

Report

Signal detection of spontaneous medical device reports over time accounting for multiple comparisons

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Table of contents

| | | |
|----------|--|-----------|
| 1 | Set up | 2 |
| 1.1 | Packages | 2 |
| 1.2 | Load data | 2 |
| 2 | Methods | 3 |
| 2.1 | Data aquisition | 3 |
| 3 | The signal detection statistics over time | 4 |
| 3.1 | Data | 4 |
| 3.2 | Reporting odds ratio (ROR) | 5 |
| 3.3 | BCPNN IC | 5 |
| 3.4 | maxSPRT | 5 |
| 4 | Analysis choices: | 6 |
| 5 | Plots | 7 |
| 6 | Session information | 12 |

1 Set up

1.1 Packages

```
suppressPackageStartupMessages({  
  library("readr")  
  library("dplyr")  
  library("tidyr")  
  library("forcats")  
  library("lubridate") # way to handle dates better than default R way  
  library("ggplot2")  
  library("ggrepel")  
  library("knitr")  
  library("gsDesign")  
  library("arrow")  
})
```

Warning: package 'dplyr' was built under R version 4.2.3

1.2 Load data

```
sra_cum_bcpnn <- read_parquet("out/sra_cum_bcpnn.parquet")  
  
bcpnn_signif <-  
  sra_cum_bcpnn %>%  
  group_by(grps, dat_type, thresh) %>%  
  arrange(dte) %>%  
  dplyr::filter(reach_sig) %>%  
  dplyr::filter(row_number() == 1) %>%  
  ungroup()
```

2 Methods

2.1 Data acquisition

The data is thanks to [curtis-murray](#) at his [MedicalDevicesNLP](#) repo

- Natural language processing of the TGA spontaneous reports of medical device database (DAEN)
- Each record has an estimate of $P(\text{topic} == \text{"pain"} \mid \text{Level}, \text{Doc})$ using hierarchical stochastic block modelling (hSBM)
- $P(\text{topic} == \text{"pain"} \mid \text{Level}, \text{Doc})$ estimates for each record are roughly interpreted as the proportion of the NLP analysed free text that is considered as using/describing words related to pain

And example record and processing values:

- [to include here]

3 The signal detection statistics over time

We will consider the three signal detection statistics below:

- Reporting Odds Ratio (ROR),
- Bayesian Confidence Propagation Neural Network Information Component (BCPNN IC with MCMC CIs), and
- the maxSPRT statistic

However, first let's outline the data available.

3.1 Data

Signal detection of disproportionate adverse events (AEs) will often have tabulated count data accumulated over time. The data at time point t can be summarised as below:

| | AE(s) X | AE(s) \bar{X} |
|---------------------|-----------|-----------------|
| Target exposure | a_t | b_t |
| Comparator exposure | c_t | d_t |

where

- AE(s) X is the set of AEs (or singular AE) of interest,
- AE(s) \bar{X} is the complementary set to the AEs of interest,
- *Target exposure* is the medical device(s) of interest,
- *Comparator exposure* is the medical devices to which the *Target exposure* is being compared, and
- a_t , b_t , c_t and d_t (all $\in \mathbb{Z}^+$) are the respective counts of AEs recorded up until (i.e., cumulative) time t .

In the motivating example of the pelvic mesh device, the contingency table can be written more specifically as

| | AEs pain | AEs not pain |
|---------------------|----------|--------------|
| Pelvic mesh | a_t | b_t |
| Comparator exposure | c_t | d_t |

where

- *AEs pain* is the count of AEs that contain “pain” themes greater or equal to some pre-specified threshold $p_t \in (0,1)$ as estimated by the hSBM (that is, $P(\text{topic} == \text{"pain"} | \text{Level}, \text{Doc}) \geq p_t$), and
- *Comparator exposure* can be any relevant set of medical devices to compare the pelvic mesh to (e.g., hernia mesh or all other mesh devices or all other devices).

3.2 Reporting odds ratio (ROR)

ROR with $100(1 - \alpha/2)\%$ confidence intervals

$$\hat{\text{ROR}} = \frac{\frac{a_t}{a_t + b_t}}{\frac{c_t}{c_t + d_t}}$$

3.3 BCPNN IC

BCPNN IC using the *maximum a posteriori* (m.a.p.) central estimate of the IC with MCMC simulation of the exact empirical distribution for $100(1 - \alpha/2)\%$ confidence (credible) regions of [Noren \(2006\)](#)

3.4 maxSPRT

The maximised log-likelihood ratio statistic

4 Analysis choices:

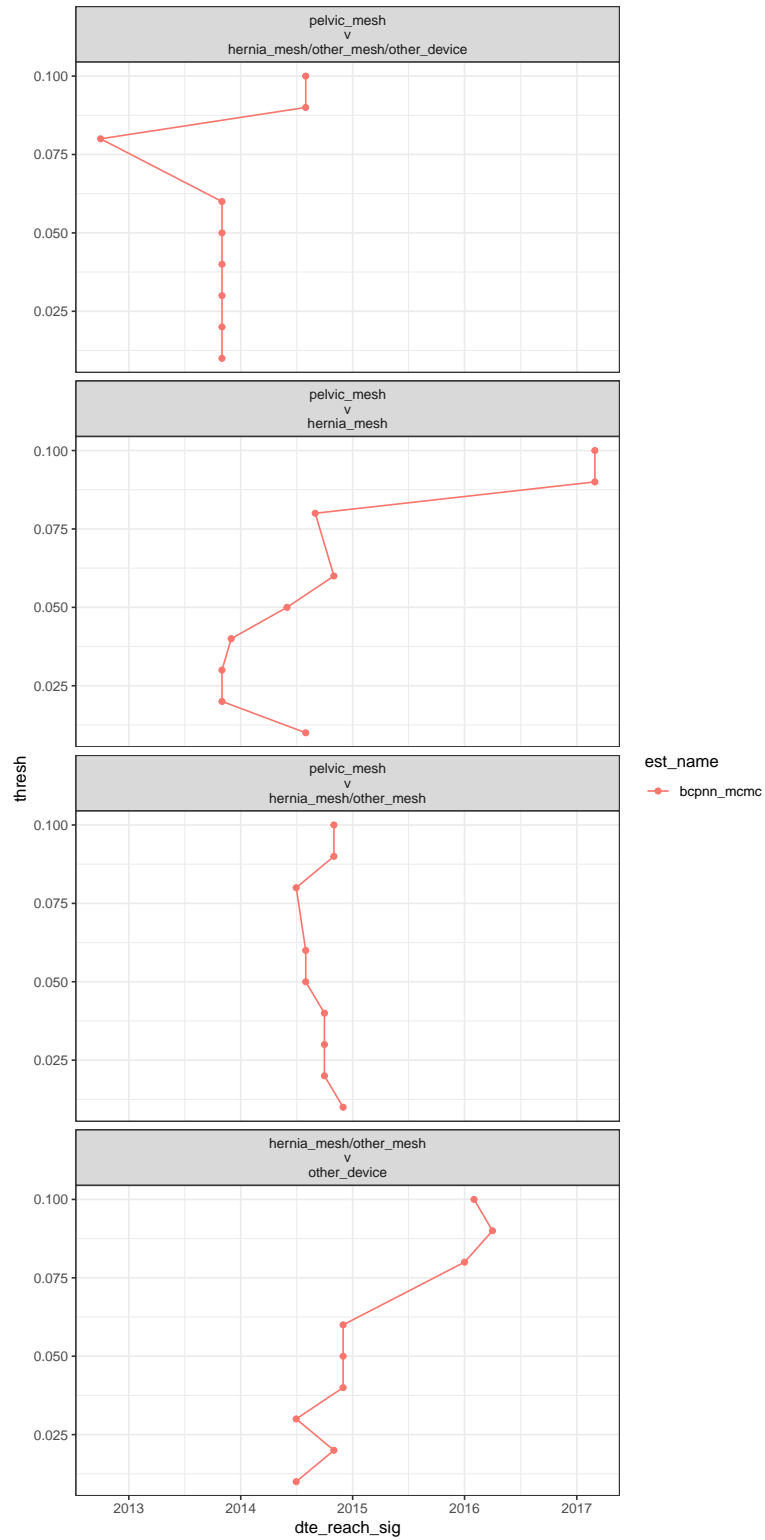
- Data structures - cumulative vs snapshot
- Threshold choose
- How many “looks”
- how to choose alpha spending
- sample size limitations for maxsprt - not an issue now can use MCMC method of `EmpiricalCalibration`

5 Plots

```
bcpnn_signif_plt <-  
  bcpnn_signif %>%  
  # keep only multiples of 0.01 (too many colours otherwise)  
  dplyr::filter(abs(100 * thresh - floor(100 * thresh)) < 1e-6) %>%  
  mutate(  
    grps = gsub(" v ", "\nv\n", grps),  
    grps = fct_inorder(grps)  
  )  
  
thresholds <- sort(unique(bcpnn_signif_plt[["thresh"]]))  
length(thresholds)
```

[1] 9

```
thresh_scale <- rev(hcl.colors(length(thresholds) + 1, "Inferno"))[-1]  
# thresh_scale <- rev(hcl.colors(length(thresholds), "SunsetDark"))  
  
bcpnn_signif_plt %>%  
  arrange(grps, thresh) %>%  
  ggplot(., aes(x = dte_reach_sig, y = thresh, col = est_name)) +  
  # ggplot(., aes(x = dte_reach_sig, y = est_name, col = factor(thresh))) +  
  geom_point() +  
  geom_path(aes(group = est_name)) +  
  # scale_colour_viridis_c(option = "B", direction = -1) +  
  # scale_colour_manual(values = thresh_scale) +  
  facet_wrap(~ grps, ncol = 1) +  
  theme_bw()
```




```

sra_cum_bcpnn_plt <-
  sra_cum_bcpnn %>%
    # keep only multiples of 0.01 (too many colours otherwise)
    dplyr::filter(abs(100 * thresh - floor(100 * thresh)) < 1e-6) %>%
    mutate(
      grps = gsub(" v ", "\nv\n", grps),
      grps = fct_inorder(grps)
    )

thresholds <- sort(unique(sra_cum_bcpnn_plt[["thresh"]]))
length(thresholds)

```

[1] 9

```

thresh_scale <- rev(hcl.colors(length(thresholds), "SunsetDark"))

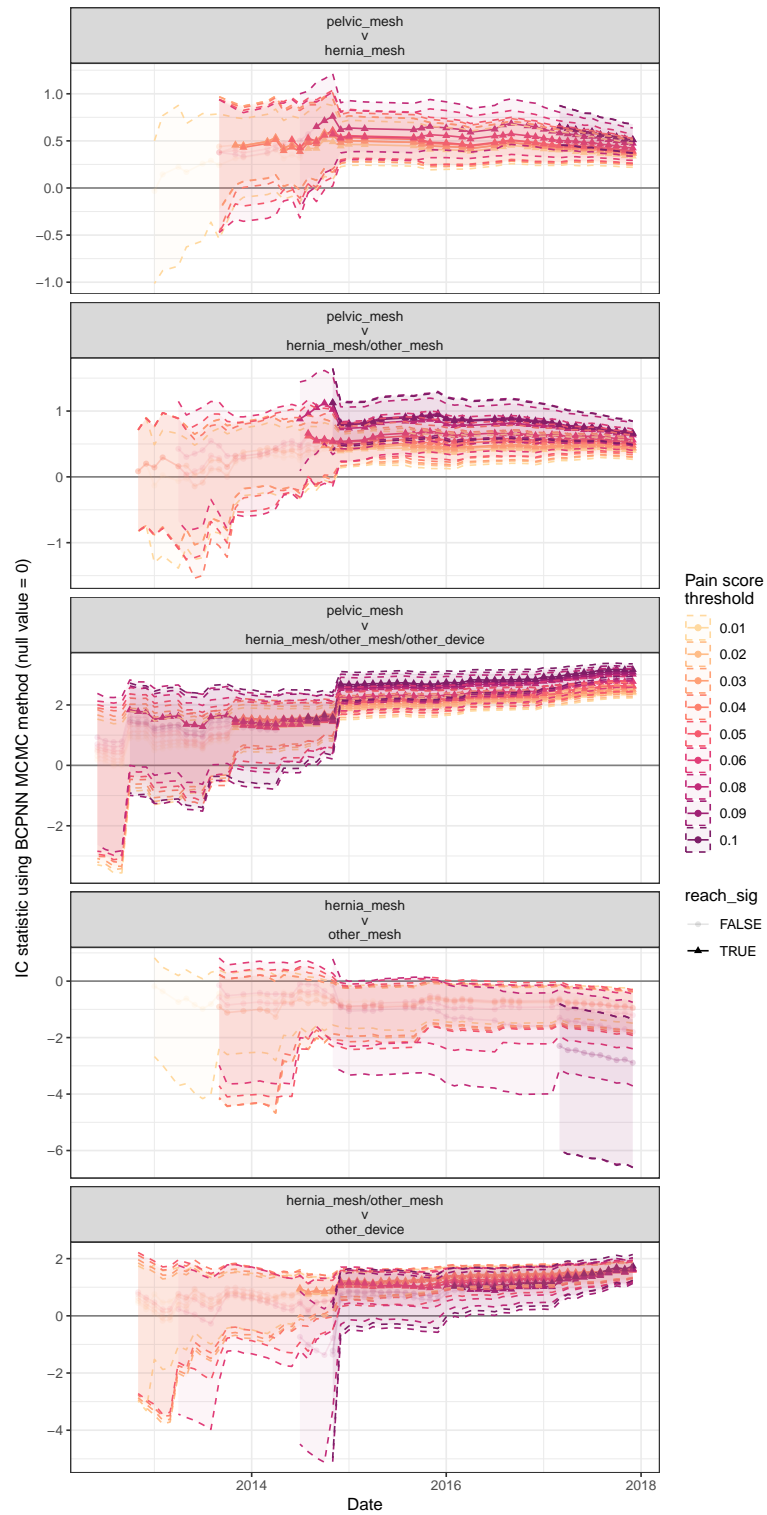
sra_cum_bcpnn_plt %>%
  ggplot(
    .,
    aes(
      dte,
      est,
      ymax = ci_hi,
      ymin = ci_lo,
      col = factor(thresh),
      fill = factor(thresh),
      group = factor(thresh),
      alpha = reach_sig,
      shape = reach_sig
    )
  ) %>%
  geom_hline(aes(yintercept = 0), col = "grey50") %>% # null value
  geom_line() %>%
  geom_point() %>%
  geom_ribbon(alpha = 0.05, lty = 2) %>%
  facet_wrap(~ grps, scales = "free_y", ncol = 1) %>%
  labs(
    subtitle = "(Calculations made on cumulative monthly data)",
    y = "IC statistic using BCPNN MCMC method (null value = 0)",
  )

```

```
x = "Date",
col = "Pain score\nthreshold",
fill = "Pain score\nthreshold"
) %>%
scale_colour_manual(values = thresh_scale, aesthetics = c("colour", "fill")) %>%
theme_bw()
```

Warning: Using alpha for a discrete variable is not advised.

(Calculations made on cumulative monthly data)



6 Session information

```
format(Sys.time(), '%d %b %Y')
```

```
[1] "17 Jul 2023"
```

```
Sys.info() %>% as.data.frame(.)
```

```
      .  
sysname      Windows  
release      10 x64  
version      build 19044  
nodename     DESKTOP-R5P5N23  
machine      x86-64  
login        ty  
user         ty  
effective_user ty
```

```
sessionInfo()
```

```
R version 4.2.2 (2022-10-31 ucrt)  
Platform: x86_64-w64-mingw32/x64 (64-bit)  
Running under: Windows 10 x64 (build 19044)
```

```
Matrix products: default
```

```
locale:
```

```
[1] LC_COLLATE=English_Australia.utf8  LC_CTYPE=English_Australia.utf8  
[3] LC_MONETARY=English_Australia.utf8 LC_NUMERIC=C  
[5] LC_TIME=English_Australia.utf8
```

```
attached base packages:
```

```
[1] stats      graphics  grDevices  utils      datasets  methods   base
```

```
other attached packages:
```

```
[1] arrow_11.0.0.2  gsDesign_3.4.0  knitr_1.42      ggrepel_0.9.3  
[5] ggplot2_3.4.1   lubridate_1.9.2 forcats_1.0.0   tidyr_1.3.0
```

[9] dplyr_1.1.2 readr_2.1.4

loaded via a namespace (and not attached):

| | | | |
|----------------------|------------------|----------------|------------------|
| [1] Rcpp_1.0.10 | pillar_1.9.0 | compiler_4.2.2 | tools_4.2.2 |
| [5] bit_4.0.5 | digest_0.6.31 | jsonlite_1.8.4 | evaluate_0.20 |
| [9] lifecycle_1.0.3 | tibble_3.2.1 | gtable_0.3.1 | timechange_0.2.0 |
| [13] pkgconfig_2.0.3 | rlang_1.1.1 | cli_3.6.0 | rstudioapi_0.14 |
| [17] yaml_2.3.7 | xfun_0.37 | fastmap_1.1.0 | withr_2.5.0 |
| [21] generics_0.1.3 | vctrs_0.6.3 | hms_1.1.2 | bit64_4.0.5 |
| [25] grid_4.2.2 | tidyselect_1.2.0 | glue_1.6.2 | R6_2.5.1 |
| [29] fansi_1.0.4 | rmarkdown_2.20 | farver_2.1.1 | tzdb_0.3.0 |
| [33] purrr_1.0.1 | magrittr_2.0.3 | scales_1.2.1 | ellipsis_0.3.2 |
| [37] htmltools_0.5.4 | assertthat_0.2.1 | xtable_1.8-4 | colorspace_2.1-0 |
| [41] labeling_0.4.2 | utf8_1.2.3 | munsell_0.5.0 | |