our lack of specific information about their ancestors. Although we are confident that microsporidia and fungi are related, the specifics of their relationship await further analysis. (Keeling and Fast, 2002)

To further investigate the evolutionary origin of Microsporidia relative to Fungi, we performed molecular phylogenetic analyses with newly determined RPB1 (DNA-dependent RNA polymerase II largest subunit) and EF-1a (translation elongation factor I alpha) including sequences from basal fungi. Although the phylogenetic position of Microsporidia in the EF-1a tree still might be misplaced due to the unusually high rate of sequence divergence of the microsporidian EF-1a gene, the phylogenies recovered based on these two protein sequences do not provide strong evidences for a close relationship between Microsporidia and Fungi. Moreover, we have identified within the EF-1a genes a characteristic two amino acid deletion conserved in all fungal sequences currently available, whereas this deletion is absent in microsporidian sequences. These results argue against the view of Microsporidia as highly degenerate fungi, and whether Microsporidia and Fungi are sister taxa remains unresolved. (Tanabe *et al.*, 2002)

Their taxo- nomic classification has evolved through time, represent- ing one of the taxa for which the position in the tree of life has been subject to most radical changes [2]. Initially described as “yeast-like fungi” [3], microsporidians were soon re-assigned to Sporozoa, an assemblage of spore- forming protozoa. Later, microsporidia were proposed to be one of the most primitive eukaryotic lineages [4] based on initial electron microscopy studies, / However, during the 1990s, a growing number of phylogenetic studies suggested a close relation- ship between microsporidia and fungi, and revealed that more basal positions of microsporidia were likely the result of long-branch attraction (LBA), a methodological artifact affecting highly divergent sequences [9]. Finally, the sequencing of Encephalitozoon cuniculi [10], and the discovery of microsporidian mitosomes [11], a relic ver- sion of mitochondria, precipitated the re-classification of microsporidia as fungi. / Most of these studies have been limited by the avail- ability of a single microsporidian genome. This, together with the extremely fast-evolving nature of microsporidian sequences, has certainly limited the strength of previous conclusions. / Altogether, our data strongly support microsporidia as the earliest diver- ging clade of sequenced fungi (Capella-Gutiérrez *et al.*, 2012) => incomplete in the choice of taxon sampling (only 3 clade)

The evolutionary relationship of Microsporidia to fungi, either as sister group or internal branch within the fungal radiation, has been extensively debated over the years. Thus, a specific relationship of microsporidians to zygomycete fungi has been suggested [73] based upon the apparent conservation of synteny in E. cuniculi and E. bieneusi, for three genes found at the sex-determining locus of zygomycetes. However, the hypothesis that microsporidia and zygomycetes are specifically related was challenged by analyses [74] showing that the TPT and Hel genes of zygomycetes and Microsporidia were not orthologous, suggesting that the observed synteny was the result of convergence. In summary, the new data from T. hominis provide no support for the hypothesis [73] that microsporidians originate from within the fungal radiation as the specific relatives of zygomycetes. (Heinz *et al.*, 2012)

We propose that Cryptomycota (Rozella allomycis) and microsporidia share a common endoparasitic ancestor (James *et al.*, 2013)

Difficulty for in-vitro experiments

Spores are expected to differ physiologically from other intracellular stages of microsporidian life cycles since this stage is adapted to survival in the environment. However, highly purified microsporidian samples suitable for biochemical studies can currently be obtained only from spores (Dolgikh *et al.*, 1997)

The purpose of this study is to investigate activities of enzymes of carbohydrate and energy metabolism in N. grylli intracellular stages. These data will allow to com- pare rate of carbohydrate metabolism, especially of glycolysis, in spores and intracellular stages. (Dolgikh, 2000)

Unfortunately, biochemical studies undertaken for determining the major molecular compo- nents, nutritional and energy requirements and biosyn- thetic capacities of microsporidia, were and are still strongly hindered by technical difficulties inherent to their obligate development within other eukaryotic cells. Indeed, no extracellular medium insuring in vitro prolif- eration of these parasites exists. Thus, most approaches were either indirect, consisting in comparative analyses of uninfected and infected host cells, or restricted to the free sporal stage, as presented here in the sections dealing with nutrient transport and metabolism. (Méténier and Vivarès, 2001)

The focal point of the microsporidian infection strategy, life history, and diagnosis is the spore, a single, highly organized cell (Figure 1). Spores are the only easily recognizable stage of microsporidia, they are the stage where species can be dif- ferentiated, and they are the only stage of microsporidia that is viable outside of a host cell. (Keeling and Fast, 2002)

The dynamics of reductive genome evolution for eukaryotes living inside other eukaryotic cells are poorly understood compared to well-studied model systems involving obligate intracellular bacteria. (Heinz *et al.*, 2012)

Interesting features

Some other intracellular parasites - Rick- ettsia prowazekii (Winkler, 1976; Plano and Winkler, 1989), Plasmodium falciparum (Kanaani and Ginsburg, 1989; Choi and Mikkelsen, 1990) and probably Toxo- plasma gondii (Sorensen et al., 1997), possess the ATP/ ADP carriers in their plasma membrane, which are similar to those found in mitochondrial membrane. Microsporidiae, the most ancient eukaryotic parasites, might also acquire such a mechanism for harvesting host’s energy. (Dolgikh, 2000)

Microsporidia are one of the more highly adapted groups of eukaryotes known: practically every major feature that distinguishes microsporidia from other eu- karyotes is an adaptation to their parasitic lifestyle. Probably the single characteristic that best defines mi- crosporidia is the severe reduction that permeates every level of the cell, from morphology, to biochemistry, to molecular biology. (Fast and Keeling, 2001)

They possess a number of unusual cytological and molecular characteristics. Their nuclear division is considered to be primitive2, they have no mitochondria3, their ribosomes and ribosomal RNAs are reported to be of prokaryotic size4,5 and their large ribosomal subunit contains no 5.8S rRNA6. (Vossbrinck *et al.*, 1987)

Here we report the DNA sequences of the 11 chromosomes of the D2.9-megabase (Mb) genome of Encephalitozoon cuniculi (1,997 potential protein- coding genes). Genome compaction is re ̄ected by reduced inter- genic spacers and by the shortness of most putative proteins relative to their eukaryote orthologues. The strong host depen- dence is illustrated by the lack of genes for some biosynthetic pathways and for the tricarboxylic acid cycle. (Katinka *et al.*, 2001)

In the 1960s the use of transmission electron micros- copy contributed to a better morphological characteriza- tion of microsporidia, appearing as a curious group of unicellular eukaryotes because of the uniqueness of the sporal invasive apparatus, the lack of mitochondria and centrioles and their Golgi apparatus without any recogniz- able dictyosome. (Méténier and Vivarès, 2001)

Microsporidia were previously considered to be primitive eukaryotes partly because they lacked several “standard” components of a eukaryotic cell (Agnew *et al.*, 2003)

The extremity of this reduc- tion demands a re-evaluation of meta- bolic processes in other microsporidia: although pathways such as glycolysis are present, comparative genomic data sug- gest they may not play the cellular role that they are generally assumed to play. (Keeling and Corradi, 2011)

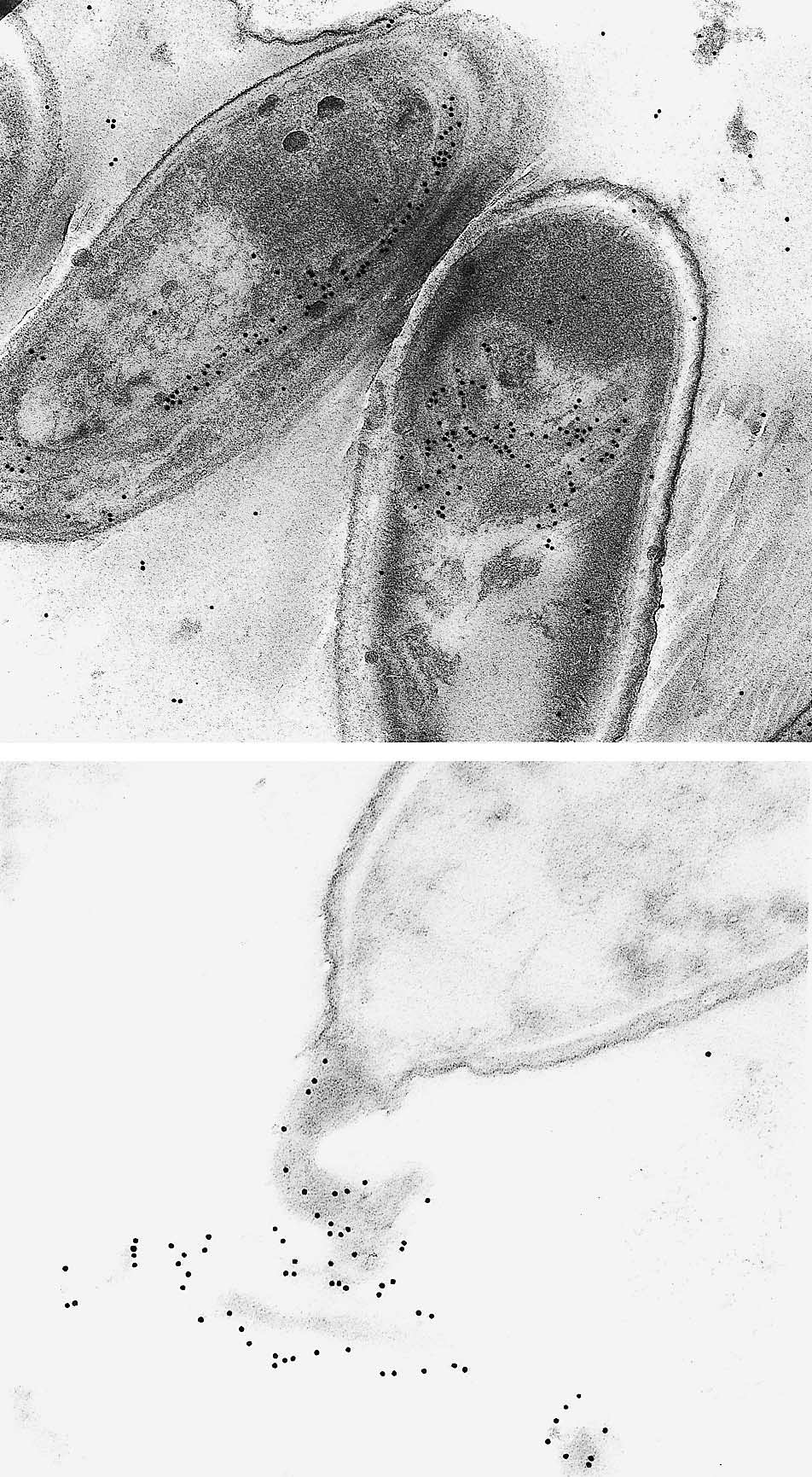


Figure A‑1: Electroimmunolacalization of polar tube proteins in the spore of E.cuniculi. (Méténier and Vivarès, 2001)

The estimation of the microsporidian LCA protein set

Introduction

Method for identifying LCA

Why we need the LCA set

Methods

Starting from 11 extant microsporidia species (Table A‑1), we used OrthoMCL (Li *et al.*, 2003) to search for homologous proteins in those 11 microsporidia species. OrthoMCL performed all-against-all BLASTP comparisons for all input data set and clustered homologous groups using Markov Cluster algorithm MCL (van Dongen, 2000).

The initial homologous groups were then extended by using HaMStR (Ebersberger *et al.*, 2009) to search for orthologs in other 24 search taxa (Table A‑2). The Hidden Markov Model (HMM) profiles for the initial homologous groups (seed sequences) crated by HaMStR were used to search in the search taxa. The obtained hits were confirmed by re-BLAST search against the protein sets of seed sequences. We took into account here also the co-orthologs for re-BLAST and we limited the HMM hits up to only the best 10 hits.

We identified a core gene set, where we found orthologs in all taxa and each taxon has exactly one orthologous protein. Using the core gene set, we reconstructed a maximum likelihood species tree for those 35 taxa. Firstly, we used ClustalW to align all orthologous groups of the core gene set. Then we created a super-alignment by concatenating the alignments of all core genes. To exclude the alignment columns that are uninformative, we removed columns that have at least 50% gaps The phyloge- netic placement of microsporidia has long been contentious and is confounded by an accelerated rate of evolution causing long branch attraction (LBA) [14 (Keeling and Fast, 2002)], but a consensus has recently emerged that microsporidia are related to or derived from within basal fungi. To test for LBA, we sequentially removed the most rapidly evolving sites (James *et al.*, 2013). We used ProtTest (Abascal *et al.*, 2005) to find the best fitting model for the tree reconstruction procedure. With the best model parameters obtained from ProtTest, we used RAxML (Stamatakis, 2014) to build the maximum likelihood species tree with 100 bootstrap replicates.

Using the principle of minimum evolution (Edwards, 1996), we filtered the orthologous group to obtain the final protein set representing the microsporidian LCA. Those final orthologous groups have to have either (1) at least one ortholog from N.parisii (the earliest branch of the microsporidia clade), or (2) at least two orthologs from microsporidia species different than N.parisii and one or more orthologs from non-microsporidia taxa.

Results

OrthoMCL gave 2904 initial homologous groups for 11 microsporidia protein sets. Out of 2904 extended groups, we found 80 groups, where all 11 microsporidia and 24 non-microsporidia taxa are present and each taxon has one representative ortholog. Those 80 groups serve as out core get set for the species tree reconstruction. The super-alignment after de-gapping has a length from 36.616 amino acids. The best model obtained from ProtTest was LG substitution model (Le and Gascuel, 2008), GAMMA distribution G , including proportion of invariable sites estimation I & empirical base frequencies F. The input model parameter for RAxML was PROTGAMMAILGF. The reconstructed species tree with bootstrap support values is shown in the Figure A‑2 below.

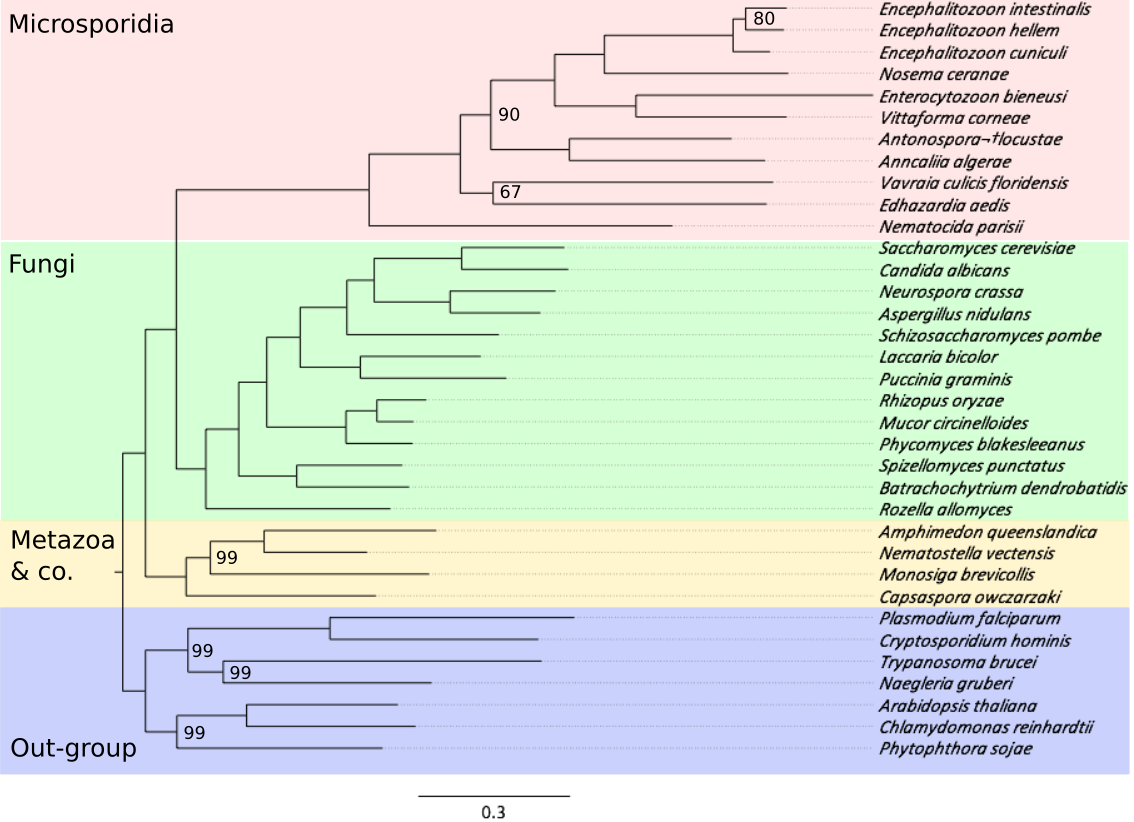


Figure A‑2: Maximum likelihood tree over 35 species. The 11 microsporidia taxa are highlighted in red. Other non-microsporidia taxa include 13 Fungi (green), 2 Metazoa and M.brevicollis, C.owczarzaki (yellow) and 7 out-group species (purple). Node labels denote the bootstrap support and only values <100 are shown.

Filtered the HaMStR result that did not match the parsimony criteria, we got at the end 1605 final orthologous groups. They present the set of microsporidian LCA proteins.

Discussion

Figure A‑3 shows the fractions of non-orthologous and orthologous proteins in 11 microsporidia species. The Encephalitozoon group is the best example for the compact genome of microsporidia. Where almost 98% of their proteome are orthologous proteins that are shared in other microsporidia species. Only 2% of genes are lineage specific proteins.

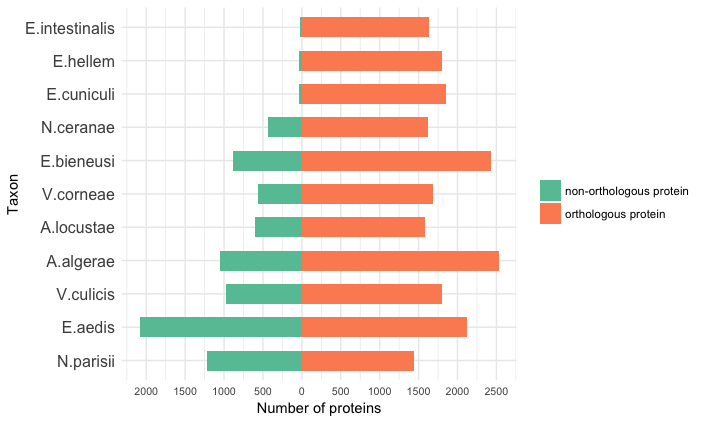


Figure A‑3: Fractions of non-orthologous (orange) and orthologous (green) proteins in different microsporidia species.

Other taxa still have orphan proteins (21% in N.ceranae up to 49% in E.aedis). We have some hypotheses for those orphan proteins.

(1) Wrong gene assignment: length of orphan proteins would be shorter than orthologous proteins.

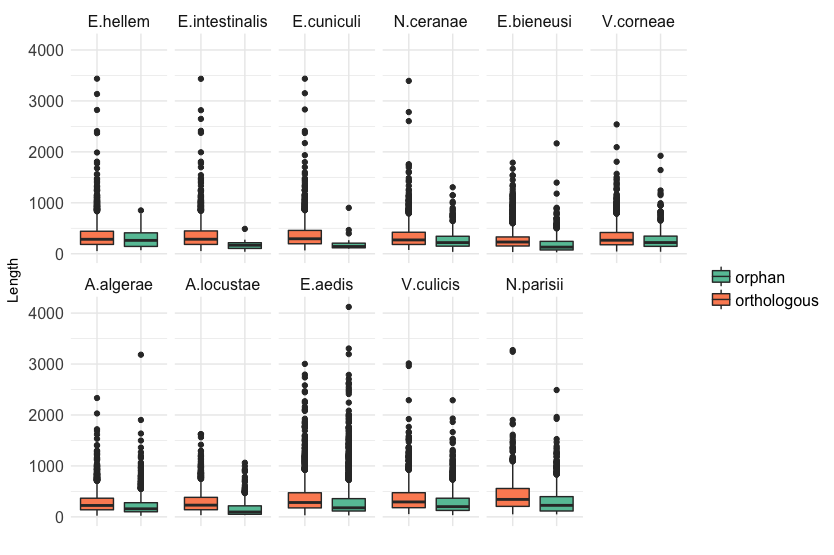


Figure A‑4: Length distribution of orthologous proteins (orange) and orphan proteins (green) in different microsporidia taxa.

Figure A‑4 shows the length distribution of orthologous and orphan proteins in 11 microsporidia taxa. We use Wilcoxon-Mann-Whitney U-Test (is it the same as paired Mann-Whitney?) to compare the two length categories. We found that the lengths of orphan proteins are significantly different (smaller) to the one of orthologous proteins with the significant level of 5%. Only for E.hellem the p-value is 0,20>0,05. But this p-value makes no sense since the number of orphan proteins in E.hellem is too small to make the comparison meaningful.

(2) New invented genes (which have no PFAM annotations), or genes from horizontal gene transfer events (which have new PFAM annotation, which are not found in orthologous proteins), or they cannot be detectable as orthologs (which have the same PFAM annotations as orthologous proteins).

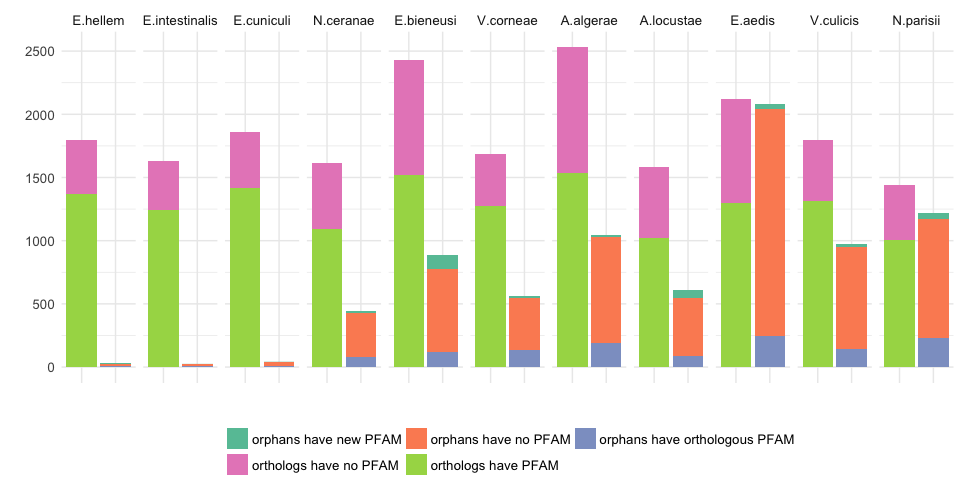


Figure A‑5: Fractions of orthologous and orphan proteins that have and do not have PFAM annotations. The left columns show the number of orthologous proteins that have PFAM annotations (light green) and do not have PFAM annotations (pink). The right columns show the proportion of orphan proteins that have new PFAM annotations that are not found in orthologous proteins (dark green), do not have any PFAM annotation (orange) and orphans that have the same PFAM annotations as orthologous proteins (purple).

A large fraction of orphan microsporidia proteins do not have any PFAM annotation as been shown in Figure A‑5 suggests that most of the orphan proteins are new invented genes after the speciation event that split fungi out of the microsporidia clade.

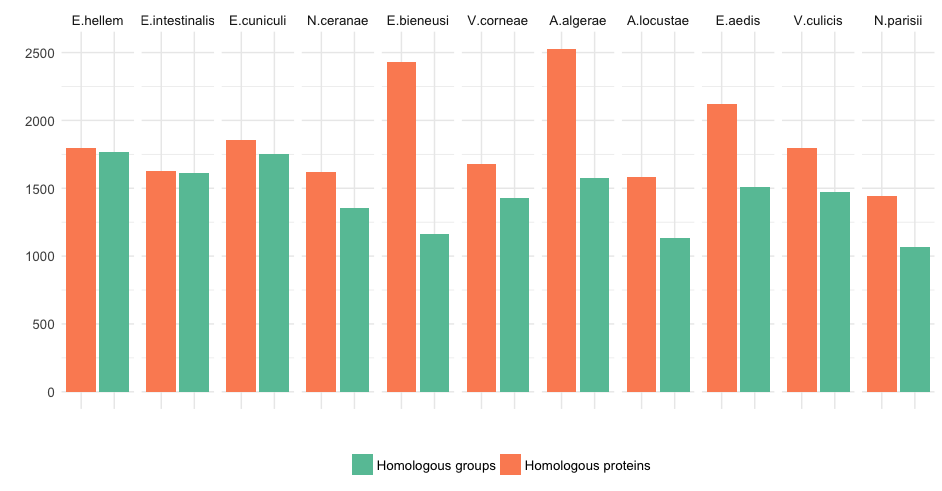


Figure A‑6: The distribution of number of homologous proteins (orange) and number of homologous groups (green) in each microsporidia taxon.

Figure A‑6 shows the relative number of homologous proteins and number of homologous groups in each microsporidia species. In some species, such as E.bieneusi or A.algerae, the number of homologous genes is substantially higher than the number of corresponding homologous groups. We check the number of in-paralogs for each microsporidia taxon in the homologous group. The result in Figure A‑7 shows that there is no evidence for whole genome duplication in any species. But there are some instances where the homologous groups contain more than 10 co-orthologs for one microsporidia species showing the effect of gene dosage (Kondrashov and Koonin, 2004).

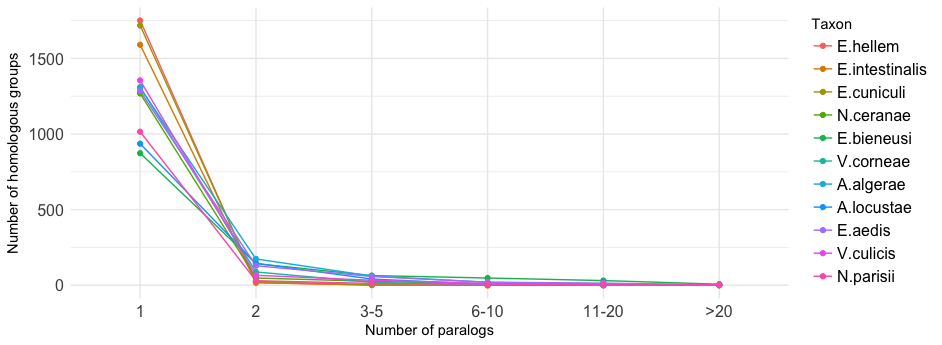


Figure A‑7: The distribution of number of homologous groups as a function of number of in-paralogs. Colors denote different microsporidia taxa.

80 core genes and the origin of microsporidia

The 80 core genes, which are used for reconstructing the species tree in Figure A‑2, shows to be a very good seed set for studying the evolutionary of fungal or even eukaryotic species. It can be used to create very good resolved species trees for a large set of fungal species (ref.) or to investigate the co-evolution of PDI/RhoGID gene clusters (protein disulfide isomerases and Rho guanine-dissociation inhibitors) across the animal phylogeny (Moretti *et al.*, 2017).

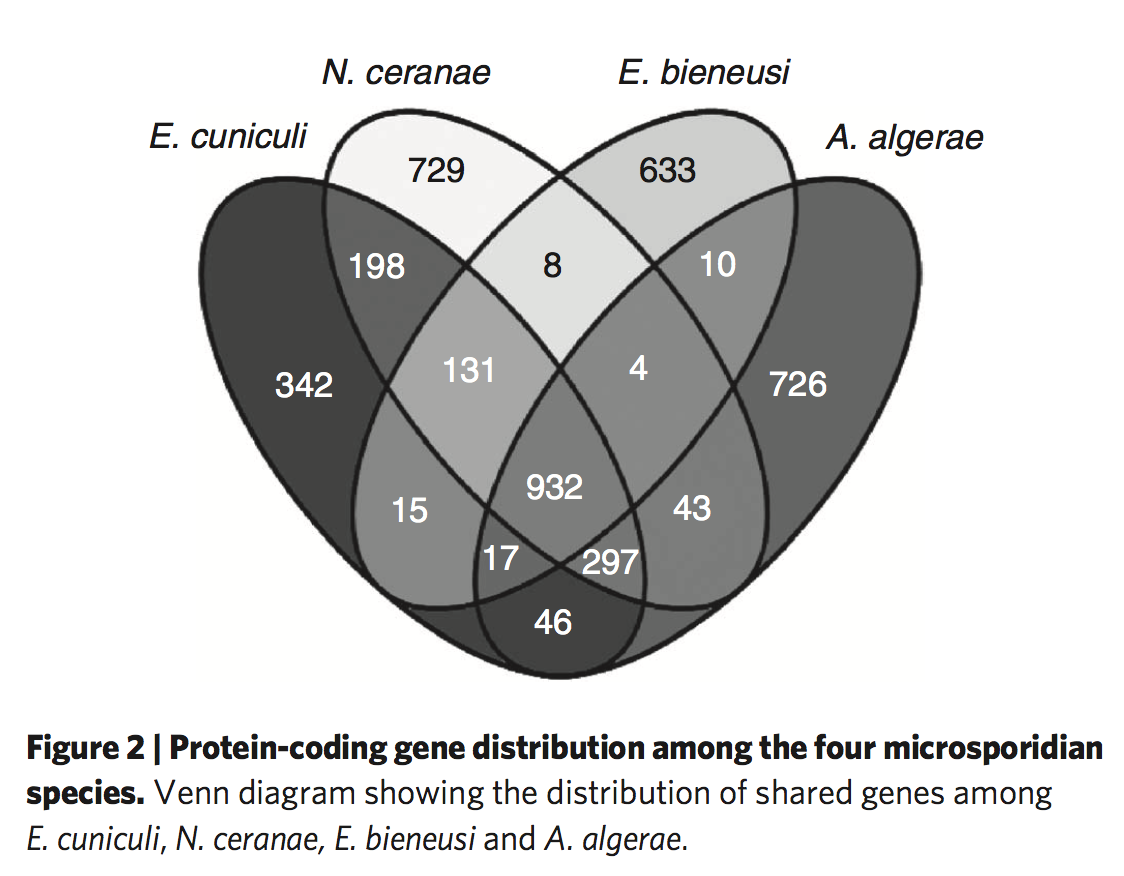
More about microsporidia origin (Keeling and Fast, 2002) => put in introduction section

The reconstructed species tree shows a stable signal for placing the microsporidia group as the earliest clade of the fungi, which are supported by the recent phylogenetic study of the taxonomy of microsporidia that hypothesis that microsporidia is the sister clade of fungi (Hirt *et al.*, 1999).

Tree of fungal analysis?

Conclusion

The estimation of microsporidian LCA proteins is the basic step for the whole downstream study. The orthology assignment result agreed with other studies about the fraction of microsporidia only proteins. The length distribution and PFAM annotation analysis indicate that most of the orphan microsporidia proteins are either resulting by wrong gene assignment or they are new invented genes in the microsporidia clade. The good resolved species tree can serve as a fundamental phylogenetic background for filtering the orthology assignment and estimating the set of 1605 proteins for the LCA of microsporidia. This tree supports the hypothesis that microsporidia is a sister group of fungi.



(Peyretaillade 2012)

Distribution analysis of the microsporidian LCA proteins

Introduction

Why: to answer the question "How old are the microsporidian LCA proteins"

Methods

We used HaMStR to search orthologs for 1605 microsporidian LCA proteins in 480 taxa across the tree of life including bacteria, archaea and eukaryote, which are grouped into 44 super taxa as you can see in this schematic species tree in the Figure A‑8. The list of all the taxa under this analysis is written in the Table A‑4. The options we used for HaMStR search are -strict, -checkCoorthologsRef, -hit\_limit=10 and -representative.

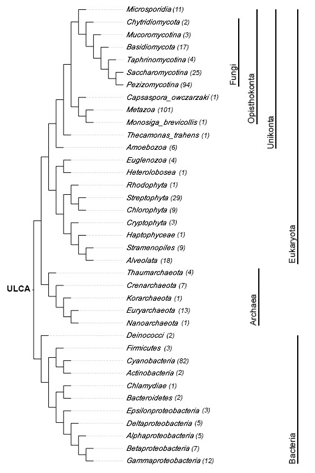


Figure A‑8: A cladogram depicts a species tree for all taxa used in the distribution analysis of microsporidian LCA proteins. The number in parenthesis next to the taxon names denotes the number of species in each supertaxon.

For a comprehensive analysis and to complement the orthology assignment, we calculated the feature architecture similarity (Koestler *et al.*, 2010) scores (FAS scores) for all pair microsporidia seed proteins and non-microsporidia orthologs. Feature architecture of a protein is the arrangement of different types of protein domains such as PFAM (Finn *et al.*, 2014) or SMART (Letunic *et al.*, 2012) domains, transmembrane domains, low complexity regions, secondary structures, etc. Comparison of feature architecture between two proteins gives a FAS score between 0 and 1. The higher the FAS score, the more similar those 2 proteins are in term of functional equivalence.

To visualize the phylogenetic profiles of 1605 microsporidian LCA proteins, we developed a tool named PhyloProfile. This tool is written in R (R Development Core Team, 2011) using the Shiny library (https://CRAN.R-project.org/package=shiny). Beside the presence / absence pattern of genes across species, PhyloProfile is able to display two additional layers of information. In particularly, PhyloProfile enables the visualization and exploration of phylogenetic profiles together with the protein feature architectures in an informative and interactive way. Implemented with the dynamic filtering option, PhyloProfile can offer a reliable analysis of phylogenetic profiles with its analysis functions.

We used PhyloProfile to estimate the evolutionary age for microsporidian LCA proteins.

Results

PhyloProfile appears to be vey handy for exploring the informative phylogenetic profile with complementary information.

Almost orthologous proteins have similar feature architectures with microsporidia proteins. It leads to a very high mean FAS score of 0.958 (see the FAS score distribution in Figure A‑9).



Figure A‑9: The distribution of FAS scores for all orthologs of 1605 microsporidian LCA proteins.

Figure A‑10 shows the full phylogenetic profile of 1607 microsporidian LCA proteins across 491 taxa that are grouped into phylum level. A large fraction of microsporidia proteins spread through all studied taxa.

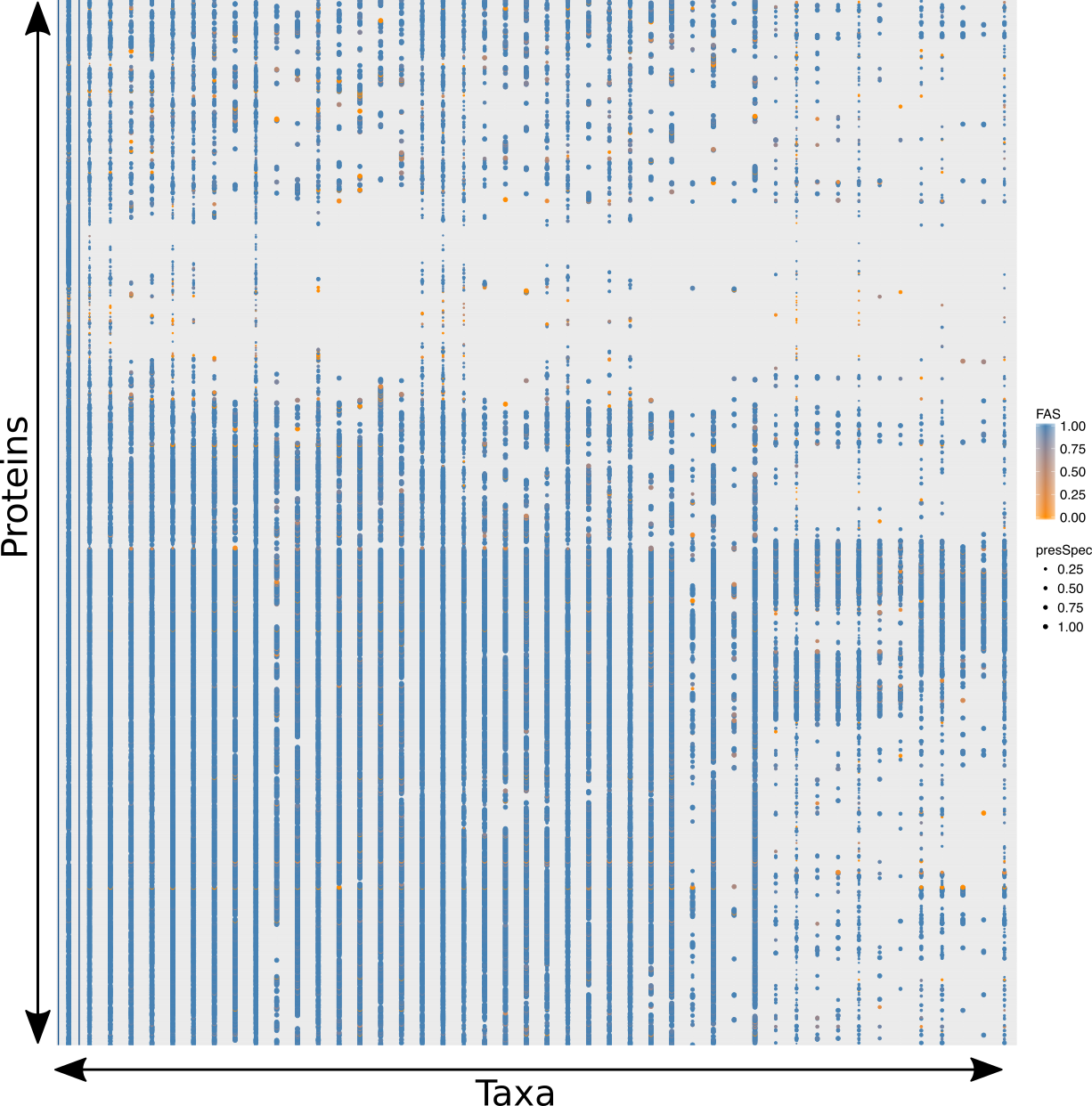


Figure A‑10: The full phylogenetic profile of 1605 microsporidian LCA protein across 491 taxa grouped in phylum level. The color of the points denotes the FAS score between microsporidia and non-microsporidia protein. The size of the points depicts the percentage of species that have orthologs in each phylum.

Using PhyloProfile we estimated the evolutionary ages for microsporidian LCA proteins. The estimation result can be seen in Figure A‑11. As expected, 94% of the proteins are as old as the last eukaryotic common ancestor, while only 3% are specific to microsporidia lineage.



Figure A‑11: Gene age estimation of 1605 microsporidian LCA proteins. The fraction and corresponding absolute number of proteins for each estimated evolutionary age are written in each block. The colors denote the estimated ages for query proteins.

Out of 42 microsporidia specific proteins, only 6 have KO annotations (see Table A‑1) or 8 if combine HamFAS and BlastKOALA.

Table A‑1: KO annotation for 42 microsporidia specific proteins using HamFAS approach

|  |  |  |
| --- | --- | --- |
| LCA protein | KO annotation | Description |
| OG\_1349 | K18592 | gamma-glutamyltranspeptidase / glutathione hydrolase / leukotriene-C4 hydrolase |
| OG\_1378 | K09485 | heat shock protein 110kDa |
| OG\_1710 | K04802 | proliferating cell nuclear antigen |
| OG\_2013 | K02155 | V-type H+-transporting ATPase 16kDa proteolipid subunit |
| OG\_2250 | K02896 | large subunit ribosomal protein L24e |
| OG\_2280 | K02180 | cell cycle arrest protein BUB3 |

Table A‑2: KO annotation for 42 microsporidia specific proteins using BlastKOALA

|  |  |  |
| --- | --- | --- |
| LCA protein | KO annotation | Description |
| OG\_1087 | K17866 | diphthamide biosynthesis protein 2 |
| OG\_1378 | K09485 |  |
| OG\_1378 | K09489 | heat shock 70kDa protein 4 |
| OG\_1515 | K08803 | death-associated protein kinase |
| OG\_1710 | K14848 | ribosome assembly protein RRB1 |
| OG\_2013 | K02155 |  |
| OG\_2250 | K02896 |  |
| OG\_2280 | K02180 |  |

Gene Ontology terms (Ashburner *et al.*, 2000) were assigned by Blast2GO v5.0.13 (Götz *et al.*, 2008).

...

Discussion

Not out of our expectation, due to the compact genomes of extant microsporidia taxa, most of the proteins in the microsporidian LCA should be evolutionary old. As 50% of the proteins are as old as the last universal common ancestor, another 44% proteins can be traced to the LCA of all eukaryotes and 3% share the age with fungal clade, only 3% (or 42 proteins) are specific to microsporidia lineage. Those microsporidia specific proteins are still a mystery because of the poor functional annotation. There is no particular function or pathway that have been enriched by those proteins as has been seen from our KO and GO assignment analysis.

Conclusion

Functional annotation

Introduction

Proteins that are orthologous to each other are likely to have similar functions. The quality of orthology-based annotation transfer methods depends strongly on the accuracy of the ortholog prediction. Here we are introducing HamFAS, a robust annotation transfer pipeline based on feature-aware orthology inference. HamFAS has been shown to have higher sensitivity and comparable specificity in comparison to two state-of-the-art annotation tools KAAS and BlastKOALA from KEGG. A feature that makes HamFAS different than BlastKOALA and KAAS is the controllable ability of the annotation process. Users can choose different methods and threshold for increase or reduce the stringency of the annotation pipeline. Besides, HamFAS can be run locally through command lines. It provides a better solution for large-scale analysis than using online tools like KAAS and BlastKOALA.

Methods

HamFAS approach

We developed a novel approach named HamFAS to transfer KO annotations based on feature-aware orthology inference. Figure A‑12 demonstrates the pipeline of HamFAS.

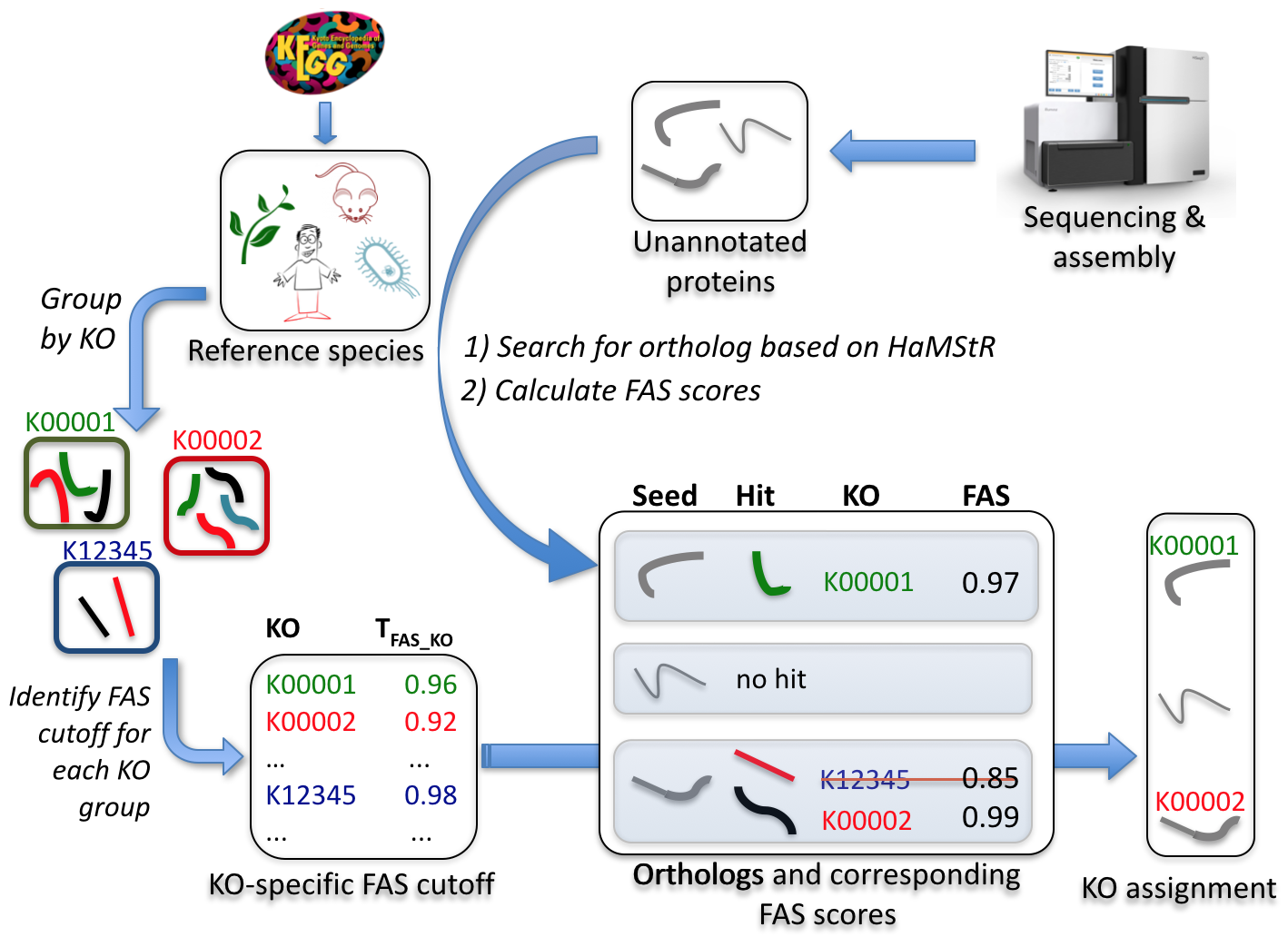


Figure A‑12: KO annotation transfer using HamFAS approach.

Protein sets of 30 manually KO-annotated reference species (Table A‑5) have been downloaded from KEGG database. Pairwise FAS scores of all reference proteins within a KO group have been calculated. A group's mean FAS score serves then as a cutoff (TFAS\_KO) that must be exceeded to warrant transfer of its KO identifier to the seed proteins.

Given a list of uncharacterized proteins (seed), we search for their orthologs in the reference species using HaMStR (with *-checkCorothologsRef*, *-rbh* options and *-hit\_limit=5*). FAS scores between seed proteins and their orthologs will be identified. If the calculated FAS scores is not smaller then the corresponding TFAS\_KO, the available KEGG identifiers of those paired orthologs will be transferred to the seed proteins.

Benchmarking HamFAS

We used *S.cerevisiae* as a test species to benchmark our approach HamFAS. The protein set of yeast has been obtained from KEGG containing 3457 KO-annotated and 3158 un-annotated sequences. The annotated proteins have been used for evaluating the accuracy of the approach, while the un-annotated set has been used for estimating its sensitivity. The output of HamFAS is also compared with KAAS and BlastKOALA.

We removed *S.cerevisiae* out of the reference species list for avoiding redundant information while doing orthology search. The same reference species have been used for KAAS approach. With BlastKOALA, however, we couldn't remove yeast annotations out of the reference data.

The ortholog search has been also performed with different parameters to find the best settings for HaMStR (*-rbh*, *-checkCoorthologRef*).

Results

Distribution of TFAS\_KO

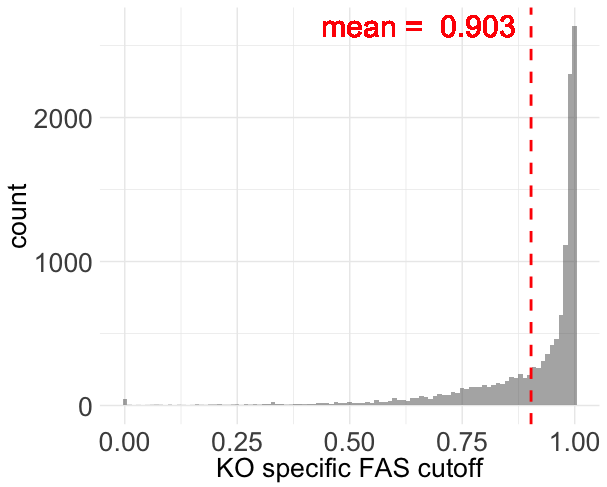


Figure A‑13: Distribution of TFAS\_KO for 12,748 KO groups

Figure A‑13 represents the distribution of all 12,748 TFAS\_KO values. Only about 3% of KOs have TFAS\_KO smaller than 0.5, 27% lie between 0.5 and 0.9, while 70% has TFAS\_KO greater than 0.9. The low TFAS\_KO values are caused mostly by the uninformative protein members. Figure A‑14 shows 2 examples, the FAS scores distribution of K00542, which represents low TFAS\_KO group, and K07888, which represents high TFAS\_KO group.

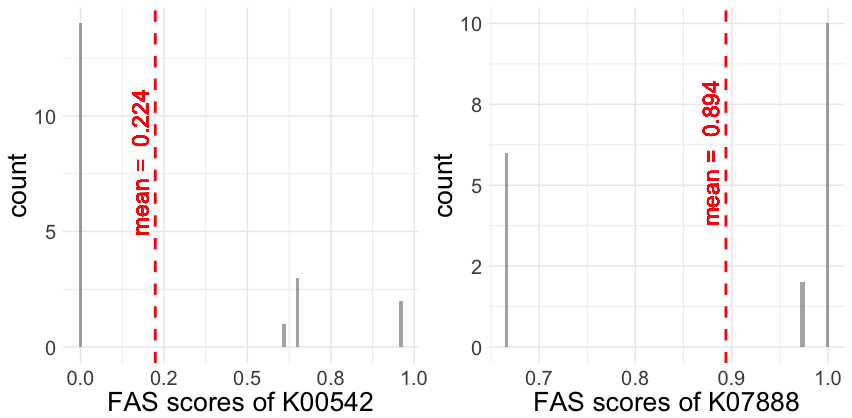


Figure A‑14: FAS score density of KO group K00542 (left) and K07888 (right)

In ortholog group K00542 (guanidinoacetate N-methyltransferase), only one protein member (rat rno:25257) has one Pfam domain (Orn\_DAP\_Arg\_deC). The lack of Pfam domain annotation of other proteins (human hsa:2593, mouse mmu:14431, zebrafish dre:796865 and *N.vectensis* nemve:1432) caused FAS scores of 0 for 14/20 pairwise comparisons and led to the low TFAS\_KO (0.224) for the whole group. On the contrary, the rich annotation of protein members of group K07888 (Ras-related protein Rab-5B) is the reason for its high TFAS\_KO.

Benchmarking result

3457 KO-annotated yeast proteins

With this data set, we tried to evaluate the accuracy of HamFAS in comparison to KAAS and BlastKOALA by calculating the recall, precision and F1 score.

recall = TP / (TP + FN)

precision = TP / (TP + FP)

F1 = (2\*precision\*recall)/(precision+recall)

Table A‑3 shows the evaluations of HamFAS, BlastKOALA and KAAS. HamFAS performed best in term of precision, while F1-score is lower then KAAS due to its lower recall. Interestingly, the latest annotation tool from KEGG, BlastKOALA, has the lowest scores in both recall and precision.

Table A‑3: Recall, precision and F1-score of HamFAS in comparison to BlastKOALA and KAAS. Second column shows values of HamFAS after filtering the orthology assignment with InParanoid's orthologs.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Approach** | **HamFAS** | **supported\_HamFAS** | **BlastKOALA** | **KAAS** |
| Recall | 0.915 | 0.861 | 0.905 | 0.931 |
| Precision | 0.985 | 0.985 | 0.979 | 0.984 |
| F1-score | 0.949 | 0.919 | 0.940 | 0.957 |

For checking the ortholog prediction result obtained by HaMStR, we evaluated the annotation transfer again using only orthologs that are supported by both HaMStR and InParanoid. Predicted KOs from HamFAS of 188 yeast proteins has been removed after filtering based on InParanoid's orthologs. It leads to the decrease of recall and F1-score. However, the precision is not affected (see Table A‑3). FAS scores of unsupported orthologs are slightly smaller than the ones of supported orthologs, with mean score of 0,918 and 0,988 respectively (see Figure A‑15).

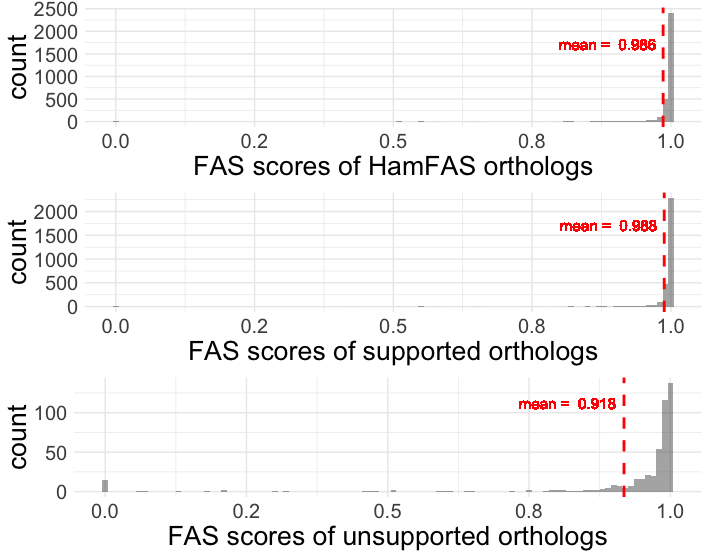


Figure A‑15: FAS score distribution of all HamFAS orthologs, only supported orthologs and unsupported orthologs

For a more detailed comparison between 3 approaches, we compare the fractions of proteins annotated by HamFAS, BlastKOALA and KAAS. 85,6% of the seed proteins has been annotated by all 3 approaches (Figure A‑16).

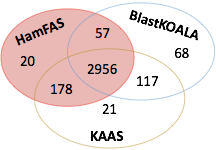


Figure A‑16: Fraction of proteins annotated by HamFAS, BlastKOALA and KAAS

There is a small difference between the KEGG identifiers annotated by each approach, which is shown in Table A‑4 below.

Table A‑4: Compare KEGG identifiers annotated by HamFAS, BlastKOALA and KAAS. Number in parentheses are the different KOs after filtered by synonymous KOs.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Approach** | **All 3 approaches** | **HamFAS +**  **BlastKOALA** | **HamFAS + KAAS** | **KAAS +**  **BlastKOALA** |
| Same KOs | 2951 | 54 | 168 | 108 |
| Diff. KOs | 5 (1) | 3 (1) | 10 (5) | 9 (6) |
| Total | 2956 | 57 | 178 | 117 |

Although those KEGG identifiers are different, most of them are "synonymous" KOs. They either have the same EC numbers, same EC classes, same GO numbers, or are the same components in KEGG pathways, responsible for the same reactions, etc.

Some examples of synonymous KOs:

* 1 KO is very general described (putative ABC transport system ATP-binding protein) while the other is specific (phospholipid/cholesterol/gamma-HCH transport system ATP-binding protein).
* Synonym/Alternative name: "septin" and "sporulation-regulated protein 3" (also septin); or "tristetraprolin" (ZFP36) and "butyrate response factor 1" (ZFP36L1).
* Involved in the same process: "cleavage stimulation factor subunit 2" and "polyadenylate-binding protein 2" are involved in 3-end formation of pre-mRNAs

3158 un-annotated yeast proteins

HamFAS could annotate 257 proteins, in which 164 proteins are HamFAS specific (HamFAS-only annotated proteins) (see Figure A‑17). In comparison to 150 and 116 annotated proteins from KAAS and BlastKOALA, HamFAS has annotated more proteins than BlastKOALA and KAAS (257 proteins versus 116 and 150 proteins, respectively)~~.~~

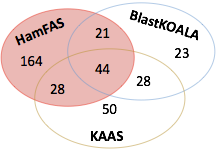


Figure A‑17: Fraction of proteins annotated by HamFAS, BlastKOALA and KAAS

Here the interesting part is the HamFAS-only proteins. So what are the differences of those proteins in comparison to others?

Discussion

The specificity of HamFAS

As we have seen from the analysis of the KO-annotation yeast protein set, HamFAS yielded the best precision regardless the supported or non-supported orthology assignment by InParanoid. It indicates the reliability of the annotation transfer result of HamFAS.

The sensitivity of HamFAS

Beside the best specificity, HamFAS also shows the highest sensitivity in comparison to BlastKOALA and KAAS with the highest number of proteins that could be annotated.

Was HaMStR so inclusive to include many false positive orthologs?

We also compared the orthology search of HamFAS with InParanoid. After removing ortholog pairs that are not predicted by InParanoid, 150 out of 257 proteins still can be annotated, 55 of them belong to HamFAS-only annotated proteins. It proved the reliability of the orthology assignment of HamFAS-only proteins.

Are HamFAS-only proteins short and uninformative?

We compared the sequence length and the informative content of protein domains between HamFAS-only proteins and proteins annotated by at least 2 approaches including HamFAS and KAAS and/or BlastKOALA.

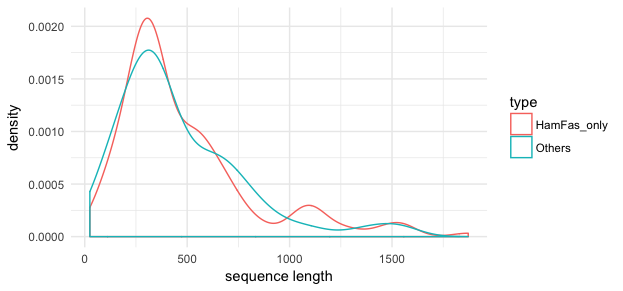


Figure A‑18: Length distribution of HamFAS-only proteins and others

Figure A‑18 and Figure A‑19 show no clear difference between those 2 protein sets. HamFAS-only proteins are not either extremely shorter or longer than other proteins. And the annotation transfer result was not driven by the uninformative domain annotation of those proteins (one Pfam domain that leads to the high FAS score).

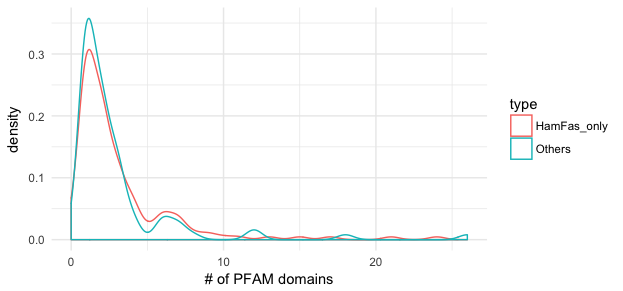


Figure A‑19: Number of Pfam domains distribution of HamFAS-only proteins and others

The distribution of FAS scores of all HamFAS orthologs in comparison to HamFAS-only orthologs shown in Figure A‑20 also confirms the rich domain annotations of HamFAS-only proteins.

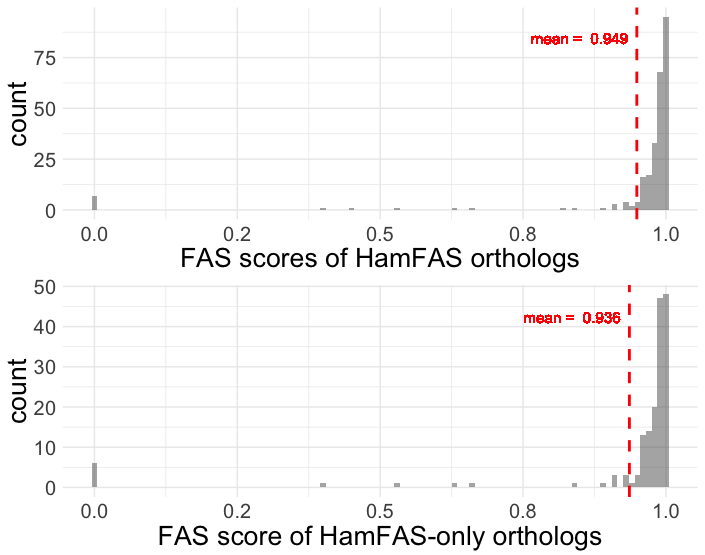


Figure A‑20: FAS score distribution of all HamFAS orthologs and HamFAS-only orthologs

How different are the phylogenetic profile of KO-annotated proteins and un-annotated protein?

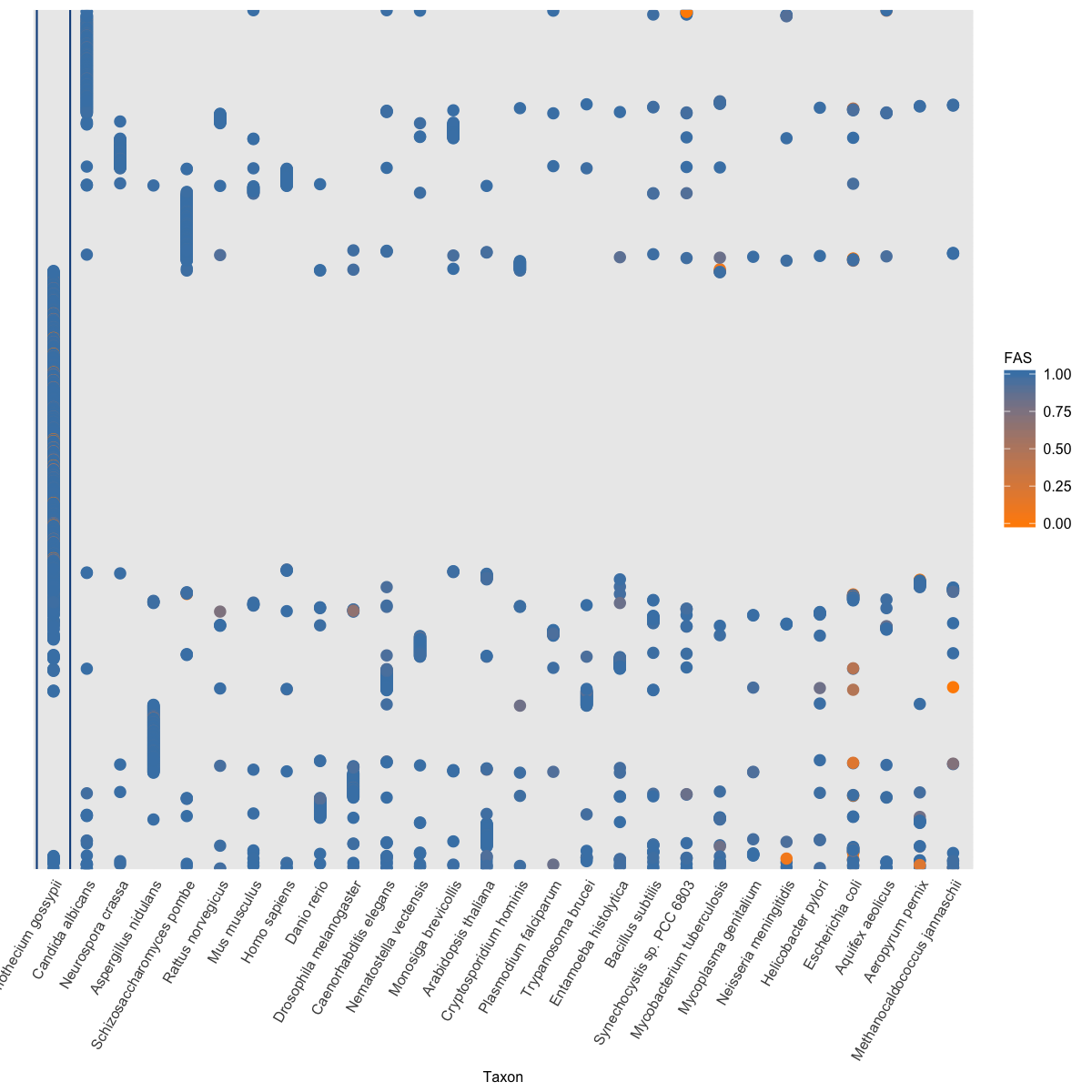


Figure A‑21: Phylogenetic profile of KO-annotated proteins

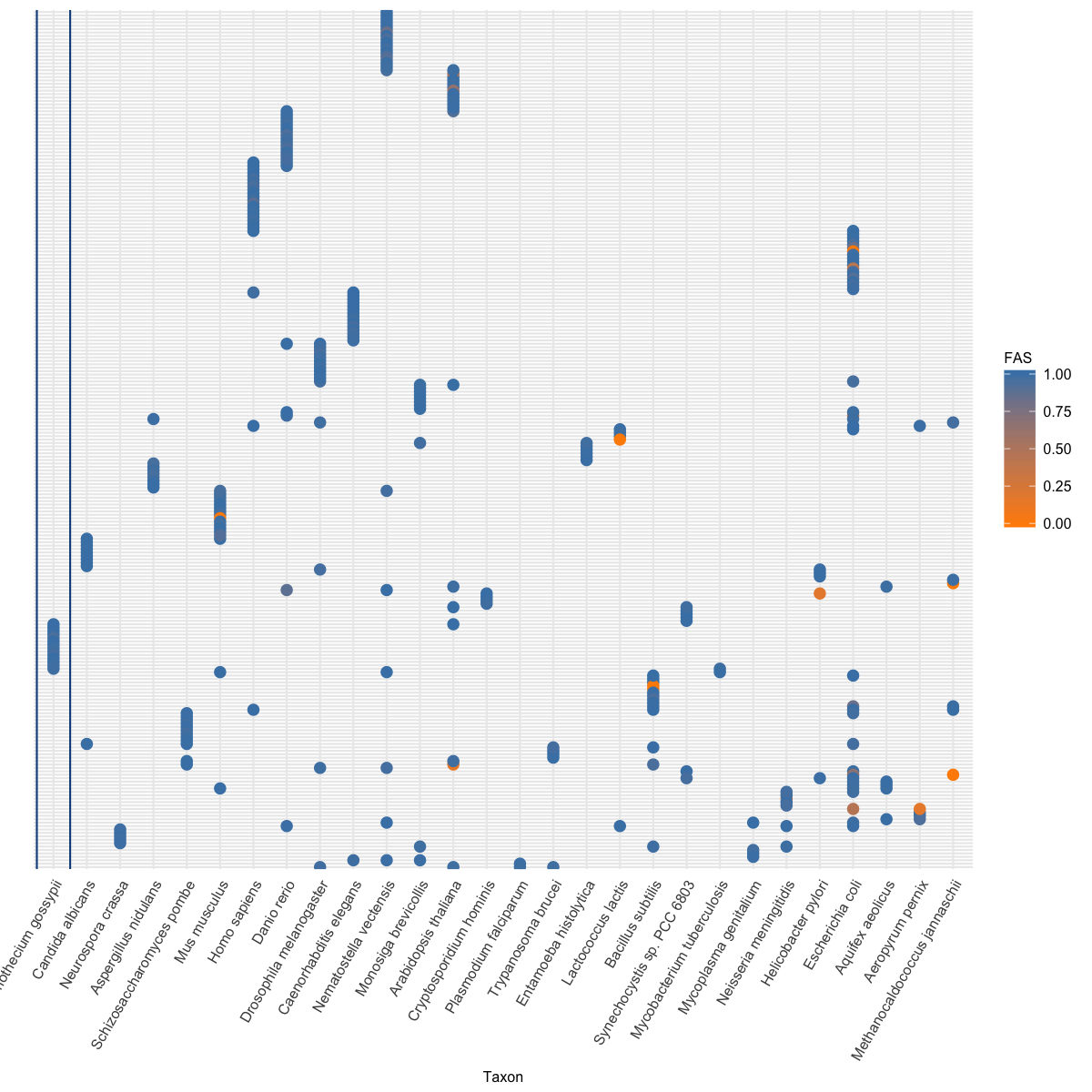


Figure A‑22: Phylogenetic profile of un-annotated proteins

Figure A‑21 and Figure A‑22 show that orthologs of un-annotated proteins are not broadly distributed like the one of annotated proteins. However, most of the proteins in both annotated and un-annotated set have only one ortholog (79% KO-annotated proteins, 80% un-annotated and 80% HamFAS-only proteins. See Figure A‑23). And more than 22% of un-annotated proteins have only orthologs in distantly related reference taxa (more detail in point **Error! Reference source not found.**).

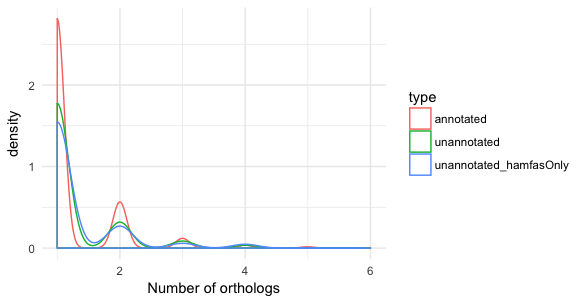


Figure A‑23: Distribution of number of orthologs for KO-annotated, un-annotated and HamFAS-only protein set

Do the annotations of HamFAS-only proteins come from distantly related species?

We checked for the origin of the annotations (i.e. the origin of reference orthologs) for all un-annotated proteins and compared with annotated set (Figure A‑24).

Figure A‑24: Origin of KO-annotations for annotated, un-annotated proteins and HamFAS-only proteins of un-annotated set

As expected, most annotations of annotated proteins come from their fungal orthologs (75%) while only few of them have obtained annotations from archaea or bacterial taxa (2,4%). In contrary, although large amount of annotations for un-annotated proteins originate from eukaryotes taxa (78%), there are still 22% (or 27% in case of HamFAS-only proteins) annotations are from distantly related taxa.

Analyzing the phylogenetic profile of proteins annotated by archaea and bacterial orthologs in Figure A‑25 and Figure A‑26, we can see that there is no difference between the HamFAS-only proteins and proteins that are annotated by both HamFAS and at least one other approach (HamFAS + BlastKOALA, HamFAS + KAAS or HamFAS + BlastKOALA + KAAS).

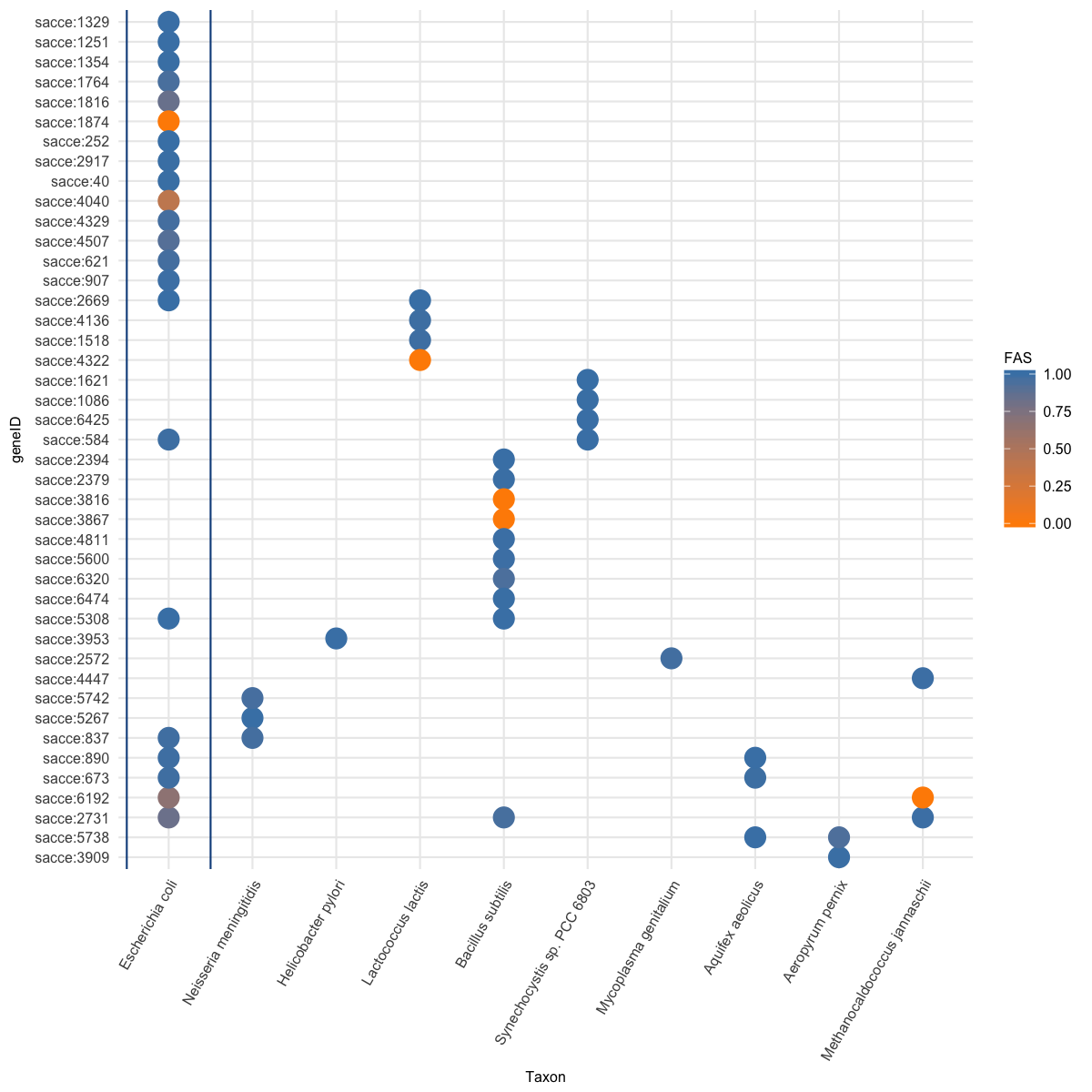


Figure A‑25: Phylogenetic profile of 44 HamFAS-only proteins that annotated based on archaea and bacterial orthologs.

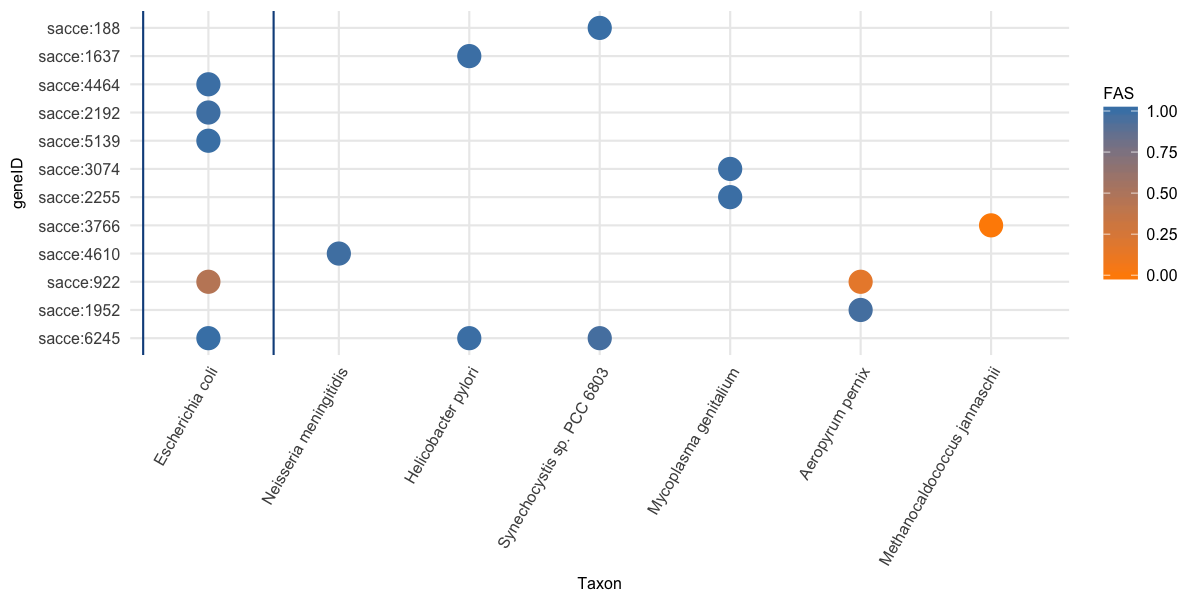


Figure A‑26: Phylogenetic profile of 12 un-annotated proteins that annotated by HamFAS and at least one other approach (BlastKOALA and/or KAAS), where their annotations originate from archaea or bacteria reference taxa.

How does the annotation result change by removing annotations from archaea and bacterial orthologs?

We filtered the annotations that originate from archaea and bacterial orthologs from both KO-annotated and un-annotated protein sets.

Table A‑5: Recall, precision and F1-score of filtered HamFAS in comparison to HamFAS, BlastKOALA and KAAS by applying on KO-annotated yeast proteins.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Approach** | **HamFAS after filtered** | **HamFAS** | **BlastKOALA** | **KAAS** |
| Recall | 0.9149 | 0.9152 | 0.905 | 0.931 |
| Precision | 0.9867 | 0.9854 | 0.979 | 0.984 |
| F1-score | 0.9496 | 0.9490 | 0.940 | 0.957 |

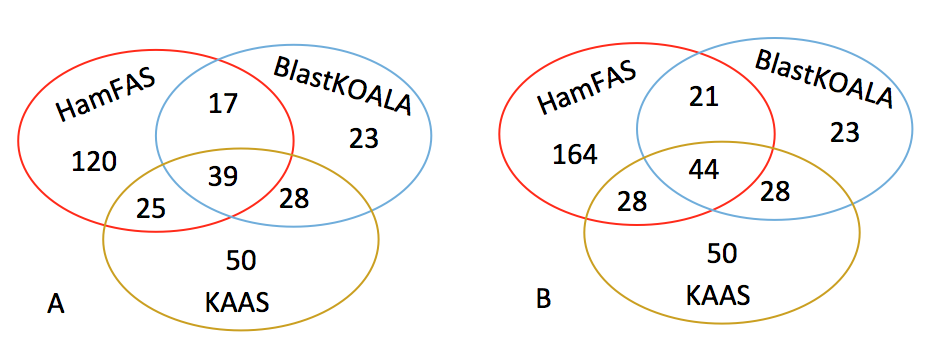
Table A‑5 shows a slightly increase in precision and F1-score of filtered HamFAS in comparison to original HamFAS due to a small number of annotations obtained from distantly related taxa.

Figure A‑27: Fraction of proteins annotated by BlastKOALA, KAAS and filtered HamFAS (A) or original HamFAS (B)

In Figure A‑27 we observe a decrease of the number of proteins annotated by HamFAS. However there are still a large amount of proteins that are annotated only by HamFAS (120 proteins) in comparison to BlastKOALA (23 proteins) and KAAS (50 proteins).

Are annotated proteins involved in PPI networks or KEGG pathways?

We analyzed the connectivity of annotated proteins and the obtained KOs by calculating the node degree of those proteins in yeast protein-protein-interaction (PPI) networks and the occurrence of the annotated KOs in KEGG pathways. PPI data are retrieved from Yeast Interactome Project (http://interactome.dfci.harvard.edu/S\_cerevisiae/) and STRING database (https://string-db.org).

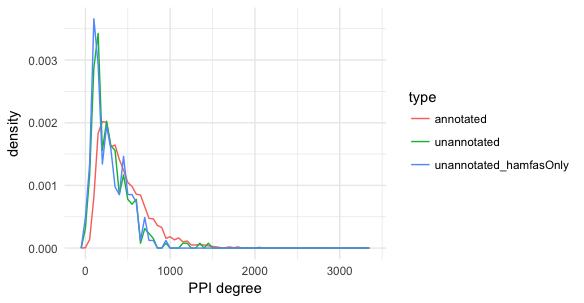


Figure A‑28: The PPI degree distribution of 3 protein sets

Figure A‑28 shows the distribution of PPI degree of KO-annotated, un-annotated and HamFAS-only proteins inside un-annotated set. KO-annotated proteins have in general more interacting partners (mean PPI degree 444) than un-annotated and HamFAS-only proteins (mean PPI degree 294 and 275 respectively). However, 99% of the proteins of un-annotated set have the PPI degree more than 10, while only 2 proteins don't have any interacting partner.

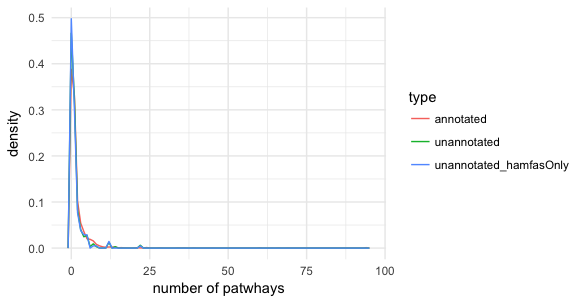


Figure A‑29: Distribution of the number of pathways in which annotated KOs are involved

For the annotated KOs, we calculate the number of pathways in which those KOs are involved. All 3 data sets show the same trend in Figure A‑29, that not less then 50% the KOs belong to at least one KEGG pathway (KO-annotated set 61%, un-annotated set 53% and HamFAS-only protein set 50%).

Are new annotations from HamFAS meaningless?

About 50% of KOs annotated only by HamFAS belongs to KEGG's pathways. Figure A‑30 shows the distribution of those KOs in different pathway categories.

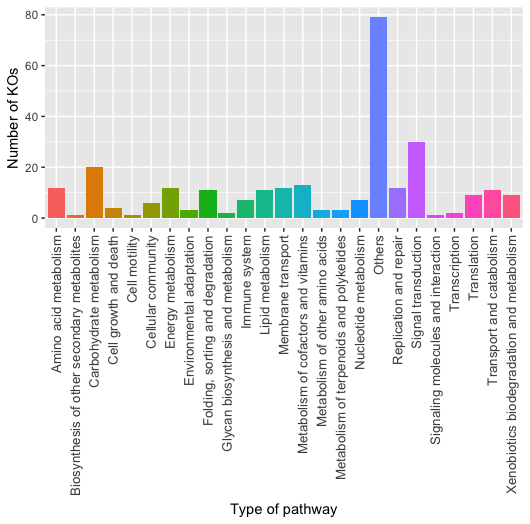


Figure A‑30: The numbers of HamFAS-only KOs distributed into different pathway categories

29 yeast pathways are further complemented by new KOs from HamFAS. (See appendix)

Why did BlastKOALA and KAAS fail to annotate HamFAS-only protein?

One reason could be, that the orthology prediction approaches used by KAAS and BlastKOALA are not as sensitive as HaMStR. The second reason is due the their "secret" filter criteria to select candidate for doing annotation transfer.

We have observed, that there are in total 86 predicted KOs that are common between annotated and unannotated data sets. In which 44 cases are also supported by InParanoid. Examples:

**K00077:** sce:4314, B.subtilis:BSU15110, B.subtilis:BSU14440 (KEGG's representative sequences)

HamFAS: sce:6474 (unannotated protein) is orthologous with B.subtilis:BSU14440

Inparanoid: 2 separate OGs: (sce:4314, B.subtilis:BSU15110) and (sce:6474, B.subtilis:BSU14440)

**K00799:** ath:AT1G02930,..., ath:AT2G30870,..., sce:5364, sce:1884

HamFAS: sce:2310 - ath:AT1G02930,...

(sce:1884 has no ortholog with *A.thaliana* according to InParanoid)

**K00877:** S.pombe:4570, S.pombe:875, S.pombe:1336, sce:1877, sce:997

HamFAS: sce:487 - S.pombe:1336

Those proteins have been probably either not predicted as orthologs or discarded after filtering through KEGG annotation pipeline.

Conclusion

The ability of identifying distantly related orthologs of HaMStR leads to the result that more proteins have been annotated by HamFAS than BlastKOALA or KAAS. There is no strong evidence to distinct the difference between HamFAS-only proteins and proteins that are annotated by both HamFAS and other approaches. We can increase the stringency of HamFAS by allowing the annotations from only close related species. However, if doing so we will lose the benefit of HaMStR and therefore we have no reason to use HaMStR instead of other more stringent orthology search approaches like OMA or InParanoid. The principle factor that affects the annotation result is the accuracy of orthology assignment method. This HamFAS approach could be supported more by the analysis of QfO from Holger.

Metabolic pathway analysis of microsporidian LCA proteins

Introduction

Metabolic analysis of microsporidia is still a challenge due to their obligate intracellular growth and short lifetime of its purified spores (Keeling and Fast, 2002). Here we compare the metabolic pathways of the microsporidian LCA with the contemporary species to verify the current hypotheses about microsporidia metabolism and investigate the differences between the metabolism of the microsporidian LCA and the extant species.

Methods

We used HamFAS approach to do KO annotation for 1605 microsporidian LCA proteins. HaMStR was used for ortholog search between microsporidian LCA (seed) and KEGG reference species (reference). Since one microsporidian LCA protein is represented by an orthologous group of several microsporidia proteins, we assigned the representative FAS score for each reference protein as the max score that protein can archive when compare with all microsporidia proteins in the corresponding orthologous group. This representative max FAS score will be then compared with the TFAS\_KO in order to decide if the annotated KO of the reference protein can be transferred to the microsporidian LCA protein.

Beside the complementary FAS scores to the orthology assignment, we also calculated the patristic distance between the reference protein and microsporidia protein to use it as a confident value for the annotation transfer. The distance between a reference protein to the microsporidian LCA protein is the minimum distance between that reference protein to all microsporidia proteins in the corresponding orthologous group. The distances in one orthologous group are normalized to a range of [0,1] by the formula (currentDist - minDist)/(maxDist - minDist).

The KO-annotated microsporidian LCA proteins were then mapped to KEGG pathways. The result was compared with E.cuniculi, E.hellem, E.intestinalis, N.ceranae, 4/11 extant microsporidia species under this study that are available in KEGG database, and with S.cerevisiae, as an example for free-living organism. The annotations for E.cuniculi, E.hellem, E.intestinalis, N.ceranae and S.cerevisiae were obtained directly from KEGG.

We also compare the connectivity network between microsporidian LCA and those contemporary species. For each reference KEGG pathway, the connectivity network nodes are enzymes (represented by their KO identifiers) in the pathway and edges are links between those nodes. KO-annotated proteins of each taxon will be then mapped to those reference networks for a connectivity analysis. This connectivity network analysis is implemented into a tool named KEGGcxn.

Results

Using HamFAS approach we have annotated 1048 out of 1605 microsporidian LCA proteins with 1344 different KO identifiers.

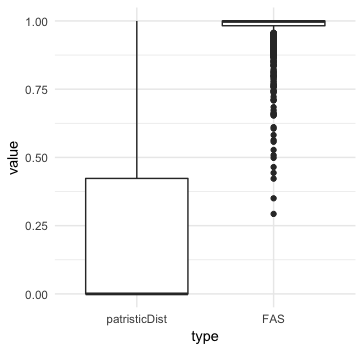


Figure A‑31: Distribution of FAS scores and patristic distances of KO-annotated microsporidian LCA proteins. Blue line represents the conditional mean of FAS score given a patristic distance value.

Figure A‑31 shows the distribution of FAS scores and patristic distances of all transferred KOs. Most of the annotated KOs have vey high FAS scores (mean FAS score is 0.97). Mean and median of scaled patristic distances are 0.22 and 0.00 respectively. The high mean FAS score and low scaled patristic distances shows that most seed and reference proteins are highly similar in term of domain architectures and a large fraction of the annotations come from the closely related species (correct for patristic distance? or it just tells us about the sequence similarity?).



Figure A‑32: Pathway enrichment of microsporidian LCA. Colors denote different pathway categories: green for cellular processes, orange for environmental information processing, purple for genetic information processing and pink for metabolism.

Pathway enrichment result of microsporidian LCA is shown in Figure A‑32. While Figure A‑33 shows the fractions of proteins take part in different pathway categories of microsporidian LCA in comparison to other extant species. Microsporidian LCA has more proteins in metabolism than the 4 extant microsporidia species (30% in comparison to 25%, respectively) but still less than the free-living S.cerevisiae (38%).



Figure A‑33: Fractions of proteins distributed in different pathway categories. (NOT NECESSARY)

A more detail of the mapped pathways and number of proteins for each pathway is shown in Figure A‑34. In general, microsporidian LCA has more proteins mapped into pathways in comparison to extent microsporidia species, especially in Cell growth and death, Signal transduction, Folding, sorting and degradation, Carbohydrate and Lipid metabolism. However, it is still very less when compare to S.cerevisiae, a representative of free-living organisms. One possible reason could be, that the number of yeast proteins in this analysis is much higher than the one from microsporidia (3534 yeast proteins versus 950 protein in average for each microsporidia species) <= put into the discussion !!



Figure A‑34: Number of proteins of each taxon (green for microsporidian LCA, orange for E.cuniculi, purple for E.hellem, pink for E.intestinalis, green for N.ceranae and yellow for S.cerevisiae) participates in different KEGG pathways.

We compare the connectivity of annotated proteins between microsporidian LCA and the contemporary species under this study for the core metabolic pathways used in (Nerima *et al.*, 2010) including glycolysis, gluconeogenesis, the Krebs cycle, pentose phosphate pathway, purine and pyrimidine metabolism, and amino acid metabolism.



Figure A‑35: Number of nodes (left) and edges (right) of core pathways for microsporidian LCA, E.cuniculi, E.hellem, E.intestinalis, N.ceranae and S.cerevisiae.

Figure A‑35 shows the comparison between number of nodes and edges in six core metabolic networks for microsporidian LCA and other 5 extant species. The average node degree, average path length and network diameter (the longest shortest paths) can be seen in Figure A‑36. In general, almost all network properties of parasite species are smaller than the free-living species S.cerevisiae, except the path length of Pentose phosphate pathway.



Figure A‑36: Density of average node degree, average path length and diameter (maximal path length) of microsporidian LCA, E.cuniculi, E.hellem, E.intestinali, N.ceranae and S.cerevisiae in 6 core pathways (Glycolysis/Gluconeogenesis, TCA cycle, Pentose phosphate pathway, purine metabolism, pyrimidine metabolism and amino acid metabolism).

Details of network properties for core pathways are shown in Table A‑6.

According to (Fast and Keeling, 2001; Agnew *et al.*, 2003; Keeling and Fast, 2002), microsporidia lacks of mitochondria. But with the presence of genes coding for heat-shock protein 70 (hsp70) in some extant microsporidia species, they suggested that microsporidia ancestor has mitochondria. Those studies also hypothesized that microsporidia will replace pyruvate dehydrogenase complex (PDH) by pyruvate ferredoxin oxidoreductase (PFOR) in order to convert pyruvate into acetyl-CoA and produce NADH. For the common ancestor, we could not find any KOs of 4 PFOR subunits (α, β, γ, δ) in microsporidian LCA, however 3 out of 4 components of PDH were found instead, i.e. pdhA (K00161, OG\_2283, EC=1.2.4.1) and pdhB (K00162, OG\_2084, EC=1.2.4.1) of E1 component, and E3 (DLD) component (K00382, OG\_3281, EC=1.8.1.4) (E2 DLAT, K00627, EC=2.3.1.12 not found). Note that E1 is also be found in N.locustae (Fast and Keeling, 2001) and Encephalitozoon (Katinka *et al.*, 2001). Figure A‑37 shows the mapped microsporidian LCA proteins into the reaction converting pyruvate into acetyl-CoA.

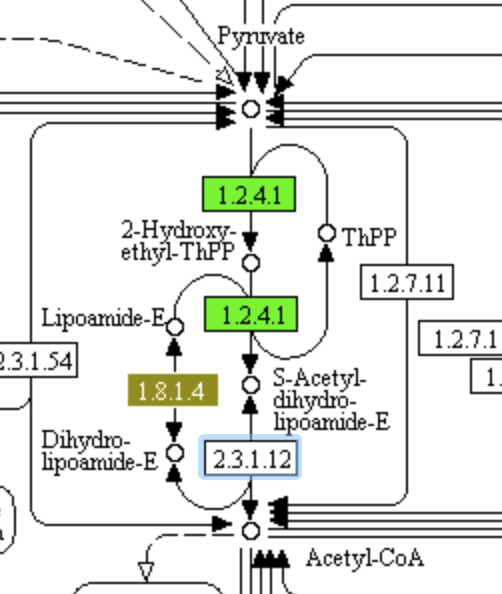


Figure A‑37: (NEED TO BE REDRAWN) The reaction Pyruvate -> Acetyl-CoA with help of pyruvate dehydrogenase complex (PDC). Proteins present in microsporidian LCA are highlighted.

The role of E1 component and the reason for the presence of hsp70 genes in extant microsporidia is still unclear (Together these genes (alpha and beta from E1) provide the first evidence for mitochondrion-derived metabolic activity in microsporidia, but it is still unclear whether they are involved in core energy metabolism in a mitochondrion as in other eukaryotes, or if they have been conscripted into some other pathway during the un- usual course of evolution in microsporidia.) (Fast and Keeling, 2001). But the presence of two subunits of E1 and the component E3 together with the hsp70 proteins (OG\_1157 and OG\_1803 with KO K03283) in the LCA emphasizes the origin hypothesis of mitochondrion in the microsporidia ancestor.

**LOST PATHWAYS**

(7) Yet in microsporidia, endoparasitic fungi living at the limits of cellular streamlining, oxidative phosphorylation has been lost: energy is obtained directly from the host or, during the dispersive spore stage, via glycolysis. It was therefore surprising when the first sequenced genome from the Enterocytozoonidae – a major family of human and animal-infecting microsporidians – appeared to have lost genes for glycolysis. (Wiredu Boakye *et al.*, 2017) See fig 1 and 2 as an example how to represent the presence/absence genes in pathways

Despite the hint about the presence of mitochondrion, it still supposes that microsporidian LCA also lack of TCA cycle, electron transport chain and oxidative phosphorylation pathway like the extant species and other amitochonriate species (Keeling, 2009; Keeling and Fast, 2002; Wiredu Boakye *et al.*, 2017). They retain only 10/13 subunits of the vacuolar H+ ATPase in the oxidative phosphorylation. Due to the lack of the main ATP supplier from the mitochondrion, the synthesis of ATPs therefore depends on other pathways like glycolysis or through ATP transport system.

Microsporidia mostly uptake ATP from host species using their ATP-binding cassette (ABC) transporters (Méténier and Vivarès, 2001; Keeling, 2009; Heinz *et al.*, 2012). Besides, (Heinz *et al.*, 2012) also found putative major facilitator superfamily (MFS) transporters in the microsporidia T.hominis. We searched for the transport proteins in the microsporidian LCA and found two MFS transporter and 6 ATP-binding cassette (ABC) transporters (see Table A‑6).

Table A‑6: Microsporidian LCA MFS and ABC transporters.

|  |  |  |
| --- | --- | --- |
| LCA protein | KO identifier | Description |
| OG\_3349 | K08139 | MFS transporter, SP family, sugar:H+ symporter |
| OG\_1075 | K08146 | MFS transporter, SP family, solute carrier family 2 (facilitated glucose transporter), member 9 |
| OG\_1019 | K06174 | ATP-binding cassette, sub-family E, member 1 |
| OG\_1050 | K06185 | ATP-binding cassette, subfamily F, member 2 |
| OG\_1034 | K06158 | ATP-binding cassette, subfamily F, member 3 |
| OG\_1082 | K05681 | ATP-binding cassette, subfamily G (WHITE), member 2 |
| OG\_1098 | K05662 | ATP-binding cassette, subfamily B (MDR/TAP), member 7 |

Beside up taking ATP using ATP transporters, microsporidia is well-known that they use glycolysis to produce ATP (Keeling and Corradi, 2011; Keeling and Fast, 2002; Heinz *et al.*, 2012).

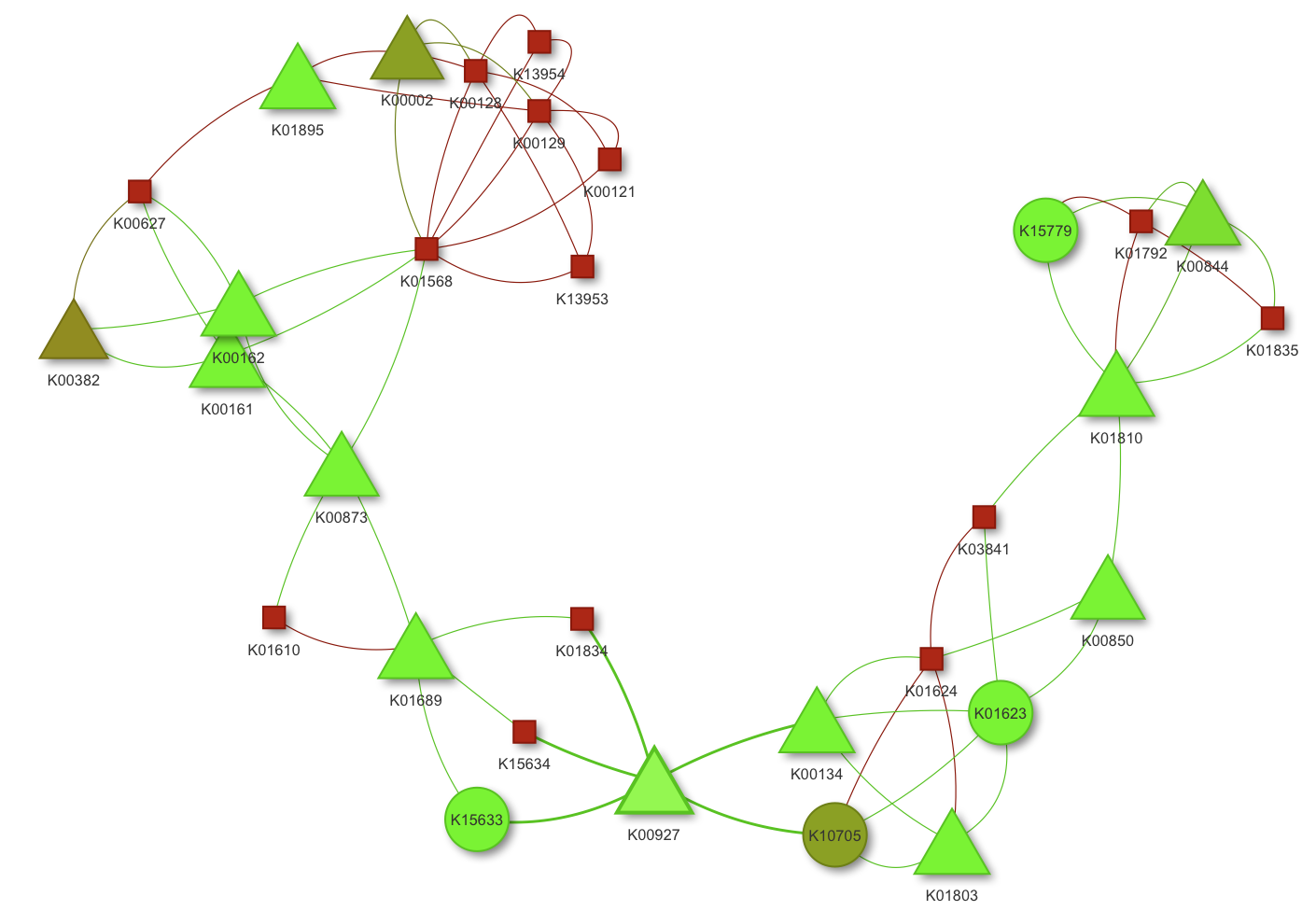


Figure A‑38: Connectivity network of annotated proteins in glycolysis/gluconeogenesis pathway. Circles are microsporidian LCA only proteins. Squares are S.cerevisiae only proteins. Triangles are mapped proteins from both microsporidian LCA and S.cerevisiae. Color denotes the patristic distance between the microsporidia seed protein and the reference ortholog, where the transferred KO annotated comes from (this color code applied only for microsporidian LCA proteins) (ADD COLOR SCALE)

The connectivity network in Figure A‑38 shows the mapped microsporidian LCA and S.cerevisiae proteins into KEGG glycolysis/gluconeogenesis pathway (unmapped nodes are removed). Except K10705 (dark green circle), all other proteins are also present in contemporary microsporidia species. Although the number of nodes of microsporidia is less than S.cerevisiae, they are highly connected and the diameter of microsporidia network is almost the same as the one of S.cerevisiae (10 vs 11). It indicates that microsporidian LCA has almost all keys enzymes for this pathway and can perform the same reactions yeast, except some reactions in one end of the network. See Figure A‑39 for the reference KEGG glycolysis/gluconeogenesis pathway mapped with microsporidian LCA proteins.

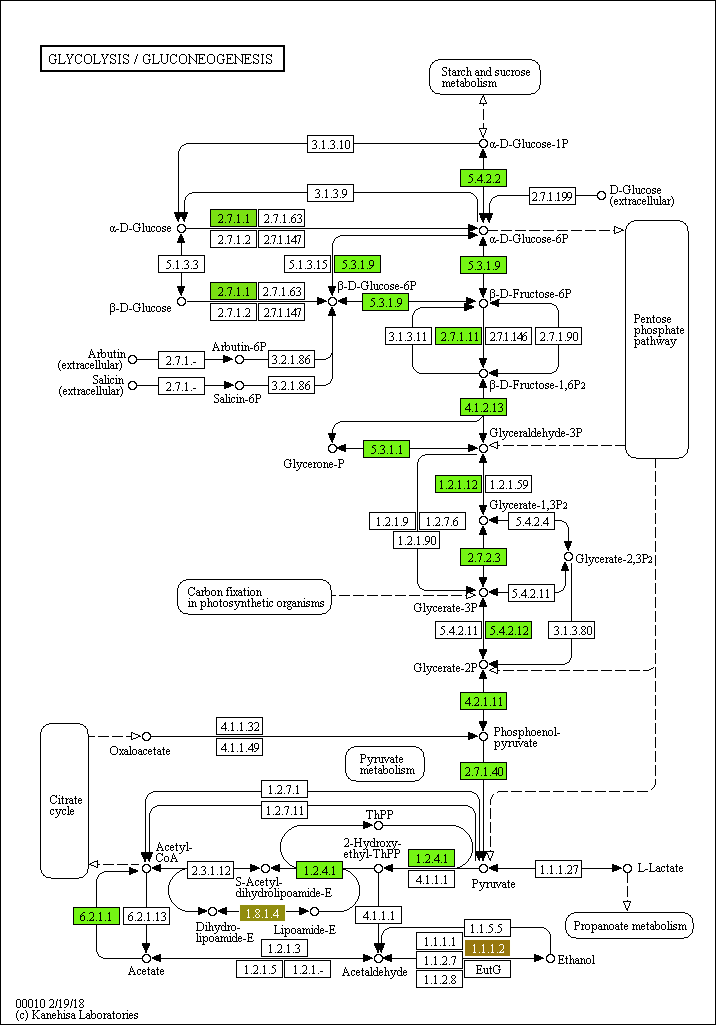


Figure A‑39: Glycolysis/Gluconeogenesis pathway. Annotated microsporidian LCA proteins are highlighted in green. Blank box are unmapped proteins in reference KEGG pathway. (REDRAWN WITH EXTANT MICROS PROTEINS AND MAYBE ALSO YEAST)

According to (Keeling and Fast, 2002; Keeling and Corradi, 2011; Heinz *et al.*, 2012), another core carbon metabolism has been confirmed to be present in microsporidia is the pentose phosphate pathway. This is also true with the LCA, since the pathway mapped with its proteins is the same as the extant microsporidia pathway (Figure A‑40).

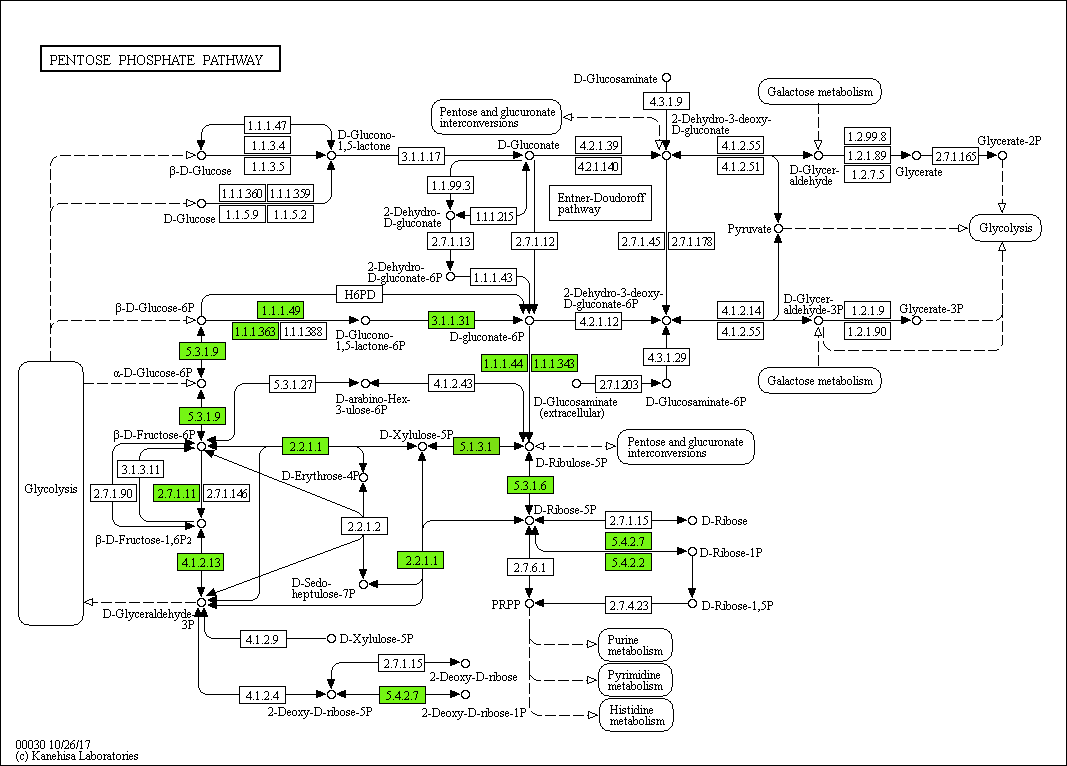


Figure A‑40: Pentose phosphate pathway

The primary carbohydrate storage trehalose is thought to be very essential for the survival and germination of microsporidian spore (Dolgikh *et al.*, 1997; Vandermeer and Gochnauer, 1971; Agnew *et al.*, 2003; Heinz *et al.*, 2012). Enzymes for trehalose synthesis and degradation in extant microsporidia (Keeling and Corradi, 2011; Vandermeer and Gochnauer, 1971; Heinz *et al.*, 2012; Méténier and Vivarès, 2001) have also been found in the LCA including the trehalose 6-phosphate synthase (EC 2.4.1.15 and 2.4.1.347) and alpha-trehalase (EC 3.2.1.28). Figure... demonstrates the starch and sucrose metabolism of microsporidia.

~~The same as contemporary microsporidia species, the LCA also has trehalose 6-phosphate synthase (K00697, EC 2.4.1.15 and 2.4.1.347) and alpha-trehalase (K01194, EC 3.2.1.28), which are involved in the synthesis and degradation of trehalose.~~ (ADD A TABLE WITH ALL ANNOTATED PROTEINS IN THIS ANALYSIS IN THE SUPPLERMENTARY)

DRAW STARCH AND SUCROSE METABOLISM OF MICROSPORIDIA

Having the obligate parasitic life-style, microsporidia tents to uptake nucleotide from the host than produce by themself (Heinz *et al.*, 2012; Dean *et al.*, 2016). Microsporidian LCA lack ribose-phosphate pyrophosphokinase (K00938, EC 2.7.6.1), which converts ribose 5-phosphate into phosphoribosyl pyrophosphate (PRPP) for the de-novo purine and pyrimidine synthesis. IMP cyclohydrolase (K11176, EC 3.5.4.10) making inosine monophosphate IMP and UMP synthetase (K13421, EC 2.4.2.10 & 4.1.1.23) making UMP from PRPP are also not found. The missing of those enzymes indicates that microsporidian LCA is unable to de-novo synthesize both purines and pyrimidines.

See Heinz 2014 for purine and pyrimidine pathway (Fig 1)

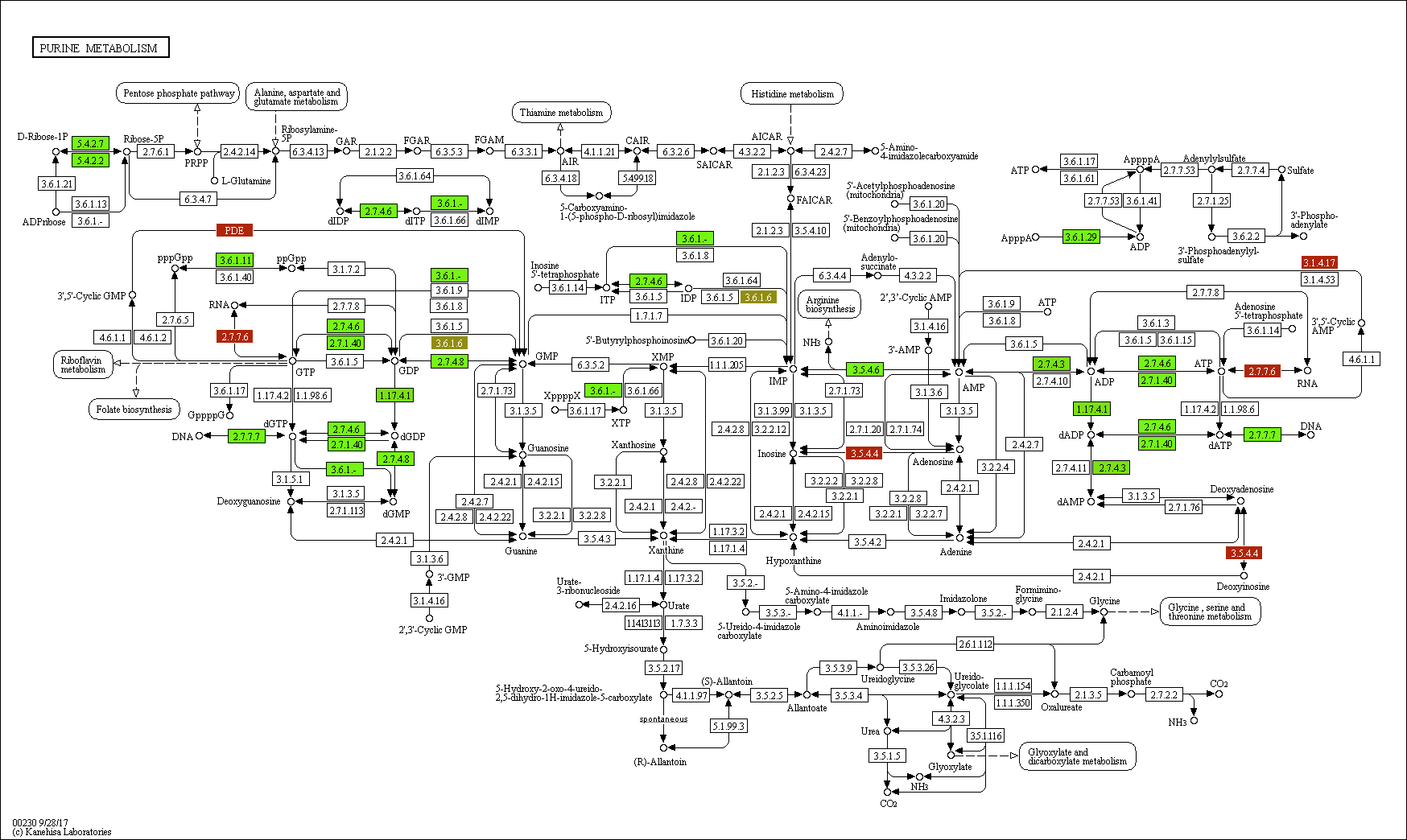


Figure A‑41: Purine

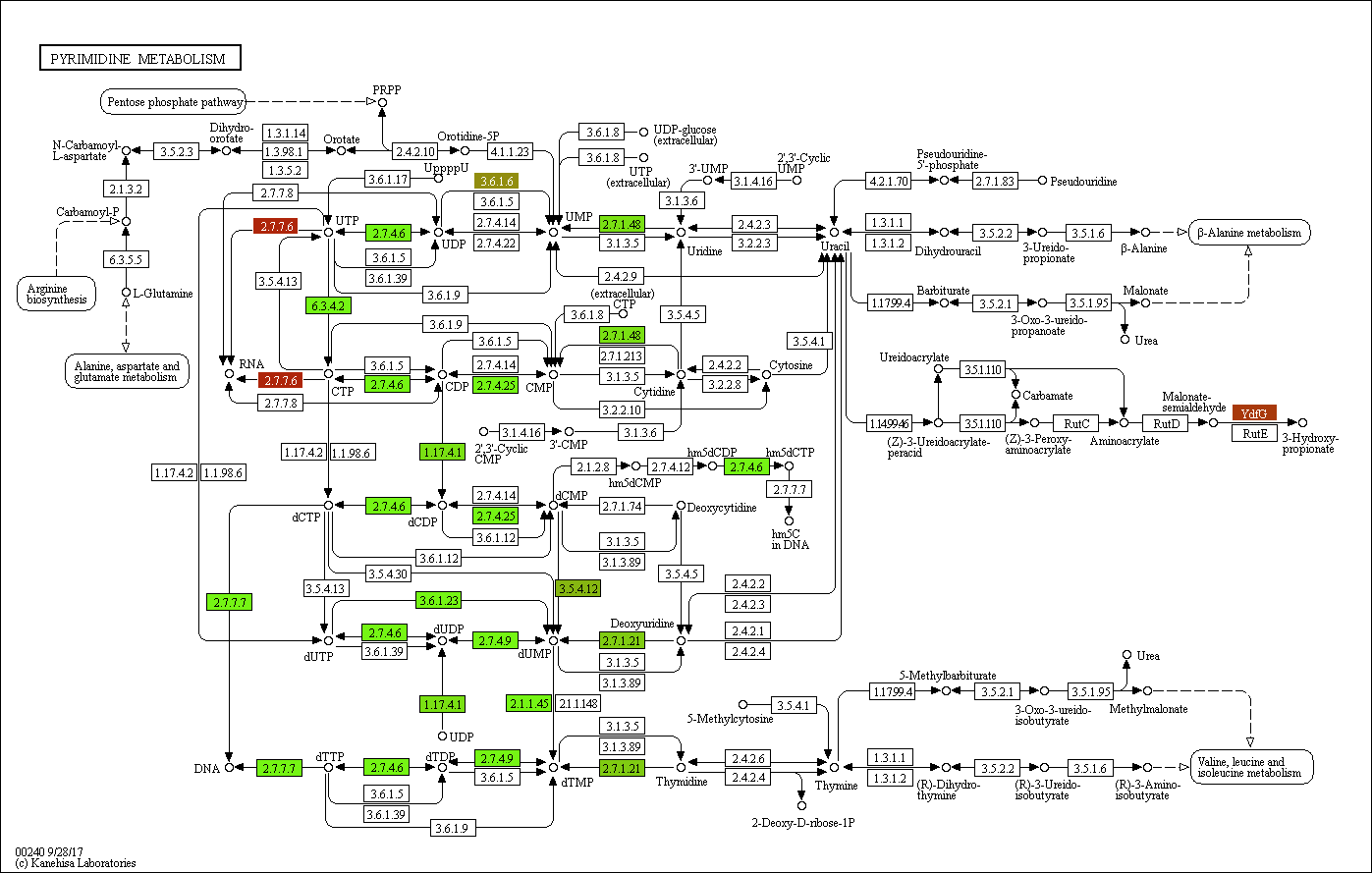


Figure A‑42: Pyrimidine

By that reason, microsporidia need to import nucleotides from the hosts using nucleotide transport (NTT) proteins. KO identifier K03301 of four NTT (NTT1, NTT2, NTT3, NTT4) proteins (Heinz *et al.*, 2014; Dean *et al.*, 2016) have been found in three different microsporidian LCA proteins. Based on studies of (Dean *et al.*, 2016; Tsaousis *et al.*, 2008; Heinz *et al.*, 2014), those NTT proteins are the result of horizontal transfer event from bacteria.



Figure A‑43: Phylogenetic profile of 3 microsporidian LCA NTT proteins

Figure A‑43 shows the phylogenetic profile of 3 microsporidian LCA NTT proteins. All three proteins have bacterial orthologs in Chlamydiae phylum. The domain annotation of a microsporidia protein in comparison with its bacterial ortholog is shown in Figure A‑44. They both contain 11-12 transmembrane domains, as commonly observed in bacterial NTT proteins (Tsaousis *et al.*, 2008; Winkler and Neuhaus, 1999).

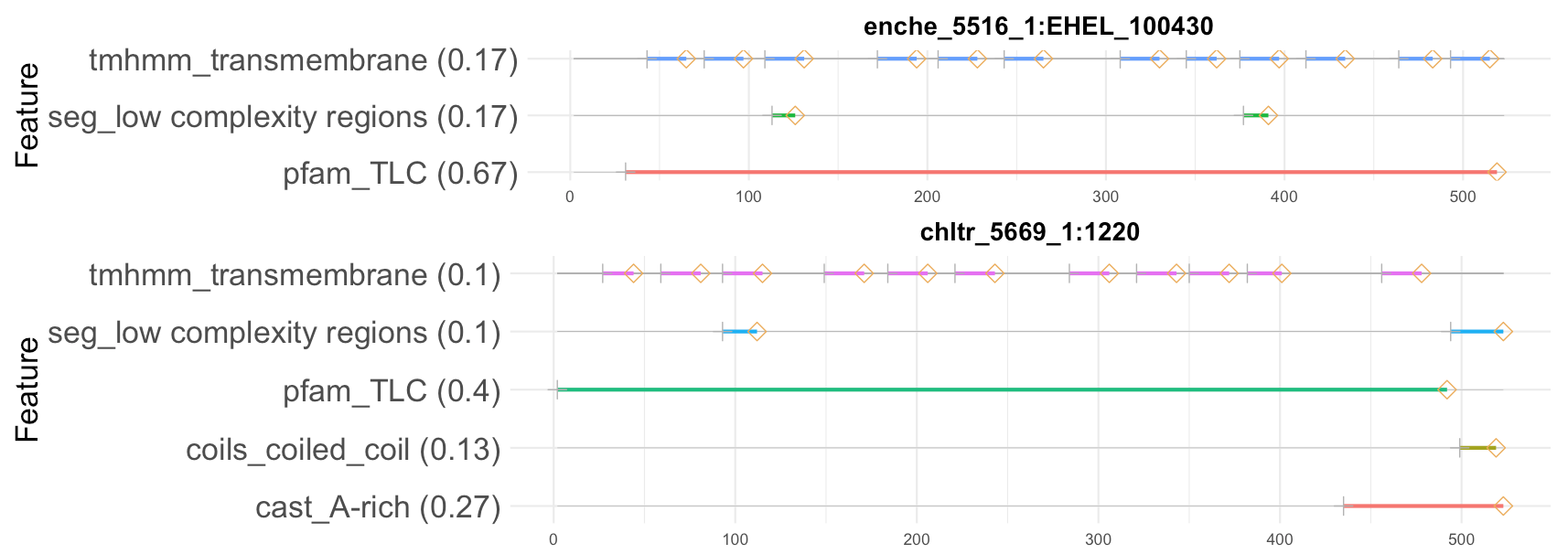


Figure A‑44: Domain architecture of E.hellem protein (enche\_5516\_1:EHEL\_100430) and its ortholog (chltr\_5669\_1:1220) of the bacteria Chlamydia trachomatis. (REMOVE WEIGHT)

4 NTT proteins (

NTT1 - Q8SRA2 - ECU08\_1300 - K03301 <=> OG\_1062,

NTT2 - Q8SUF9 - ECU10\_0540 - K03301 <=> OG\_1062,

NTT3 - Q8SUG0 - ECU10\_0520 - K03301 <=> OG\_3238,

NTT4 - Q8SUG7 - ECU10\_0420 - K03301 <=> OG\_3237 Heinz, Hacker & others 2014)

Check the origin of NTT (also in other species) => see fig 2C (Nakjang 2013)

(9) Genes encoding a fatty acid synthase complex are lacking, which supports the uptake of host-derived fatty acids (El Alaoui *et al.*, 2001; Katinka *et al.*, 2001).

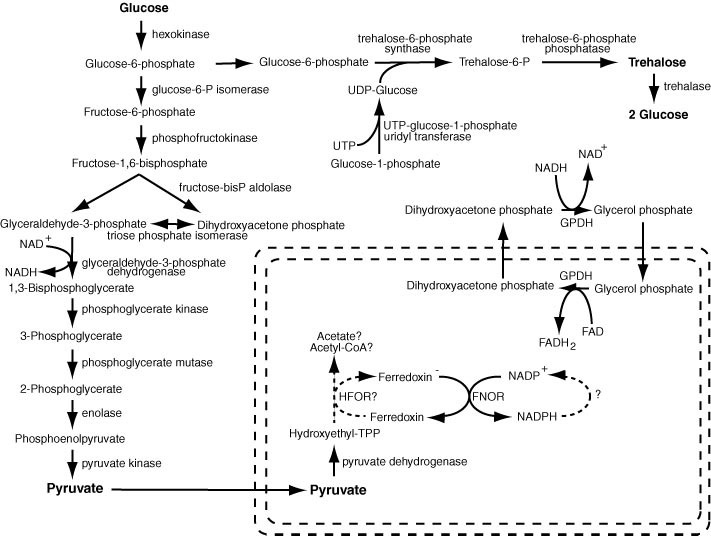


Figure A‑45: carbon pathway (Keeling and Fast, 2002)

Discussion

Conclusion

Our analysis of microsporidian LCA metabolic pathways acquired the consistent results with the finding other studies. Microsporidian LCA, as well as the contemporary species, obligatorily depends on the host species for their survival. The presence of transport proteins supplements the lack of some main pathways for producing energy and other important compounds. Trehalose seems to be the main carbohydrate storage for microsporidia since the enzymes for de-novo trehalose synthesis and degradation are also be found in the LCA. However, the reason for the existence of mitochondria in the LCA is still unclear, since the pathways that take place in mitochondria are missing. This analysis demonstrates a novel approach for in-silico studying the metabolic network of microsporidia or any other species.

can replace extant microsporidia by contemporary microsporidia

scheme (used for figure, e.g. scheme of possible energy metabolism in microsporidia)

interpretation

investigation

largely poorly understood

indeed

In essence

illustrating

microsporidian inventions

opisthokont ancestor

# Appendix

Tables

Table A‑1: Eleven extant microsporidia species

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| No. | Name | Source | Strain | Number of proteins |
| 1 | *Encephalitozoon hellem* | JGI | ATCC 50504 | 1827 |
| 2 | *Encephalitozoon intestinalis* | Broad Inst | ATCC 50506 | 1657 |
| 3 | *Encephalitozoon cuniculi* | Broad Inst | GB-M1 | 1896 |
| 4 | *Nosema ceranae* | Broad Inst | BRL01 | 2057 |
| 5 | *Enterocytozoon bieneusi* | JGI | H348 | 3312 |
| 6 | *Vittaforma corneae* | Broad Inst | ATCC 50505 | 2243 |
| 7 | *Anncaliia algerae* | Broad Inst | PRA339 | 3576 |
| 8 | *Antonospora locustae* | JGI | HM-2013 | 2191 |
| 9 | *Edhazardia aedis* | Broad Inst | USNM 41457 | 4208 |
| 10 | *Vavraia culicis* subsp. floridensis | Broad Inst |  | 2775 |
| 11 | *Nematocida parisii* | Broad Inst | ERTm1 | 2659 |

Table A‑2: 24 taxa used for extent the initial homologous groups

|  |  |  |  |
| --- | --- | --- | --- |
| No. | Name | No. | Name |
| 1 | *S.pombe* | 14 | *M.brevicollis* |
| 2 | *C.albicans* | 15 | *N.vectensis* |
| 3 | *S.cerevisiae* | 16 | *A.queenslandica* |
| 4 | *N.crassa* | 17 | *C.owczarzaki* |
| 5 | *A.nidulans* | 18 | *T.brucei* |
| 6 | *L.bicolor* | 19 | *N.gruberi* |
| 7 | *P.graminis* | 20 | *A.thaliana* |
| 8 | *M.circinelloides* | 21 | *C.reinharditii* |
| 9 | *R.oryzae* | 22 | *P.sojae* |
| 10 | *P.blakesleeanus* | 23 | *C.hominis* |
| 11 | *B.dendrobatidis* | 24 | *P.falciparum* |
| 12 | *S.punctatus* |  |  |
| 13 | *R.allomyces* |  |  |

Table A‑3: Mean length of orthologous and orphan proteins in 11 microsporidia. We used Wilcoxon-Mann-Whitney U-Test to compare the length of those two protein groups. P-value is less then 0.05 meaning that the length of orthologous proteins are significant different to the one of orphan proteins.

|  |  |  |  |
| --- | --- | --- | --- |
| Taxon | Mean length of orthologous proteins | Mean length of orphans | P\_value |
| E.hellem | 358,507 | 305,250 | 0,1966 |
| E.intestinallis | 358,931 | 174,630 | 9,11E-07 |
| E.cuniculi | 368,688 | 187,100 | 1,14E-10 |
| N.ceranae | 339,184 | 279,514 | 2,32E-09 |
| E.bieneusi | 274,151 | 182,634 | p < 2,2E-16 |
| V.corneae | 330,872 | 283,743 | 5,05E-08 |
| A.algerae | 284,651 | 223,355 | p < 2,2E-16 |
| A.locustae | 295,033 | 157,594 | p < 2,2E-16 |
| E.aedis | 380,879 | 319,525 | p < 2,2E-16 |
| V.culicis | 370,504 | 294,433 | p < 2,2E-16 |
| N.parisii | 421,400 | 302,794 | p < 2,2E-16 |

Table A‑4: (AGAIN!!) List of species we used for the distribution analysis of microsporidian LCA proteins.

|  |  |  |  |
| --- | --- | --- | --- |
| No. | Full name | supertaxa | group |
| 1 | Ashbya gossypii | Saccharomycotina | fungi |
| 2 | Candida albicans | Saccharomycotina | fungi |
| 3 | Candida dubliniensis CD36 | Saccharomycotina | fungi |
| 4 | Candida glabrata | Saccharomycotina | fungi |
| 5 | Candida parapsilosis | Saccharomycotina | fungi |
| 6 | Candida tropicalis | Saccharomycotina | fungi |
| 7 | Clavispora lusitaniae | Saccharomycotina | fungi |
| 8 | Debaryomyces hansenii CBS767 | Saccharomycotina | fungi |
| 9 | Kluyveromyces lactis | Saccharomycotina | fungi |
| 10 | Kluyveromyces thermotolerans | Saccharomycotina | fungi |
| 11 | Kluyveromyces waltii | Saccharomycotina | fungi |
| 12 | Lodderomyces elongisporus NRRL YB-4239 | Saccharomycotina | fungi |
| 13 | Pichia guilliermondii | Saccharomycotina | fungi |
| 14 | Pichia pastoris GS115 | Saccharomycotina | fungi |
| 15 | Pichia stipitis CBS 6054 | Saccharomycotina | fungi |
| 16 | Saccharomyces bayanus 623-6C | Saccharomycotina | fungi |
| 17 | Saccharomyces castelli | Saccharomycotina | fungi |
| 18 | Saccharomyces cerevisiae | Saccharomycotina | fungi |
| 19 | Saccharomyces kluyveri | Saccharomycotina | fungi |
| 20 | Saccharomyces kudriavzevii | Saccharomycotina | fungi |
| 21 | Saccharomyces mikatae | Saccharomycotina | fungi |
| 22 | Saccharomyces paradoxus | Saccharomycotina | fungi |
| 23 | Vanderwaltozyma polyspora | Saccharomycotina | fungi |
| 24 | Yarrowia lipolytica | Saccharomycotina | fungi |
| 25 | Zygosaccharomyces rouxii | Saccharomycotina | fungi |
| 26 | Acidomyces richmondensis | Pezizomycotina | fungi |
| 27 | Aulographum hederae | Pezizomycotina | fungi |
| 28 | Baudoinia compniacensis uamh 10762 | Pezizomycotina | fungi |
| 29 | Botryosphaeria dothidea | Pezizomycotina | fungi |
| 30 | Cenococcum geophilum 1.58 | Pezizomycotina | fungi |
| 31 | Cladonia grayi | Pezizomycotina | fungi |
| 32 | Cochliobolus carbonum 26-r-13 | Pezizomycotina | fungi |
| 33 | Cochliobolus heterostrophus c5 3332 | Pezizomycotina | fungi |
| 34 | Cochliobolus heterostrophus c5 5759 | Pezizomycotina | fungi |
| 35 | Cochliobolus lunatus m118 | Pezizomycotina | fungi |
| 36 | Cochliobolus miyabeanus atcc 44560 | Pezizomycotina | fungi |
| 37 | Cochliobolus victoriae fi3 | Pezizomycotina | fungi |
| 38 | Cucurbitaria berberidis cbs 394.84 | Pezizomycotina | fungi |
| 39 | Dissoconium aciculare | Pezizomycotina | fungi |
| 40 | Dothistroma septosporum nze10 | Pezizomycotina | fungi |
| 41 | Dothidotthia symphoricarpi | Pezizomycotina | fungi |
| 42 | Hysterium pulicare | Pezizomycotina | fungi |
| 43 | Leptosphaeria maculans | Pezizomycotina | fungi |
| 44 | Lepidopterella palustris | Pezizomycotina | fungi |
| 45 | Lophiostoma macrostomum | Pezizomycotina | fungi |
| 46 | Macrophomina phaseolina ms6 | Pezizomycotina | fungi |
| 47 | Melanomma pulvis-pyrius | Pezizomycotina | fungi |
| 48 | Myriangium duriaei cbs 260.36 | Pezizomycotina | fungi |
| 49 | Neofusicoccum parvum ucrnp2 | Pezizomycotina | fungi |
| 50 | Piedraia hortae | Pezizomycotina | fungi |
| 51 | Pleomassaria siparia | Pezizomycotina | fungi |
| 52 | Pyrenophora teres f. teres | Pezizomycotina | fungi |
| 53 | Pyrenophora tritici-repentis pt-1c-bfp 3136 | Pezizomycotina | fungi |
| 54 | Pyrenophora tritici-repentis pt-1c-bfp 5809 | Pezizomycotina | fungi |
| 55 | Rhytidhysteron rufulum | Pezizomycotina | fungi |
| 56 | Septoria musiva so2202 | Pezizomycotina | fungi |
| 57 | Septoria populicola | Pezizomycotina | fungi |
| 58 | Thermomyces stellatus cbs 241.64 | Pezizomycotina | fungi |
| 59 | Trypethelium eluteriae | Pezizomycotina | fungi |
| 60 | Zasmidium cellare atcc 36951 | Pezizomycotina | fungi |
| 61 | Zopfia rhizophila | Pezizomycotina | fungi |
| 62 | Cladosporium fulvum | Pezizomycotina | fungi |
| 63 | Cochliobolus sativus nd90pr | Pezizomycotina | fungi |
| 64 | Didymella exigua cbs 183.55 | Pezizomycotina | fungi |
| 65 | Lentithecium fluviatile | Pezizomycotina | fungi |
| 66 | Patellaria atrata | Pezizomycotina | fungi |
| 67 | Polychaeton citri | Pezizomycotina | fungi |
| 68 | Setosphaeria turcica et28a | Pezizomycotina | fungi |
| 69 | Sporormia fimetaria | Pezizomycotina | fungi |
| 70 | Xanthoria parietina | Pezizomycotina | fungi |
| 71 | Ajellomyces capsulatus NAmI WU24 | Pezizomycotina | fungi |
| 72 | Ajellomyces dermatitidis ER-3 | Pezizomycotina | fungi |
| 73 | Alternaria brassicicola | Pezizomycotina | fungi |
| 74 | Ascosphaera apis | Pezizomycotina | fungi |
| 75 | Aspergillus clavatus | Pezizomycotina | fungi |
| 76 | Aspergillus fischeri | Pezizomycotina | fungi |
| 77 | Aspergillus flavus | Pezizomycotina | fungi |
| 78 | Aspergillus fumigatus | Pezizomycotina | fungi |
| 79 | Aspergillus kawachii | Pezizomycotina | fungi |
| 80 | Aspergillus nidulans 2095 | Pezizomycotina | fungi |
| 81 | Aspergillus nidulans 1855 | Pezizomycotina | fungi |
| 82 | Aspergillus oryzae | Pezizomycotina | fungi |
| 83 | Aspergillus terreus | Pezizomycotina | fungi |
| 84 | Botrytis cinerea | Pezizomycotina | fungi |
| 85 | Chaetomium globosum | Pezizomycotina | fungi |
| 86 | Coccidioides immitis RS | Pezizomycotina | fungi |
| 87 | Coccidioides posadasii RMSCC\_3488 | Pezizomycotina | fungi |
| 88 | Cryphonectria parasitica 3352 | Pezizomycotina | fungi |
| 89 | Cryphonectria parasitica 4119 | Pezizomycotina | fungi |
| 90 | Fusarium graminearum ph1 | Pezizomycotina | fungi |
| 91 | Fusarium oxysporum lycopersici | Pezizomycotina | fungi |
| 92 | Fusarium verticillioides | Pezizomycotina | fungi |
| 93 | Magnaporthe grisea | Pezizomycotina | fungi |
| 94 | Microsporum canis CBS 113480 | Pezizomycotina | fungi |
| 95 | Microsporum gypseum CBS 118893 | Pezizomycotina | fungi |
| 96 | Mycosphaerella fijiensis | Pezizomycotina | fungi |
| 97 | Mycosphaerella graminicola | Pezizomycotina | fungi |
| 98 | Nectria haematococca MPVI | Pezizomycotina | fungi |
| 99 | Neurospora crassa | Pezizomycotina | fungi |
| 100 | Neurospora discreta FGSC 8579 mat A | Pezizomycotina | fungi |
| 101 | Neurospora tetrasperma FGSC 2508 mat A | Pezizomycotina | fungi |
| 102 | Paracoccidioides brasiliensis Pb03 | Pezizomycotina | fungi |
| 103 | Penicillium chrysogenum | Pezizomycotina | fungi |
| 104 | Penicillium marneffei ATCC 18224 | Pezizomycotina | fungi |
| 105 | Podospora anserina | Pezizomycotina | fungi |
| 106 | Sclerotinia sclerotiorum | Pezizomycotina | fungi |
| 107 | Stagonospora nodorum | Pezizomycotina | fungi |
| 108 | Talaromyces stipitatus | Pezizomycotina | fungi |
| 109 | Thielavia terrestris | Pezizomycotina | fungi |
| 110 | Trichoderma atroviride | Pezizomycotina | fungi |
| 111 | Trichophyton equinum CBS127.97 | Pezizomycotina | fungi |
| 112 | Trichoderma reesei | Pezizomycotina | fungi |
| 113 | Trichoderma virens Gv29-8 | Pezizomycotina | fungi |
| 114 | Tuber melanosporum | Pezizomycotina | fungi |
| 115 | Uncinocarpus reesii 5820 | Pezizomycotina | fungi |
| 116 | Uncinocarpus reesii 2939 | Pezizomycotina | fungi |
| 117 | Verticillium albo-atrum VaMs.102 | Pezizomycotina | fungi |
| 118 | Verticillium dahliae VdLs.17 | Pezizomycotina | fungi |
| 119 | Phaeosphaeria nodorum SN15 | Pezizomycotina | fungi |
| 120 | Schizosaccharomyces japonicus | Taphrinomycotina | fungi |
| 121 | Schizosaccharomyce octosporus | Taphrinomycotina | fungi |
| 122 | Schizosaccharomyces pombe | Taphrinomycotina | fungi |
| 123 | Schizosaccharomyces sp. OY26 | Taphrinomycotina | fungi |
| 124 | Coprinopsis cinerea | Basidiomycota | fungi |
| 125 | Cryptococcus neoformans JEC21 | Basidiomycota | fungi |
| 126 | Gelatoporia subvermispora | Basidiomycota | fungi |
| 127 | Heterobasidion annosum | Basidiomycota | fungi |
| 128 | Laccaria bicolor | Basidiomycota | fungi |
| 129 | Malassezia globosa CBS 7966 | Basidiomycota | fungi |
| 130 | Melampsora laricis-populina | Basidiomycota | fungi |
| 131 | Moniliophthora perniciosa FA553 | Basidiomycota | fungi |
| 132 | Phanerochaete chrysosporium P-78 | Basidiomycota | fungi |
| 133 | Pleurotus ostreatus PC15 | Basidiomycota | fungi |
| 134 | Postia placenta | Basidiomycota | fungi |
| 135 | Puccinia graminis | Basidiomycota | fungi |
| 136 | Schizophyllum commune | Basidiomycota | fungi |
| 137 | Serpula lacrymans S7\_3 | Basidiomycota | fungi |
| 138 | Sporobolomyces roseus | Basidiomycota | fungi |
| 139 | Tremella mesenterica Fries | Basidiomycota | fungi |
| 140 | Ustilago maydis | Basidiomycota | fungi |
| 141 | Mucor circinelloides | Mucoromycotina | fungi |
| 142 | Phycomyces blakesleeanus | Mucoromycotina | fungi |
| 143 | Rhizopus oryzae | Mucoromycotina | fungi |
| 144 | Batrachochytrium dendrobatidis | Chytridiomycota | fungi |
| 145 | Spizellomyces punctatus | Chytridiomycota | fungi |
| 146 | Encephalitozoon hellem | microsporidia | microsporidia |
| 147 | Encephalitozoon intestinalis | microsporidia | microsporidia |
| 148 | Encephalitozoon cuniculi | microsporidia | microsporidia |
| 149 | Nosema ceranae | microsporidia | microsporidia |
| 150 | Enterocytozoon bieneusi | microsporidia | microsporidia |
| 151 | Antonospora locustae | microsporidia | microsporidia |
| 152 | Edhazardia aedis | microsporidia | microsporidia |
| 153 | Vavraia culicis floridensis | microsporidia | microsporidia |
| 154 | Nematocida parisii | microsporidia | microsporidia |
| 155 | Anncaliia algerae PRA339 | microsporidia | microsporidia |
| 156 | Vittaforma corneae | microsporidia | microsporidia |
| 157 | Anas platyrhynchos | Metazoa | unikonta |
| 158 | Latimeria chalumnae | Metazoa | unikonta |
| 159 | mustela putorius furo | Metazoa | unikonta |
| 160 | Linepithema humile | Metazoa | unikonta |
| 161 | Pelodiscus sinensis | Metazoa | unikonta |
| 162 | Acropora digitifera | Metazoa | unikonta |
| 163 | Acyrthosiphon pisum | Metazoa | unikonta |
| 164 | Aedes aegypti | Metazoa | unikonta |
| 165 | Ailuropoda melanoleuca | Metazoa | unikonta |
| 166 | Amphimedon queenslandica | Metazoa | unikonta |
| 167 | Anolis carolinensis | Metazoa | unikonta |
| 168 | Anopheles gambiae | Metazoa | unikonta |
| 169 | Apis mellifera | Metazoa | unikonta |
| 170 | Bombyx mori | Metazoa | unikonta |
| 171 | Bos taurus | Metazoa | unikonta |
| 172 | Branchiostoma floridae | Metazoa | unikonta |
| 173 | Caenorhabditis brenneri 2851 | Metazoa | unikonta |
| 174 | Caenorhabditis brenneri 70 | Metazoa | unikonta |
| 175 | Caenorhabditis elegans | Metazoa | unikonta |
| 176 | Caenorhabditis japonica | Metazoa | unikonta |
| 177 | Caenorhabditis remanei | Metazoa | unikonta |
| 178 | Callithrix jacchus | Metazoa | unikonta |
| 179 | Canis familiaris | Metazoa | unikonta |
| 180 | Capitella capitata | Metazoa | unikonta |
| 181 | Cavia porcellus | Metazoa | unikonta |
| 182 | Choloepus hoffmanni | Metazoa | unikonta |
| 183 | Ciona intestinalis | Metazoa | unikonta |
| 184 | Ciona savignyi | Metazoa | unikonta |
| 185 | Culex pipiens quinquefasciatus | Metazoa | unikonta |
| 186 | Danio rerio | Metazoa | unikonta |
| 187 | Daphnia pulex | Metazoa | unikonta |
| 188 | Dasypus novemcinctus | Metazoa | unikonta |
| 189 | Dipodomys ordii | Metazoa | unikonta |
| 190 | Drosophila ananassae | Metazoa | unikonta |
| 191 | Drosophila erecta | Metazoa | unikonta |
| 192 | Drosophila grimshawi | Metazoa | unikonta |
| 193 | Drosophila melanogaster | Metazoa | unikonta |
| 194 | Drosophila mojavensis | Metazoa | unikonta |
| 195 | Drosophila persimilis | Metazoa | unikonta |
| 196 | Drosophila pseudoobscura | Metazoa | unikonta |
| 197 | Drosophila sechellia | Metazoa | unikonta |
| 198 | Drosophila simulans | Metazoa | unikonta |
| 199 | Drosophila virilis | Metazoa | unikonta |
| 200 | Drosophila willistoni | Metazoa | unikonta |
| 201 | Drosophila yakuba | Metazoa | unikonta |
| 202 | Echinops telfairi | Metazoa | unikonta |
| 203 | Equus caballus | Metazoa | unikonta |
| 204 | Erinaceus europaeus | Metazoa | unikonta |
| 205 | Felis catus | Metazoa | unikonta |
| 206 | Takifugu rubripes | Metazoa | unikonta |
| 207 | Gadus morhua | Metazoa | unikonta |
| 208 | Gallus gallus | Metazoa | unikonta |
| 209 | Gasterosteus aculeatus | Metazoa | unikonta |
| 210 | Gorilla gorilla | Metazoa | unikonta |
| 211 | Helobdella robusta | Metazoa | unikonta |
| 212 | Homo sapiens | Metazoa | unikonta |
| 213 | Hydra magnipapillata | Metazoa | unikonta |
| 214 | Ixodes scapularis | Metazoa | unikonta |
| 215 | Lama pacos | Metazoa | unikonta |
| 216 | Lepisosteus oculatus | Metazoa | unikonta |
| 217 | Loa loa | Metazoa | unikonta |
| 218 | Lottia gigantea | Metazoa | unikonta |
| 219 | Loxodonta africana | Metazoa | unikonta |
| 220 | Macropus eugenii | Metazoa | unikonta |
| 221 | Macaca mulatta | Metazoa | unikonta |
| 222 | Microcebus murinus | Metazoa | unikonta |
| 223 | Monodelphis domestica | Metazoa | unikonta |
| 224 | Mus musculus | Metazoa | unikonta |
| 225 | Myotis lucifugus | Metazoa | unikonta |
| 226 | Nasonia vitripennis | Metazoa | unikonta |
| 227 | Nematostella vectensis | Metazoa | unikonta |
| 228 | Nomascus leucogenys | Metazoa | unikonta |
| 229 | Ochotona princeps | Metazoa | unikonta |
| 230 | Ornithorhynchus anatinus | Metazoa | unikonta |
| 231 | Oryctolagus cuniculus | Metazoa | unikonta |
| 232 | Oryzias latipes | Metazoa | unikonta |
| 233 | Otolemur garnettii | Metazoa | unikonta |
| 234 | Pan troglodytes | Metazoa | unikonta |
| 235 | Pediculus humanus | Metazoa | unikonta |
| 236 | Petromyzon marinus | Metazoa | unikonta |
| 237 | Pongo pygmaeus | Metazoa | unikonta |
| 238 | Pristionchus pacificus | Metazoa | unikonta |
| 239 | Procavia capensis | Metazoa | unikonta |
| 240 | Pteropus vampyrus | Metazoa | unikonta |
| 241 | Rattus norvegicus | Metazoa | unikonta |
| 242 | Sarcophilus\_harrisii | Metazoa | unikonta |
| 243 | Schistosoma mansoni | Metazoa | unikonta |
| 244 | Sorex araneus | Metazoa | unikonta |
| 245 | Spermophilus tridecemlineatus | Metazoa | unikonta |
| 246 | Strongylocentrotus purpuratus | Metazoa | unikonta |
| 247 | Sus scrofa | Metazoa | unikonta |
| 248 | Taeniopygia guttata | Metazoa | unikonta |
| 249 | Tarsius syrichta | Metazoa | unikonta |
| 250 | Tetraodon nigroviridis | Metazoa | unikonta |
| 251 | Trichoplax adhaerens | Metazoa | unikonta |
| 252 | Tribolium castaneum | Metazoa | unikonta |
| 253 | Tupaia belangeri | Metazoa | unikonta |
| 254 | Tursiops truncatus | Metazoa | unikonta |
| 255 | Wuchereria bancrofti | Metazoa | unikonta |
| 256 | Xenopus tropicalis | Metazoa | unikonta |
| 257 | Callorhinchus milii | Metazoa | unikonta |
| 258 | Monosiga brevicollis | Monosiga\_brevicollis | unikonta |
| 259 | Capsaspora owczarzaki | Capsaspora\_owczarzaki | unikonta |
| 260 | Thecamonas trahens | Thecamonas\_trahens | unikonta |
| 261 | Bigelowiella natans | Amoebozoa | unikonta |
| 262 | Dictyostelium discoideum AX4 | Amoebozoa | unikonta |
| 263 | Dictyostelium purpureum QSDP1 | Amoebozoa | unikonta |
| 264 | Entamoeba dispar SAW760 | Amoebozoa | unikonta |
| 265 | Entamoeba histolytica | Amoebozoa | unikonta |
| 266 | Polysphondylium pallidum | Amoebozoa | unikonta |
| 267 | Leishmania braziliensis | Euglenozoa | eukaryota |
| 268 | Leishmania infantum | Euglenozoa | eukaryota |
| 269 | Leishmania major strain Friedlin | Euglenozoa | eukaryota |
| 270 | Trypanosoma brucei | Euglenozoa | eukaryota |
| 271 | Naegleria gruberi | Heterolobosea | eukaryota |
| 272 | Aquilegia coerulea | Streptophyta | eukaryota |
| 273 | Arabidopsis lyrata | Streptophyta | eukaryota |
| 274 | Arabidopsis thaliana | Streptophyta | eukaryota |
| 275 | Brachypodium distachyon | Streptophyta | eukaryota |
| 276 | Brassica rapa | Streptophyta | eukaryota |
| 277 | Capsella rubella | Streptophyta | eukaryota |
| 278 | Citrus clementina | Streptophyta | eukaryota |
| 279 | Citrus sinensis | Streptophyta | eukaryota |
| 280 | Cucumis sativus | Streptophyta | eukaryota |
| 281 | Eucalyptus grandis | Streptophyta | eukaryota |
| 282 | Glycine max | Streptophyta | eukaryota |
| 283 | Linum usitatissimum | Streptophyta | eukaryota |
| 284 | Malus x domestica | Streptophyta | eukaryota |
| 285 | Manihot esculenta | Streptophyta | eukaryota |
| 286 | Medicago truncatula | Streptophyta | eukaryota |
| 287 | Mimulus guttatus | Streptophyta | eukaryota |
| 288 | Oryza sativa sp. japonica | Streptophyta | eukaryota |
| 289 | Phaseolus vulgaris | Streptophyta | eukaryota |
| 290 | Physcomitrella patens sp. patens | Streptophyta | eukaryota |
| 291 | Populus trichocarpa | Streptophyta | eukaryota |
| 292 | Prunus persica | Streptophyta | eukaryota |
| 293 | Ricinus communis | Streptophyta | eukaryota |
| 294 | Selaginella moellendorffii | Streptophyta | eukaryota |
| 295 | Setaria italica | Streptophyta | eukaryota |
| 296 | Solanum lycopersicum | Streptophyta | eukaryota |
| 297 | Sorghum bicolor | Streptophyta | eukaryota |
| 298 | Vitis vinifera | Streptophyta | eukaryota |
| 299 | Zea mays | Streptophyta | eukaryota |
| 300 | Thellungiella halophila | Streptophyta | eukaryota |
| 301 | Chlorella sp. NC64A | Chlorophyta | eukaryota |
| 302 | Chlamydomonas reinhardtii | Chlorophyta | eukaryota |
| 303 | Micromonas sp. CCMP490 | Chlorophyta | eukaryota |
| 304 | Micromonas pusilla sp. RCC299 | Chlorophyta | eukaryota |
| 305 | Ostreococcus lucimarinus | Chlorophyta | eukaryota |
| 306 | Ostreococcus sp. RCC809 | Chlorophyta | eukaryota |
| 307 | Ostreococcus tauri | Chlorophyta | eukaryota |
| 308 | Volvox carteri f. nagariensis | Chlorophyta | eukaryota |
| 309 | Coccomyxa subellipsoidea | Chlorophyta | eukaryota |
| 310 | Cyanidioschyzon merolae | Rhodophyta | eukaryota |
| 311 | Aureococcus anophagefferens | Stramenopiles | eukaryota |
| 312 | Ectocarpus siliculosus | Stramenopiles | eukaryota |
| 313 | Fragilariopsis cylindrus CCMP 1102 | Stramenopiles | eukaryota |
| 314 | Phaeodactylum tricornutum | Stramenopiles | eukaryota |
| 315 | Phytophthora infestans | Stramenopiles | eukaryota |
| 316 | Phytophthora ramorum | Stramenopiles | eukaryota |
| 317 | Phytophthora sojae | Stramenopiles | eukaryota |
| 318 | Saprolegnia parasitica | Stramenopiles | eukaryota |
| 319 | Thalassiosira pseudonana | Stramenopiles | eukaryota |
| 320 | Babesia bovis | Alveolata | eukaryota |
| 321 | Cryptosporidium hominis ATCC BAA-381 | Alveolata | eukaryota |
| 322 | Eimeria tenella | Alveolata | eukaryota |
| 323 | Neospora caninum | Alveolata | eukaryota |
| 324 | Paramecium tetraurelia | Alveolata | eukaryota |
| 325 | Perkinsus marinus | Alveolata | eukaryota |
| 326 | Plasmodium berghei | Alveolata | eukaryota |
| 327 | Plasmodium chabaudi | Alveolata | eukaryota |
| 328 | Plasmodium falciparum | Alveolata | eukaryota |
| 329 | Plasmodium gallinaceum | Alveolata | eukaryota |
| 330 | Plasmodium knowlesi | Alveolata | eukaryota |
| 331 | Plasmodium reichenowi | Alveolata | eukaryota |
| 332 | Plasmodium vivax | Alveolata | eukaryota |
| 333 | Plasmodium yoelii | Alveolata | eukaryota |
| 334 | Tetrahymena thermophila | Alveolata | eukaryota |
| 335 | Theileria annulata | Alveolata | eukaryota |
| 336 | Theileria parva | Alveolata | eukaryota |
| 337 | Toxoplasma gondii | Alveolata | eukaryota |
| 338 | Emiliania huxleyi CCMP1516 | Haptophyceae | eukaryota |
| 339 | Hemiselmis andersenii | Cryptophyta | eukaryota |
| 340 | Guillardia theta | Cryptophyta | eukaryota |
| 341 | Hemiselmis andersenii | Cryptophyta | eukaryota |
| 342 | Archaeoglobus fulgidus | Euryarchaeota | archaea |
| 343 | Methanococcoides burtonii | Euryarchaeota | archaea |
| 344 | Methanopyrus kandleri | Euryarchaeota | archaea |
| 345 | Methanocorpusculum labreanum | Euryarchaeota | archaea |
| 346 | Natronomonas pharaonis | Euryarchaeota | archaea |
| 347 | Haloferax volcanii DS2 | Euryarchaeota | archaea |
| 348 | Methanosarcina barkeri str. Fusaro | Euryarchaeota | archaea |
| 349 | Methanocaldococcus jannaschii DSM 2661 | Euryarchaeota | archaea |
| 350 | Methanothermobacter thermautotrophicus str. Delta H | Euryarchaeota | archaea |
| 351 | Picrophilus torridus DSM 9790 | Euryarchaeota | archaea |
| 352 | Pyrococcus horikoshii | Euryarchaeota | archaea |
| 353 | Thermoplasma acidophilum DSM 1728 | Euryarchaeota | archaea |
| 354 | Thermococcus kodakarensis KOD1 | Euryarchaeota | archaea |
| 355 | Nanoarchaeum equitans | Nanoarchaeota | archaea |
| 356 | Candidatus Korarchaeum cryptofilum OPF8 | Korarchaeota | archaea |
| 357 | Aeropyrum pernix K1 | Crenarchaeota | archaea |
| 358 | Ignicoccus hospitalis | Crenarchaeota | archaea |
| 359 | Metallosphaera sedula | Crenarchaeota | archaea |
| 360 | Pyrobaculum neutrophilum | Crenarchaeota | archaea |
| 361 | Thermofilum pendens | Crenarchaeota | archaea |
| 362 | Caldivirga maquilingensis | Crenarchaeota | archaea |
| 363 | Sulfolobus solfataricus P2 | Crenarchaeota | archaea |
| 364 | Candidatus Caldiarchaeum subterraneum | Thaumarchaeota | archaea |
| 365 | Cenarchaeum symbiosum | Thaumarchaeota | archaea |
| 366 | Nitrosopumilus maritimus | Thaumarchaeota | archaea |
| 367 | Candidatus Nitrososphaera gargensis Ga9.2 | Thaumarchaeota | archaea |
| 368 | Deinococcus proteolyticus MRP | Deinococci | bacteria |
| 369 | Marinithermus hydrothermalis DSM 14884 | Deinococci | bacteria |
| 370 | Clostridium tetani E88 | Firmicutes | bacteria |
| 371 | Coprothermobacter proteolyticus DSM 5265 | Firmicutes | bacteria |
| 372 | Desulfotomaculum acetoxidans DSM 771 | Firmicutes | bacteria |
| 373 | Acaryochloris marina | Cyanobacteria | bacteria |
| 374 | Acaryochloris marina | Cyanobacteria | bacteria |
| 375 | Anabaena cylindrica | Cyanobacteria | bacteria |
| 376 | Anabaena sp. | Cyanobacteria | bacteria |
| 377 | Anabaena variabilis ATCC 29413 | Cyanobacteria | bacteria |
| 378 | Arthrospira platensis | Cyanobacteria | bacteria |
| 379 | Calothrix sp. 5685 | Cyanobacteria | bacteria |
| 380 | Calothrix sp. 5686 | Cyanobacteria | bacteria |
| 381 | Chamaesiphon minutus | Cyanobacteria | bacteria |
| 382 | Chlorogloeopsis fritschii | Cyanobacteria | bacteria |
| 383 | Chlorogloeopsis sp. | Cyanobacteria | bacteria |
| 384 | Chroococcidiopsis thermalis | Cyanobacteria | bacteria |
| 385 | Crinalium epipsammum | Cyanobacteria | bacteria |
| 386 | Cyanobacterium aponinum | Cyanobacteria | bacteria |
| 387 | Cyanothece ATCC 51142 | Cyanobacteria | bacteria |
| 388 | Cyanobium gracile | Cyanobacteria | bacteria |
| 389 | Cyanothece sp. 5693 | Cyanobacteria | bacteria |
| 390 | Cyanothece sp. 5694 | Cyanobacteria | bacteria |
| 391 | Cyanothece sp. 5695 | Cyanobacteria | bacteria |
| 392 | Cyanothece sp. 5696 | Cyanobacteria | bacteria |
| 393 | Cyanothece sp. 5697 | Cyanobacteria | bacteria |
| 394 | Cyanothece sp. 5698 | Cyanobacteria | bacteria |
| 395 | Cyanobacterium stanieri | Cyanobacteria | bacteria |
| 396 | Cyanobacterium UCYN-A | Cyanobacteria | bacteria |
| 397 | Cylindrospermum stagnale | Cyanobacteria | bacteria |
| 398 | Dactylococcopsis salina | Cyanobacteria | bacteria |
| 399 | Fischerella muscicola 5744 | Cyanobacteria | bacteria |
| 400 | Fischerella muscicola 5745 | Cyanobacteria | bacteria |
| 401 | Fischerella sp. | Cyanobacteria | bacteria |
| 402 | Geitlerinema sp. | Cyanobacteria | bacteria |
| 403 | Gloeocapsa sp. | Cyanobacteria | bacteria |
| 404 | Gloeobacter violaceus 4698 | Cyanobacteria | bacteria |
| 405 | Gloeobacter violaceus 5702 | Cyanobacteria | bacteria |
| 406 | Halothece sp. | Cyanobacteria | bacteria |
| 407 | Leptolyngbya sp. | Cyanobacteria | bacteria |
| 408 | Microcystis aeruginosa NIES 843 | Cyanobacteria | bacteria |
| 409 | Microcoleus sp. | Cyanobacteria | bacteria |
| 410 | Nostoc azollae 0708 | Cyanobacteria | bacteria |
| 411 | Nostoc punctiforme PCC 73102 | Cyanobacteria | bacteria |
| 412 | Nostoc sp. 5707 | Cyanobacteria | bacteria |
| 413 | Nostoc sp. 5708 | Cyanobacteria | bacteria |
| 414 | Nostoc sp. 5709 | Cyanobacteria | bacteria |
| 415 | Oscillatoria acuminata | Cyanobacteria | bacteria |
| 416 | Oscillatoria nigro-viridis | Cyanobacteria | bacteria |
| 417 | Pleurocapsa sp. | Cyanobacteria | bacteria |
| 418 | Prochlorococcus marinus AS9601 4702 | Cyanobacteria | bacteria |
| 419 | Prochlorococcus marinus AS9601 5713 | Cyanobacteria | bacteria |
| 420 | Prochlorococcus marinus AS9601 5714 | Cyanobacteria | bacteria |
| 421 | Prochlorococcus marinus AS9601 5715 | Cyanobacteria | bacteria |
| 422 | Prochlorococcus marinus AS9601 5716 | Cyanobacteria | bacteria |
| 423 | Prochlorococcus marinus AS9601 5717 | Cyanobacteria | bacteria |
| 424 | Prochlorococcus marinus AS9601 5718 | Cyanobacteria | bacteria |
| 425 | Prochlorococcus marinus AS9601 5719 | Cyanobacteria | bacteria |
| 426 | Prochlorococcus marinus AS9601 5720 | Cyanobacteria | bacteria |
| 427 | Prochlorococcus marinus AS9601 5721 | Cyanobacteria | bacteria |
| 428 | Prochlorococcus marinus AS9601 5722 | Cyanobacteria | bacteria |
| 429 | Prochlorococcus marinus AS9601 5723 | Cyanobacteria | bacteria |
| 430 | Prochlorococcus marinus AS9601 5724 | Cyanobacteria | bacteria |
| 431 | Pseudanabaena sp. | Cyanobacteria | bacteria |
| 432 | Rivularia sp. | Cyanobacteria | bacteria |
| 433 | Scytonema hofmanni | Cyanobacteria | bacteria |
| 434 | Stanieria cyanosphaera | Cyanobacteria | bacteria |
| 435 | Synechococcus elongatus PCC 7942 4703 | Cyanobacteria | bacteria |
| 436 | Synechococcus elongatus PCC 7942 4704 | Cyanobacteria | bacteria |
| 437 | Synechococcus\_sp\_JA-2-3Ba\_2-13 4694 | Cyanobacteria | bacteria |
| 438 | Synechococcus\_sp\_JA-2-3Ba\_2-13 4695 | Cyanobacteria | bacteria |
| 439 | Synechocystis sp. 5728 | Cyanobacteria | bacteria |
| 440 | Synechocystis sp. 5729 | Cyanobacteria | bacteria |
| 441 | Synechocystis sp. 5730 | Cyanobacteria | bacteria |
| 442 | Synechocystis sp. 5731 | Cyanobacteria | bacteria |
| 443 | Synechocystis sp. 5731 | Cyanobacteria | bacteria |
| 444 | Synechocystis sp. 5733 | Cyanobacteria | bacteria |
| 445 | Synechocystis sp. 5734 | Cyanobacteria | bacteria |
| 446 | Synechocystis sp. 5735 | Cyanobacteria | bacteria |
| 447 | Synechocystis sp. 5736 | Cyanobacteria | bacteria |
| 448 | Synechocystis sp. 5737 | Cyanobacteria | bacteria |
| 449 | Synechocystis sp. 5738 | Cyanobacteria | bacteria |
| 450 | Synechocystis sp. 5739 | Cyanobacteria | bacteria |
| 451 | Synechocystis sp. 5740 | Cyanobacteria | bacteria |
| 452 | Thermosynechococcus elongatus 4705 | Cyanobacteria | bacteria |
| 453 | Thermosynechococcus elongatus 5741 | Cyanobacteria | bacteria |
| 454 | Trichodesmium erythraeum IMS101 | Cyanobacteria | bacteria |
| 455 | Clavibacter michiganensis subsp. michiganensis NCPPB 382 | Actinobacteria | bacteria |
| 456 | Conexibacter woesei DSM 14684 | Actinobacteria | bacteria |
| 457 | Chlamydophila psittaci 6BC | Chlamydiae | bacteria |
| 458 | Candidatus Azobacteroides pseudotrichonymphae genomovar. CFP2 | Bacteroidetes | bacteria |
| 459 | Candidatus Sulcia muelleri DMIN | Bacteroidetes | bacteria |
| 460 | Campylobacter curvus 525.92 | Epsilonproteobacteria | bacteria |
| 461 | Nitratiruptor sp. SB155-2 | Epsilonproteobacteria | bacteria |
| 462 | Sulfurovum sp. NBC37-1 | Epsilonproteobacteria | bacteria |
| 463 | Bdellovibrio bacteriovorus HD100 | Deltaproteobacteria | bacteria |
| 464 | Desulfovibrio vulgaris DP4 | Deltaproteobacteria | bacteria |
| 465 | Geobacter sulfurreducens PCA | Deltaproteobacteria | bacteria |
| 466 | Sorangium cellulosum So ce 56 | Deltaproteobacteria | bacteria |
| 467 | Syntrophus aciditrophicus SB | Deltaproteobacteria | bacteria |
| 468 | Agrobacterium fabrum | Alphaproteobacteria | bacteria |
| 469 | Caulobacter crescentus CB15 | Alphaproteobacteria | bacteria |
| 470 | Ehrlichia canis str. Jake | Alphaproteobacteria | bacteria |
| 471 | Maricaulis maris MCS10 | Alphaproteobacteria | bacteria |
| 472 | Zymomonas mobilis subsp. mobilis ZM4 | Alphaproteobacteria | bacteria |
| 473 | Bordetella petrii DSM 12804 | Betaproteobacteria | bacteria |
| 474 | Chlamydia trachomatis G/9301 | Betaproteobacteria | bacteria |
| 475 | Dechloromonas aromatica RCB | Betaproteobacteria | bacteria |
| 476 | Methylobacillus flagellatus KT | Betaproteobacteria | bacteria |
| 477 | Neisseria gonorrhoeae FA 1090 | Betaproteobacteria | bacteria |
| 478 | Nitrosomonas europaea ATCC 19718 | Betaproteobacteria | bacteria |
| 479 | Thiobacillus denitrificans ATCC 25259 | Betaproteobacteria | bacteria |
| 480 | Aeromonas hydrophila subsp. hydrophila ATCC 7966 | Gammaproteobacteria | bacteria |
| 481 | Baumannia cicadellinicola str. Hc (Homalodisca coagulata) | Gammaproteobacteria | bacteria |
| 482 | Candidatus Carsonella ruddii PV | Gammaproteobacteria | bacteria |
| 483 | Coxiella burnetii RSA 331 | Gammaproteobacteria | bacteria |
| 484 | Dichelobacter nodosus VCS1703A | Gammaproteobacteria | bacteria |
| 485 | Escherichia coli str. K-12 substr. MG1655 | Gammaproteobacteria | bacteria |
| 486 | Haemophilus influenzae 10810 | Gammaproteobacteria | bacteria |
| 487 | Marinomonas mediterranea MMB-1 | Gammaproteobacteria | bacteria |
| 488 | Methylococcus capsulatus str. Bath | Gammaproteobacteria | bacteria |
| 489 | Nitrosococcus oceani ATCC 19707 | Gammaproteobacteria | bacteria |
| 490 | Pseudomonas putida F1 | Gammaproteobacteria | bacteria |
| 491 | Candidatus Ruthia magnifica str. Cm (Calyptogena magnifica) | Gammaproteobacteria | bacteria |

Table A‑5: List of 30 manually KO-annotated reference taxa

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| No. | Name | No. | Name | No. | Name |
| 1 | *A.gossypii* | 11 | *D.melanogaster* | 21 | *A.pernix* |
| 2 | *S.pombe* | 12 | *C.elegans* | 22 | *E.coli* |
| 3 | *C.albicans* | 13 | *M.brevicollis* | 23 | *N.meningtidis* |
| 4 | *S.cerevisiae* | 14 | *N.vectensis* | 24 | *H.pylori* |
| 5 | *N.crassa* | 15 | *E.histolytica* | 25 | *B.subtilis* |
| 6 | *A.nidulans* | 16 | *T.brucei* | 26 | *L.lactis* |
| 7 | *H.sapiens* | 17 | *A.thaliana* | 27 | *M.genitalium* |
| 8 | *M.musculus* | 18 | *P.falciparum 3D7* | 28 | *M.tuberculosis* |
| 9 | *R.norvegicus* | 19 | *C.hominis* | 29 | *Synechocystis sp.* |
| 10 | *D.rerio* | 20 | *M.jannaschii* | 30 | *A.aeolicus* |

Table A‑6: Network properties of core pathways for microsporidian LCA, 4 extant microsporidia species and *S.cerevisiae*.

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| Pathway | Source | Nodes | Edges | Avg. degree | Max degree | Avg. path length | Max path length (diameter) |
| Glycolysis / Gluconeogenesis | LCA microsporidia | 17 | 19 | 2,24 | 4 | 4,04 | 10 |
| Glycolysis / Gluconeogenesis | E.cuniculi | 14 | 14 | 2,00 | 3 | 3,95 | 9 |
| Glycolysis / Gluconeogenesis | E.hellem | 14 | 14 | 2,00 | 3 | 3,95 | 9 |
| Glycolysis / Gluconeogenesis | E.intestinalis | 14 | 14 | 2,00 | 3 | 3,95 | 9 |
| Glycolysis / Gluconeogenesis | N.ceranae | 13 | 12 | 1,85 | 3 | 3,77 | 9 |
| Glycolysis / Gluconeogenesis | S.cerevisiae | 27 | 48 | 3,56 | 9 | 4,59 | 11 |
| TCA cycle | LCA microsporidia | 3 | 3 | 2,00 | 2 | 1,00 | 1 |
| TCA cycle | E.cuniculi | 2 | 1 | 1,00 | 1 | 1,00 | 1 |
| TCA cycle | E.hellem | 2 | 1 | 1,00 | 1 | 1,00 | 1 |
| TCA cycle | E.intestinalis | 2 | 1 | 1,00 | 1 | 1,00 | 1 |
| TCA cycle | N.ceranae | 2 | 1 | 1,00 | 1 | 1,00 | 1 |
| TCA cycle | S.cerevisiae | 20 | 39 | 3,90 | 5 | 2,53 | 4 |
| Pentose phosphate pathway | LCA microsporidia | 10 | 15 | 3,00 | 6 | 1,93 | 4 |
| Pentose phosphate pathway | E.cuniculi | 10 | 15 | 3,00 | 6 | 1,93 | 4 |
| Pentose phosphate pathway | E.hellem | 10 | 15 | 3,00 | 6 | 1,93 | 4 |
| Pentose phosphate pathway | E.intestinalis | 10 | 15 | 3,00 | 6 | 1,93 | 4 |
| Pentose phosphate pathway | N.ceranae | 9 | 13 | 2,89 | 5 | 1,92 | 4 |
| Pentose phosphate pathway | S.cerevisiae | 15 | 32 | 4,27 | 10 | 2,01 | 4 |
| Purine metabolism | LCA microsporidia | 47 | 131 | 5,57 | 40 | 1,85 | 2 |
| Purine metabolism | E.cuniculi | 40 | 107 | 5,35 | 36 | 1,84 | 2 |
| Purine metabolism | E.hellem | 41 | 107 | 5,22 | 36 | 1,84 | 2 |
| Purine metabolism | E.intestinalis | 41 | 110 | 5,37 | 37 | 1,84 | 2 |
| Purine metabolism | N.ceranae | 30 | 55 | 3,67 | 27 | 1,85 | 2 |
| Purine metabolism | S.cerevisiae | 82 | 310 | 7,56 | 55 | 2,80 | 9 |
| Pyrimidine metabolism | LCA microsporidia | 46 | 85 | 3,70 | 40 | 2,05 | 3 |
| Pyrimidine metabolism | E.cuniculi | 38 | 66 | 3,47 | 35 | 1,99 | 3 |
| Pyrimidine metabolism | E.hellem | 39 | 67 | 3,44 | 35 | 2,08 | 4 |
| Pyrimidine metabolism | E.intestinalis | 40 | 69 | 3,45 | 36 | 2,04 | 4 |
| Pyrimidine metabolism | N.ceranae | 31 | 51 | 3,29 | 29 | 1,88 | 2 |
| Pyrimidine metabolism | S.cerevisiae | 65 | 161 | 4,95 | 51 | 2,61 | 8 |
| Amino acid metabolism | LCA microsporidia | 22 | 11 | 1,00 | 3 | 1,33 | 3 |
| Amino acid metabolism | E.cuniculi | 6 | 1 | 0,33 | 1 | 1,00 | 1 |
| Amino acid metabolism | E.hellem | 8 | 1 | 0,25 | 1 | 1,00 | 1 |
| Amino acid metabolism | E.intestinalis | 7 | 1 | 0,29 | 1 | 1,00 | 1 |
| Amino acid metabolism | N.ceranae | 8 | 2 | 0,50 | 1 | 1,00 | 1 |
| Amino acid metabolism | S.cerevisiae | 146 | 299 | 4,10 | 19 | 5,16 | 13 |

