Bio Resources at CARC

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University of Southern California
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Bio Resources (updated twice a year)

https://carc.usc.edu/user-information/bio-resources ENTER SEARCH TERMS USC | USC ITS | Office of the CIO | Office of Research USC | Advanced Research Computing
Enabling scientific breakthroughs at scale **User Suppor** Education & Outreach News & Events About Services User Information **Getting Started CARC User Portal** complete and comprehensive user Accounts And Allocations **CARC User Forum CARC OnDemand** System Information **Frequently Asked Questions User Guides** Condo Cluster Program **Brings Cryo-EM to USC** Phase 1 of the cryogenic electron microscopy (cryo-EM) project is now complete **Read More**

Bio Resources (updated twice a year)

User Information

Getting Started
Accounts and Allocations
System Information
User Guides
CARC User Portal
CARC User Forum
CARC OnDemand
Condo Cluster Program

Bio Resources

Using USC's Cryo-EM Instruments Frequently Asked Questions

Bio Resources

Reference genomes, protein and nucleotide sequences databases, and other bio resources are now available on Discovery and Endeavour. If you need a specific release that is not currently included in the pages below, please submit a help ticket and we will try to make those resources available to you.

Genomes

A set of ready-to-use reference sequences and annotations for commonly analyzed organisms, sourced from iGenomes.

Genbank

The GenBank sequence database is an open access, annotated collection of all publicly available nucleotide sequences and their protein translations.

Genome Taxonomy Database (GTDB)

The Genome Taxonomy Database (GTDB) is an initiative to establish a standardized microbial taxonomy based on genome phylogeny.

Pfam Database

The Pfam database is a large collection of protein families, each represented by multiple sequence alignments and hidden Markov models (HMMs).

TIGRFAMs

TIGRFAMs is a resource consisting of curated multiple sequence alignments, Hidden Markov Models (HMMs) for protein sequence classification, and associated information designed to support automated annotation of (mostly prokaryotic) proteins.

UniProt

The Universal Protein Resource (UniProt), a collaboration between the European Bioinformatics Institute (EBI), the SIB Swiss Institute of Bioinformatics, and the Protein Information Resource (PIR)

Bio Resources (updated twice a year)

https://carc.usc.edu/user-information/bio-resources

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- Genbank collection of all public nucleotide sequences and their protein translations
- Genome Taxonomy Database (GTDB) an initiative to establish a standardized microbial taxonomy based on genome phylogeny
- <u>Pfam Database</u> large collection of protein families, each represented by multiple sequence alignments and hidden Markov models (HMMs)
- <u>TIGRFAMs</u> curated multiple sequence alignments, Hidden Markov Models (HMMs) for protein sequence classification
- <u>UniProt</u> UniProtKB (curated protein information), UniRef (closely related sequences),
 UniParc (all protein sequences, consisting only of unique identifiers and sequences)

Bio Resources (through command line)

https://carc.usc.edu/user-information/bio-resources

- Biogeotraces set of metagenomes, collected under the auspices of the bioGEOTRACES component of the international GEOTRACES program
 /project/biodb/biogeotraces
- TaraOceans marine microbial metagenomes sampled across space and time
 /project/biodb/taraoceans
- Variant Effect Predictor cache VEP can use a variety of annotation sources to retrieve the
 transcript models used to predict consequence types. Cache contains all transcript models,
 regulatory features and variant data for a species and allows for an offline use of VEP
 /project/biodb/vep-cache

Log into CARC OnDemand

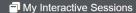
- Open the web browser and go to:
 - https://carc-ondemand.usc.edu
- Enter your username and password
- Choose an option in Duo-2FA, and confirm your access
- Done. You should see the CARC OnDemand dashboard

CARC OnDemand

iles▼

Jobs▼

ers ▼ Interactive Apps ▼





Logged in as osinski





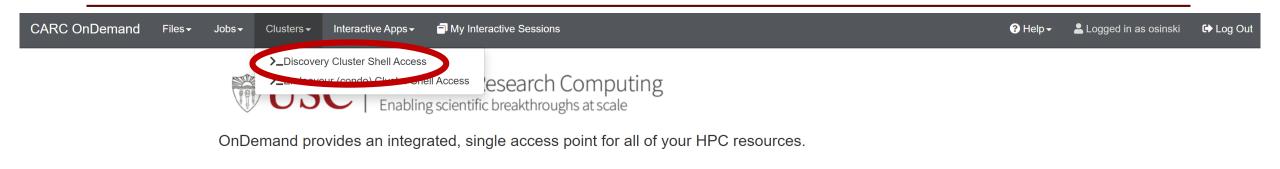
OnDemand provides an integrated, single access point for all of your HPC resources.

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OPEN ON Demand



OnDemand version: v1.8.18



powered by

OPEN ON Demand



OnDemand version: v1.8.18

```
Duo two-factor login for osinski
Enter a passcode or select one of the following options:
1. Duo Push to XXX-XXX-4132
 2. Phone call to XXX-XXX-4132
3. SMS passcodes to XXX-XXX-4132
Passcode or option (1-3): 1
Success. Logging you in...
Last login: Thu Mar 3 15:16:56 2022 from 10.21.74.187
         Welcome to the Center for Advanced Research Computing (CARC)
                at the University of Southern California (USC)
        CARC website : https://www.carc.usc.edu
       User support : https://www.carc.usc.edu/user-support
        User portal : https://hpcaccount.usc.edu
              ** Unauthorized use/access is prohibited **
If you log on to this computer system, you acknowledge your awareness of and
concurrence with the USC CARC Acceptable Use Policy. USC will prosecute
violators to the full extent of the law.
[osinski@discovery2 ~]$
```

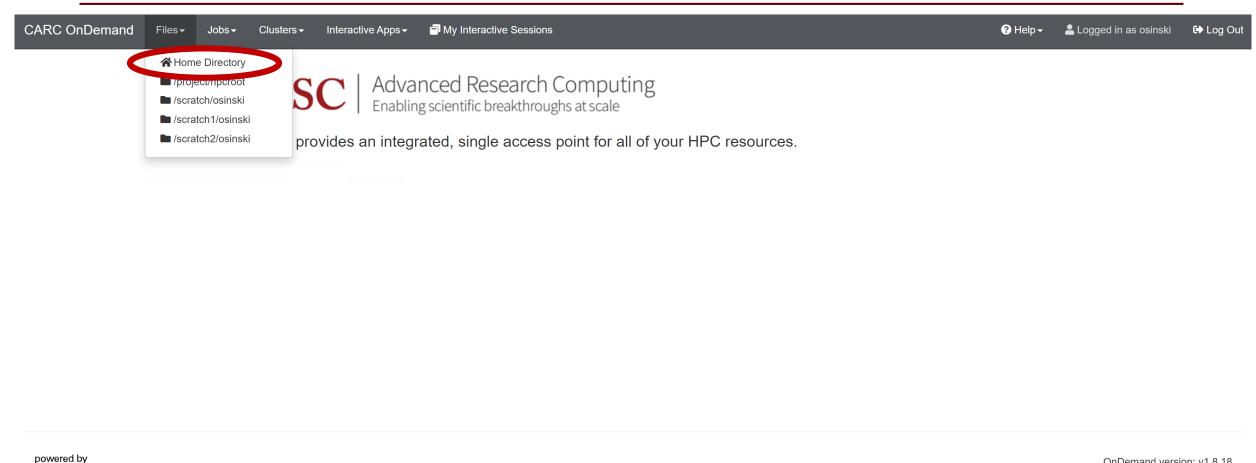
Copy the text below:

git clone https://github.com/uschpc/workshop-bioresources.git and paste it in the terminal by pressing Shift+Ins keys simultaneously, then press Enter



```
Duo two-factor login for osinski
Enter a passcode or select one of the following options:
1. Duo Push to XXX-XXX-4132
2. Phone call to XXX-XXX-4132
3. SMS passcodes to XXX-XXX-4132
Passcode or option (1-3): 1
Success. Logging you in...
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               ** Unauthorized use/access is prohibited **
If you log on to this computer system, you acknowledge your awareness of and concurrence with the USC CARC Acceptable Use Policy. USC will prosecute
violators to the full extent of the law.
[osinski@discovery2 ~]$ git clone https://github.com/uschpc/workshop-bioresources.git
Cloning into 'workshop-bioresources'...
remote: Enumerating objects: 22, done.
remote: Total 22 (delta 0), reused 0 (delta 0), pack-reused 22
Unpacking objects: 100% (22/22), done.
Checking out files: 100% (23/23), done.
[osinski@discovery2 ~]$
```

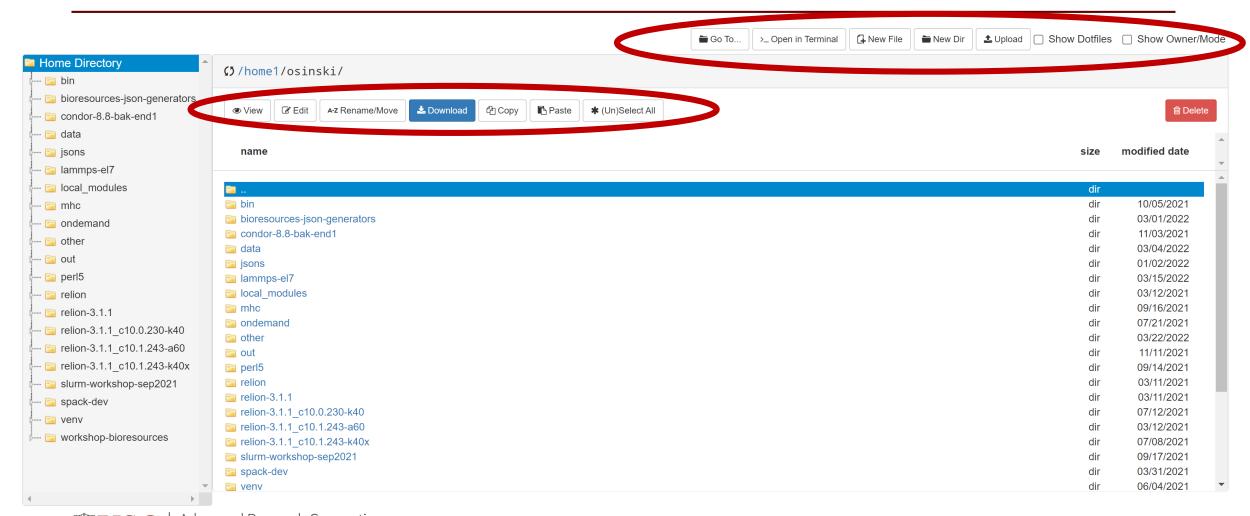
CARC OnDemand: File Management





OPEN On Demand

CARC OnDemand: File Management



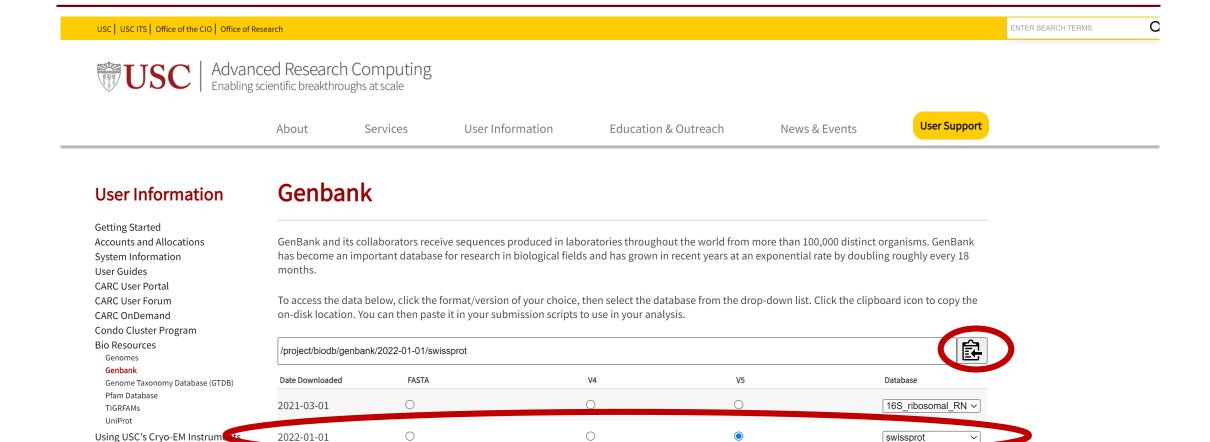
Create the BLAST Job Script

Replace swissprot with the path to the v5 of swissprot db obtained from

https://carc.usc.edu/user-information/bio-resources/genbank

```
#!/bin/bash
#SBATCH --nodes 1
#SBATCH --ntasks 10
#SBATCH --partition debug
#SBATCH --time 00:05:00
#SBATCH --account=ttrojan_001
#SBATCH --chdir /homel/ttrojan/workshop-bioresources
module purge
module load gcc/9.2.0
module load blast-plus
echo "Start BLAST Job"
blastp -db swissprot -query blast/query.txt -out results/blast/results.txt -num_threads
$SLURM_NTASKS
echo "Finish BLAST Job"
```

Create the BLAST Job Script





Frequently Asked Questions

CARC OnDemand: Job Composer



OnDemand provides an integrated, single access point for all of your HPC resources.

powered by

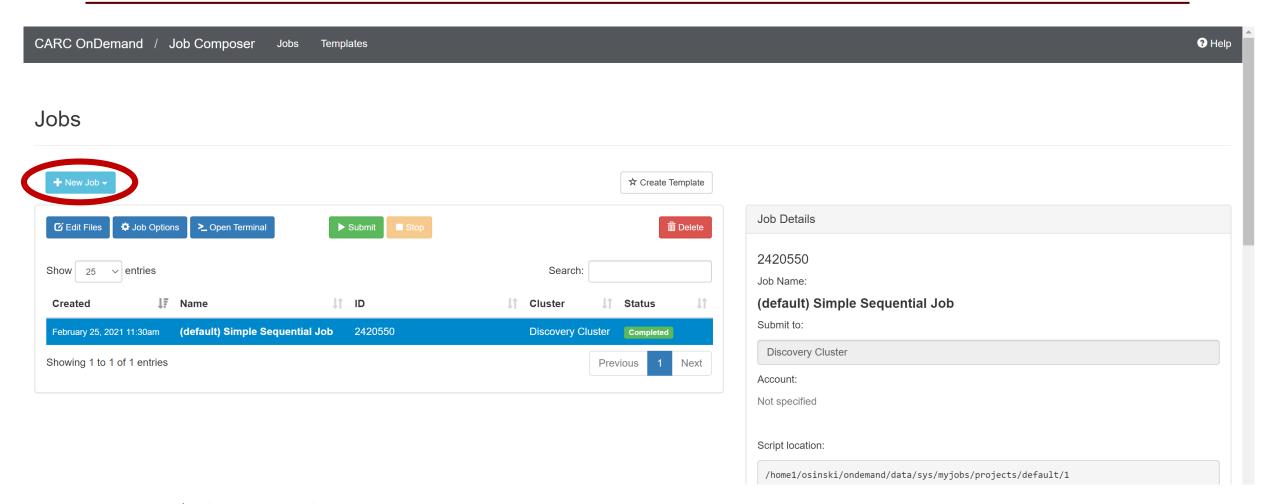
OPEN ONDemand

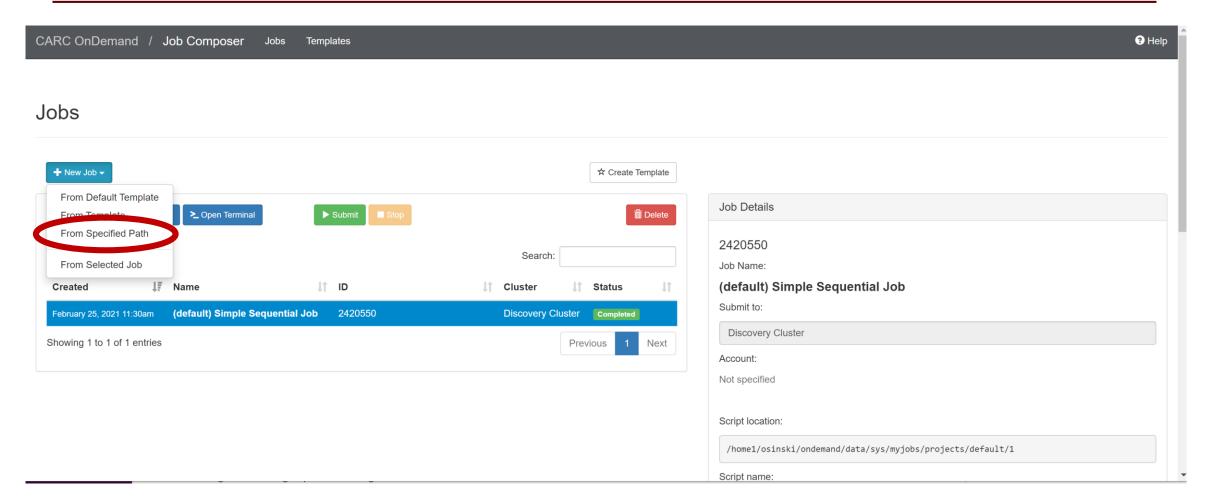


OnDemand version: v1.8.18

🚣 Logged in as osinski

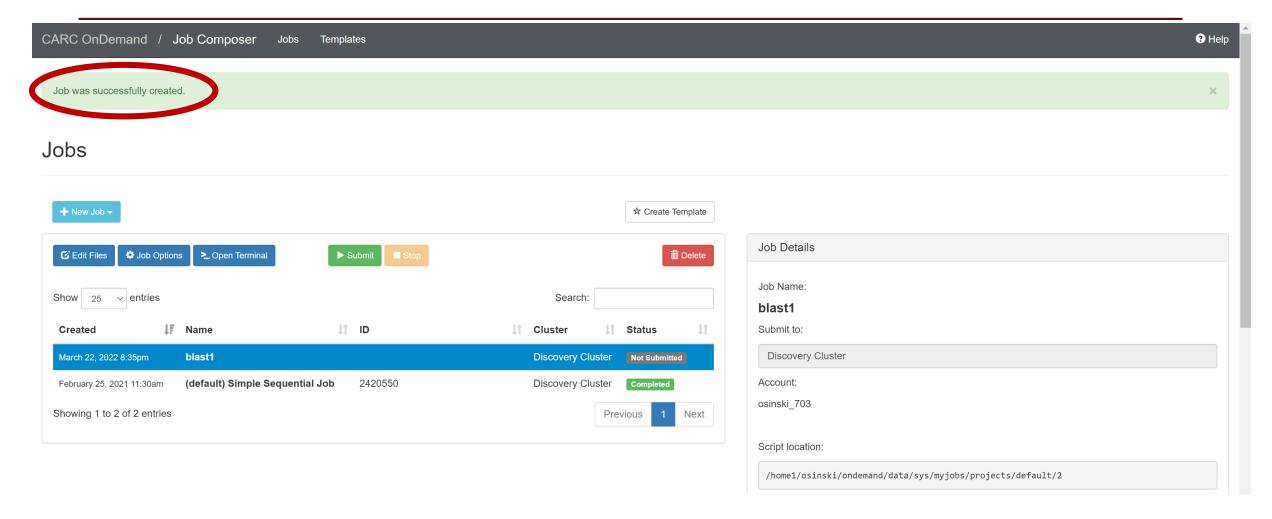
C→ Log Out





Create a new job from a path Path to source (Required) Source path /home1/osinski/workshop-bioresources/jobs/ Enter the path to a directory on the file system. The contents of this path will be copied to a new workflow. Job Attributes (Optional) Name blast1 Script name blast1.job Cluster: **Discovery Cluster** Account osinski_703 Account is an optional field. If not set, the account may be auto-set by the submit filter.

Back



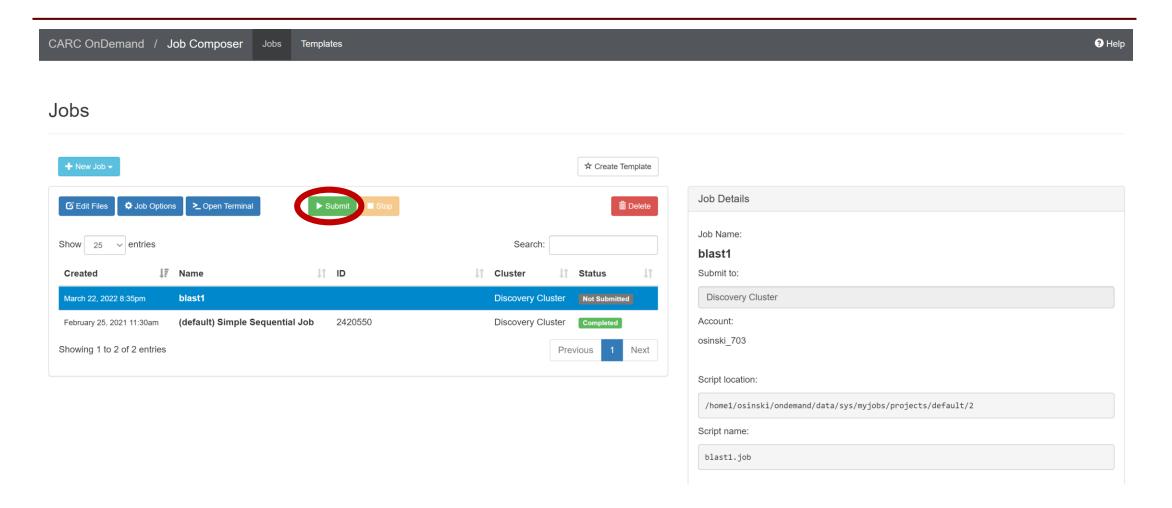
CARC OnDemand: Editing a Job

blast1.job					
fastqc1.job					
fastqc2.job					
fastqc_numbe	ered_array.job				
fastqc_unnun	nbered_array.job				
Submit Script					
olast1.job					
#!/bin/bash #SBATCHno: #SBATCHnt: #SBATCHti: #SBATCHti: #SBATCHti	asks 10 htition debug me=00:05:00 count=ttrojan_001	/workshop-bioresource	s		
#!/bin/bash #SBATCHno: #SBATCHnt: #SBATCHti: #SBATCHcti: #SBATCHcci: module purge module load ; module load ; echo "Example sleep 20	esks 10 rtition debug ne=00:05:00 count=ttrojan_001 dir /home1/ttrojan gcc/9.2.0 plast-plus p blast start"	/workshop-bioresource ank/2021-03-01/swissp		a/blast/query.t	∴xt -ou

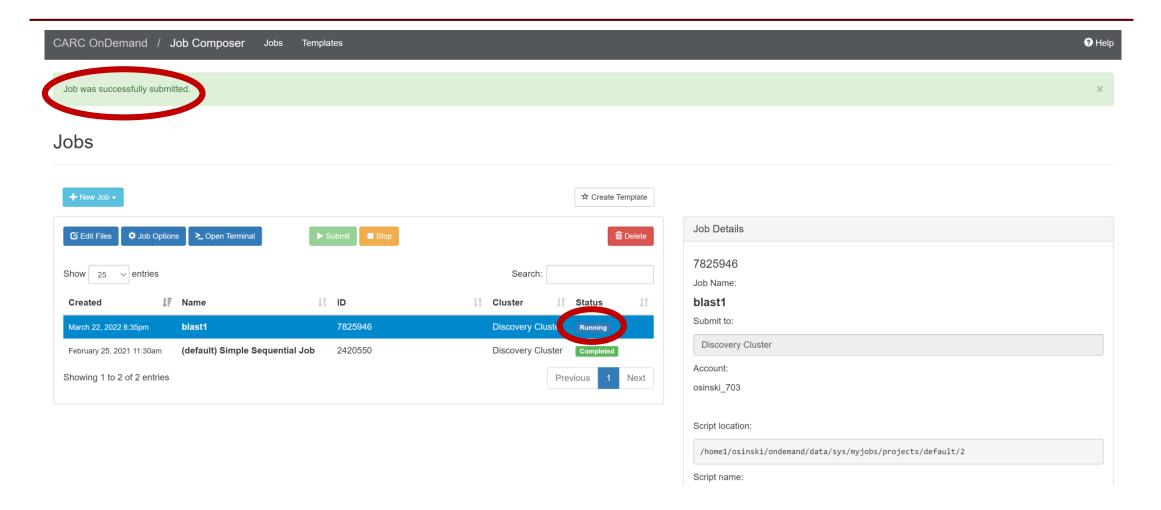
CARC OnDemand: Editing a Job



CARC OnDemand: Submitting a Job



CARC OnDemand: Submitting a Job



Submit a Job – shell

Submit the job

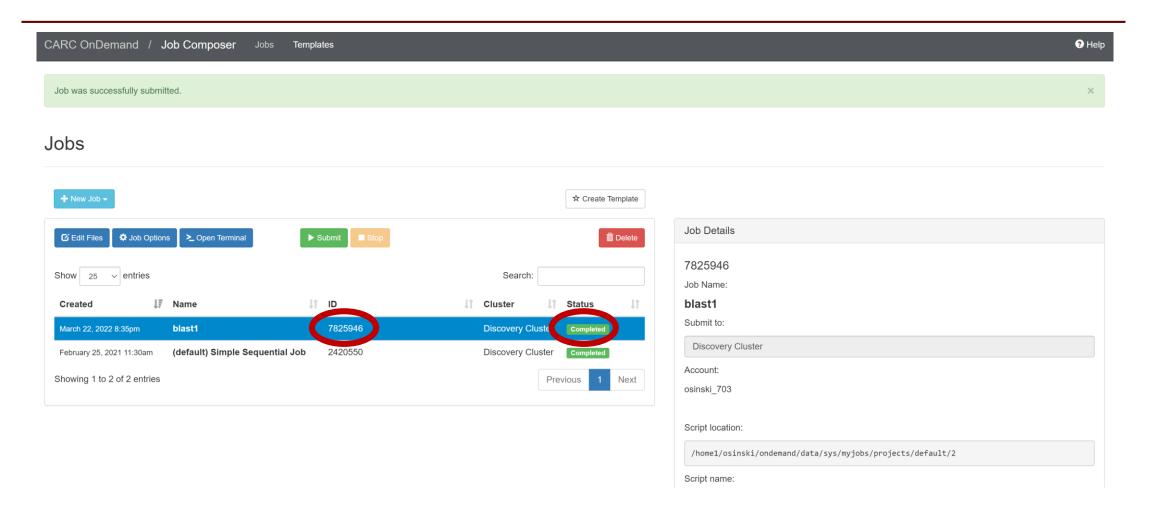
```
[ttrojan@discovery1 ~]$ sbatch blast1.job
Submitted batch job 4773117
```

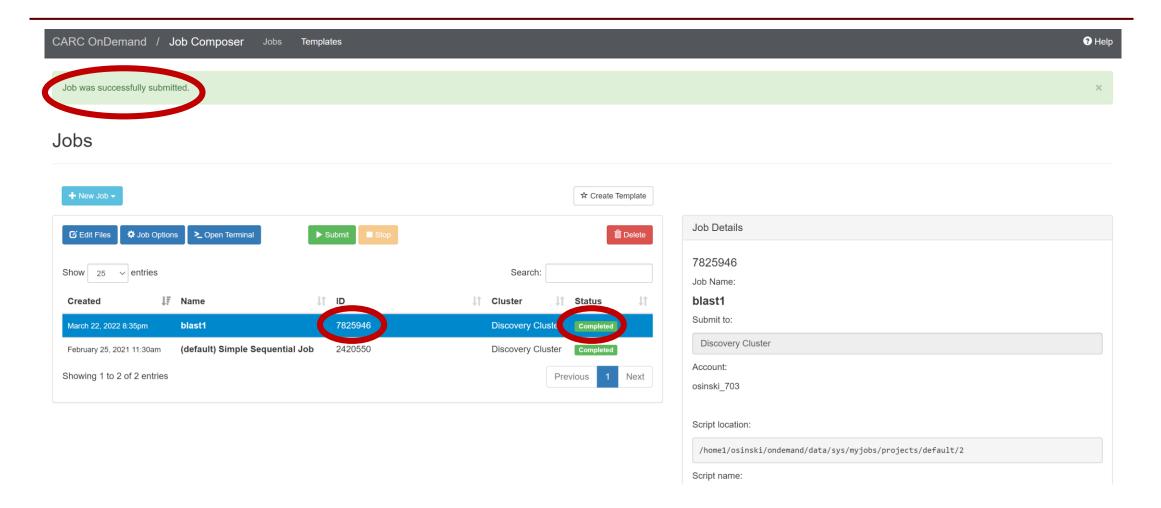
Check the status of the job

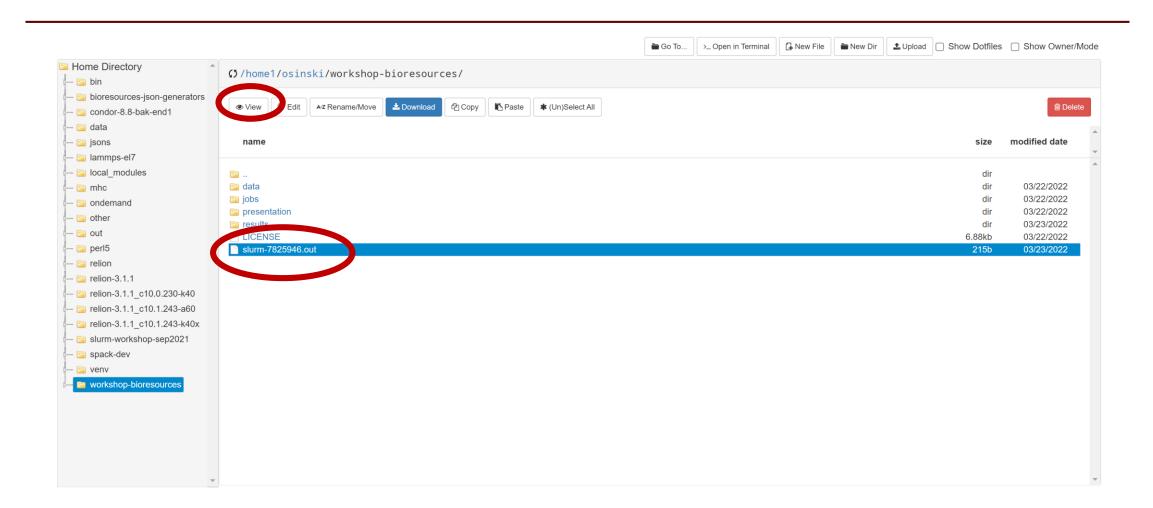
```
[ttrojan@discovery1 ~]$ squeue -u ttrojan

JOBID PARTITION NAME USER ST TIME NODES NODELIST(REASON)

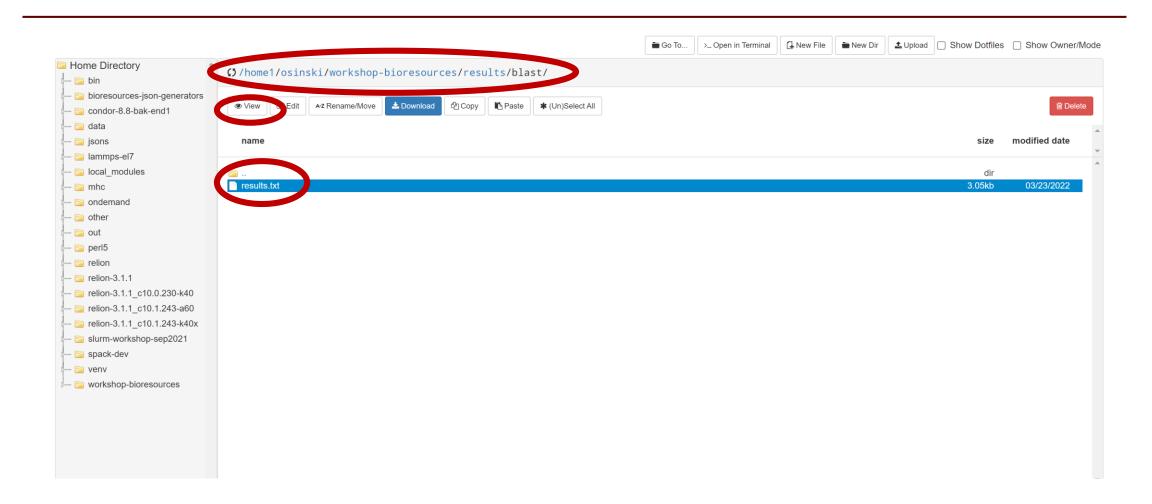
4773117 Main blast1.j ttrojan R 0:02 1 a02-d11
```







```
SLURM_JOB_ID = 7825946
SLURM_JOB_NODELIST = e09-18
TMPDIR = /tmp/SLURM_7825946
"Example blast start"
"Example blast end"
```



```
Query=
Length=15
                                                                     Score
Sequences producing significant alignments:
                                                                    (Bits) Value
Q9JK11.1 RecName: Full=Reticulon-4; AltName: Full=Foocen; AltName... 35.0
                                                                             0.001
Q99P72.2 RecName: Full=Reticulon-4; AltName: Full=Neurite outgrow... 35.0
                                                                            0.001
Q9NQC3.2 RecName: Full=Reticulon-4; AltName: Full=Foocen; AltName... 33.9
                                                                            0.004
>09JK11.1 RecName: Full=Reticulon-4; AltName: Full=Foocen; AltName: Full=Glut4
vesicle 20 kDa protein; AltName: Full=Neurite outgrowth
inhibitor; Short=Nogo protein [Rattus norvegicus]
Length=1163
Score = 35.0 bits (79), Expect = 0.001, Method: Composition-based stats.
Identities = 15/15 (100%), Positives = 15/15 (100%), Gaps = 0/15 (0%)
Query 1
            HYLGLANKSVKDAMA 15
            HYLGLANKSVKDAMA
Sbjct 1135 HYLGLANKSVKDAMA 1149
>Q99P72.2 RecName: Full=Reticulon-4; AltName: Full=Neurite outgrowth inhibitor;
Short=Nogo protein [Mus musculus]
Length=1162
Score = 35.0 bits (79), Expect = 0.001, Method: Composition-based stats.
```

Genome mapping and tools: Read mapping

- Aim: to find coordinates of reads in the reference genome.
- Challenges:
 - Millions of short sequences
 - Sequences are often paired
 - Errors are not randomly distributed
- Most popular programs are <u>bow</u>tie and <u>bwa</u> (both use <u>Burrows-Wheeler</u> Transform algorithm). Two-step approach:
 - Create an index for the reference genome (one time for one genome).
 - Map reads to the reference genome using this index

Genome mapping and tools – overview I

- FastQC
 - FastQC is a quality control application for high throughput sequence data
 - Checks the quality of their sequence data
 - Generates an HTML report

http://www.bioinformatics.babraham.ac.uk/projects/fastqc/

Genome mapping and tools – overview II

- bowtie
 - The first version of bowtie [Langmead et al. 2009] is optimal for:
 - short reads (under 50 bp)
 - reads without indels (insertions/deletions)
- bowtie2
 - The second version of bowtie2 [Langmead & Salzberg 2012] is optimal for:
 - long reads (more than 50 bp)
 - reads with indels
 - various alignment options
- Each version has its own index file format (bowtie-build / bowtie2-build tools).
- A popular RNA-seq analysis toolset (tophat, cufflinks) is based on bowtie / bowtie2

http://bowtie-bio.sourceforge.net

Genome mapping and tools – overview III

- bwa
 - bwa backtrack [Li, Durbin 2009]:
 - for short reads (< 100bp)
 - bwa bwasw [Li, Durbin 2010]:
 - for long reads (70bp 1Mbp)
 - short indels
 - bwa mem [Li 2013]:
 - for long reads (70bp 1Mbp)
 - faster and more efficient

Genome mapping and tools – overview IV

- samtools package A set of utilities for processing SAM/BAM files
- samtools view
 - convert a bam file into a sam file samtools view sample.bam > sample.sam
 - Convert a sam file into a bam file samtools view -bS sample.sam > sample.bam
 - Extract all the reads aligned to the range specified. An index of the input file is required

```
samtools view -h -b sample sorted.bam "chr1:10-13" > tiny sorted.bam
```

samtools sort

samtools sort unsorted_in.bam sorted_out

samtools index

samtools index sorted.bam (creates an index file, sorted.bam.bai)

http://samtools.sourceforge.net



Genome mapping and tools – overview IV

- samtools package A set of utilities for processing SAM/BAM files
- samtools view

- -b: output BAM -S: read SAM
- convert a bam file into a sam file samtools view sample.bam > sample.sam
- Convert a sam file into a bam file samtools view -bS sample.sam > sample.bam
- Extract all the reads aligned to the range specified. An index of the input file is required •

```
samtools view -h -b sample_sorted.bam "chr1:10-13" > tiny_sorted.bam
```

samtools sort

```
samtools sort unsorted in.bam sorted out
```

samtools index

```
samtools index sorted.bam (creates an index file, sorted.bam.bai)

<a href="http://samtools.sourceforge.net">http://samtools.sourceforge.net</a>
```



Genome mapping and tools – overview IV

samtools flagstat – report basic statistics

```
samtools flagstat sample.bam
```

An example of output:

```
4198456 + 0 in total (QC-passed reads + QC-failed reads)
0 + 0 duplicates
4022089 + 0 mapped (95.80%:-nan%)
4198456 + 0 paired in sequencing
2099228 + 0 read1
2099228 + 0 read2
3796446 + 0 properly paired (90.42%:-nan%)
4013692 + 0 with itself and mate mapped
8397 + 0 singletons (0.20%:-nan%)
167574 + 0 with mate mapped to a different chr
72008 + 0 with mate mapped to a different chr (mapQ>=5)
```

samtools faidx – index a FASTA file

samtools faidx ref.fasta (creates an index file ref.fasta.fai)

samtools merge – merge several BAM files into one

samtools merge out.bam in1.bam in2.bam

Genome mapping and tools – overview V

BedTools package

- bamtobed Convert BAM alignments to BED (& other) formats
- bamtofastq Convert BAM records to FASTQ records
- bedtobam Convert intervals to BAM records
- closest Find the closest, potentially non-overlapping interval
- complement Extract intervals _not_ represented by an interval file
- coverage Compute the coverage over defined intervals
- genomecov Compute the coverage over an entire genome
- getfasta Use intervals to extract sequences from a FASTA file
- intersect Find overlapping intervals in various ways
- shuffle Randomly redistribute intervals in a genome
- sort Order the intervals in a file

Genome mapping and tools – overview V

bedtools intersect

Report the intervals that represent overlaps between your two files:

bedtools intersect -a cpg.bed -b exons.bed

Report the original feature in each file:

bedtools intersect -a cpg.bed -b exons.bed -wa -wb

How many base pairs of overlap were there?

bedtools intersect -a cpg.bed -b exons.bed -wo

Counting the number of overlapping features:

bedtools intersect -a cpg.bed -b exons.bed -c

Find features that DO NOT overlap:

bedtools intersect -a cpg.bed -b exons.bed -v



Exercise

 There are paired reads of some DNA sequencing experiment of the human sample:

```
bio-bootcamp/R1.fastq.gz
bio-bootcamp/R2.fastq.gz
```

- You will study some particular region of the human genome
- Map reads to the human reference genome (version hg19 find path on our Bio Resources)
- Extract reads that map to your region only
- Upload the reads to UCSC genome browser as a custom track
- count the number of insertions and deletions in SAM file

How To: Mapping

load bowtie2 program:

```
module purge
module load gcc/9.2.0
module load bowtie2
```

- Copy sequence of a chromosome your region is located at as a FASTA file
 - find the path on our website in Homo sapiens > UCSC > hg19 > Chromosome 21
 - https://carc.usc.edu/user-information/bio-resources/reference-genomes
 - Add chr*.fa at the end of the path
 - cp path_above /home1/ttrojan/bio-bootcamp/results/read-mapping
- Map reads to this chromosome using bowtie2 with the standard parameters

Don't forget to make an index (bowtie-build2) of the chromosome before mapping!

- You will get a SAM file as an output, convert to BAM (samtools view)
- Count the number of insertions and deletions in SAM file (use cut for field 6, and grep)

How To: Extracting reads

load BedTools:

```
module purge
module load gcc/9.2.0
module load bedtools2
```

• Create a tab-delimited BED file with the coordinates of your region:

```
chr21 10000000 20000000
```

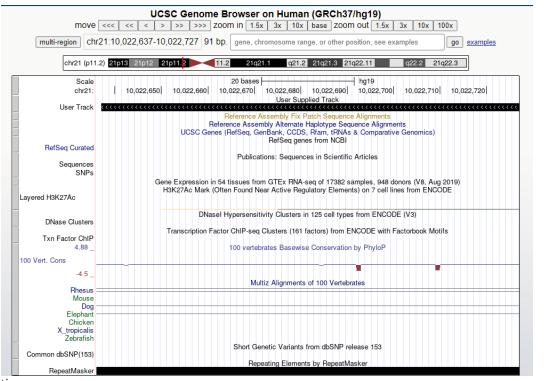
- Convert SAM file with mapped reads to BAM file using samtools view
- Use bedtools intersect to extract the reads from the BAM file

You'll need a BED file to upload the result to UCSC genome browser, so figure out how to make bedtools intersect to produce an output in BED format.

How To: UCSC custom track

Upload the BED file to UCSC genome browser

'Add custom track' button \rightarrow Choose file \rightarrow Submit \rightarrow Go



Thanks to the whole CARC team:

BD Kim
Bill Jendrzejek
Jimi Chu
James K Hong
Derek Strong
Cesar Sul
Marco Olguin
Hao Ji
Iman Rahbari

Reference material:

- HPCBio (Holmes J., Clark L., Drnevicch J., Valizadegan N.)
- CNRG (Davidson D., Leigh J.)
- Skoltech (Khrameeva E.)

Resources

- CARC home page
 - https://carc.usc.edu
- Bio Resources at CARC
 - https://carc.usc.edu/user-information/bio-resources
- CARC User Forum
 - https://hpc-discourse.usc.edu/categories
- SLURM tutorials
 - https://slurm.schedmd.com/tutorials.html
- SLURM quick reference
 - https://slurm.schedmd.com/pdfs/summary.pdf



Mapping exercise: Answers

```
#!/bin/bash
#SBATCH --partition main
#SBATCH --nodes 1
#SBATCH --ntasks 20
#SBATCH --time 01:00:00
#SBATCH --mem 4a
#SBATCH --account=ttrojan 001
#SBATCH --chdir /home1/ttrojan/workshop-bioresources
module purge
module load gcc/9.2.0
module load bowtie2
module load samtools
module load bedtools2
mkdir results/read-mapping
cp data/R*.gz results/read-mapping
gunzip results/read-mapping/R1.fastg.gz
gunzip results/read-mapping/R2.fastg.gz
cp -v /project/biodb/genomes/Homo sapiens/UCSC/hq19/Sequence/Chromosomes/chr21.fa results/read-mapping/
bowtie2-build --threads $SLURM NTASKS results/read-mapping/chr21.fa results/read-mapping/chr21index
bowtie2 --threads $SLURM NTASKS -x results/read-mapping/chr21index -q results/read-mapping/R1.fastq > results/read-mapping/R1.sam
bowtie2 --threads $SLURM NTASKS -x results/read-mapping/chr21index -q results/read-mapping/R2.fastq > results/read-mapping/R2.sam
samtools view results/read-mapping/R1.bam | cut -f 6 | grep -c 'D' > results/read-mapping/R1.no of deletions.txt
samtools view results/read-mapping/R1.bam | cut -f 6 | grep -c 'I' > results/read-mapping/R1.no of insertions.txt
samtools view results/read-mapping/R2.bam | cut -f 6 | grep -c 'D' > results/read-mapping/R2.no of deletions.txt
samtools view results/read-mapping/R2.bam | cut -f 6 | grep -c 'I' > results/read-mapping/R2.no of insertions.txt
samtools view -bS results/read-mapping/R1.sam > results/read-mapping/R1.bam
samtools view -bS results/read-mapping/R2.sam > results/read-mapping/R2.bam
samtools sort results/read-mapping/R1.bam > results/read-mapping/R1 sorted.bam
samtools sort results/read-mapping/R2.bam > results/read-mapping/R2 sorted.bam
samtools view -h -b results/read-mapping/R1 sorted.bam "chr21:10000000-20000000" > results/read-mapping/R1 sorted region.bam
samtools view -h -b results/read-mapping/R2 sorted.bam "chr21:10000000-20000000" > results/read-mapping/R2 sorted region.bam
bamToBed -i results/read-mapping/R1 sorted region.bam > results/read-mapping/R1 sorted region.bed
bamToBed -i results/read-mapping/R2 sorted region.bam > results/read-mapping/R2 sorted region.bed
bedtools intersect -a results/read-mapping/R1 sorted region.bed -b results/read-mapping/R2 sorted region.bed > results/read-mapping/reads.bed
```