rawdata_normalization

```
## -- Attaching packages ------ tidyverse 1.3.1 --
## v ggplot2 3.3.5 v purrr 0.3.4
## v tibble 3.1.6 v dplyr 1.0.8
## v tidyr 1.2.0 v stringr 1.4.0
## v readr 2.1.2 v forcats 0.5.1
## -- Conflicts ------ tidyverse_conflicts() --
## x tidyr::extract() masks magrittr::extract()
## x dplyr::filter() masks stats::filter()
## x dplyr::lag() masks stats::lag()
## x purrr::set_names() masks magrittr::set_names()
Reading in data
# Reading in raw data
rawdata <-
   read.table(
      "Core_facility_results/data/gene_counts.tsv",
      header = TRUE,
      sep = "\t",
      check.names = FALSE
   )
## Removal of pre activated AMs
rawdata <-
 rawdata %>% dplyr::select(-c("16AMUntreated", "17AMUntreated", "19AMMtbAUX", "20AMMtbAUX"))
rownames(rawdata) <- rawdata[, 1]</pre>
# Reading in Gene info
gene.info <- read_tsv("Core_facility_results/data/gene_info.tsv")</pre>
# gene.info <- read.csv("Core_facility_results/data/gene_info.tsv", sep = "\t")
\# names(gene.info)[1:(ncol(gene.info)-1)] <- names(gene.info)[2:ncol(gene.info)]
# gene.info <- gene.info %>% rownames_to_column("Gene_ID")
#Changing first column to Gene ID for joining
colnames(gene.info)[1] <- "Gene_ID"</pre>
#Reading in Sample info
sample.info <-</pre>
```

```
read.csv("Core_facility_results/data/sample_info.tsv", sep = "\t")
## Removal of pre activated AMs from the metainfo
sample.info \leftarrow sample.info[-c(16, 17, 19, 20), ]
# Add condition colum to sample.info containing group and treatment
sample.info[, "Condition"] <-</pre>
   factor(paste(sample.info$Sample_Group,
                 sample.info$Treatment, sep = "."))
genes <- semi_join(gene.info, rawdata, by = "Gene_ID")</pre>
rawdata <- rawdata %>% dplyr::select(-Gene ID)
rawdata <- as.matrix(rawdata)</pre>
DGE Object
# Defining group for DGE object
group <- sample.info$Condition</pre>
# Creating dge object which will contain read counts, sample info and gene info
dge_object2 <- DGEList(rawdata, group = group)</pre>
#adding treatment to dge$samples
Treatment <- factor(sample.info$Treatment)</pre>
dge_object2$samples$Treatment <- Treatment</pre>
#adding cell type
Cell type <- factor(sample.info$Sample Group)</pre>
dge_object2$samples$Cell_type <- Cell_type</pre>
# Removing duplicate gene entries
genes <- genes[!duplicated(genes$Gene_ID), ]</pre>
# Adding gene info to dge object
dge_object2$genes <- genes
# For later use
samplenames <- colnames(rawdata)</pre>
# Removing duplicate gene entries
genes <- genes[!duplicated(genes$Gene_ID),]</pre>
# Adding gene info to dge object
dge_object2$genes <- genes</pre>
Filtering low counts
# Creating a model matrix without intercept
mm <- model.matrix(~0 + group)</pre>
# Naming the columns in the model matrix
colnames(mm) <- gsub("group", "", colnames(mm))</pre>
```

```
# Finding genes to remove using edgeR flterByExpr()
keep.exprs <- filterByExpr(dge_object2, mm)
dge_object2 <- dge_object2[keep.exprs,, keep.lib.sizes=FALSE]</pre>
```

Calculating normalization factors

```
unormalized_dge <- dge_object2

# TMM normalization
dge_object2 <- calcNormFactors(dge_object2, method = "TMM")

dge_object2$samples</pre>
```

```
##
                            group lib.size norm.factors Treatment Cell_type
## 1iMACUntreated iMACs.Untreated 15998962
                                             1.0891601 Untreated
                                                                     iMACs
## 2iMACUntreated iMACs.Untreated 17409260
                                             1.1559715 Untreated
                                                                     iMACs
## 3iMACUntreated iMACs.Untreated 14740955
                                             1.1034883 Untreated
                                                                     iMACs
## 4iMACMtbAUX
                     iMACs.MtbAUX 18017027
                                            0.9419256
                                                          MtbAUX
                                                                     iMACs
## 5iMACMtbAUX
                     iMACs.MtbAUX 16224426 1.0040381
                                                          MtbAUX
                                                                     iMACs
## 6iMACMtbAUX
                    iMACs.MtbAUX 16440071
                                             0.7976455
                                                          MtbAUX
                                                                     iMACs
## 7iMACLPS
                        iMACs.LPS 19233878
                                             0.7138140
                                                             I.PS
                                                                     iMACs
                    MDM.Untreated 16222615 0.9473939 Untreated
                                                                       MDM
## 8MDMUntreated
## 9MDMUntreated
                    MDM.Untreated 17017126 0.9431496 Untreated
                                                                       MDM
                    MDM.Untreated 19136785
## 10MDMUntreated
                                             0.9304014 Untreated
                                                                       MDM
## 11MDMMtbAUX
                       MDM.MtbAUX 16783933
                                             0.9766687
                                                          MtbAUX
                                                                       MDM
## 12MDMMtbAUX
                       MDM.MtbAUX 19244732
                                                          MtbAUX
                                                                       MDM
                                             0.9203157
## 13MDMMtbAUX
                       MDM.MtbAUX 15293231
                                             0.9369498
                                                          MtbAUX
                                                                       MDM
## 14MDMLPS
                          MDM.LPS 15441454
                                             0.9400197
                                                             LPS
                                                                       MDM
## 15AMUntreated
                     AM.Untreated 13566293
                                             1.0642814 Untreated
                                                                        ΔM
## 18AMMtbAUX
                        AM.MtbAUX 13066588
                                            1.0040790
                                                          MtbAUX
                                                                        ΑM
## 21AMLPS
                           AM.LPS 13983059
                                             1.0074879
                                                             LPS
                                                                        AM
## 22THP1Untreated THP1.Untreated 18334982
                                             1.2554687 Untreated
                                                                      THP1
## 23THP1LPS
                         THP1.LPS 14535895
                                                             I.PS
                                                                      THP1
                                             1.2765425
## 24THP1MtbAUX
                      THP1.MtbAUX 18204303
                                             1.1770835
                                                          MtbAUX
                                                                      THP1
```

```
# Counts per million
tmm <- cpm(dge_object2)

# Log Counts per million
## This is the normalized counts used for WGCNA analysis
norm_exp_matrix_am_rm <- cpm(dge_object2, log = TRUE, prior.count = 1)

norm_exp_matrix_am_rm_notlog <- cpm(dge_object2, log = FALSE, prior.count = 1)</pre>
```

Making different versions of the normalized data for use in other scripts

```
norm_exp_matrix_am_rm_notlog %>% as.data.frame() %>% rownames_to_column("Gene_ID")
avg_norm_exp_as_df_am_rm <- norm_exp_as_df_am_rm %>%
  mutate(
      "iMACs.Untreated" = rowMeans(norm exp as df am rm[2:4]),
      "iMACs.MtbAUX" = rowMeans(norm_exp_as_df_am_rm[5:7]),
      "MDM.Untreated" = rowMeans(norm exp as df am rm[9:11]),
      "MDM.MtbAUX" = rowMeans(norm exp as df am rm[12:14])
   ) %>%
  dplyr::select(c(-2:-7,-9:-14)) \%>\%
  relocate(10:13, .after = 1) %>% relocate(6, .after = 3)
colnames(avg_norm_exp_as_df_am_rm) <- c("Gene_ID", as.vector(unique(group)))</pre>
#notlog
avg_norm_exp_as_df_am_rm_notlog <- norm_exp_as_df_am_rm_notlog %>%
  mutate(
      "iMACs.Untreated" = rowMeans(norm_exp_as_df_am_rm_notlog[2:4]),
      "iMACs.MtbAUX" = rowMeans(norm_exp_as_df_am_rm_notlog[5:7]),
      "MDM. Untreated" = rowMeans(norm_exp_as_df_am_rm_notlog[9:11]),
      "MDM.MtbAUX" = rowMeans(norm_exp_as_df_am_rm_notlog[12:14])
   dplyr::select(c(-2:-7,-9:-14)) \%>\%
   relocate(10:13, .after = 1) %>% relocate(6, .after = 3)
colnames(avg_norm_exp_as_df_am_rm_notlog) <- c("Gene_ID", as.vector(unique(group)))</pre>
z_transformed_norm_exp_am_rm <-</pre>
    t(scale(
      t(
        norm_exp_as_df_am_rm %>% as.tibble() %>% column_to_rownames(var = "Gene_ID") %>%
          as.matrix()
      )
    ))
  z_transformed_avg_norm_exp_am_rm <-</pre>
    t(scale(
      t(
        avg_norm_exp_as_df_am_rm %>% as.tibble() %>%
          column_to_rownames(var = "Gene_ID") %>%
          as.matrix()
      )
    ))
```