

Microbiota amplicOn
CHaracterization Implement

MOCHI

User Guide

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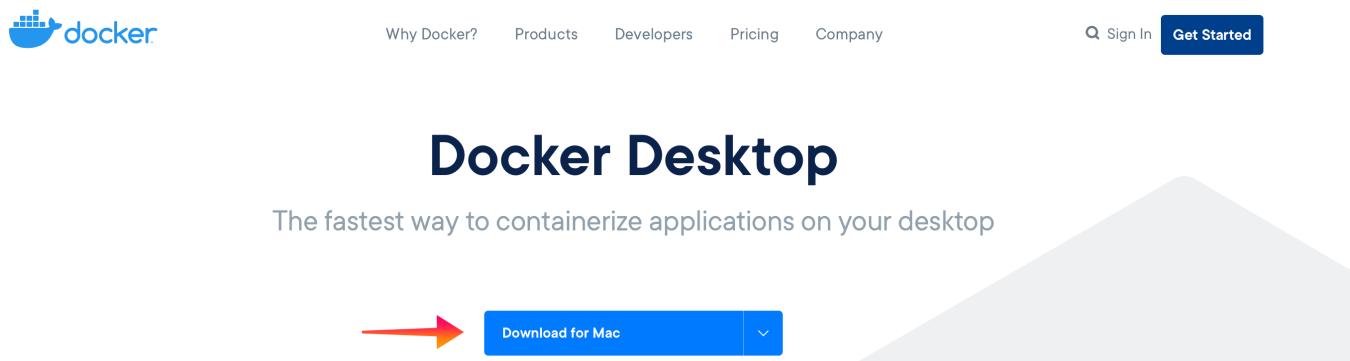
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CHAPTER 1: INSTALLATION OF LOCAL SERVICE

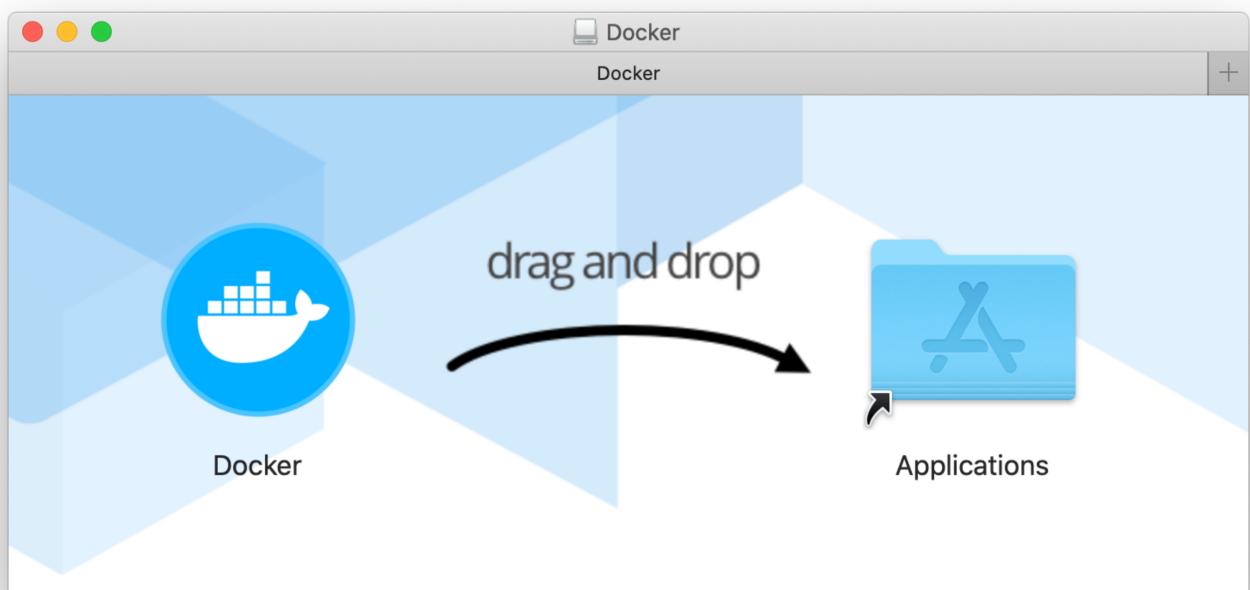
MacOS

(A) Install Docker

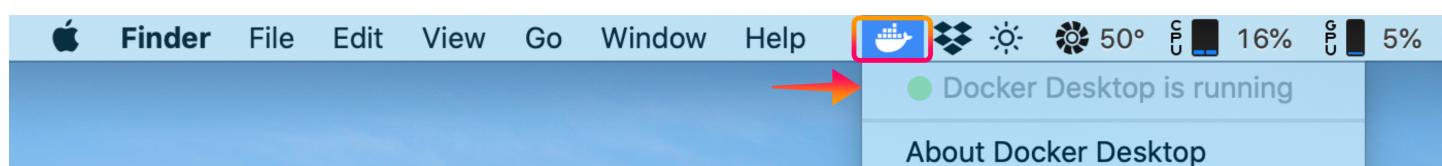
1. Download [Docker Desktop](#).



2. Open "**Docker.dmg**" file. Drag **Docker Desktop** app to the Applications folder.



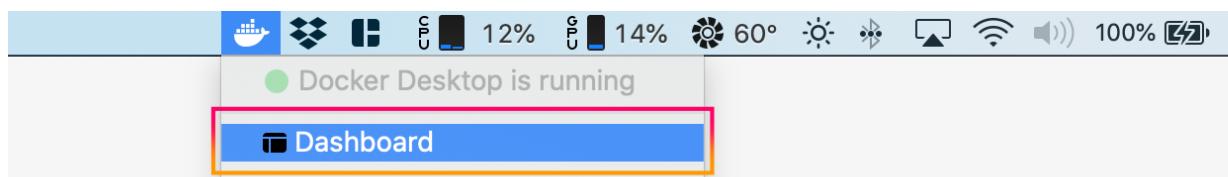
3. Start the Docker service by double clicking the Docker app. Wait for a few seconds to load, the docker icon should appear in the status bar.



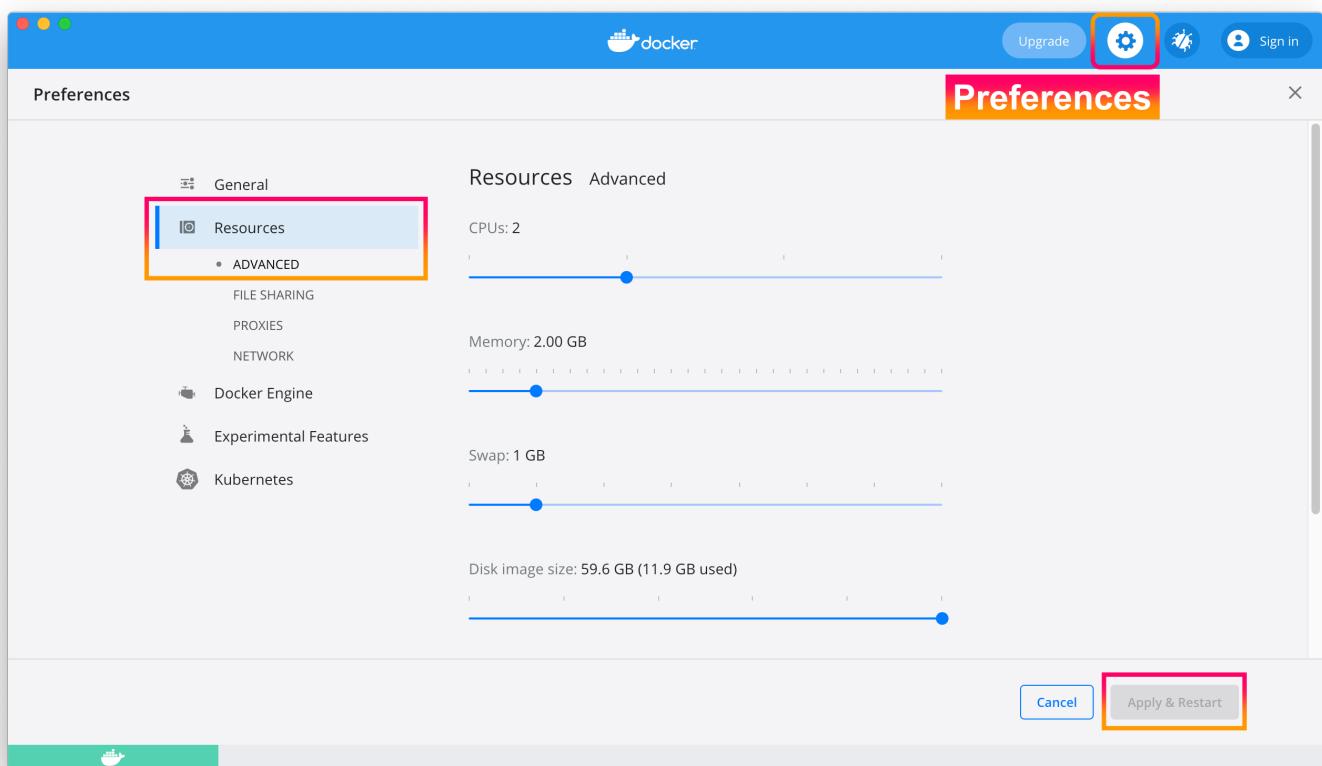
4. (Optional) In MacOS, the computational resources are preset in Docker app. To optimize the efficiency of analysis, the user can adjust the settings with the following instruction. We recommend settings **above 4 CPUs** and **8-16 GB memory** (by default, MOCHI only uses a maximum of 16 GB memory).

⚠ Assigning all of the resources to Docker may cause your system to delay or crash.

4.1. Open the Docker dashboard from the drop-down menu in the status bar.



4.2. Go to **Preference / Resources / Advanced**. Adjust the resources using the rolling bar. Press "Apply & Restart". Waiting for Docker to restart.

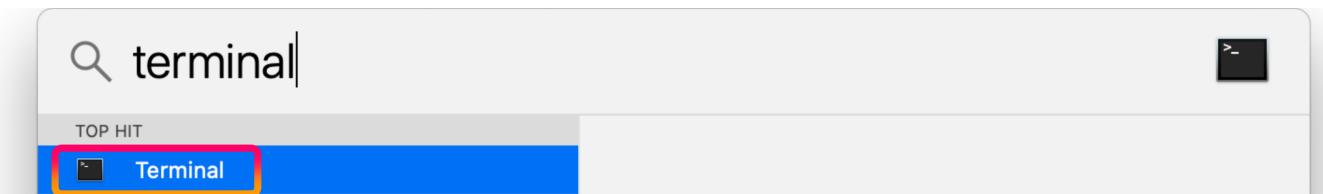


(B) Start MOCHI service

1. Download “**docker-compose.yml**” from [NCTU website](#), and place it under a folder named “**MOCHI**”.



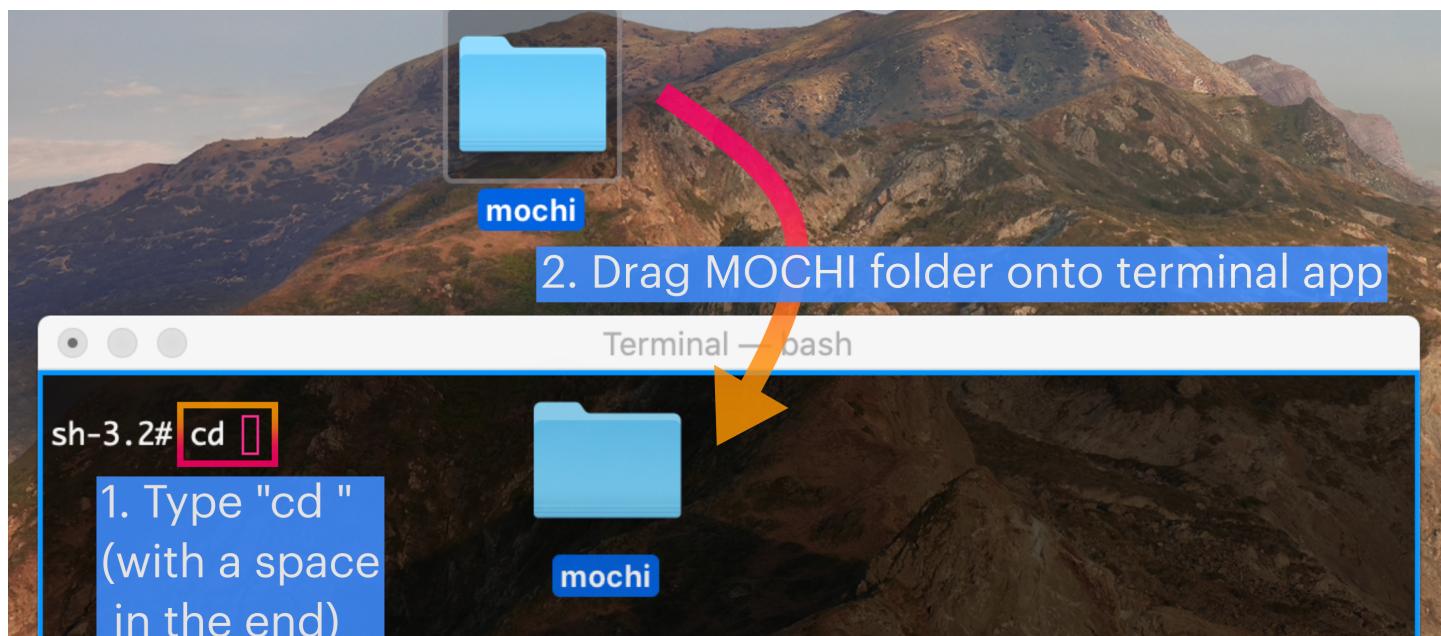
2. Open the **Terminal** app. (Press “Command + Space” to open Spotlight and type “terminal”.)



3. Run **cd /path/to/MOCHI** to navigate to the MOCHI directory.

```
sh-3.2# cd /Users/Mac/Desktop/mochi
```

Hint: If you do not know the folder path of MOCHI, type **cd** and then a space, and then drag the MOCHI folder into the terminal window. The folder path should appear in the terminal automatically.



4. Run `docker-compose up -d` to download and start the MOCHI image from Docker Hub. (The download process will only run during the first setup. The size of the MOCHI image is around 10 GB, and the running time depends on the download speed.)

```
[powang@MacBook-Pro--PoWang mochi % docker-compose up -d
WARNING: Some services (mochi_server) use the 'deploy' key, which will be ignored. Compose does not support 'deploy' configuration - use `docker stack deploy` to deploy to a swarm.
Creating network "mochi_default" with the default driver
Pulling mochi_server (dockerjjz/mochi_local:...
latest: Pulling from dockerjjz/mochi_local
f15005b0235f: Pull complete
1901fd813023: Pull complete
a92940affedf: Pull complete
dbebda29cb22: Pull complete
3c63b26b92fd: Pull complete
e0c15c0b4e0b: Pull complete
Digest: sha256:1501a145eb826f9f799239964eb064170fdc5be8abcd7b04fd9a61c888b9dee
Status: Downloaded newer image for dockerjjz/mochi_local:latest
Creating mochi_server ... done]
```

5. Open the browser, and type **127.0.0.1:3811** in the address bar. A MOCHI interactive webpage will appear and you will be able to begin your analysis.

Welcome to MOCHI! (Microbiota amplicon CHaracterization Implement)

MOCHI is a 16S or 18S microbiota amplicon rRNA analytical tool for microbiota based primarily on QIIME2 with a friendly web interface powered by the R package of Shiny. MOCHI may also be downloaded and operated locally.

Overview of MOCHI

The MOCHI pipeline consists of three main steps:

- Sequence Preprocessing:** This step includes:
 - Step 1. Sequence summary:** Shows read counts and read quality.
 - Step 2. Sequence denoising (DADA2):** Shows sequence table and alpha rarefaction.
 - Step 3. Taxonomy classification:** Shows taxonomy prediction table.
- Taxonomy Analysis:** This step includes various plots and tables:
 - Krona chart.
 - Bar plot.
 - Heatmap.
 - Taxonomy table.
 - Alpha diversity plot.
 - Box plot.
 - PCoA plot.
 - Comparison between groups.
 - Post hoc analysis.
 - ANCOM.
- Function Analysis:** This step includes:
 - FAPROTAX.
 - Functional annotation table.
 - Bar plot.

The advantages of MOCHI

- Friendly user interface: point-and-click and fill-in inputs, no programming required.
- Cross-platform: simple set-up with Docker containers on Linux, Windows, or Mac OS.
- Local computing resource: runs on your premise with privacy, not subject to network reliability and limitation.
- Compatible with other downstream analytical tools
- Publishable plots and charts generated with chosen parameters

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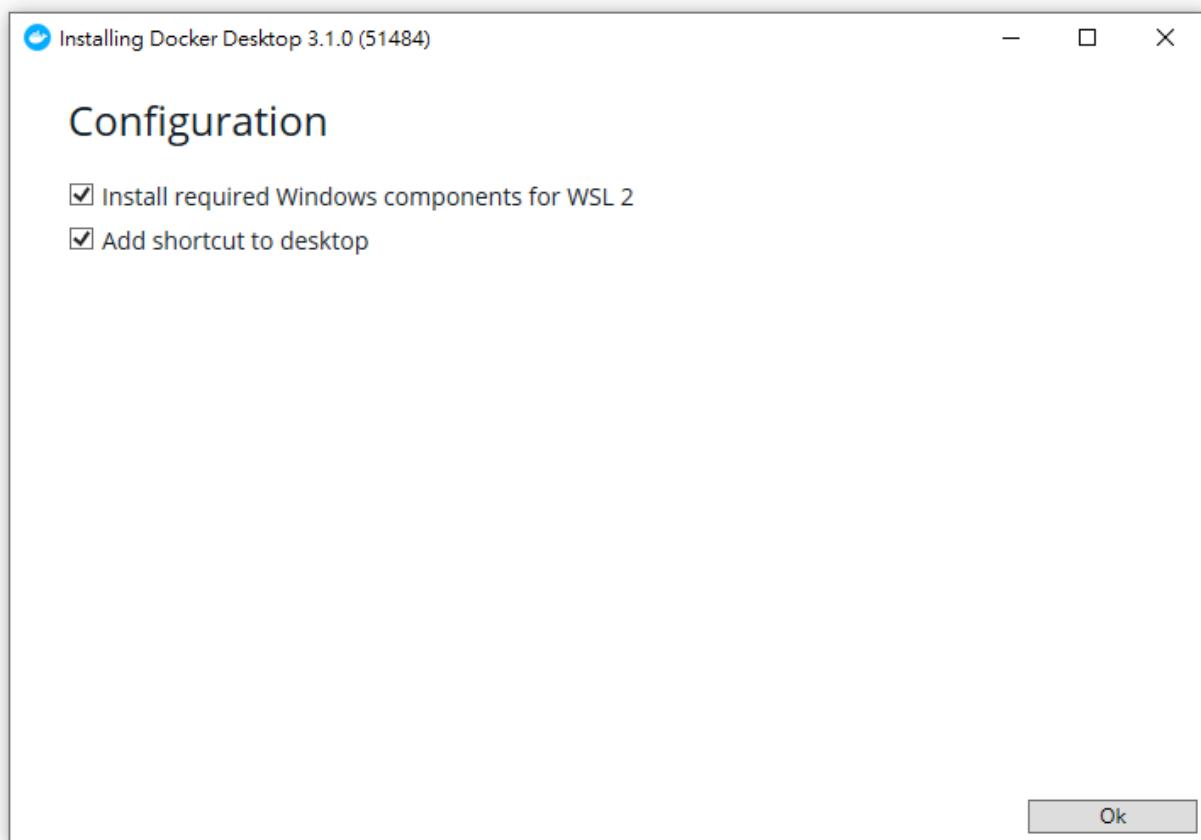
Windows 10

(A) Install Docker

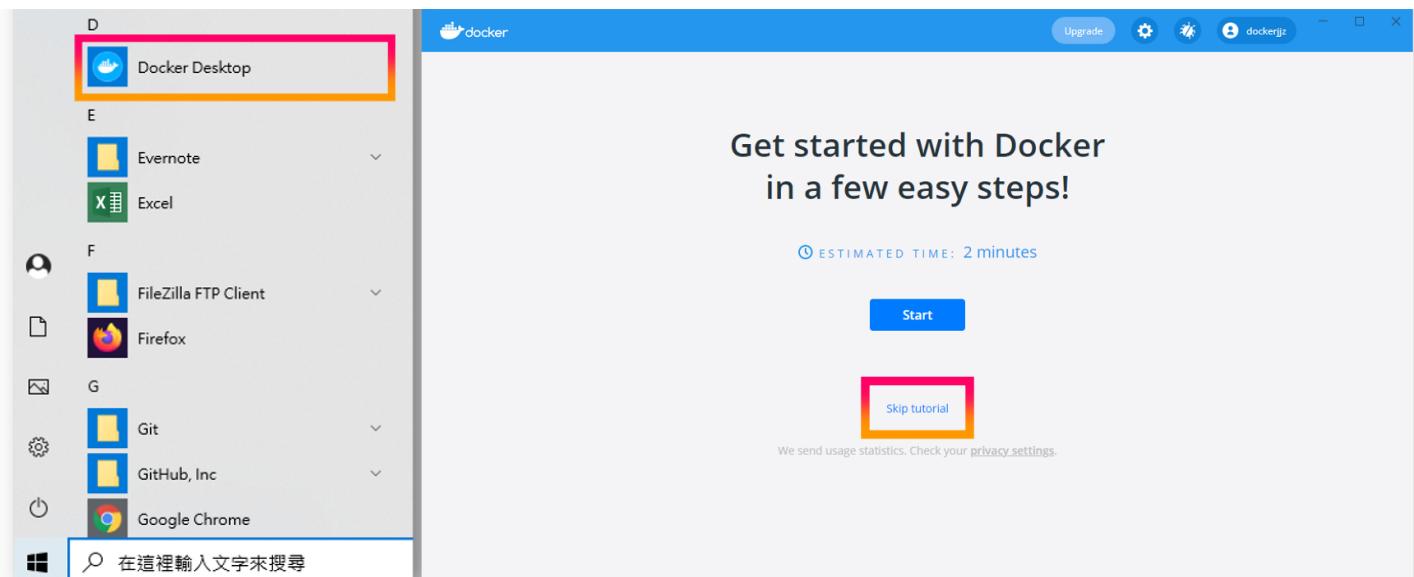
6. Download [Docker Desktop](#).



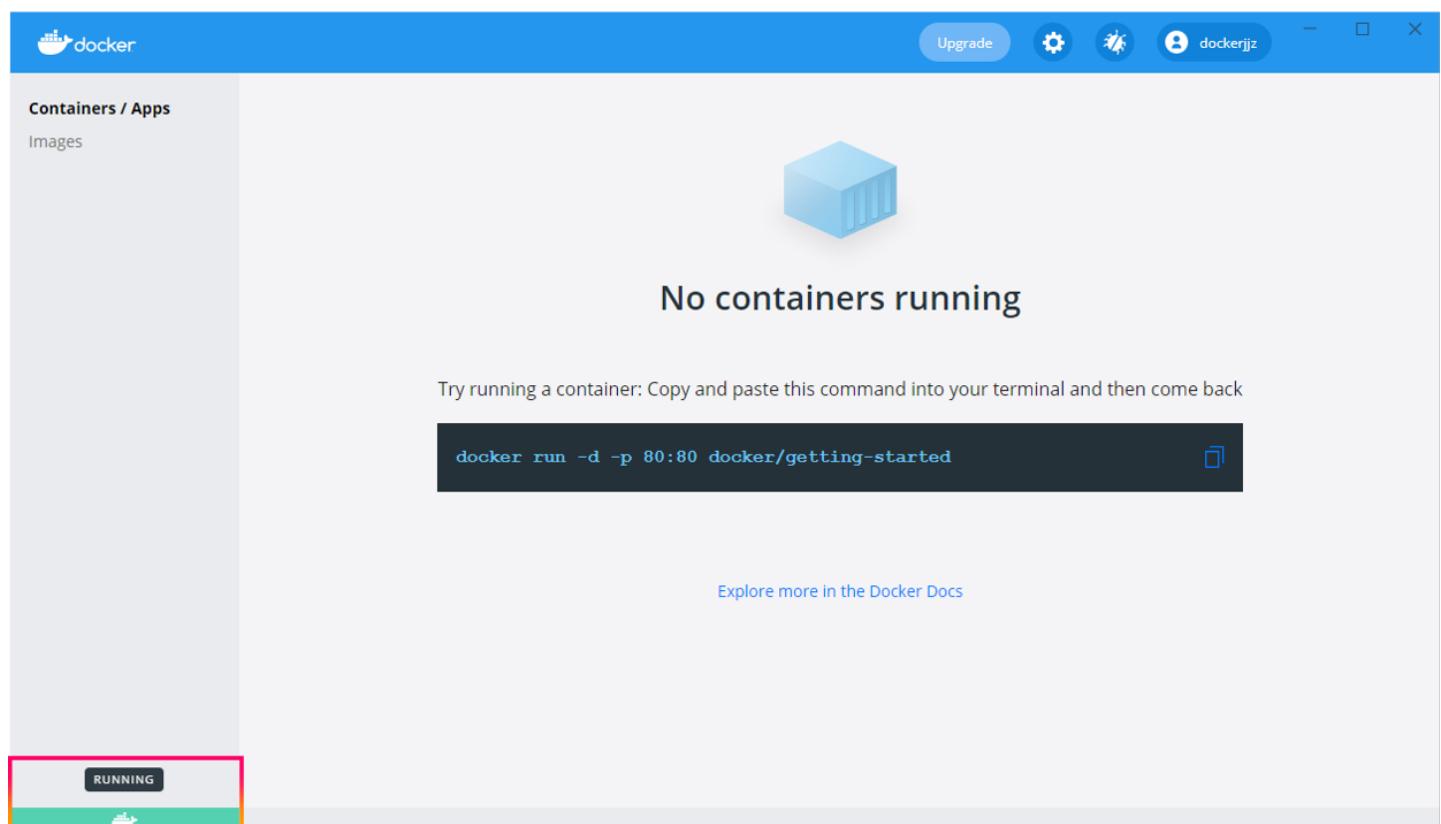
7. Run "**Docker Desktop Installer.exe**" and follow the instructions for setup. (You may be asked whether to install WSL2 Engine. By default, it is installed along with Docker to speed up the performance but is not required for running MOCHI. Use the default setting if you don't know what to choose.)



8. Start Docker by **clicking the Docker icon**. Press “Skip tutorial”.



9. If the Docker service has been successfully turned on, a green indicator will appear on the bottom-left in the Docker window.

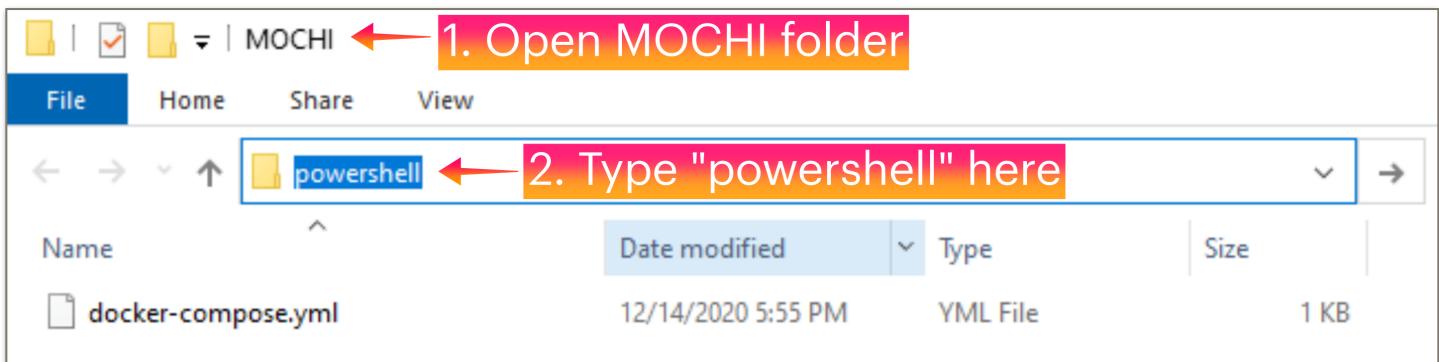


(B) Start MOCHI service

1. Download “**docker-compose.yml**” from the NCTU website, and place it in a folder named “**MOCHI**”.



2. Open **MOCHI** folder, type “**powershell**” in the address bar, and press enter. This will open a command-line shell under MOCHI directory.



3. In Powershell, run **docker-compose up -d** to download and start the MOCHI image from Docker Hub. (The download process will only run at the first setup. The size of the MOCHI image is about 10 GB, and the running time depends on the download speed.)

```
PS C:\Users\dodolab\Desktop\MOCHI> docker-compose up -d
Creating network "mochi_default" with the default driver
Pulling mochi_server (dockerjjz/mochi_local_version:)...latest: Pulling from dockerjjz/mochi_local_version
f15005b0235f: Pull complete
1901fd813023: Pull complete
a92940affedf: Pull complete
dbebda29cb22: Pull complete
3c63b26b92fd: Pull complete
e4191a297544: Pull complete
Digest: sha256:794909d921dfe9cc55edba44a3fe66701e43845c5c5578bf1e194064e071cbe
Status: Downloaded newer image for dockerjjz/mochi_local_version:latest
Creating mochi_server_version ... done
```

4. Open a browser, and type **127.0.0.1:3811** in the address bar. A MOCHI interactive webpage is now available to begin your analysis.

The screenshot shows the MOCHI web application interface. At the top, there is a navigation bar with links for Home, Sequence Preprocessing, Taxonomy Analysis, Function Analysis, and Tutorial. Below the navigation bar, a main title says "Welcome to MOCHI! (Microbiota amplicon CHaracterization Implement)". A brief description follows: "MOCHI is a 16S or 18S microbiota amplicon rRNA analytical tool for microbiota based primarily on QIIME2 with a friendly web interface powered by the R package Shiny. MOCHI may also be downloaded and operated locally." The central part of the page is titled "Overview of MOCHI". It illustrates the analytical workflow:

- Sequence Preprocessing:** This section is divided into three steps:
 - Step 1. Sequence summary:** Shows "read counts" and "read quality" plots.
 - Step 2. Sequence denoising (DADA2):** Shows a "sequence table" and an "alpha rarefaction" curve.
 - Step 3. Taxonomy classification:** Shows a "taxonomy prediction table".
- Taxonomy Analysis:** This section contains various plots and tables:
 - A taxonomy table showing hierarchical taxonomic distribution.
 - A bar plot showing abundance at different taxonomic levels.
 - A heatmap showing abundance across samples.
 - A circular "Krona" chart.
 - An "alpha diversity" plot.
 - A "box plot" comparing alpha diversity.
 - A "PCoA plot" showing samples grouped by sample ID.
 - A "comparison between groups" section with an ANCOM plot.
 - A "post hoc analysis" table for Tukey test results.
- Function Analysis:** This section shows results from FAPROTAX:
 - A "functional annotation table" listing genes and their counts across samples.
 - A "bar plot" showing abundance of specific functional annotations.

On the left side of the main content area, there are icons for "Sequence data", "Metadata for samples", and "Taxonomy database", each with an arrow pointing to its respective input step in the sequence processing flow.

The advantages of MOCHI

- Friendly user interface: point-and-click and fill-in inputs, no programming required.
- Cross-platform: simple set-up with Docker containers on Linux, Windows, or MacOS.
- Local computing resource: runs on your premise with privacy, not subject to network reliability and limitation.
- Compatible with other downstream analytical tools
- Publishable plots and charts generated with chosen parameters

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Linux

(A) Install Docker

- ▶ Please follow the [official guidance](#) for setup and start Docker Engine on Ubuntu.

(B) Start MOCHI service

1. Download “**docker-compose.yml**” from [NCTU website](#), and place it in a folder named “**MOCHI**”.



2. Navigate to the MOCHI directory.

```
$ cd /path/to/place/MOCHI
```

3. Download and start the MOCHI image from Docker Hub. (The download process will only run during the first setup. The size of the MOCHI image is about 10 GB, and the running time depends on the download speed.)

```
$ docker-compose up -d
```

4. Open a browser, and type **127.0.0.1:3811** in the address bar. A MOCHI interactive webpage is now available to begin your analysis.

Additional Information

* Stop, Restore or Remove MOCHI service

- ▶ To temporarily pause MOCHI service, please run `docker-compose stop`. This will create a paused MOCHI container, and can be restored with command `docker-compose start`.

NOTE: Please be aware that interrupting the MOCHI service when an analysis is running will not save the running process. The MOCHI container is automatically paused once you restart your computer or docker service.

- ▶ To permanently close MOCHI service, please run `docker-compose down`. This will remove all the data generated by MOCHI. Please save the results in advance. If you wish to open the service again, please run `docker-compose up -d`.
- ▶ To uninstall MOCHI image from your computer, please run `docker images`:

REPOSITORY	TAG	IMAGE ID	CREATED	SIZE
dockerjjz/mochi_local	latest	441fe987dce8	2 weeks ago	9.05GB

and look for “IMAGE ID” of the repository named “**dockerjjz/mochi_local**”. Then, run `docker rmi [IMAGE ID]` (e.g., `docker rmi 441`; type partial or complete image ID are both acceptable.)

* Remember to navigate under the MOCHI folder (where “`docker-compose.yml`” is located) before starting, stopping or restoring MOCHI service.

- ▶ For MacOS, Please see [MacOS / step B-3](#).
- ▶ For Windows 10, Please see [Windows 10 / step B-2](#).
- ▶ For Linux, Please see [Linux / step B-2](#).

* After installing MOCHI, two folders are created under the MOCHI folder:

1. **seqs_folder**

- ▶ This folder is used to store the user’s sequence data which will be loaded in “Sequence preprocessing - Sequence summary”.
- ▶ The file type of sequence data needs to be **fastq.gz** or **fq.gz**.
- ▶ The filename of sequence data needs to satisfy Casava 1.8 demultiplexed format or the following example format **[SampleID]_[direction of reads]**:
 - ✓ Forward read: LS105_R1 or LS105_1
 - ✓ Reverse read: LS105_R2 or LS105_2

2. **taxa_database**

- ▶ This folder is used to store the taxonomy database, such as Greengenes, Silva and PR2. Please see "Sequence preprocessing - Taxonomy classification".

- * If you wish to change the default path to the sequence data and the taxonomic database, open the "**docker-compose.yml**" file, replace the texts "**./seqs_folder**" and "**./taxa_database**" with new paths to the sequence data and the taxonomic database, respectively. Relative path is allowed.
- * The default maximum memory used by MOCHI is 16GB. To increase, please modify the resource limit in the "**docker-compose.yml**" file.

```
version: '3.7'

services:

  mochi_server:
    # build: .
    image: dockerjjz/mochi_local
    ports:
      - "3811:3838"
      - "8011:80"
    volumes:
      - ./seqs_folder:/home/imuser/raw_data/:rw
      - ./taxa_database/greengenes:/home/imuser/taxa_database/greengenes:rw
      - ./taxa_database/silva:/home/imuser/taxa_database/silva:rw
      - ./taxa_database/PR2/18S/seqs:/home/imuser/taxa_database/PR2/18S/seqs:rw
      - ./taxa_database/PR2/18S/taxonomy:/home/imuser/taxa_database/PR2/18S/taxonomy:rw
    container_name: mochi_server

    deploy:
      resources:
        limits:
          #cpus: '8'
          memory: 16G

      environment:
        - shiny_port=3811
        - nginx_port=8011
```

CHAPTER 2: ANALYSIS

Sequence preprocessing

(A) Sequence summary

1. Select “**Sequence Preprocessing**” in the top bar, choose “**Step 1. Sequence summary**”.

The screenshot shows the MOCHI web interface. At the top, there is a navigation bar with links for Home, Sequence Preprocessing (which is currently selected), Taxonomy Analysis, Function Analysis, and Tutorial. Below the navigation bar, a blue header bar says "Welcome to MOCHI". On the right side of the header, there are three steps listed: "Step 1. Sequence summary" (with a red arrow pointing to it), "Step 2. Sequence denoising", and "Step 3. Taxonomy classification". A note below the steps states: "MOCHI is a 16S or 18S microbiota amplicon rRNA analysis tool based on QIIME2 with a friendly web interface powered by the R package of Shiny. MOCHI may also be downloaded and operated locally."

2. Press the “**Select the directory**” button to open the selection window. Alternatively, you can press the “Example sequences” button to download the example files. The parameters for example analysis are set once you have pressed the “Example sequences” button.

! Important: The sequence files should be placed under “seqs_folder” where MOCHI was installed.

The screenshot shows the "Sequence files" section of the MOCHI web interface. It asks to choose a directory containing sequence files (*.fastq.gz or *.fq.gz). There are two buttons: "Select the directory" (highlighted with a red box) and "Example sequences". To the right, there is a section titled "1. Sequences summary (for single end)" with a sub-section "(1) Summarize the single-end sequences" and a "Start!" button.

3. Select the directory containing the sequence files.

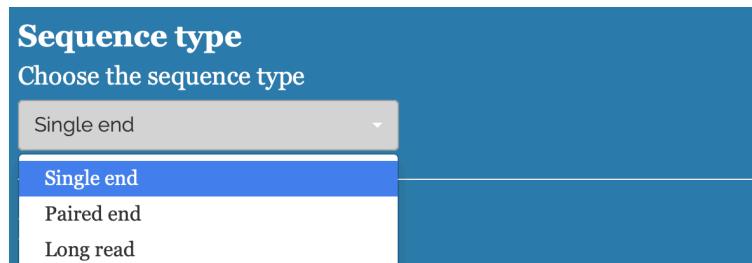
The screenshot shows a file selection dialog box titled "Please select a directory". It has a toolbar with "Create new folder", "Sort content", and a search bar. The main area shows a tree view of "Directories" and a list of "Content". The "raw_data" directory is expanded, showing a folder named "single_seqs_demo" (highlighted with a red box and labeled "Select folder"). To the right is a table of files with their sizes. At the bottom are buttons for "Confirm selection" (highlighted with a red box), "Cancel", and "Select" (highlighted with a blue box).

	Content	Size
1	L1S105_9_L001_R1_001.fastq.gz	889.1 kB
1	L1S140_6_L001_R1_001.fastq.gz	675.3 kB
1	L1S208_10_L001_R1_001.fastq.gz	861.9 kB
1	L1S257_11_L001_R1_001.fastq.gz	633.7 kB
1	L1S281_5_L001_R1_001.fastq.gz	685.7 kB
1	L1S57_13_L001_R1_001.fastq.gz	896.4 kB
1	L1S76_12_L001_R1_001.fastq.gz	767.0 kB
1	L1S88_8_L001_R1_001.fastq.gz	887.4 kB
1	L2S155_25_L001_R1_001.fastq.gz	770.0 kB
1	L2S175_27_L001_R1_001.fastq.gz	873.6 kB
1	L2S204_1_L001_R1_001.fastq.gz	615.1 kB
1	L2S222_23_L001_R1_001.fastq.gz	736.1 kB
1	L2S240_7_L001_R1_001.fastq.gz	929.2 kB
1	L2S309_33_L001_R1_001.fastq.gz	314.7 kB
1	L2S357_15_L001_R1_001.fastq.gz	502.5 kB
1	L2S382_34_L001_R1_001.fastq.gz	792.2 kB
1	L3S242_19_L001_R1_001.fastq.gz	149.3 kB

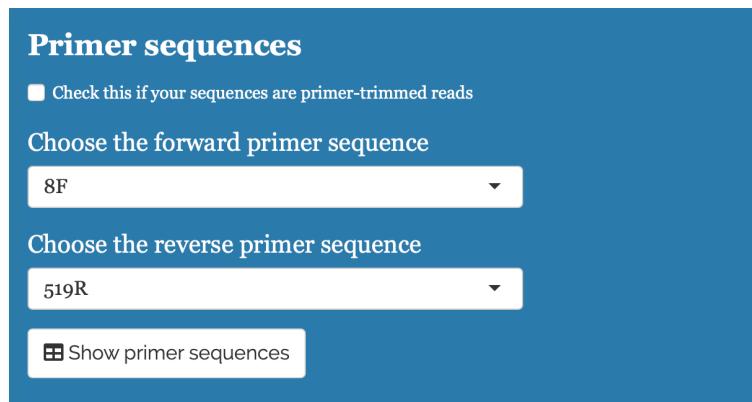
4. Sequence type: The settings for “**Step 1. Sequence summary**” and “**Step 2. Sequence denoising**” are different based on the sequence type chosen.

4.a. **Single-end or Paired-end**

- 4.a.1. The sequence type is automatically detected. If not correct, please choose manually.



- 4.a.2. Choose the **primers** or check the box when using primer-trimmed reads.



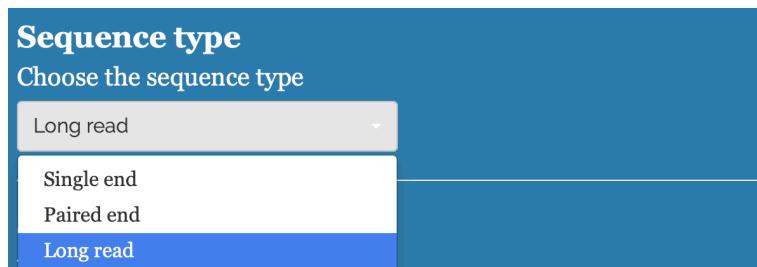
- 4.a.3. Set the number of threads for running the analysis. If zero, all available cores will be used. If you do not know the number to enter, leave it at the default number (all threads - 2).



- 4.a.4. Click on the “**Start!**” button.

4.b. Long-read

- 4.b.1. The sequence type will not automatically be detected. **Please select manually.**



- 4.b.2. Set the number of threads for running the analysis. If zero, all available cores will be used. If you do not know the number to enter, leave it at the default number (all threads - 2).



- 4.b.3. Click on the "**Start!**" button.

5. Please wait while it is running. When complete, a popup window will be displayed.

* running status

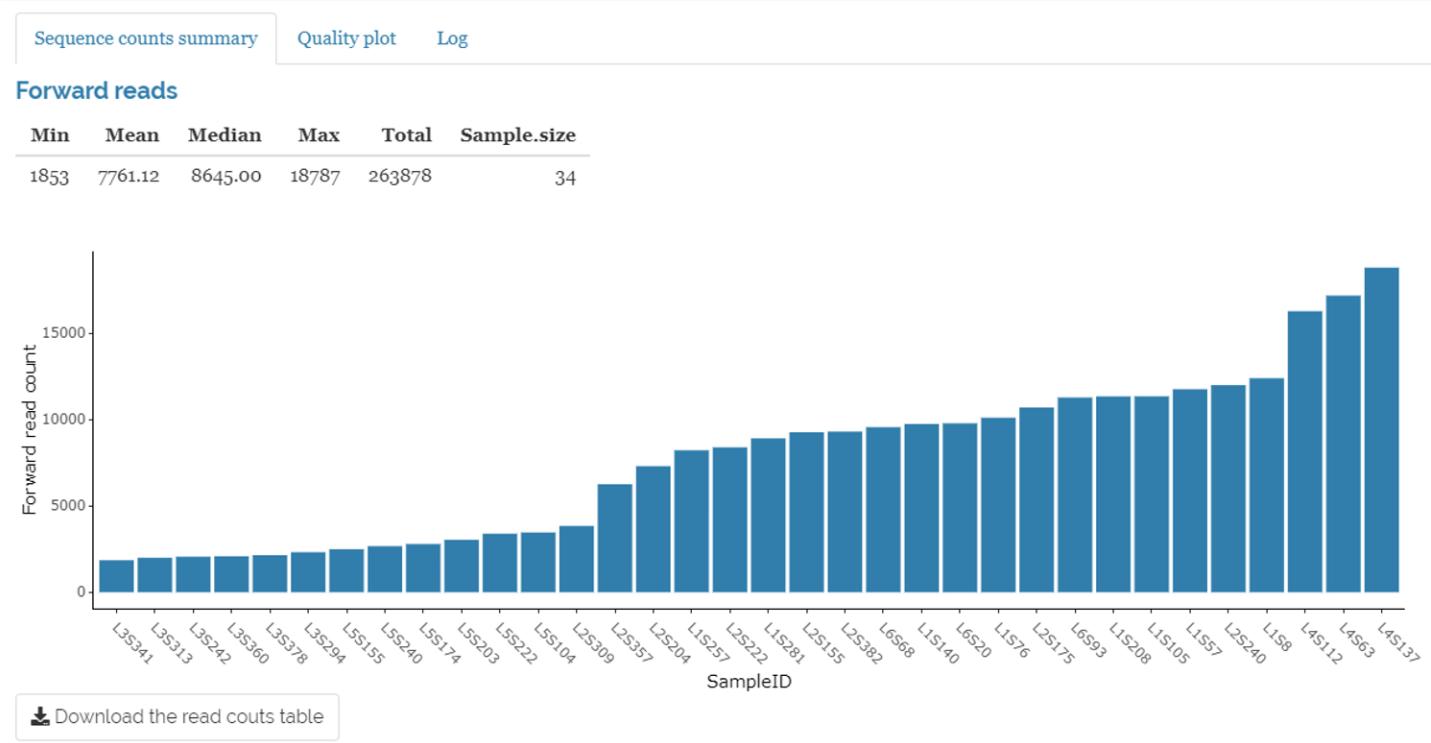


* complete status

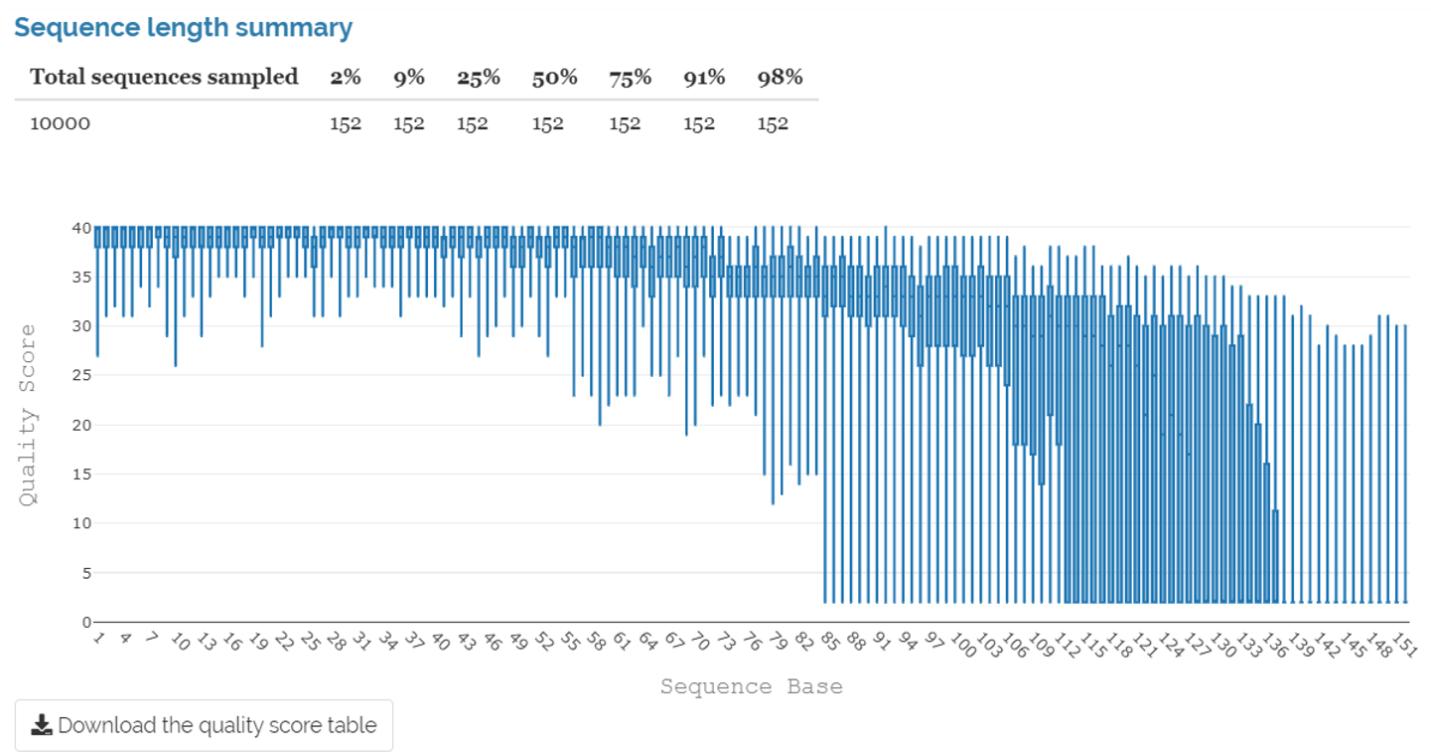
Successful!

This analysis took 11.5 secs. You can inspect the results now.

5.1. Demo results — “Sequence counts summary” summarizes the read counts of all samples.



5.2. Demo results — "Quality plot" summarizes the sequence length and show the distribution of quality score at each sequence base.



5.3. Demo results — "Log" records the used parameters and provides a button to download the table.

Record	Value
time	2021-05-31 01:44:22
duration	11.16 secs
sequence_type	Single end
sample_size	34
primer_trimmed	TRUE
forward_primer	515F
reverse_primer	806R
computing_setting	6

 Download

(B) Sequence denoising

1. Select “**Sequence Preprocessing**” in the top bar, choose “**Step 2. Sequence denoising**”.

The screenshot shows the MOCHI web interface. At the top, there is a navigation bar with links for Home, Sequence Preprocessing (which is currently selected and highlighted in blue), Taxonomy Analysis, Function Analysis, and Tutorial. Below the navigation bar, a banner reads "Welcome to MOCHI". Under the banner, there are three steps listed: Step 1. Sequence summary, Step 2. Sequence denoising, and Step 3. Taxonomy classification. A red arrow points to the "Sequence denoising" link. To the right of the steps, a note states: "MOCHI is a 16S or 18S microbiota amplicon rRNA MOCHI may also be downloaded and operated locally on QIIME2 with a friendly web interface powered by the R package of Shiny. Characterization Implement)"

2. Depending on sequence type selected in “**Step 1. Sequence summary**”, the settings for denoising will be different.

2.a. Single-end or Paired-end

- 2.a.1. Set the position and the quality score for trimming the sequences.

The screenshot shows the "Sequence trimming" section of the MOCHI web interface. It includes fields for "The start position" (set to 0) and "The end position" (set to 0). Below these are "learn more" and "Example" buttons. The "Quality score filtering" section follows, featuring a "Quality score threshold" field (set to 0) and a "learn more" button.

* Starting and ending position:

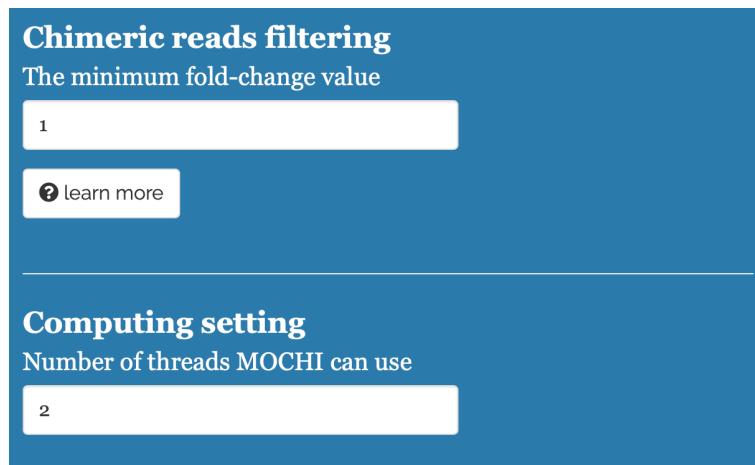
Base pairs below the starting position and above the ending position will be trimmed off. For instance, setting the starting position to 5 and the ending position to 120 will obtain sequences from 5 to 120 bp with 115 bp long.

In addition, reads shorter than the ending position will be discarded. In above setting, sequences less than 120 bp will be discarded. If the ending position is set to 0, no truncation or length trimming will be performed.

* Quality score:

Nucleotides with quality score less than or equal to specified value are trimmed off. The truncated reads shorter than the ending position are discarded.

- 2.a.2. Set the parameter of the **chimera**, **computing setting** and upload the **metadata**.



* **Chimeric reads filter:**

A chimeric read is a sequence originated from multiple parent sequences. Chimeric reads are generally considered contaminant. Whereas a chimeric read can be interpreted as a novel sequence, it is in fact an artifact. The higher this value is, the more chimeric reads will be used in the analysis. For most cases, 1 is the default value.

* **Computing setting:**

Specify the number of threads to speed up the analytical process. Increasing the number of threads will decrease the running time. When zero is specified, all available cores will be used. If you do not know the number to enter, leave it at the default number.

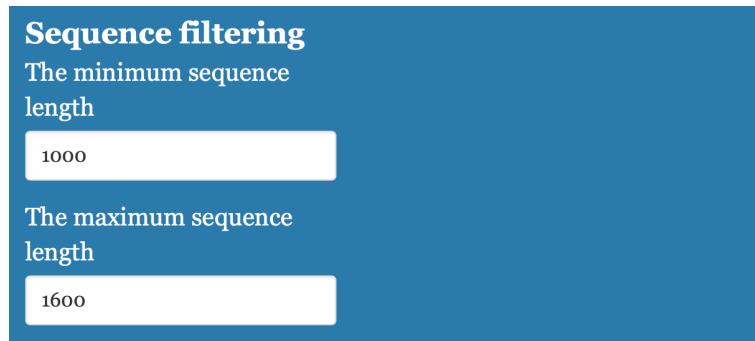
2.b. Long-read

- 2.b.1. Set the minimum and maximum sequence lengths allowed for analysis. Sequences below the minimum length and above the maximum length will be discarded.

Sequence filtering

The minimum sequence length
1000

The maximum sequence length
1600



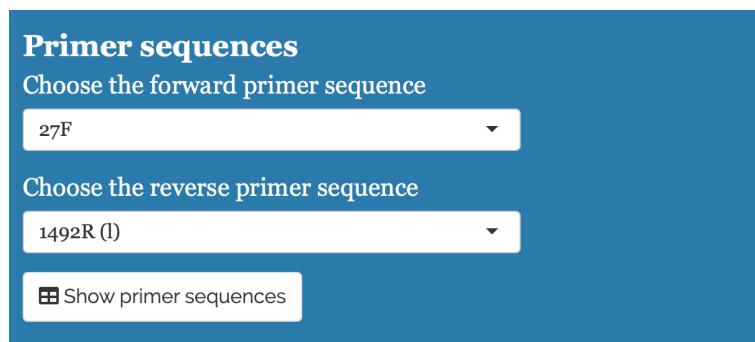
- 2.b.2. Choose the primers.

Primer sequences

Choose the forward primer sequence
27F

Choose the reverse primer sequence
1492R (l)

Show primer sequences



- 2.b.3. Assign the number of threads permitted for denoising.

Computing setting

Number of threads MOCHI can use
2



3. Click on the “**Start!**” button.

2. Sequence denoising (DADA2) for Single end

(1) Start to denoise.

 **Start!**

4. Please wait while it is running. When complete, a popup window will be displayed.

Denoising successfully!

This analysis took 1.25 mins. You can inspect the results!

- 4.1. Demo results — "Summary" summarizes the read counts of all samples.

Summary	Filter info	Sequence info	Rarefaction plot	Table	Log												
Sample read count summary																	
<table border="1"> <thead> <tr> <th>Min</th><th>Mean</th><th>Median</th><th>Max</th><th>Total</th><th>Sample.size</th></tr> </thead> <tbody> <tr> <td>897.00</td><td>4523.03</td><td>4010.50</td><td>9820.00</td><td>153783.00</td><td>34</td></tr> </tbody> </table>						Min	Mean	Median	Max	Total	Sample.size	897.00	4523.03	4010.50	9820.00	153783.00	34
Min	Mean	Median	Max	Total	Sample.size												
897.00	4523.03	4010.50	9820.00	153783.00	34												
Sample summary table																	
Show 10 <input type="button" value="▼"/> entries			Search: <input type="text"/>														
SampleID	Read count	Number of ASVs observed in															
1 L1S105	9820	63															
2 L1S140	9743	65															
3 L1S208	8701	84															
4 L1S257	8339	81															
5 L1S281	8146	72															
6 L1S57	7866	70															
7 L1S76	7780	61															

4.2. Demo results — "Summary" summarizes the read counts of all ASVs.

ASV read count summary

Min	Mean	Median	Max	Total	Number.of.ASVs
0.00	199.46	23.00	11371.00	153783.00	771

ASV summary table

ASV		Read count	Number of samples observed in
1	4b5eeb300368260019c1fbc7a3e718fc	11371	13
2	fe3offof71a38a39cf1717ec2be3a2fc	8929	16
3	d29fe3c70564fc0f69f2c03e0d1e5561	8621	25
4	868528ca947bc57b69ffd83e6b73bae	7660	10
5	154709e160e8cada6fb21115acc80f5	7410	13
6	1d2e5f3444ca750c85302ceee2473331	7185	23
7	0305a4993ecfd8ef4149fdfc7592603	5389	11
8	cb2fe0146e2fbcb101050edb996aoee2	4645	15

4.3. Demo results — "Filter info" shows filtered read counts of each sample at every step of DADA2.

Summary	Filter info	Sequence info	Rarefaction plot	Table	Log
Show 10 entries				Search:	
SampleID	Input read	Filtered read	Filtered read (%)	Denoised read	Denoised read (%)
1	L1S105	11340	8571	75.58	8499
2	L1S140	9736	7676	78.84	7604
3	L1S208	11335	9260	81.69	9146
4	L1S257	8216	6705	81.61	6627
5	L1S281	8904	7066	79.36	6975
6	L1S57	11750	9298	79.13	9259
7	L1S76	10100	8394	83.11	8336
8	L1S8	12386	7662	61.86	7623
9	L2S155	9261	4112	44.4	3932

4.4. Demo results — "Sequence info" summarizes the length and the bases of denoised sequences.

Summary Filter info Sequence info Rarefaction plot Table Log

Sequence Length Statistics

Sequence.count	Min.length	Mean.length	Max.length	Range	Standard.deviation
770	120	120	120	0	0

Seven-Number Summary of Sequence Lengths

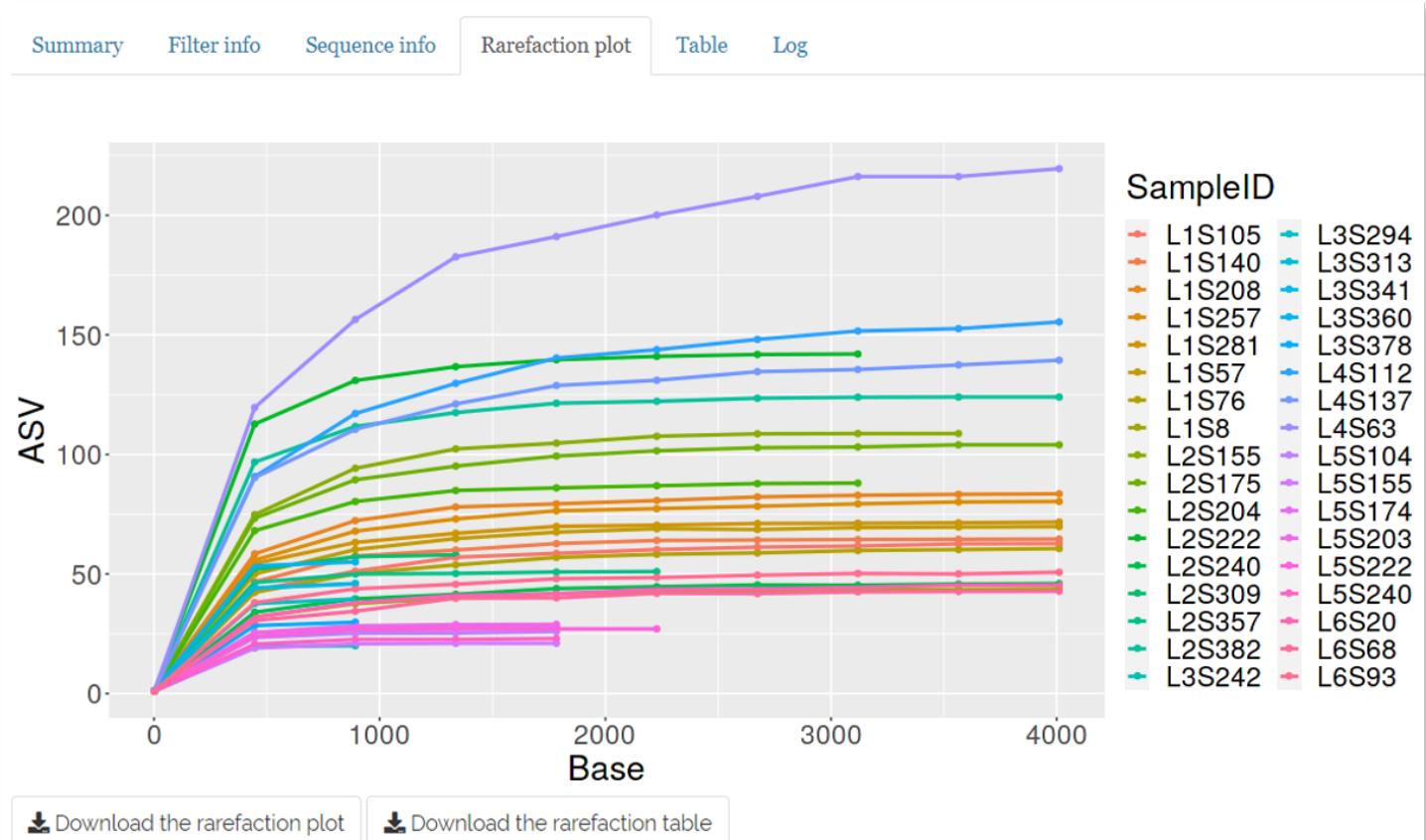
Percentile:	2%	9%	25%	50%	75%	91%	98%
Length (nts):	120	120	120	120	120	120	120

Sequence table

Show 10 entries Search:

ASV	Sequence.length	Sequence
1 4b5eeb300368260019c1fb7a3c718fc	120	TACGGAGGATCCGAGCGTTATCCGGATTATGGGTTAAAGGGAGCGTAGATGGATGTTA
2 fe3offff71a38a39cf1717ec2be3a2fc	120	TACGTAGGGTGCAGCGTTAACCGAATTACTGGCGTAAAGCGAGCGCAGACGGTTACTTA
3 d29fe3c70564fc0f69f2c03e0d1e5561	120	TACGTAGGTCCCGAGCGTTGTCCGGATTATGGCGTAAAGCGAGCGCAGGCGGTTAGATA
4 868528ca947bc57b69fdf83e6b73bae	120	TACGGAGGATCCGAGCGTTATCCGGATTATGGGTTAAAGGGAGCGTAGATGGATGTTA
5 154709e160e8cada6fb2115acc80f5	120	TACGGAGGATCCGAGCGTTATCCGGATTATGGGTTAAAGGGAGCGTAGGTGGATTGTTA
6 1d2e5f3444ca750c85302ceee2473331	120	TACGGAGGGTGCAGCGTTAACCGAATAACTGGCGTAAAGGGCACCGCAGGCGGTGACTT
7 0305a4993ecf2d8ef4149fdfc7592603	120	TACGGAGGATCCGAGCGTTATCCGGATTATGGGTTAAAGGGAGCGTAGGCGGACGCTTA

4.5. Demo results — "Rarefaction plot" shows the ASV number of every sample at different sampling depth.



4.6. Demo results — "Table" shows the read counts of each ASV in every sample. =

Summary	Filter info	Sequence info	Rarefaction plot	Table	Log				
Show 10 entries					Search: <input type="text"/>				
ASV	L1S105	L1S140	L1S208	L1S257	L1S281	L1S57	L1S76	L1S8	L2S155
1 4b5eeb300368260019c1fbc7a3c718fc	2175	0	0	0	0	2806	3308	2594	10
2 fe3offof71a38a39cf1717ec2be3a2fc	5	0	0	0	0	0	0	0	0
3 d29fe3c70564fc0f69f2c03e0d1e5561	0	0	0	0	0	0	0	0	353
4 868528ca947bc57b69ffd83e6b73bae	0	2248	2107	1177	1721	0	0	0	0
5 154709e160e8cada6fb21115acc80f5	802	1174	694	406	242	1081	930	1623	0
6 1d2e5f3449ca750c85302ceee2473331	0	0	5	0	0	0	0	5	27
7 0305a4993ecf2d8ef4149fdfc7592603	1197	647	964	909	531	414	334	243	0
8 cb2fe0146e2fbcb101050edb996a0ee2	0	0	0	7	0	0	0	0	82
9 997056ba80681bbbdd5d09aa591eadco	0	0	0	0	0	0	0	3	0
10 3c9c437f27aca05f8db167cd08off1ec	0	0	0	0	0	0	0	0	0

Showing 1 to 10 of 770 entries

Previous [1](#) [2](#) [3](#) [4](#) [5](#) ... [77](#) Next

4.7. Demo results — "Log" records the used parameters and provides a button to download the table.

Summary	Filter info	Sequence info	Rarefaction plot	Table	Log
Record					Value
time					2021-05-31 01:55:54
duration					1.87 mins
sequence_type					Single end
start_position_trim					0
end_position_trim					120
quality_score_truncate					2
chimeric_reads_min_fold_change					1
metadata_upload					TRUE
computing_setting					6

 Download

(C) Taxonomy classification

1. Select “**Sequence Preprocessing**” in the top bar, choose “**Step 3. Taxonomy classification**”.

The screenshot shows the MOCHI web interface. At the top, there is a navigation bar with links for Home, Sequence Preprocessing (which is currently selected), Taxonomy Analysis, Function Analysis, and Tutorial. Below the navigation bar, a blue header bar says "Welcome to MOCHI". On the right side of the header, there is a dropdown menu with options: Step 1. Sequence summary, Step 2. Sequence denoising, Step 3. Taxonomy classification, and a link to QIIME2. A red arrow points to the "Step 3. Taxonomy classification" link. Below the header, there is a text area stating: "MOCHI is a 16S or 18S microbiota amplicon rRNA MOCHI may also be downloaded and operated locally...".

2. Download and select **database** (Silva, Greengene, or PR2) to predict taxa.

The screenshot shows a "Database" selection page. The title is "Database" and the subtitle is "Select the reference database for taxonomy classification." There is a section titled "Choose the database" with a dropdown menu set to "Silva (Not detected)". Below the dropdown is a button labeled "Auto download database". At the bottom is a "Example" button.

2.1. Automatically download database:

- * Select a database from the drop-down menu “**Choose the database**”. Press the “**Auto download database**” button. The latest database will be pulled from the server. The downloading process may take a while depending on the file size and the network speed.

2.2. Manually download the database:

- * Silva: Follow the [link](#). Choose a version to download. Decompress the downloaded file. Copy the two folders “**rep_set**” and “**taxonomy**” to the folder “**taxa_database/silva**”.
- * Greengene: Follow the [link](#). Choose a version and download the corresponding “**otus.tar.gz**”. Decompress the downloaded files. Copy the two folders “**rep_set**” and “**taxonomy**” into the folder “**taxa_database/greengenes**”.
- * PR2: Follow the [link](#). Choose a version and download the corresponding “**pr2_version_X.XX.X_16S_mothur.fasta.gz**” and “**pr2_version_X.XX.X_16S_mothur.tax.gz**”. Decompress the downloaded files. Copy the file “**pr2_version_X.XX.X_16S_mothur.fasta**” into the folder “**taxa_database/PR2/18S/seqs**” and the file “**pr2_version_X.XX.X_16S_mothur.tax**” into the folder “**taxa_database/PR2/18S/taxonomy**”.

3. Check if your **primers** are correct.

Reference sequence filtering

1. Check primers
If incorrect, go to 'Step 1. Sequence summary' to select the correct primer.

Your forward primer is **8F** now.

Your reverse primer is **519R** now.

4. Set the minimum and maximum lengths for filtering the reference sequence.

2. Filter the reference sequence by length

Minimum length

Maximum length

[learn more](#)

*** Minimum and maximum length:**

Reference sequences not in range of the specified values will be discarded. The default values are the minimum and the maximum lengths of the denoised sequences. To disable length filtering, set the values to zero.

5. Set the number of threads for running the analysis. If zero is specified, all available cores will be used. If you do not know the number to enter, leave it at the default number (all threads - 2).

Computing setting

Number of threads MOCHI can use

6. Click on the "**Start!**" button.

3. Taxonomy classification

(1) Classify taxonomy

 **Start!**

7. Please wait while it is running. When complete, a popup window will be displayed.

Taxonomy classification has been finished!

This analysis took 55.92 secs. You can inspect the results!

8. Demo results — “taxonomy results” shows the ASV and assigned taxonomy. The three buttons at the bottom are for downloading the files to conduct the subsequent analyses.

Inspect the taxonomy classification result.

Taxonomy result

[Log](#)

ASV	Taxon	Confidence
1 4b5eeb300368260019cfbc7a3c718fc	k__Bacteria; p__Bacteroidetes; c__Bacteroidia; o__Bacteroidales; f__Bacteroidaceae; g__Bacteroides; s__	0.99340978592142
2 fe30ff0f71a38a39cf1717ec2be3a2fc	k__Bacteria; p__Proteobacteria; c__Betaproteobacteria; o__Neisseriales; f__Neisseriaceae; g__Neisseria; s__	0.89490583340447
3 868528ca947bc57b69ffdf83e6b73bae	k__Bacteria; p__Bacteroidetes; c__Bacteroidia; o__Bacteroidales; f__Bacteroidaceae; g__Bacteroides; s__	0.98368032171919
4 154709e160e8cada6fb21115acc80f5	k__Bacteria; p__Bacteroidetes; c__Bacteroidia; o__Bacteroidales; f__Bacteroidaceae; g__Bacteroides; s__	0.980528382140841
5 d29fe3c70564fc0f69f2c03e0d1e5561	k__Bacteria; p__Firmicutes; c__Bacilli; o__Lactobillales; f__Streptococcaceae; g__Streptococcus; s__	0.999999998690498
6 1d2e5f3444ca750c85302ceee2473331	k__Bacteria; p__Proteobacteria; c__Gammaproteobacteria; o__Pasteurellales; f__Pasteurellaceae; g__Haemophilus; s__parainfluenzae	0.96830683630633
7 0305a4993ecf2d8ef4149fdfc7592603	k__Bacteria; p__Bacteroidetes; c__Bacteroidia; o__Bacteroidales; f__Bacteroidaceae; g__Bacteroides; s__uniformis	0.996031457767898
8 997056ba80681bbbdd5d09aa591eadco	k__Bacteria; p__Fusobacteria; c__Fusobacteriia; o__Fusobacteriales; f__Fusobacteriaceae; g__Fusobacterium; s__	0.907315818016303
9 3e9e437f27aca05f8db167cd08off1ec	k__Bacteria; p__Bacteroidetes; c__Bacteroidia; o__Bacteroidales; f__Prevotellaceae; g__Prevotella; s__melaninogenica	0.999998818423878
10 bfbbed36e63b69fec4627424163d20118	k__Bacteria; p__Firmicutes; c__Clostridia; o__Clostridiales; f__Ruminococcaceae; g__Faecalibacterium; s__prausnitzii	0.999996821028142

Showing 1 to 10 of 501 entries

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[Download](#)

(3) Download the files for the next step.

[The taxonomic table](#)

[The ASV table](#)

[The seqs data](#)

9. Demo results — "Log" records the used parameters and provides a button to download the table.

Inspect the taxonomy classification result.	
Taxonomy result	Log
Record	Value
time	2021-03-18 05:53:21
duration	1.39 mins
sequence_type	Single end
database	Greengenes_16S_88
forward_primer	515F
reverse_primer	806R
min_length	100
max_length	400
computing_setting	6

 Download

Taxonomy analysis

(A) Upload files

1. Select “**Taxonomy analysis**” in the top bar.



Welcome to MOCHI! (Microbiota amplicOn CHaracterization Implement)

MOCHI is a 16S or 18S microbiota amplicon rRNA analytical tool for microbiota based primarily on [QIIME2](#) with a friendly web interface powered by the R package of [Shiny](#). MOCHI may also be downloaded and operated locally.

2. In the left panel, press the “Browse” button to upload the **metadata, taxonomic table** and **ASV table** files. Then, select the format of uploaded files, e.g., “.qza” or “.txt”.

These files can be downloaded from the “Sequence Preprocessing - Taxonomic classification” section. Please see [Sequence preprocessing / Taxonomic classification / step 8](#). Alternatively, you can press the “Demo” button to download the example files first and then upload the files. If sequences are 18S rRNA, please check the “18S rRNA” box.

* **Metadata (.tsv):**

The first column name must be **SampleID**.

* **Taxonomic table file (.qza):**

You can upload self-derived taxonomic table file (FeatureTable[Frequency], level: 7) from QIIME2.

* **ASV table (.qza):**

An “amplicon sequence variant” table is a higher-resolution analogue of the traditional OTU table.

* **ASV table (.txt):**

The table should include the information of ASV in the first column and Taxon in the last column.

3. Click on the “**Start!**” button to run the analysis. (Or, click on the “**Reset**” button to re-upload the files).
4. The results will be displayed in the right panel after pressing the “**Start!**” button.

Kingdom (K=3)	Phylum (K=20)	Class (K=45)	Order (K=72)	Family (K=126)	Genus (K=190)	Species (K=198)	LiS8	LiS57	LiS76	LiS105	LiS155	LiS175	LiS204
Bacteria	Bacteroidetes	Bacteroidia	Bacteroidales	Bacteroidaceae	Bacteroides	Unassigned	4217	3887	4238	2977	10	10	3
Bacteria	Proteobacteria	Betaproteobacteria	Neisseriales	Neisseriaceae	Unassigned	Unassigned	0	0	0	5	51	109	43
Bacteria	Firmicutes	Bacilli	Lactobacillales	Streptococcaceae	Streptococcus	Unassigned	30	5	0	0	377	929	126
Bacteria	Proteobacteria	Gammaproteobacteria	Pasteurellales	Pasteurellaceae	Gallibacterium	Unassigned	5	0	0	0	27	51	0
Bacteria	Bacteroidetes	Bacteroidia	Bacteroidales	Bacteroidaceae	Bacteroides	uniformis	260	553	530	1439	0	0	0
Bacteria	Firmicutes	Bacilli	Bacillales	Bacillaceae	Unassigned	Unassigned	0	0	0	0	82	227	96
Bacteria	Fusobacteria	Fusobacteriia	Fusobacteriales	Fusobacteriaceae	Fusobacterium	Unassigned	3	0	0	31	31	53	0
Bacteria	Bacteroidetes	Bacteroidia	Bacteroidales	Prevotellaceae	Prevotella	Unassigned	0	0	0	80	16	88	10
Bacteria	Proteobacteria	Gammaproteobacteria	Unassigned	Unassigned	Unassigned	Unassigned	0	0	0	0	97	41	1376
Bacteria	Firmicutes	Clostridia	Clostridiales	Ruminococcaceae	Faecalibacterium	prausnitzii	129	693	906	910	0	6	19

(B) Inspect results

- MOCHI displays the results in eight approaches: (1) Taxonomic table, (2) Taxonomic barplot, (3) Taxonomic heatmap, (4) Krona, (5) Alpha diversity, (6) Beta diversity, (7) Phylogenetic diversity, and (8) ANCOM:

1. Taxonomic table

A table describes taxonomy information and read count.

Choose the group

- SampleID
- barcode.sequence
- body.site
- year
- month
- day
- subject
- reported.antibiotic.usage
- days.since.experiment.start

Show 10 entries

Taxonomy information

Kingdom (K=3)	Phylum (K=21)	Class (K=42)	Order (K=69)	Family (K=134)	Genus (K=281)	Species (K=356)	L1S8	L1S57	L1S76	L1S105	L2S155	L2S175
Unassigned	Unassigned	Unassigned	Unassigned	Unassigned	Unassigned	Unassigned	0	0	0	0	0	0
Archaea	Crenarchaeota	Thaumarchaeota	Nitrosphaerales	Nitrosphaeraceae	Candidatus Nitrosphaera	Unassigned	0	0	0	0	0	0
Archaea	Crenarchaeota	Thaumarchaeota	Nitrosphaerales	Nitrosphaeraceae	Candidatus Nitrosphaera	SCA1145	0	0	0	0	0	0
Bacteria	Unassigned	Unassigned	Unassigned	Unassigned	Unassigned	Unassigned	0	0	0	0	25	13
Bacteria	Acidobacteria	[Chloracidobacteria]	RB41	Ellin6075	Unassigned	Unassigned	0	0	0	0	0	0
Bacteria	Actinobacteria	Acidimicrobia	Acidimicrobiales	Unassigned	Unassigned	Unassigned	0	0	0	0	0	0
Bacteria	Actinobacteria	Actinobacteria	Actinomycetales	Unassigned	Unassigned	Unassigned	0	0	0	0	7	0
Bacteria	Actinobacteria	Actinobacteria	Actinomycetales	Actinomycetaceae	Unassigned	Unassigned	0	0	0	0	0	0
Bacteria	Actinobacteria	Actinobacteria	Actinomycetales	Actinomycetaceae	Actinomyces	Unassigned	0	0	0	0	0	0
Bacteria	Actinobacteria	Actinobacteria	Actinomycetales	Actinomycetaceae	Actinomyces	Unassigned	0	0	0	0	0	0

Showing 1 to 10 of 385 entries

Previous 1 2 3 4 5 ... 39 Next

Download button

* Choose the group:

Select a group provided in the metadata to categorize read count (see below).

* Taxonomy information:

The left part of the table (the first 7 columns) represents taxonomy information. The column name indicates taxonomy levels. K denotes the number of taxa at a given level.

* Read count:

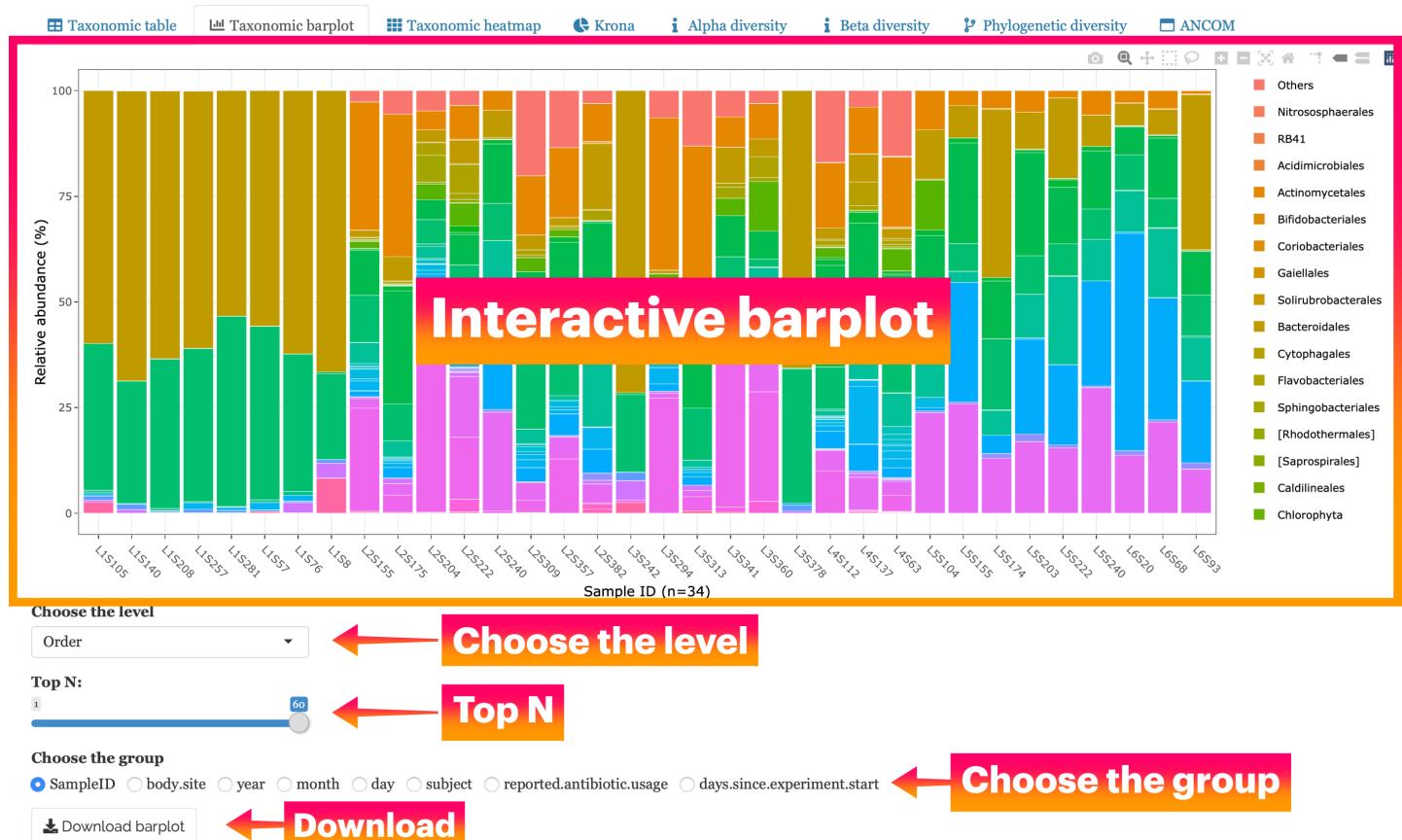
The right part of the table represents read count. The values of read count are categorized by the selected group. The column name indicates the variables of the selected group.

* Download the taxonomic table:

Click on the "Download Taxonomic table" button to download the displayed table.

2. Taxonomic barplot

An interactive barplot showing the percentage of taxa in all sample. Each taxon is represented by a sub-bar with different colors.



* Interactive barplot:

When the cursor hovers over the bar region, the information of species will be presented. Click and drag on the plot to zoom in and out. Double click on the plot to zoom back.

* Choose the level:

The taxa in the plot will be presented at the selected taxonomic level.

* Top N:

Control the numbers of taxa displayed in the plot. When you select value N, the plot will show the union of the top N relatively abundant taxa in each sample. For example, if N = 2 is selected and the top 2 abundant taxa in Sample A and Sample B are “taxa_1 and taxa_2” and “taxa_1 and taxa_3”, respectively, the plot will show the relative abundance of taxa_1, taxa_2 and taxa_3.

* Choose the group:

The barplot will be grouped based on the selected metadata.

* Download the barplot:

Click on the “Download barplot” button to download the barplot. Alternatively, click on the camera icon in the top-right region of the barplot.

3. Taxonomic heatmap

An interactive heatmap showing the log10-transformation percentage of taxa in all samples. To prevent taking logarithms of zero, a small value of 0.01 is added to all percentage values before the transformation. The transformed values are shown in color gradient.



* Interactive heatmap:

When the cursor hovers over the heatmap, the information of transformed value will be presented. Click and drag on the plot to zoom in and out. Double click on the plot to zoom back. Click on the camera icon in the top-right region of the heatmap to download the plot.

* Choose the level:

The taxa in the plot will be presented at the selected taxonomic level.

* Choose the group:

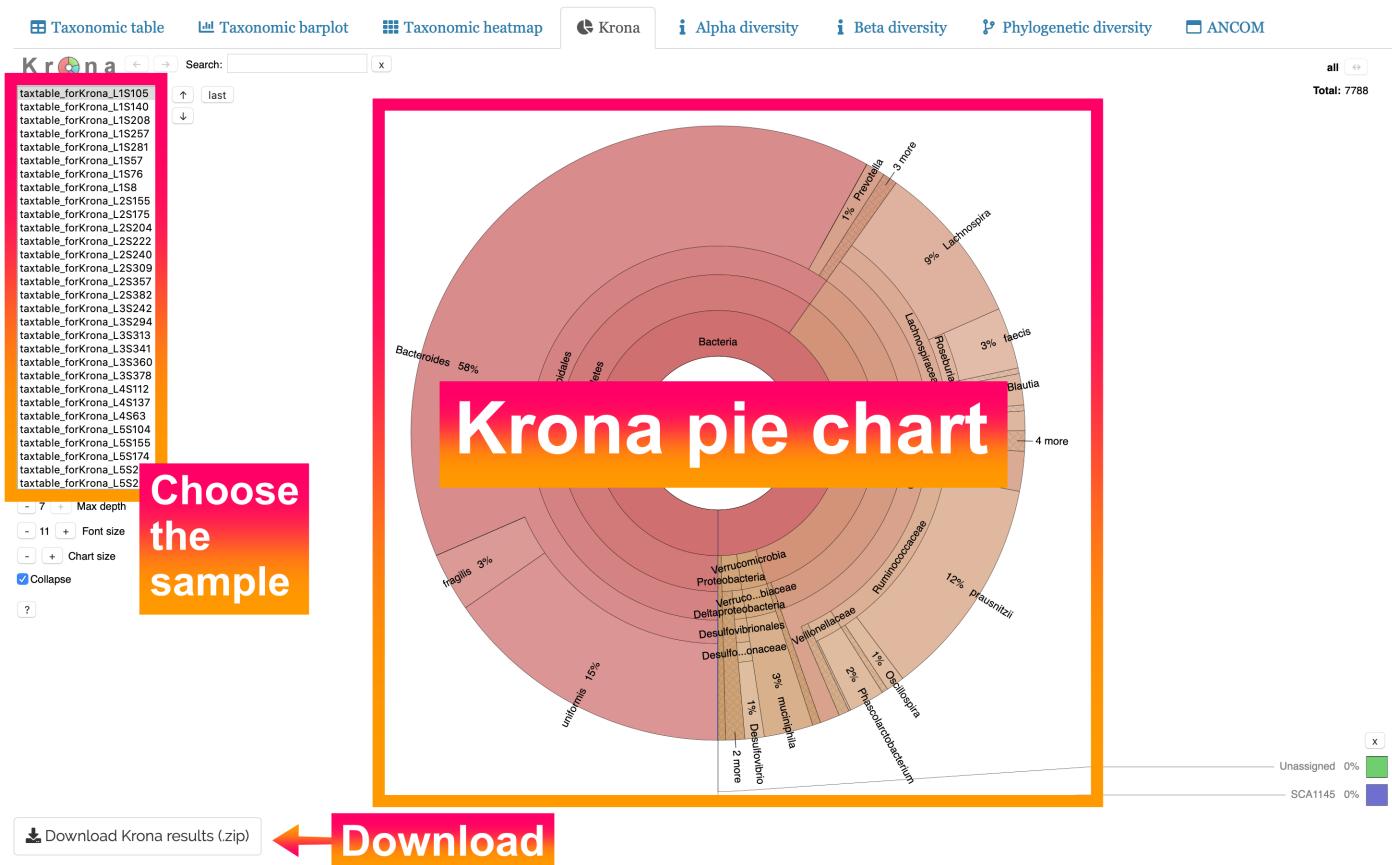
The heatmap will be grouped based on the selected metadata.

* Download the heatmap matrix:

Click on the "Download Heatmap matrix" button to download the heatmap matrix data.

4. Krona

A visualization tool allowing hierarchical data to be explored with zooming, multi-layered pie charts. [Get more information](#).



- * **Krona pie chart:**
An interactive pie plot. Single click a taxon to show the ratio of the selected taxon over different taxonomy level. Double click a taxon to zoom in the selected taxonomy level. To zoom back, click the backspace button in the top-left region.
 - * **Choose the sample:**
Select the sample to switch to the corresponding pie plot.
 - * **Download Krona results:**
Click on the “Download Krona results (.zip)” button to download the interactive pie plot (html files with Javascript).

5. Alpha diversity

Evaluation of species diversity within samples. In MOCHI, we adapt 5 indexes (ACE, Shannon diversity, InvSimpson diversity, Shannon evenness, and Simpson evenness).

5.1. Table

Table							
Show 10 entries							
	Sample	ACE	Shannon_diversity	Simpson_diversity	InvSimpson_diversity	Shannon_evenness	Simpson_evenness
1	L1S105	63	2.6823	0.8708	7.7386	0.6474	0.1228
2	L1S140	65	2.6612	0.8519	6.7521	0.6375	0.1039
3	L1S208	84	3.1173	0.8998	9.979	0.7035	0.1188
4	L1S257	81	3.2625	0.9261	13.5272	0.7424	0.1671
5	L1S281	72	3.1500	0.9150	11.5000	0.7458	0.1515
6	L1S57	70	2.9300	0.8900	11.0000	0.684	0.1067
7	L1S76	61	2.4893	0.7962	4.9065	0.6055	0.0804
8	L1S8	44	2.2034	0.7941	4.8565	0.5823	0.1104
9	L2S155	109	3.5545	0.9388	16.3338	0.7577	0.1499
10	L2S175	104	3.4393	0.9222	12.86	0.7405	0.1237

Showing 1 to 10 of 34 entries

Previous 1 2 3 4 Next

[!\[\]\(03756646c5f31ecb5e60d8bb445ffe36_img.jpg\) Download Alpha Diversity Table](#) [Download](#)

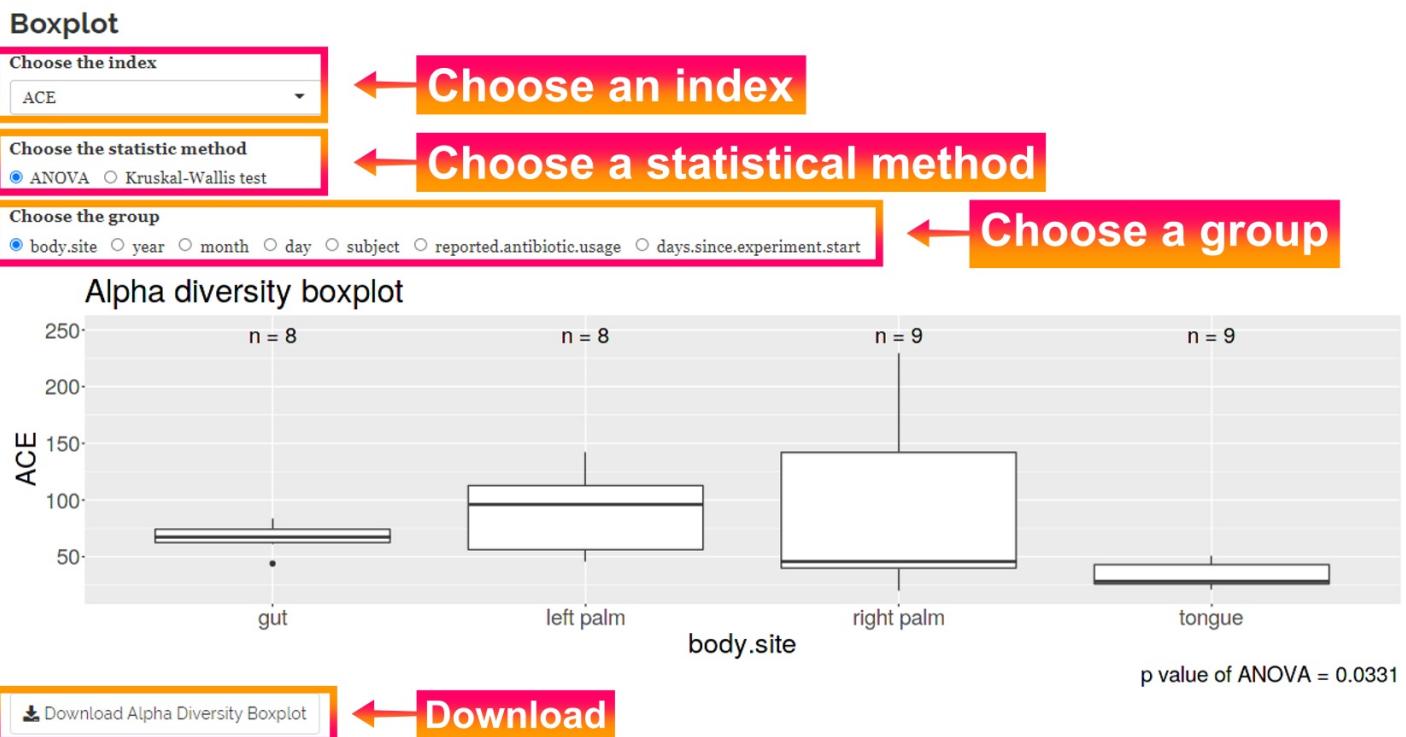
* Alpha diversity table:

This table shows the values of 5 alpha diversity indexes.

* Download the Alpha diversity table:

Click on the “Download Alpha Diversity Table” button to download the table.

5.2. Boxplot



* Choose the index:

A boxplot will be presented with the selected index.

* Choose the statistic method:

Select ANOVA (a parametric method) or Kruskal-Wallis (a nonparametric method) to test whether the distribution of the index is significantly different among the groups.

* Choose the group:

The values of the index in the boxplot will be grouped based on the selected metadata.

* Download the Alpha diversity boxplot:

Click on the “Download Alpha Diversity Boxplot” button to download the boxplot.

5.3. Post hoc analysis

Post hoc analysis

Tukey test

Group A	Group B	Diff	P value
tongue	right palm	-52.78	0.06
tongue	left palm	-57.46	0.04
tongue	gut	-34.96	0.34
right palm	left palm	-4.68	1.00
right palm	gut	17.82	0.82
left palm	gut	22.50	0.72

 Download Alpha Diversity statistical result

Post hoc analysis

Dunn test

Group A	Group B	Z	P value
gut	left palm	-0.58	0.28
gut	right palm	0.76	0.22
gut	tongue	2.89	0.00
left palm	right palm	1.35	0.09
left palm	tongue	3.49	0.00
right palm	tongue	2.20	0.01

 Download Alpha Diversity statistical result

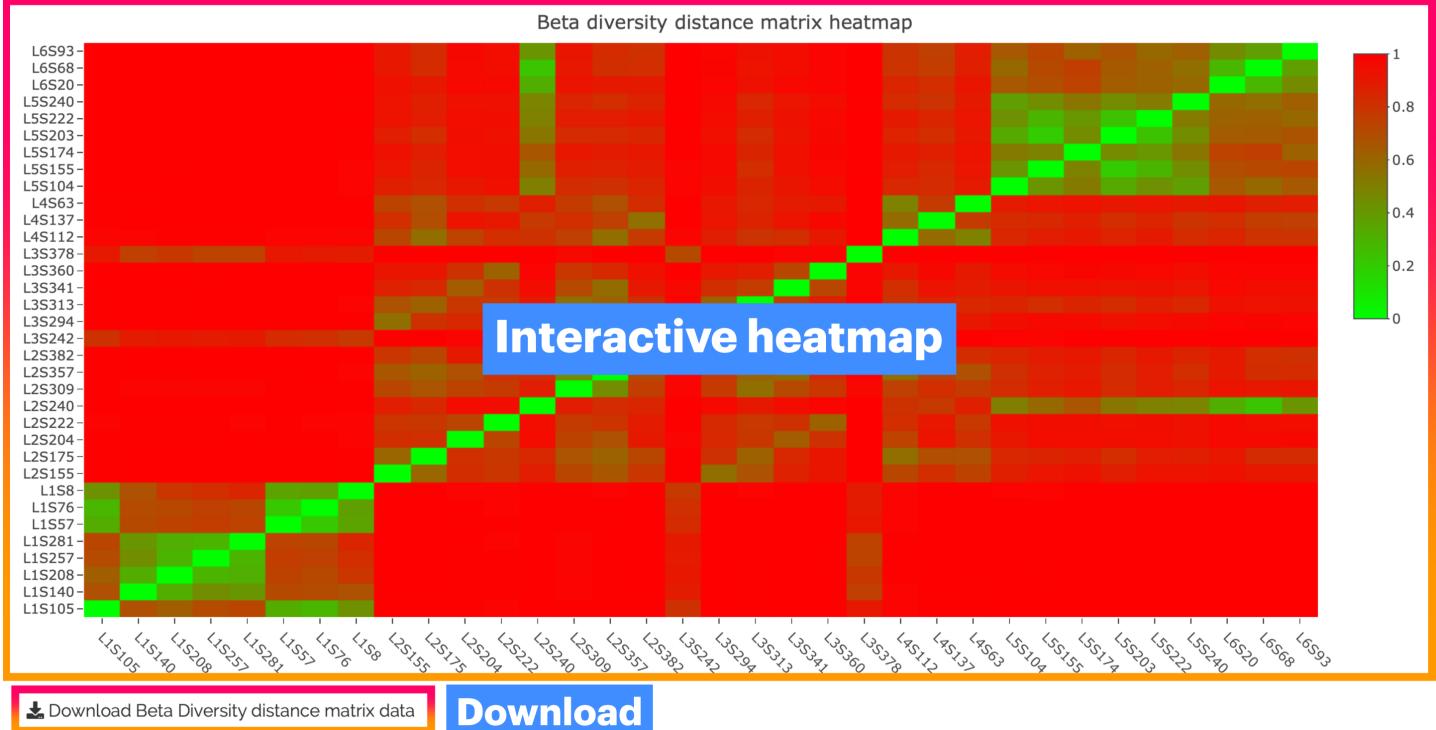
- * If ANOVA is selected when creating the boxplot, the Tukey test will be used for the post-hoc test. If Kruskal-Wallis is selected, then the Dunn test will be used.
- * Download the Alpha diversity post hoc test result:
Click on the “Download Alpha Diversity statistical result” button to download the post-hoc test result.

6. Beta diversity

Evaluation of species diversity between samples. In MOCHI, we use the Bray-Curtis index.

6.1. Distance matrix

Beta diversity table (Bray-Curtis)



* Interactive heatmap:

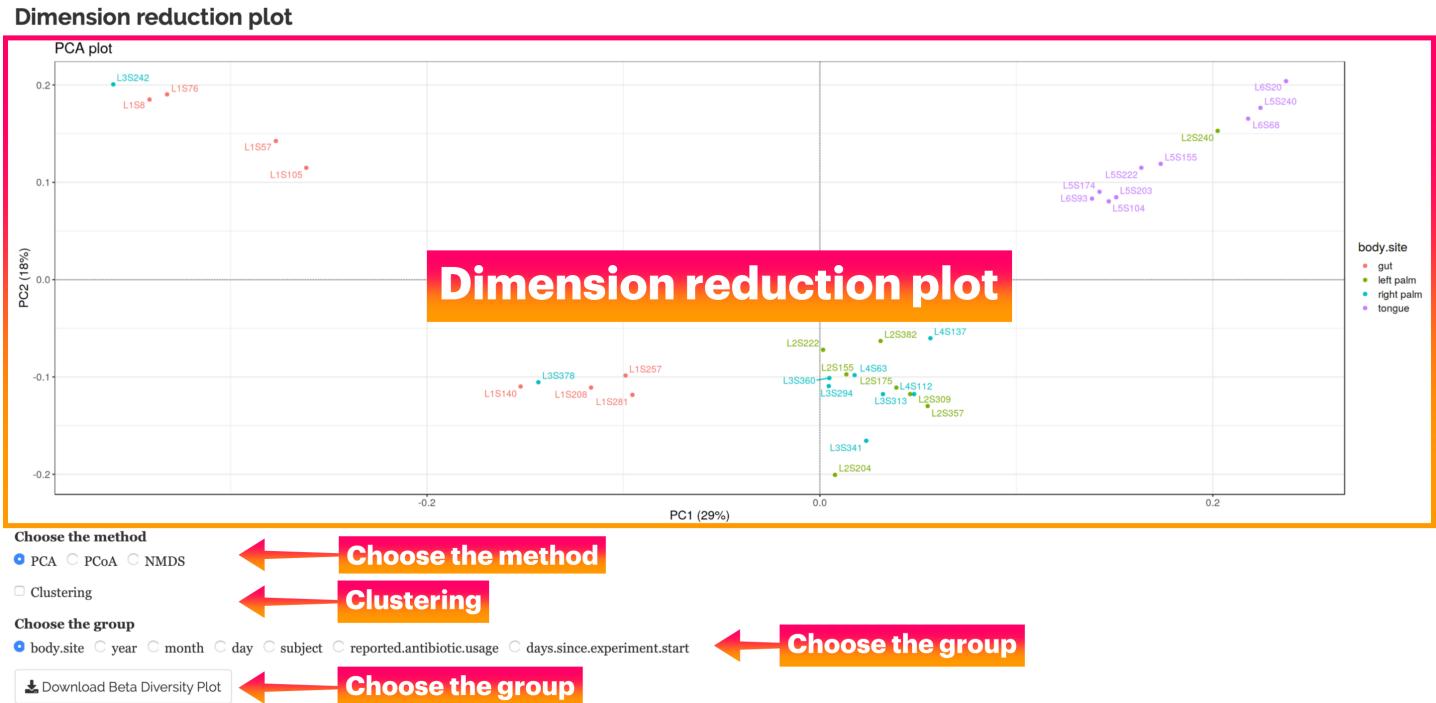
When a cursor hovers over the heatmap, the distance between species will be presented. Click and drag on the plot to zoom in and out. Double click on the plot to zoom back. Click on the camera icon in the top-right region of the heatmap to download the plot.

* Download distance matrix:

Click on the button "Download Beta Diversity distance matrix data" to download the matrix data.

* The values shown in the heatmap are the original values plus 0.01 and then natural log transformed.

6.2. Dimension-reduction plot



- * We provide three dimension-reduction methods for visualization of beta diversity: PCA (Principal Component Analysis), PCoA (Principal Co-ordinates Analysis) and NMDS (Non-metric Multidimensional Scaling) for visualization of beta diversity.
- * Choose the method:
Select a dimension reduction method.
- * Clustering:
When checking the box, samples in the same group will be surrounded by a circle.
- * Choose the group:
Samples will be labeled in colors based on the selected metadata.
- * Download the Beta diversity plot:
Click on the “Download Beta Diversity Plot” button to download the plot.

6.3. Statistical analysis

Statistical analysis

PERMANOVA		ANOSIM		MRPP							
R ²	P value	R	P value	A	P value						
0.3999	0.001	0.6855	0.001	0.2085	0.001						
Download PERMANOVA table		Download ANOSIM table		Download MRPP table							
Pairwise PERMANOVA		Pairwise ANOSIM		Pairwise MRPP							
Comparisons	R ²	P value	Adjusted P value	Comparisons	R	P value	Adjusted P value	Comparisons	A	P value	Adjusted P value
gut - left palm	0.3983	0.001	0.0012	gut - left palm	1	0.001	0.0015	gut - left palm	0.2055	0.001	0.0012
gut - right palm	0.2834	0.001	0.0012	gut - right palm	0.6686	0.001	0.0015	gut - right palm	0.1456	0.001	0.0012
gut - tongue	0.5474	0.001	0.0012	gut - tongue	1	0.002	0.0024	gut - tongue	0.3046	0.001	0.0012
left palm - right palm	0.0585	0.544	0.544	left palm - right palm	-0.0538	0.78	0.78	left palm - right palm	-0.0018	0.543	0.543
left palm - tongue	0.2985	0.001	0.0012	left palm - tongue	0.6953	0.001	0.0015	left palm - tongue	0.1373	0.001	0.0012
right palm - tongue	0.276	0.001	0.0012	right palm - tongue	0.5343	0.001	0.0015	right palm - tongue	0.1412	0.001	0.0012
Download pairwise PERMANOVA table		Download pairwise ANOSIM table		Download pairwise MRPP table							

- * We provide three statistical methods, including PerMANOVA (Permutational Multivariate Analysis of Variance), ANOSIM (Analysis of Similarities) and MRPP (Multiple Response Permutation Procedure), to test whether beta diversity is significantly different among groups or between pairs of groups. The adjusted p-values are multiple testing corrected using the Benjamini-Hochberg method.
- * Download the table of statistical results:
Click on the button below the table to download the statistical results.

7. Phylogenetic diversity

A species diversity considers the genetic distance. In MOCHI, we use Faith PD (a kind of alpha diversity which considers the genetic distance) and Unifrac distance (a kind of beta diversity which considers the genetic distance).

Phylogenetic diversity is a measure of diversity that take the genetic distance between species into consideration.

Upload the sequence file ⓘ

Browse... No file selected

Sampling depth

897

learn more

Number of threads MOCHI can use

6

The default value is (number of threads on the system -2).

Start!

Start

example files

* Upload the sequence file:

Upload the sequence file (.qza). If you have already finished the “Sequence Preprocessing” steps, download the file from “Sequence Preprocessing - Taxonomic classification” section and upload. Please see [Sequence preprocessing / Taxonomic classification / step 8](#).

* Sampling depth:

Samples with total count smaller than set value will be dropped from the diversity analysis. The default value is the smallest total count among samples where no sample will be dropped.

* Number of threads:

The number of threads to use for multithreaded process. The default value is all threads minus two.

* Start:

Click on the “Start!” button to execute the analysis after the above files and parameters have been uploaded and set.

- 7.1. Faith PD table: Faith PD (Faith's Phylogenetic Diversity) is a commonly used phylogenetic index. PD is a species diversity that considers genetic distance among species.

Faith PD table

Show **10** entries Search:

	SampleID	FaithPD
1	L1S105	7.03504527906064
2	L1S140	6.81348963332276
3	L1S208	7.56734619259508
4	L1S257	6.85786737645975
5	L1S281	6.75666778936291
6	L1S57	6.63365160132782
7	L1S76	6.33300452179527
8	L1S8	5.36655055450142
9	L2S155	18.3846874539932
10	L2S175	16.260657028738

Showing 1 to 10 of 34 entries

Previous 2 3 4 Next

 Download Faith PD table

Download

* Faith PD table:

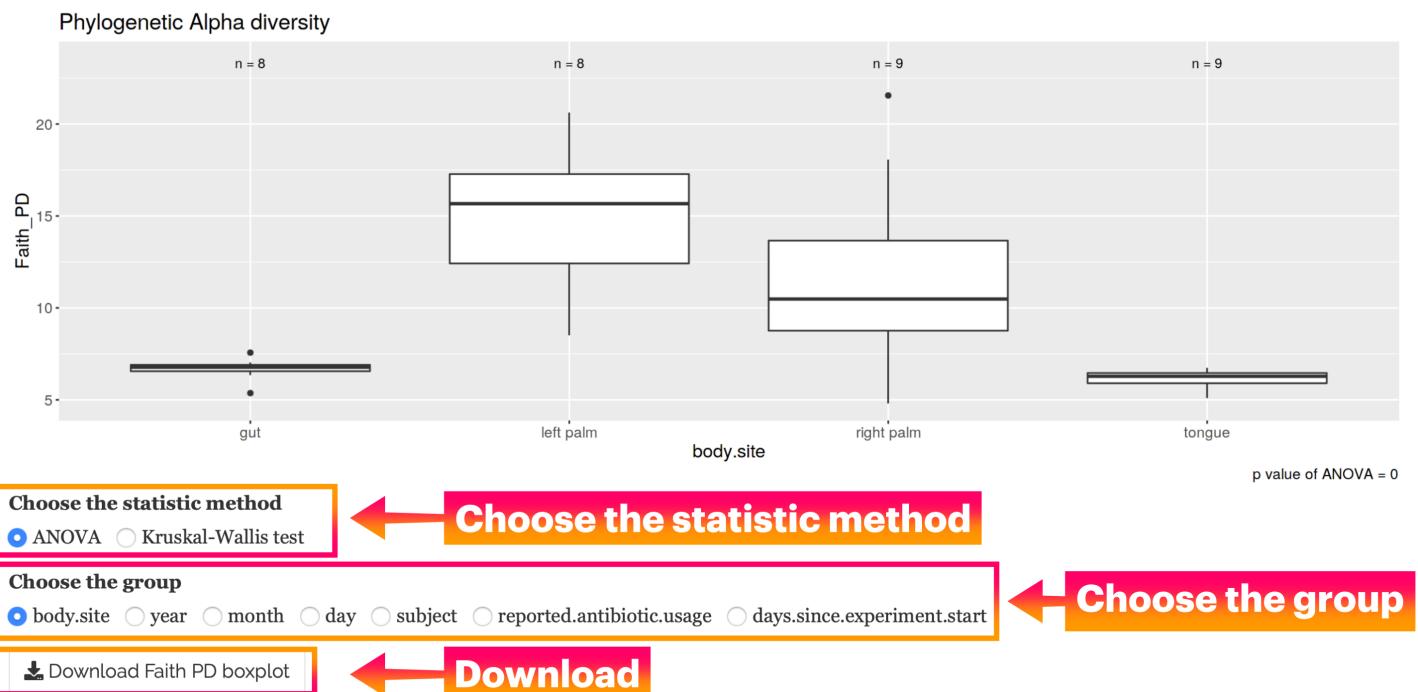
This table shows the Faith PD (phylogenetic diversity) of all samples.

* Download the Faith PD table:

Click on the “Download Faith PD table” button to download the table.

7.2. Faith PD boxplot: The distribution of Faith PD values using a boxplot.

Faith PD boxplot



* Choose the statistic method:

Select ANOVA (parametric method) or Kruskal-Wallis (nonparametric method) to test whether the distribution of Faith PD is significantly different among the groups.

* Choose the group:

Faith PD will be grouped based on the selected metadata.

* Download the Faith PD boxplot:

Click on the “Download Faith PD Boxplot” button to download the boxplot.

7.3. Post hoc analysis

Post hoc analysis

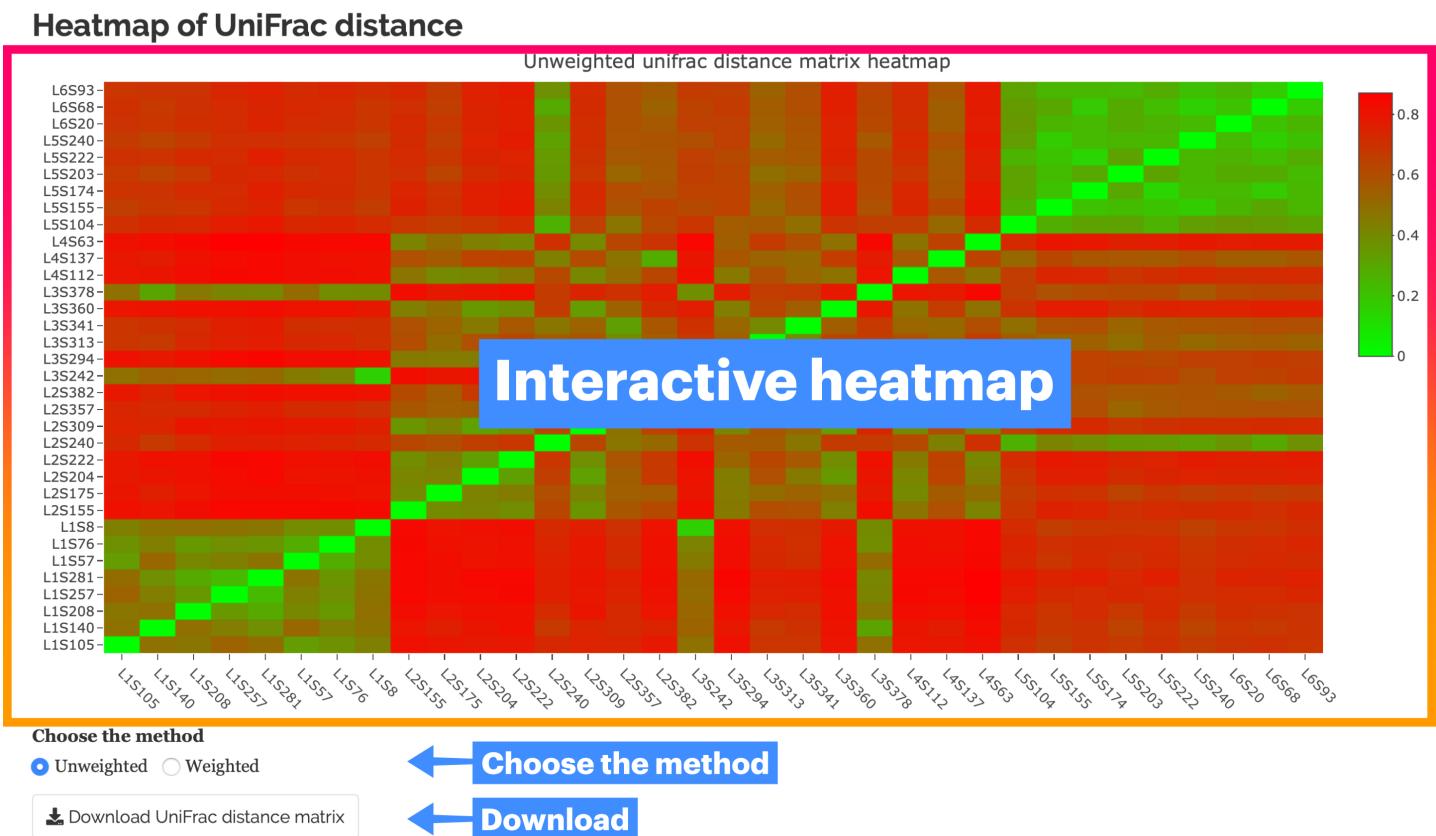
Tukey test

Group A	Group B	Diff	P value
tongue	right palm	-5.60	0.01
tongue	left palm	-8.65	0.00
tongue	gut	-0.50	0.99
right palm	left palm	-3.05	0.30
right palm	gut	5.10	0.03
left palm	gut	8.15	0.00

 Download Faith PD post hoc result

- * If ANOVA is selected when creating the Faith PD boxplot, the Tukey test will be used for the post-hoc test. If Kruskal-Wallis is selected, then the Dunn test will be used.
 - * Download the Faith PD post hoc result:
Click on the “Download Faith PD post hoc result” button to download the result.
-

7.4. Heatmap of UniFrac distance



* **Interactive heatmap:**

When the cursor hovers over the heatmap, the information of species will be presented. Click and drag on the plot to zoom in and out. Double click on the plot to zoom back. Click on the camera icon in the top-right region of the heatmap to download the plot.

* **Choose the method:**

Select unweighted UniFrac (not consider the richness of taxa) or weighted UniFrac (consider the richness of taxa).

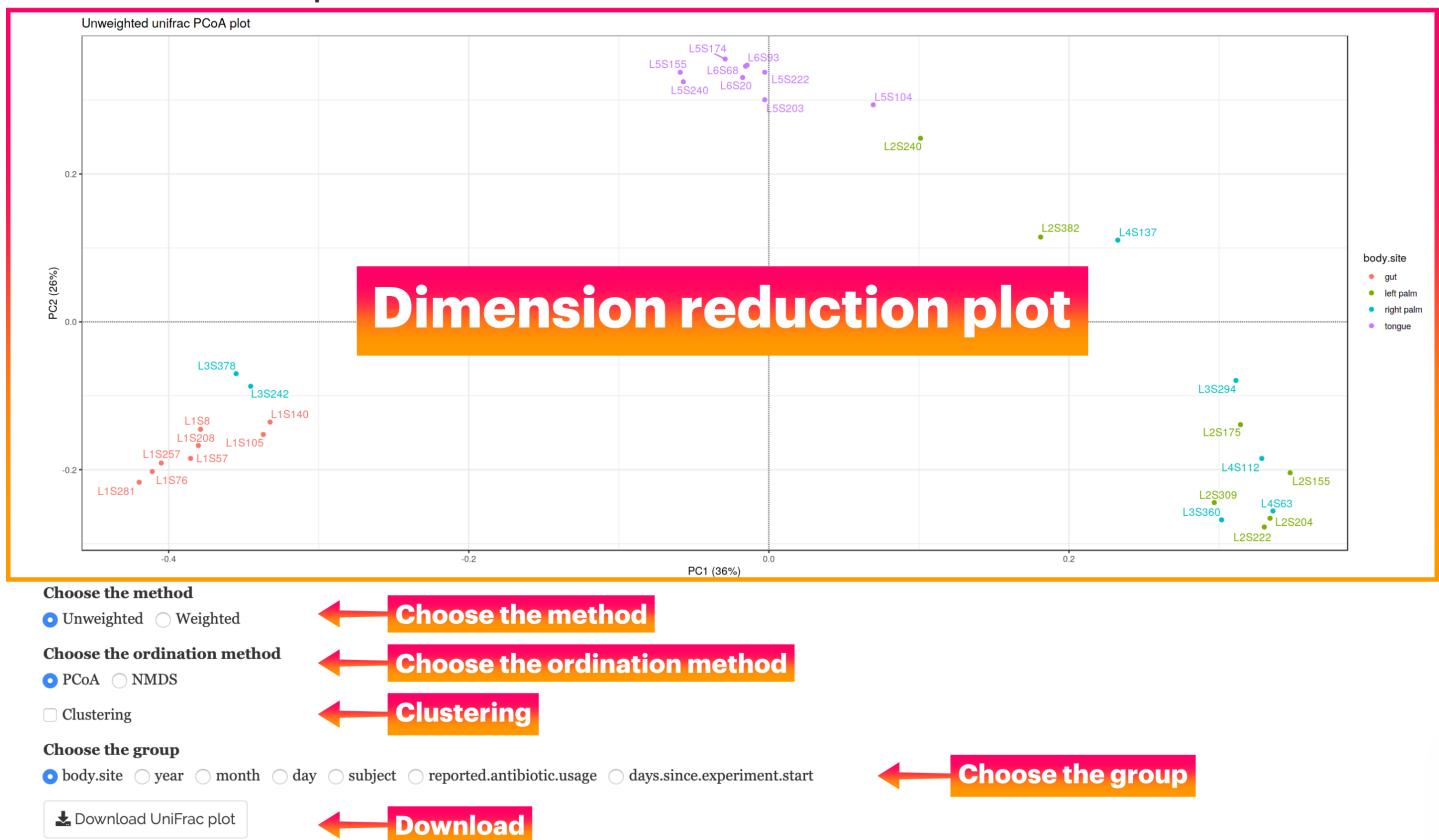
* **Download heatmap matrix:**

Click on the “Download UniFrac distance matrix” button to download the matrix data.

* **The values shown in the heatmap are the original values plus 0.01 and then natural log transformed.**

7.5. Dimension-reduction plot of UniFrac distance

Dimension reduction plot of UniFrac distance



- * We provide two dimension-reduction methods for visualizing UniFrac distance: PCoA (Principal Co-ordinates Analysis) and NMDS (Non-metric Multidimensional Scaling).
- * Choose the method:
Select unweighted UniFrac (not consider the richness of taxa) or weighted UniFrac (consider the richness of taxa).
- * Choose the ordination method:
Select a dimension reduction method.
- * Clustering:
When checking the box, samples in the same group will be surrounded by a circle.
- * Choose the group:
The samples in the plot will be labeled using colors based on the selected metadata.
- * Download the UniFrac plot:
Click on the “Download UniFrac Plot” button to download the plot.

7.6. Statistical analysis

Statistical analysis

PERMANOVA

R^2	P value
0.1567	0.042

[Download PERMANOVA table](#)

ANOSIM

R	P value
0.1219	0.027

[Download ANOSIM table](#)

MRPP

A	P value
0.0488	0.02

[Download MRPP table](#)

Pairwise PERMANOVA

Comparisons	R^2	P value	Adjusted P value
gut - left palm	0.5049	0.001	0.0012
gut - right palm	0.3261	0.001	0.0012
gut - tongue	0.6587	0.001	0.0012
left palm - right palm	0.0649	0.394	0.394
left palm - tongue	0.4563	0.001	0.0012
right palm - tongue	0.3071	0.001	0.0012

[Download pairwise PERMANOVA table](#)

Pairwise ANOSIM

Comparisons	R	P value	Adjusted P value
gut - left palm	0.9933	0.002	0.0024
gut - right palm	0.5742	0.001	0.0015
gut - tongue	1	0.001	0.0015
left palm - right palm	-0.0191	0.504	0.504
left palm - tongue	0.7509	0.001	0.0015
right palm - tongue	0.4767	0.001	0.0015

[Download pairwise ANOSIM table](#)

Pairwise MRPP

Comparisons	A	P value	Adjusted P value
gut - left palm	0.2643	0.001	0.0012
gut - right palm	0.1556	0.001	0.0012
gut - tongue	0.3811	0.001	0.0012
left palm - right palm	0.0008	0.45	0.45
left palm - tongue	0.2476	0.001	0.0012
right palm - tongue	0.1856	0.001	0.0012

[Download pairwise MRPP table](#)

- * We provide three statistical methods, including PerMANOVA (Permutational Multivariate Analysis of Variance), ANOSIM (Analysis of Similarities) and MRPP (Multiple Response Permutation Procedure), to test whether UniFrac distance is significantly different among groups or between pairs of groups. The adjusted p-values are multiple testing corrected using the Benjamini-Hochberg method.

- * Download statistical result table:

Click on the button below the table to download the statistical results.

8. ANCOM

Analyze composition of microbiomes. Used for comparing the composition of microbiomes in two or more populations. [Get more information](#).

 Taxonomic table  Taxonomic barplot  Taxonomic heatmap  Krona  Alpha diversity  Beta diversity
 Phylogenetic diversity  ANCOM

ANCOM (Analysis of composition of microbiomes) is used for comparing the composition of microbiomes in two or more populations.

Select an attribute comparison

Select an attribute comparison.

body.site year month day subject reported.antibiotic.usage days.since.experiment.start

Choose the level

Phylum

▼

Choose the level

 Start!

Start

* Select an attribute comparison:

Select a group. ANCOM will then find significantly different abundant taxa among subgroups in that group.

* Choose the level:

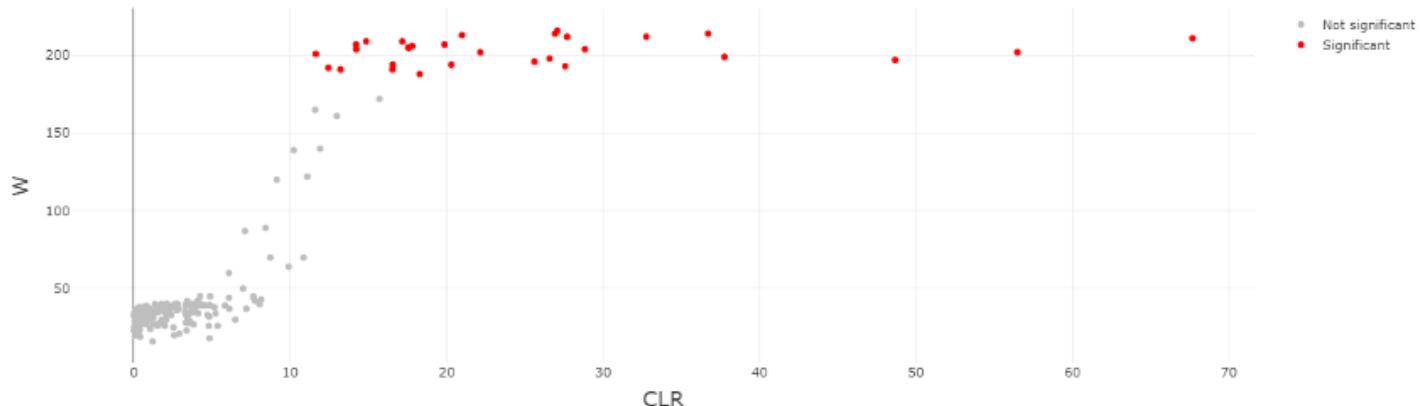
Select a taxonomic level for comparison.

* Start:

Click on the “Start!” button to execute the analysis.

- 8.1. ANCOM volcano plot: An interactive plot which shows the significantly different abundant taxa (red). When a cursor hovers over the dot, the taxa information will be presented.

ANCOM Volcano Plot (body.site-Genus)



The W value is the number of sub-hypotheses that have rejected for a given taxon in ANCOM analysis.
The clr represents log-fold change relative to the average microbe.

- * Download the ANCOM volcano plot:
Click on the camera icon to save the plot.
-

8.2. ANCOM statistical results: A table shows the W values for all taxa.

ANCOM results (Taxa with significant W value)

Show 10 entries		Search:						
	Kingdom	Phylum	Class	Order	Family	Genus	Species	W
1	Bacteria	Fusobacteria						20
2	Bacteria	Proteobacteria						20
3	Bacteria	Actinobacteria						20
4	Bacteria	Cyanobacteria						20
5	Bacteria	Firmicutes						19
6	Bacteria	Bacteroidetes						19
7	Bacteria	Verrucomicrobia						18

Showing 1 to 7 of 7 entries

Previous 1 Next

 Download the ANCOM result table (Contain all species)

* Download the ANCOM result table:

Click on the “Download the ANCOM result table” button to download the results. The table will contain the W values for all taxa.

Function analysis

The database [FAPROTAX](#) is used to predict the function of microbiota.

(A) Upload files

2. Select “**Function analysis**” in the top bar.



Welcome to MOCHI! (Microbiota amplicOn CHaracterization Implement)

MOCHI is a 16S or 18S microbiota amplicon rRNA analytical tool for microbiota based primarily on [QIIME2](#) with a friendly web interface powered by the R package of [Shiny](#). MOCHI may also be downloaded and operated locally.

3. In the left panel, press the “**Browse**” buttons to upload metadata file and taxonomic table. Alternatively, you can press the “Example files” button to download the example files first and then upload.

The image shows the MOCHI upload interface. It has two main sections: "Upload the metadata file" and "Upload the taxonomic table file". Each section contains a "Browse..." button and a "No file selected" message. Below these are dropdown menus for "Choose file format" (with options for MOCHI/QIIME2 output (.qza) and Plain text table (.txt), where Plain text table is selected). At the bottom of each section is a large orange "Upload" button with a white arrow pointing to it. The "Start!" button in the footer is also highlighted with a red box. Other buttons in the footer include "Reset", "learn more", and "Example files". To the right of the upload sections, there are links for "Function annotation table" and "Function barplot".

4. Click on the “**Start!**” button to conduct the analysis. (Or, click on the “**Reset**” button to re-upload the files.)

(B) Inspect result

1. Function annotation table

Display reads of the function types in every sample.

Function annotation table

Show 10 entries Search:

Type	L1S105	L1S140	L1S208	L1S257	L1S281	L1S57	L1S76	L1S8	L2S155	L2S175	L2S204
1 methanol_oxidation	0	0	0	0	0	0	0	0	34	12	6
2 methylotrophy	0	0	0	0	0	0	0	0	34	12	6
3 aerobic_ammonia_oxidation	0	0	0	0	0	0	0	0	0	0	0
4 nitrification	0	0	0	0	0	0	0	0	0	0	0
5 sulfate_respiration	79								0	0	2
6 sulfur_respiration	0								0	0	0
7 thiosulfate_respiration	0	0	0	0	0	0	0	0	0	0	0
8 respiration_of_sulfur_compounds	79	40	21	27	24	35	29	62	0	0	2
9 arsenate_detoxification	0	0	0	0	0	0	0	0	0	0	0
10 dissimilatory_arsenate_reduction	0	0	0	0	0	0	0	0	0	0	0

Showing 1 to 10 of 54 entries Previous [1](#) [2](#) [3](#) [4](#) [5](#) [6](#) Next

[Download the function annotation table](#) [Download](#)

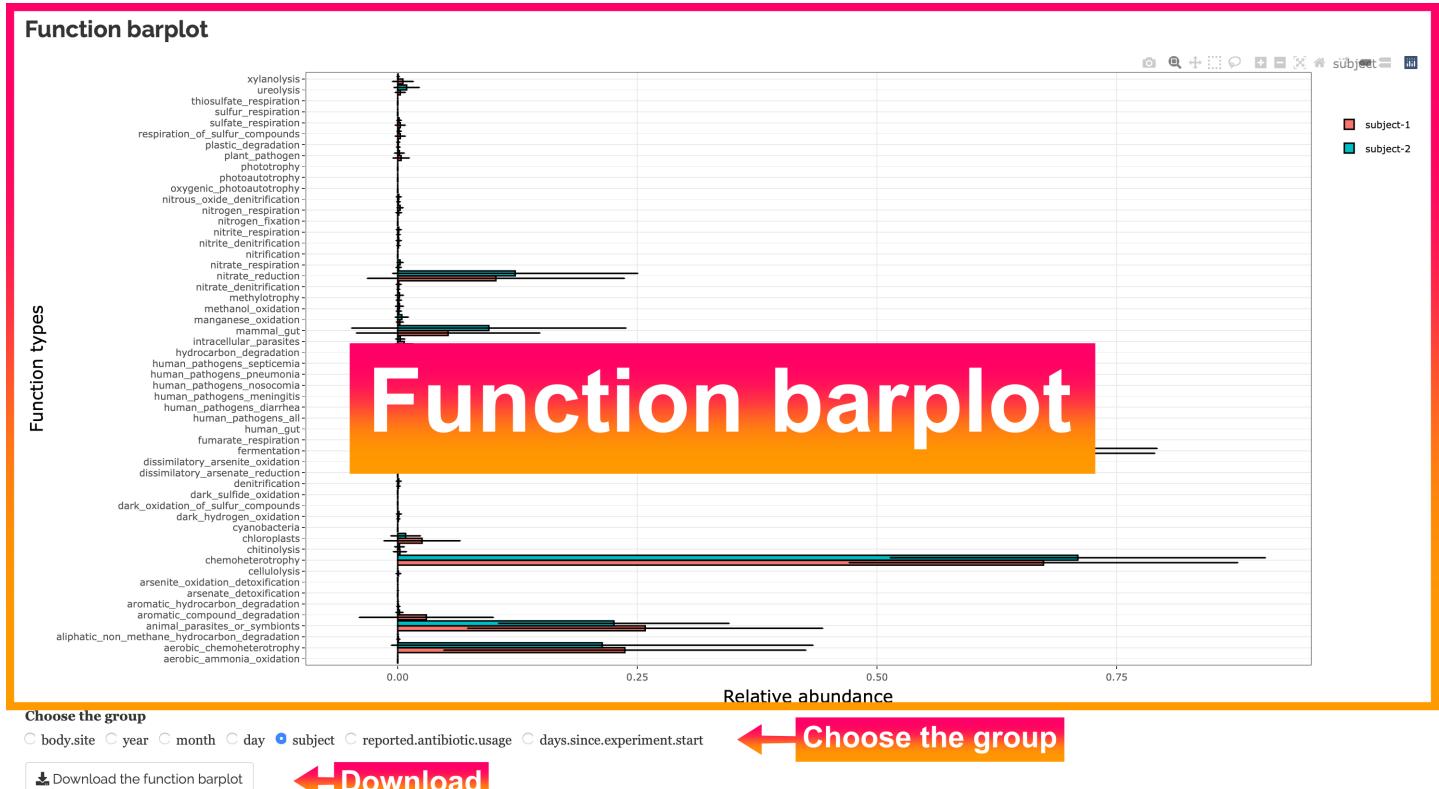
* Summary: Essential information regarding the function prediction.

* Download the function table:

Click on the “Download the function table” button to download the table.

2. Function barplot

The horizontal bars indicate reads of each function and are grouped based on the metadata.



- * Choose the group: The bars will be categorized based on the selected metadata.
- * Download the function barplot:
Click on the “Download the function barplot” button to download the barplot.