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# Assembling the drosophila genome with IPA and HiFi data

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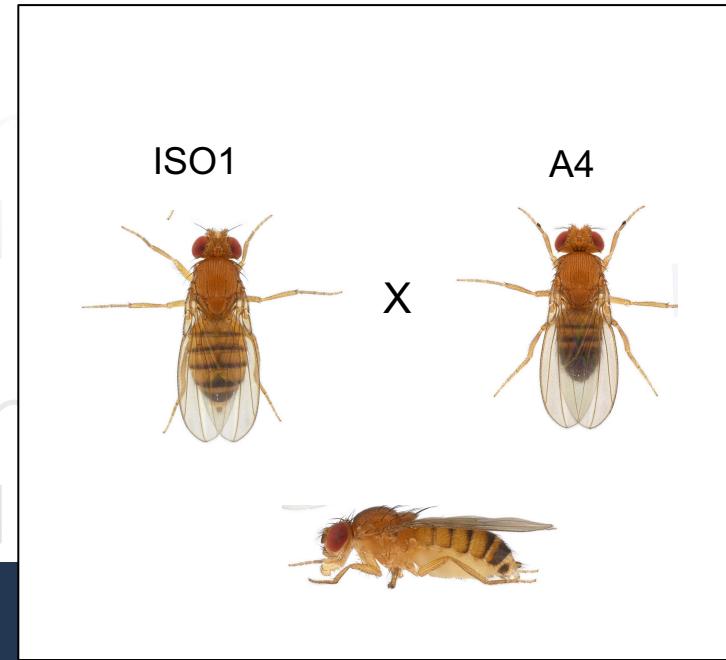
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# AGENDA

- Introduction
- Setup PacBio software
- Learn about command line interface
- Run an IPA assembly



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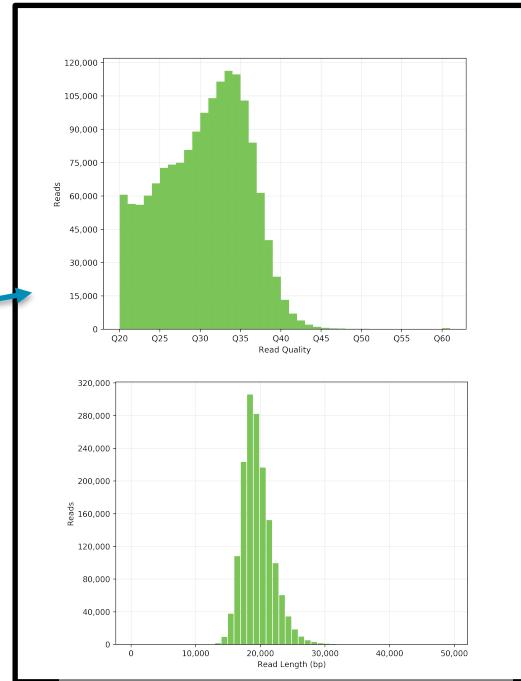
# Today's dataset

Drosophila

# DROSOPHILA DATASET INFO

## 19 Kb dataset

- Processing of PacBio data
  - Circular consensus algorithm was done with SMRT Link
  - Data was subsampled down to 38x depth of coverage (DOC)
  
- Short read data
  - Standard procedures
  - Both parental strains were sequenced (70-90x DOC)
  - Utility
    - Trio binning
    - Phasing evaluation



# SETUP CONDA – A SOFTWARE MANAGEMENT SYSTEM

- Google: `conda install`
  - <https://docs.conda.io/projects/conda/en/latest/user-guide/install/linux.html>
- Follow link for download (linux x86)
- Follow install directions, **setup BASH shell!**
- You will likely need to source your .bashrc if conda isn't in your path after setup

- Why does PacBio use Conda?
  - Central repository of many bioinformatic tools
  - We can distribute binary code
  - We can post updates quickly
  - Many more...

## SETUP IPA



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- Google: `pacbio bioconda`
  - <https://github.com/PacificBiosciences/pbbioconda>
- Familiarize yourself with available command line tools available for download.
- Go to IPA bioconda wiki:
  - <https://github.com/PacificBiosciences/pbbioconda/wiki/Improved-Phased-Assembler>



## SETUP IPA



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```
conda create -n ipa -c  
bioconda -c conda-forge -c  
defaults conda activate ipa  
conda install pbipa
```



UC DAVIS SETUP



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```
eval "$(~/share/biocore/shunter/2020-  
07-15-IPA-tests/conda/bin/conda  
shell.bash hook)"
```



# COMMAND LINE



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```
(ipa) zevk@tadpole:~$ ipa -h
usage: ipa [-h] [--version] {local,dist,validate} ...

Improved Phased Assembly tool for HiFi reads.

optional arguments:
  -h, --help            show this help message and exit
  --version             show program's version number and exit

subcommands:
  One of these must follow the options listed above and may be followed by sub-command specific options.

  {local,dist,validate}
    sub-command help
    local           Run IPA on your local machine.
    dist            Distribute IPA jobs to your cluster.
    validate        Check dependencies.

Try "ipa local --help".
Or "ipa validate" to validate dependencies.
https://github.com/PacificBiosciences/pbbioconda/wiki/Improved-Phased-Assembler
```

# KNOW YOUR VERSIONS



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- We commonly update IPA
- Before running assembly update IPA
- Keep track of your versions
- Avoid updating mid-assembly

```
(ipa) zevk@tadpole:~$ ipa validate
INFO: /home/zevk/anaconda3/envs/ipa/bin/ipa validate
Checking dependencies ...
/home/zevk/anaconda3/envs/ipa/bin/python3
/home/zevk/anaconda3/envs/ipa/bin/ipa2-task
/home/zevk/anaconda3/envs/ipa/bin/falconc
/home/zevk/anaconda3/envs/ipa/bin/nighthawk
/home/zevk/anaconda3/envs/ipa/bin/pancake
/home/zevk/anaconda3/envs/ipa/bin/pblayout
/home/zevk/anaconda3/envs/ipa/bin/racon
/home/zevk/anaconda3/envs/ipa/bin/samtools
snakemake version=5.20.1
Machine name: 'Linux'
ipa2-task 0.2.0 (commit 33ccb062c1db781cd9aa10e4341c670430b1e575)
falconc version=1.5.1+git.895d7f33113c17b399428ff45dce127f7aa635ef, nim-version=1.2.0
Nighthawk 0.1.0 (commit df65ce5*)
pancake 0.1.0 (commit 3a4146f*)
pblayout 0.1.0 (commit 5257a1a*)
racon version=v1.4.13
samtools 1.9
Using htslib 1.9
```

RUN IPA ON THE CLUSTER



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```
ipa dist -i ../hifi_long_read_data/ELF_19kb.m64001_190914_015449.Q20.38X.fasta \
--nthreads 24 --njobs 30 --cluster-args 'sbatch -J zev-ipa.{rule} -t 45 \
-c {params.num_threads} -e stderr -o stdout --get-user-env \
--chdir pacbio_2020_data_drosophila/hifi_long_read_diploid_ipa_assembly_cluster '
```

RUN IPA LOCALLY



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```
ipa local --nthreads 48 --njobs 2 -i ELF_19kb.m64001_190914_015449.Q20.38X.fasta
```



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## STAGES OF IPA

```
drwxrwsr-x 2 zevk genome_workshop    7 Jul 14 14:17 01-generate_config  
drwxrwsr-x 2 zevk genome_workshop 10 Jul 14 14:17 02-build_db  
drwxrwsr-x 3 zevk genome_workshop    3 Jul 14 14:18 03-ovl_asym_prepare  
drwxrwsr-x 5 zevk genome_workshop    5 Jul 14 14:21 04-ovl_asym_run  
drwxrwsr-x 2 zevk genome_workshop    8 Jul 14 14:24 05-ovl_asym_merge  
drwxrwsr-x 3 zevk genome_workshop  3 Jul 14 14:24 06-phasing_prepare  
drwxrwsr-x 8 zevk genome_workshop    8 Jul 14 14:43 07-phasing_run  
drwxrwsr-x 2 zevk genome_workshop 10 Jul 14 14:49 08-phasing_merge  
drwxrwsr-x 2 zevk genome_workshop 111 Jul 14 14:52 09-ovl_filter  
drwxrwsr-x 2 zevk genome_workshop 36 Jul 14 14:57 10-assemble  
drwxrwsr-x 3 zevk genome_workshop    3 Jul 14 14:57 11-polish_prepare  
drwxrwsr-x 5 zevk genome_workshop    5 Jul 14 15:04 12-polish_run  
drwxrwsr-x 2 zevk genome_workshop   7 Jul 14 15:10 13-polish_merge  
drwxrwsr-x 2 zevk genome_workshop   4 Jul 14 15:10 14-final  
-rw-rw-r-- 1 zevk genome_workshop 210 Jul 14 14:17 config.json  
-rw-rw-r-- 1 zevk genome_workshop 140 Jul 14 14:17 config.yaml  
-rw-rw-r-- 1 zevk genome_workshop 124 Jul 14 14:17 input.fofn  
-rw-rw-r-- 1 zevk genome_workshop 307 Jul 14 14:17 ipa.log  
drwxrwsr-x 2 zevk genome_workshop    2 Jul 14 14:17 qsub_log
```

# GETTING ASM STATS



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```
module load assembly_stats/1.0.1
```

```
assembly-stats -t final.p_ctg.fasta  
final.a_ctg.fasta | column -t
```

filename	total_length	number	mean_length	longest	shortest	N_count	Gaps	N50	N50n	N70	N70n	N90	N90n
..../hifi_long_read_diploid_ipa_assembly/RUN/14-final/final.p_ctg.fasta	232197789	208	1116335.52	23464522	50222	0	0	7970785	9	2631370	18	461442	54
..../hifi_long_read_diploid_ipa_assembly/RUN/14-final/final.a_ctg.fasta	36577756	287	127448.63	9995912	4424	0	0	1540414	5	649690	11	28408	128

## LOOKING AT YOUR ASM



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Make GFA

```
pblayout subgraph --tp all_ctg.tp sg_edges_list tp_graph
```

Remove

```
perl -lane '$_ =~ s/:B|:E//g; print' tp_graph.csv >
```

Double

```
tp_graph_single.csv ; perl -lane '$_ =~ s/:B|:E//g;
```

Edges

```
print' tp_graph.gfa > tp_graph_single.gfa
```

# LOOKING AT YOUR ASM



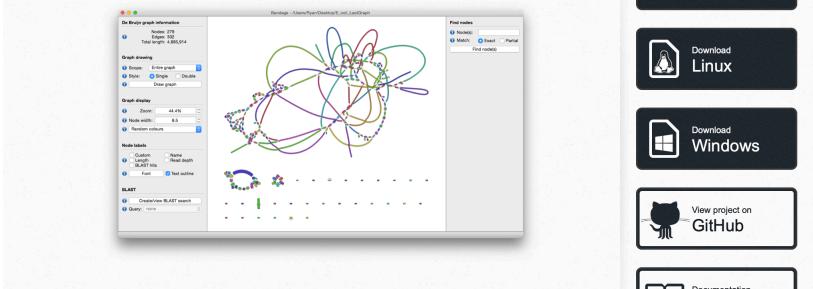
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## Bandage

a Bioinformatics Application for Navigating *De novo* Assembly Graphs Easily

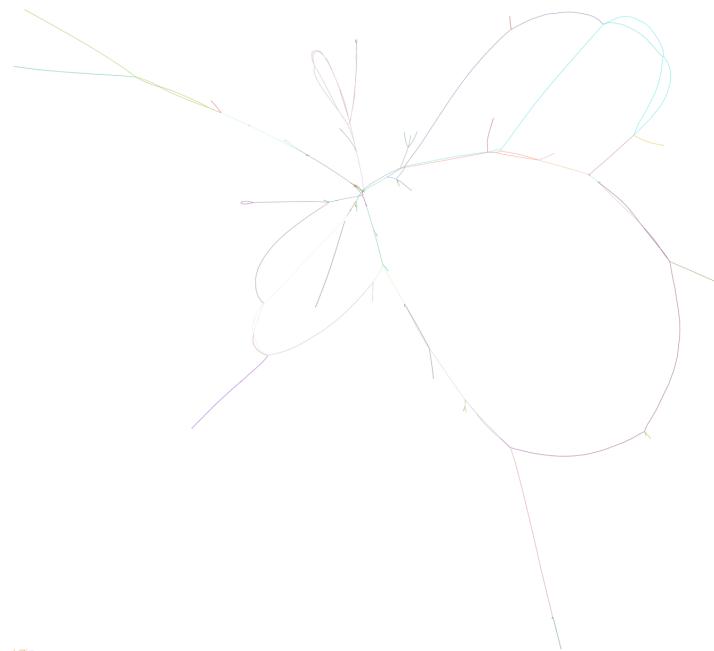
Bandage is a program for visualising *de novo* assembly graphs. By displaying connections which are not present in the contigs file, Bandage opens up new possibilities for analysing *de novo* assemblies.



The screenshot shows the Bandage software interface. On the left is a large window titled "De novo graph information" displaying a complex assembly graph with many nodes and edges. The graph is composed of various colors (green, blue, red, purple) representing different contigs or scaffolds. On the right side of the interface, there are several control panels and toolbars for navigating the graph, including "Order drawing" (with options "Euler", "Single", and "Double"), "Node labels" (with options "None", "Label", "Label & ID", and "Font"), and "BLAST" (with options "Create new BLAST search" and "BLAST"). Below the main window are four dark rectangular buttons with white text and icons:

- Download Mac
- Download Linux
- Download Windows
- View project on GitHub

At the bottom right is a small "Documentation" link.





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