

Introduction to Microbial Sequencing

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General rules for preparing and experiment/samples

- Prepare more samples than you are going to need, i.e. expect some will be of poor quality, or fail
- Preparation stages should occur across all samples at the same time (or as close as possible) and by the same person
- Spend time practicing a new technique to produce the highest quality product you can, reliably
- Quality should be established using Fragment analysis traces (pseudo-gel images, RNA RIN > 7.0)
- DNA/RNA should not be degraded
 - 260/280 ratios for RNA should be approximately 2.0 and 260/230 should be between 2.0 and 2.2. Values over 1.8 are acceptable
- Quantity should be determined with a Fluorometer, such as a Qubit.

Sample preparation

In high throughput biological work (Microarrays, Sequencing, HT Genotyping, etc.), what may seem like small technical details introduced during sample extraction/preparation can lead to large changes, or technical bias, in the data.

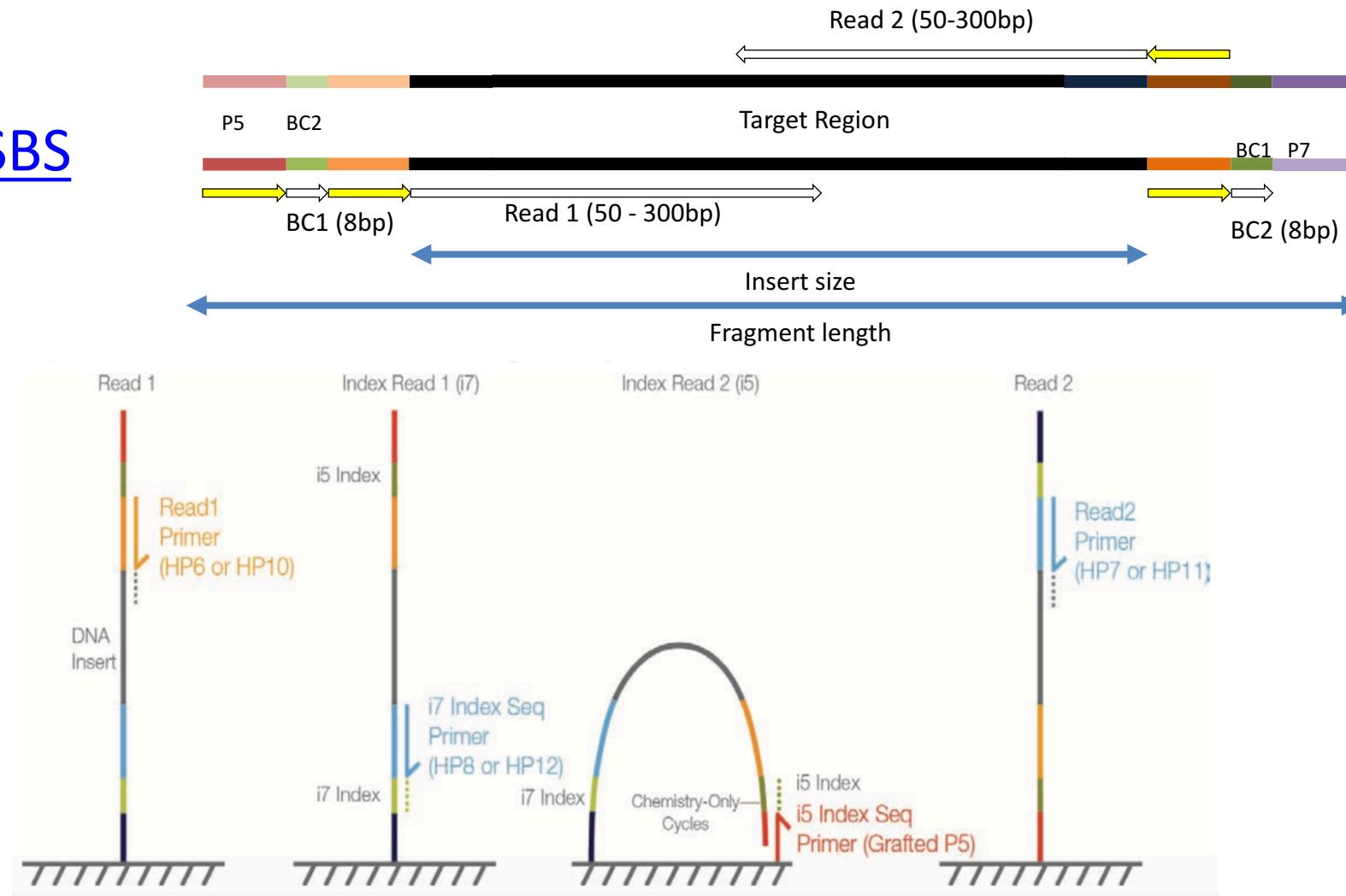
Not to say this doesn't occur with smaller scale analysis such as Sanger sequencing or qRT-PCR, but they do become more apparent (seen on a global scale) and may cause significant issues during analysis.

Be Consistent

BE CONSISTENT ACROSS ALL SAMPLES!!!

Illumina sequencing

- Illumina SBS



<https://www.illumina.com/systems/sequencing-platforms/miseq/specifications.html>

Illumina MISEQ SEQUENCING

Cluster Generation and Sequencing

	MiSeq Reagent Kit v2					MiSeq Reagent Kit v3	
Read Length	1 x 36 bp	2 x 25 bp	2 x 150 bp	2 x 250 bp	2 x 75 bp	2 x 300 bp	
Total Time*	~4 hrs	~5.5 hrs	~24 hrs	~39 hrs	~21 hrs	~56 hrs	
Output	540–610 Mb	750–850 Mb	4.5–5.1 Gb	7.5–8.5 Gb	3.3–3.8 Gb	13.2–15 Gb	

	MiSeq Reagent Kit v2 Micro		MiSeq Reagent Kit v2 Nano	
Read Length	2 x 150 bp	2 x 250 bp	2 x 150 bp	2 x 150 bp
Total Time*	~19 hrs	~28 hrs	~17 hrs	~17 hrs
Output	1.2 Gb	500 Mb	300 Mb	300 Mb

* Total time includes cluster generation, sequencing, and base calling on a MiSeq System enabled with dual-surface scanning.

Reads Passing Filter**

	MiSeq Reagent Kit v2	MiSeq Reagent Kit v3	MiSeq Reagent Kit v2 Micro	MiSeq Reagent Kit v2 Nano
Single Reads	12–15 million	22–25 million	4 million	1 million
Paired-End Reads	24–30 million	44–50 million	8 million	2 million

** Install specifications based on Illumina PhiX control library at supported cluster densities (865–965 k/mm² clusters passing filter for v2 chemistry and 1200–1400 k/mm² clusters passing filter for v3 chemistry). Actual performance parameters may vary based on sample type, sample quality, and clusters passing filter.

Quality Scores†

MiSeq Reagent Kit v2	MiSeq Reagent Kit v3
> 90% bases higher than Q30 at 1 x 36 bp	> 85% bases higher than Q30 at 2 x 75 bp
> 90% bases higher than Q30 at 2 x 25 bp	> 70% bases higher than Q30 at 2 x 300 bp
> 80% bases higher than Q30 at 2 x 150 bp	
> 75% bases higher than Q30 at 2 x 250 bp	

† A quality score (Q-score) is a prediction of the probability of an error in base calling. The percentage of bases > Q30 is averaged across the entire run.



<https://www.illumina.com/systems/sequencing-platforms/hiseq-3000-4000/specifications.html>

Illumina HiSeq Sequencing

Performance Parameters

	HiSeq 3000 System	HiSeq 4000 System
No. of Flow Cells per Run	1	1 or 2
Data Yield - 2 × 150 bp	650–750 Gb	1300–1500 Gb
Data Yield - 2 × 75 bp	325–375 Gb	650–750 Gb
Data Yield - 1 × 50 bp	105–125 Gb	210–250 Gb
Clusters Passing Filter (8 lanes per flow cell)	up to 2.5B single reads or 5B paired end reads	up to 5B single reads or 10B PE reads
Quality Scores - 2 × 50 bp	≥ 85% bases above Q30	≥ 85% bases above Q30
Quality Scores - 2 × 75 bp	≥ 80% bases above Q30	≥ 80% bases above Q30
Quality Scores - 2 × 150 bp	≥ 75% bases above Q30	≥ 75% bases above Q30
Daily Throughput	> 200 Gb	> 400 Gb
Run Time	< 1–3.5 days	< 1–3.5 days
Human Genomes per Run*	up to 6	up to 12
Exomes per Run†	up to 48	up to 96
Transcriptomes per Run‡	up to 50	up to 100

Install specifications based on Illumina PhiX control library at supported cluster densities (between 1310–1524 K/mm² passing filter). Run times correspond to sequencing only. Performance may sample quality, cluster density, and other experimental factors.

*Assumes >30x coverage of a human genome.

†Assumes 100x coverage with 80% on target using 2 × 75 bp reads.

‡Assumes 50 million reads per sample.



Sequencing Depth

- The first and most basic question is how many base pairs of sequence data will I get

Factors to consider are:

- 1. Number of reads being sequenced
- 2. Read length (if paired consider them as individuals)
- 3. Number of samples being sequenced
- 4. Expected percentage of usable data

$$bpPerSample = \frac{readLength * readCount}{sampleCount} * 0.8$$

- The number of reads and read length data are best obtained from the manufacturer's website (search for specifications) and always use the lower end of the estimate.

Genomic Coverage

Once you have the number of base pairs per sample you can then determine expected coverage

Factors to consider then are:

1. Length of the genome
2. Any extra-genomic sequence (ie mitochondria, virus, plasmids, etc.). For bacteria in particular, these can become a significant percentage

$$\frac{\text{ExpectedCoverage}}{\text{sample}} = \frac{\frac{(\text{readLength} * \text{numReads}) * 0.8}{\text{numSamples}} * \text{num.lanes}}{\text{TotalGenomicContent}}$$

Sequencing Depth – Counting based experiments

- Coverage is determined differently for "Counting" based experiments (RNAseq, amplicons, etc.) where an expected number of reads per sample is typically more suitable.
- The first and most basic question is how many reads per sample will I get
Factors to consider are (per lane):
 1. Number of reads being sequenced
 2. Number of samples being sequenced
 3. Expected percentage of usable data
 4. Number of lanes being sequenced

$$\frac{\text{reads}}{\text{sample}} = \frac{\text{reads.sequenced} * 0.8}{\text{samples.pooled}} * \text{num.lanes}$$

- Read length, or SE vs PE, does not factor into sequencing depth.

Amplicon Sequencing (Communities, genotyping)

Considerations

- Number of reads being sequenced
- Proportion that is diversity sample (e.g. PhiX)
- Number of samples being pooled in the run

The back of the envelope calculation

$$\frac{\text{reads}}{\text{sample}} = \frac{\text{reads_sequenced} * (1 - \text{diversity_sample})}{\text{num_samples}}$$

example

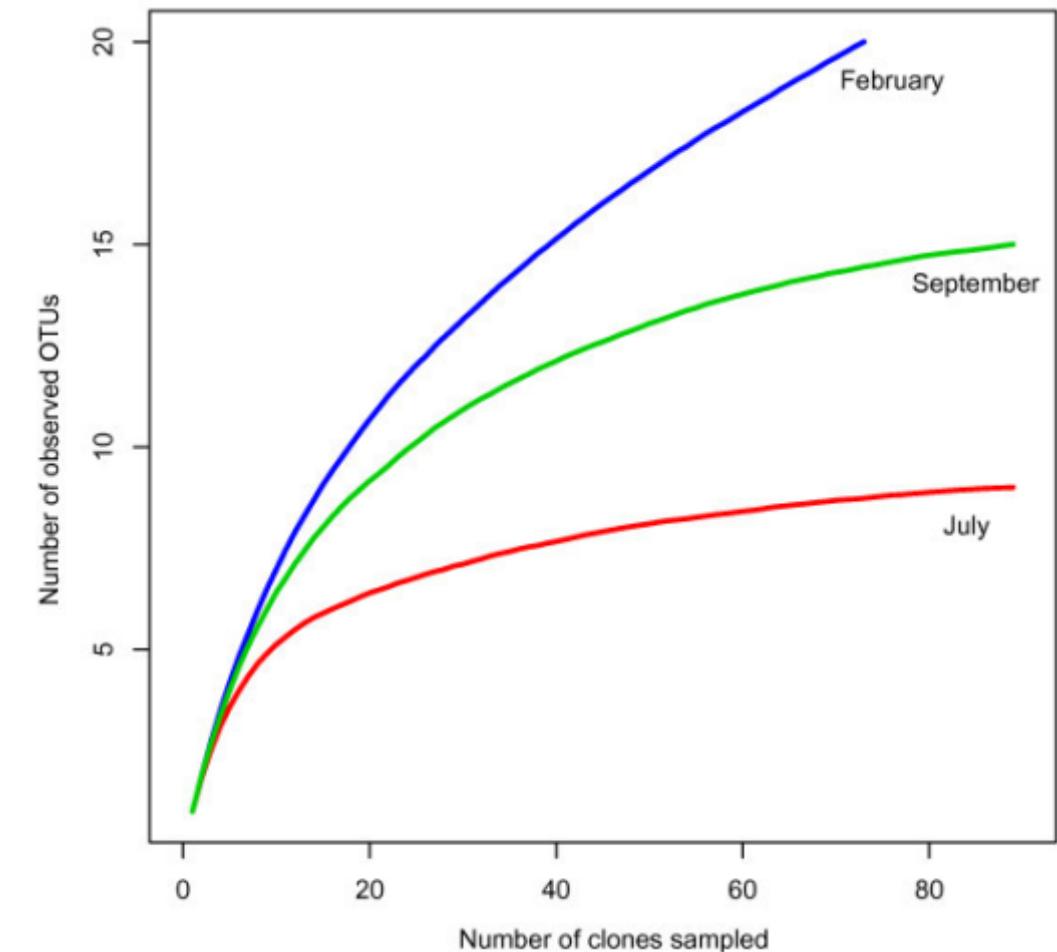
$$\frac{102,000}{\text{sample}} = \frac{18e6 * (1 - 0.15)}{150}$$

Recommendations

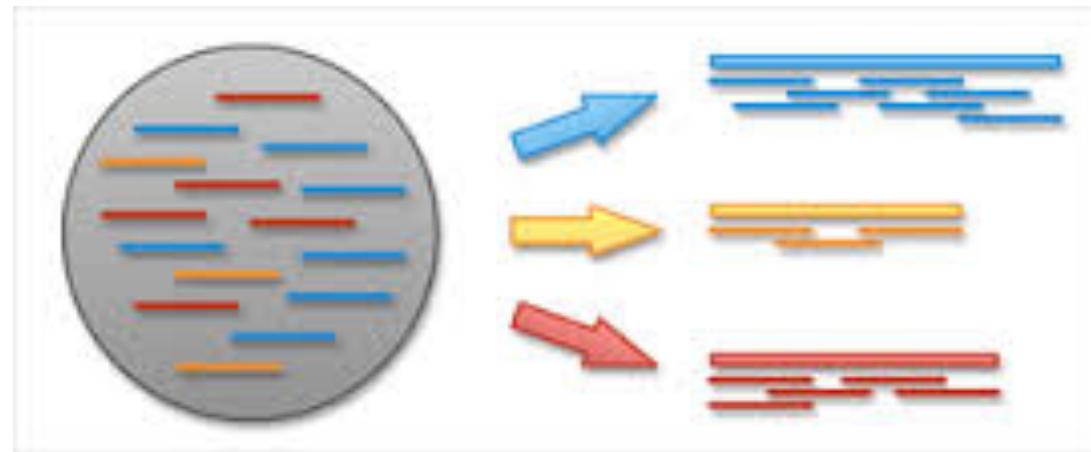
- Illumina ‘recommends’ 100K per sample
- I’ve used 30K per sample historically, others are fine with 3K per sample
- Really should have as many reads as your experiment needs

How Much? Community Rarefaction curves

- 'Deep' sequence a number of test samples
amplicons: ~ 1M+ reads.
metagenomics: 1 full HiSeq lane
- Plot rarefactions curves of organism identification, to determine if saturation is achieved



Metagenomics assembly



To determine if you've sequenced 'enough' to re-assemble 'most' of the community member's genetic content, look to what is left over - proportionally

Amplicons vs. Metagenomics

- Metagenomics
 - Shotgun libraries intended to sequence random genomic sequences from the entire bacterial community.
 - Can be costly per sample (\$500 to multi thousands per sample)
 - Better resolution and sensitivity to characterize the sample
 - Due to cost, can only do relatively few samples
- Amplicon community profiling
 - Sequence only one regions of one gene (e.g. 16s, ITS, LSU)
 - Cheap per sample (at scale, down to \$20/sample)
 - Due to cost, can do many hundreds of samples make more global inferences

Community Sequencing Designs

- Taxonomic Identification
 - Amplicon based (e.g. 16s variable regions)
 - Shotgun Metagenomics
- Functional Characterization
 - Shotgun Metagenomics
 - Shotgun Metatranscriptomics (active)
- Genome Assembly, Function and Variation
 - Shotgun Metagenomics
 - Shotgun Metatranscriptomics

Cost Estimation

- DNA/RNA extraction and QA/QC (Bioanalyzer/Gels)
- Metatranscriptomes: Enrichment of RNA of interest and RNA library preparation
 - Library QA/QC (Bioanalyzer and Qubit)
 - Pooling (\$10/library)
- Metagenomes: DNA library preparation
 - Library QA/QC (Bioanalyzer and Qubit)
 - Pooling (\$10/library)
- Community Profiling: PCR reactions
 - Library QA/QC (Bioanalyzer and Qubit/microplate reader)
 - Pooling
- Sequencing (Number of Lanes / runs)
- Bioinformatics (General rule is to estimate the same amount as data generation, i.e. double your budget)

<http://dnatech.genomecenter.ucdavis.edu/prices/>

Bioinformatics Costs

Bioinformatics includes:

1. Storage of data
2. Access and use of computational resources and software
3. System Administration time
4. Bioinformatics Data Analysis time
5. Back and forth consultation/analysis to extract biological meaning

Rule of thumb:

Bioinformatics can and should cost as much (sometimes more) as the cost of data generation.

Cost Estimation

- Amplicons
 - 384 Samples
 - Amplicon generation (\$20/sample)= $384 * 20 = \$7,680$
 - Sequencing PE300, target 30K reads per sample
 - Bioinformatics
- Metagenome
 - 12 samples (DNA)
 - Expectations: Host Proportion 40%, use average genome size of eColi,
Target the 1% and coverage of 20
 - Sequencing PE150
 - Bioinformatics

Take Homes

- Experience and/or literature searches (other peoples experiences) will provide the best justification for estimates on needed depth.
- ‘Longer’ reads are better than short reads.
- Paired-end reads are more useful than single-end reads
- Libraries can be sequenced again, so do a pilot, perform a preliminary analysis, then sequence more accordingly.

Why do we utilize 16S sequencing?

- All bacteria have a 16S gene
- Highly conserved because it is necessary for ribosomes to translate mRNA
- Because this gene has changed very little over time we can construct phylogenies to examine the evolution of bacteria and their relationships to one another
- This conserved nature also allows us to construct universal primers that will amplify gene sequences across many different bacterial groups

What 16s Is and Is NOT

IS	IS NOT
Tool for Generating New Hypotheses	Tool for Generating the Final Answer
Powerful when combined with other techniques like qPCR and Gene Expression data	A standalone technique that requires no other validation
Qualitative with some quantitative elements depending on experimental design	A purely quantitative tool that will solve all of your problems and wrap them up with a nice little bow
The Beginning	The End

16S sequencing is a valuable resource IF you understand its limitations

