Section II

The Structure of Macromolecules: *Human muscarinic M3 and bovine rhodopsin sequences alignment - tutorial*

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Objective

To learn how to use T-Coffee for sequences alignment.

Brief info

A sequence alignment consists in the primary sequences arrangement of proteins, DNA or RNA, in order to identify regions of similarity that may be a result of functional, structural, or evolutionary relationships between those sequences. Aligned sequences are typically represented as rows within a matrix, and identical or similar characters must be aligned in successive columns. Gaps may be inserted between the residues if it is necessary. There are several web-based programs that can be used for sequences alignment. The main ones are <u>BLAST</u>^{1,2}, <u>ClustalW</u>, and <u>T-Coffee</u>³.

T-Coffee runs by assembling a library - a list of potential pairs of residues and then turns this library into an alignment. Each library line is a constraint and the purpose is to assemble the alignment that accommodates the more all the constraints.

For this tutorial, the human muscarinic M3 (hM3) and bovine rhodopsin (RHO) sequences were chosen to be aligned using T-Coffee. The bovine rhodopsin is the only protein belonging to the same family with hM3 receptor (G-protein coupled receptors, GPCR) having the 3D structure available from experimental studies (X-RAY or RMN studies).

How to

- 1. Obtain the proteins sequences
 - go to <u>Swiss-PROT</u>^{4,5} (the protein sequence database).
 - enter the protein name (e.g. muscarinic M3 receptor) in the query field see Figure 1; you will be directed to a new page where you can find the results of your query.

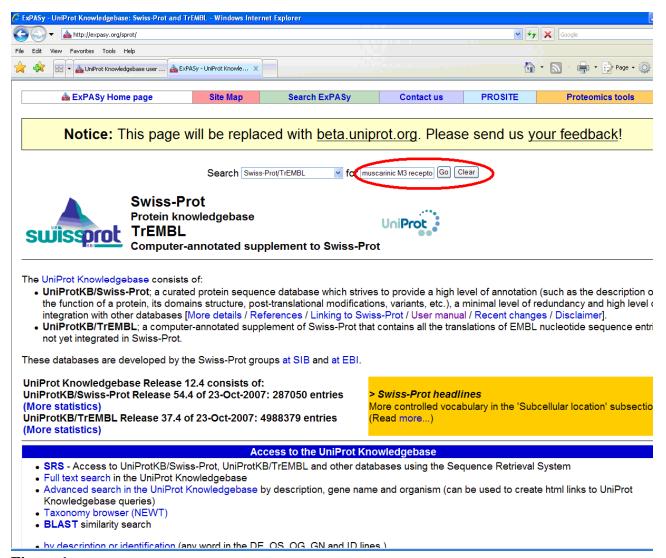


Figure 1

• select/open the record for the human M3 sequence (SWISS PROT ID: P20309) – see Figure 2. At the end of this page, in the sequence section, you have the option to view the protein sequence as FASTA format – see Figure 3.

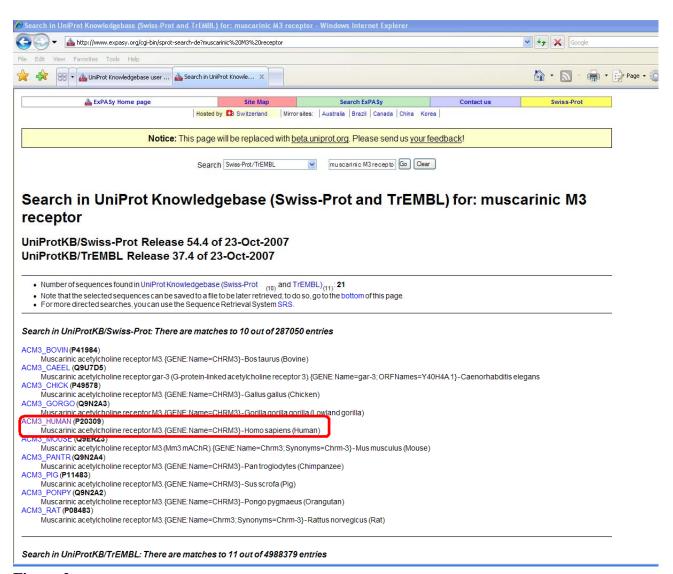


Figure 2

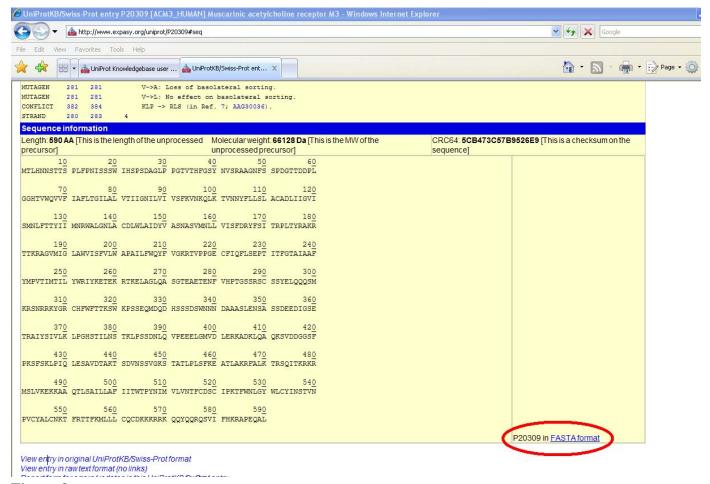


Figure 3

- save the sequence (in FASTA format) as a text file.
- repeat the above-given steps for bovine rhodopsin and save the sequence OPSD_BOVIN (SWISS PROT ID: P02699)
- merge the two text files into a single one and save it as "sequences.txt"

2. Align the proteins sequences

• access the T-Coffee server

Note that the following five modules are available for proteins sequences alignment: (i) T-Coffee, (ii) Expresso (it replaces 3DCoffee), (iii) M-Coffee, (iv) Rcoffee (beta version), and (v) Combine. *T-Coffee* computes a multiple sequence alignment and the associated phylogenetic tree. *Expresso* computes **structure** based multiple sequence alignments by running a BLAST alignment between every sequence in the query against the PDB database. If it finds one structure similar enough to a sequence in your dataset (>60% identity), it will use it as a template for your sequence. *M-Coffee* computes a multiple sequence alignment and the associated phylogenetic tree by combining the output of several multiple sequence alignment packages (PCMA, Poa, Mafft, Muscle, T-Coffee, ClustalW, ProbCons, DialignT). *Rcoffee* computes multiple sequence alignment of **non coding** RNA sequences using RNAplfold predicted secondary structures. *Combine* combines two (or more) multiple sequence alignments into a single one. Details about each of these modules can be found by following the link "cite" to the original papers.

• go to T-Coffee > Advanced and input the sequences in Fasta format. This can be done in two ways: (i) either upload the text file containing both sequences in Fasta format (sequences.txt), or (ii) paste both proteins sequences in Fasta format (no empty line between the sequences). Keep the default options for the "Alignment computation" and the "Output", as can be seen in Figure 4:

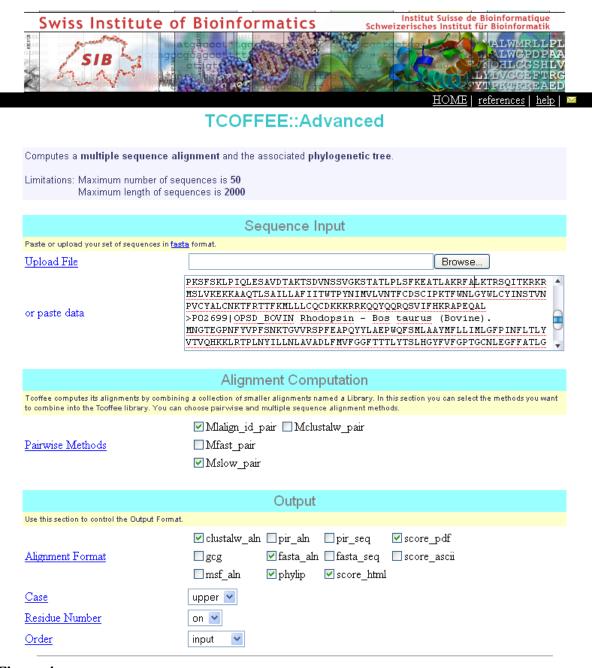


Figure 4

• press the submit button and wait until the sequence alignment will be computed. The results are given in different formats – see Figure 5. The high-similarity regions in the alignment are marked yellow/red in the HTML or PDF format.

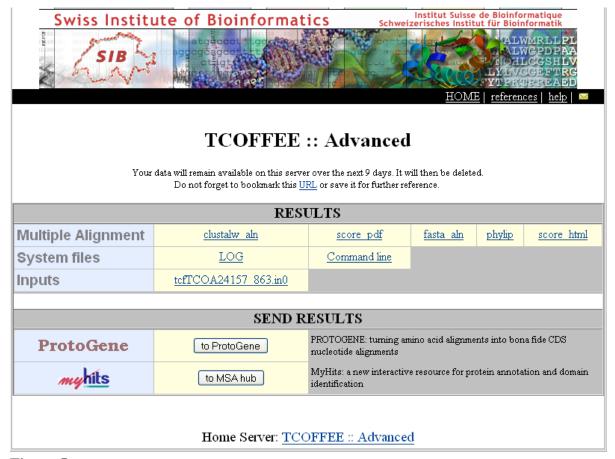


Figure 5

The sequence alignment is the most critical step in developing a homology model, for example a misalignment by just one residue will cause an error around 4Å in the model. For an accurate alignment the **structurally-conserved regions** (SCRs) have to be identified in both sequences. Structurally-conserved regions within a family proteins refer to the fragments for which an average structure or framework can be constructed for these regions of the proteins.

In rhodopsin-like family the highly conserved amino acids are⁶:

- ➤ Gly17, Asn 18 and Val21 on helix I,
- Asn or Ser9, Leu10, Ala11, Ala or Ser13, and Asp14 on helix II,
- > Ser 14, Leu 18, Ile 21, Ser or Ala22, Asp or Glu24, Arg 25, Tyr 26, Ile or Val29, on helix III,
- > Trp11, Ser or Ala14 and Pro20 on helix IV,
- ➤ Phe11, Pro14, Ile or Met18, Tyr22 and Ile or Val25 on helix V,
- Lys or Arg0, Phe12, Cys15, Trp16 and Pro18 on helix VI,
- Asn or Ser13, Ser or Cys14, Asn or Asp17, Pro18, Tyr21, Phe or Tyr28 and Arg or Lys29 on helix VII.

After highly conserved residues identification in the alignment (yellow background in figure 6) one can observe that T-coffee fails to find the common motif in the helices 5 (green background in figure 6). This is due to the fact that the third intracellular loops have obviously different lengths (218 residues in M3 receptor versus 25 residues in rhodopsin). To avoid this inconvenient, a new alignment is necessary, this time using a short sequence for hM3 receptor (the short sequence does not contain all the amino acid residues from the third loop). This deletion will not have a bad influence on following studies (e.g. docking) because the binding site is

located in the transmembranar region. In Figure 6 bold letters indicate the transmembranar domains; the fifth transmembrane is pointed out with bold and italic letters.

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CLUSTAL FORMAT for T-COFFEE Version_5.13 [http://www.tcoffee.org], CPU=0.09 sec, SCORE=63, Nseq=2, Len=595
P02699|OPSD_BOVIN ----MNGTEGPNF-----YVPFSNKTGV------VRSPFEAPQYYLAEP- 34
P20309 ACM3_HUMAN MTLHNNSTTSPLFPNISSSWIHSPSDAGLPPGTVTHFGSYNVSRAAGNFSSPDGTTDDPL 60
                    *.* .* *
                              :: ...:*:
                             TM I
                                                          TM II
P02699 OPSD_BOVIN -----WQFSMLAAYMFLLIMLGFPINFLTLYVTVQHKKLRTPLNYILLNLAVADLFMVFG 89
P20309 ACM3_HUMAN GGHTVWQVVFIAFLTGILALVTIIGNILVIVSFKVNKQLKTVNNYFLLSLACADL--IIG 118
                   **. ::* :* :: *:*.: :*:*:* **:** **: ::*
                                              TM III
P02699 OPSD_BOVIN GFTTTLYTS--LHGYFVFGPTGCNLEGFFATLGGEIALWSLVVLAIERYVVVCKPMS-NF 146
P20309 ACM3_HUMAN VISMNLFTTYIIMNRWALGNLACDLWLAIDYVASNASVMNLLVISFDRYFSITRPLTYRA 178
                :: .*:*: : . :.:* .*:* : :..: :: .*::::**. : :*:: .
                           TM IV
P02699 OPSD_BOVIN RFGENHAIMGVAFTWVMALACAAPPLVGWSRYI------ 179
P20309 ACM3_HUMAN KRTTKRAGVMIGLAWVISFVLWAPAILFWQYFVGKRTVPPGECFIQFLSEPTITFGTAIA 238
                  ::* : :.::**:::. **.:: *. ::
P02699|OPSD BOVIN ------ 189
P20309|ACM3_HUMAN AFYMPVTIMTILYWRIYKETEKRTKELAGLQASGTEAETENFVHPTGSSRSCSSYELQQQ 298
P20309 ACM3_HUMAN SMKRSNRRKYGRCHFWFTTKSWKPSSEQMDQDHSSSDSWNNNDAAASLENSASSDEEDIG 358
P20309|ACM3_HUMAN SETRAIYSIVLKLPGHSTILNSTKLPSSDNLQVPEEELGMVDLERKADKLQAQKSVDDGG 418
               .*: .** :*:::
P02699|OPSD_BOVIN ------AQQQE 239
P20309|ACM3_HUMAN SFPKSFSKLPIQLESAVDTAKTSDVNSSVGKSTATLPLSFKEATLAKRFALKTRSQITKR 478
                                        : . . * ::.**:
                                TM VT
P02699 OPSD_BOVIN SATTQKAEKEVTRMVIIMVIAFLICWLPYAGVAFYIFTHQGSDFGPIFMTIPAFFAKT<mark>SA</mark> 299
P20309 ACM3_HUMAN KRMSLVKEKKAAQTLSAILLAFIITWTPY-NIMVLVNTFCDSCIPKTFWNLGYWLCYINS 537
                     **:.:: : :::**:* * ** .: . : * . : * .: ::. .:
            TM VII
P02699 OPSD_BOVIN VYNPVIYIMMNKQFRNCMVTTLCC-----GKNPLGDDEASTTVSKTETSQVAPA 348
P20309 ACM3_HUMAN TVNPVCYALCNKTFRTTFKMLLLCQCDKKKRRKQQYQQRQSVIFHKRAPEQA--L 590
               . *** * : ** **. : * * * :: ::. *. . * ..*.
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Figure 6

Therefore the improved alignment must be accomplished so that the SCRs from the two sequences are perfectly aligned. In the same time, each transmembrane alignment must not contain insertion or deletion at their level. If that happens, they have to be moved in loop regions for the sake of the accuracy of the alignment. As one can see from Figure 6 such operations have to be performed at helix 2 and 6 levels. In order to avoid the induction of perturbations it is better to eliminate the insertions before helix 1 and after helix 8. Also helices 5 must be manually realigned. Based on these modifications and by removing a part of CIII loop a new alignment is obtained – see Figure 7.

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CLUSTAL FORMAT for T-COFFEE Version_5.13 [http://www.tcoffee.org], CPU=0.07 sec, SCORE=64, Nseq=2, Len=390

TM I

P02699 | OPSD_BOVIN MNGTEGPNFYVPFSNKTGVVRSPFEAPQYYLAEPWQFSMLAAYMFLLIMLGFPINFLTLYVTV

P20309 | ACM3_HUMAN NNSTTSPLFWIHSPSDAGLAAGNFSSPDGTTDDPWQVVFIAFLTGILALVTIIGNILVIVSFK
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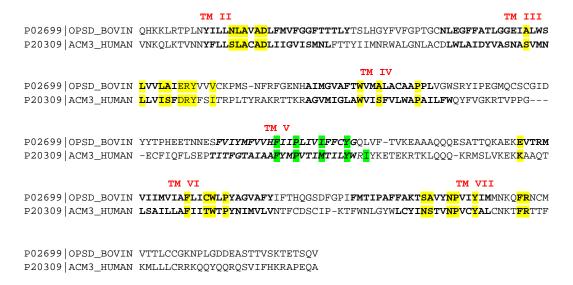


Figure 7

- compare your newly obtained short sequence of amino acids for hM3 receptor with this one [insert link]
- correct the errors
- save the corrected sequence as text file, i.e. h3M_short_seq.txt (!keep the first line from the initial text file!), because you will need it in the next section of the course.

Altschul S.F., Madden T.L., Schaffer A.A., Zhang J., Zhang Z., Miller W., Lipman D.J., *Nucleic Acids Res* 25 (1997) 3389-3402

Jaroszewski L., Rychlewski L., Zhang B., Godzik A., *Protein Sci* 7 (1998) 1431-1440

C. Notredame, D. Higgins, J. HeringaJournal of Molecular Biology, 302, 205-217, (2000)

Bairoch A., Apweiler R., *Nucleic Acids Res.*, 28 (**2000**) 45-48

⁵ http://www.expasy.org/sprot/sprot-top.html

Baldwin, J.M.; Schertler, G.F.X.; Unger, V.M. J Mol Biol 1997, 272, 144–164