

# Introduction to variant calling

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Norway

BIOS-IN 5K/9K  
24<sup>th</sup> of Oct 2022

@archaeogenomics 



UiO • University of Oslo



# Evolutionary Biologist

specialize in ancient DNA

Archaeogenomics group  
(10+ MSc, PhDs & Postdocs)

@archaeogenomics



Multidisciplinary research:  
Archaeology

Biology  
Ecology

**Molecular methods/sequencing  
Genomics  
Bioinformatics**

Today:

- 1) Introduction: variant calling, why do we want to do this, and what it is?

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- 2) Variant calling pipelines/methods and limitations

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- 1) Introduction: variant calling, why do we want to do this, and what it is?
- 2) Variant calling pipelines/methods and limitations
- 3) Practical session, going through (parts of) a SNP calling pipeline and interpret biological results

# Introduction

Genetic variation (genomic differences between individuals) is everywhere



# Genetic variation at different scales:

- 1) Biological differences (phenotypes) between species



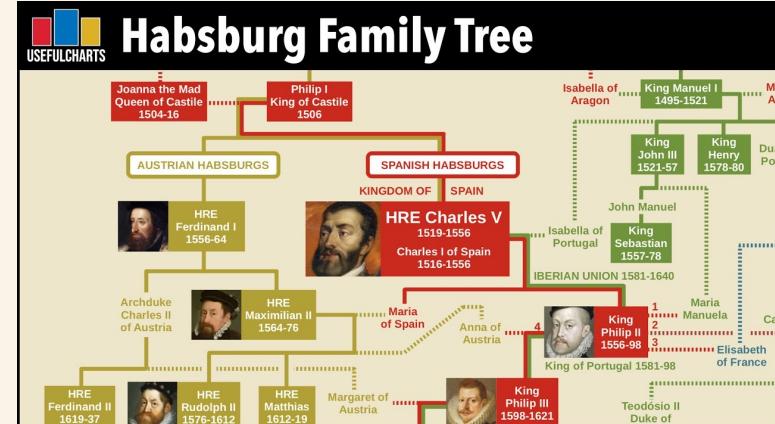
# Genetic variation at different scales :

- 1) Biological differences (phenotypes) between species
- 2) Biological differences within species



## Genetic variation at different scales :

- 1) Biological differences (phenotypes) between species
  - 2) Biological differences within species
  - 3) Patterns of relatedness between individuals/ populations (23 and me)



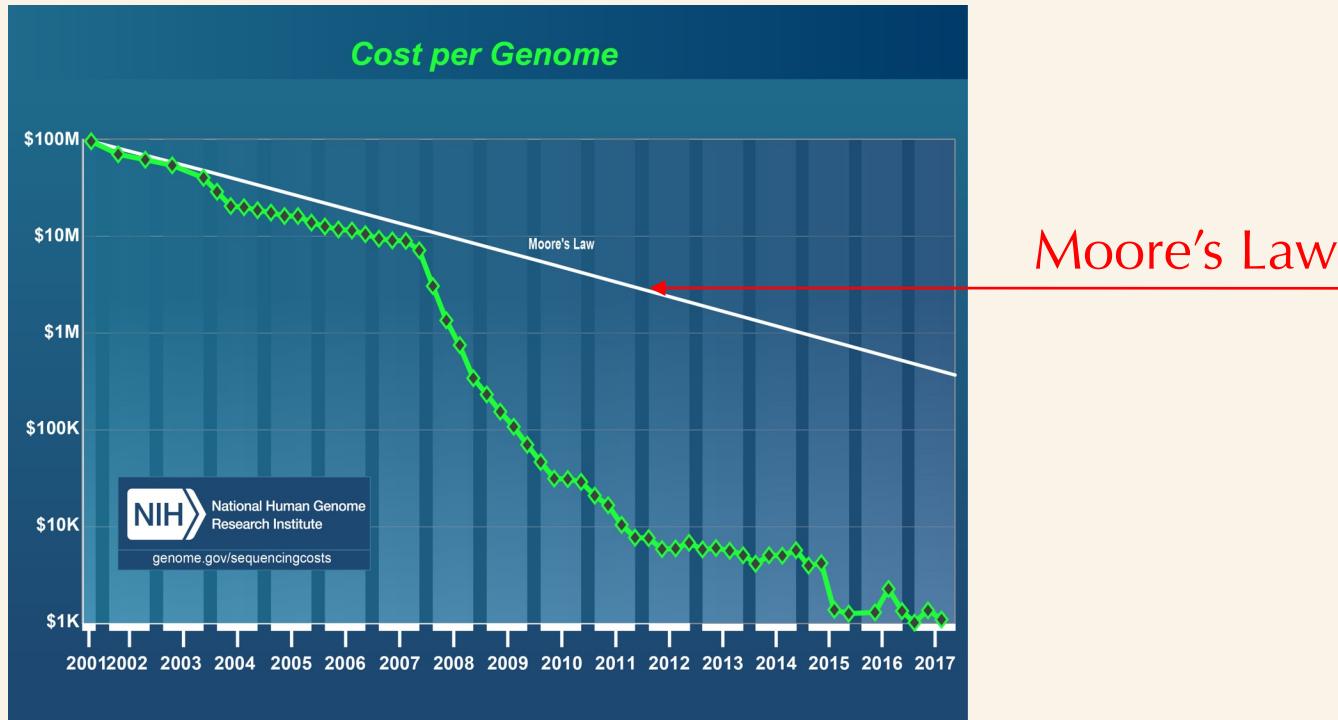
Genetic variation explains many observations within biology

Genetic variation explains many observations within biology

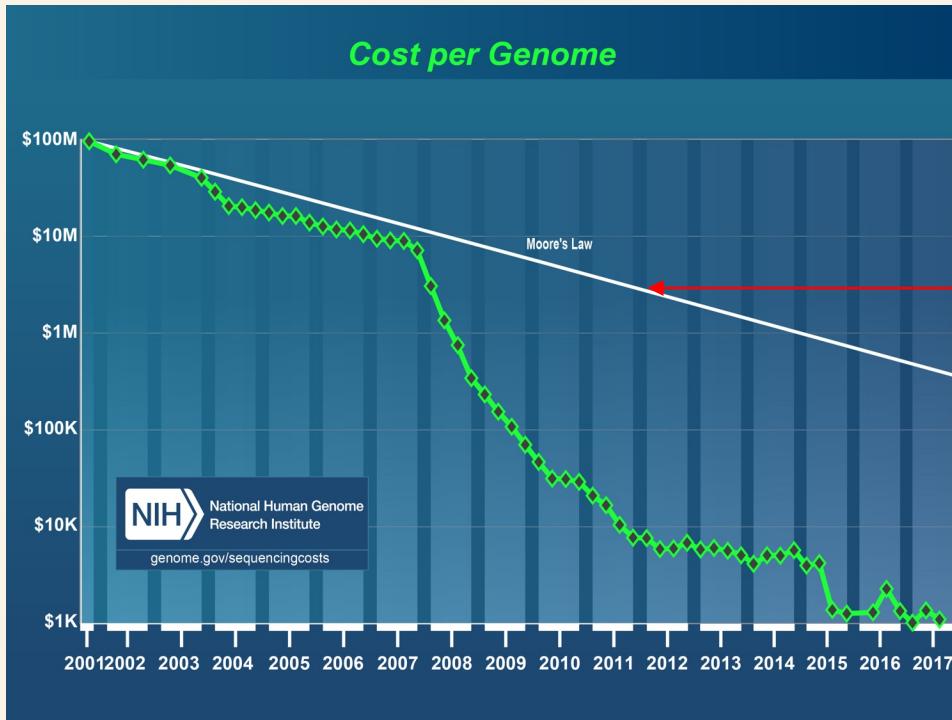
*Knowing patterns of/quantifying genetic variation has enormous potential for a wide range of applications in society*

*(e.g. personal medicine, forensic sciences, biodiversity assessments, crop improvement, animal breeding, conservation management, history & genealogy, etc etc)*

# Why are we here?



# Why are we here?

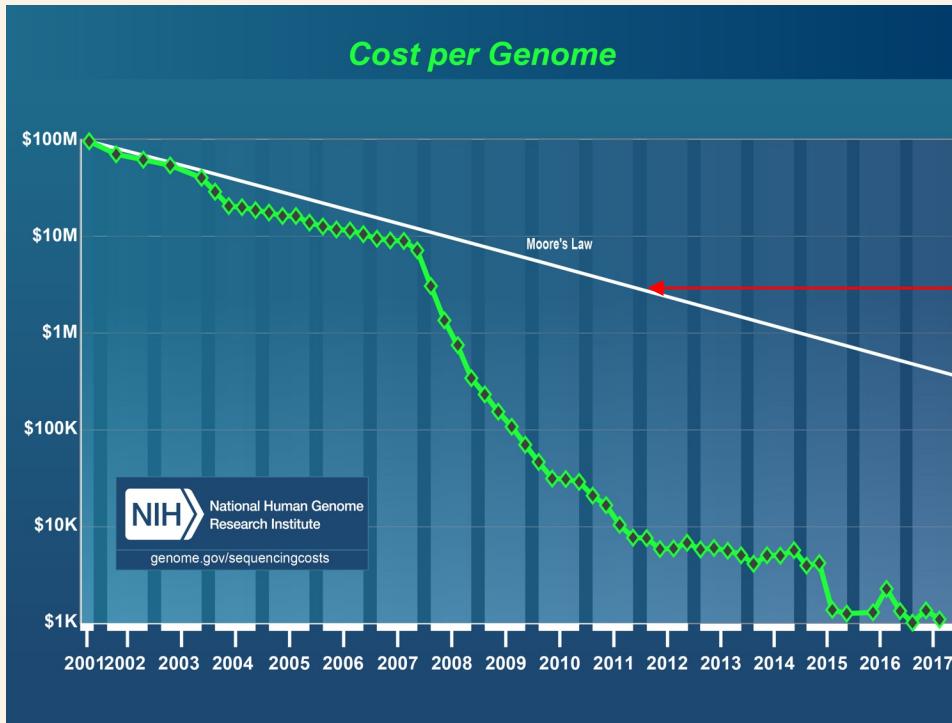


2000



2017

# Why are we here? ***Phenomenal*** technological advances



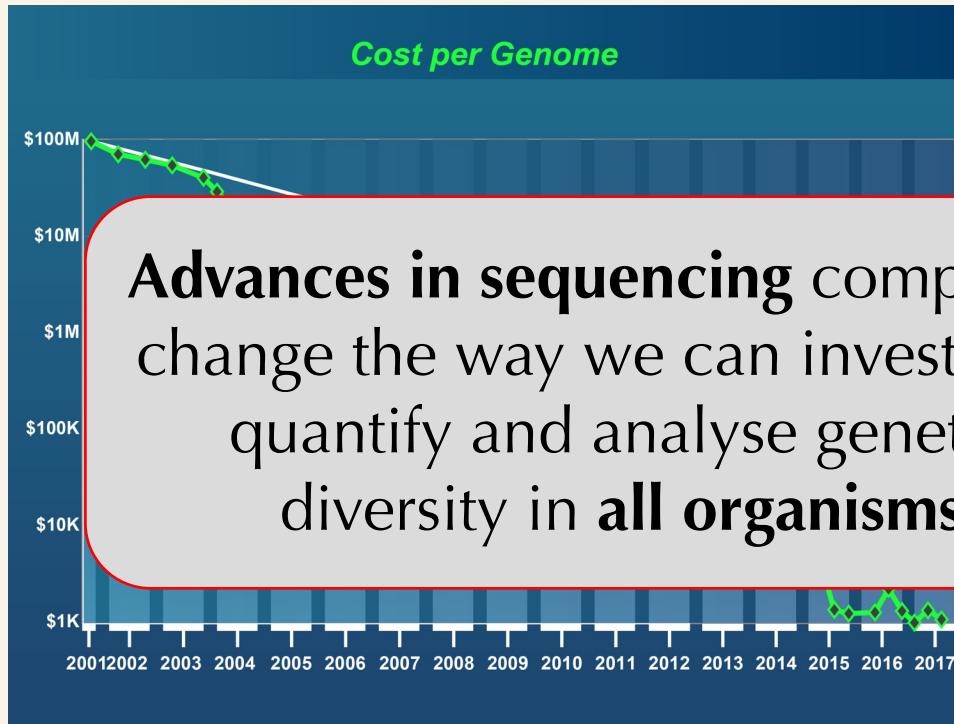
2000



2017

Technological revolution that has *fundamentally* changed the way we do biology

# Why are we here? ***Phenomenal*** technological advances



**Advances in sequencing** completely change the way we can investigate, quantify and analyse genetic diversity in **all organisms**



2017

Technological revolution that has *fundamentally* changed the way we do biology

# How has sequencing changed and is changing the world?

*Changed healthcare*

Sequencing (genome and exome) funded solely by *healthcare* systems

2012

~1%

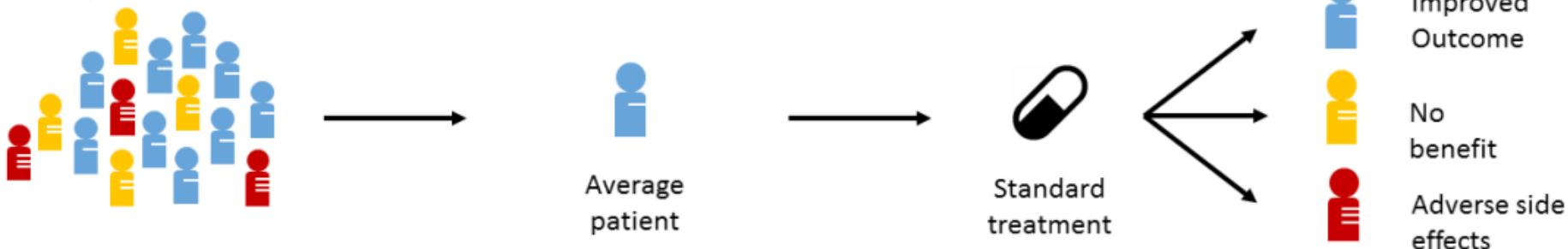
2017

~20%

2022

>80%

## Today

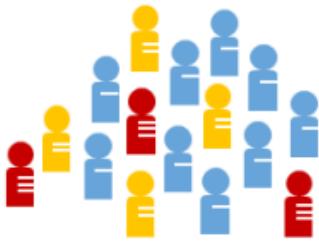


## 2030

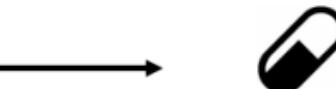
### Precision medicine



Today



Average



Standard



Improved  
Outcome



No  
benefit



Adverse side  
effects

2030

Precision medicine



Whole genome sequencing is  
actively used in Norwegian  
healthcare *and* provides clinical  
solutions



Population  
health data



Data  
analytics



Personalised  
treatment



Improved  
Outcome

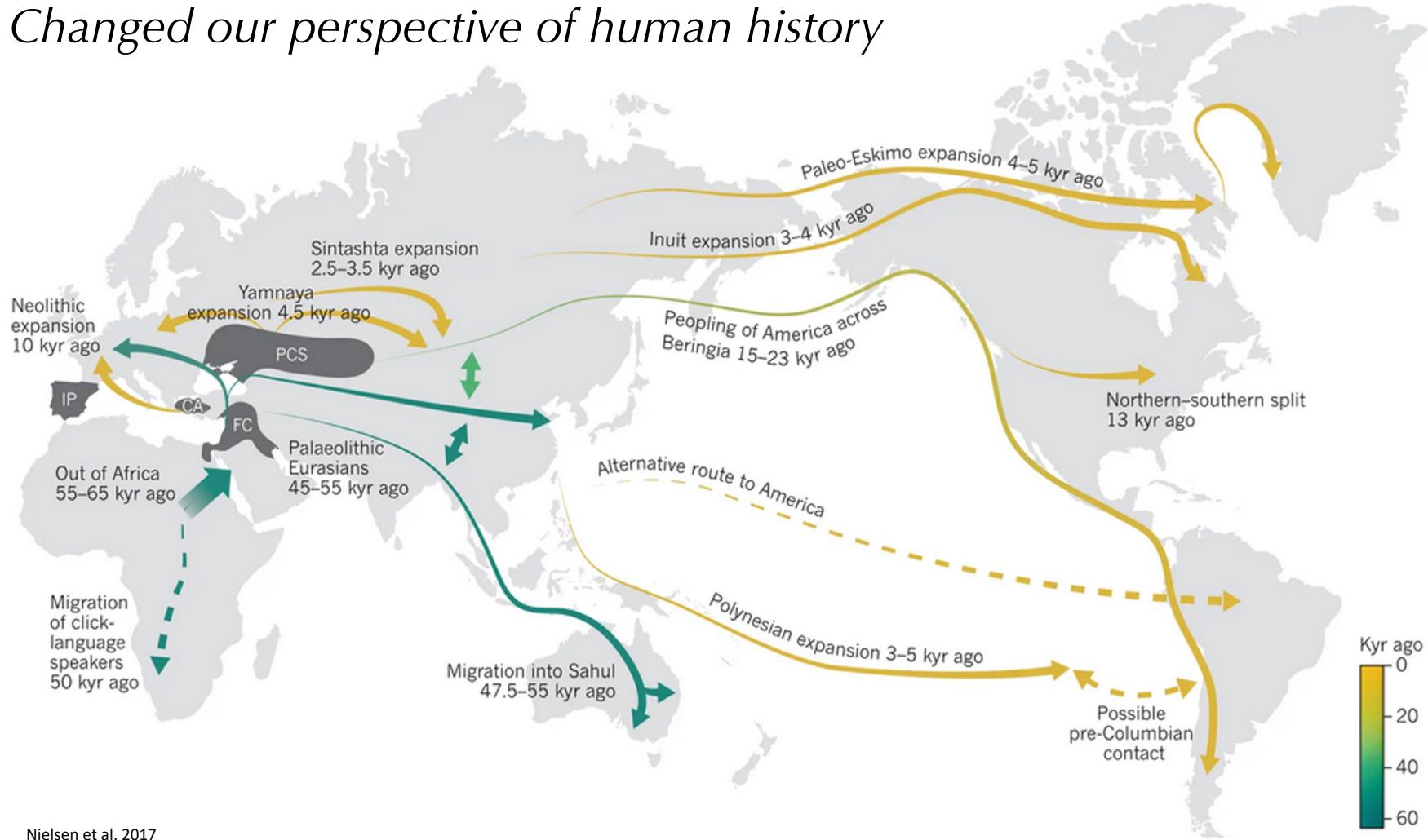


Improved  
Outcome



Reduced  
side  
effects

# Changed our perspective of human history



## *Changed forensic capabilities*

Using continuously expanding public genomic databases (e.g. 23 and me)...

The New York Times

### ***Genealogists Turn to Cousins' DNA and Family Trees to Crack Five More Cold Cases***

Police arrested a D.J. in Pennsylvania and a nurse in Washington State this week, the latest examples of the use of an open-source ancestry site since the break in the Golden State killer case.

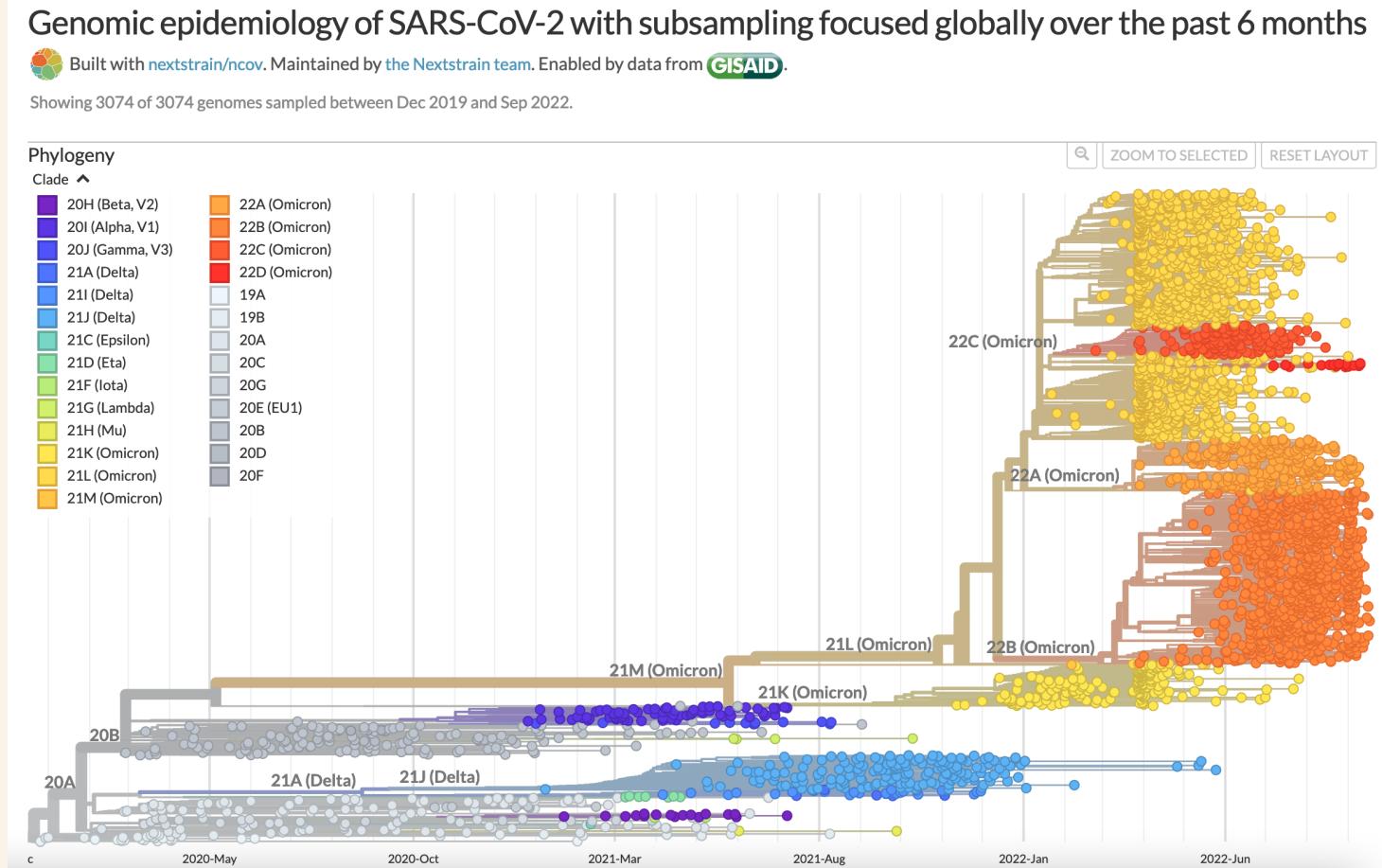
## *Changed forensic capabilities*

Or by the genetic testing of thousands of people!

As the *Times* reports, that law paved the way for a prosecutor in the Verstappen case to call for the voluntary DNA sampling of 21,500 Dutchmen, and the obligatory sampling of 1,500 men who were of “special interest” to investigators.

The alleged killer, 55-year-old Jos Brech, was one of those 1,500 men who were mandated to provide a DNA sample. He never showed up. Dutch officials grew suspicious and took DNA samples from Brech’s relatives. The results matched the DNA

# *Changed vaccine development and disease tracking*



# *Changed improvement and selection of commercial crops*

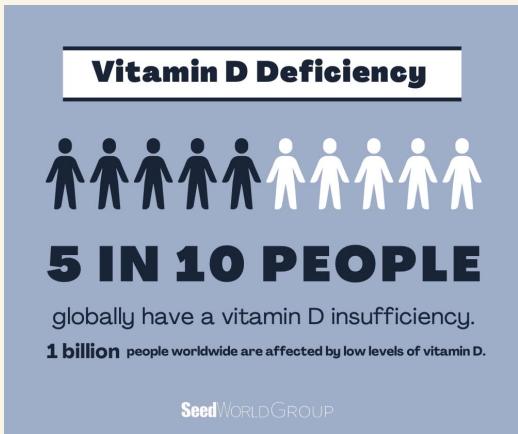
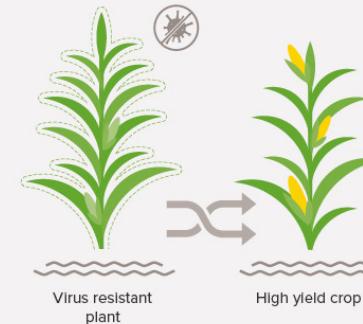
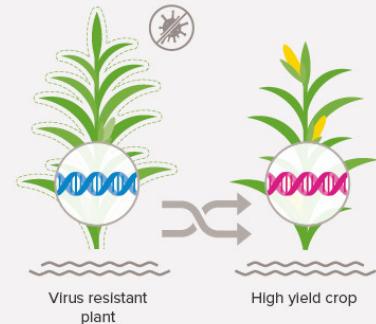


FIGURE 3 Differences between conventional breeding and GM

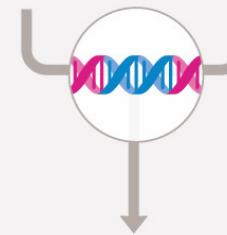
Conventional breeding



Genetic modification



Virus resistant and high yield crop



Virus resistant and high yield crop

# Changed our understanding of the human microbiome

NIH Human Microbiome Project



Characterization of the microbiomes of healthy human subjects at five major body sites, using 16S and metagenomic shotgun sequencing.

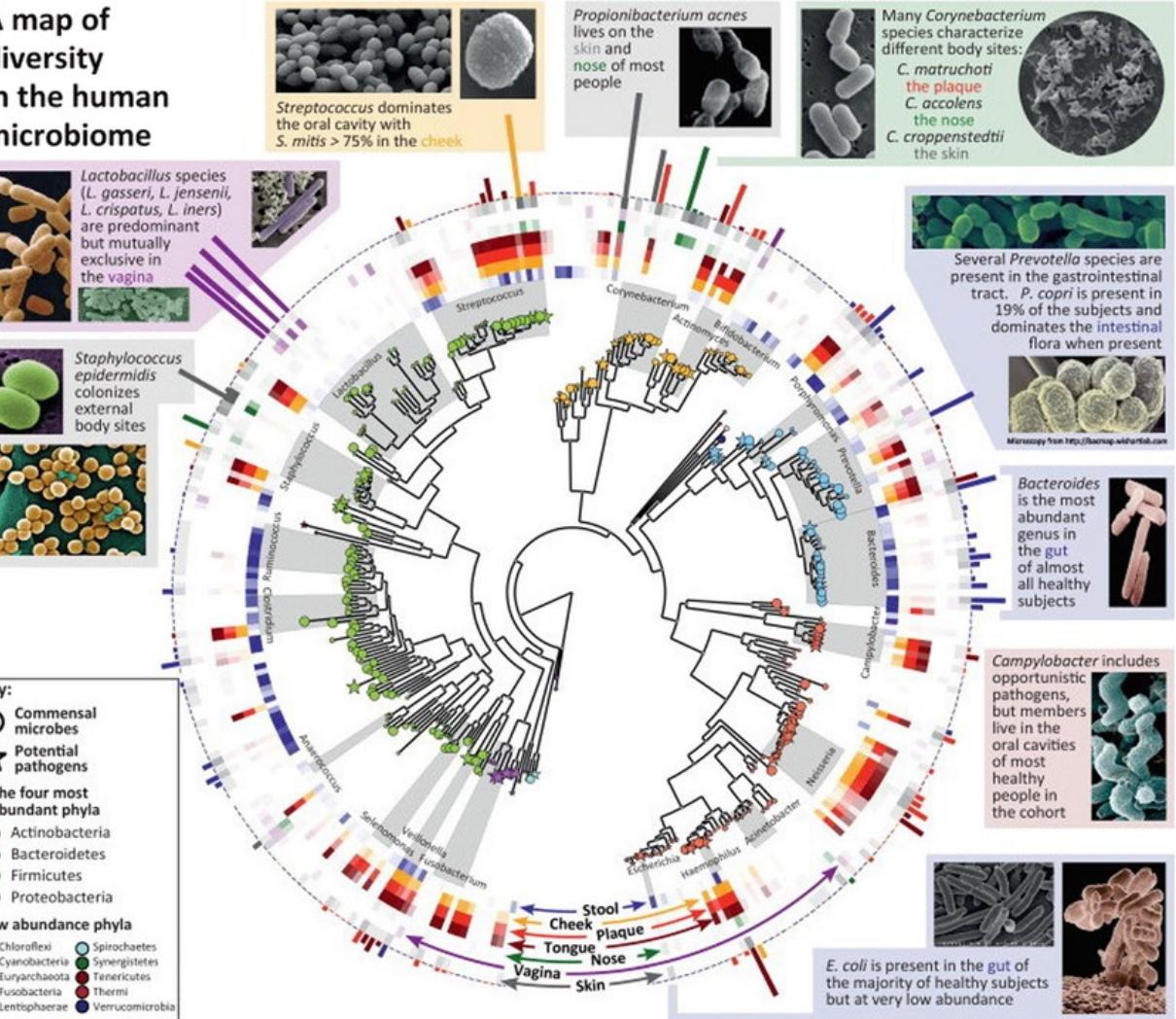
Enter HMP



Characterization of microbiome and human host from three cohorts of microbiome-associated conditions, using multiple 'omics technologies.

Enter iHMP

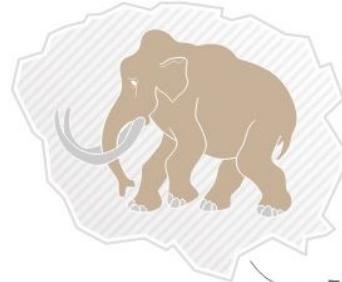
## A map of diversity in the human microbiome



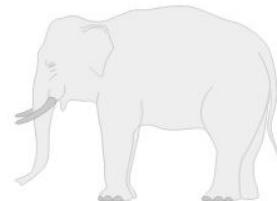
# *Changing our perspective of extinct species*

## **How Woolly mammoths could be brought back from extinction**

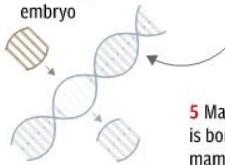
**1** DNA extracted from mammoth found in permafrost.



**2** Identify genes which separate them from elephants, such as those which code for a shaggy coat, big ears and antifreeze blood.



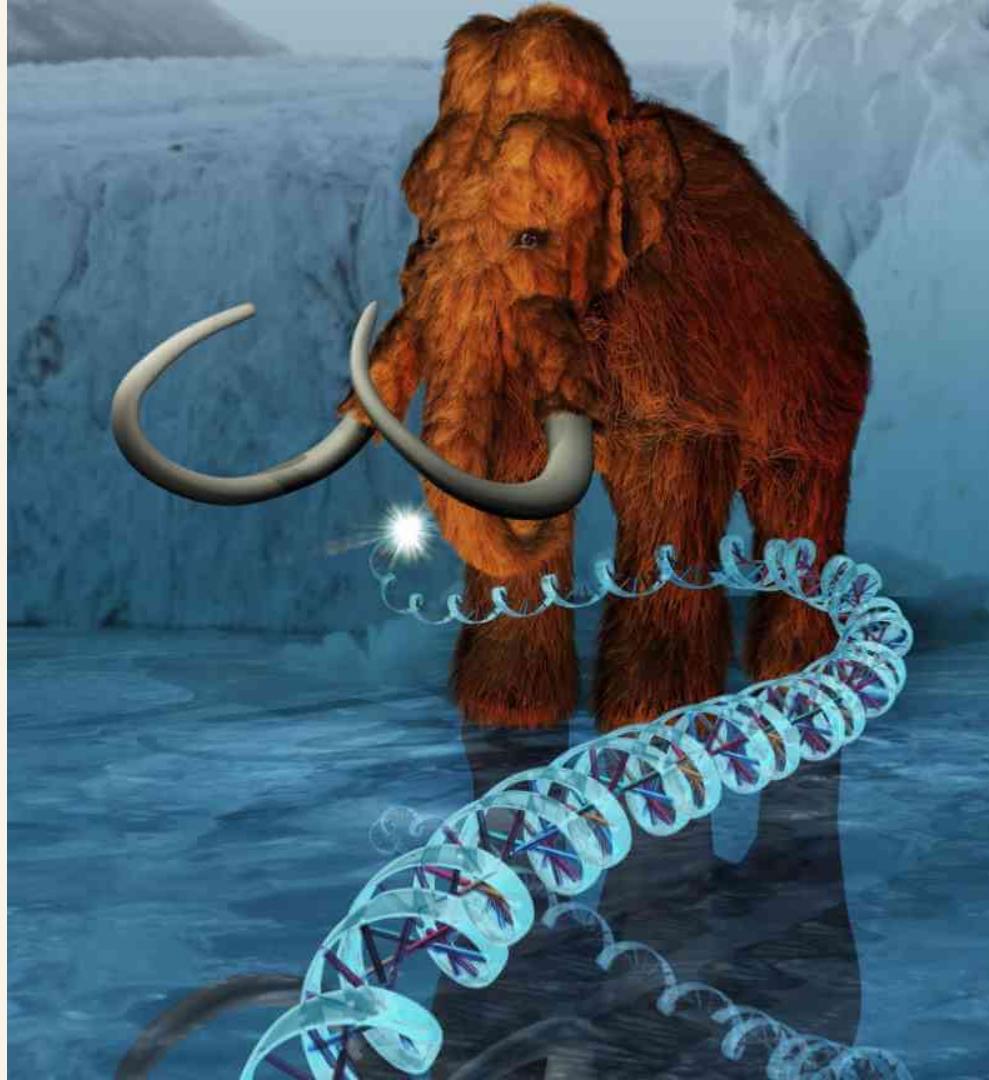
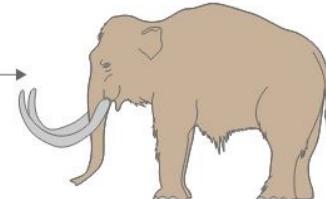
**3** Splice the mammoth gene into the genome of an elephant embryo



**4** Grow the mammoth embryo within an artificial womb



**5** Mammoth-elephant hybrid is born with a number of mammoth traits



# *Changing our perspective of extinct species*

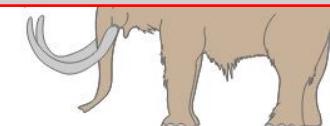
## **How Woolly mammoths could be brought back from extinction**

**1** DNA extracted from mammoth found in permafrost.

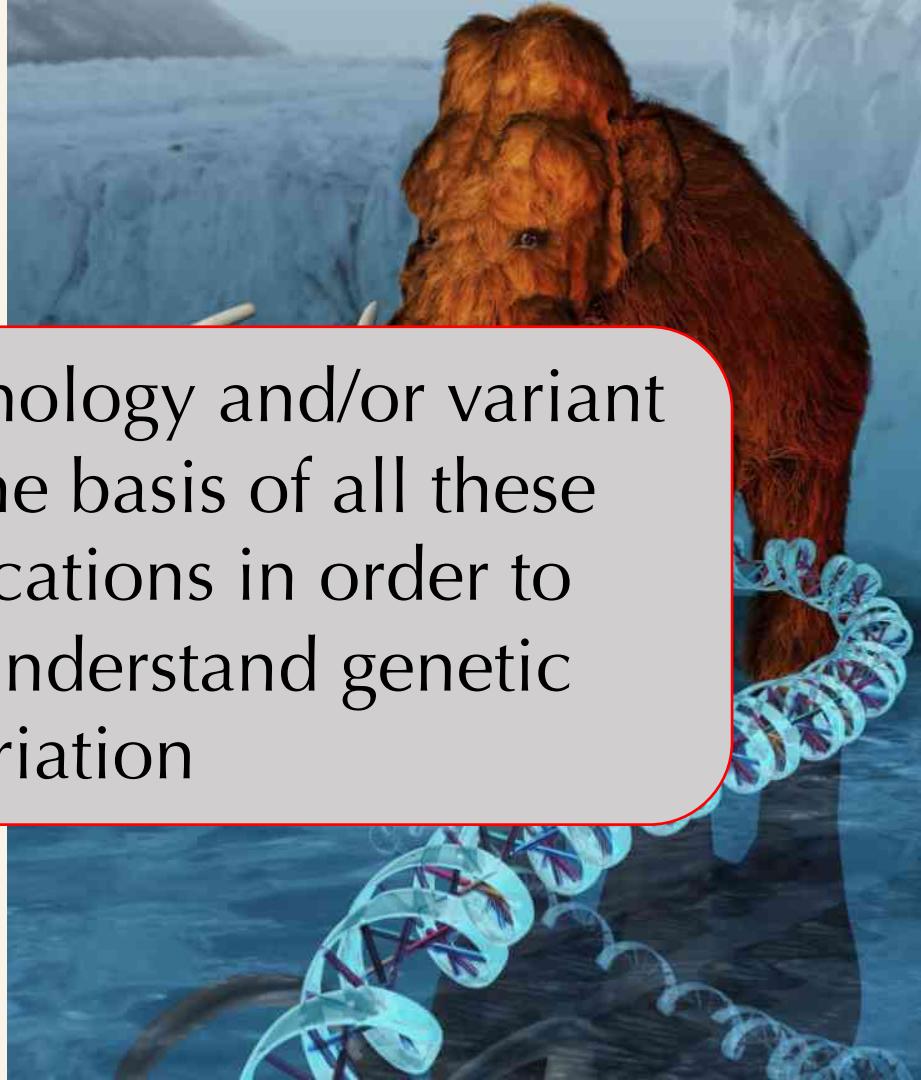
**2** Identify genes which separate them from elephants, such as those which code for a shaggy coat, big ears and antifreeze blood.



**4** Grow the mammoth embryo within an artificial womb



Sequencing technology and/or variant calling are at the basis of all these different applications in order to quantify and understand genetic variation



Available to  
high school  
students!  
21.10 2022

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## High school student is first to sequence the angelfish genome

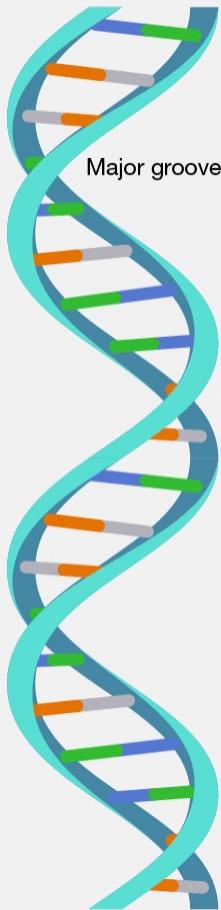
17-year-old Indeever Madireddy sequenced the genome of his pet angelfish after it died – the first time this species has been sequenced



LIFE 21 October 2022

By Michael Le Page

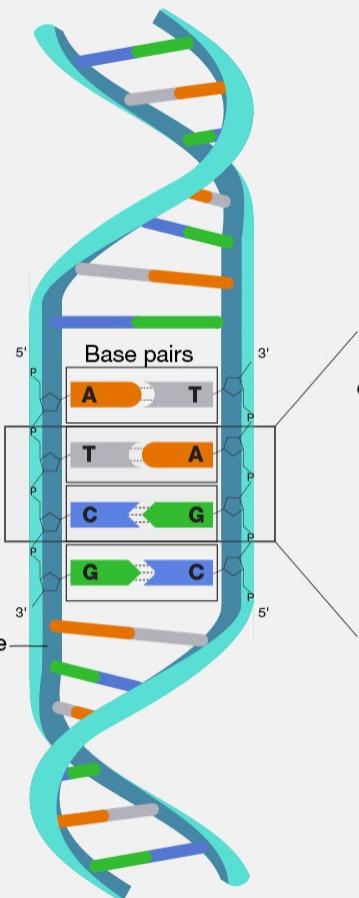




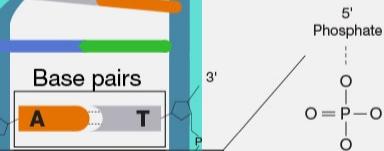
Minor groove

Major groove

Sugar-phosphate backbone



Base pairs



Phosphate

5'

Thymine



Nucleotide

Adenine

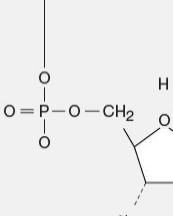
3'

Nitrogenous  
base

Sugar

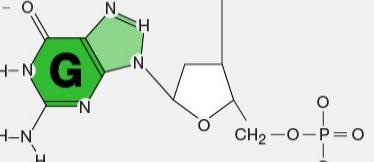
Phosphate  
group

Cytosine



3'

Guanine



5'

Phosphate

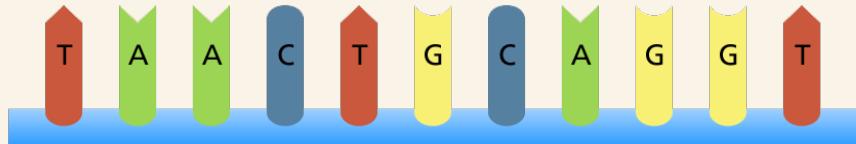
# DNA

# What does genetic variation look like?

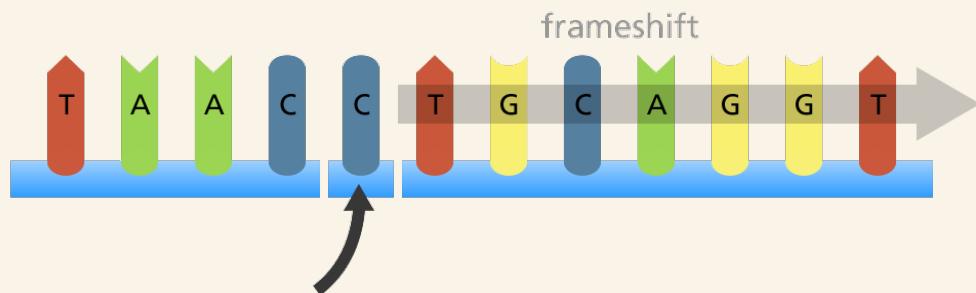
- 1) DNA (nucleotides) can be inserted or deleted (*indels*).

# 1) Insertion/Deletion (Indel)

Original sequence



Insertion



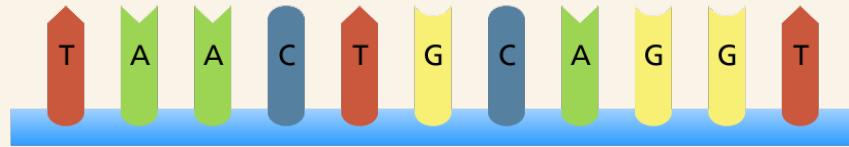
Can range from 1 base-pair (bp) to many bp

# What does genetic variation look like?

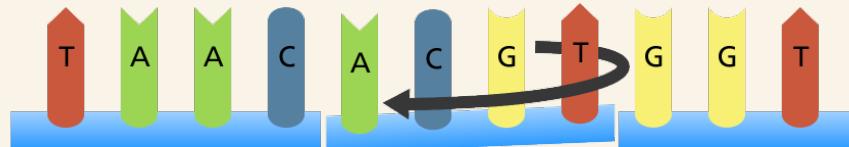
- 1) DNA (nucleotides) can be inserted or deleted (*indels*).
- 2) DNA can be *structurally rearranged* (inversions/translocations)

## 2) Structural rearrangements (inversions/translocations)

Original sequence



Inversion



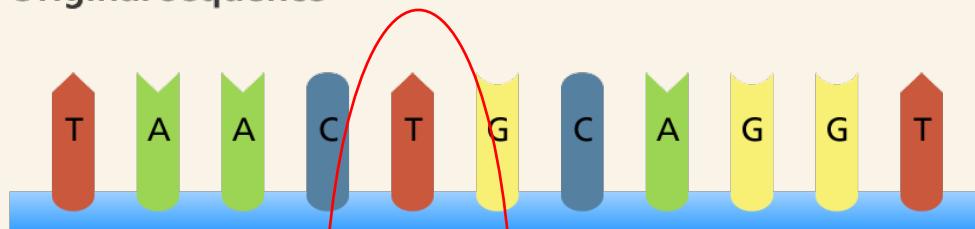
Can be MILLIONS of bp long affecting the order of many genes simultaneously  
(*Supergenes*)

# What does genetic variation look like?

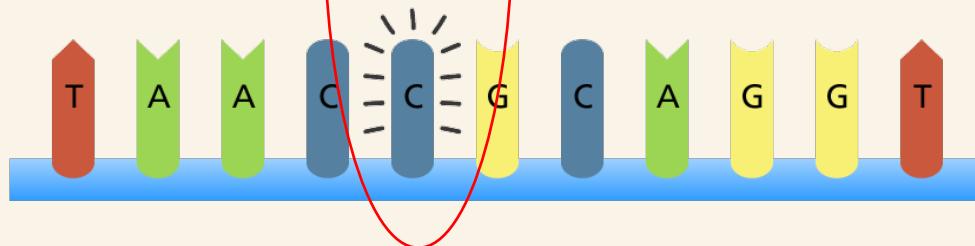
- 1) DNA (nucleotides) can be inserted or deleted (*indels*).
- 2) DNA can be *structurally rearranged* (inversions/translocations)
- 3) DNA can be *altered* at a single base pair (Single Nucleotide Polymorphism or SNP)

### 3) Single Nucleotide Polymorphism (SNP)

Original sequence



Point mutation



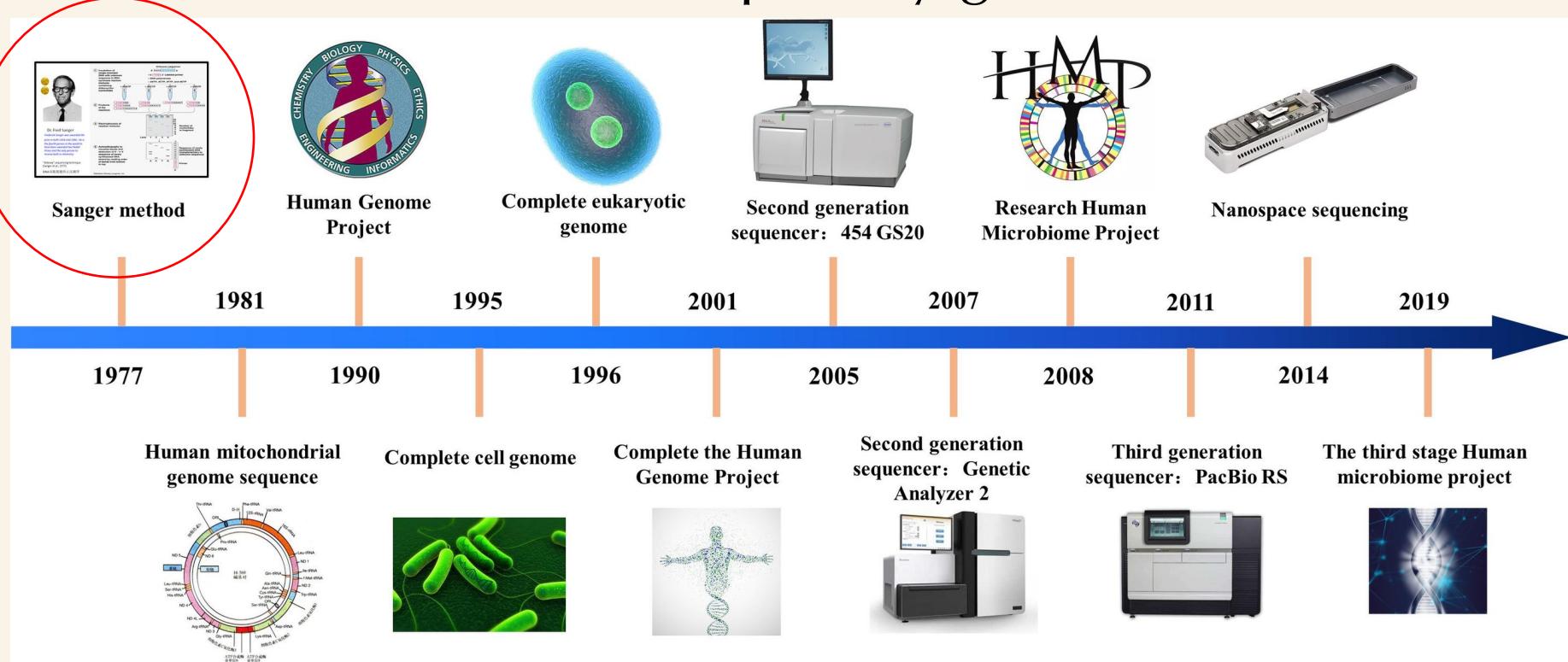
Extremely common

> 150 million known SNPs  
in humans (2015)

~100 SNPs unique in  
EVERY human

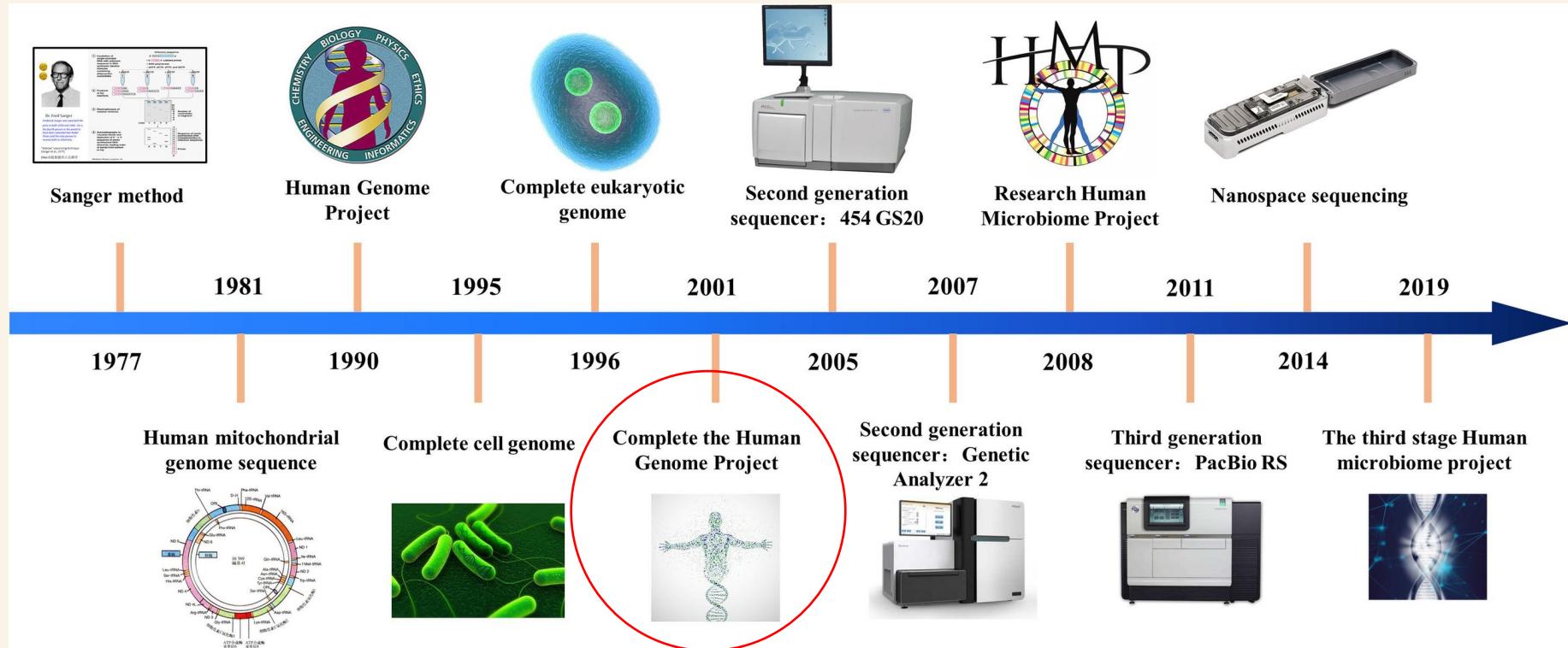
*Putatively EVERY base in  
the human genome*

# How do we observe and quantify genetic variation?



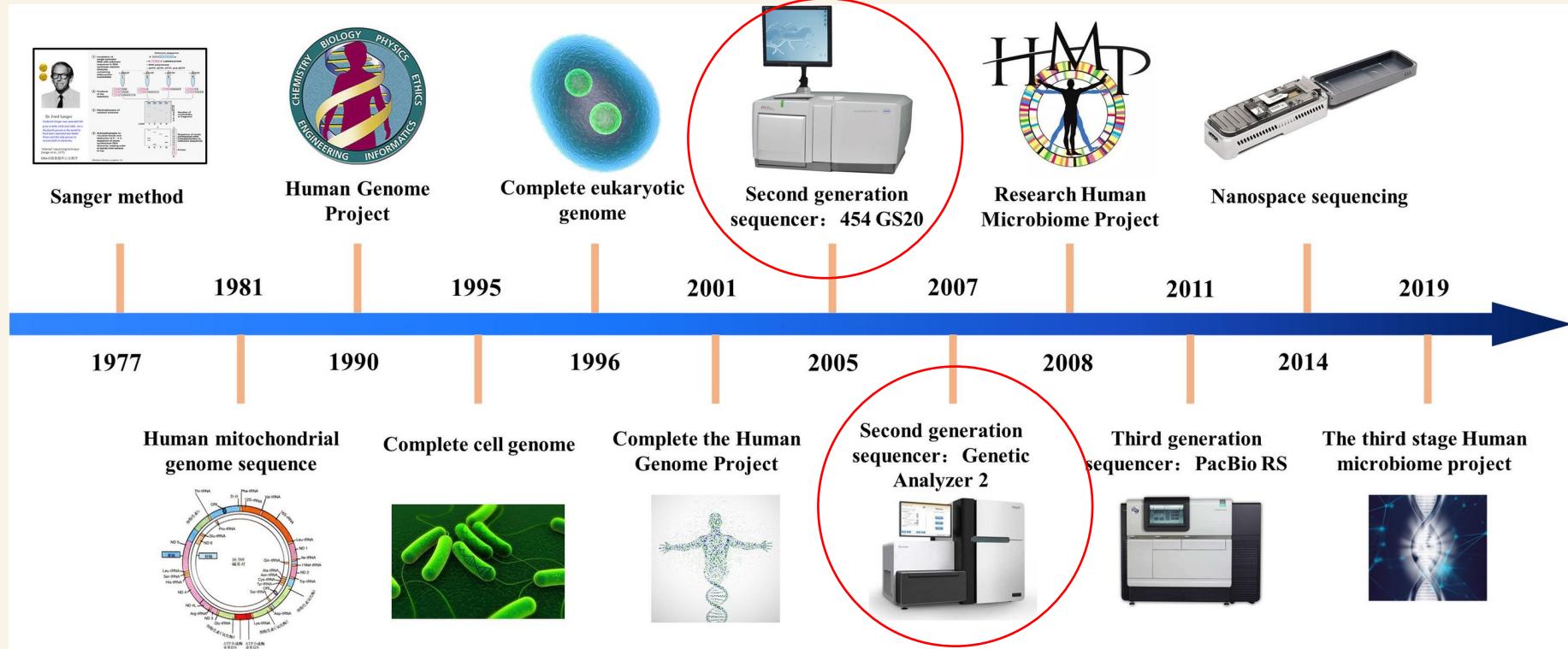
**Sanger sequencing – leading sequencing technology for decades**

# How do we observe and quantify genetic variation?



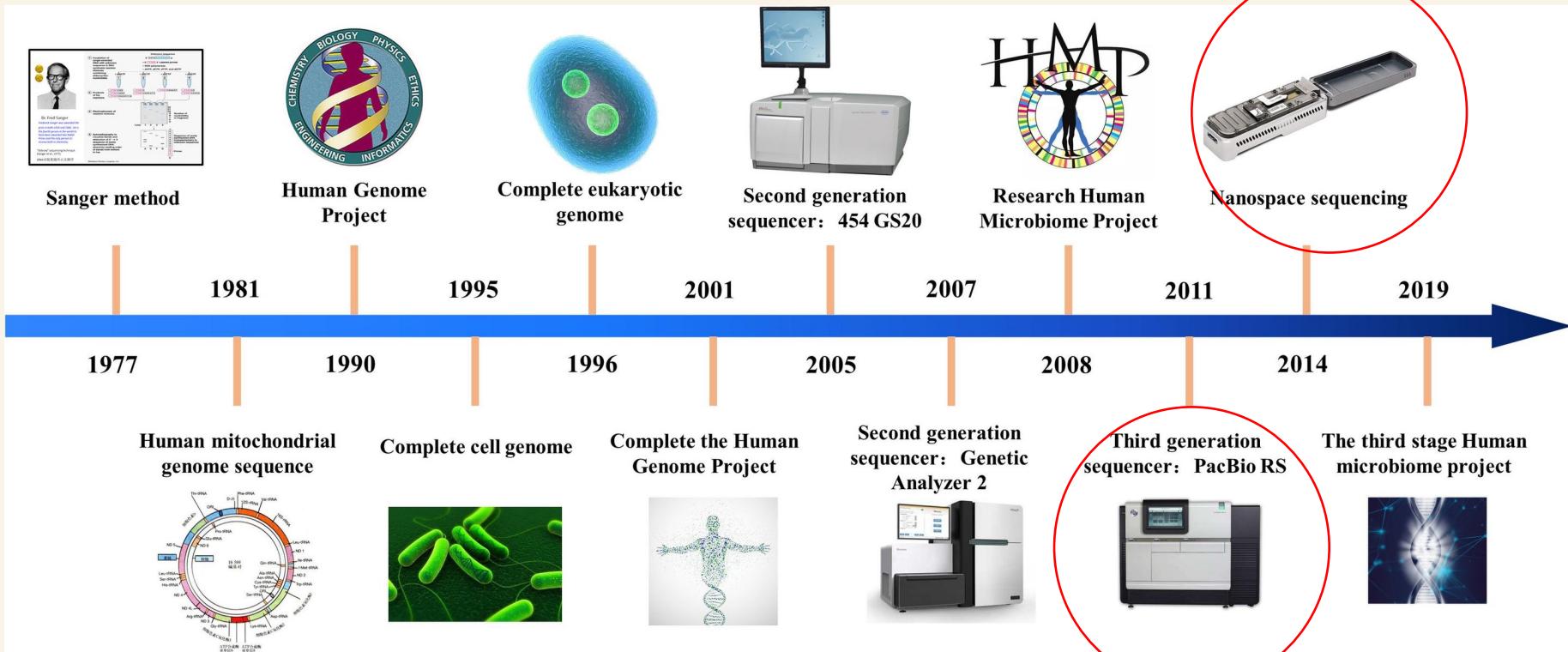
Human genome project: sparked a novel industry

# How do we observe and quantify genetic variation?



**“New” sequencing technologies (already outdated!)**

# How do we observe and quantify genetic variation?



Latest sequencing technologies that focus on long read sequencing

# Two dominant technologies today



PacBio

Long read length (10k bp +)

More expensive

Specific applications

# Two dominant technologies today



PacBio

Long read length (10k bp +)

More expensive

Specific applications



Illumina

Short read length (150-250 bp)

Cheap!

Workhorse of sequencing

# Practical considerations: size matters!



PacBio  
Long read length (10k bp +)  
More expensive  
Specific applications



Illumina  
Short read length (150-250 bp)  
Cheap!  
Workhorse of sequencing

# What variation can you assess with these different types of reads?

Type of variant	Short reads	Long reads
Indel	Only if small (~few bp)	Yes
Structural (inversion)	Difficult	Yes
SNP	Yes	Yes

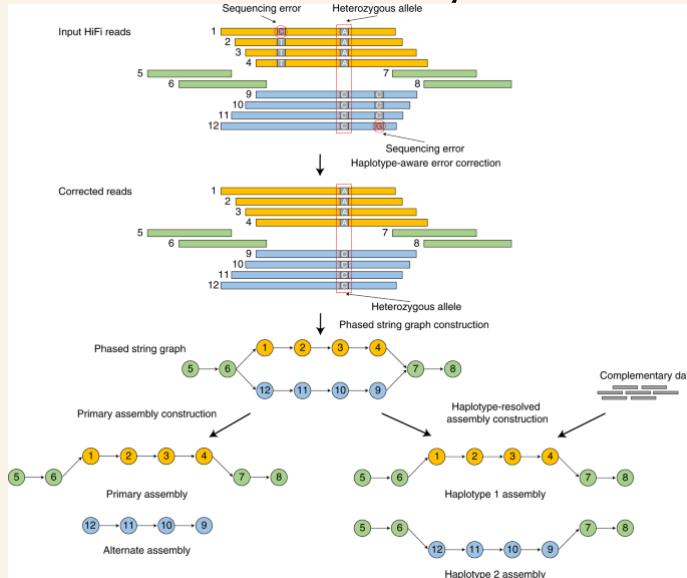
# What variation can you assess with these different types of reads?

Type of variant	Short reads	Long reads
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Structural (inversion)	Difficult	Yes
SNP	Yes	Yes

Illumina **re-sequencing** domination means that SNPs are most reliably targeted and are most studied type of genetic variation

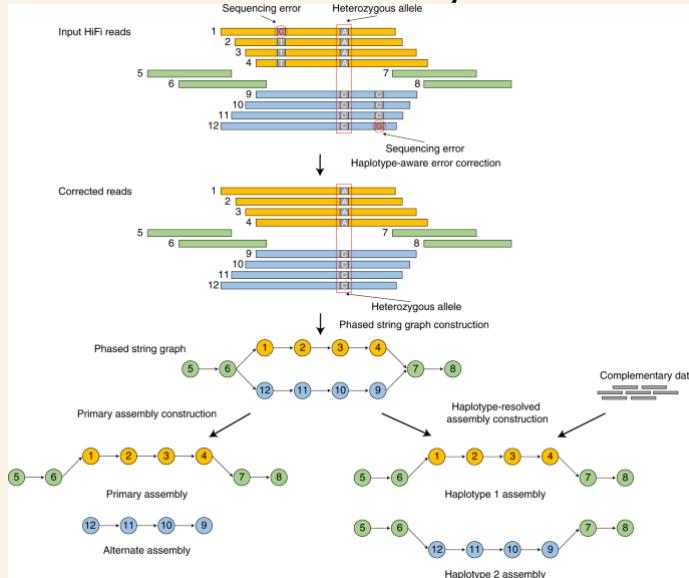
# Yet there *are* different ways of assessing genetic variation

i.e. *de novo*  
haplotype aware  
assembly

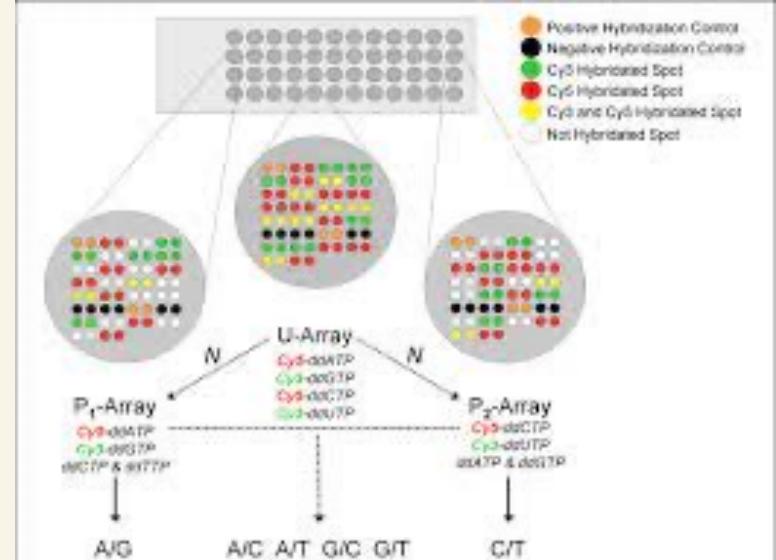


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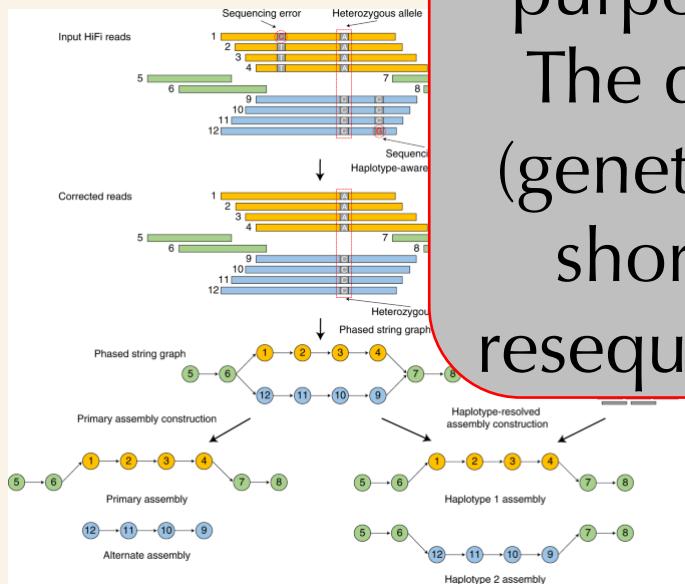


## SNP Chip/DNA micro array genotyping

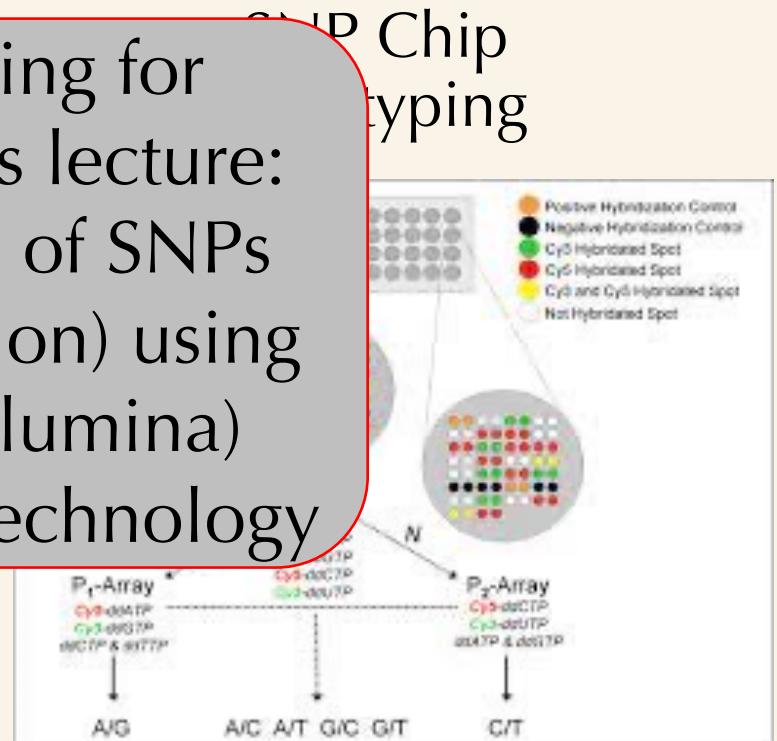


# Yet there are different ways of assessing genetic variation

i.e. *de novo*  
haplotype  
assembly



Variant calling for  
purpose of this lecture:  
The detection of SNPs  
(genetic variation) using  
short read (Illumina)  
resequencing technology



# Questions?

WHO? WHERE?  
WHEN? WHY? HOW?  
**WHAT?**  
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**WHAT?**  
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## 2) Variant calling pipelines/methods and limitations

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Variant calling **always** starts with a reference genome



Assembly of the first complex vertebrate genome  
Human genome assembly project (2003)  
Not easily repeated: it was massive task  
Nowadays; much cheaper and faster

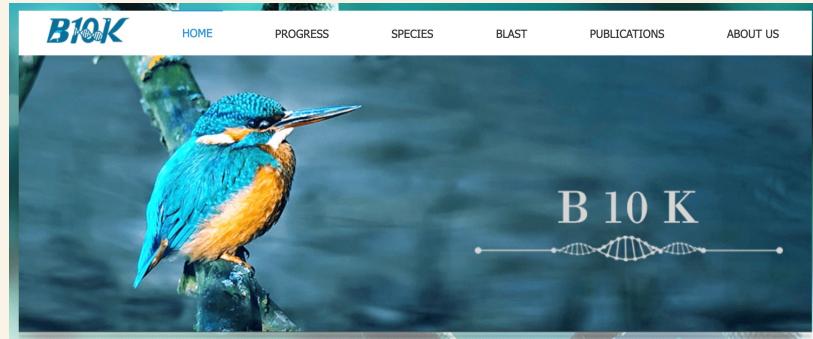
## 2) Variant calling pipelines/methods and limitations

Variant calling **always** starts with a reference genome



Assembly of the first complex vertebrate genome  
Human genome assembly project (2003)  
Not easily repeated: it was massive task  
Nowadays; much cheaper and faster

**Great push** to provide reference genomes for many organisms!



# B10 K: 10.000 bird genomes

*Deep evolutionary understanding  
of the entire living avian class*

<https://b10k.genomics.cn/>



# The *DNA* Zoo

*facilitates conservation  
efforts by releasing high-  
quality genomics resources.*

The DNA ZOO

Search for organism or taxonomic group, e.g. Artiodactyla, giraffe, Suricata suricatta...

Cheetah	Acropora milleporosa	Addax	Yellow fever mosquito	Hoary bat
Bicolored striped s...	Common mushroom	Red panda	Allen's Swamp Monkey	American alligator
Chinese alligator	Green anole	Asian small-clawed otter	California sea hare	Golden eagle
Wild peanut	Peanut	Wild peanut	Guadalupe fur seal	Jamaican fruit bat
Pure green sweat...	Golden green sweat...	Hog deer	North Sulawesi babirusa	Fin whale

<https://www.dnazoo.org>

# The most ambitious: Earth Biogenome Project



ABOUT EBP GOVERNANCE COMMITTEES REPORTS MEDIA CONTACT

CREATING A NEW FOUNDATION FOR BIOLOGY

**Sequencing Life for the  
Future of Life**

<https://www.earthbiogenome.org/>

# The most ambitious: Earth Biogenome Project



EARTH  
BIOGENOME  
PROJECT

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*EBP: moonshot for biology, aims to*

*characterize the genomes of all of Earth's eukaryotic biodiversity over a period of ten years.*

# The most ambitious: Earth Biogenome Project



EARTH  
BIOGENOME  
PROJECT

ABOUT EBP GOVERNANCE COMMITTEES REPORTS MEDIA CONTACT

*EBP: moonshot for biology, aims to characterize the genomes of all of Earth's eukaryotic biodiversity over a period of ten years.*

The vision: to create a new foundation for biology, with new solutions for preserving biodiversity and sustaining human societies.

# But what is a reference genome?

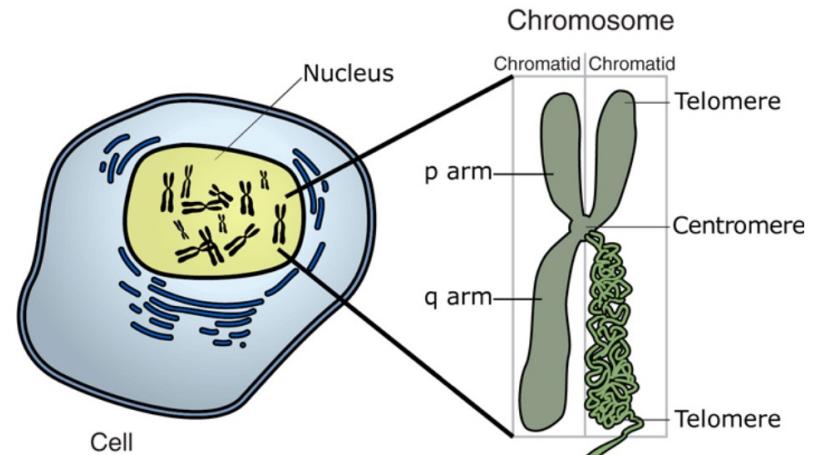
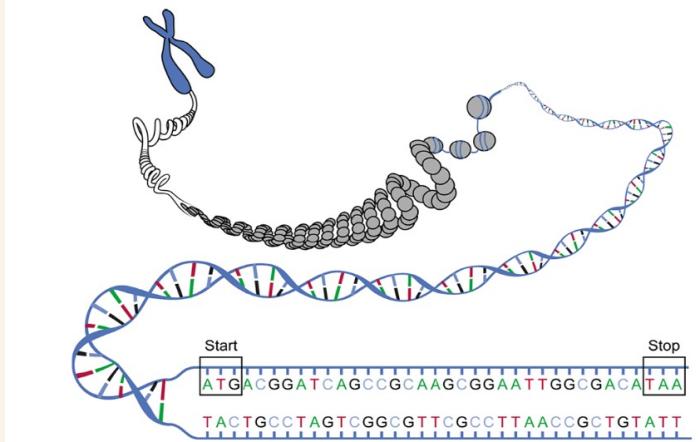
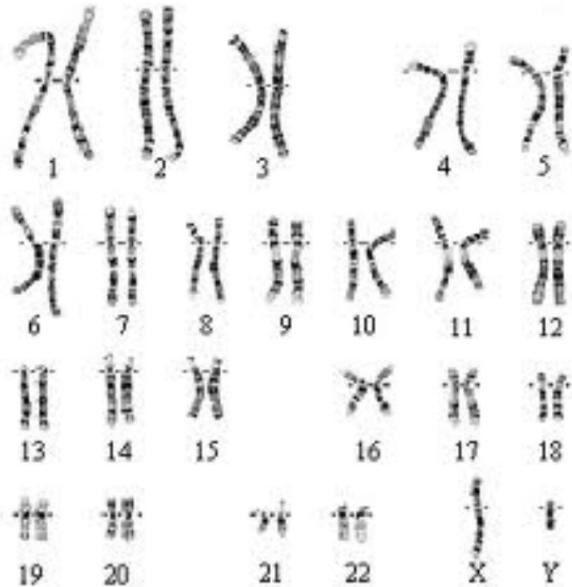


Image adapted from: National Human Genome Research Institute.



# But what is a reference genome?

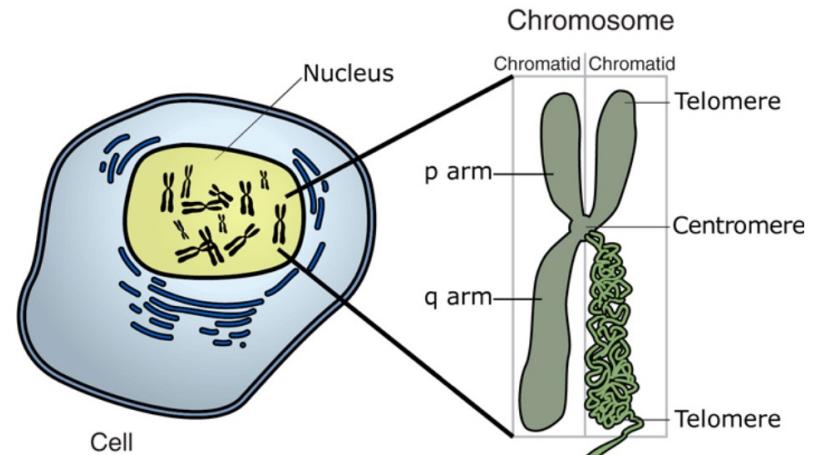
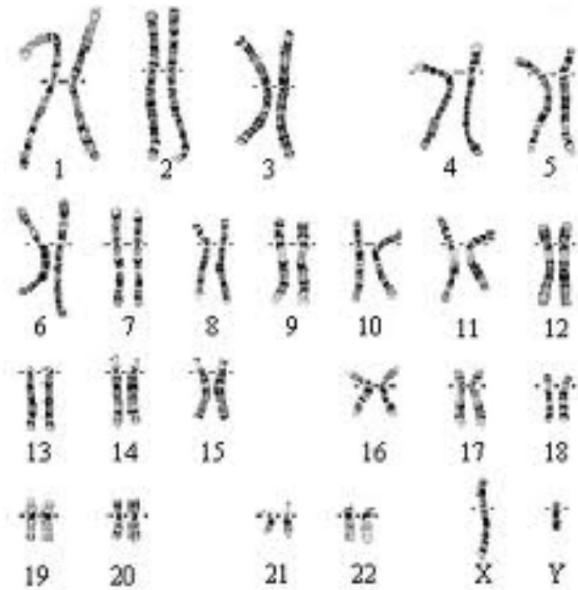


Image adapted from: National Human Genome Research Institute.



Digital representation /  
abstraction of a physical,  
biological phenomenon

# A reference genome is ...

Usually from a single individual

Result of a genome assembly process -> errors are introduced

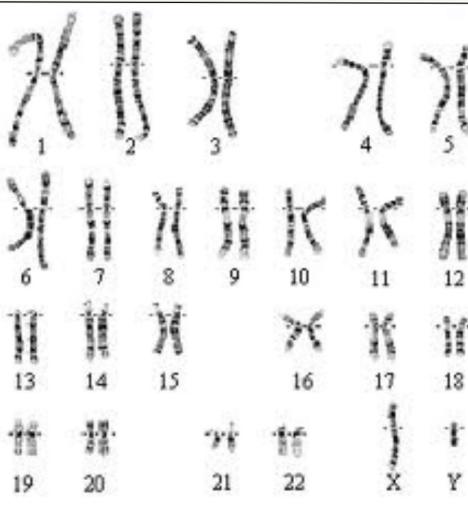
Of varying quality, that can vary from organism to organism

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Digital version of the genome

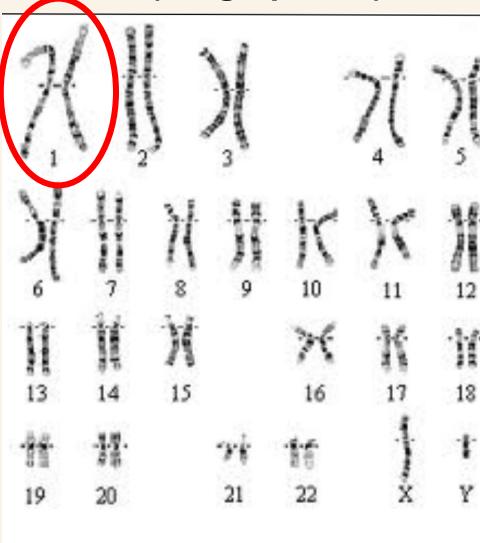
```
>Chr01  
ACTACGTATATAGCATGATCATGCATGATACTGGCTAGT...  
>Chr02  
ATCATGCATGATACTGGCTAGTACTACGTATATAGCATG...  
>Chr03  
ATGATCATGCATGATAACTACGTATATAGCCATGGCTAGT...  
>Chr04  
CGTATATAGCATGATCATGACTACATGATACTGGCTAGT...  
... ...
```

# A reference genome is ...

Usually from a single individual

Result of a genome assembly process -> errors are introduced

Of varying quality, that can vary from organism to organism



Sequencing  
and  
assembly



>Chr01

ACTACGTATATAGCATGATCATGCATGATAACATGGCTAGT...

>Chr02

ATCATGCATGATAACATGGCTAGTACTACGTATATAGCATG...

>Chr03

ATGATCATGCATGATAACTACGTATATAGCCATGGCTAGT...

>Chr04

CGTATATAGCATGATCATGACTACATGATAACATGGCTAGT...

... ...

Digital version of the genome

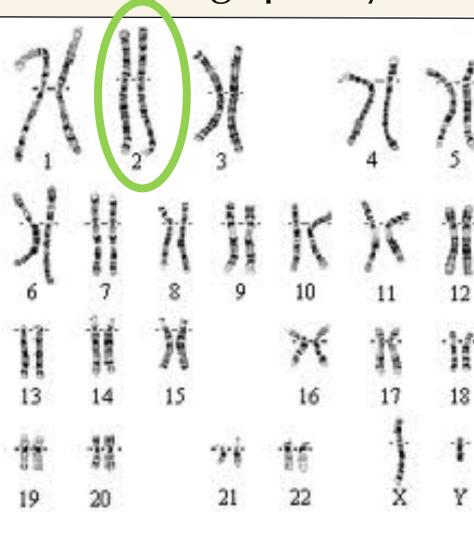


# A reference genome is ...

Usually from a single individual

Result of a genome assembly process -> errors are introduced

Of varying quality, that can vary from organism to organism



Sequencing  
and  
assembly



Digital version of the genome

>Chr01  
ACTACGTATATAGCATGATCATGCATGATACTGGCTAGT...  
>Chr02  
ATCATGCATGATACTGGCTAGTACTACGTATATAGCATG...  
>Chr03  
ATGATCATGCATGATAACTACGTATATAGCCATGGCTAGT...  
>Chr04  
CGTATATAGCATGATCATGACTACATGATACTGGCTAGT...  
... ...

# Quality scale of reference genomes

Poor

Good



- Chromosomes unclear
- Thousands of loose fragments
- Gaps (*nnnnn*) in sequences
- Missing nucleotides

# Quality scale of reference genomes

Poor

Good



Chromosomes unclear  
Thousands of loose fragments  
Gaps (*nnnnn*) in sequences  
Missing nucleotides

Chromosomes resolved  
Continuous sequences  
No gaps  
Most nucleotides covered,  
including centromeres and  
repetitive regions

# A reference genome has a 2D coordinate system

>Chr01

ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATAACATGGCTAGT...

123456789.....



Millions of nucleotides/bases

Note: some different coordinate systems exist (i.e. starting at 0 or 1)  
or using the base or space as “location”

# A reference genome has a 2D coordinate system

>Chr01

ACTACGTATATAGCATGATCATGCATGATGATCATGCATGATA  
123456789.....

Note: some different coordinate systems exist (i.e. starting at 0 or 1)  
or using the base or space as “location”

i.e. A-C-T-A-C-G-T-A

1 2 3 4 5 6 7 8  
1 2 3 4 5 6 7

Such different systems are usually automatically recognized by different software

# A multiple alignment towards a reference

>Chr01

Start (12) Stop (31)

ACTACGTATATA  
GCATGATCATGCATGATGATGATGCATGATGATA  
CATGGCTAGT...

Read 1 AGC ATGATCATGCATGATGA

Read 2 GCATGATGATCATGCATGATGATA  
CATGG

Read 3 TGATGATCATGCATGATGATA  
CATGGCTAGT



Short read sequencing data is compared to the reference (looking for a "match")

We first need such alignment before we can analyse variation

Read variation is analysed within this alignment context

The diagram illustrates a DNA sequence across three reads. The sequence starts with 'ACTACGTATATA' and ends with 'GGCTAGT...'. A red arrow labeled 'Start' points to the first 'A' in 'ACTACGTATATA'. A green arrow labeled 'SNP (G/T, 23)' points to the second 'G' in 'GCATGATGATCATGCAT...', which is highlighted in green. A red arrow labeled 'Stop' points to the second 'G' in 'GATAACATGGCTAGT...', which is also highlighted in green. A green arrow labeled 'SNP (G/A, 40)' points to the third 'G' in 'GATAACATGGCTAGT...', which is highlighted in green. Below the sequence, three reads are shown: 'Read 1 AGCATGATCATTCATGATGA' (with the 'T' highlighted in green), 'Read 2 GCATGATGATCATGCATAATACATGG' (with the 'A' highlighted in green), and 'Read 3 TGATGATCATGCATAATACATGGCTAGT' (with the 'A' highlighted in green).

An accurate alignment is *essential* before we can trust any variant

Read variation is analysed within this alignment context

>Chr01

Start

SNP (G/T, 23)

SNP (G/A, 40)

ACTACGTATATA~~G~~CATGATCAT~~G~~CATGATGATGATGCAT~~G~~A~~T~~ACATGGCTAGT...

Read 1 AGCATGATCAT~~T~~CATGATGA

Read 2 GCATGATGATCATGCAT~~A~~~~T~~ACATGG

Read 3 TGATGATCATGCAT~~A~~~~T~~ACATGGCTAGT

We usually analyse ***millions to billions*** of reads and compare these to reference genomes that consist of ***billions*** of nucleotides/bases (Human genome ~3Gb)

Read variation is analysed within this alignment context

>Chr01  
ACTACGTATATAAGC...  
Read 1 AGC

Start

SNP (G/T, 23)

SNP (G/A, 40)

SNP calling for large datasets is computationally intensive -> work on remote HPC clusters

We usually compare thousands of reads and that consist of billions of nucleotides/bases (Human genome ~3Gb)

TACATGGCTAGT...  
TACATGG  
TACATGGCTAGT

# Read variation is analysed within this alignment context

Incredibly efficient software has been designed to take care of this task!

Reference



Reference



Alignment program  
BWA  
BowTie

Unaligned reads

Aligned reads (nicely sorted  
and tiled)

# Read variation is analysed within this alignment context

Incredibly efficient software has been designed to take care of this task!

Reference



Alignment program  
*BWA*  
*BowTie*

Reference

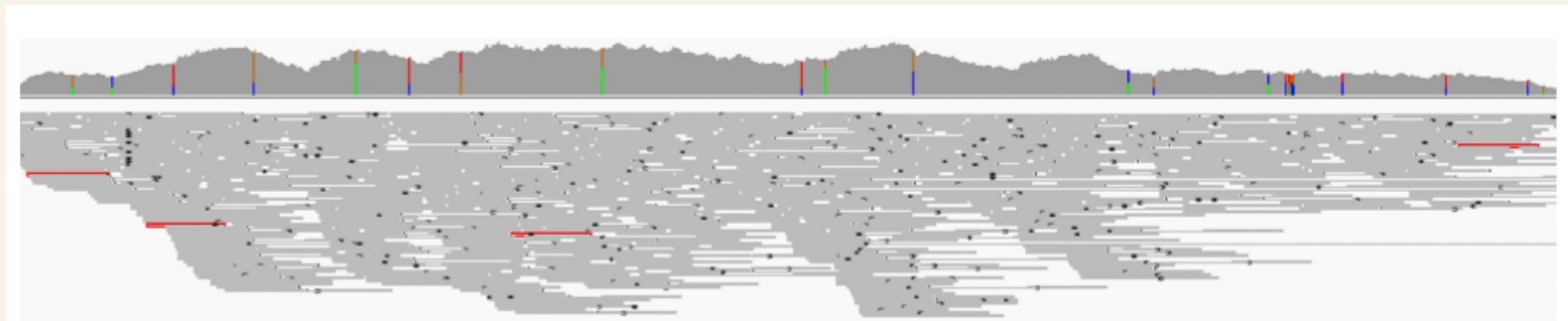


Unaligned reads

Aligned reads (nicely sorted  
and tiled)

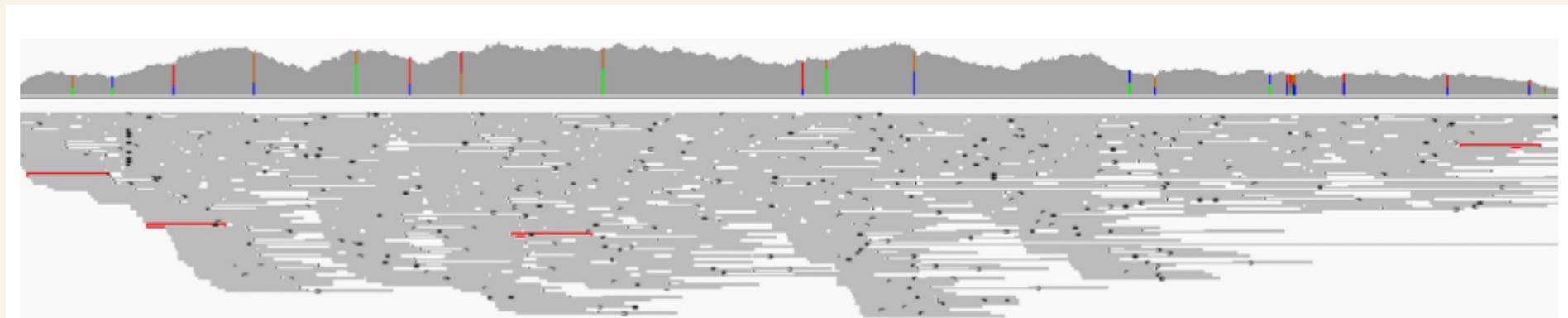
Standard program settings are usually sufficient

# Visualisation of thousands of reads



# Visualisation of thousands of reads

Genetic variation (SNPs) colours reflect which bases are variable (A-C, A-T, G-C, etc)



After aligning, we need another program to determine which bases are variable:

A SNP caller

# SNP calling programs

Table 1. A brief summary of different tools.

caller	Bcftools	16GT	Freebayes	VarScan2	GATK
Code	C	Perl	C++	Java	Java
Model	HMM & MAQ	16-genotype probabilistic	Bayesian	heuristic algorithm	Bayesian
Sampling	Single & multiple	Single	Single	Single & multiple	Single & multiple
Variants	SNPs & indels	SNPs & indels	SNPs & indels&MNP	SNPs & indels	SNPs & indels
Features	Sorting, indexing, etc.	easy to use, timesaving	straightforward	meet desired thresholds for read depth, base quality, variant allele frequency, and statistical significance	Realignment, per base recalibration, VQSR
Reference	Danecek et al., 2017 [15]	Luo et al., 2017 [19]	Garrison and Marth, 2012 [18]	Koboldt et al., 2012 [16]	Mckenna et al., 2010 [14]

<https://doi.org/10.1371/journal.pone.0262574.t001>

Liu J, Shen Q, Bao H (2022)

Many programs exist, and there is *continuous* development  
For instance Bcftools/16GT are now recommended  
Yet use of GATK is wide-spread (oldest, developed by Broad institute, good documentation)

# What does a variant caller do?

Aims to provide statistical confidence in observing TRUE genetic variation

>Chr01

ACTACGTATATAGCATGATCAT**G**ATGATGATCATGCATGATA  
Read 1 AGCATGATCAT**T**ATGATGA

Is this real or not?

# What does a variant caller do?

Aims to provide statistical confidence in observing TRUE genetic variation

```
>Chr01  
ACTACGTATATAGCATGATCATGATGATGATGCATGATACATGGCTAGT...  
Read 1 AGCATGATCATTATGATGA
```

Is this real or not?

Sequencing data (as any type of data) comes with errors (wrong bases called) and/or uncertainty (low quality of bases) in the call

Solution? Generate LOTS more data!

# What does a variant caller do?

With more data (read), more certainty is obtained: ***fold coverage***

>Chr01

ACTACGTATATA~~G~~CATGATCAT~~G~~CATGATGATCATGCATGATA~~C~~ATGGCTAGT...

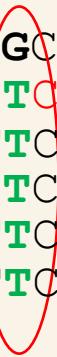
Read 1 AGC~~A~~TGATCAT~~T~~CATGATGA

Read 2 ATGATCAT~~T~~CATGATGATCAT

Read 3 GATCAT~~T~~CATGATGATCATGCATGAT

Read 4 TCAT~~T~~CATGATGATCATGCAT

Read 5 CATT~~T~~CATGATGATCATGCATGATA~~C~~ATGG



**5-fold** coverage, all the same, we are pretty certain about this call (note: we usually strive for ~20 fold coverage)

# What does a variant caller do?

Another example

>Chr01

ACTACGTATATA~~G~~CATGATCAT**G**CATGATGATGATGCATGATA~~C~~ATGGCTAGT...

Read 1 AGC~~A~~TGATCAT**T**CATGATGA

Read 2 ATGATCAT**T**CATGATGATGATCAT

Read 3 GATCATT**T**CATGATGATGATCATGCATGAT

Read 4 TCAT**A**CATGATGATGATCATGCAT

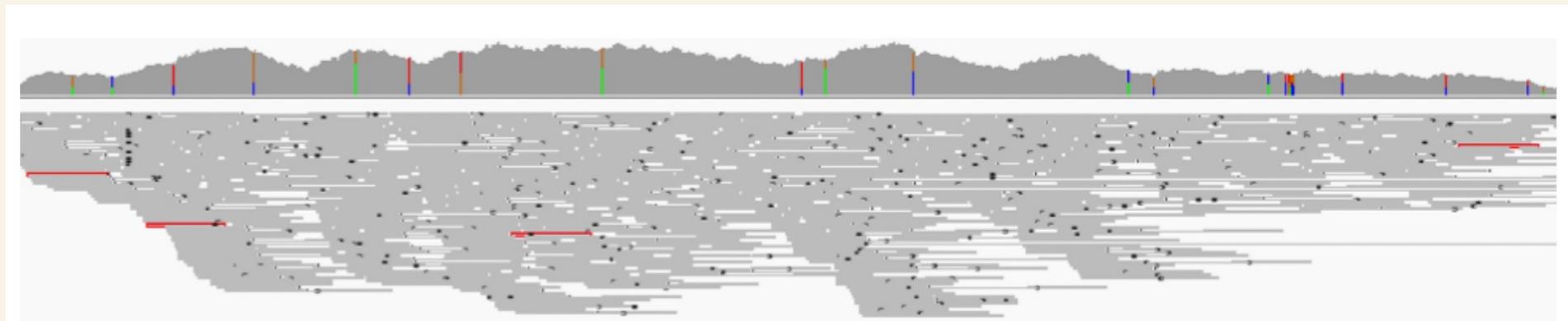
Read 5 CAT**T**CATGATGATGATCATGCATGATA~~C~~ATGG



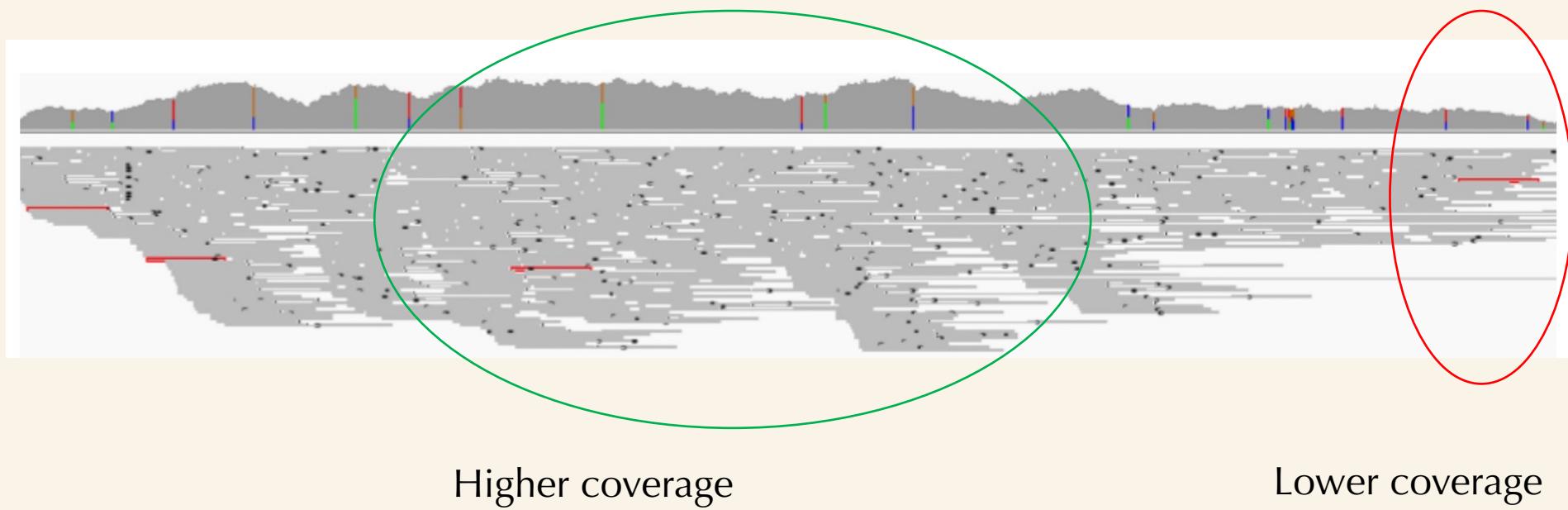
We cannot be so certain about the A, until we get more data

**Coverage is the most important determinant for the quality of your data**

Yet along a reference, you'll obtain variable coverage due to random processes, assembly quality, or genomic complexity



Yet along a reference, you'll obtain variable coverage due to random processes, assembly quality, or genomic complexity



Yet along a reference, you'll obtain variable coverage due to random complexity

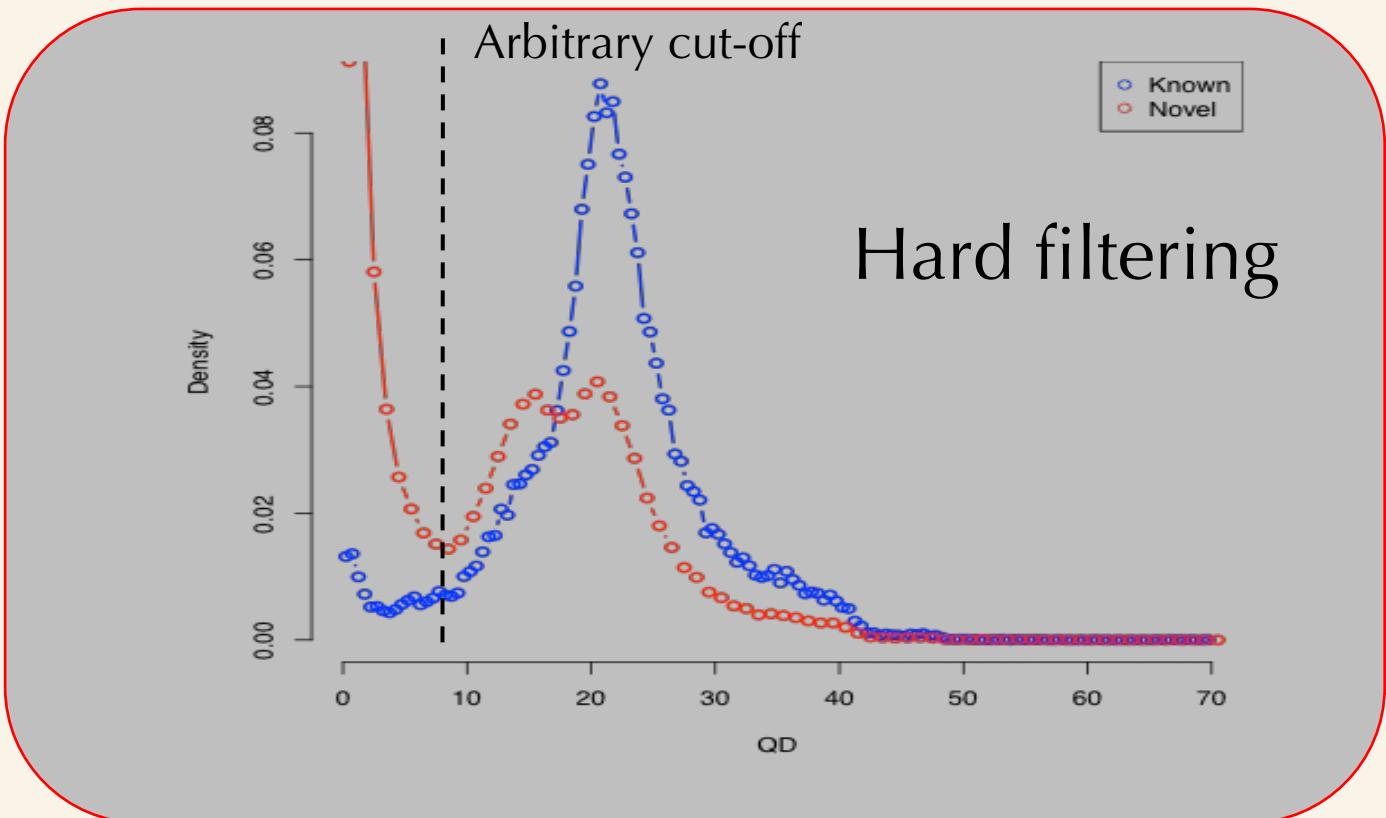
SNP callers run complex statistical models (e.g. Bayesian or HMM models) to provide confidence in SNP calls and if they are “TRUE”. They often assume correct read alignment **and** require sufficient read coverage in order to provide high-quality calls

Higher coverage

Lower coverage

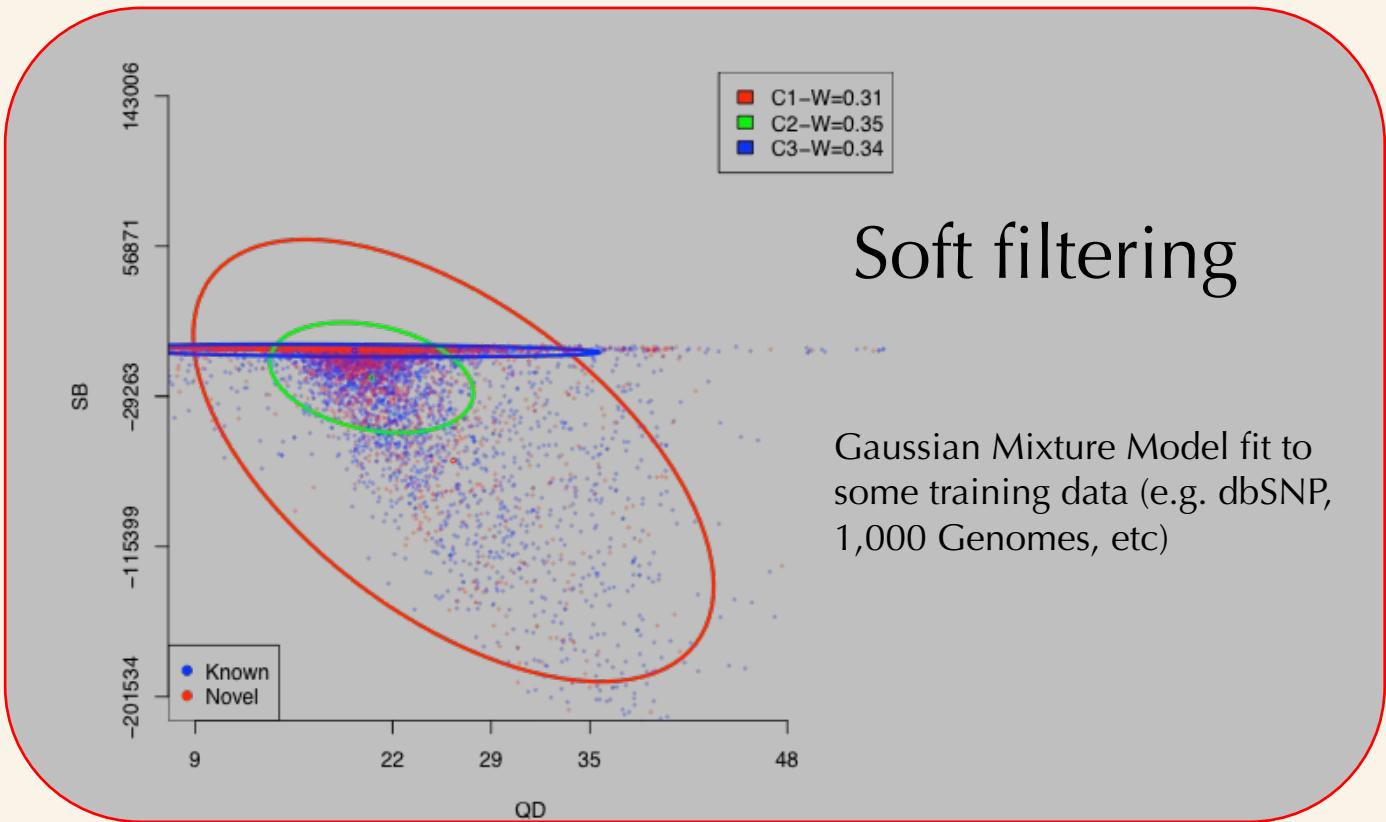
SNP callers will ALSO yield a large numbers of SNPs of which many will NOT be true (false positives)

We need to **filter** our data to only retain the high quality part of the data

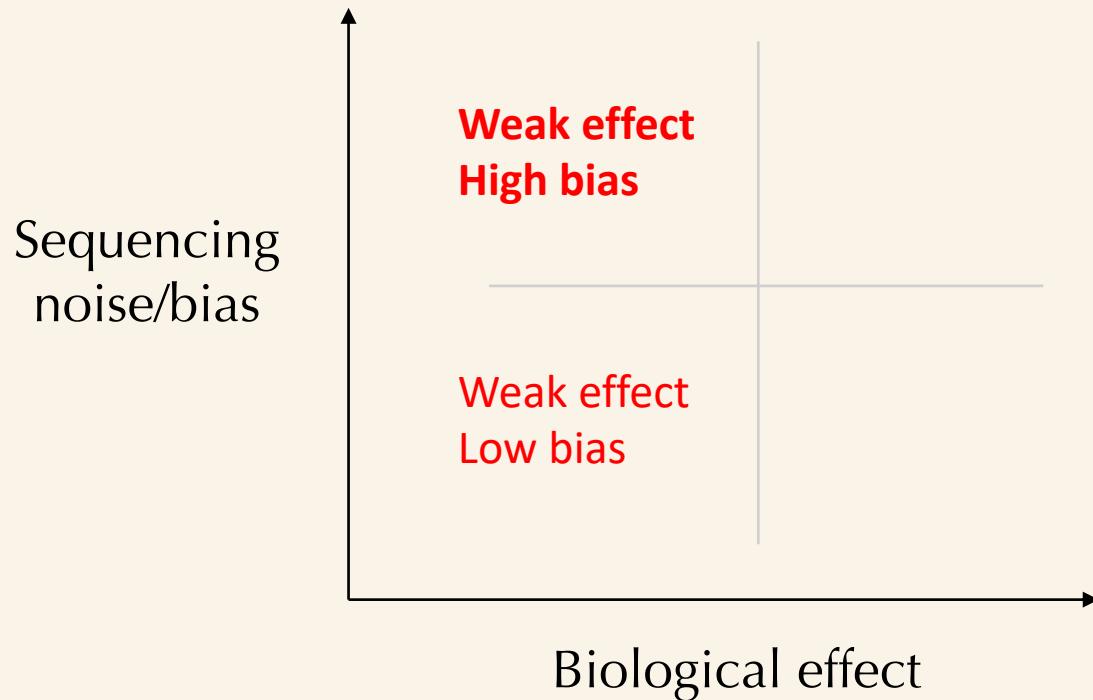


SNP callers will ALSO yield a large numbers of SNPs of which many will NOT be true (false positives)

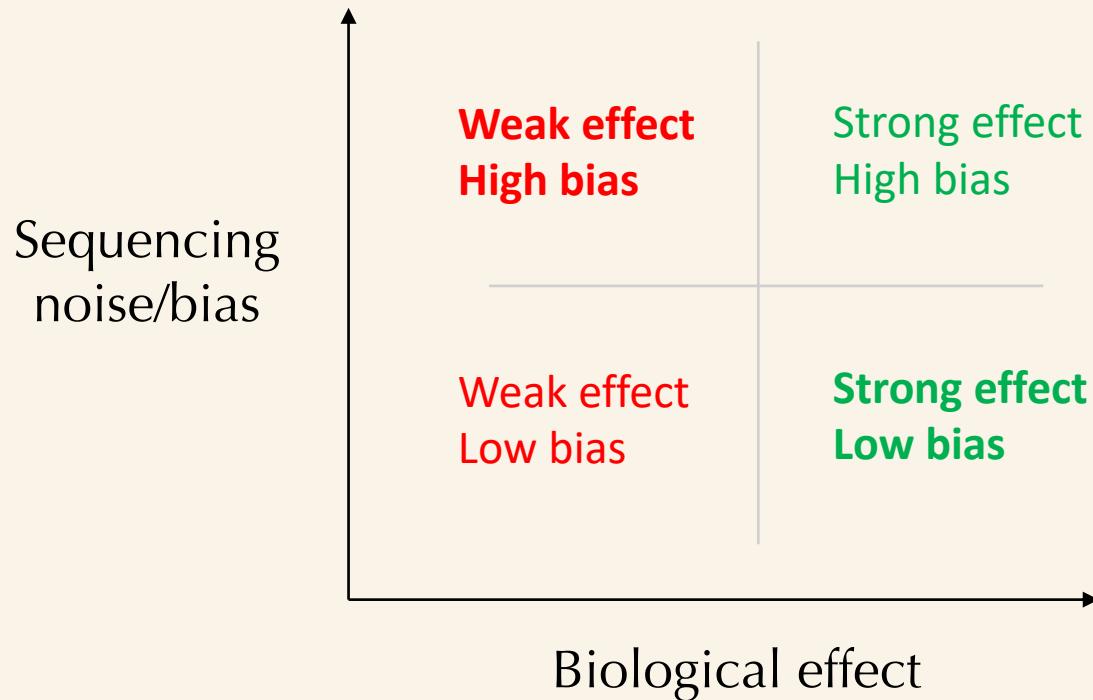
We need to **filter** our data to only retain the high quality part of the data



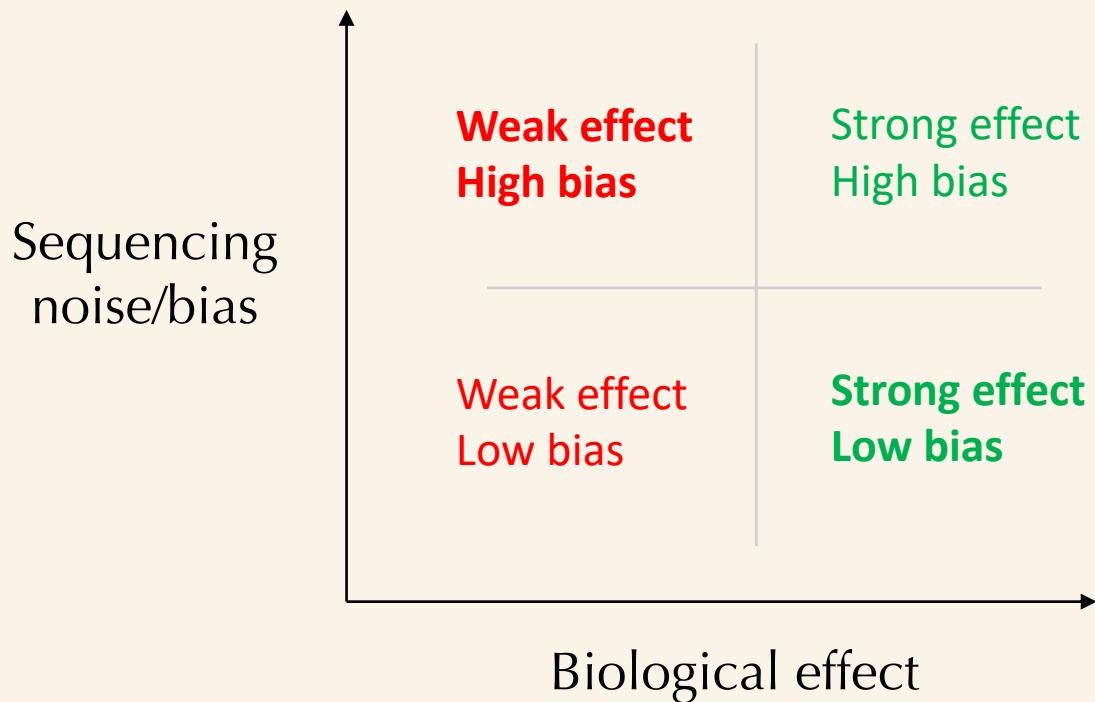
Yet there is no “fixed” approach to filtering your data



Yet there is no “fixed” approach to filtering your data



# Yet there is no “fixed” approach to filtering your data



It is not always clear from the outset where you are! You need to explore your data and use preliminary analyses

# Questions?

WHO? WHERE?  
WHEN? WHY? HOW?  
**WHAT?**  
WHO? WHERE?  
WHAT?  
WHO?  
WHERE?  
WHAT?  
WHY?  
WHO?  
WHERE?  
WHAT?  
WHEN?  
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WHEN?

WHO?  
WHAT?  
WHERE?  
WHY?  
HOW?  
WHEN?  
**WHAT?**  
WHO?  
WHERE?  
WHY?  
HOW?  
WHEN?  
WHERE?  
WHAT?  
WHEN?

# After all this, what does a variant calling pipeline look like?



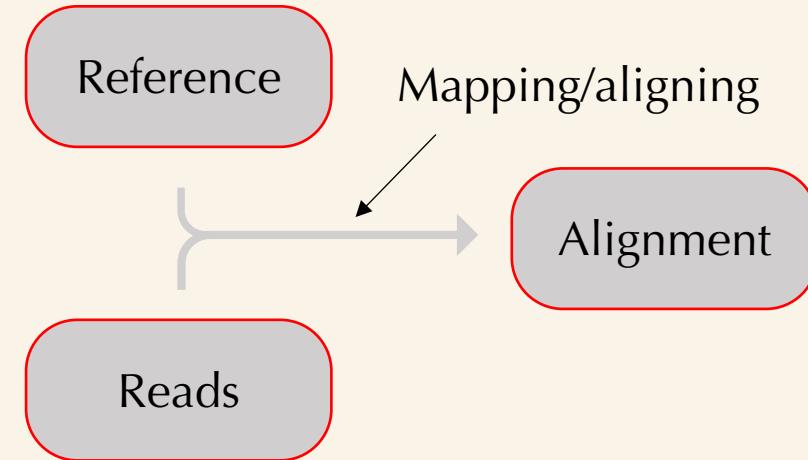
Reference

Reads



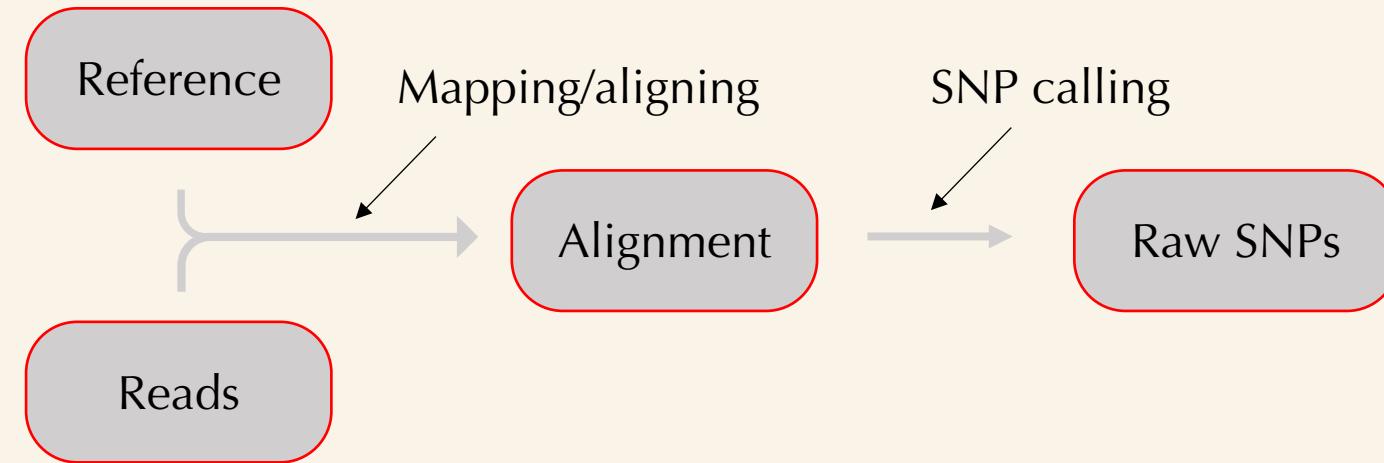
e.g.  
population  
data

# After all this, what does a variant calling pipeline look like?



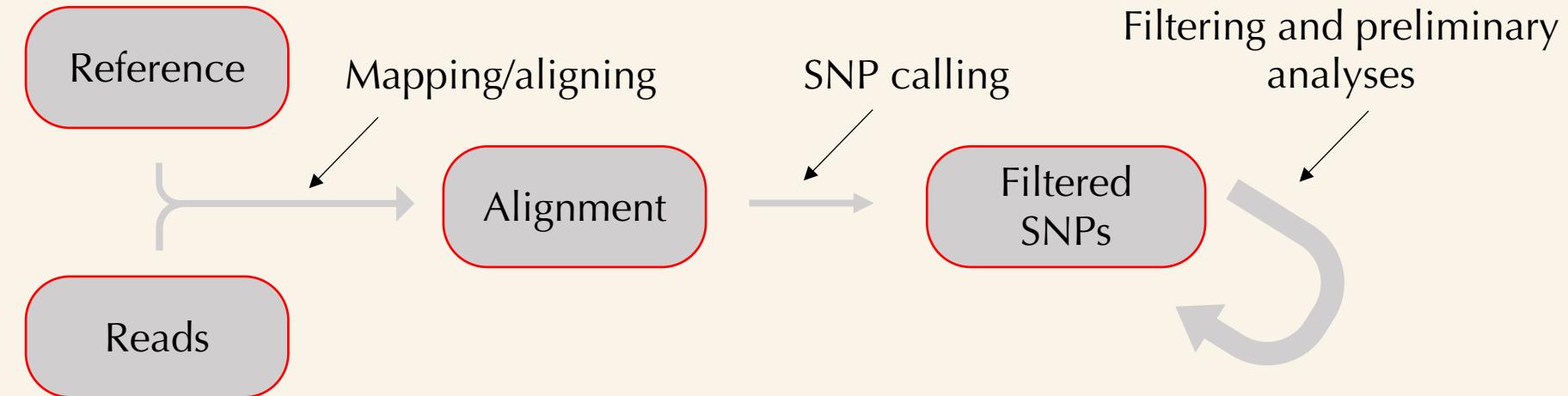
e.g.  
population  
data

# After all this, what does a variant calling pipeline look like?



e.g.  
population  
data

# After all this, what does a variant calling pipeline look like?

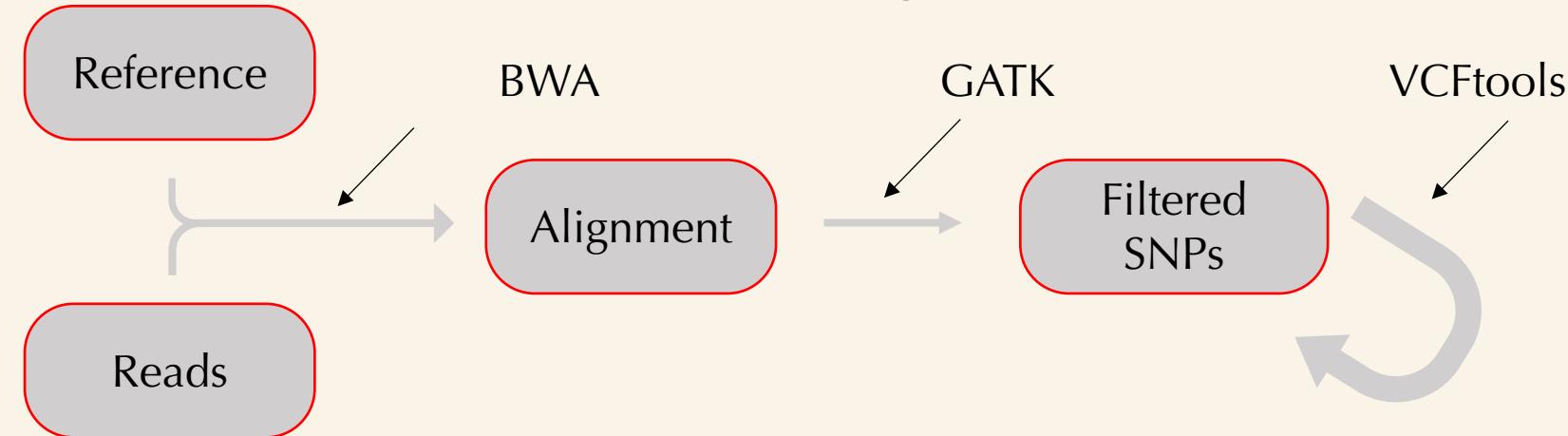


e.g.  
population  
data

# After all this, what does a variant calling pipeline look like?

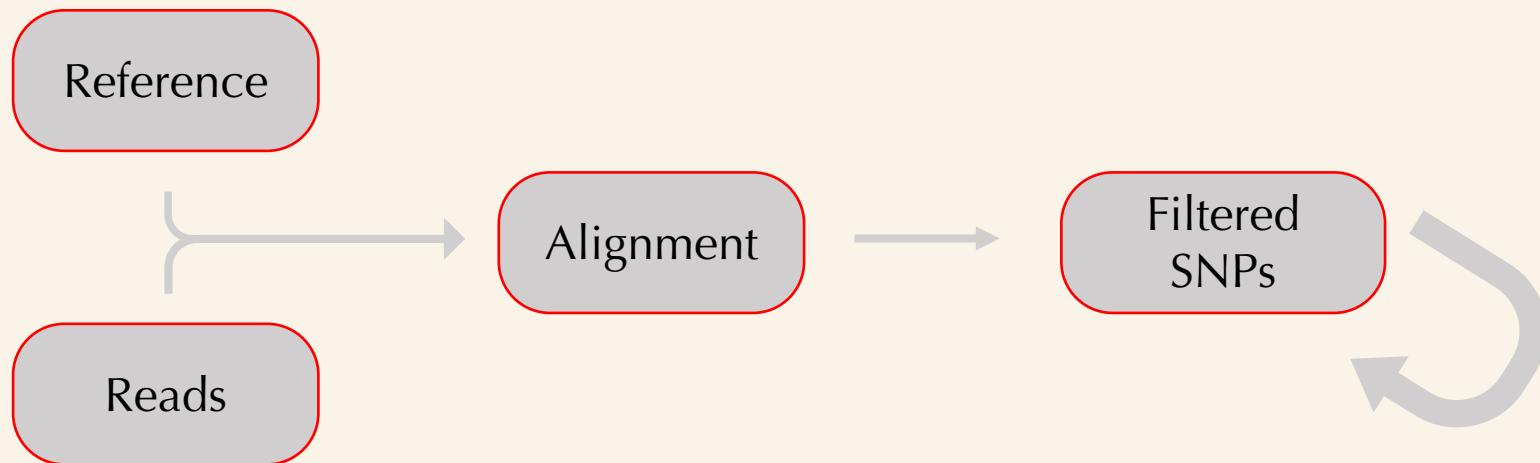


A selection of programs that can be used

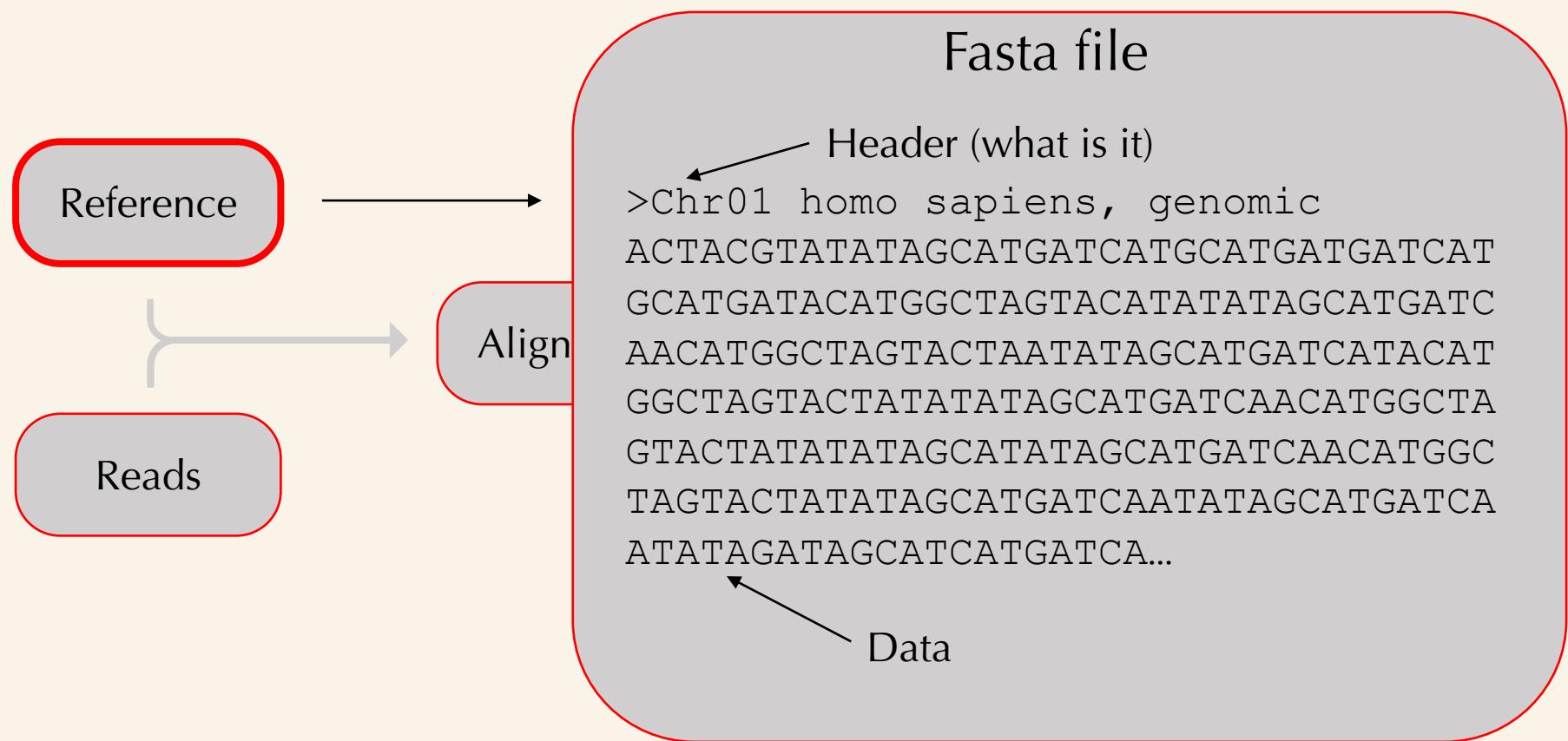


e.g.  
population  
data

Each of these steps requires specific files to work with!



Each of these steps requires specific files to work with!



Each of these steps requires specific files to work with!



# Each of these steps requires specific files to work with!

Alignment



## BAM file (binary alignment file)

Kind of data

```
@HD VN:1.5 GO:none SO:coordinate  
@SQ SN:NC_004029.2 LN:16565  
@RG ID:L1i1_AGAACCG SM:WLR001  
@RG PL:ILLUMINA PG:bwa  
@PG ID:bwa PN:bwa VN:0.7.17-r1188 CL:bwa samse Orosv1mt.fasta  
@PG ID:GATK IndelRealigner VN:3.6-0-g89b7209 CL:knownAlleles=[] targetIntervals=WLR001/Wal_m...  
@PG ID:samtools CL:samtools view -H WLR001.Wal_mt.realigned.bam
```

“Header” with information about the file

Reference

Sample name

Each of these steps requires specific files to work with!

Alignment

BAM file (binary alignment file)

Readname

Follow by data with information about each read alignment

M\_D00564:55:C9FG3ANXX:7 0  
M\_D00564:55:C9FG3ANXX:7 0  
M\_D00564:55:C9FG3ANXX:7 0

NC\_004029.2  
NC\_004029.2  
NC\_004029.2

419  
474  
515

37  
37  
37

91M  
58M  
56M

TAAAAAAGCTGCCGCTAATACAAAATATACTACGAAAGTACT  
TTACACGACAGCTAACGACCCAAACTGGGATTAGATACCCAC  
CTATGCTTAGCCATAAACACAAATAATTGCACAACAAAATT

Reference name

Start of alignment

Matching bases

Quality of alignment (37 is max)

CIGAR string (56 matching bases)

Each of these steps requires specific files to work with!

## SNP data

# VCF file (Variant call format)

Each of these steps requires specific files to work with!

SNP data



VCF file (Variant call format)  
Followed by the data:

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO
NC_004029.2	131	.	T	C	356.22	.	AC=1;AF=0.022;AN=45;DP=143;FS=0.000;MLEAC=1;MLEAF=0.022;PValue=0.000;RankSum=-0.000
NC_004029.2	162	.	T	C	18479.23	.	AC=15;AF=0.333;AN=45;BaseQRankSum=0.00;DP=543;FS=0.000;MLEAC=15;MLEAF=0.333;PValue=0.000;RankSum=-0.000
NC_004029.2	198	.	C	T	608.22	.	AC=1;AF=0.022;AN=45;DP=410;FS=0.000;MLEAC=1;MLEAF=0.022;PValue=0.000;RankSum=-0.000
NC_004029.2	387	.	G	A	547.22	.	AC=1;AF=0.022;AN=45;DP=408;FS=0.000;MLEAC=1;MLEAF=0.022;PValue=0.000;RankSum=-0.000
NC_004029.2	616	.	T	C	235.62	.	AC=1;AF=0.022;AN=45;DP=406;FS=0.000;MLEAC=1;MLEAF=0.022;PValue=0.000;RankSum=-0.000
NC_004029.2	741	.	C	T	819.22	.	AC=1;AF=0.022;AN=45;DP=412;FS=0.000;MLEAC=1;MLEAF=0.022;PValue=0.000;RankSum=-0.000
NC_004029.2	743	.	C	T	819	.	AC=1;AF=0.022;AN=45;DP=413;FS=0.000;MLEAC=1;MLEAF=0.022;PValue=0.000;RankSum=-0.000



Reference name

Each of these steps requires specific files to work with!

SNP data

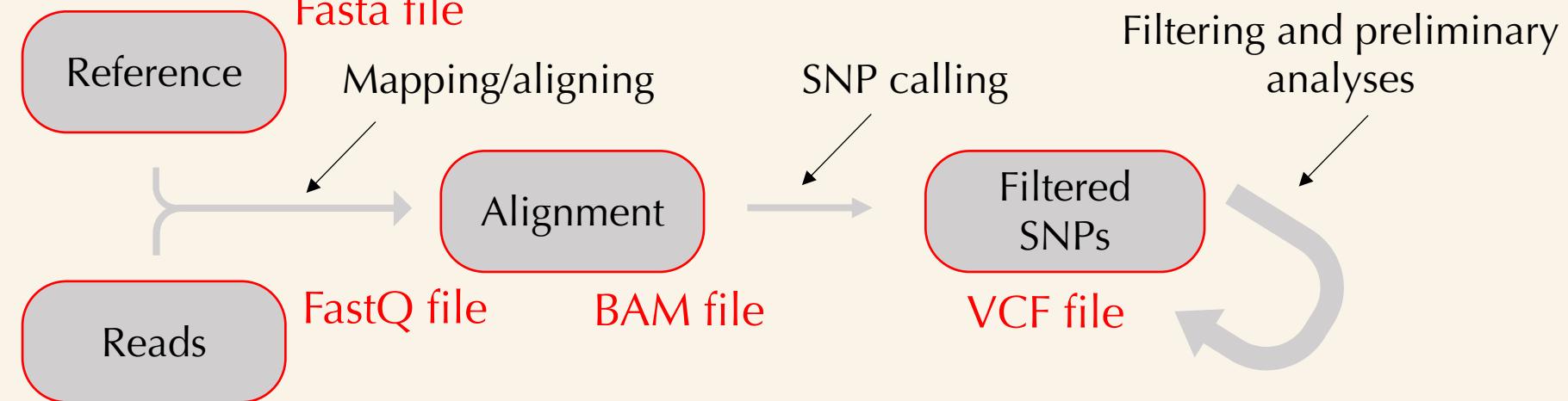
## VCF file (Variant call format)

Followed by the data:

**GenoType: Allele Depth: Read Depth (DP): Genotype Quality: Phred-scaled Likelihood**

FORMAT	WLR001	WLR002	WLR003	WLR004	WLR005
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:2,0:2:90:0,90	0:0,0:0:0:0,0	0:0,0:0:0:0,0	0:0,0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:4,0:4:99:0,135	0:1,0:1:0:0,0	0:1,0:1:45:0,45	0:1,0:1:45:0,45
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:5,0:5:46:0,46	0:0,0:0:0:0,0	0:2,0:2:90:0,90	0:2,0:2:90:0,90
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:3,0:3:99:0,135	0:0,0:0:0:0,0	0:2,0:2:45:0,45	0:2,0:2:45:0,45
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:0,0:0:0:0,0	0:0,0:0:0:0,0	0:0,0:0:0:0,0	0:0,0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:3,0:3:99:0,128	0:0,0:0:0:0,0	0:1,0:1:45:0,45	0:1,0:1:45:0,45
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:3,0:3:99:0,128	0:0,0:0:0:0,0	0:1,0:1:45:0,45	0:1,0:1:45:0,45
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:3,0:3:99:0,128	0:0,0:0:0:0,0	0:1,0:1:45:0,45	0:1,0:1:45:0,45
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:3,0:3:99:0,135	0:0,0:0:0:0,0	0:1,0:1:42:0,42	0:1,0:1:42:0,42
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0,0	0:1,0:1:42:0,42	0:1,0:1:42:0,42
GT:AD:DP:GQ:PL	0:1,0:1:45:0,45	0:1,0:1:45:0,45	0:0,0:0:0:0,0	0:3,0:3:99:0,119	0:3,0:3:99:0,119
GT:AD:DP:GQ:PL	0:1,0:1:45:0,45	0:1,0:1:45:0,45	0:0,0:0:0:0,0	0:3,0:3:99:0,119	0:3,0:3:99:0,119
GT:AD:DP:GQ:PL	0:1,0:1:0:0,0	0:1,0:1:0:0,0	0:0,0:0:0:0,0	0:0,0:0:0:0,0	0:0,0:0:0:0,0
GT:AD:DP:GQ:PL	0:1,0:1:0:0,0	0:1,0:1:0:0,0	0:0,0:0:0:0,0	0:0,0:0:0:0,0	0:0,0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0,0	0:0,0:0:0:0,0	0:0,0:0:0:0,0
GT:AD:DP:GQ:PL	0:0,0:0:0:0,0	0:1,0:1:45:0,45	0:0,0:0:0:0,0	0:0,0:0:0:0,0	0:0,0:0:0:0,0

# After all this, what does a variant calling pipeline look like?



e.g.  
population  
data

# Questions?

WHO? WHERE?  
WHEN? WHY? HOW?  
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WHEN?  
**WHAT?**  
WHO?  
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WHAT?  
WHEN?

# Today:

- 1) Introduction: variant calling, why do we want to do this, and what it is?
- 2) Variant calling pipelines/methods and pitfalls
- 3) Practical session, going through (parts of) a SNP calling pipeline and interpret biological results

