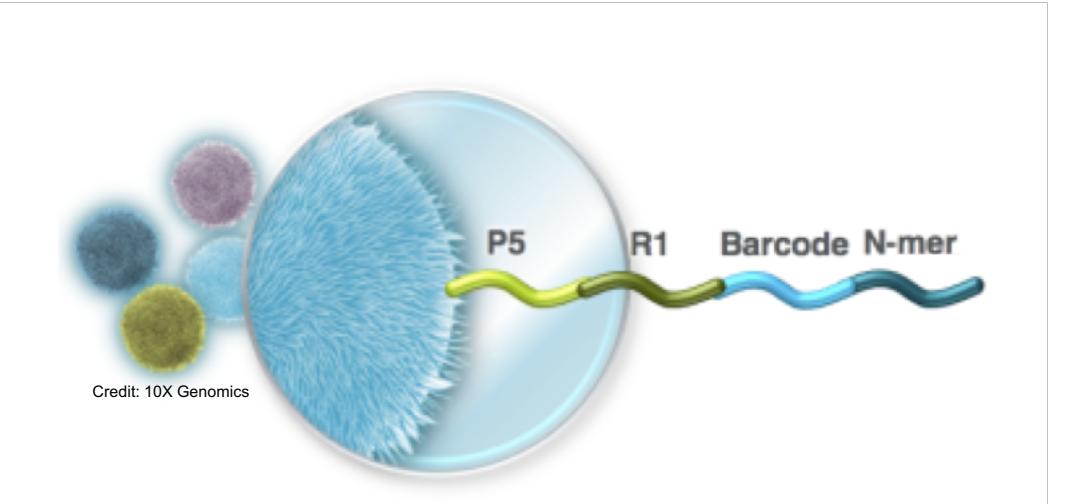


10X Genome Assembly Technology and Single Cell CNV



Diana Burkart-Waco
DNA Technologies and Expression Analysis
Cores

12-19-2018

10X Chromium Genome

linked read assembly

...providing *de novo* genome assembly, variant calling, and genome structure information...

- Upstream sample preparation
 - Sample QC guidelines
- 10X Chromium Genome
 - Technology
 - Applications
 - UC Davis projects
- NEW: Copy Number Variant kit

DNA Quality and Applications

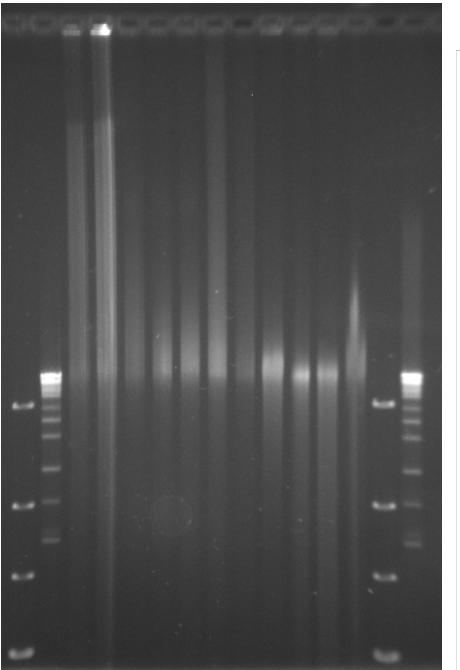
DNA Quality Level	DNA size (reported by 10x Chromium Genome pipeline)	Applications
5	>80 kb	De novo assembly with Supernova and Long Ranger analysis
4	60-80 kb	De novo assembly with Supernova and Long Ranger analysis
3	40-60 kb	Long Ranger analysis
2	20-40 kb	Long Ranger analysis [‡]
1	<20 kb	Long Ranger analysis possible, performance not thoroughly characterized [‡]

10X Technical note: "Single-stranded DNA Damage and its Effects on Chromium Genome Application Performance"

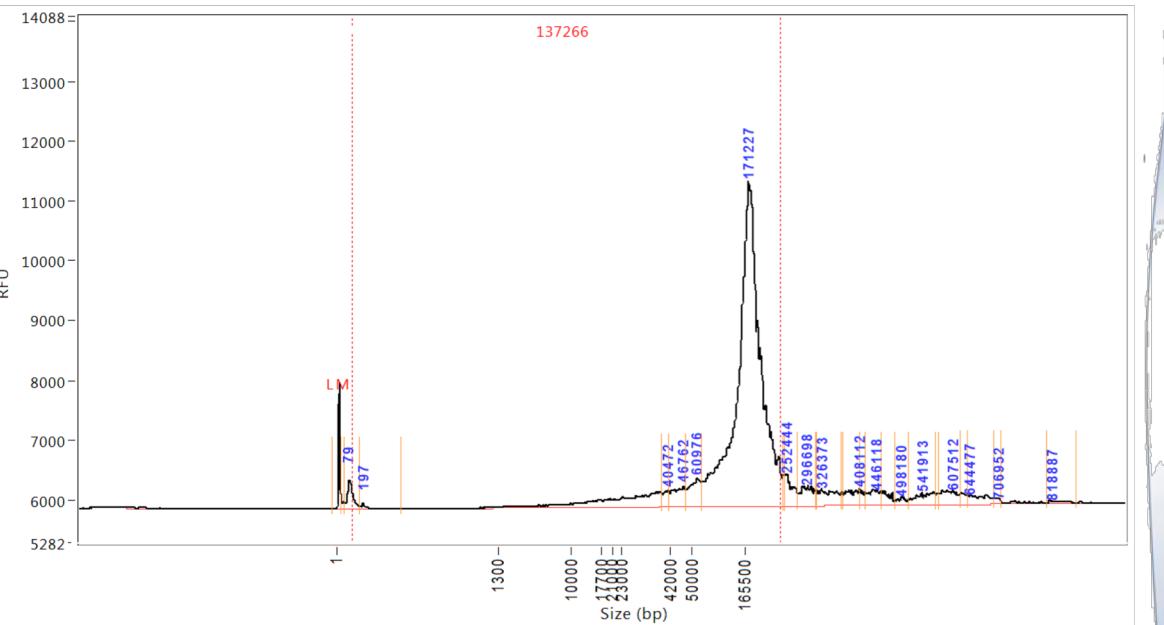
QC options

- Fragment analysis needed to determine size and degree of degradation.

➤ Pulsed-Field Gel Electrophoresis

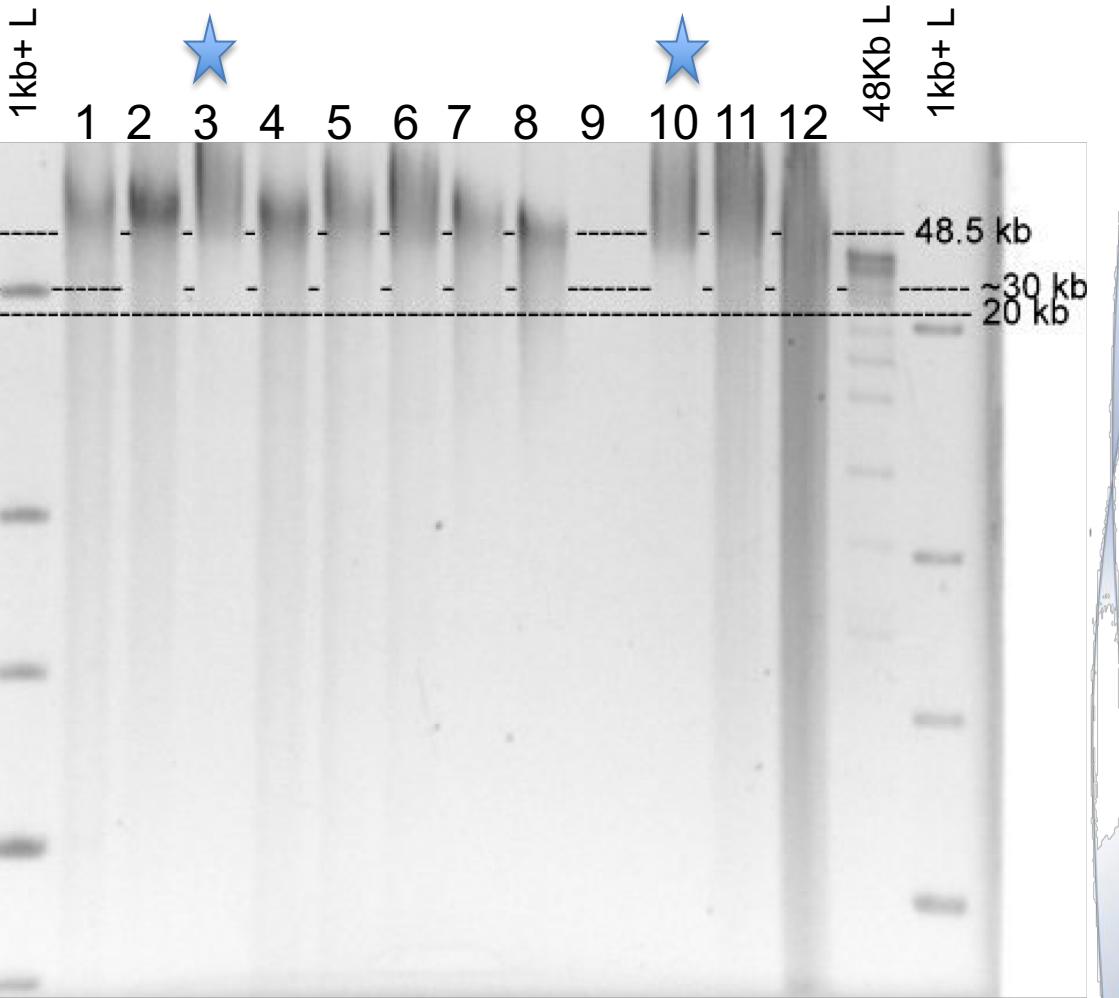


➤ Femto pulse



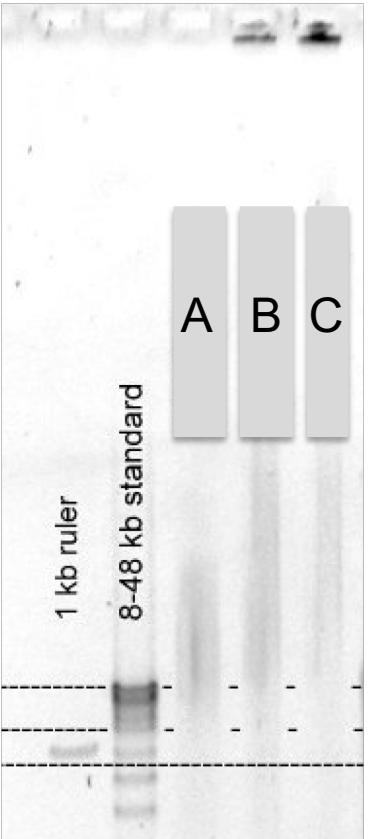
HMW gDNA QC guidelines

- Above 40kb!
- No smear below 20kb.
- Free of RNA, protein, and carbohydrates.
- Nanodrop ratio (2.0) for both 260/230 and 260/280.



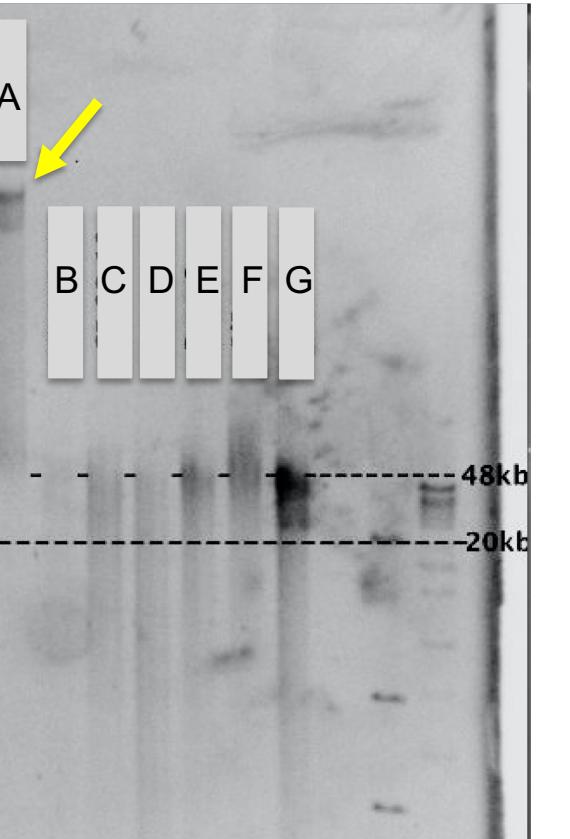
QC Examples

Example #1



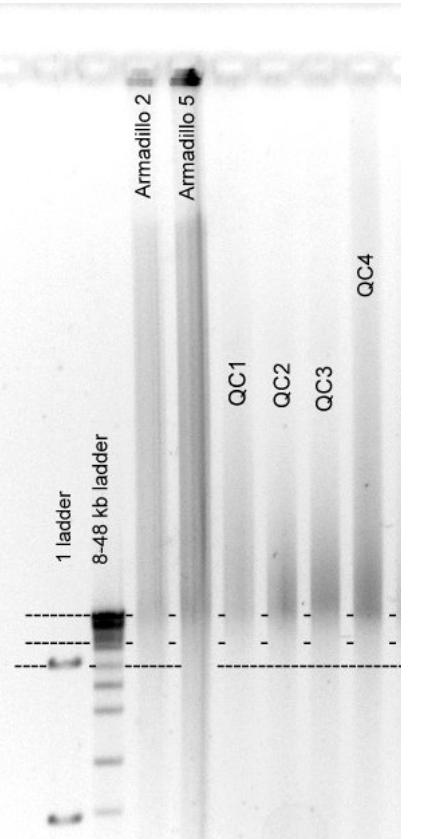
Look at loading wells.

Example #2



Bands are better than smear.

Example #3



Loading amount impacts QC.

Sample requirements

- Input into library prep 0.6ng-1.25ng.
 - Input depends on genome size.
- Additional 200 ng for QC.
- 40kb minimum, but 60kb better.
 - Don't size select (new reco from us), DNA damage repair optional.



TECHNICAL NOTE

Sample Preparation Recommendations for the Chromium™ Genome Kit

<https://support.10xgenomics.com/>

10X Chromium Genome

linked read assembly

...providing *de novo* genome assembly, variant calling, and genome structure information...

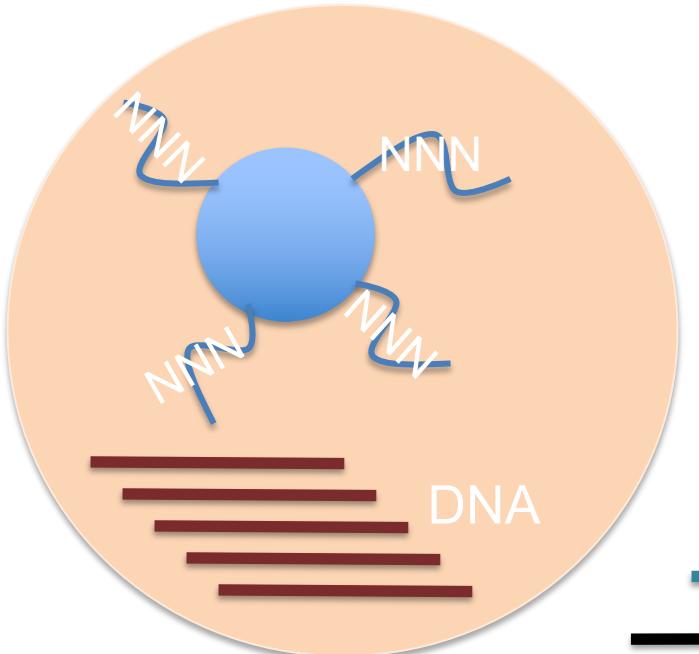
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10X Genomics

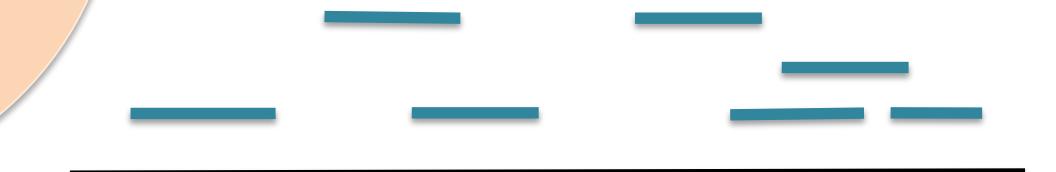
(genomic DNA analysis, CNV, and SC)



GemCode technology



- Barcoded amplicons generated in gel beads provide building blocks of genome.
- Droplet-based technology. Subset of genome partitioned in oil droplets with beads with a millions of barcodes.



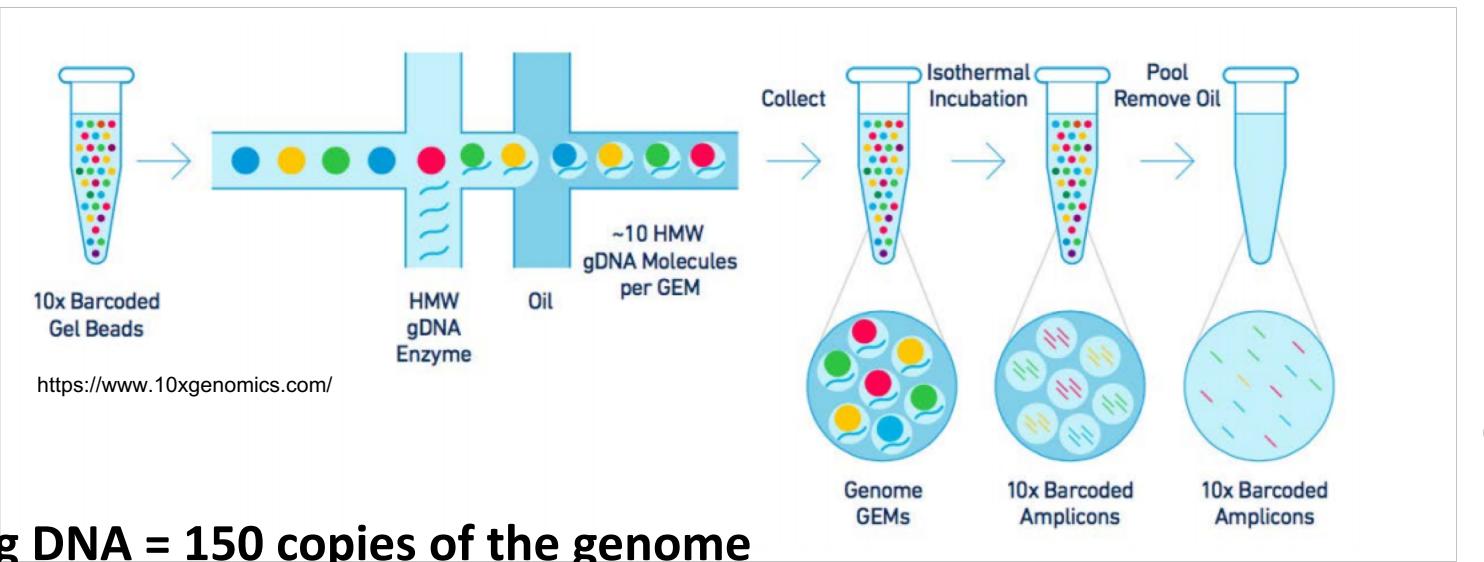
GEM 1



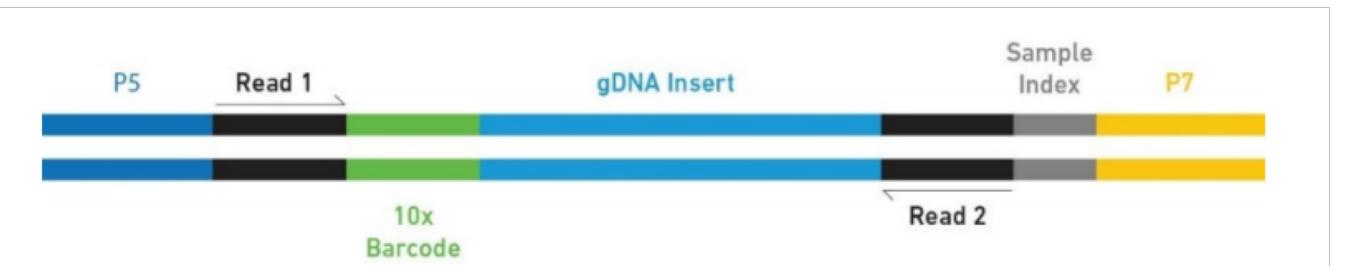
GEM 2

➤ “Read clouds”: molecules inferred linked reads

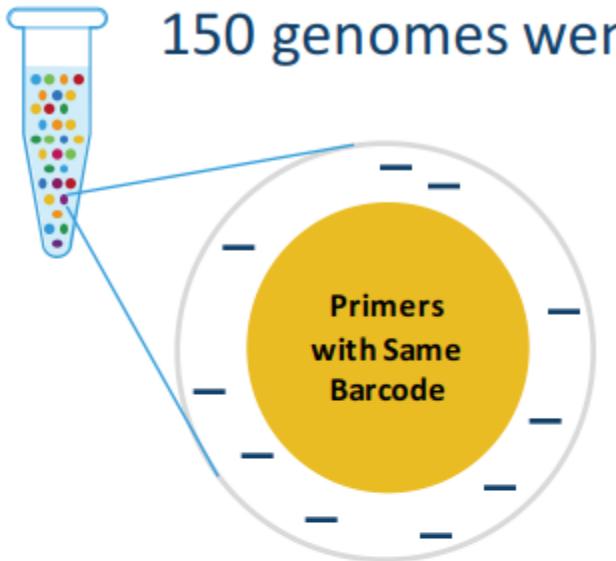
From gDNA to library



**0.5ng DNA = 150 copies of the genome
partitioned into ~1M GEMs.**



Molecule partitioning – human



150 genomes went into 1M partitions

Each GEM contains:

- One barcode (many copies)
- 1/6000 of the genome (500 Kb)
- At 50Kb length, 10 molecules

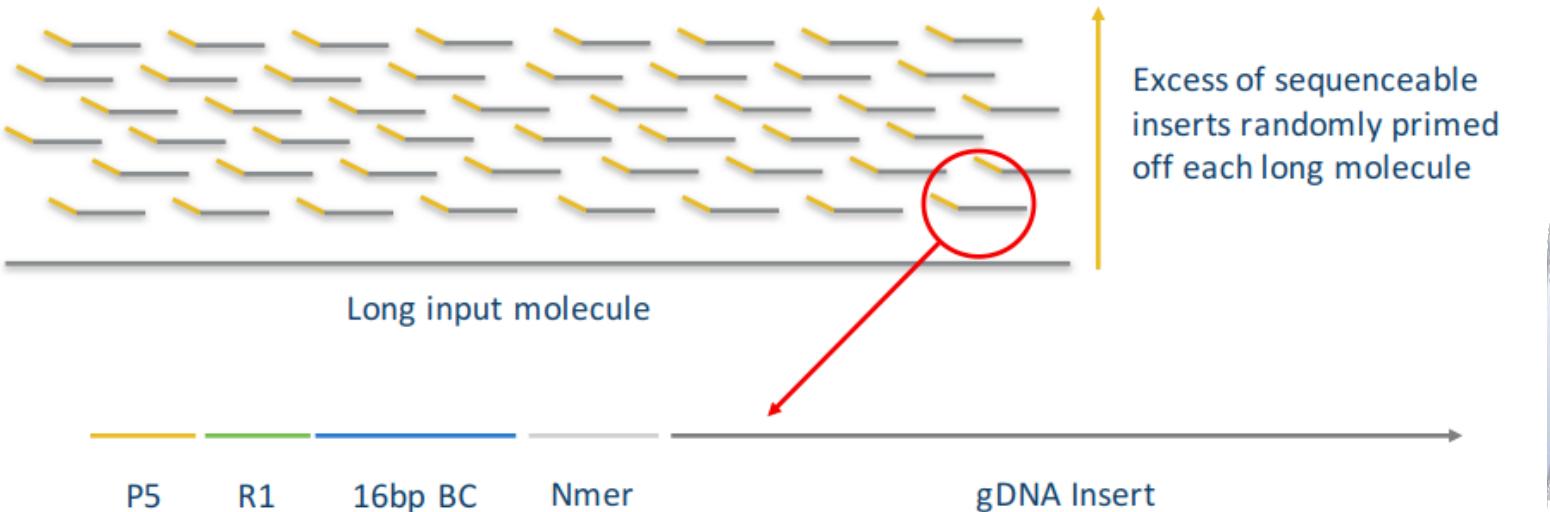
Chance that 2 molecules covering a locus are in same GEM:

1 in 6000

Percent unique barcodes at any genomic locus:

99.98%

Molecule coverage



At 30X read coverage, ~35 library fragments
will end up sequenced from each 50Kb input molecule

$35 \times 2 \times 150\text{bp} \approx 10\text{Kb}$, or 0.2X read coverage per molecule

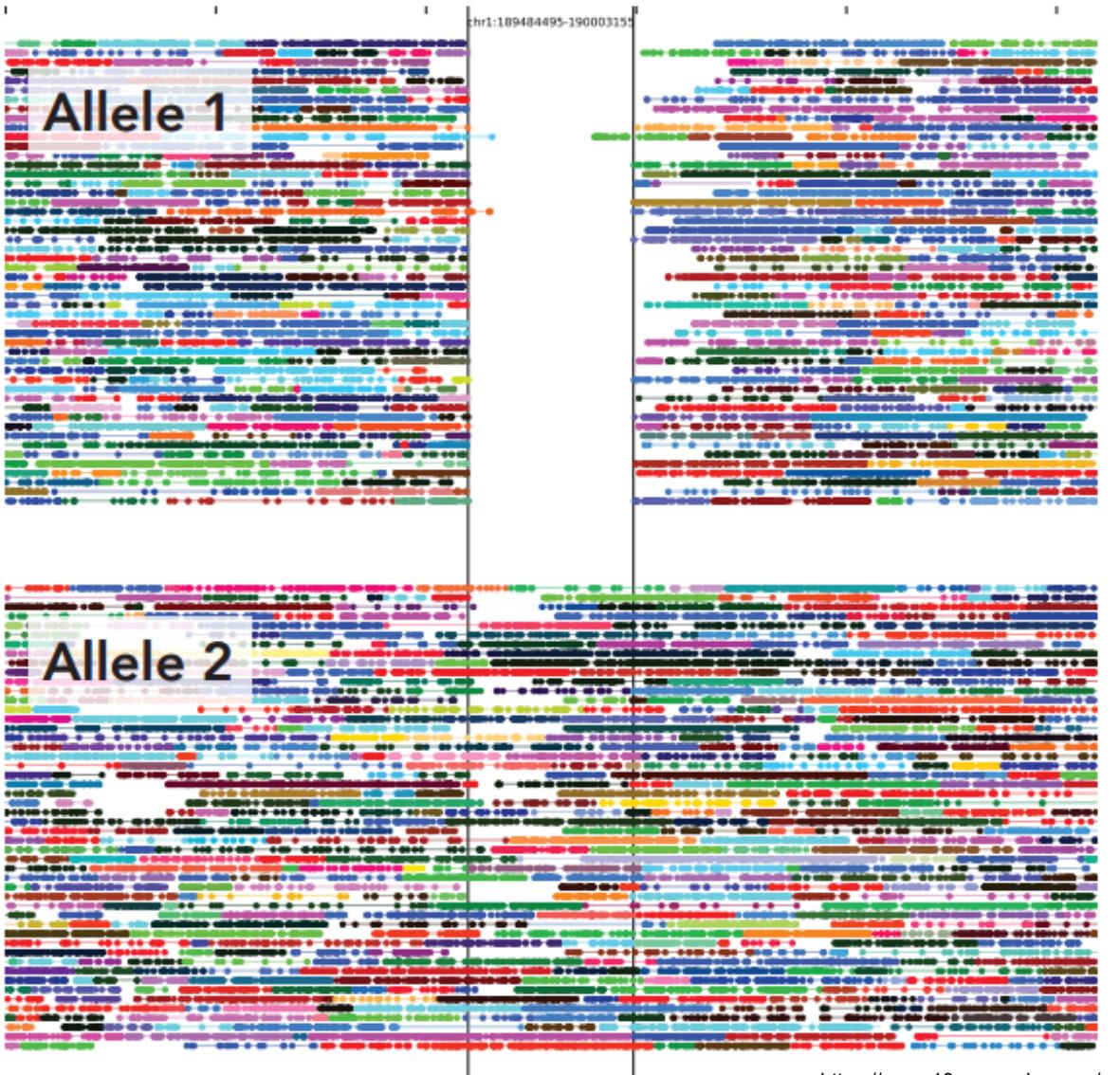
Reads from the same input molecule are called “Linked-Reads”

- Very little gDNA loaded into GEMs (some lost).
- Because so little gDNA added, unlikely that two haplotypes will have same barcode.

Read coverage recommendations

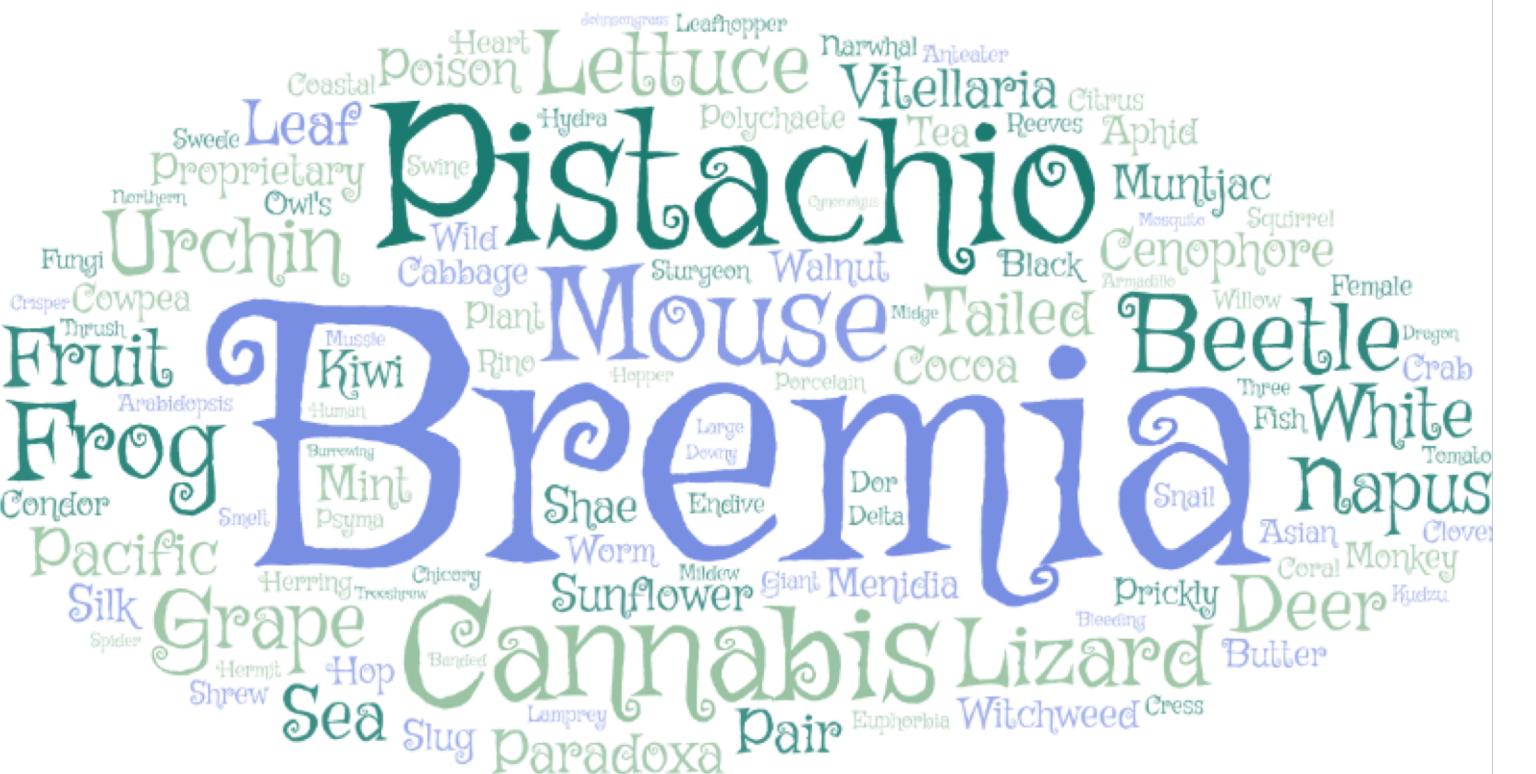
- Genome assembly: 60X coverage
- Structural variants: 25X coverage
- Too many reads doesn't improve assembly.
 - Worth running multiple assemblies with subsets of reads.

Structural variant detection



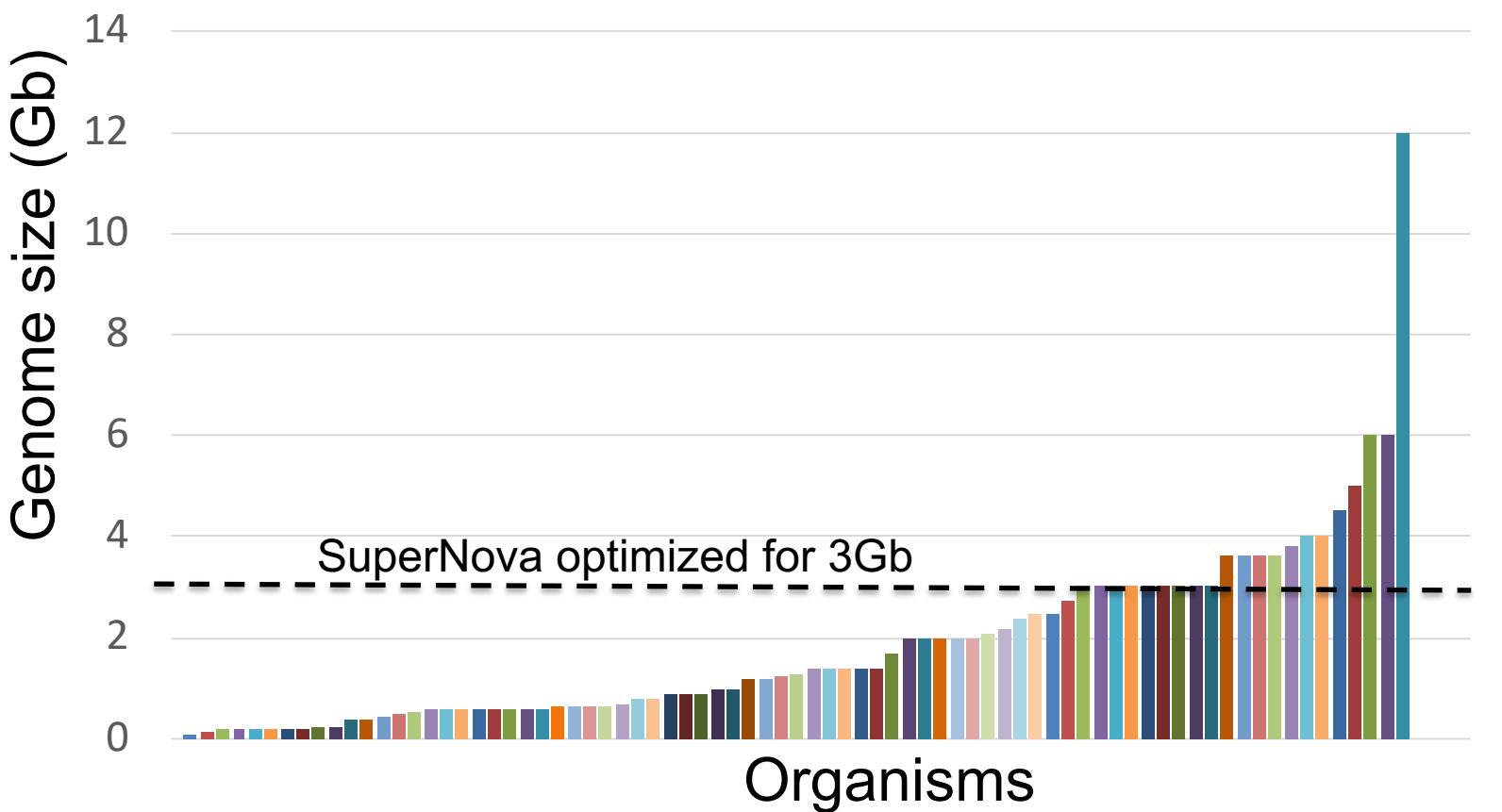
- Each colored line represents linked read.
 - Linked reads used to infer alleles.
- 60 Kb deletion visible.

DNA Tech 10X Genome Assemblies



De novo genome assembly

- 120 genomes to date.
- Smallest genome: 78Mb (Oomycete)
- Largest genome: 12Gb (frog, way too big!)



Assembly Stats - Best

- Mammals, birds, and reptiles.
- Example #1 (3.01 Gb genome)
 - Assembly size: 2.49 Gb
 - Molecule length: 174.31 Kb
 - Contig N50: 334.53 Kb
 - Scaffold N50: 38.80 Mb (entire chromosome arms)
- Example #2 (3.00 Gb genome)
 - Assembly size: 2.3 Gb
 - Molecule length: 118.08 Kb
 - Contig N50: 87.32 Kb
 - Scaffold N50: 7.41 Mb

Assembly Stats - Suboptimal

- Insects, marine life, plants (variable)
 - Depends on genome architecture, gut contents, metabolites, heterozygosity / variant density, ploidy.
- Example #1 (400 Mb genome)
 - Assembly size: 200 Mb
 - Molecule length: 13.42 Kb
 - Contig N50: 13.86 Kb
 - Scaffold N50: 40 Kb
- Example #2 (790 Mb genome)
 - Assembly size: 369.98 Mb
 - Molecule length: 64.70 Kb
 - Contig N50: 16.60 Kb
 - Scaffold N50: 90.45 Kb

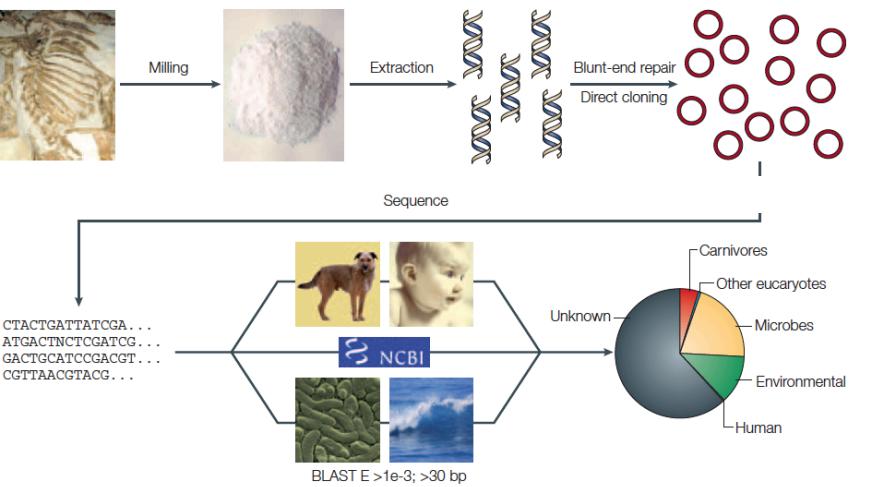
10X Chromium Genome

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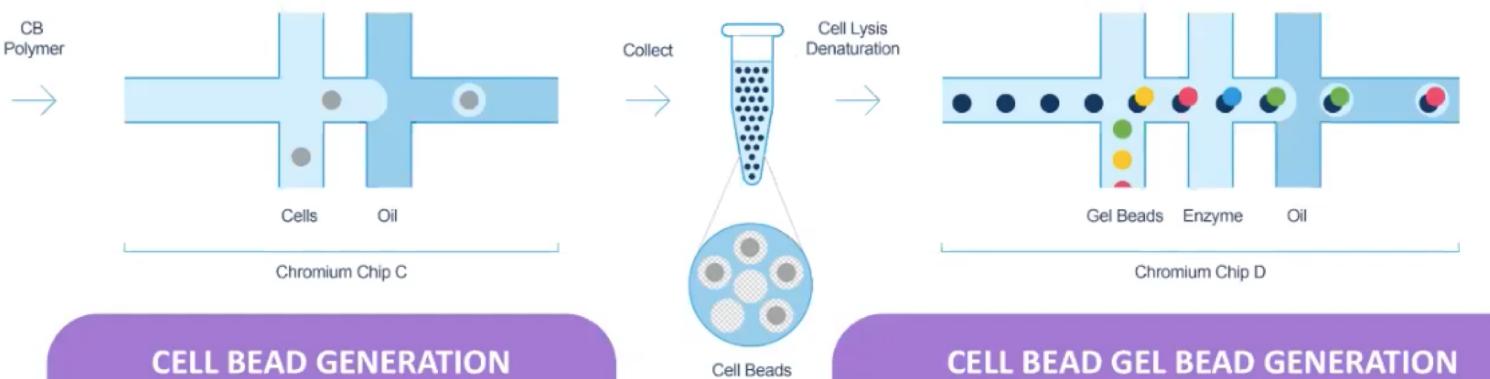
Summary of 10X Genome



- 10X great option if you are a human, bird, lizard, or diploid.
- Max genome size = 7.5 Gb / 2.14 B reads.
- 120 *de novo* genomes in core with linked reads.
 - High N50 = >300 Kb. Low N50 = 8 Kb (DNA damage).
- Plants are risky, but can still provide better assemblies.

Copy Number Variation

- Capture 100-1000s of single cell → copy number information.
- Calls single cell (or nuclei) CNV at 2 Mb resolution.
- Important tool to study dosage imbalances → changes in traits.
 - CNVs determine phenotypes more than SNPs.



CELL BEAD GENERATION

Input

- Cell Bead Polymer
- Cell Suspension in Cell Matrix

Output

- Cell Bead Emulsion

CELL BEAD GEL BEAD GENERATION

Input

- Cell Beads
- Gel Beads
- Enzymes

Output

- Cell Bead Gel Bead Emulsion (CBGBs)



PACIFIC
BIOSCIENCES™

<http://pacificbiosciences.com>



- Read long molecules in real-time with polymerase.
- Very long reads.
 - Subread N50: up to 35kb.
 - Polymerase read length: up to 100Kb for CCS.
 - Yield: up to 50 Gb for CCS.
 - High error rate for raw data (~13%), but random (unlike Nanopore).

Iso-Seq Pacbio

- Sequence full length transcripts
 - Using TeloPrime protocol for mostly full length transcripts.
 - No assembly required.
- High accuracy – CCS data.
- More than 95% of genes show alternate splicing.
- On average more than 5 isoforms/gene.
- Precise delineation of transcript isoforms.
(PCR artifacts? chimeras?).
- Ideal for gene annotation.

Please contact Oanh Nguyen (ohnguyen@ucdavis.edu)

Post Short Read Assemblies

- The future of sequencing is longer and longer reads.
 - Price dropping significantly.
 - Do 10X first because cheap?
- If 10X alone doesn't work, use combined assemblies (PacBio + 10X + Hi-C).
- Even suboptimal 10X data can be used for scaffolding with ARKS.
- Focusing on high molecular weight DNA can help obtain longer read lengths.
 - Junk in is junk out.
- But now we have to figure out how to use these data!

Price List – UC Rate

custom projects

- 10X Genome
 - Library prep: \$918.
 - Sequencing: \$1,500 for each 1.5Gb genome (NovaSeq, PE150).
- HMW gDNA extraction
 - Labor: \$792 (plants, 1-4 samples)
 - Reagents: \$100 per sample.
- 10X Single Cell CNV
 - TBD. But currently \$\$\$\$, but looking for testers.
- PromethION
 - \$2,880 per experiment (library prep and sequencing).
- Hi-C
 - \$1,690 (library prep only) + 100 million reads per 1.0 Gb genome (HiSeq4000 PE150).

Thank you!

“Safety first”



“Davis smog days”



From left to right:

Lutz – *Core Director*

Oanh – *PacBio*

Siranoosh – *HiSeq4000, MiSeq, and smallRNA*

Vanessa – *MiSeq, Genotyping*

Emily – *Library prep*

Ruta – *Nanopore, HMW gDNA extraction, Hi-C*