Data Wrangling and Processing for Genomics - Parte 4

Evidencia:

Nueva carpeta y descargamos el genoma de referencia de E. coliREL606:

```
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop/data/trimmed_fastq$ cd ~/dc workshop
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ mkdir -p data/ref_genome estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ sudo curl -L -o data/ref_genome/ecoli_rel606.fasta
.gz ftp://ftp.ncbi.nlm.nih.gov/genomes/all/GCA/000/017/985/GCA 000017985.1 ASM1798v1/GCA 00001
7985.1 ASM1798v1 genomic.fna.gz
[sudo] password for estuardo8u14:
 % Total
          % Received % Xferd Average Speed
                                          Time
                                                 Time
                                                         Time Current
                            Dload Upload
                                                         Left Speed
                                          Total
                                                 Spent
100 1343k 100 1343k 0
                          0
                             625k
                                      0 0:00:02 0:00:02 --:-- 625k
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ gunzip data/ref_genome/ecoli_rel606.fasta.gz
estuardo8u14@LAPTOP-5IN4BIR3:~/dc workshop$ head data/ref genome/ecoli_rel606.fasta
>CP000819.1 Escherichia coli B str. REL606, complete genome
GTTACCTGCCGTGAGTAAATTAAAATTTTATTGACTTAGGTCACTAAATACTTTAACCAATATAGGCATAGCGCACAGAC
AGATAAAAATTACAGAGTACACAACATCCATGAAACGCATTAGCACCACCACTACCACCATCACCATTACCACAGGT
AACGAGGTAACAACCATGCGAGTGTTGAAGTTCGGCGGTACATCAGTGGCAAATGCAGAACGTTTTCTGCGGGTTGCCGA
CGATGATTGAAAAAACCATTAGCGGCCAGGATGCTTTACCCAATATCAGCGATGCCGAACGTATTTTTGCCGAACTTTTG
ACGGGACTCGCCGCCCAGCCGGGATTCCCGCTGGCGCAATTGAAAACTTTCGTCGATCAGGAATTTGCCCAAATAAA
ACATGTCCTGCATGGCATTAGTTTGTTGGGGGCAGTGCCCGGATAGCATCAACGCTGCGCTGATTTTGCCGTGGCGAGAAAA
```

Nuestro primer paso es indexar el genoma de referencia para que lo utilice BWA:

```
estuardo8u14@LAPTOP-5IN4BIR3:~/dc workshop$ sudo curl -L -o sub.tar.gz https://ndownloader.tig
share.com/files/14418248
             % Received % Xferd Average Speed
 % Total
                                                   Time
                                                           Time
                                                                     Time Current
                                 Dload Upload Total
                                                                     Left Speed
                                                           Spent
                                           0 --:--: 0:00:01 --:--:
                        0
                               0
       0
           0
                   0
                                    0
                             0 3492k
100 109M 100 109M
                        0
                                             0 0:00:32 0:00:32 --:-- 3599k
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ tar xvf sub.tar.gz
sub/SRR2584866 1.trim.sub.fastq
sub/SRR2589044 1.trim.sub.fastq
sub/SRR2589044 2.trim.sub.fasta
sub/SRR2584863 2.trim.sub.fastq
sub/SRR2584866_2.trim.sub.fastq
sub/SRR2584863 1.trim.sub.fastq
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ mv sub/ ~/dc_workshop/data/trimmed_fastq_small
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ mkdir -p results/sam results/bam results/bcf resul
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ bwa index data/ref genome/ecoli rel606.fasta
[bwa index] Pack FASTA... 0.15 sec
[bwa index] Construct BWT for the packed sequence...
[bwa index] 2.21 seconds elapse.
[bwa index] Update BWT... 0.06 sec
[bwa index] Pack forward-only FASTA... 0.02 sec
[bwa_index] Construct SA from BWT and Occ... 1.32 sec
main] Version: 0.7.17-r1188
[main] CMD: bwa index data/ref_genome/ecoli_rel606.fasta
main] Real time: 4.928 sec; CPU: 3.824 sec
```

Comenzaremos alineando las lecturas de solo una de las muestras en nuestro conjunto de datos (SRR2584866) y tras correr la línea bwa mem data/ref_genome/ecoli_rel606.fasta data/trimmed_fastq_small/SRR2584866_1.trim.sub.fastq data/trimmed_fastq_small/SRR2584866 2.trim.sub.fastq > results/sam/SRR2584866.aligned.sam :

```
M::mem pestat] analyzing insert size distribution for orientation FF...
[M::mem_pestat] (25, 50, 75) percentile: (235, 720, 1382)
[M::mem_pestat] low and high boundaries for computing mean and std.dev: (1, 3676)
[M::mem pestat] mean and std.dev: (770.19, 676.46)
[M::mem pestat] low and high boundaries for proper pairs: (1, 4823)
[M::mem_pestat] analyzing insert size distribution for orientation FR...
[M::mem_pestat] (25, 50, 75) percentile: (219, 350, 560)

[M::mem_pestat] low and high boundaries for computing mean and std.dev: (1, 1242)

[M::mem_pestat] mean and std.dev: (404.41, 221.74)

[M::mem_pestat] low and high boundaries for proper pairs: (1, 1583)
[M::mem_pestat] skip orientation RF as there are not enough pairs
[M::mem_pestat] analyzing insert size distribution for orientation RR...
[M::mem_pestat] (25, 50, 75) percentile: (325, 557, 721)
[M::mem pestat] low and high boundaries for computing mean and std.dev: (1, 1513)
[M::mem_pestat] mean and std.dev: (501.66, 309.62)
[M::mem_pestat] low and high boundaries for proper pairs: (1, 1909)
[M::mem_pestat] skip orientation FF
[M::mem_pestat] skip orientation RR
[M::mem_process_seqs] Processed 45746 reads in 3.633 CPU sec, 3.443 real sec
[main] Version: 0.7.17-r1188
[main] CMD: bwa mem data/ref genome/ecoli rel606.fasta data/trimmed fastq small/SRR2584866 1.t
rim.sub.fastq data/trimmed fastq small/SRR2584866 2.trim.sub.fastq
[main] Real time: 28.329 sec; CPU: 28.838 sec
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ samtools view -S -b results/sam/SRR2584866.aligned
.sam > results/bam/SRR2584866.aligned.bam
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ samtools sort -o results/bam/SRR2584866.aligned.so
rted.bam results/bam/SRR2584866.aligned.bam
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ samtools flagstat results/bam/SRR2584866.aligned.s
orted.bam
351169 + 0 in total (QC-passed reads + QC-failed reads)
0 + 0 secondary
1169 + 0 supplementary
0 + 0 duplicates
351103 + 0 mapped (99.98%: N/A)
350000 + 0 paired in sequencing
175000 + 0 read1
175000 + 0 read2
346688 + 0 properly paired (99.05% : N/A)
349876 + 0 with itself and mate mapped
58 + 0 singletons (0.02% : N/A)
0 + 0 with mate mapped to a different chr
0 + 0 with mate mapped to a different chr (mapQ>=5)
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ [
```

Llamado de variantes

Paso 1: Calcule la cobertura de lectura de posiciones en el genoma

```
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ bcftools mpileup -0 b -o results/bcf/SRR2584866_ra
w.bcf \
> -f data/ref_genome/ecoli_rel606.fasta results/bam/SRR2584866.aligned.sorted.bam
[mpileup] 1 samples in 1 input files
[mpileup] maximum number of reads per input file set to -d 250
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$
```

Paso 2: detectar los polimorfismos de un solo nucleótido (SNP)

Viendo los datos:

```
###IltRecDiptos_Noscription=All filters passed">
###IltRecDiptos_Noscription=All filters passed |
###IltRecDiptos_Noscription=All filtrecomposed |
###IltRecDiptos_Noscription=All filtr
```

Use the grep and wc commands you've learned to assess how many variants are in the vcf file.

Respuesta:

```
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ bcftools call --ploidy 1 -m -v -o results/bcf/SRR2584866_variants.vcf results/bcf/SRR2584866
6_raw.bcf
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ vcfutils.pl varFilter results/bcf/SRR2584866_variants.vcf > results/vcf/SRR2584866_final_v
ariants.vcf
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ less -S results/vcf/SRR2584866_final_variants.vcf
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$ grep -v "#" results/vcf/SRR2584866_final_variants.vcf | wc -l
767
estuardo8u14@LAPTOP-5IN4BIR3:~/dc_workshop$
```