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BIOSCIENCES®



# Single-Cell RNA Sequencing using the Iso-Seq Method

Elizabeth Tseng, Principal Scientist, PacBio

 @Magdoll

# SINGLE-CELL ISO-SEQ METHOD: WHY?

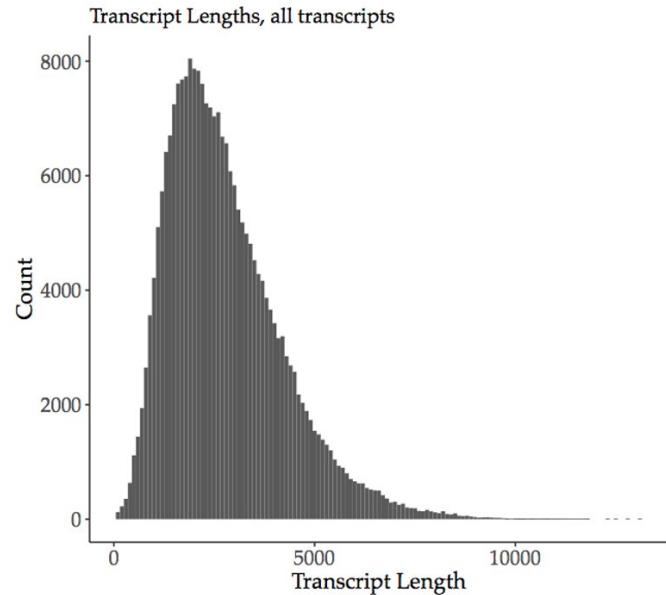
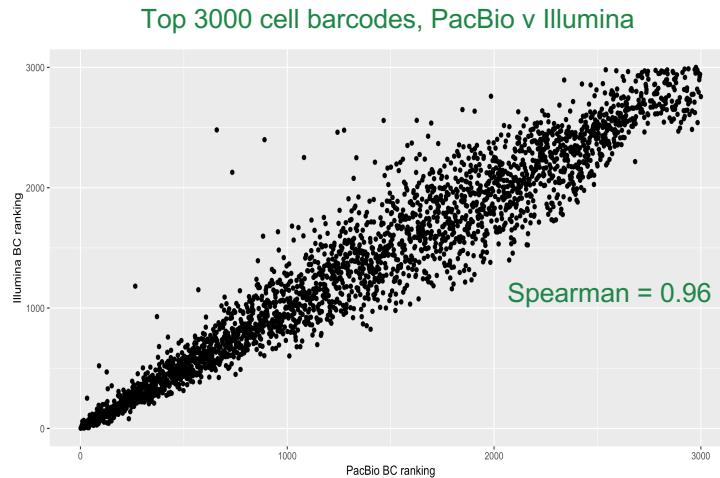


Cell type specific  
mRNA splicing

Not captured with 3'-end  
short-read scRNA seq

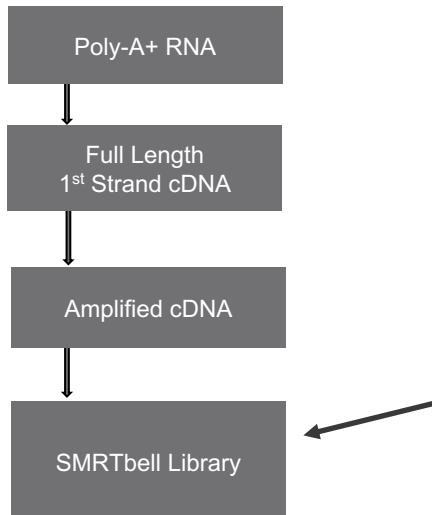
# PACBIO HIFI READS ARE ACCURATE FOR SINGLE-CELL

HiFi reads ~100% concordant with matching Illumina data

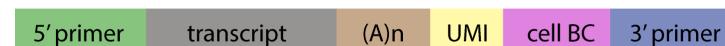
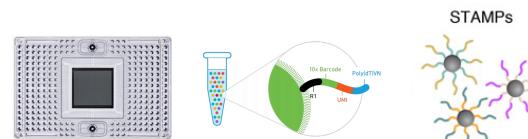


# COMPATIBLE WITH FULL-LENGTH SINGLE-CELL PLATFORMS

## Bulk RNA-seq on PacBio Systems



Any single-cell platform that generates full-length cDNA



# SINGLE-CELL ISO-SEQ WORKFLOW

- Compatible with any full-length single-cell platform
- Detailed workflow guidance
- Standard Iso-Seq library preparation & sequencing workflow
- Can generate matching short read data from same sample
- ~3 million HiFi reads per SMRT Cell 8M
- Flexible barcoding & multiplexing based on desired number of reads per cell

 PACBIO®

**Procedure & Checklist – Preparing Single-Cell Iso-Seq™ Libraries Using SMRTbell® Express Template Prep Kit 2.0**

**Before You Begin**

The Sequel Systems generate long reads that are well-suited for characterizing full-length transcripts produced from Single-Cell platforms. This document describes a method for constructing Single-Cell Iso-Seq SMRTbell® libraries for sequencing.

Generating Single-Cell Iso-Seq SMRTbell libraries is a two-step process. Initially, the intact RT-PCR product from a typical Single-Cell preparation is reamplified to increase the mass. Then the SMRTbell Express Template Prep Kit 2.0 is used for SMRTbell library preparation.

For best analytical results, we recommend combining matching (i.e., the same exact library) short-read and Iso-Seq datasets. We recommend that the reamplification yield allow for parallel processing of both short-read sequencing and SMRT® Sequencing. The Sequel System requires >80 ng of DNA, while the Sequel II System requires >160 ng DNA. These are target amounts for the reamplification steps for the Iso-Seq Express workflow.

Single-Cell library amplification primers for specific platform (primers may be ordered from any oligo synthesis company)	10x Chromium Single Cell 3' Solution V2 and V3: cDNA Forward primer (e.g., PCR Primer 1): 5'- CTACACGACGCTTCCGATCT -3' cDNA Reverse primer (e.g., PCR Primer 2): 5'- AAGCAGTGGTATCAACGCAGAGT-3'
Single cell cDNA amplification kit from your single cell vendor†	Any Single-Cell Vendor
Additional Single-Cell library amplification primers for specific platform may be required for parallel reamplification	For example, Drop-Seq: Both Primers are the same sequence for PCR Primer 1 and 2: 5'- AAGCAGTGGTATCAACGCAGAGT -3' [Macosko, et al., Cell, 2015:pp1202-14]  Commercial vendors may have different sequences for their PCR Primers. Please consult your vendor for the primer(s) sequence(s).

† Pacific Biosciences does not sell a kit for carrying out the Single-Cell RNA Sequencing method. Use of these methods may require rights to third-party owned intellectual property.



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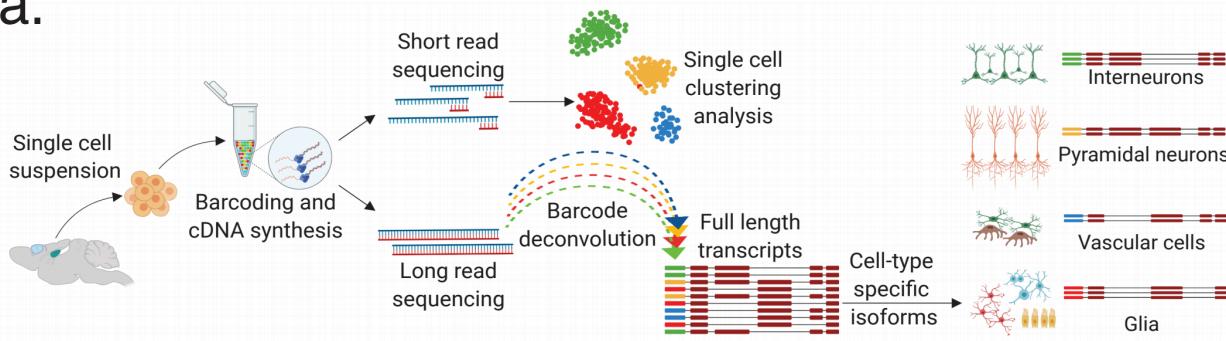
# Single-Cell Iso-Seq Research Highlights

# SINGLE-CELL ISO-SEQ METHOD ON DIFFERENT PLATFORMS

Single-cell Platform	Publications/Presentations	Research Areas
10x Genomics	<a href="#">Zheng et al. <i>biorxiv</i> (2020)</a>	Corneal epithelium
	<a href="#">Tilgner <i>SMRT Leiden</i> (2020)</a>	Brain
	<a href="#">Joglekar et al. <i>biorxiv</i> (2020)</a>	Brain
	<a href="#">Mincarelli et al. <i>biorxiv</i> (2020)</a>	Immune repertoire
	<a href="#">Russell et al. <i>J Virology</i> (2019)</a>	Influenza
Celsee (Bio-Rad)	Underwood <i>RNA Society</i> (2020)	Human/mouse cell cycle
Dolomite Bio	<a href="#">Underwood <i>AGBT</i> (2019)</a>	Primate organoids
Berkeley Lights	<a href="#">Zost et al. <i>biorxiv</i> (2020)</a>	COVID-19 antibodies
Various	<a href="#">Conessa <i>SMRT Leiden</i> (2020)</a>	Bioinformatics tools

# COMBINING SHORT- AND LONG-READ SINGLE CELL DATA

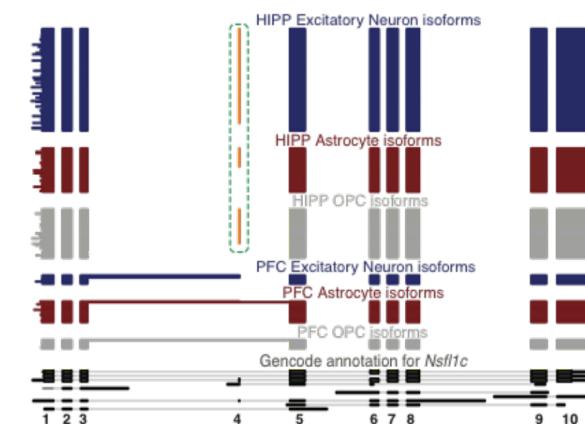
a.



Short read data identifies cell clusters

Long read data identifies full-length isoforms

Matching (UMI, BC) links isoforms to cell types



## CHIMP-HUMAN TSS DIFFERENCE IN SINGLE-CELL ORGANOIDS

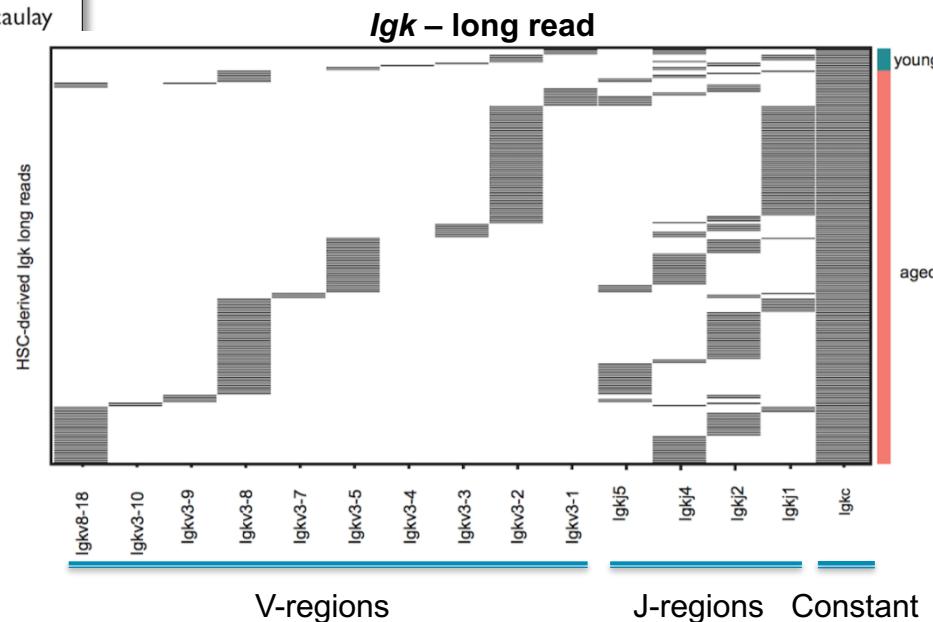
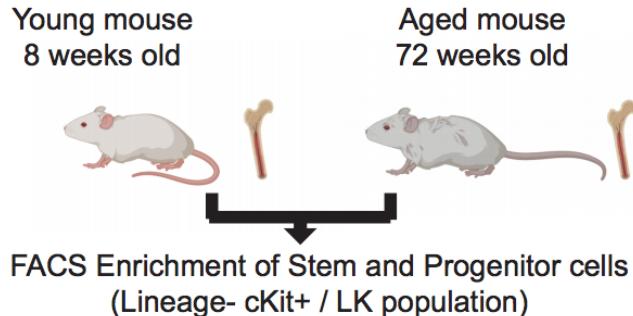


# AGED MICE SHOW MORE DIVERSE V(D)J RECOMBINATION

bioRxiv  
THE PREPRINT SERVER FOR BIOLOGY

## Combined single-cell gene and isoform expression analysis in haematopoietic stem and progenitor cells

by Laura Mincarelli, Vladimir Uzun, Stuart A. Rushworth, Wilfried Haerty, Iain C. Macaulay  
doi: <https://doi.org/10.1101/2020.04.06.202747>



# CONCATENATION OF SINGLE-CELL TRANSCRIPTS

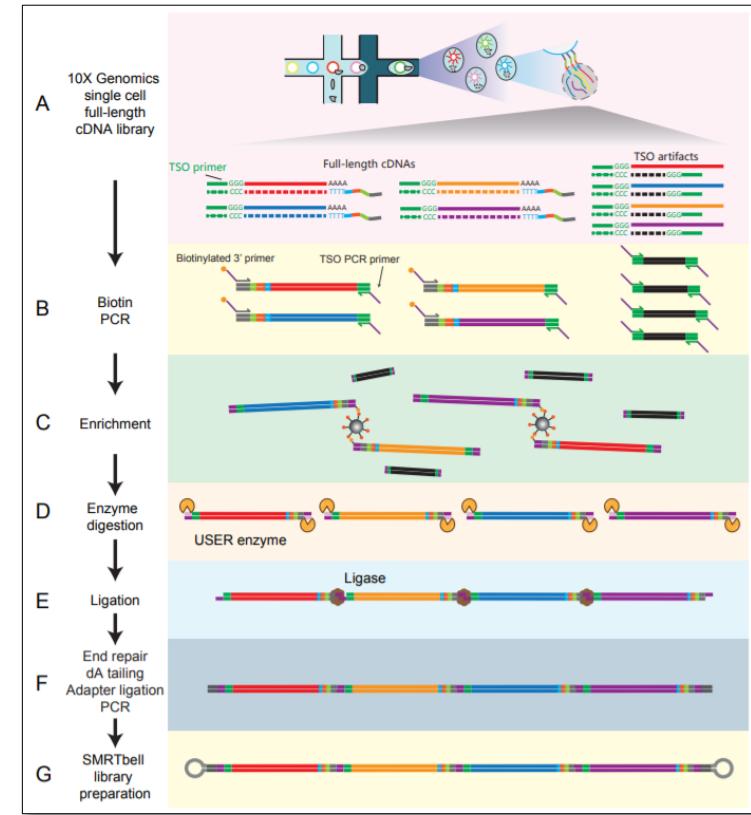
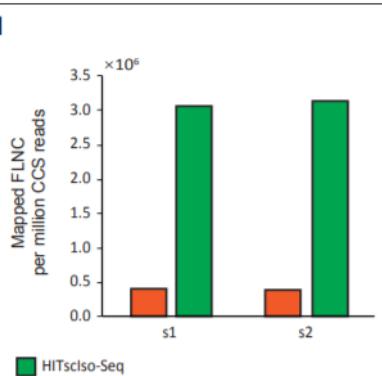
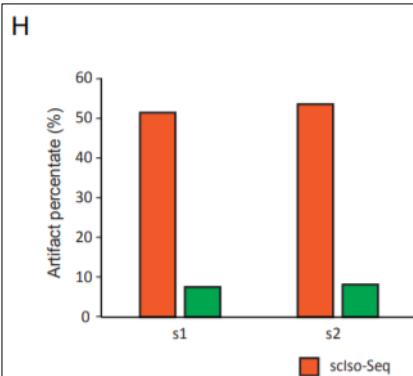
New Results

[Comment on this paper](#)

## HIT-sclIsoSeq: High-throughput and High-accuracy Single-cell Full-length Isoform Sequencing for Corneal Epithelium

Ying-Feng Zheng, Zhi-Chao Chen, Zhuo-Xing Shi, Kun-Hua Hu, Jia-Yong Zhong, Chun-Xiao Wang, Wen Shi, Ying Chen, Shang-Qian Xie, Feng Luo, Xiao-Chen Bo, Chong Tang, Yi-Zhi Liu, Chuan-Le Xiao

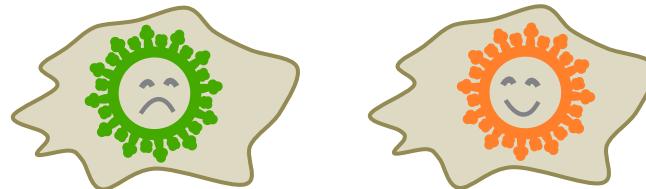
doi: <https://doi.org/10.1101/2020.07.27.222349>



<https://medium.com/@magdoll>

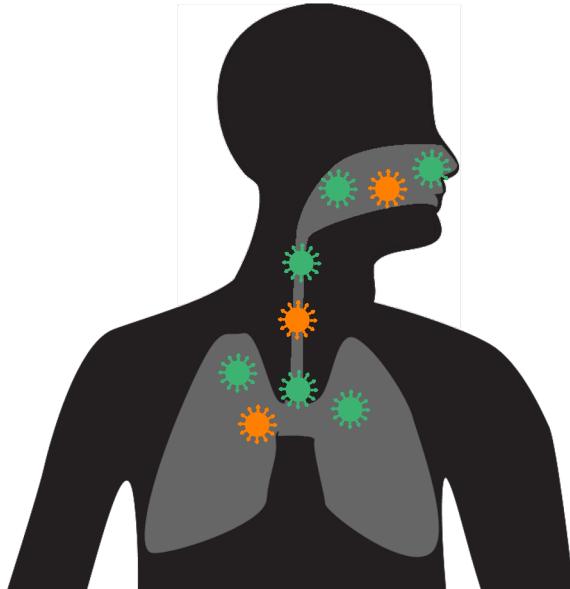


## A Tale of Mutants: Sequencing the Full-Length Influenza Virus at the Single Cell Level



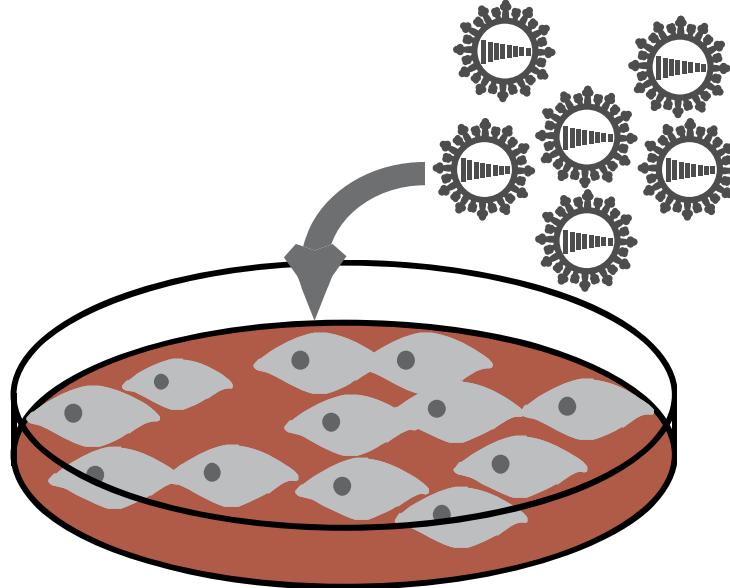
*(slides hereafter courtesy of Jesse Bloom)*

# Human influenza infections are initiated by just a few virions

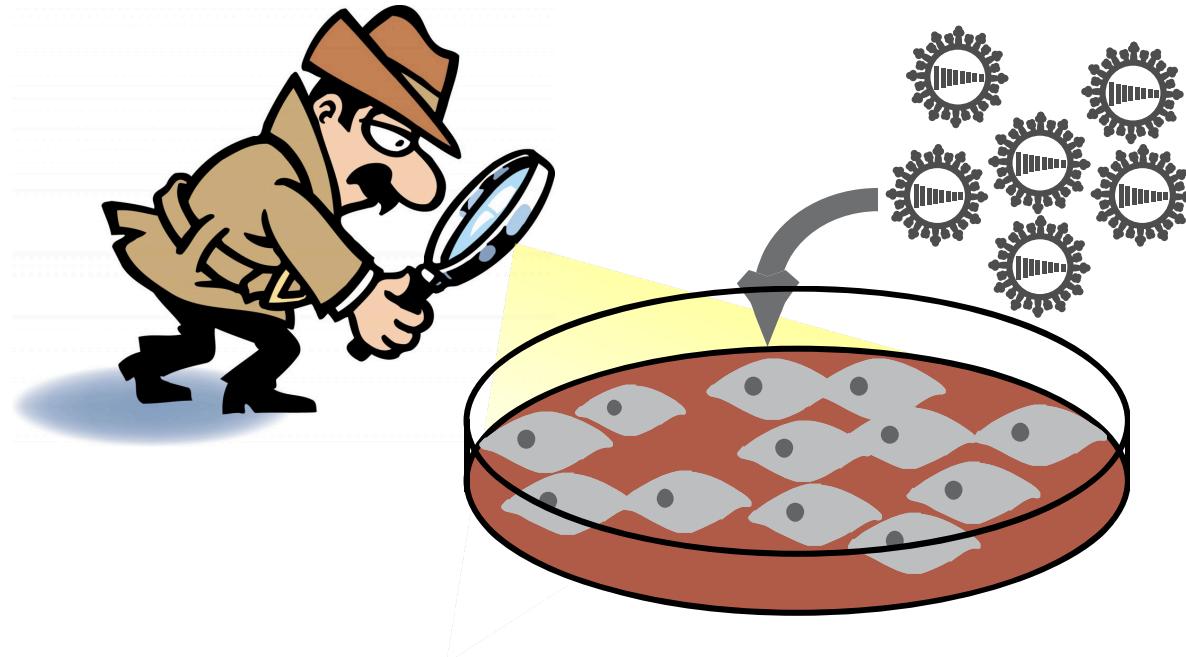


McCrone, John T., Robert J. Woods, Emily T. Martin, Ryan E. Malosh, Arnold S. Monto, and Adam S. Lauring. 2018. "Stochastic Processes Constrain the within and between Host Evolution of Influenza Virus." *eLife* 7 (May). <https://doi.org/10.7554/eLife.35962>.  
Xue, Katherine S., and Jesse D. Bloom. 2020. "Linking Influenza Virus Evolution within and between Human Hosts." *Virus Evolution* 6 (1): veaa010.

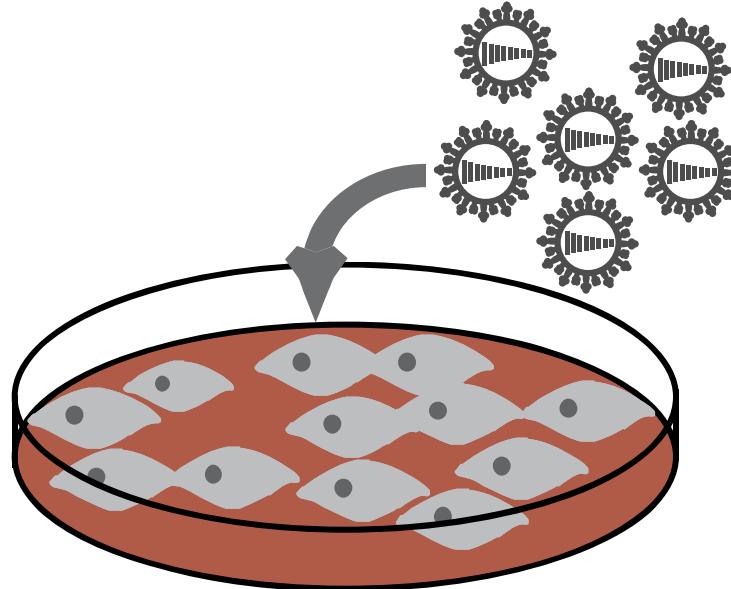
We take many cells, and infect them with many virions



We usually **average** over this entire process to study infection



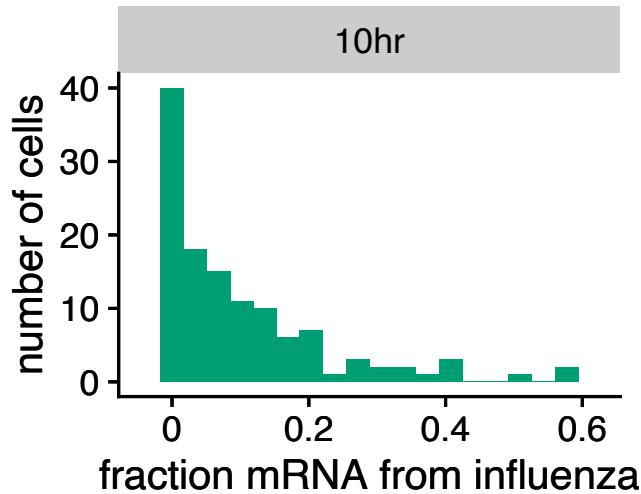
We infect A549 cells at low multiplicity of infection (MOI)



This yields a large **cell-gene matrix** that we can analyze computationally

	Cellular gene 1	Cellular gene 2	...	Cellular gene <b>19,960</b>	Cellular gene <b>19,961</b>	Viral gene 1	...	Viral gene 8
<b>Cell 1</b>	23	1	...	120	0	0	...	0
<b>Cell 2</b>	17	0	...	88	3	967	...	588
...	...	...	...	...	...	...	...	...
<b>Cell 9,999</b>	35	2	...	159	1	32	...	16
<b>Cell 10,000</b>	22	0	...	102	0	0	...	0

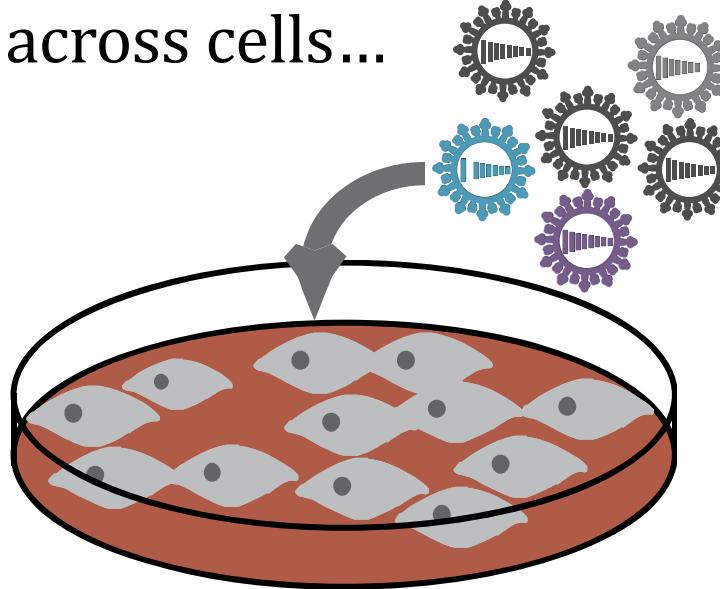
# Extreme variation across single cells



I said that we used a stock of "wildtype" virus

Actually, ***all*** viral stocks contain mutants

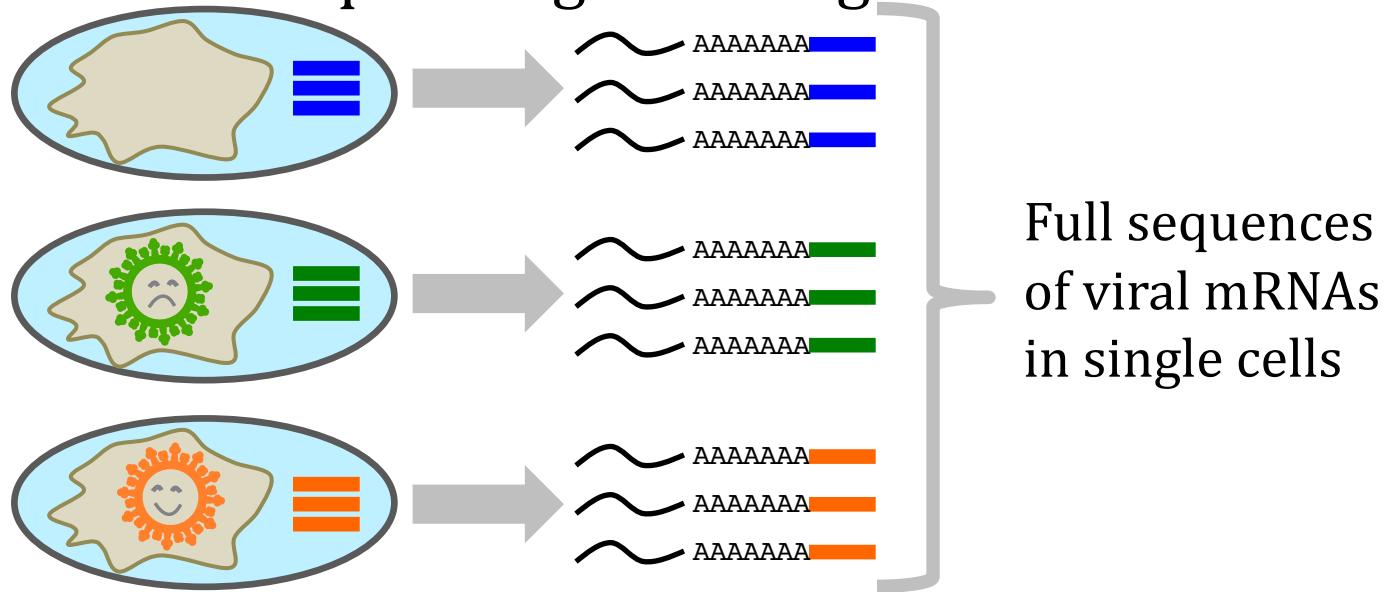
Maybe mutations in the virions explain variation across cells...



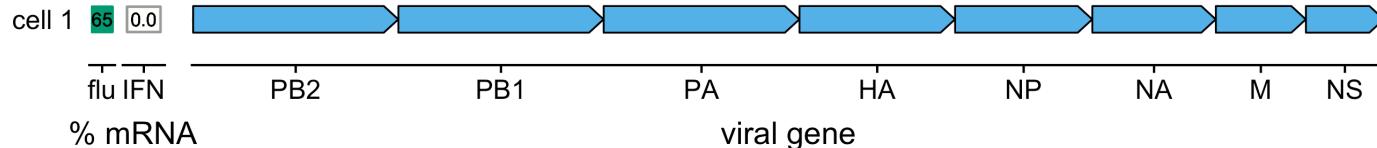
# Single-cell transcriptomics counts mRNAs, it doesn't tell us if they have mutations

	Cellular gene 1	Cellular gene 2	...	Cellular gene <b>19,960</b>	Cellular gene <b>19,961</b>	Viral gene 1	...	Viral gene 8
<b>Cell 1</b>	23	1	...	120	0	0	...	0
<b>Cell 2</b>	17	0	...	88	3	967	...	588
...	...	...	...	...	...	...	...	...
<b>Cell 9,999</b>	35	2	...	159	1	32	...	16
<b>Cell 10,000</b>	22	0	...	102	0	0	...	0

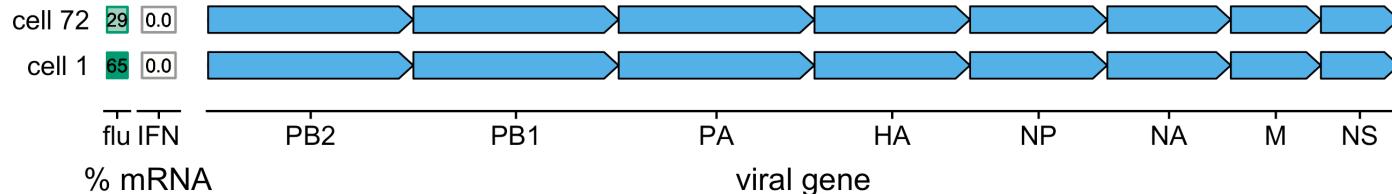
We start with the same process as for  
single-cell transcriptomics  
But we also perform full-length PacBio  
sequencing on viral genes



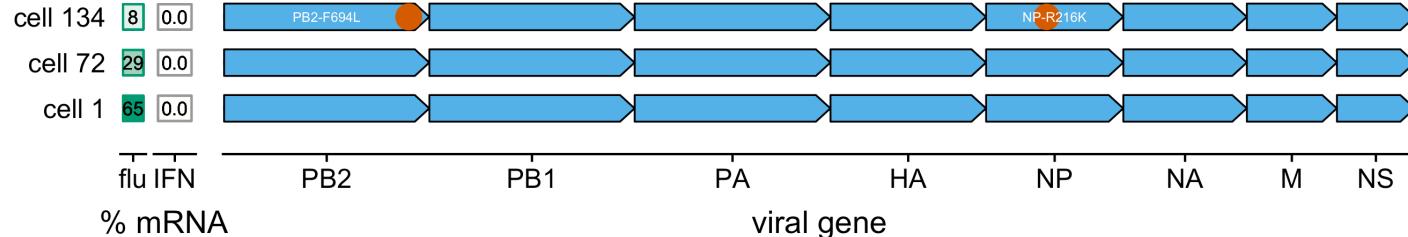
# Cells infected by wildtype virions often produce lots of viral mRNA



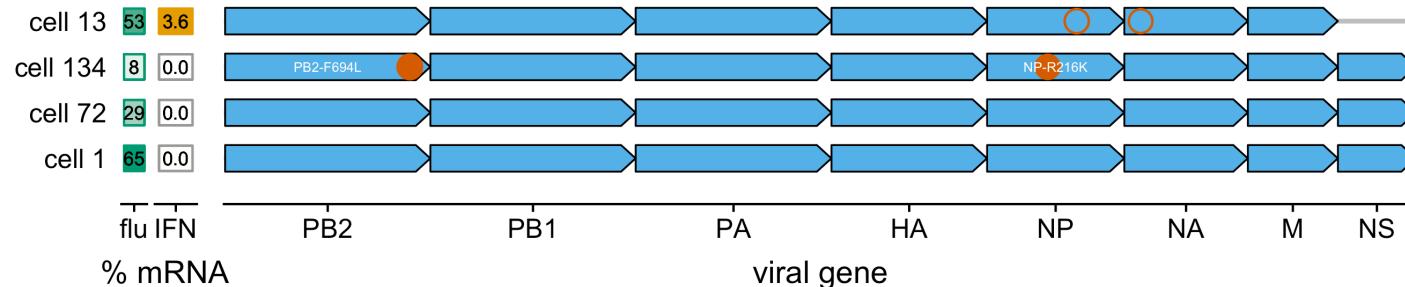
Although some wildtype virions produce less viral mRNA than others



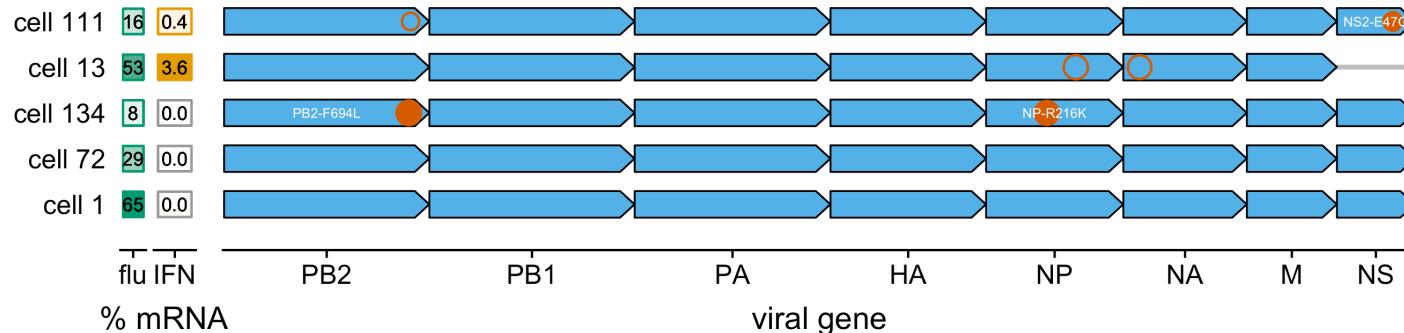
# Virions with mutations sometimes produce little viral mRNA



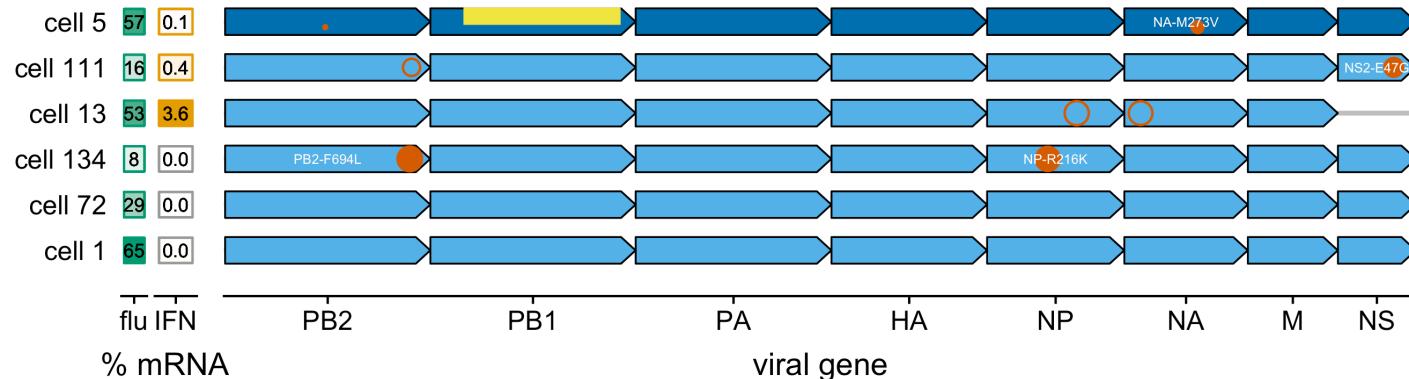
# Virions with defects sometimes produce IFN: fails to express NS



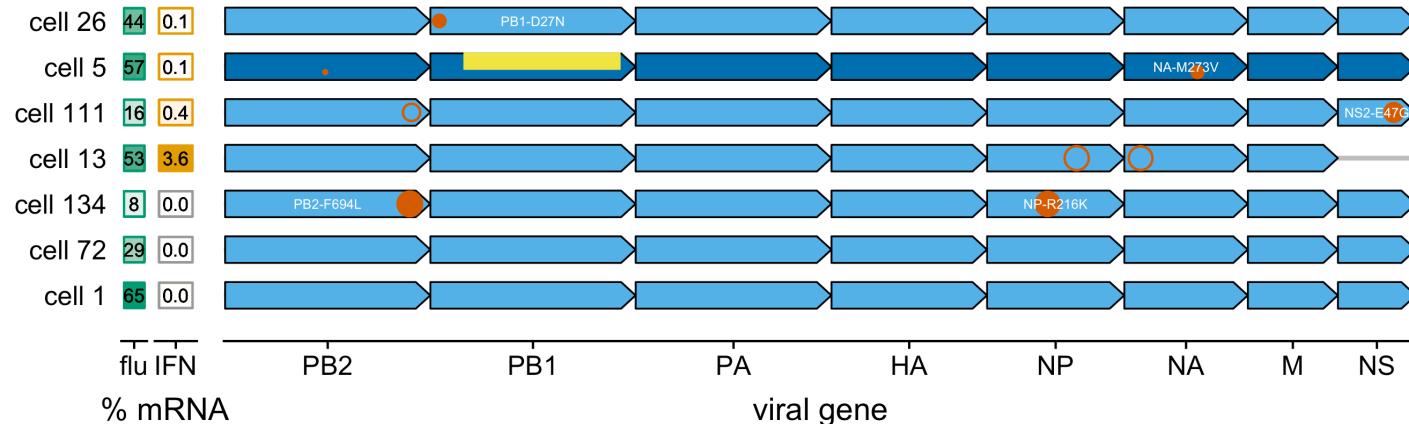
# Virions with defects sometimes produce IFN: point mutation in NS



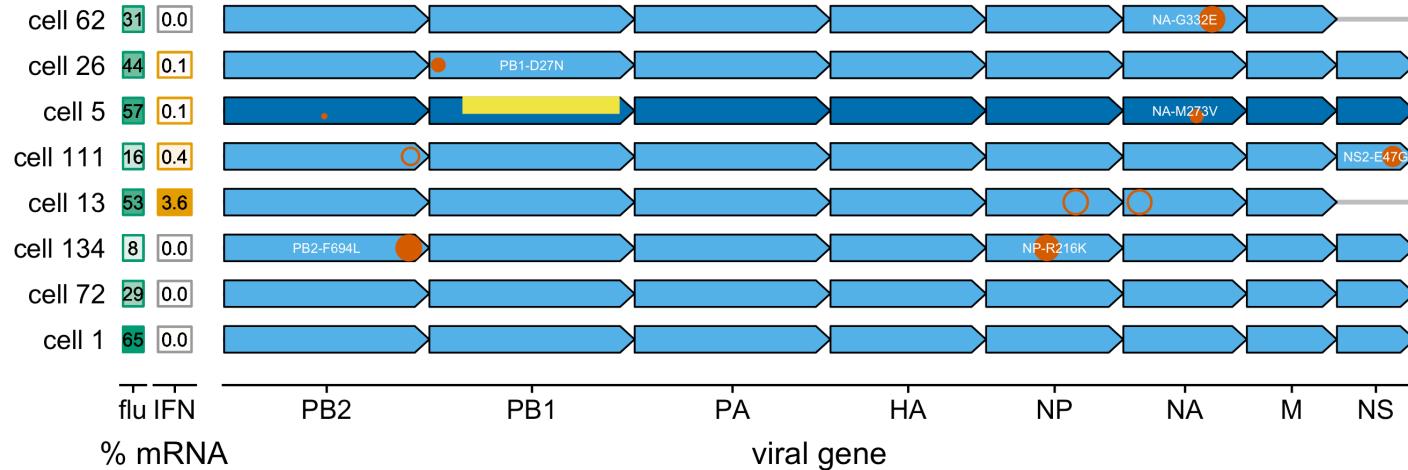
# Virions with defects sometimes produce IFN: internal deletion in PB1



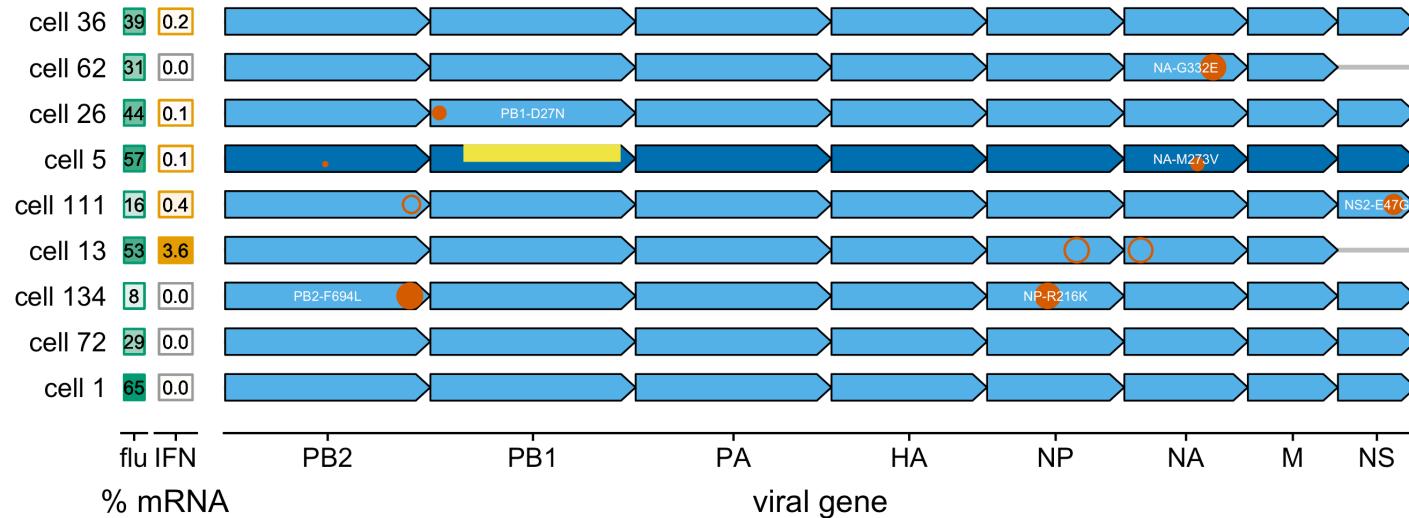
# Virions with defects sometimes produce IFN: point mutation in PB1



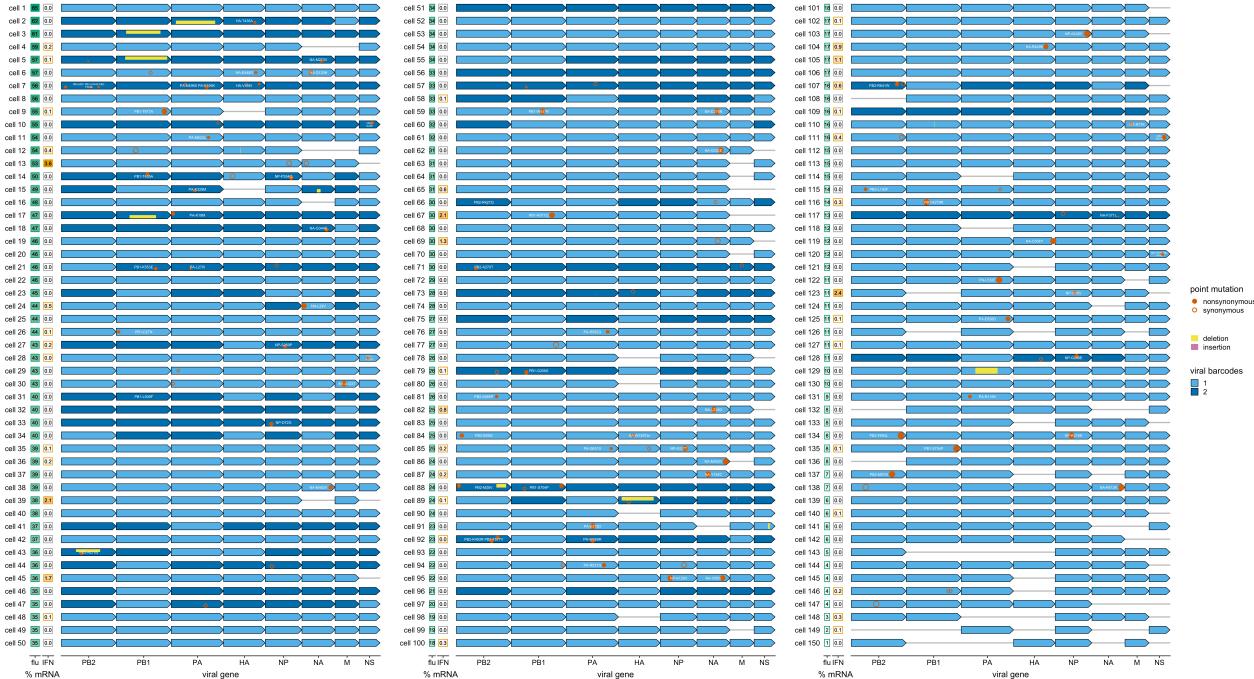
# But even virions lacking NS do not always induce IFN



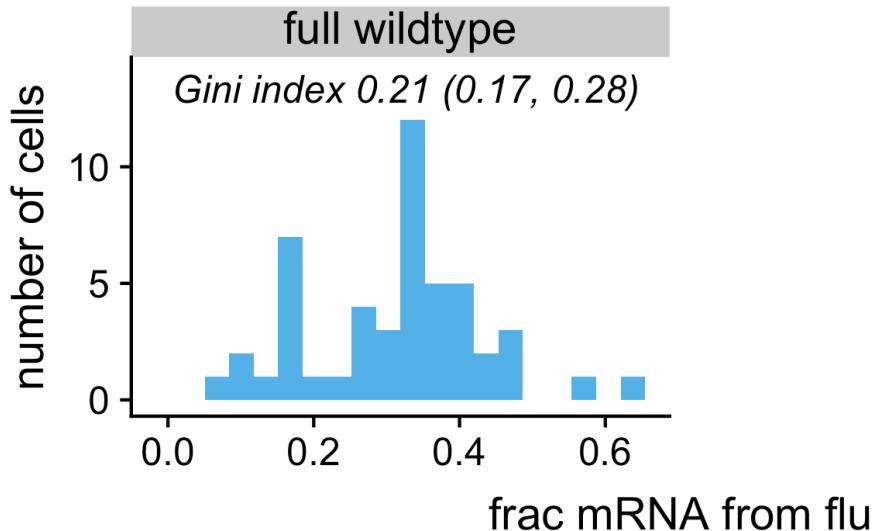
# And sometimes wildtype virions induce IFN



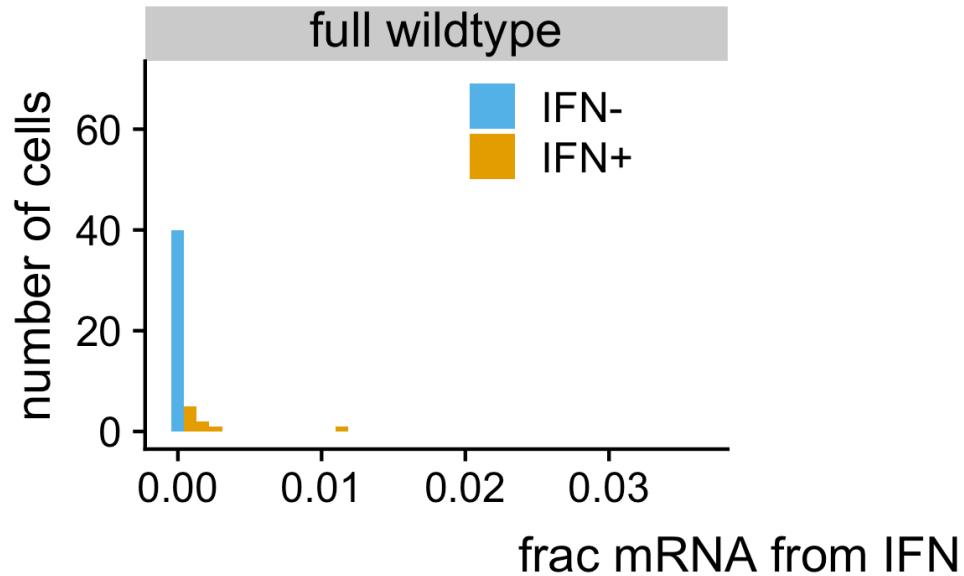
# 150 cells infected, only 49 by wildtype virions



Infections by wildtype virions are less heterogeneous than ones by mutant ones



# Infections by mutant virions induce more IFN



# SINGLE-CELL VIRAL SEQUENCING

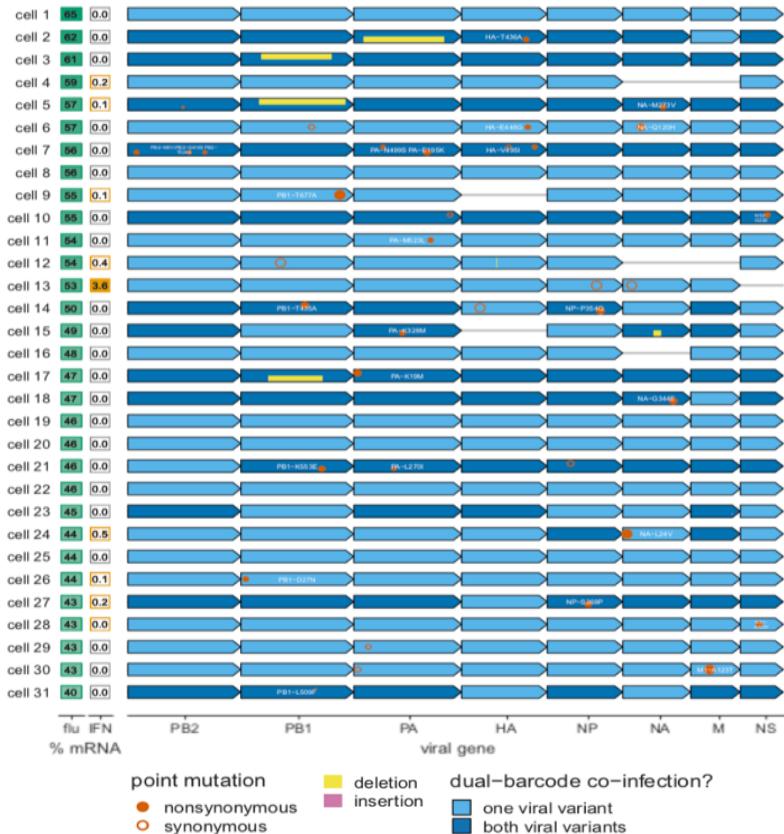
- Single-cell sequencing of H1N1 influenza
- Full-length viral transcripts reveal cell-to-cell variation on mutational landscape
- 10hr post-infection, only 49 of 150 infected cells remain wild type
- Mutations linked to differences in viral load and the innate immune response

Liz T in PacBio  
Jun 17, 2019 · 6 min read

<https://medium.com/@magdoll>



A Tale of Mutants: Sequencing the Full-Length Influenza Virus at the Single Cell Level





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# Single-Cell Iso-Seq Bioinformatics

# SINGLE-CELL ISO-SEQ BIOINFORMATICS WORKFLOW

HiFi reads



# SINGLE-CELL ISO-SEQ BIOINFORMATICS WORKFLOW

HiFi reads



Remove cDNA primers



Extract UMI and BC

transcript

# SINGLE-CELL ISO-SEQ BIOINFORMATICS WORKFLOW

HiFi reads

Remove cDNA primers

Extract UMI and BC

Cluster by (UMI,BC)

Classify Transcripts

Remove Artifacts

<b>id</b>	<b>length</b>	<b>UMI</b>	<b>BC</b>	<b>count</b>
molecule/34465	833	ATGGTATA	GAACGTGGTCGA	1
molecule/34597	807	GTACGATG	CAGGATTAAGAG	3
molecule/34598	807	ACCTCCTG	GACCAGACTGGA	1
molecule/34638	807	AAAGTCAC	TATATGGTAGGT	2

# SINGLE-CELL ISO-SEQ BIOINFORMATICS WORKFLOW

HiFi reads

Remove cDNA primers

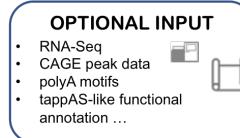
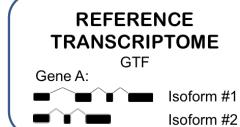
Extract UMI and BC

Cluster by UMI

Classify Transcripts

Remove Artifacts

## SQANTI3: QUALITY CONTROL OF TRANSCRIPTOMES

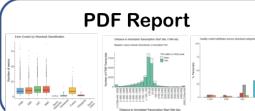


Classification file

ID	Gene	Transcript	Category
PB.1.1	Ctnnd1	ENSMUST00000067232	FSM
PB.1.2	Ctnnd1	novel	NIC
PB.2.1	Novel	novel	Intergenic

Junction file

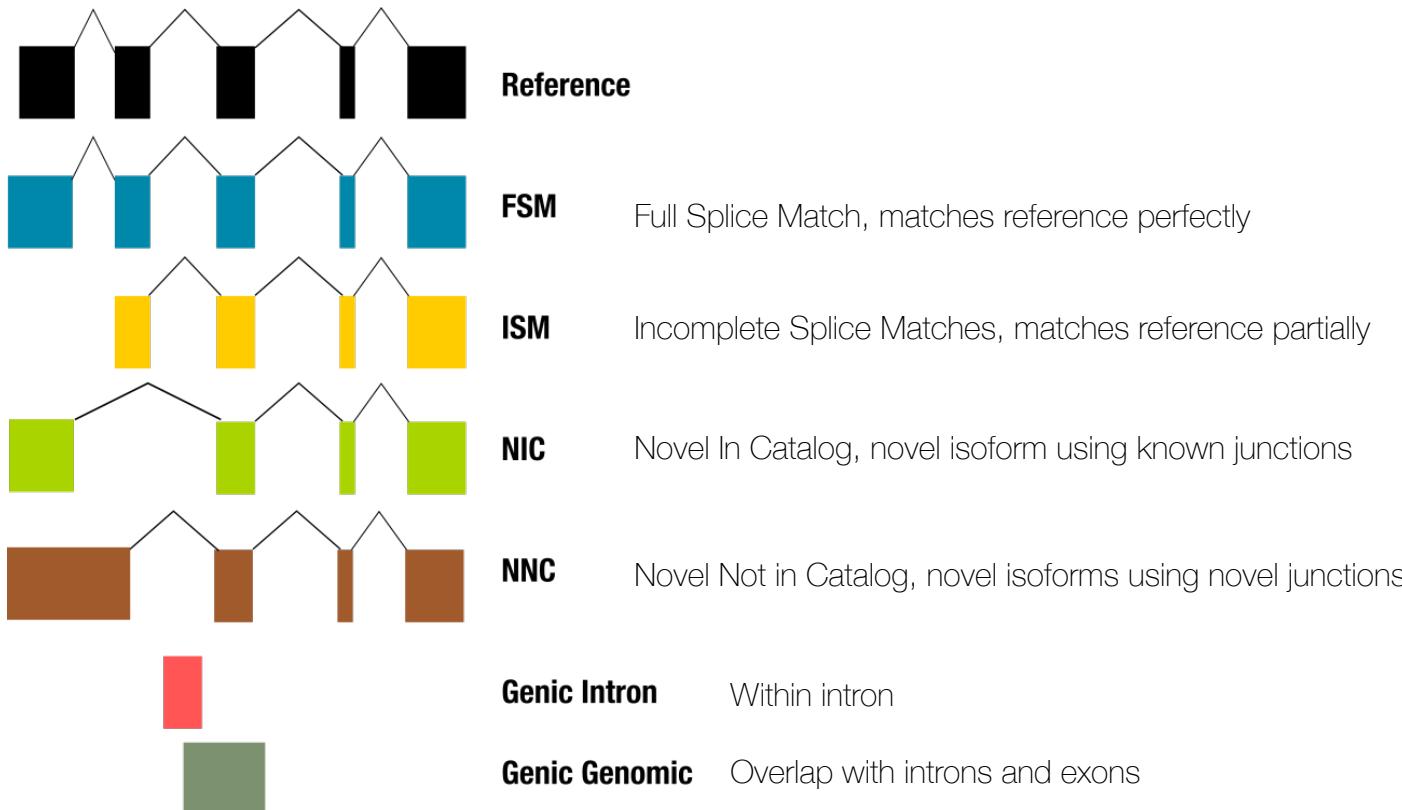
Junction	Isoform	Splice site	Known
Junction1	PB.1.2	GT-AG	True
Junction2	PB.1.2	GC-AG	True
Junction3	PB.1.2	GT-AF	False



- Complementary files
- Genome-corrected transcriptome
  - ORF prediction (FASTA)
  - CDS-annotated GTF
  - tappAS-compatible GFF3

<https://github.com/ConesaLab/SQANTI3>

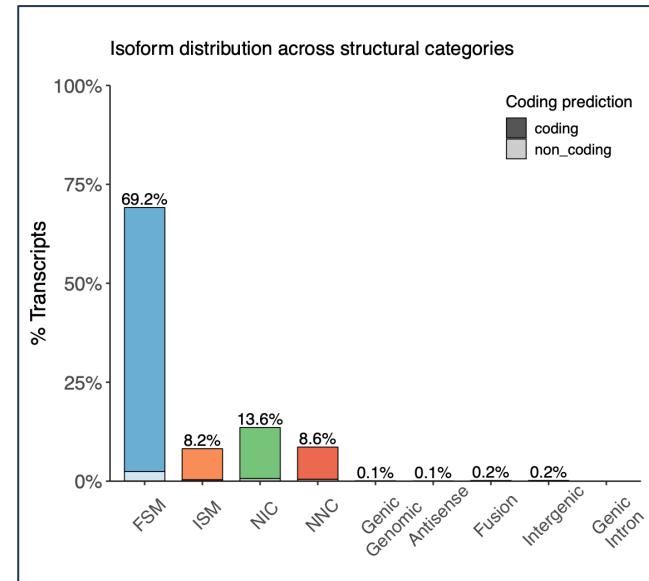
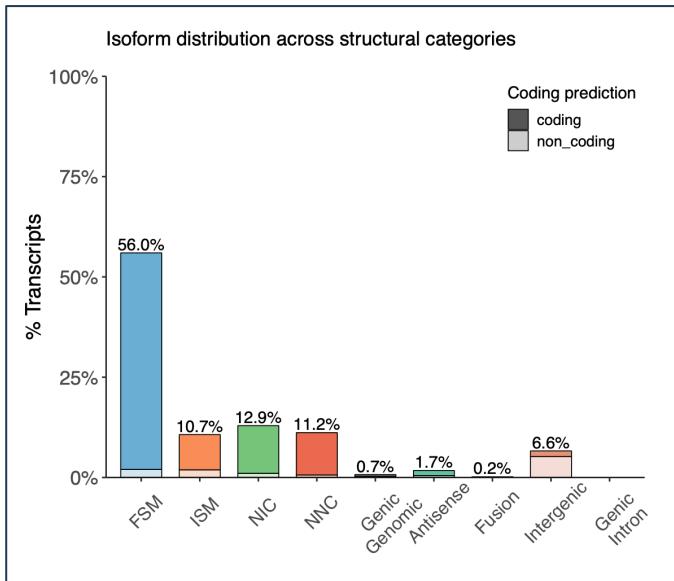
# TRANSCRIPT CLASSIFICATION BY SQANTI



# SQANTI3: BEFORE AND AFTER FILTERING

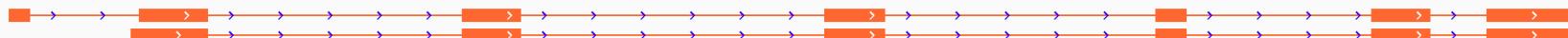
After SQANTI3 filtering, percentage of FSMs increase

Most filtered transcripts are genic, genomic, or intergenic



# SINGLE-CELL ISO-SEQ BIOINFORMATICS WORKFLOW

<b>id</b>	<b>length</b>	<b>transcript</b>	<b>gene</b>	<b>category</b>	<b>UMI</b>	<b>BC</b>
molecule/34465	833	ENST00000392221.5	ENSG00000114942.14	full-splice_match	ATGGTATA	GAACGTGGTCGA
molecule/34597	807	ENST00000392222.7	ENSG00000114942.14	full-splice_match	GTACGATG	CAGGATTAAGAG
molecule/34598	807	ENST00000392222.7	ENSG00000114942.14	full-splice_match	ACCTCCTG	GACCAGACTGGA
molecule/34638	807	ENST00000392222.7	ENSG00000114942.14	full-splice_match	AAAGTCAC	TATATGGTAGGT



Cell:	1	2	...	N
Transcript 1	10	1		5
Transcript 2	8	6		0
.	.	.		.
.	.	.		.
Transcript M	3	0		12



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