Q-TRAY MEDIA

1. Prepare LB media of 200ml aliquot in 500 milliliter bottles/flask with 1.5% agar. (8g LB/Agar plus 200ml water)- shake well.
2. Autoclave and shake them right after the autoclaving. Be careful not to make bubbles.
3. Cool down to 50 C (cool until warm to the touch- do not cool too much, but be sure that it is cool enough to add antibiotic).
4. Add ingredients required and mix well without getting any **bubbles**!!!

	Stock	Final conc	Per 200ml
IPTG	200 mg/ml	90 ug/ml	90 ul
X-Gal	20 mg/ml	90 ug/ml	0.9 ml
Carbenicillin	50 mg/ml	100 ug/ml	400 ul
Ampicillin	100 mg/ml	150 ug/ml	300 ul
Chloramphenicol(Plasmid)	50 mg/ml	30 ug/ml	120 ul
Chloramphenicol(BAC)	50 mg/ml	25 ug/ml	100 ul

5. Pour 200 ml per Q-Tray on a **"perfect-leveled surface"** in a laminar flow hood.
6. Remove bubbles, if any, with either pipette tip or burned needle.
7. Leave lids off until solidified to prevent condensation on lids. It will take about 30-60 minutes
8. Keep media out of direct light to prevent degradation of X-gal, if used .
9. Wrap with Saran wrap and keep it in 4C until you need.

* The medium surface should be smooth and flat.

** Proper titration is 3,000 colonies/Q-tray