# What is single-cell RNA-Seq, and why is it useful?

SINGLE-CELL RNA-SEQ WITH BIOCONDUCTOR IN R

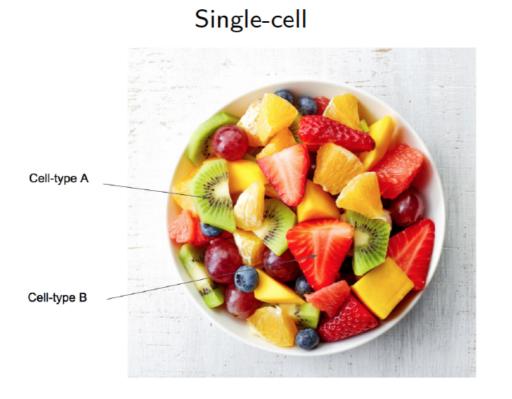


Fanny Perraudeau Senior Data Scientist, Whole Biome



#### Milkshake or fruit salad?.

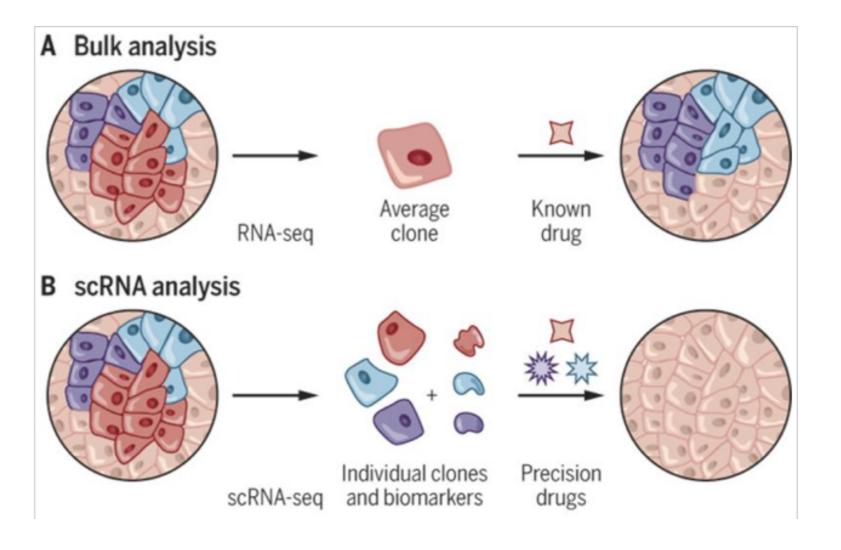
scRNA-seq is capturing gene expression at the cellular level



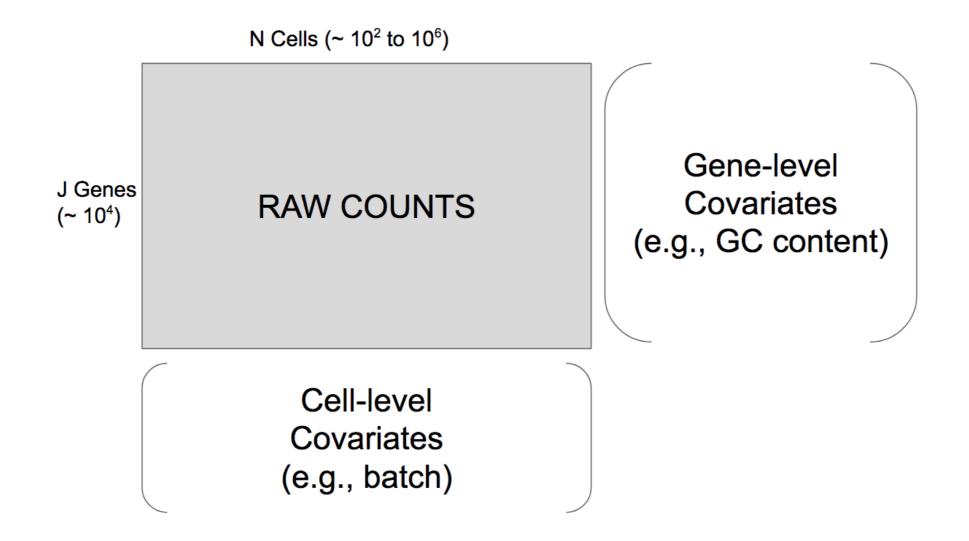


Shalek and Regev (2016)

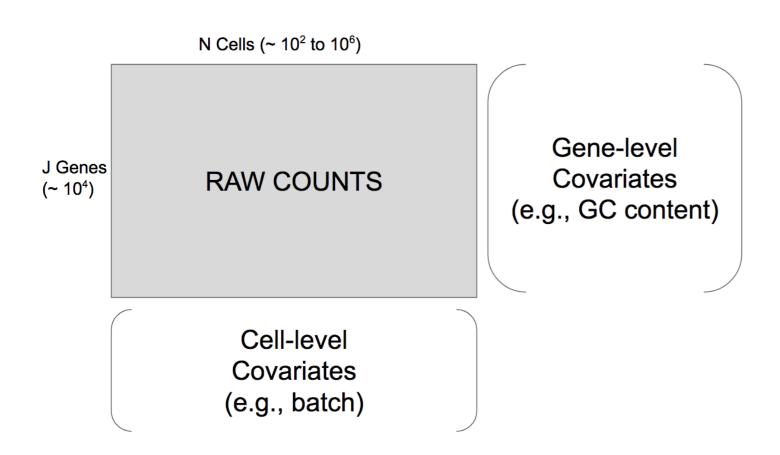
# scRNA-Seq could revolutionize personalized medicine in cancer



#### Data structure



### Zero inflation in single-cell transcriptome sequencing



- Biological zeros (e.g., cell cycle genes).
- Technical (false) zeros: dropouts.

# Let's practice!

SINGLE-CELL RNA-SEQ WITH BIOCONDUCTOR IN R



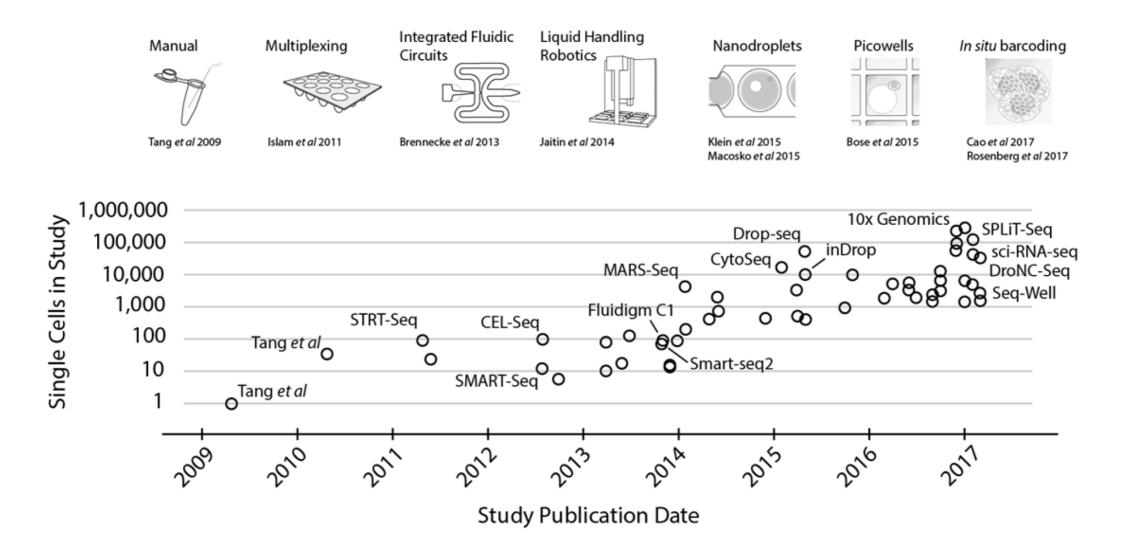
SINGLE-CELL RNA-SEQ WITH BIOCONDUCTOR IN R



Fanny Perraudeau Senior Data Scientist, Whole Biome



#### Exponential scaling of scRNA-Seq in the last decade



<sup>&</sup>lt;sup>1</sup> "Exponential scaling of single cell RNAseq in the last decade". Valentine Svensson, Roser Vento <sup>2</sup> Tormo, Sarah A Teichmann



#### Aspects of scRNE-Seq methods

- Quantification: determines types of analyses
  - Full-length protocols -- uniform coverage of RNA seq
  - Tag-based protocols -- one of the ends of each RNA
- Capture: determines throughput
  - microwell-based
  - microfluidic-based
  - droplet-based

<sup>&</sup>lt;sup>1</sup> https://hemberg <sup>2</sup> lab.github.io/scRNA.seq.course/introduction <sup>3</sup> to <sup>4</sup> single <sup>5</sup> cell <sup>6</sup> rna <sup>7</sup> seq.html

#### scRNE-seq workflow

- 1. Quality control
- 2. Normalization
- 3. Dimensionality reduction
- 4. Clustering
- 5. Differential expression analysis



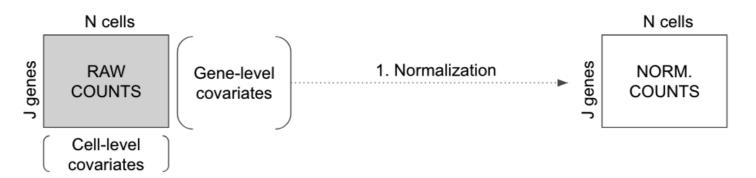
#### First step: quality control

Filter out low-quality cells:

- by library size: total number of reads aligned to each cell (a library refers to a cell)
- by cell coverage: average number of expressed genes in each cell

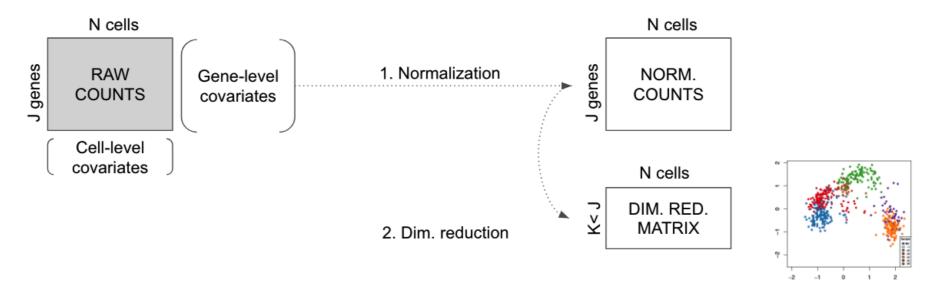
<sup>1</sup> "A step <sup>2</sup> by <sup>3</sup> step workflow for low <sup>4</sup> level analysis of single <sup>5</sup> cell RNA <sup>6</sup> seq data". Lun ATL, McCarthy DJ and Marioni JC





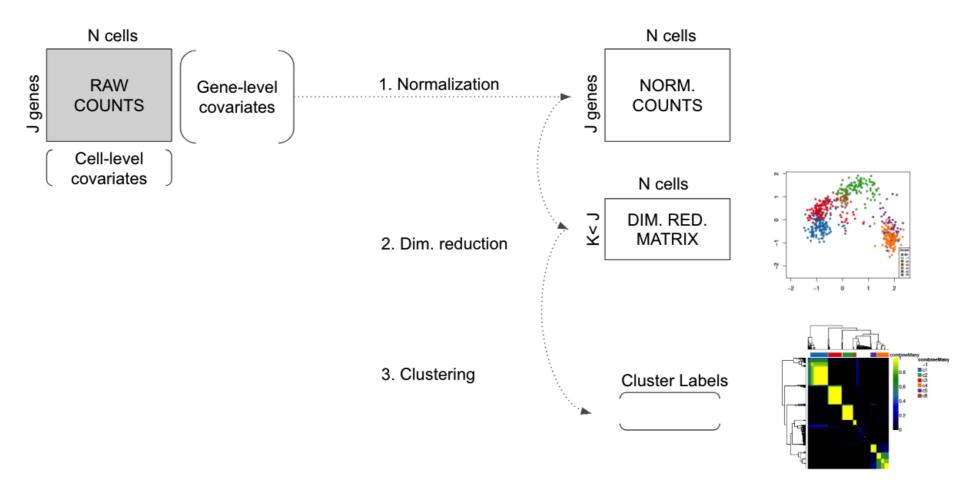
<sup>&</sup>lt;sup>1</sup> "Bioconductor workflow for single <sup>2</sup> cell RNA sequencing". Perraudeau F, Risso D, Street K et al





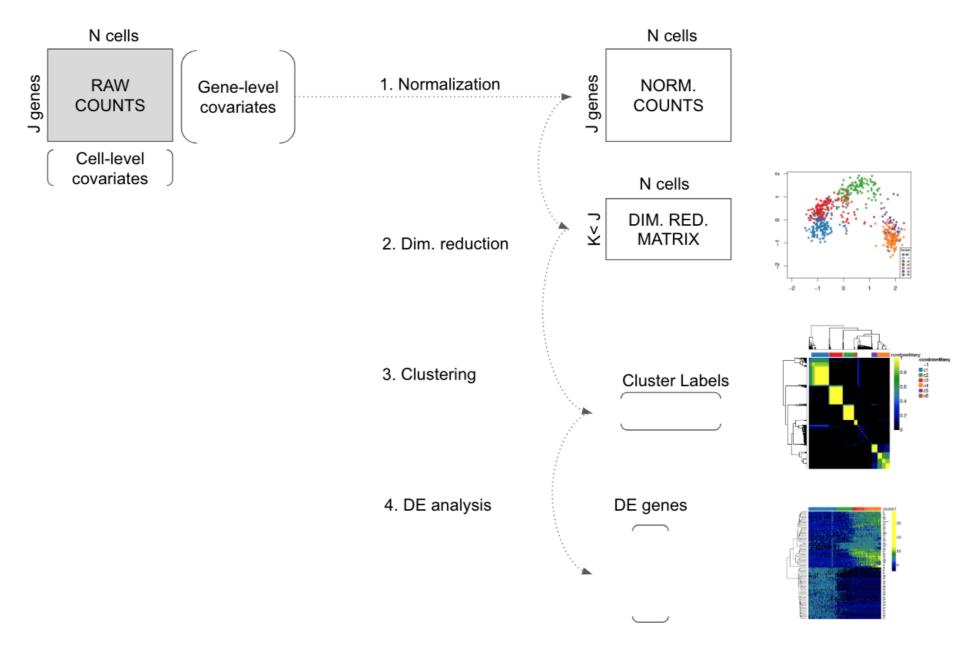
<sup>&</sup>lt;sup>1</sup> "Bioconductor workflow for single <sup>2</sup> cell RNA sequencing". Perraudeau F, Risso D, Street K et al





<sup>&</sup>lt;sup>1</sup> "Bioconductor workflow for single <sup>2</sup> cell RNA sequencing". Perraudeau F, Risso D, Street K et al





<sup>&</sup>lt;sup>1</sup> "Bioconductor workflow for single <sup>2</sup> cell RNA sequencing". Perraudeau F, Risso D, Street K et al



# Let's practice!

SINGLE-CELL RNA-SEQ WITH BIOCONDUCTOR IN R



# Load, create, and access single-cell datasets in R

SINGLE-CELL RNA-SEQ WITH BIOCONDUCTOR IN R



Fanny Perraudeau Senior Data Scientist, Whole Biome



#### SingleCellExperiment class

SingleCellExperiment (SCE) is a S4 class for storing data from single-cell experiments.

#### Can store and retrieve:

- matrix of counts
- cell and gene information
  - o spike-in information,
  - dimensionality reduction coordinates,
  - size factors for each cell,
  - usual metadata for genes and cells.

#### in a single R object!

<sup>1</sup> https://bioconductor.org/packages/3.9/bioc/html/SingleCellExperiment.html (by Aaron Lun and Davide Risso)



#### Load and install

Install SingleCellExperiment package

```
source("https://bioconductor.org/biocLite.R")
biocLite("SingleCellExperiment")
```

Load SingleCellExperiment package

library(SingleCellExperiment)

#### SCE object from a counts matrix

```
# create a counts matrix from Poisson distribution
counts <- matrix(rpois(8, lambda = 10), ncol = 2, nrow = 4)
# assign row and column names of counts matrix
rownames(counts) <- c("Lamp5", "Fam19a1", "Cnr1", "Rorb") #genes
colnames(counts) <- c("SRR2140028", "SRR2140022") #cells
# print the counts matrix
counts</pre>
```

	SRR2140028	SRR2140022
Lamp5	13	3
Fam19a1	9	10
Cnr1	8	10
Rorb	5	7

```
class: SingleCellExperiment
dim: 4 2
metadata(0):
assays(1): counts
rownames(4): Lamp5 Fam19a1 Cnr1 Rorb
rowData names(1): gene
colnames(2): SRR2140028 SRR2140022
colData names(1): cell
reducedDimNames(0):
spikeNames(0):
```



### SCE object from SummarizedExperiment

```
# create a SummarizedExperiment object from the counts matrix
se <- SummarizedExperiment(assays = list(counts = counts))
# convert to SingleCellExperiment
sce <- as(se, "SingleCellExperiment")
sce</pre>
```

```
class: SingleCellExperiment
dim: 4 2
metadata(0):
assays(1): counts
rownames(4): Lamp5 Fam19a1 Cnr1 Rorb
rowData names(0):
colnames(2): SRR2140028 SRR2140022
colData names(0):
reducedDimNames(0):
spikeNames(0):
```

<sup>&</sup>lt;sup>1</sup> SummarizedExperiment package: https://bioconductor.org/packages/3.9/bioc/html/SummarizedExperiment.html



#### Real single-cell dataset

```
# load the allen dataset from scRNAseq
library(scRNAseq)
data(allen)
# print allen
allen
```

```
class: SummarizedExperiment
dim: 20908 379
metadata(2): SuppInfo which_qc
assays(4): tophat_counts cufflinks_fpkm rsem_counts rsem_tpm
rownames(20908): 0610007P14Rik 0610009B22Rik ... Zzef1 Zzz3
rowData names(0):
colnames(379): SRR2140028 SRR2140022 ... SRR2139341 SRR2139336
colData names(22): NREADS NALIGNED ... Animal.ID passes_qc_checks_s
```

<sup>&</sup>lt;sup>1</sup> Tasic et al "Adult mouse cortical cell taxonomy revealed by single cell transcriptomics"



```
# covert to a SingleCellExperiment
sce <- as(allen, "SingleCellExperiment")

#print the sce object
sce</pre>
```

```
class: SingleCellExperiment
dim: 20908 379
metadata(2): SuppInfo which_qc
assays(4): tophat_counts cufflinks_fpkm
  rsem_counts rsem_tpm
rownames(20908): 0610007P14Rik 0610009B22Rik
  ... Zzef1 Zzz3
rowData names(0):
colnames(379): SRR2140028 SRR2140022 ...
  SRR2139341 SRR2139336
colData names(22): NREADS NALIGNED ...
 Animal.ID passes_qc_checks_s
reducedDimNames(0):
spikeNames(0):
```



# Let's practice!

SINGLE-CELL RNA-SEQ WITH BIOCONDUCTOR IN R

