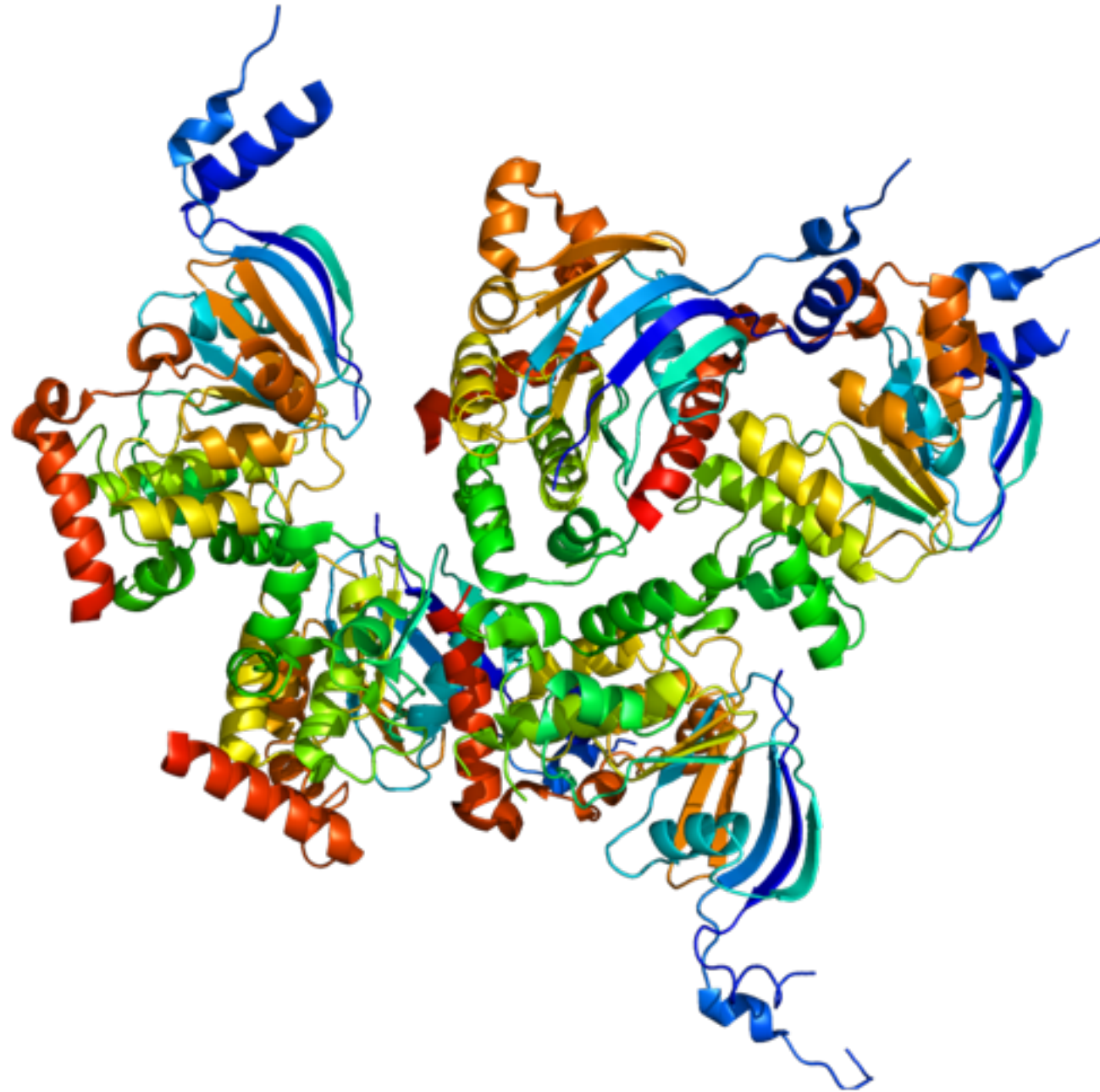


# CFTR



(Cystic Fibrosis Transmembrane Conductance  
Regulator)

# What is this thing?

- CFTR is a transmembrane channel protein. It exists in all jawed vertebrates, and modulates the passage of chloride [3] and thiocyanate ions [1] through phospholipid bilayers.
- Mutations in CFTR are responsible for the etiology of cystic fibrosis. There are ~2000 documented mutations, but most belong to a general subset of CFTR dysfunctions.
- Cystic Fibrosis (CF) is an autosomal recessive disease characterized by an overproduction of mucus in all of an afflicted individual's mucosa, causing male sterility, pancreatic/cardiovascular/digestive dysfunction, pulmonary infection and failure.
- Interesting as both an etiological factor and a model for understanding autosomal disorders.

# Its History

- Much like other homozygous diseases, there exists a set of evolutionary explanations for CFTR mutations.
- CFTR discovered and characterized 1989 by Messrs. Riordan, Rommens, Kerem, et al. by means of cDNA expression measurement in epithelial cells of CF patients. They discover the most common mutation,  $\Delta F508$ . [3]
- Better methodologies and increased interest spur the characterization of hundreds of other mutant CFTR.
- The natural history: CFTR appears in aquatic vertebrates (exact time uncertain) for osmoregulation, and take a secondary role in organ development of epithelial systems. The gene propagated to the other chordates. The phylogeny illuminates function. [1]

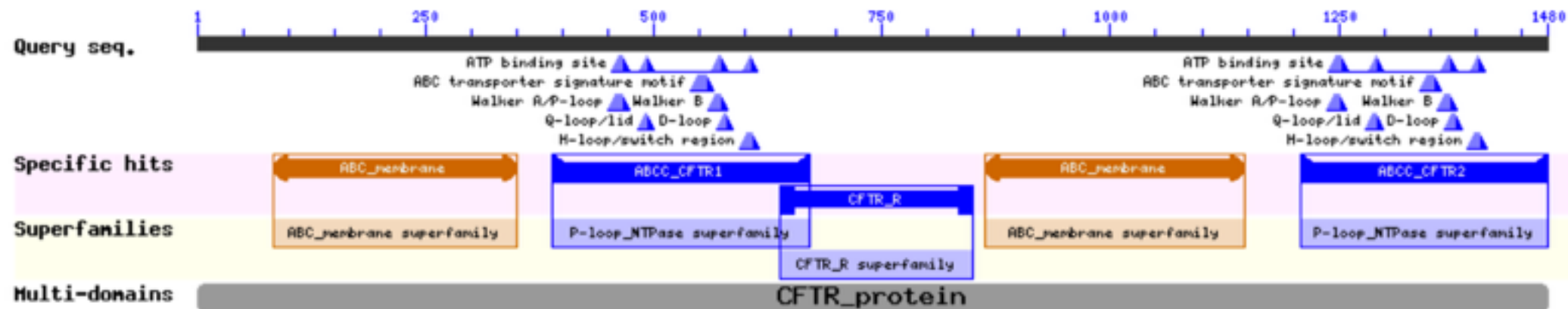
# $\Delta F508$

- $\Delta F508$  is a CFTR mutation characterized by a deletion of a single phenylalanine residue at position 508.
- Heterozygosity may be evolutionarily advantageous—cholera toxin, diarrhea, asthma hypotheses.
  - ...but homozygosity causes CF and is fatal.
  - And the data suggest that negative selection may also explain the prevalence of the mutation. [2]
- Single residue deletion in transmembrane region, mutant-type CFTR cannot be exported from endoplasmic reticulum to epithelial cell membranes.

# The Interesting Parts

- CFTR [https://www.youtube.com/watch?v=\\_j99-xgOlaw](https://www.youtube.com/watch?v=_j99-xgOlaw)
- Structure: Two transmembrane domains, two nucleotide binding domains (ATP), a regulatory site.
- Mutation types:
  - Splice error: Most protein produced is nonfunctional and cannot embed into ER for trafficking to apical membrane.
  - Premature stop codon: Truncated, nonfunctional protein.
  - Trafficking defect:  $\Delta F508$  and others. Protein misfolds, never leaves ER.
  - Gating defect & narrow chloride channel:  $\text{Cl}^-$  cannot pass through the protein as in wild-type
  - Decreased stability: Proteins don't stay in the cell membrane as long as wild-type, and are degraded.
- Net effect is osmotic dysregulation: hyperabsorption of some ions in some tissues, underabsorption of some ions in others (depending upon how CFTR functions in the cell)

# The Families & Their Structures



- ABC Transporter Superfamily & membrane superfamily, also its own personal superfamily
- cAMP-dependent Chloride Channel (NB: the regulatory site)
- ATP binding are well-conserved, ABC transporters have a signature motif
- <http://www.ncbi.nlm.nih.gov/protein/P13569.3>

# Why Are The Interesting Parts Interesting?

- Development of novel therapies.
  - Potentiators
    - A class of pharmaceuticals modifies gate defects CFTR such that the protein no longer requires ATP to mediate flux. Improves pancreatic function in human models. [6]
  - CRISPR-Cas9
    - Homozygous hereditary illnesses are sensitive to gene therapies.
    - CRISPR-Cas9 + HIV-derived factors + small hairpin RNA can knockdown CFTR mutant expression in airway cells. [5]

# Question Time

- Say we use a CRISPR-Cas9 knockdown therapy in individuals with CF, and use a CRISPR-Cas9 therapy inducing expression of wild-type CFTR. Would lung function return to normal?
  - Animal models (sorry)
- Would organismal knockdown of the trait adversely effect downstream systemic functionality?
- Can CF CRISPR-Cas9 gene therapeutic model be abstracted to the other autosomal diseases? (Huntington's, Tay-Sachs, etc.)



# References.

1. Exploiting species differences to understand the CFTR Cl<sup>-</sup> channel. Samuel J. Bose, Toby S. Scott-Ward, Zhiwei Cai, David N. Sheppard Biochemical Society Transactions Oct 09, 2015, 43 (5) 975-982; DOI: 10.1042/BST20150129
2. CARSTEN WIUF (2001). Do  $\Delta F508$  heterozygotes have a selective advantage?. Genetical Research, 78, pp 41-47. doi:10.1017/S0016672301005195.
3. A new model of cystic fibrosis pathology: Lack of transport of glutathione and its thiocyanate conjugates. Melanie Childers, , George Eckel, Alan Himmel, Jim Caldwell. doi:10.1016/j.mehy.2006.06.020
4. Riordan JR, Rommens JM, Kerem B, Alon N, Rozmahel R, Grzelczak Z, Zielenski J, Lok S, Plavsic N, Chou JL (1989). "Identification of the cystic fibrosis gene: cloning and characterization of complementary DNA". Science 245 (4922): 1066–73.
5. Curr Gene Ther. 2015;15(5):447-59. CFTR inactivation by lentiviral vector-mediated RNA interference and CRISPR-Cas9 genome editing in human airway epithelial cells. PMID: 26264708
6. CFTR potentiator therapy ameliorates impaired insulin secretion in CF patients with a gating mutation. Reuven Tsabaria, Hila Iron Elyashara, Malena Cohen Cymberknowha, Oded Breuera, Shoshana Armonia, Galit Livnatc, Eitan Kerema, David Haim Zangen. J Cyst Fibros. 2015 Nov 4. pii: S1569-1993(15)00255-6. doi: 10.1016/j.jcf.2015.10.012. PMID: 26547591
7. NCBI Protein, Nucleotide, Blast