# Report

# Signal detection of spontaneous medical device reports over time accounting for multiple comparisons

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## 1 Set up

#### 1.1 Packages

```
suppressPackageStartupMessages({
    library("readr")
    library("dplyr")
    library("forcats")
    library("lubridate") # way to handle dates better than default R way
    library("stringr")
    library("ggplot2")
    library("ggthemes")
    library("ggrepel")
    library("grepel")
    library("gsDesign")
    library("arrow")
})
```

Warning: package 'dplyr' was built under R version 4.2.3

```
col_pal <- c("darkorange", "cyan4", "purple")</pre>
```

#### 1.2 Load data

```
sra_cum_bcpnn <- read_parquet("out/sra_cum_bcpnn.parquet")
sra_cum_bcpnn_mc_adj <- read_parquet("out/sra_cum_bcpnn_mc_adj.parquet")
sra_cum_maxsprt <- read_parquet("out/sra_cum_maxsprt.parquet")

sra <-
bind_rows(
    sra_cum_bcpnn %>% mutate(stat = "BCPNN"),
    sra_cum_bcpnn_mc_adj %>% mutate(stat = "BCPNN (MCadj)"),
    sra_cum_maxsprt %>% mutate(stat = "maxSPRT")
) %>%
select(stat, everything())
```

```
sra <-
    sra %>%
    mutate(
    test_stat = if_else(stat == "maxSPRT", maxllr, ci_lo),
    test_thresh = if_else(stat == "maxSPRT", cv, 0),
    rr_stat = if_else(stat == "maxSPRT", rre, 2 ^ est)
)

thresholds <- sort(unique(sra[["thresh"]]))
length(thresholds)</pre>
```

[1] 19

### 2 Methods

#### 2.1 Data aquisition

The data is thanks to curtis-murray at his MedicalDevicesNLP repo

- Natural language processing of the TGA spontaneous reports of medical device database (DAEN)
- Each record has an estimate of P(topic == "pain" | Level, Doc) using hierarchical stochastic block modelling (hSBM)
- P(topic == "pain" | Level, Doc) estimates for each record are roughly interpreted as the proportion of the NLP analysed free text that is considered as using/describing words related to pain

And example record and processing values:

• [to include here]

#### 2.2 Analysis data

Signal detection of disproportionate adverse events (AEs) will often have tabulated count data accumulated over time. The data at time point t can be summarised as below:

	AE(s) X	$AE(s) \bar{X}$
Target exposure	$a_t$	$b_t$
Comparator exposure	$c_t$	$d_t$

where

- AE(s) X is the set of AEs (or singular AE) of interest,
- AE(s) X is the complementary set to the AEs of interest,
- Target exposure is the medical device(s) of interest,
- Comparator exposure is the medical devices to which the Target exposure is being compared, and
- $a_t$ ,  $b_t$ ,  $c_t$  and  $d_t$  (all  $\in \mathbb{Z}^+$ ) are the respective counts of AEs recorded up until (i.e., cumulative) time t.

In the motivating example of the pelvic mesh device, the contingency table can be written more specifically as

	AEs pain	AEs not pain
Pelvic mesh	$a_t$	$b_t$
Comparator exposure	$c_t$	$d_t$

where

- $AEs\ pain$  is the count of AEs that contain "pain" themes greater or equal to some pre-specified threshold  $p_t\in(0,1)$  as estimated by the hSBM (that is,  $P(\texttt{topic} == "pain" | \texttt{Level}, \texttt{Doc}) \geq p_t)$ , and
- Comparator exposure can be any relevant set of medical devices to compare the pelvic mesh to (e.g., hernia mesh or all other mesh devices or all other devices).

## 3 The signal detection statistics over time

We will consider the three signal detection statistics below:

• Proportional reporting ratio (PRR),

- Bayesian Confidence Propagation Neural Network Information Component (BCPNN IC with MCMC CIs), and
- the maxSPRT statistic

As signal detection is being undertaken repeatedly as data are being accumulated, alpha spending needs to be considered. The below table classifies the aforementioned signal detection methods by their null hypothesis as well as whether they control for the family-wise error rate (FWER)

non-FWER version	FWER version
PRR	binary, group sequential maxSPRT
IC	IC with $\alpha$ -spending scheme
	PRR

We will demonstrate how the group sequential binary maxSPRT, as described in previous work, is equivalent to a FWER-controlled PRR method of signal detection.

#### 3.1 Proportional reporting ratio (PRR)

The PRR estimate is calculated

$$\hat{PRR} = \frac{\frac{a_t}{a_t + b_t}}{\frac{c_t}{c_t + d_t}}$$

with approximate  $100(1-\alpha/2)\%$  confidence intervals

$$\left( \hat{\text{PRR}} \times e^{-Z_{\alpha/2}^* \sqrt{\frac{1}{a_t} + \frac{1}{a_t + b_t} + \frac{1}{c_t} + \frac{1}{c_t + d_t}}}, \ \ \hat{\text{PRR}} \times e^{+Z_{\alpha/2}^* \sqrt{\frac{1}{a_t} + \frac{1}{a_t + b_t} + \frac{1}{c_t} + \frac{1}{c_t + d_t}}} \right)$$

where  $Z_{\alpha/2}^*$  is the  $(1-\alpha/2)^{\rm th}$  quantile of the standard normal distribution.

#### 3.2 BCPNN IC

BCPNN IC using the maximum a posteriori (m.a.p.) central estimate of the IC with MCMC simulation of the exact empirical distribution for  $100(1-\alpha/2)\%$  confidence (credible) regions of Noren (2006)

#### 3.3 maxSPRT

Kulldorff et al. (2011) outlined that the relative risk (RR) at a given point-in-time for accumulated binary data (that is, "success"/"failure" events or AE of interest or not) of a target group relative to a comparator has the maximum likelihood estimate of

$$\hat{RR} = z \frac{C_n}{n - C_n}$$

where

- z is the ratio of the total AEs for the comparator to the total AEs for the target,
- $C_n$  is the count of target exposure AEs in X,
- n is the count of all AEs in X (target and comparator exposure), and
- $n-C_n$  is therefore the count of comparator exposure AEs in X.

In the context of our data, the values z,  $C_n$  and n are the quantilties  $\frac{c_t + d_t}{a_t + b_t}$ ,  $a_t$  and  $a_t + c_t$ , respectively, at time t.

Therefore the RR maximum likelihood estimate at time t can be re-written

$$\hat{RR}_t = \frac{c_t + d_t}{a_t + b_t} \times \frac{a_t}{c_t}$$

$$= \frac{\frac{1}{a_t + b_t}}{\frac{1}{c_t + d_t}} \times \frac{a_t}{c_t}$$

$$= \frac{\frac{a_t}{a_t + b_t}}{\frac{c_t}{c_t + d_t}}$$

which is the PRR estimate as before.

The (maximised) log-likelihood ratio statistic of  $\hat{RR}_t$  can be determined calculated as

$$\mathrm{LLR}_t = a_t \ln \left( \frac{a_t}{a_t + c_t} \right) + c_t \ln \left( \frac{c_t}{a_t + c_t} \right) - a_t \ln \left( \frac{a_t + b_t}{a_t + b_t + c_t + d_t} \right) - c_t \ln \left( \frac{c_t + d_t}{a_t + b_t + c_t + d_t} \right)$$

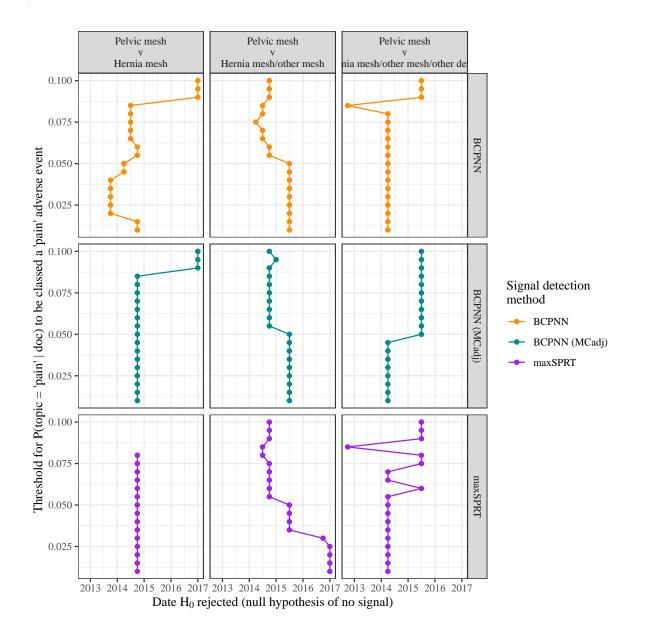
## 4 Analysis choices:

- Data structures cumulative vs snapshot
- Threshold choose
- How many "looks"
- how to choose alpha spending
- $\bullet$  sample size limitations for maxsprt not an issue now can use MCMC method of EmpiricalCalibration

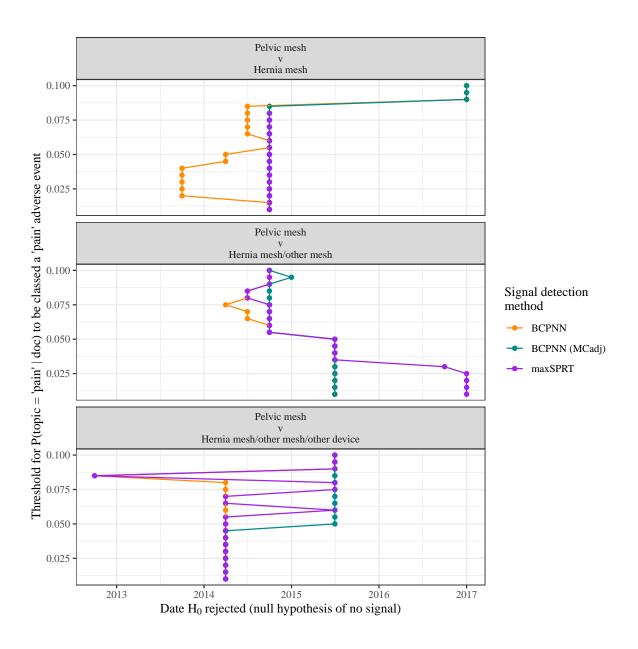
#### 5 Plots

```
date_signif_dat <-
    sra %>%
    group_by(stat, grps, dat_type, thresh) %>%
    arrange(dte) %>%
    dplyr::filter(reach_sig) %>%
    dplyr::filter(row number() == 1) %>%
    ungroup() %>%
    arrange(stat, grps, dat_type, thresh)
  signif plt <-
    date_signif_dat %>%
    ### only keep pelvic mesh as target vs whatever comparator
    dplyr::filter(grepl("^.*pelvic.* v ", grps)) %>%
    mutate(
      grps = gsub("\([a-z]\)", "", grps),
      grps = gsub("_", " ", grps),
      grps = gsub("pelvic mesh", "Pelvic mesh", grps),
      grps = gsub("hernia mesh", "Hernia mesh", grps),
      # grps = str_to_sentence(grps),
      grps = gsub(" v ", "\nv\n", grps, fixed = TRUE),
      grps = fct_inorder(grps)
  levels(signif_plt$grps)
[1] "Pelvic mesh\nv\nHernia mesh"
[2] "Pelvic mesh\nv\nHernia mesh/other mesh"
[3] "Pelvic mesh\nv\nHernia mesh/other mesh/other device"
  signif_plt %>%
    arrange(grps, thresh) %>%
    ggplot(., aes(x = dte_reach_sig, y = as.numeric(thresh), col = stat)) +
    geom_point() +
    geom_path(aes(group = stat)) +
    scale_colour_manual(values = col_pal) +
    # scale_colour_tableau(palette = "Color Blind", direction = -1) +
    facet_grid(stat ~ grps) +
```

```
# facet_wrap( ~ grps, ncol = 1) +
theme_bw() +
theme(text = element_text(family = "serif")) +
labs(
    x = expression("Date" ~ H[0] ~ "rejected (null hypothesis of no signal)"),
    y = "Threshold for P(topic = 'pain' | doc) to be classed a 'pain' adverse event",
    col = "Signal detection\nmethod"
)
```



```
ggsave(
 filename = "fig/time_to_signal_method_facets.png",
 dpi = 900, width = 9, height = 9
signif_plt %>%
 arrange(grps, thresh) %>%
 ggplot(., aes(x = dte_reach_sig, y = as.numeric(thresh), col = stat)) +
  geom_point() +
  geom_path(aes(group = stat)) +
 scale_colour_manual(values = col_pal) +
  # scale_colour_tableau(palette = "Color Blind", direction = -1) +
  # facet_grid(stat ~ grps) +
 facet_wrap( ~ grps, ncol = 1) +
 theme_bw() +
 theme(text = element_text(family = "serif")) +
 labs(
    x = expression("Date" ~ H[0] ~ "rejected (null hypothesis of no signal)"),
   y = "Threshold for P(topic = 'pain' | doc) to be classed a 'pain' adverse event",
   col = "Signal detection\nmethod"
  )
```



```
ggsave(
  filename = "fig/time_to_signal_method_overlay.png",
  dpi = 900, width = 5, height = 9
)
```

```
sra_stat_plt <-
    sra %>%

# keep only subset of thresholds (too many colours otherwise)
dplyr::filter(thresh %in% sprintf("%0.3f", seq(0.02, 0.08, by = 0.02))) %>%
### only keep pelvic mesh as target vs whatever comparator
# dplyr::filter(grepl("^.*pelvic.* v ", grps)) %>%
dplyr::filter(grepl("^\\(a\\))", grps)) %>%
mutate(
    grps = gsub(" v ", "\nv\n", grps),
    grps = gsub("\\([a-z]\\) ", "", grps),
    grps = gsub("_", " ", grps),
    grps = fct_inorder(grps)
)

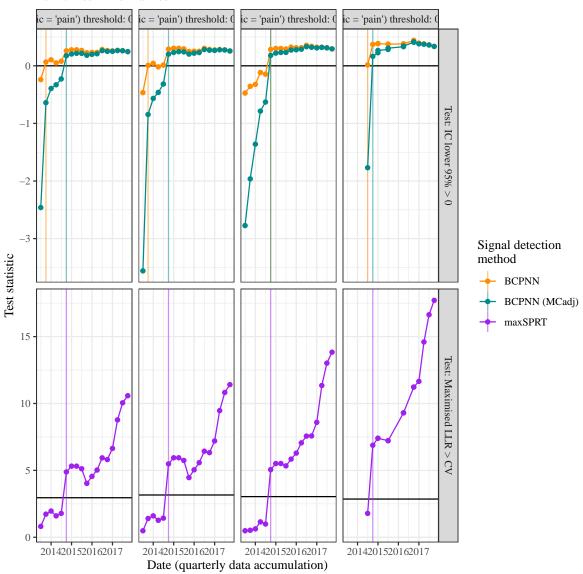
thresholds <- sort(unique(sra_stat_plt[["thresh"]]))
length(thresholds)</pre>
```

#### [1] 4

```
thresh scale <- rev(hcl.colors(length(thresholds), "SunsetDark"))
# thresh scale <- rev(hcl.colors(length(thresholds) + 1, "Inferno"))[-1]
sra_stat_plt %>%
 mutate(
    reach_sig_alpha = ifelse(reach_sig, 1, 0.8),
    'P(topic = 'pain') threshold' = thresh,
    `Test` = if_else(stat == "maxSPRT", "Maximised LLR > CV", "IC lower 95% > 0")
  ) %>%
  ggplot(
    ٠,
   aes(
     x = dte
     y = test_stat,
     col = stat,
      group = interaction(stat, thresh)
      # alpha = reach_sig_alpha
  ) %+%
  geom_hline(aes(yintercept = test_thresh), col = "black") %+% # null value
  geom_vline(aes(xintercept = dte_reach_sig, col = stat), alpha = 0.5) %+% # sig first red
```

```
geom_line() %+%
geom_point() %+%
# facet_wrap(~ grps, scales = "free_y", ncol = 1) %+%
facet_grid(
 `Test` ~ `P(topic = 'pain') threshold`,
 scales = "free_y",
 labeller = label_both
) %+%
labs(
  subtitle = "Pelvic mesh v hernia mesh",
 y = "Test statistic",
 x = "Date (quarterly data accumulation)",
 col = "Signal detection\nmethod"
) %+%
# scale_y_continuous(limits = c(NA, 6)) %+%
scale_colour_manual(values = col_pal) %+%
# scale_colour_tableau(palette = "Color Blind", direction = -1) +
theme_bw() %+%
theme(text = element_text(family = "serif"))
```

#### Pelvic mesh v hernia mesh

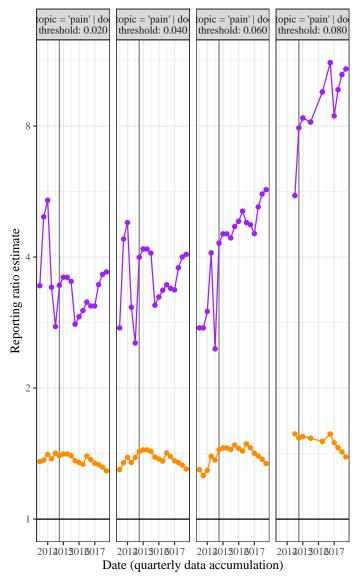


```
ggsave(
  filename = "fig/pelvic_v_hernia_sig_detect_over_time.png",
  dpi = 900, width = 10, height = 8
)
thresh_lablr <- function(string) paste0("P(topic = 'pain' | doc)\nthreshold: ", string)</pre>
```

```
sra_stat_plt %>%
 dplyr::filter(stat != "BCPNN") %>%
 mutate(
   stat2 =
     paste0(
       ifelse(stat == "maxSPRT", "RR", "2^IC =\nP(Pain AE & Pelvic)/{P(Pain AE)P(Pelvic)}
       "\n[", stat, "]"
     ),
   reach_sig_alpha = ifelse(reach_sig, 1, 0.8),
   `P(topic = 'pain') threshold` = thresh,
   `Test` = if_else(stat == "maxSPRT", "Maximised LLR > CV", "IC lower 95% > 0"),
   `Statistic calculation method` = stat
 ) %>%
 ggplot(
   ٠,
   aes(
     x = dte
     y = rr_stat,
     col = stat2,
     group = stat2
     # group = interaction(stat2, thresh)
     # alpha = reach sig alpha
     # ymin = ci_lo,
     # ymax = ci_hi
   )
 ) %+%
 geom_hline(aes(yintercept = 1), col = "black") %+% # null value
 geom_vline(aes(xintercept = dte_reach_sig), alpha = 0.5) %+% # sig first reached
 geom_line() %+%
 geom_point() %+%
 # geom_ribbon(alpha = 0.05) %+%
 facet_wrap(~ thresh, nrow = 1, labeller = as_labeller(thresh_lablr)) %+%
 # facet_grid(
 # `Statistic calculation method` ~ `P(topic = 'pain') threshold`,
 # labeller = label_both
 # ) %+%
 labs(
   subtitle = "Pelvic mesh v hernia mesh",
   y = "Reporting ratio estimate",
   x = "Date (quarterly data accumulation)",
   col = "Reporting ratio estimate\n[signal detection method]"
```

```
) %+%
scale_y_continuous(trans = "log2") %+%
scale_colour_manual(values = col_pal[-2]) %+%
theme_bw() %+%
theme(
  text = element_text(family = "serif"),
  legend.key.height = unit(3, units = "line")
)
```

#### Pelvic mesh v hernia mesh



## Reporting ratio estimate [signal detection method]

- 2^IC =
  P(Pain AE & Pelvic)/{P(Pain AE)P(Pelvic)}
  [BCPNN (MCadj)]
- 22
- RR [maxSPRT]

```
ggsave(
  filename = "fig/pelvic_v_hernia_rr_est_over_time.png",
  dpi = 900, width = 12, height = 5
)
```

#### 6 Session information

```
# Sys.info()[!(names(Sys.info()) %in% c("login", "nodename"))] %>%
  # as.data.frame(.)
  format(Sys.time(), '%d %b %Y')
[1] "31 Aug 2023"
  sessionInfo()
R version 4.2.2 (2022-10-31 ucrt)
Platform: x86_64-w64-mingw32/x64 (64-bit)
Running under: Windows 10 x64 (build 19045)
Matrix products: default
locale:
[1] LC_COLLATE=English_Australia.utf8 LC_CTYPE=English_Australia.utf8
[3] LC_MONETARY=English_Australia.utf8 LC_NUMERIC=C
[5] LC_TIME=English_Australia.utf8
attached base packages:
[1] stats
              graphics grDevices utils
                                            datasets methods
                                                                 base
other attached packages:
 [1] arrow 11.0.0.2 gsDesign 3.4.0 knitr 1.42
                                                      ggrepel_0.9.3
 [5] ggthemes_4.2.4 ggplot2_3.4.1
                                     stringr_1.5.0
                                                      lubridate_1.9.2
                     tidyr 1.3.0
                                                      readr_2.1.4
 [9] forcats 1.0.0
                                     dplyr 1.1.2
loaded via a namespace (and not attached):
                                         compiler_4.2.2
 [1] Rcpp_1.0.10
                       pillar_1.9.0
                                                            tools_4.2.2
 [5] bit_4.0.5
                       digest_0.6.31
                                         jsonlite_1.8.4
                                                            evaluate_0.20
 [9] lifecycle_1.0.3
                       tibble_3.2.1
                                         gtable_0.3.1
                                                            timechange_0.2.0
[13] pkgconfig_2.0.3
                                                            rstudioapi_0.14
                       rlang_1.1.1
                                         cli_3.6.0
[17] yaml_2.3.7
                       xfun_0.37
                                         fastmap_1.1.0
                                                            withr_2.5.0
[21] systemfonts_1.0.4 generics_0.1.3
                                         vctrs_0.6.3
                                                            hms_1.1.2
[25] bit64_4.0.5
                       grid_4.2.2
                                         tidyselect_1.2.0
                                                            glue_1.6.2
[29] R6_2.5.1
                       textshaping_0.3.6 fansi_1.0.4
                                                            rmarkdown_2.20
[33] farver_2.1.1
                       tzdb_0.3.0
                                         purrr_1.0.1
                                                            magrittr_2.0.3
```

[37] scales_1.2.1	ellipsis_0.3.2	htmltools_0.5.4	assertthat_0.2.1
[41] xtable_1.8-4	colorspace_2.1-0	ragg_1.2.5	labeling_0.4.2
[45] utf8 1.2.3	stringi 1.7.12	munsell 0.5.0	