

lickcalc: Easy analysis of lick microstructure in experiments of rodent ingestive behaviour

K. Linnea Volcko¹ and James E. McCutcheon¹

¹ Dept. of Psychology, UiT The Arctic University of Norway, Tromsø, Norway  Corresponding author

DOI: [10.xxxxxx/draft](https://doi.org/10.xxxxxx/draft)

Software

- [Review](#)
- [Repository](#)
- [Archive](#)

Editor: [Open Journals](#)

Reviewers:

- [@openjournals](#)

Submitted: 01 January 1970

Published: unpublished

License

Authors of papers retain copyright and release the work under a Creative Commons Attribution 4.0 International License ([CC BY 4.0](#))

Summary

Lick microstructure is a term used to describe the information that can be obtained from a detailed study of when individual licks occur, when a rodent is drinking. Rather than simply recording total intake (volume consumed), lick microstructure examines how licks are grouped, and the spacing of these groups of licks. This type of analysis can provide important insight into why an animal is drinking, such as if it is driven by taste or as a response to the consequences of consumption (e.g. feeling "full"). The simplicity of using lickcalc, requiring only a drag-and-drop of files, will make microstructural analysis accessible to any who wish to use it while providing sophisticated analyses with high scientific value.

Statement of need

lickcalc is a software suite that performs microstructural lick analysis on data files containing timestamps of lick onset and offset, in MedAssociates or csv/txt format. In addition to providing an overview of individual files (corresponding to a particular session for a single subject), results of the analysis can be added to a table that is exportable to Excel.

Microstructural analysis was first described in John D. Davis & Smith (1992) and has since then been used understand diverse phenomena. In-depth reviews on many of these, and microstructural parameters used to study them, are available (Alexander W. Johnson, 2018; Naneix et al., 2020; Smith, 2001). Briefly, although much of the foundational work on drinking microstructure was on licking for nutritive solutions (e.g., sucrose solutions), microstructural analysis can also be used to study intake of water (McKay & Daniels, 2013; Santollo et al., 2021), ethanol (Patwell et al., 2021), and other tastants such as non-caloric artificial sweeteners, sodium and quinine (Lin et al., 2012; Spector & St. John, 1998; Verharen et al., 2019). Lick microstructure has been used to shed light on, for example, how licking is affected by neuropeptides (McKay & Daniels, 2013), enzymes in the mouth (Chometton et al., 2022), ovarian hormones (Santollo et al., 2021), nutrient restriction (Naneix et al., 2020), response to alcohol (Patwell et al., 2021), and diet (Alexander W. Johnson, 2012). The number of lick bouts over a session are thought to reflect post-ingestive feedback from the consumed fluid, whereas the number of licks in a bout are thought to reflect palatability of the solution.

Lick microstructure can provide a great deal of interesting information about why an animal is drinking. Often, changes in microstructure are accompanied by changes in total intake, but this is not always the case: sometimes, equal intake will be achieved by quite different licking patterns that indicate changes in orosensory and post-ingestive feedback (A. W. Johnson et al., 2010; Volcko et al., 2020). Analyzing lick microstructure is therefore highly valuable in understanding how a manipulation affects appetite; if X causes an animal to feel more sated after drinking, that may lead to a different interpretation than if X were to reduce the palatability of the solution. Because of the value of microstructural data, many labs habitually

42 record and analyze it. There are many others, however, that have not yet begun collecting these
43 data. Investing in lickometers can be costly, but there are an increasing number of alternatives
44 to commercial products such as those produced by Med Associates. Several open-source
45 lickometer designs are now available (e.g. Frie & Khokhar (2024); Monfared et al. (2024);
46 Petersen et al. (2024); Silva et al. (2024); Raymond et al. (2018)).

47 Recording individual licks with high temporal resolution is necessary for microstructural analysis
48 of drinking behavior, but another barrier to reporting microstructure is its analysis. This
49 problem can now easily be solved by `lickcalc`. `lickcalc` does not require any special software
50 or coding knowledge: all the user has to do is drag a file with timestamps of lick onset
51 (and, ideally, offset) into the program and `lickcalc` will generate a detailed microstructural
52 analysis, with a high degree of user control over key parameters. After loading a file and
53 setting parameters, `lickcalc` displays results, the detail of which are an additional advantage
54 of using the program. Tables show values for number of licks, number of bursts, and burst
55 size (among others) – the values that are often reported and used to draw inferences about
56 postigestive and orosensory feedback of the solution. But importantly, there are several charts
57 displayed that show information that helps with quality control of the data and challenges the
58 user to think critically about which parameters they have chosen. In short, `lickcalc` makes
59 microstructural analysis accessible to any who have timestamps of lick onsets, while providing
60 a lot of information on the data underlying the analysis which helps in choosing the appropriate
61 parameters.

62 Key features

63 `lickcalc` has several features that make it exceptionally user-friendly at the same time as
64 providing a sophisticated and detailed microstructural analysis. Some of these features include:

- 65 ■ **ease of use:** a file in Med Associates or CSV/TXT format simply needs to be dragged
66 into the `lickcalc` software in order to perform the analyses. Parameters can be set
67 manually using sliders, and results exported to Excel with the push of a button.
- 68 ■ **flexibility:** the user sets key parameters appropriate for their experimental setup and
69 data. Data can be examined by whole-session or different epochs.
- 70 ■ **customization:** by using the configuration file, users can change default settings to match
71 their preferences and avoid manually changing settings for each file loaded.
- 72 ■ **results compilation:** data from multiple sessions and/or individuals can be exported into
73 a single Excel file, which streamlines the analysis process.
- 74 ■ **detail of analysis:** one of the benefits of using `lickcalc` is the level of detail it provides.
75 In addition to the properties often reported (e.g. burst number, burst size), `lickcalc`
76 computes and displays attributes of the data that are important in establishing the quality
77 of the data and determining appropriate parameters for its microstructural analysis. Four
78 charts are:

- 79 1) *intraburst lick frequency*, or how often certain interlick intervals within a burst of
80 licking occur. While a rodent is licking, its tongue makes rhythmic protrusions that
81 are under the control of a central pattern generator (Travers et al., 1997). Rats
82 typically lick 6–7 times per second (John D. Davis & Smith, 1992), while mice
83 lick at a slightly higher rate (A. W. Johnson et al., 2010). In addition to these
84 species differences, there are also strain differences (A. W. Johnson et al., 2010; ?).
85 Because intraburst lick rate is under the control of the central pattern generator, it
86 should remain relatively stable across mice and conditions (unless a manipulation is
87 expected to cause changes in the central pattern generator). A typical chart for a
88 mouse might show a sharp peak around an intra-burst ILI of around 129, which
89 corresponds to a lick rate of 7.75 Hz. Much smaller peaks are often present at
90 the harmonics of the intra-burst ILI (e.g., a primary peak at 129 will have smaller

- 91 peaks at 258, 387, and so on), often because of “missed licks” in which the mouse
 92 attempts to lick but its tongue misses the spout. A large number of these, or
 93 other differences from the expected pattern of results, may indicate problems with
 94 the experimental setup (e.g. if the animal fails to reach the spout frequently, then
 95 perhaps the spout is too far away).
- 96 2) *lick length* is only available when lick offset is included in the data file. As with
 97 intraburst lick frequency, lick length should show little variability and the graph
 98 will have a sharp peak. Occasionally a lickometer will register longer licks than
 99 normal. This may be because a “lick” is registered by something other than a
 100 tongue, such as if a rodent grabs the spout with its paws, or if a fluid droplet
 101 hangs between the spout and the cage and is thus able to complete the electrical
 102 circuit. Concerns about data quality may be warranted with increasing number and
 103 duration of long licks. `lickcalc` displays both the number and maximum duration
 104 (s) of licks above the threshold that the user has set. There is also an option to
 105 remove these problematic licks from the dataset.
- 106 3) *burst frequency*, or how often certain burst sizes occurred. This is informative
 107 because burst size, by virtue of being a mean (mean licks per burst), does not
 108 take into account potentially relevant information about the distribution of how
 109 many licks are in a burst. For example, a burst size of 80 could result from bursts
 110 all containing between 70 and 90 licks, or from many single licks and one or two
 111 burst with a lot of licks. The latter case might raise some questions about how
 112 reliable the burst size value is. Although single licks occur, they can also be caused
 113 by non-tongue contact with the lickometer. Changing the minimum licks/burst
 114 parameter can perhaps filter out some of these suspect “licks.”
- 115 4) *Weibull probability*. The Weibull analysis, as described in J. D. Davis (1996), uses
 116 a mathematical equation to fit the data to a survival function. Although used by
 117 some (Aja et al., 2001; Moran et al., 1998; Spector & St. John, 1998), it is still
 118 relatively rare to find Weibull probabilities in microstructural analyses. The Weibull
 119 function can be used on several aspects of data, such as lick rate across a session,
 120 but in the `lickcalc` program the Weibull probability is calculated for burst size.
 121 It plots the probability that, given n licks, the mouse will continue to lick. This
 122 makes it sensitive to the number of licks per burst parameter that is set by the
 123 user. The Weibull alpha means.... And the Weibull beta means...

124 Design and usage

125 `lickcalc` is hosted by UiT The Arctic University of Norway and can be accessed at <https://lickcalc.uit.no>. Alternatively, it can be installed locally following instructions in the repository. To
 126 use `lickcalc`, the user drags in a file in Med Associates or CSV/TXT format and indicates
 127 which column contains the lick onsets and, if applicable, the lick offsets. A plot is automatically
 128 generated that displays a histogram of licks across the session. Session length defaults to the
 129 time of the last licks. This can be manually changed, or set in the optional “config file”. Session
 130 length can be set in seconds, minutes, or hours. The bin size (licks per unit of time) can be
 131 changed manually or in the config file. The user can toggle between the default histogram and
 132 a plot showing cumulative licks.

133 A microstructural analysis is, in essence, a division of individual licks into groups of licks. To
 134 perform this grouping, the user must set several parameters. One of these is the inter-lick
 135 interval (ILI), which is the minimum amount of time licks must be separated by in order to be
 136 considered separate groups. Early studies identified ILI of 251-500 ms as separating “bursts”
 137 of licking, and pauses of more than 500 ms as separating “clusters” of licking (i.e. a cluster
 138 of licks is made up of several bursts of licking). Others have argued that ILI of 1 s better
 139 reflects separation of lick bursts (Spector & St. John, 1998). In `lickcalc`, the user may set

141 the ILI between 250 ms and 2 s. Another parameter that needs to be decided prior to the
142 lick analysis is the minimum number of licks per burst. `lickcalc` allows between 1 and 5 licks.
143 The appropriate number of minimum licks/burst may vary depending on experimental set up,
144 and the likelihood that a single lick represents a lick rather than, for example, a tail touching
145 the spout. Finally, in `lickcalc`, the user must set a “long-lick threshold” between 0.1 and 1 s.
146 This parameter is only available when lick offset is included. Licks that are longer than the set
147 threshold are counted as “long” and may indicate a problem (e.g. the mouse holding the spout
148 with its paws) rather than a true lick. The user can decide whether to remove “long licks”
149 or not. All of these parameters can be set manually or through the config file. Four plots
150 are generated (see Key Features section above), and tables are displayed showing values of
151 several properties: total licks, intraburst frequency, number of long licks, maximum duration
152 of long licks, number of bursts, mean licks per burst, weibull alpha, weibull beta, and weibull
153 r-squared.

154 To save these data, the user has two options. The first is to export an excel file for the
155 given data file. The user sets the animal ID and chooses which data should be exported
156 in addition to the summary of the microstructural analysis. Data for “session histogram,”
157 “intraburst frequency,” “lick length,” “burst histogram,” “burst probability,” and “burst details”
158 are available. These allow the user to recreate the plots displayed in `lickcalc`. The second
159 option for saving the data is to add the microstructural data to “results summary” table in
160 the app. The results in this table remain even as new data files are loaded, so the data from
161 many sessions (and/or individual animals) can be exported into a single Excel file. In addition
162 to the data from the whole session, the user can choose to divide the session into epochs,
163 or to examine only the first n bursts, or perform a trial-based analysis (e.g., for Davis rig
164 experiments). Each of these analysis epochs can be added to the results summary table. The
165 table contains the data as well as the parameters (e.g., minimum burst size) used to generate
166 them. Finally, a batch process feature is available allowing multiple files to be analysed using
167 the same parameters.

168 Acknowledgements

169 We acknowledge contributions from colleagues in the field of ingestive behaviour who have
170 thought deeply about the meaning of patterns of licking. In particular, the following have
171 either contributed data for us to test or have advised on the design of the program and analysis:
172 (in alphabetical order) Derek Daniels, Samantha Fortin, Kevin Myers, Jess Santollo, Lindsey
173 Schier, Alan Spector.

174 References

- 175 Aja, S., Schwartz, G. J., Kuhar, M. J., & Moran, T. H. (2001). Intracerebroventricular CART
176 peptide reduces rat ingestive behavior and alters licking microstructure. *American Journal
177 of Physiology-Regulatory, Integrative and Comparative Physiology*, 280(6), R1613–R1619.
<https://doi.org/10.1152/ajpregu.2001.280.6.R1613>
- 178 Chometton, S., Jung, A.-H., Mai, L., Dal Bon, T., Ramirez, A. O., Pittman, D. W., & Schier, L.
179 A. (2022). A glucokinase-linked sensor in the taste system contributes to glucose appetite.
180 *Molecular Metabolism*, 64, 101554. <https://doi.org/10.1016/j.molmet.2022.101554>
- 181 Davis, J. D. (1996). Deterministic and probabilistic control of the behavior of rats ingesting
182 liquid diets. *American Journal of Physiology-Regulatory, Integrative and Comparative
183 Physiology*, 270(4), R793–R800. <https://doi.org/10.1152/ajpregu.1996.270.4.R793>
- 184 Davis, John D., & Smith, G. P. (1992). Analysis of the microstructure of the rhythmic tongue
185 movements of rats ingesting maltose and sucrose solutions. *Behavioral Neuroscience*,
186 106(1), 217–228. <https://doi.org/10.1037/0735-7044.106.1.217>

- 188 Frie, J. A., & Khokhar, J. Y. (2024). FARESHARE: An open-source apparatus for assessing
189 drinking microstructure in socially housed rats. *NPP—Digital Psychiatry and Neuroscience*,
190 2(1), 1. <https://doi.org/10.1038/s44277-024-00002-z>
- 191 Johnson, Alexander W. (2012). Dietary manipulations influence sucrose acceptance in diet
192 induced obese mice. *Appetite*, 58(1), 215–221. <https://doi.org/10.1016/j.appet.2011.09.015>
- 194 Johnson, Alexander W. (2018). Characterizing ingestive behavior through licking microstructure:
195 Underlying neurobiology and its use in the study of obesity in animal models. *International
196 Journal of Developmental Neuroscience: The Official Journal of the International Society for
197 Developmental Neuroscience*, 64, 38–47. <https://doi.org/10.1016/j.ijdevneu.2017.06.012>
- 198 Johnson, A. W., Sherwood, A., Smith, D. R., Wosiski-Kuhn, M., Gallagher, M., & Holland, P.
199 C. (2010). An analysis of licking microstructure in three strains of mice. *Appetite*, 54(2),
200 320–330. <https://doi.org/10.1016/j.appet.2009.12.007>
- 201 Lin, J.-Y., Amodeo, L. R., Arthurs, J., & Reilly, S. (2012). Taste neophobia and palatability:
202 The pleasure of drinking. *Physiology & Behavior*, 106(4), 515–519. <https://doi.org/10.1016/j.physbeh.2012.03.029>
- 204 McKay, N. J., & Daniels, D. (2013). Glucagon-like peptide-1 receptor agonist administration
205 suppresses both water and saline intake in rats. *Journal of Neuroendocrinology*, 25(10),
206 929–938. <https://doi.org/10.1111/jne.12086>
- 207 Monfared, M., Mascret, Q., Marroquin-Rivera, A., Blanc-Árabe, L., Lebouleux, Q., Lévesque,
208 J., Gosselin, B., & Labonté, B. (2024). High-throughput low-cost digital lickometer system
209 for the assessment of licking behaviours in mice. *Journal of Neuroscience Methods*, 410,
210 110221. <https://doi.org/10.1016/j.jneumeth.2024.110221>
- 211 Moran, T. H., Katz, L. F., Plata-Salaman, C. R., & Schwartz, G. J. (1998). Disordered
212 food intake and obesity in rats lacking cholecystokinin A receptors. *American Journal
213 of Physiology-Regulatory, Integrative and Comparative Physiology*, 274(3), R618–R625.
214 <https://doi.org/10.1152/ajpregu.1998.274.3.R618>
- 215 Naneix, F., Peters, K. Z., & McCutcheon, J. E. (2020). Investigating the Effect of Physiological
216 Need States on Palatability and Motivation Using Microstructural Analysis of Licking.
217 *Neuroscience*, 447, 155–166. <https://doi.org/10.1016/j.neuroscience.2019.10.036>
- 218 Patwell, R., Yang, H., Pandey, S. C., & Glover, E. J. (2021). An operant ethanol self-
219 administration paradigm that discriminates between appetitive and consummatory behaviors
220 reveals distinct behavioral phenotypes in commonly used rat strains. *Neuropharmacology*,
221 201, 108836. <https://doi.org/10.1016/j.neuropharm.2021.108836>
- 222 Petersen, N., Adank, D. N., Quan, Y., Edwards, C. M., Hallal, S. D., Taylor, A., Winder, D. G.,
223 & Doyle, M. A. (2024). A Novel Mouse Home Cage Lickometer System Reveals Sex- and
224 Housing-Based Influences on Alcohol Drinking. *eNeuro*, 11(10), ENEURO.0234–24.2024.
225 <https://doi.org/10.1523/ENEURO.0234-24.2024>
- 226 Raymond, M. A., Mast, T. G., & Breza, J. M. (2018). An open-source lickometer and
227 microstructure analysis program. *HardwareX*, 4, e00035. <https://doi.org/10.1016/j.hwx.2018.e00035>
- 229 Santollo, J., Edwards, A. A., Howell, J. A., & Myers, K. E. (2021). Bidirectional effects
230 of estradiol on the control of water intake in female rats. *Hormones and Behavior*, 133,
231 104996. <https://doi.org/10.1016/j.yhbeh.2021.104996>
- 232 Silva, A., Carriço, P., Fernandes, A. B., Saraiva, T., Oliveira-Maia, A. J., & Da
233 Silva, J. A. (2024). High-Precision Optical Fiber-Based Lickometer. *Eneuro*, 11(7),
234 ENEURO.0189–24.2024. <https://doi.org/10.1523/ENEURO.0189-24.2024>
- 235 Smith, G. P. (2001). John Davis and the meanings of licking. *Appetite*, 36(1), 84–92.

- 236 <https://doi.org/10.1006/appc.2000.0371>
- 237 Spector, A. C., & St. John, S. J. (1998). Role of taste in the microstructure of quinine
238 ingestion by rats. *American Journal of Physiology-Regulatory, Integrative and Comparative*
239 *Physiology*, 274(6), R1687–R1703. <https://doi.org/10.1152/ajpregu.1998.274.6.R1687>
- 240 Travers, J. B., Dinardo, L. A., & Karimnamazi, H. (1997). Motor and Premotor Mechanisms
241 of Licking. *Neuroscience & Biobehavioral Reviews*, 21(5), 631–647. [https://doi.org/10.1016/S0149-7634\(96\)00045-0](https://doi.org/10.1016/S0149-7634(96)00045-0)
- 243 Verharen, J. P. H., Roelofs, T. J. M., Menting-Henry, S., Luijendijk, M. C. M., Vanderschuren,
244 L. J. M. J., & Adan, R. A. H. (2019). Limbic control over the homeostatic need for sodium.
245 *Scientific Reports*, 9(1), 1050. <https://doi.org/10.1038/s41598-018-37405-w>
- 246 Volcko, K. L., Brakey, D. J., Przybysz, J. T., & Daniels, D. (2020). Exclusively drinking
247 sucrose or saline early in life alters adult drinking behavior by laboratory rats. *Appetite*,
248 149, 104616. <https://doi.org/10.1016/j.appet.2020.104616>

DRAFT