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Antibody-dependent enhancement of severe dengue disease in humans

Leah C. Katzelnick,¹ Lionel Gresh,² M. Elizabeth Halloran,^{3,4} Juan Carlos Mercado,⁵ Guillermmina Kuan,⁶ Aubree Gordon,⁷ Angel Balmaseda,⁵ Eva Harris^{1*}

¹Division of Infectious Diseases and Vaccinology, School of Public Health, University of California, Berkeley, USA. ²Sustainable Sciences Institute, Managua, Nicaragua.

³Department of Biostatistics, University of Washington, USA. ⁴Vaccine and Infectious Disease Institute, Hutchinson Research Center, Seattle, Washington, USA.

⁵Laboratorio Nacional de Virología, Centro Nacional de Diagnóstico y Referencia, Ministry of Health, Managua, Nicaragua. ⁶Centro de Salud Sócrates Flores Vivas, Ministry of Health, Managua, Nicaragua. ⁷Department of Epidemiology, School of Public Health, University of Michigan, USA.

*Corresponding author. Email: eharris@berkeley.edu

For dengue viruses (DENV1-4), a specific range of antibody titer has been shown to enhance viral replication in vitro and severe disease in animal models. Although suspected, such antibody-dependent enhancement (ADE) of severe disease has not been shown to occur in humans. Using multiple statistical approaches to study a long-term pediatric cohort in Nicaragua, we show that risk of severe dengue disease is highest within a narrow range of pre-existing anti-DENV antibody titers. By contrast, we observe protection from all symptomatic dengue disease at high antibody titers. Thus, immune correlates of severe dengue must be evaluated separately from correlates of protection against symptomatic disease. These results have implications for studies of dengue pathogenesis and for vaccine development, because enhancement, not just lack of protection, is of concern.

Dengue viruses 1–4 (DENV1–4) are mosquito-borne flaviviruses that cause 50–100 million cases of Dengue Fever (DF) and ~500,000 hospitalizations annually (1, 2). Dengue Hemorrhagic Fever/Dengue Shock Syndrome (DHF/DSS) is the most severe form of dengue disease and is characterized by vascular leakage, hemorrhagic manifestations, thrombocytopenia, and hypotensive shock, which can lead to organ failure and death (3). Heterotypic secondary DENV infection (with a DENV type distinct from the primary infecting type) is the greatest risk factor for DHF/DSS (4–6). Age, interval between infections, antibody characteristics, and host-specific genetics are contributing factors (4–6). The theory of antibody-dependent enhancement (ADE) posits that at a specific concentration, heterotypic antibodies bind but do not neutralize virions of the subsequent infecting DENV type. These virus-immune complexes are recognized by Fc_Y receptors that facilitate virus entry and replication in target immune cells. This initiates an immune cascade that results in vascular leak and severe dengue disease (6, 7). In vitro and in animal models, a peak enhancement titer has been observed, i.e., a specific concentration of antibodies that most efficiently enhances DENV infection. In contrast, higher antibody concentrations effectively neutralize virions while lower concentrations poorly enhance infection (8, 9).

Yet there is no conclusive evidence in humans of a peak enhancement titer associated with the greatest risk of severe dengue disease. A recent phase 3 clinical trial observed that young dengue vaccine recipients had elevated risk of dengue

hospitalization >1 year after vaccination compared with placebo controls (10), raising concerns, but not confirming, that vaccination of DENV-naïve individuals induced poorly neutralizing anti-DENV antibodies that increased the risk of severe dengue disease (11). Further, the unexpected number of DHF/DSS cases in 6–12 month-old infants, when maternal derived-antibodies have decayed below neutralizing levels (12–18), is consistent with the concept of a peak enhancement titer for DHF/DSS; however, attempts to relate in vitro peak enhancement titer to disease severity in infants or older children have been unsuccessful (13, 15, 17–19).

Here, we directly studied the relationship between pre-existing anti-DENV binding antibodies (DENV-Abs) and dengue disease severity in a large, well-characterized pediatric cohort study in Managua, Nicaragua (20, 21). From August 2004 to April 2016, 8002 children ages 2–14 years were enrolled; 6684 children had at least one DENV-Ab titer measurement and were included in our study (41,302 samples in total, fig. S1 and table S1). DENV-Ab titers were measured using the inhibition enzyme-linked immunosorbent assay (iELISA) and estimated as the geometric mean of replicate titrations (quality control and reproducibility data in figs. S2 and S3) (22). In the iELISA, serially diluted serum antibodies competed with DENV-specific peroxidase-conjugated IgG for binding to a balanced mixture of DENV1–4 antigens (2). The iELISA measured antibodies binding cross-reactive epitopes such as the fusion loop in the envelope protein as well as the prM protein (table S2) that induce ADE in vitro and in vivo

(8, 9, 23). For comparison, iELISA titers are reliably (Pearson's correlation $r = 80$) a two-fold dilution higher than hemagglutination inhibition assay titers (24) and are correlated (Pearson's $r = 0.80$ [95% CI: 0.77-0.83]) with the geometric mean of neutralizing antibody titers to DENV1-4 and to heterologous DENV types (figs. S4 and S5 and table S3). As per the cohort protocol, children who became febrile visited the study health center, and those meeting the case definition for dengue or presenting with undifferentiated febrile illness were tested for dengue using molecular and serological diagnostic methods; those who developed warning signs for severe dengue disease were referred to the study hospital (618 dengue cases studied in total, table S4) (20). Disease severity was initially classified using 1997 World Health Organization (WHO) criteria of DF and DHF/DSS (table S5) (3).

We first compared individuals with different levels of pre-existing DENV-Ab titers (binned by four-fold serum dilution) to DENV-naïve individuals using a Cox proportional hazards model (25) adjusted for sex, epidemic season, age, and number of previous infections. Hazard ratio estimates of DHF/DSS across the range of DENV-Ab titers resembled the canonical ADE curve obtained in vitro (Figs. 1 and 2 and table S6) (27). The hazard of DHF/DSS was similar in children with no (DENV-naïve) or high ($>1:1280$) DENV-Ab titers. However, in children with pre-existing DENV-Ab levels of 1:21-1:80, the hazard of DHF/DSS was 7.64-fold higher [95% CI: 3.19-18.28] (Fig. 1A). These effects remained significant when adjusted for age or number of previous infections (fig. S6), and when analyzed with alternative DENV-Ab titer binning methods or sampling of individual iELISA titer measurements (figs. S7 to S9). During the 12 years of the cohort studied, a child with pre-existing DENV-Ab titers of 1:21-1:80 had a cumulative hazard of 11.4% for DHF/DSS. This is nearly twice as high as for a child with a prior DENV infection but low DENV-Ab titers ($<1:21$), who had a cumulative hazard of 6.6% of developing DHF/DSS (Fig. 1B). For both DENV-naïve children and children with high DENV-Ab titers ($>1:1280$), the cumulative hazard was 1.6% and 1.5%, respectively, indicating that high antibody levels did not provide any greater protection against DHF/DSS than having no pre-existing DENV-Ab. On average, in the PDCS, the DENV-Ab half-life was 4.00 years [95% CI: 3.81-4.20] and by three years post-infection, an estimated 22% of children had DENV-Ab titers of 1:21-1:80 (table S7). Children with subsequent severe dengue cases had lower, but not more rapidly decaying, DENV-Ab titers (table S8).

In 2009, WHO revised the classification guidelines for severe dengue to improve clinical management of dengue patients and to capture other complications. The 2009 guidelines replace the category of DHF/DSS with Dengue with Warning Signs (Dengue+Warning) and Severe Dengue (table S5) (2). We evaluated whether there is also a peak enhancement titer for Dengue+Warning Signs/Severe Dengue.

Again, we observed that the highest hazard ratio, 1.75 [95% CI: 1.11-2.74], occurred among children with DENV-Ab titers of 1:21-1:80 (Fig. 1, C and D and table S9). At higher antibody levels (1:321-1:1280 and $>1:1280$), the hazard ratios of Dengue+Warning Signs/Severe Dengue were less than those for DENV-naïve children, indicating that a protective effect against Dengue+Warning Signs/Severe Dengue is also antibody titer-dependent. We also estimated hazard ratios for hospitalization admissions with dengue. Similarly, a peak enhancement titer was observed at DENV-Ab titers of 1:21-1:80, with protection observed at higher titers (Fig. 1, E and F, table S10; previous infections only model, $p < 0.05$, fig. S6).

Hence, the magnitude of the enhancement effect observed related to how specific the definition of severe dengue disease was to the classical pathophysiological classification of DHF/DSS (26, 27). When we relaxed the case definition criteria further and modeled the hazard of having any dengue case, we did not observe a peak enhancement titer: children with a prior DENV infection and DENV-Ab titers $<1:21$ or 1:21-1:80 had comparable hazard ratios of dengue as DENV-naïve children (Fig. 1, G and H, and table S11). However, a protective effect was evident at DENV-Ab titers at and above 1:80-1:320.

Continuous hazard ratio curves for DHF/DSS rise and fall symmetrically around a peak hazard ratio of 5.95 [95% CI: 1.86-19.06], which occurred at a DENV-Ab titer of 1:34 (Fig. 2A) (28). When we controlled for prior DENV infection, children with DENV-Ab titers below this peak enhancement titer still had a lower hazard of DHF/DSS than those at the peak enhancement titer (Fig. 2B and fig. S10). Hazard ratio curves modeled on the basis of the WHO 2009 guidelines of Dengue+Warning Signs/Severe Dengue and hospitalization also peaked at 1:30, although the magnitude of the effect was smaller than for the DHF/DSS classification; again, protection was seen at higher DENV-Ab titers (Fig. 2, A and B).

To further test whether pre-existing DENV-Ab levels explained our observations of a peak enhancement titer, we compared severe secondary dengue cases each with five matched controls drawn randomly from the cohort. Controls were matched to cases by sex, age, and evidence of prior DENV infection, but had not experienced dengue in the epidemic season of the case. Conditional logistic regression was used to compare the pre-existing DENV-Ab titers of the severe cases with those of matched controls having titers $>1:320$. Again, a peak enhancement titer was observed at DENV-Ab titers of 1:21-1:80, with reduced odds ratios observed at lower and higher DENV-Ab titers (Fig. 3A and figs. S11 and S12) and the strongest enhancement effect seen for the DHF/DSS classification (Fig. 3, A to C, titer distributions, Fig. 3, D to F). In contrast, when non-severe secondary cases were compared to their own matched controls, no peak en-

hancement titer was observed (Fig. 3, A to C). We also observed the same peak enhancement titers when we directly compared severe to non-severe dengue cases according to pre-infection DENV-Ab titers, using logistic regression and accounting for known covariates (Fig. 4 and tables S12 to S14).

Thus, we have established that antibody titer does predict enhancement of severe dengue disease in humans. This association is seen most strongly for DHF/DSS, which is defined by the pathophysiological entity of DENV-induced vascular leak. In contrast, the 2009 WHO criteria, which are intentionally broader than DHF/DSS as they are meant improve triage and case management, encompass severe dengue cases caused by mechanisms other than vascular leak syndrome.

The correlate of risk for DHF/DSS and severe dengue disease (DENV-Ab titer 1:21-1:80) is distinct from the correlate of protection (DENV-Ab titers at and above 1:80-1:320) against symptomatic dengue. To date, dengue vaccine manufacturers have only been required to demonstrate induction of detectable neutralizing antibodies. With recent evidence of possible vaccine-enhanced dengue disease (10), this assumption is being revisited. A vaccine that induces antibody titers at, or near, the peak enhancement titer, may place vaccines at greater risk of severe dengue than if they had never been vaccinated (11). Further, immune correlates of severe dengue need to be estimated separately from correlates of protection against any dengue disease in vaccine trials and natural infection studies, an observation that may also be relevant to studies of Zika disease and vaccines.

We show that the iELISA, a simpler assay than neutralization tests (29, 30), is associated with elevated risk of severe disease as well as protection against symptomatic disease, making it a promising alternative method for measuring biologically predictive serological responses. Further, the iELISA measures antibodies targeting cross-reactive epitopes implicated in ADE in vitro and in vivo (8, 9, 23). The iELISA may directly measure the mechanistic correlate of enhancement and protection or may measure antibodies indirectly associated with the causal underlying immune determinants (31). Critical next steps toward identifying mechanistic correlates of protection and enhancement as well as of safe, protective dengue vaccines include development of serological assays that distinguish protective from enhancing antibodies, determination of how the sequence of infecting DENV types modifies disease, and integrated evaluation of cellular, innate, and humoral immunity to DENV infection and disease (32, 33).

In sum, we verify enhancement of dengue disease in humans and show that the level of pre-existing anti-DENV antibodies is directly associated with the severity of secondary dengue disease in humans. We also show that the immune correlate for enhanced severe dengue disease is distinct from

that for protection. These observations are important for future dengue and Zika vaccine trial design and evaluation, as well as for further studies on the mechanisms of ADE in relation to severe dengue and Zika disease.

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SUPPLEMENTARY MATERIALS

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Methods

Figs. S1 to S12

Tables S1 to S14

References (34–103)

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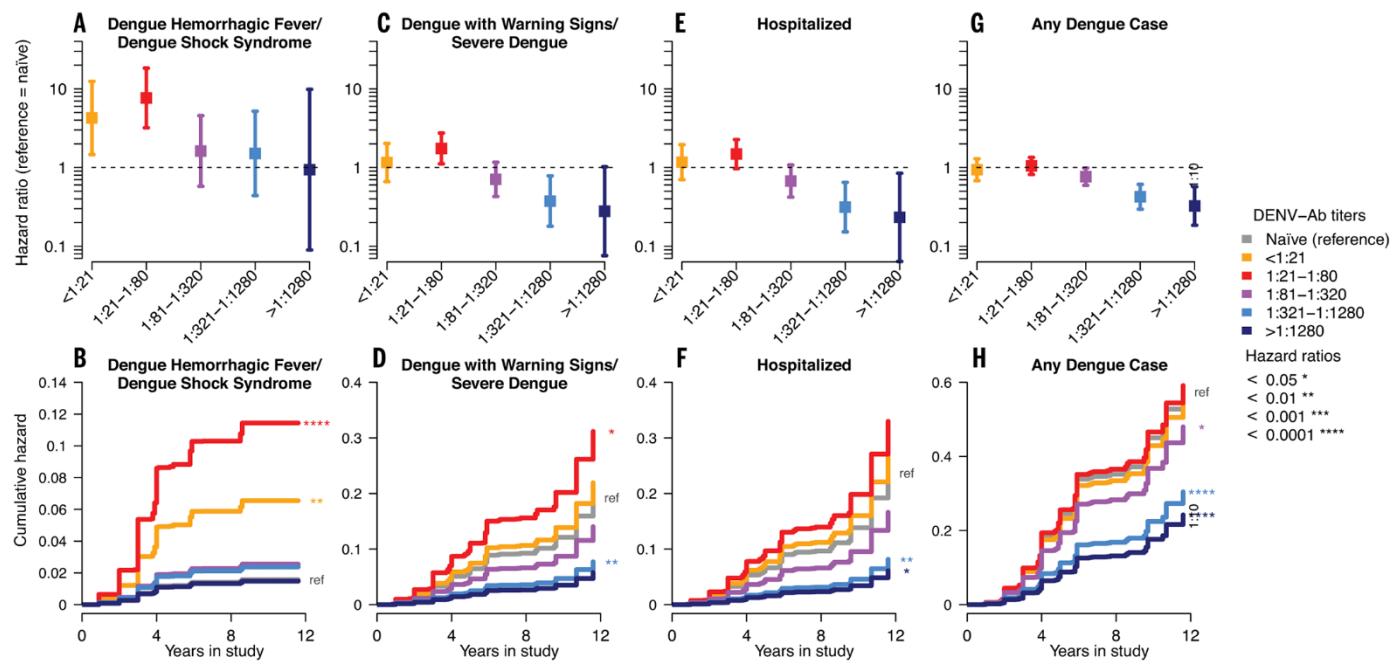


Fig. 1. Longitudinal analyses of the hazard of severe dengue disease or any dengue case by pre-existing DENV-Ab titer for the full Pediatric Dengue Cohort. Hazard ratios with 95% confidence intervals (A, C, E, and G) and cumulative hazard for an average child (B, D, F, and H) with pre-existing DENV-Ab titers binned by four-fold dilution. Cox proportional hazard models were adjusted for sex, epidemic season, age, and number of previous DENV infections. Average child = female, age 5–9, 2007–2008 epidemic season, and one previous DENV infection.

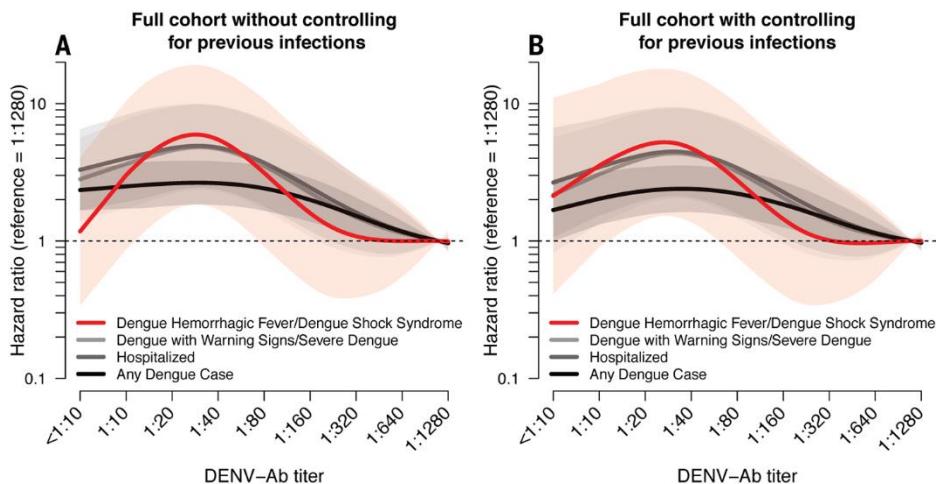


Fig. 2. Continuous hazard ratio curves for severe dengue disease or any dengue case by pre-existing DENV-Ab titer for the Pediatric Dengue Cohort. Cox proportional hazard models were fit without (A) or with (B) control for number of previous infections. Models were also adjusted for sex, epidemic season, and age.

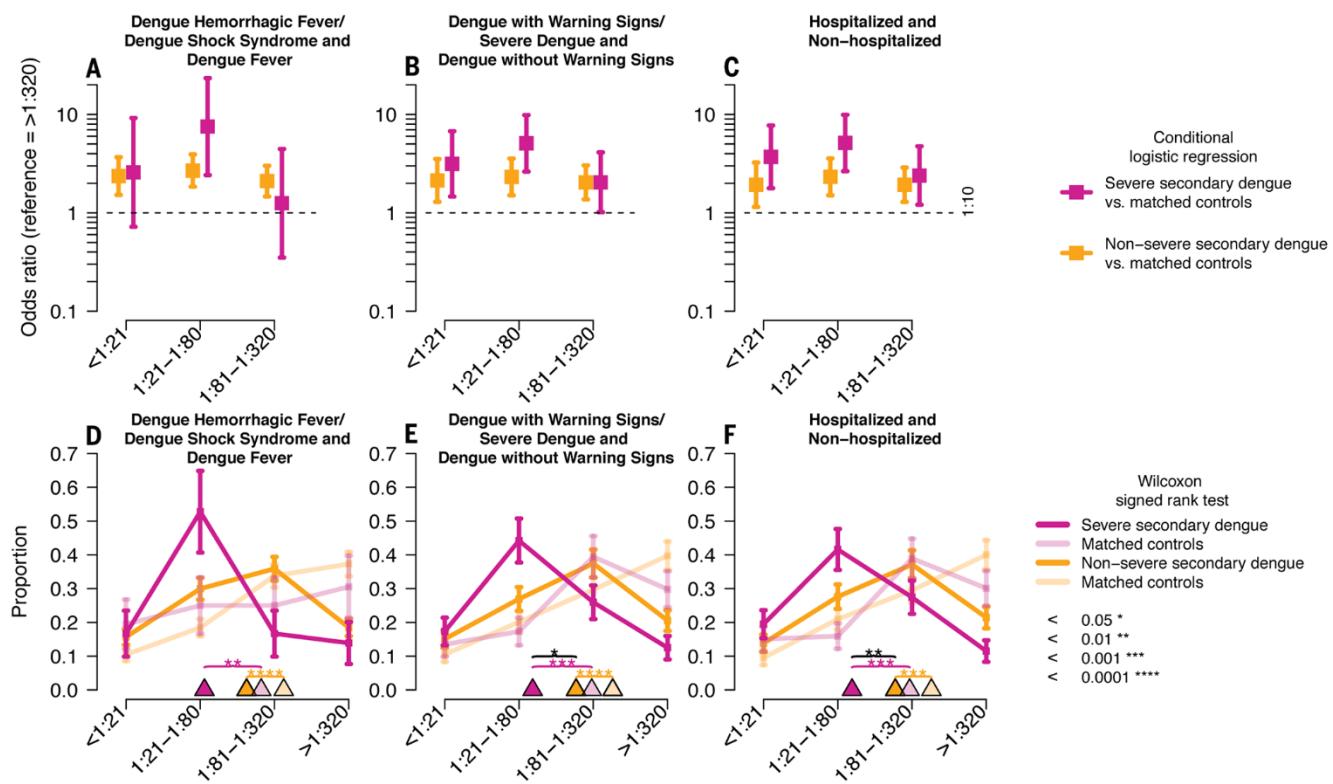


Fig. 3. Pre-existing DENV-Ab titers in severe or non-severe secondary dengue cases compared to matched controls drawn randomly from the Pediatric Dengue Cohort. (A to C) Five controls were matched to each case, and were of the same sex and age, had evidence of prior DENV infection, provided a blood sample within 1–2 months of the case’s pre-infection sample, but did not have a dengue case that year. Conditional logistic regression was used to compare pre-existing DENV-Ab titers of severe cases and non-severe cases each to matched controls, with titers $>1:320$ as reference. (D to F) Distributions of pre-existing DENV-Ab titers for severe and non-severe secondary dengue cases and matched controls (one control for each case). Error bars show one standard deviation, triangles show distribution medians, and brackets indicate significant differences in medians (severe and non-severe cases compared with Wilcoxon rank sum test, black bracket).

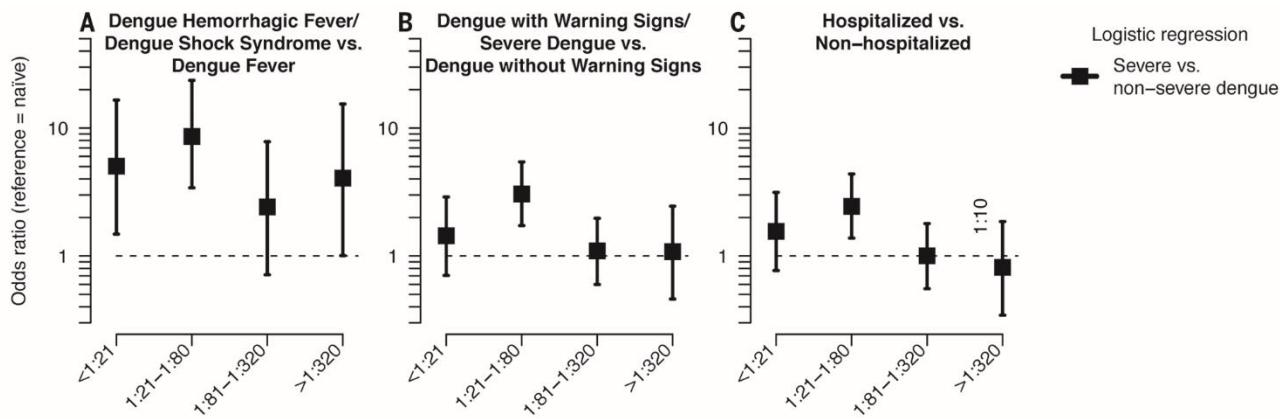


Fig. 4. Odds ratios for severe as compared to non-severe dengue by pre-infection DENV-Ab titer. (A to C) Logistic regression models were adjusted for sex, epidemic season, infecting DENV type, age, and number of previous DENV infections. DENV-naïve children were used as the reference group.

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