

R Boot Camp Problem Set

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Establishing reliable biomarkers for assessing and validating clinical diagnosis at early prodromal stages of Parkinson's disease is crucial for developing therapies to slow or halt disease progression. This data set uses whole blood gene expression profiling from over 500 individuals where we will attempt to find a gene signature. This repository contains the gene expression profiles collected in the GENEPARK consortium. The main study sought a classifier for IPD. These data contain 233 healthy controls, 205 IPD patients, and 48 patients with other neurodegenerative diseases (NDD). Other samples are available in the data and can be used for additional analyses. The largest class of these additional samples are 22 samples from genetic unaffected controls and 41 genetic PD patients.

Note: the original study which uploaded this data to NIH Geo is not yet published.

Data Wrangling

Let's start by loading in our data sets. Download these from the sharepoint site, and make a new folder for R bootcamp. We'll switch to this directory here.

Note that we have both a phenotype file, as well as a file which includes the normalized and log transformed expression values. We can use the read.csv function to load in these files.

```
library(dplyr)

##
## Attaching package: 'dplyr'

## The following objects are masked from 'package:stats':
##
##   filter, lag

## The following objects are masked from 'package:base':
##
##   intersect, setdiff, setequal, union
```

```
library(stringr)

pheno<-read.csv("parkPheno.csv")
expr<-read.csv("simulatedData.csv")
```

We should start by summarizing both these files. Try the following functions: head(), and View(). Note that while the dimensions on our phenotype file are reasonable, we have 552 columns in our expression file. Just summarize the first 10 columns of this file.

```
## Enter your own code here
```

```
head(pheno)
```

```
##      geo_accession submission_date last_update_date type      tissue      organism
## 1    GSM2631171      May 17 2017      May 20 2017  RNA Whole blood Homo sapiens
## 2    GSM2631309      May 17 2017      May 20 2017  RNA Whole blood Homo sapiens
## 3    GSM2631219      May 17 2017      May 20 2017  RNA Whole blood Homo sapiens
## 4    GSM2630775      May 17 2017      May 20 2017  RNA Whole blood Homo sapiens
## 5    GSM2631147      May 17 2017      May 20 2017  RNA Whole blood Homo sapiens
## 6    GSM2630853      May 17 2017      May 20 2017  RNA Whole blood Homo sapiens
##      subject_id disease_label      sex mutated_pd_genes age_at_exam age_at_symptoms
## 1      L2899    ATYPICAL_PD    Male             NONE             NA             53
## 2      L2872    ATYPICAL_PD    Male             NONE             NA             64
## 3      L2131    ATYPICAL_PD    Male             NONE             NA             NA
## 4      L2573             CBD    Female             NONE             NA             60
## 5      L2697             CBD    Female             NONE             NA             66
## 6      L3031    CONTROL      Male             NONE             NA             41
##      updrs updrs_ii updrs_iii_score_on updrs_iii_score_off updrs_iv hoehn_yahr_on
## 1      1      4      19      0      0      2
## 2      0      0      0      0      0      9
## 3      0      0      0      0      0      0
## 4      0      0      0      0      0      9
## 5      0      0      30      0      0      9
## 6      0      0      1      0      0      8
##      hoehn_yahr_off moca_score
## 1      0      21
## 2      0      0
## 3      0      0
## 4      0      0
## 5      0      0
## 6      0      30
```

```
View(pheno)
```

```
summary(expr[1:20668,1:10, drop=FALSE])
```

```
##      X      GeneName      GSM2631171      GSM2631309
## Min.   : 1      A1BG      : 1      Min.   : -5.223788      Min.   : -6.09018
## 1st Qu.: 5168    A1BG-AS1: 1      1st Qu.: -0.960423      1st Qu.: -0.92906
## Median :10334    A1CF      : 1      Median : -0.004842      Median : 0.01385
## Mean   :10334    A2M      : 1      Mean   : -0.009648      Mean   : 0.01249
## 3rd Qu.:15501    A2M-AS1 : 1      3rd Qu.: 0.953228      3rd Qu.: 0.95912
## Max.   :20668    (Other) :20662      Max.   : 5.766301      Max.   : 5.66627
##      NA's      : 1
##      GSM2631219      GSM2630775      GSM2631147      GSM2630853
## Min.   : -6.39097      Min.   : -5.206869      Min.   : -5.27578      Min.   : -6.115736
## 1st Qu.: -0.97337      1st Qu.: -0.981831      1st Qu.: -0.96379      1st Qu.: -0.944666
## Median : -0.01097      Median : 0.001772      Median : 0.01906      Median : -0.007942
## Mean   : -0.00354      Mean   : -0.000010      Mean   : 0.00298      Mean   : -0.009892
## 3rd Qu.: 0.95324      3rd Qu.: 0.971013      3rd Qu.: 0.98545      3rd Qu.: 0.945826
## Max.   : 6.56118      Max.   : 5.275719      Max.   : 5.18612      Max.   : 5.570111
##
```

```
##      GSM2630769      GSM2631196
## Min.      :-5.608142 Min.      :-6.303044
## 1st Qu.: -0.968002 1st Qu.: -0.970730
## Median : -0.001583 Median : -0.004689
## Mean    : 0.014813 Mean     : -0.006484
## 3rd Qu.: 0.987677 3rd Qu.: 0.977216
## Max.    : 5.591597 Max.     : 5.434250
##
```

Try summarizing the phenotype data:

```
## Enter your own code here
```

```
summary(pheno)
```

```
##      geo_accession      submission_date      last_update_date      type
## GSM2630758: 1      May 17 2017:550      May 20 2017:550      RNA:550
## GSM2630759: 1
## GSM2630760: 1
## GSM2630761: 1
## GSM2630762: 1
## GSM2630763: 1
## (Other)      :544
##      tissue      organism      subject_id      disease_label
## Whole blood:550      Homo sapiens:550      B25      : 1      CONTROL      :233
##      B27      : 1      IPD      :205
##      B28      : 1      GPD      : 41
##      B29      : 1      GENETIC_UNAFFECTED: 22
##      B32      : 1      HD      : 19
##      B36      : 1      MSA      : 8
##      (Other):544      (Other)      : 22
##      sex      mutated_pd_genes      age_at_exam      age_at_symptoms
##      : 45      NONE :428      Min.      :30.00      Min.      :10.00
## Female:281      : 48      1st Qu.:54.75      1st Qu.:45.00
## Male :224      PARKIN: 22      Median :61.00      Median :55.00
##      PINK1 : 21      Mean :60.56      Mean :53.61
##      NMF : 12      3rd Qu.:68.25      3rd Qu.:64.00
##      LRRK2 : 11      Max. :82.00      Max. :78.00
##      (Other): 8      NA's :266      NA's :325
##      updrs      updrs_ii      updrs_iii_score_on      updrs_iii_score_off
## Min.      : 0.000      Min.      : 0.000      0      :198      0      :381
## 1st Qu.: 0.000      1st Qu.: 0.000      :152      :108
## Median : 0.000      Median : 0.000      1      : 13      1      : 11
## Mean : 1.171      Mean : 4.593      2      : 12      2      : 10
## 3rd Qu.: 2.000      3rd Qu.: 7.000      15     : 9      17     : 4
## Max. :36.000      Max. :35.000      18     : 8      19     : 3
## NA's :122      NA's :123      (Other):158      (Other): 33
##      updrs_iv      hoehn_yahr_on      hoehn_yahr_off      moca_score
## Min.      : 0.000      0      :164      0      :406      0      :191
## 1st Qu.: 0.000      :148      :101      30     : 95
## Median : 0.000      8      : 57      ND     : 8      29     : 76
## Mean : 1.236      1      : 43      1      : 8      28     : 47
## 3rd Qu.: 1.000      2      : 37      2      : 6      26     : 30
```

```
## Max.      :14.000    3      : 31    4      : 6    27      : 24
## NA's      :118      (Other): 70    (Other): 15    (Other): 87
```

We make the following observations.

1. We have some unnecessary data in this file. We aren't interested in the submission and last update date. We can reduce the dimensions of this file so it handles nicer from now on.
2. We have a LOT of missing data. You'll learn how to handle this in some of your biostats classes! For now, we'll run what analyses we can given the data we have.
3. Some of our scores have been read in as character values (and they should be numbers). If you investigate this further, you'll find that some values have been recorded as "ND", which we'll assume means "no data". We will need to record these as NA values in R.

Our next step is to address item one. We will reduce the dimensions of our pheno data frame to include only that information that we're interested in modelling. We can exclude the dates, type (as it's all RNA), tissue (all whole blood), organism (all homo sapiens), and subject ID (we will be using geo_accession as our unique indicator). As well, we will exclude mutated_pd_genes, as we intend to define our own gene signature later this week.

Subset your pheno data frame to include columns 1,8,9,11:20.

```
## Enter your own code here
```

```
library(tidyverse)
```

```
## -- Attaching packages -----
```

```
## v ggplot2 3.3.0      v readr    1.3.1
## v tibble  3.0.1      v purrr   0.3.4
## v tidyr   1.1.0      v forcats 0.5.0
```

```
## Warning: package 'tibble' was built under R version 3.6.2
```

```
## Warning: package 'tidyr' was built under R version 3.6.2
```

```
## Warning: package 'purrr' was built under R version 3.6.2
```

```
## -- Conflicts -----
```

```
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()     masks stats::lag()
```

```
## Enter your own code here FIX
```

```
pheno <- pheno %>% select(1,8,9,11:20)
pheno
```

```
##      geo_accession      disease_label      sex age_at_exam age_at_symptoms updrs
## 1      GSM2631171      ATYPICAL_PD      Male           NA           53         1
## 2      GSM2631309      ATYPICAL_PD      Male           NA           64         0
## 3      GSM2631219      ATYPICAL_PD      Male           NA           NA         0
## 4      GSM2630775              CBD      Female           NA           60         0
```

## 5	GSM2631147	CBD	Female	NA	66	0
## 6	GSM2630853	CONTROL	Male	NA	41	0
## 7	GSM2630769	CONTROL	Female	NA	43	0
## 8	GSM2631196	CONTROL	Male	40	NA	NA
## 9	GSM2631194	CONTROL	Male	42	NA	NA
## 10	GSM2631197	CONTROL	Male	43	NA	NA
## 11	GSM2631195	CONTROL	Male	49	NA	NA
## 12	GSM2631198	CONTROL	Male	40	NA	NA
## 13	GSM2631306	CONTROL		NA	NA	0
## 14	GSM2631162	CONTROL		NA	NA	0
## 15	GSM2631172	CONTROL		NA	NA	0
## 16	GSM2631241	CONTROL	Female	NA	NA	0
## 17	GSM2631252	CONTROL	Female	NA	NA	0
## 18	GSM2630927	CONTROL	Female	NA	NA	0
## 19	GSM2630928	CONTROL	Female	NA	NA	0
## 20	GSM2631227	CONTROL	Female	NA	NA	0
## 21	GSM2631231	CONTROL	Female	NA	NA	0
## 22	GSM2631235	CONTROL	Female	NA	NA	0
## 23	GSM2631236	CONTROL	Female	NA	NA	0
## 24	GSM2631238	CONTROL	Female	NA	NA	0
## 25	GSM2631239	CONTROL	Female	NA	NA	0
## 26	GSM2631243	CONTROL	Female	NA	NA	0
## 27	GSM2630771	CONTROL	Female	NA	NA	0
## 28	GSM2630783	CONTROL	Female	NA	NA	0
## 29	GSM2630830	CONTROL	Female	NA	NA	0
## 30	GSM2630857	CONTROL	Female	NA	NA	0
## 31	GSM2630868	CONTROL	Female	NA	NA	0
## 32	GSM2630818	CONTROL	Female	NA	NA	0
## 33	GSM2630907	CONTROL	Female	NA	NA	0
## 34	GSM2630909	CONTROL	Female	NA	NA	0
## 35	GSM2630916	CONTROL	Female	NA	NA	0
## 36	GSM2630923	CONTROL	Female	NA	NA	0
## 37	GSM2630925	CONTROL	Female	NA	NA	0
## 38	GSM2630929	CONTROL	Female	NA	NA	0
## 39	GSM2630930	CONTROL	Female	NA	NA	0
## 40	GSM2630932	CONTROL	Female	NA	NA	0
## 41	GSM2631221	CONTROL	Male	NA	NA	0
## 42	GSM2631230	CONTROL	Male	NA	NA	0
## 43	GSM2631232	CONTROL	Male	NA	NA	0
## 44	GSM2631234	CONTROL	Male	NA	NA	0
## 45	GSM2631237	CONTROL	Male	NA	NA	0
## 46	GSM2631240	CONTROL	Male	NA	NA	0
## 47	GSM2631242	CONTROL	Male	NA	NA	0
## 48	GSM2631248	CONTROL	Male	NA	NA	0
## 49	GSM2630899	CONTROL	Male	NA	NA	0
## 50	GSM2630905	CONTROL	Male	NA	NA	0
## 51	GSM2630906	CONTROL	Male	NA	NA	0
## 52	GSM2630917	CONTROL	Male	NA	NA	0
## 53	GSM2630922	CONTROL	Male	NA	NA	0
## 54	GSM2630924	CONTROL	Male	NA	NA	0
## 55	GSM2631298	CONTROL		NA	NA	0
## 56	GSM2631300	CONTROL		NA	NA	0
## 57	GSM2631301	CONTROL		NA	NA	0
## 58	GSM2631304	CONTROL		NA	NA	0

## 491	GSM2630762	IPD	Female	NA	NA	0
## 492	GSM2630817	IPD	Female	NA	NA	0
## 493	GSM2630826	IPD	Female	NA	NA	0
## 494	GSM2630895	IPD	Female	NA	NA	0
## 495	GSM2630915	IPD	Female	NA	NA	0
## 496	GSM2630838	IPD	Female	NA	NA	0
## 497	GSM2630900	IPD	Female	NA	NA	0
## 498	GSM2631224	IPD	Male	NA	NA	0
## 499	GSM2631244	IPD	Male	NA	NA	0
## 500	GSM2631245	IPD	Male	NA	NA	0
## 501	GSM2631246	IPD	Male	NA	NA	0
## 502	GSM2631247	IPD	Male	NA	NA	0
## 503	GSM2630794	IPD	Male	NA	NA	0
## 504	GSM2630870	IPD	Male	NA	NA	0
## 505	GSM2630890	IPD	Male	NA	NA	0
## 506	GSM2630901	IPD	Male	NA	NA	0
## 507	GSM2630902	IPD	Male	NA	NA	0
## 508	GSM2630904	IPD	Male	NA	NA	0
## 509	GSM2630764	IPD	Male	NA	NA	0
## 510	GSM2630781	IPD	Male	NA	NA	0
## 511	GSM2630791	IPD	Male	NA	NA	0
## 512	GSM2630862	IPD	Male	NA	NA	0
## 513	GSM2630898	IPD	Male	NA	NA	0
## 514	GSM2630903	IPD	Male	NA	NA	0
## 515	GSM2630931	IPD	Male	NA	NA	0
## 516	GSM2631150	IPD		NA	NA	0
## 517	GSM2631151	IPD		NA	NA	0
## 518	GSM2631155	IPD		NA	NA	0
## 519	GSM2631156	IPD		NA	NA	0
## 520	GSM2631157	IPD		NA	NA	0
## 521	GSM2631160	IPD		NA	NA	0
## 522	GSM2631169	IPD		NA	NA	0
## 523	GSM2631154	IPD		NA	NA	0
## 524	GSM2631164	IPD		NA	NA	0
## 525	GSM2631166	IPD		NA	NA	0
## 526	GSM2631168	IPD		NA	NA	0
## 527	GSM2631170	IPD		NA	NA	0
## 528	GSM2631178	IPD		NA	NA	0
## 529	GSM2631019	IPD	Female	60	NA	3
## 530	GSM2630886	IPD	Female	NA	NA	0
## 531	GSM2630765	IPD	Male	NA	NA	0
## 532	GSM2630831	MSA	Male	NA	51	0
## 533	GSM2631315	MSA	Male	66	60	0
## 534	GSM2630814	MSA	Female	NA	64	1
## 535	GSM2630776	MSA	Male	NA	68	0
## 536	GSM2631204	MSA	Female	75	71	5
## 537	GSM2630943	MSA	Male	76	72	6
## 538	GSM2631199	MSA	Female	81	74	1
## 539	GSM2631299	MSA		NA	NA	0
## 540	GSM2631223	PD_DEMENTIA	Male	NA	69	1
## 541	GSM2631233	PD_DEMENTIA	Male	NA	NA	0
## 542	GSM2631313	PSP	Male	63	52	NA
## 543	GSM2630808	PSP	Female	NA	53	0
## 544	GSM2631314	PSP	Male	76	66	36

## 545	GSM2630836	PSP	Female	NA	72	2
## 546	GSM2631216	PSP	Female	NA	72	0
## 547	GSM2630889	PSP		NA	NA	0
## 548	GSM2631174	PSP		NA	NA	0
## 549	GSM2630894	PSP	Male	NA	NA	0
## 550	GSM2630792	Vascular dementia		NA	NA	0
##	updrs_ii	updrs_iii_score_on	updrs_iii_score_off	updrs_iv	hoehn_yahr_on	
## 1	4	19	0	0	2	
## 2	0	0	0	0	9	
## 3	0	0	0	0	0	
## 4	0	0	0	0	9	
## 5	0	30	0	0	9	
## 6	0	1	0	0	8	
## 7	0	0	0	0	8	
## 8	NA		0	0	0	
## 9	NA		0	0	0	
## 10	NA		0	0	0	
## 11	NA		0	0	0	
## 12	NA		0	NA		
## 13	0	0	0	0	0	
## 14	0	0	0	0	0	
## 15	0	0	0	0	0	
## 16	0	0	0	0	0	
## 17	0	0	0	0	0	
## 18	0	0	0	0	0	
## 19	0	0	0	0	0	
## 20	0	0	0	0	0	
## 21	0	0	0	0	0	
## 22	0	0	0	0	0	
## 23	0	0	0	0	0	
## 24	0	0	0	0	0	
## 25	0	0	0	0	0	
## 26	0	0	0	0	0	
## 27	0	0	0	0	0	
## 28	0	0	0	0	0	
## 29	0	0	0	0	0	
## 30	0	0	0	0	0	
## 31	0	0	0	0	0	
## 32	0	0	0	0	0	
## 33	0	0	0	0	0	
## 34	0	0	0	0	0	
## 35	0	0	0	0	0	
## 36	0	0	0	0	0	
## 37	0	0	0	0	0	
## 38	0	0	0	0	0	
## 39	0	0	0	0	0	
## 40	0	0	0	0	0	
## 41	0	0	0	0	0	
## 42	0	0	0	0	0	
## 43	0	0	0	0	0	
## 44	0	0	0	0	0	
## 45	0	0	0	0	0	
## 46	0	0	0	0	0	
## 47	0	0	0	0	0	

## 534	16	35	0	6	3
## 535	0	46	0	0	9
## 536	28		64	1	
## 537	25	43		0	4
## 538	26		55	1	
## 539	0	0	0	0	0
## 540	7	26	0	0	3
## 541	0	2	0	0	8
## 542	NA		26	NA	
## 543	0	37	0	0	9
## 544	NA	75		0	5
## 545	15	31	0	0	3
## 546	0	0	0	0	9
## 547	0	0	0	0	0
## 548	0	0	0	0	0
## 549	0	2	0	1	8
## 550	0	0	0	0	0
##	hoehn_yahr_off	moca_score			
## 1	0	21			
## 2	0	0			
## 3	0	0			
## 4	0	0			
## 5	0	0			
## 6	0	30			
## 7	0	0			
## 8	0	0			
## 9	0	0			
## 10	0	0			
## 11	0	0			
## 12	0	30			
## 13	0	0			
## 14	0	0			
## 15	0	0			
## 16	0	0			
## 17	0	0			
## 18	0	0			
## 19	0	0			
## 20	0	0			
## 21	0	0			
## 22	0	0			
## 23	0	0			
## 24	0	0			
## 25	0	0			
## 26	0	0			
## 27	0	0			
## 28	0	0			
## 29	0	0			
## 30	0	0			
## 31	0	0			
## 32	0	0			
## 33	0	0			
## 34	0	0			
## 35	0	0			
## 36	0	0			


```
## 523      0      0
## 524      0      0
## 525      0      0
## 526      0      0
## 527      0      0
## 528      0      0
## 529      2     26
## 530      0     30
## 531      0     30
## 532      0      0
## 533      4
## 534      0     23
## 535      0      0
## 536      5
## 537             24
## 538      5     27
## 539      0      0
## 540      0     20
## 541      0     29
## 542      2.5
## 543      0      0
## 544
## 545      0      0
## 546      0      0
## 547      0      0
## 548      0      0
## 549      0     29
## 550      0      0
```

Next we need to correct the columns which contain “ND”. You can use the “which” function to find the index of of the matrices which are “ND”, and then set these to NA. Set columns 8,9,11,12,13 to numeric values using the “as.numeric” function inside a “sapply” loop. Run a summary of the data frame again.

```
index<-which(pheno == " ND",arr.ind = T)
pheno[index]<-NA
j<-c(8,9,11,12,13)
pheno[,j]<-sapply(unlist(pheno[,j]),as.numeric)
summary(pheno)
```

```
##      geo_accession      disease_label      sex      age_at_exam
## GSM2630758: 1      CONTROL      :233      : 45      Min.      :30.00
## GSM2630759: 1      IPD      :205      Female:281      1st Qu.:54.75
## GSM2630760: 1      GPD      : 41      Male  :224      Median :61.00
## GSM2630761: 1      GENETIC_UNAFFECTED: 22      Mean   :60.56
## GSM2630762: 1      HD      : 19      3rd Qu.:68.25
## GSM2630763: 1      MSA      : 8      Max.   :82.00
## (Other)      :544      (Other)      : 22      NA's    :266
## age_at_symptoms      updrs      updrs_ii      updrs_iii_score_on
## Min.      :10.00      Min.      : 0.000      Min.      : 0.000      Min.      : 1.000
## 1st Qu.:45.00      1st Qu.: 0.000      1st Qu.: 0.000      1st Qu.: 1.000
## Median :55.00      Median : 0.000      Median : 0.000      Median : 3.000
## Mean   :53.61      Mean   : 1.171      Mean   : 4.593      Mean   : 9.593
## 3rd Qu.:64.00      3rd Qu.: 2.000      3rd Qu.: 7.000      3rd Qu.:12.250
```

```
## Max. :78.00 Max. :36.000 Max. :35.000 Max. :53.000
## NA's :325 NA's :122 NA's :123 NA's :2
## updrs_iii_score_off updrs_iv hoehn_yahr_on hoehn_yahr_off
## Min. : 1.000 Min. : 0.000 Min. : 1.00 Min. : 1.00
## 1st Qu.: 3.000 1st Qu.: 0.000 1st Qu.: 1.00 1st Qu.: 3.00
## Median : 3.000 Median : 0.000 Median : 3.00 Median : 3.00
## Mean : 4.829 Mean : 1.236 Mean :15.63 Mean : 4.45
## 3rd Qu.: 3.000 3rd Qu.: 1.000 3rd Qu.:26.00 3rd Qu.: 3.00
## Max. :58.000 Max. :14.000 Max. :60.00 Max. :60.00
## NA's :118 NA's :10 NA's :8
## moca_score
## Min. : 1.00
## 1st Qu.: 3.00
## Median :22.00
## Mean :16.09
## 3rd Qu.:25.00
## Max. :27.00
## NA's :9
```

Let's look at a summary of the first 10 columns of expression data set.

```
## Enter your own code here
```

```
summary(expr[1:20668,1:10, drop=FALSE])
```

```
##      X      GeneName      GSM2631171      GSM2631309
## Min. : 1 A1BG : 1 Min. :-5.223788 Min. :-6.09018
## 1st Qu.: 5168 A1BG-AS1: 1 1st Qu.: -0.960423 1st Qu.: -0.92906
## Median :10334 A1CF : 1 Median :-0.004842 Median : 0.01385
## Mean :10334 A2M : 1 Mean :-0.009648 Mean : 0.01249
## 3rd Qu.:15501 A2M-AS1 : 1 3rd Qu.: 0.953228 3rd Qu.: 0.95912
## Max. :20668 (Other) :20662 Max. : 5.766301 Max. : 5.66627
## NA's : 1
## GSM2631219 GSM2630775 GSM2631147 GSM2630853
## Min. :-6.39097 Min. :-5.206869 Min. :-5.27578 Min. :-6.115736
## 1st Qu.: -0.97337 1st Qu.: -0.981831 1st Qu.: -0.96379 1st Qu.: -0.944666
## Median :-0.01097 Median : 0.001772 Median : 0.01906 Median :-0.007942
## Mean :-0.00354 Mean :-0.000010 Mean : 0.00298 Mean :-0.009892
## 3rd Qu.: 0.95324 3rd Qu.: 0.971013 3rd Qu.: 0.98545 3rd Qu.: 0.945826
## Max. : 6.56118 Max. : 5.275719 Max. : 5.18612 Max. : 5.570111
##
## GSM2630769 GSM2631196
## Min. :-5.608142 Min. :-6.303044
## 1st Qu.: -0.968002 1st Qu.: -0.970730
## Median :-0.001583 Median :-0.004689
## Mean : 0.014813 Mean :-0.006484
## 3rd Qu.: 0.987677 3rd Qu.: 0.977216
## Max. : 5.591597 Max. : 5.434250
##
```

We don't need the X1 variable - this is just remaining row labels in the csv file. Let's remove this variable.

```
## Enter your own code here
```

```
expr <- expr %>%  
  select(-X)
```

We don't see any evidence of missing values in our summary, but we should check all of the columns (excluding the ProbeID and GeneName). You can check this with the "anyNA" function.

```
## Enter your own code here
```

```
expr_na_CheckPrep = subset(expr, select = -c(GeneName)) # temp dropping these for N/A check  
anyNA(expr_na_CheckPrep, recursive = FALSE)
```

```
## [1] FALSE
```

```
#anyNA(expr)
```

Let's identify how big this problem is, and where it occurs.

```
which(is.na(expr), arr.ind = T)
```

```
##           row col  
## [1,] 20668    1
```

So one of our gene names is NA! This isn't useful, so let's remove this row.

```
## Enter your own code here
```

```
expr <- expr[-20668,]
```

We should see if the unique identifiers in our two data sets match. Check for a perfect match using the "identical" function.

```
identical(colnames(expr[, -1]), as.character(pheno[, 1]))
```

```
## [1] TRUE
```

Question: why is the '-1' necessary here? Answer below!

The -1 is necessary, because exclude GeneNames

So that we don't lose any work, let's clean up our workspace to include only our cleaned expression and pheno data sets, which we can reload later.

Exploratory Data Analysis

In this section we are going to explore some of the data we have, and maybe develop a diagnostic signature for Parkinson's disease.

First, load in your data from yesterday.

Let's re-examine our pheno data set with the summary function again.

```
## Enter your own code here
```

```
summary(pheno)
```

```
##      geo_accession      disease_label      sex      age_at_exam
## GSM2630758: 1      CONTROL      :233      : 45      Min.      :30.00
## GSM2630759: 1      IPD      :205      Female:281      1st Qu.:54.75
## GSM2630760: 1      GPD      : 41      Male :224      Median :61.00
## GSM2630761: 1      GENETIC_UNAFFECTED: 22      Mean :60.56
## GSM2630762: 1      HD      : 19      3rd Qu.:68.25
## GSM2630763: 1      MSA      : 8      Max. :82.00
## (Other) :544      (Other) : 22      NA's :266
## age_at_symptoms      updrs      updrs_ii      updrs_iii_score_on
## Min. :10.00      Min. : 0.000      Min. : 0.000      Min. : 1.000
## 1st Qu.:45.00      1st Qu.: 0.000      1st Qu.: 0.000      1st Qu.: 1.000
## Median :55.00      Median : 0.000      Median : 0.000      Median : 3.000
## Mean :53.61      Mean : 1.171      Mean : 4.593      Mean : 9.593
## 3rd Qu.:64.00      3rd Qu.: 2.000      3rd Qu.: 7.000      3rd Qu.:12.250
## Max. :78.00      Max. :36.000      Max. :35.000      Max. :53.000
## NA's :325      NA's :122      NA's :123      NA's :2
## updrs_iii_score_off      updrs_iv      hoehn_yahr_on      hoehn_yahr_off
## Min. : 1.000      Min. : 0.000      Min. : 1.00      Min. : 1.00
## 1st Qu.: 3.000      1st Qu.: 0.000      1st Qu.: 1.00      1st Qu.: 3.00
## Median : 3.000      Median : 0.000      Median : 3.00      Median : 3.00
## Mean : 4.829      Mean : 1.236      Mean :15.63      Mean : 4.45
## 3rd Qu.: 3.000      3rd Qu.: 1.000      3rd Qu.:26.00      3rd Qu.: 3.00
## Max. :58.000      Max. :14.000      Max. :60.00      Max. :60.00
## NA's :118      NA's :10      NA's :8
## moca_score
## Min. : 1.00
## 1st Qu.: 3.00
## Median :22.00
## Mean :16.09
## 3rd Qu.:25.00
## Max. :27.00
## NA's :9
```

We need to further delve into our disease label in order to simplify some of this analysis. Attach your pheno data frame using the attach function, and then summarize the disease label vector.

```
## Enter your own code here
```

```
attach(pheno)
```

```
summary(disease_label)
```

```
##      ATYPICAL_PD      CBD      CONTROL      DRD
##      3      2      233      3
##      DRD-DYT5      GENETIC_UNAFFECTED      GPD      HD
##      3      22      41      19
##      IPD      MSA      PD_DEMENTIA      PSP
##      205      8      2      8
```

```
## Vascular dementia
## 1
```

Here we have the counts of all the diseases in our data set. If you look at the actual excel file (not the csv), I've put in a dictionary for these acronyms if you're curious. Here, our controls and our genetic unaffected are both considered to be healthy controls. Any label which contains PD is some subset of Parkinson's Disease, and the other labels represent other neurological disorders. We need to make a variable which records a 1 for our cases, and a 0 for our controls. Here, since we are interested in a signature that distinguishes PD from our other disease, the other diseases are technically part of the control set.

Try to set your case control vector using the grep function to find the indices which contain "PD". At the end, sum your case vector to check that it worked. Make another variable of the words "case" and "control"

```
## Enter your own code here
pheno <- pheno %>%
  mutate(case = if_else(str_detect(disease_label, "PD"), 1, 0))
#Case Set
case <- pheno %>%
  mutate(case = if_else(str_detect(disease_label, "PD"), 1, 0)) %>%
  subset(case == 1)
#Control
control <- pheno %>%
  mutate(case = if_else(str_detect(disease_label, "PD"), 1, 0)) %>%
  subset(case == 0)
```

We need to find differentially expressed genes. You'll learn more about this later. For now, feel free to use some of my code. Start by downloading the limma package

```
## If using Windows, first go to https://cran.rstudio.com/bin/windows/Rtools/ and install the appropriate Rtools
if (!requireNamespace("BiocManager", quietly = TRUE))
  install.packages("BiocManager")

#uncomment to load limma for first run through

BiocManager::install("limma")
library(limma)
```

We will use the following code. Please add comments to every line to tell me what its doing!

```
## Subset our data for a training and test set
set.seed(2) #two random numbers generated for simulation
prob<-runif(ncol(expr)-2) #generates random deviates of the uniform distribution for expr
k<-which(prob>=0.3333333) #stores indexes that have a prob > 1/3
#Subsets out the GeneName column
eset<-expr[,2:ncol(expr)] # Explanation Below
eset<-eset[,k] #Stores the rows that meet the threshold of prob > 1/3
rownames(eset)<-expr[,1] #take the row names from expr dataframe and put them in eset dataframe
design <- model.matrix(~0+as.factor(pheno$case[k])) #creating a design matrix and getting the independent variables
fit <- eBayes(lmFit(eset,design)) #fitting the model with the parkinsons labels
topTable(fit, coef=2) # 2 coef is the optimal coef
```

##	logFC	AveExpr	t	P.Value	adj.P.Val	B
## EX0C3L4	3.142306	1.471867	29.51102	4.199361e-149	4.759985e-145	329.8083

```
## FAM132A      -3.159535 -1.461491 -29.50594 4.606362e-149 4.759985e-145 329.7162
## MDM2         3.127701  1.426391  29.45147 1.239815e-148 8.541085e-145 328.7308
## CCR3        -3.145072 -1.547047 -29.41096 2.588618e-148 1.337474e-144 327.9981
## MYO9A       -3.172079 -1.571136 -29.27526 3.042541e-147 1.257604e-143 325.5456
## GADD45GIP1  -3.086818 -1.391784 -29.09131 8.553065e-146 2.946103e-142 322.2251
## ANXA2       -3.106348 -1.517455 -29.04409 2.011963e-145 5.940178e-142 321.3737
## CCNJ        -3.123721 -1.454289 -28.99841 4.602051e-145 1.188882e-141 320.5503
## EMC6        3.070413  1.590585  28.97850 6.599845e-145 1.515544e-141 320.1914
## GEMIN4      3.100428  1.427101  28.89106 3.211859e-144 6.637949e-141 318.6165
```

```
results<-topTable(fit, coef=2, number=Inf) # showing the inferential stats
```

Here, we have our gene names, our log fold change for expression, average expression, t statistic, pvalue, adjusted pvalue (for multiple testing!!), and the log odds of differential expression.

Next, we select those genes that have adjusted p-values below 0.001. Again, add comments to every line to describe what the code is doing.

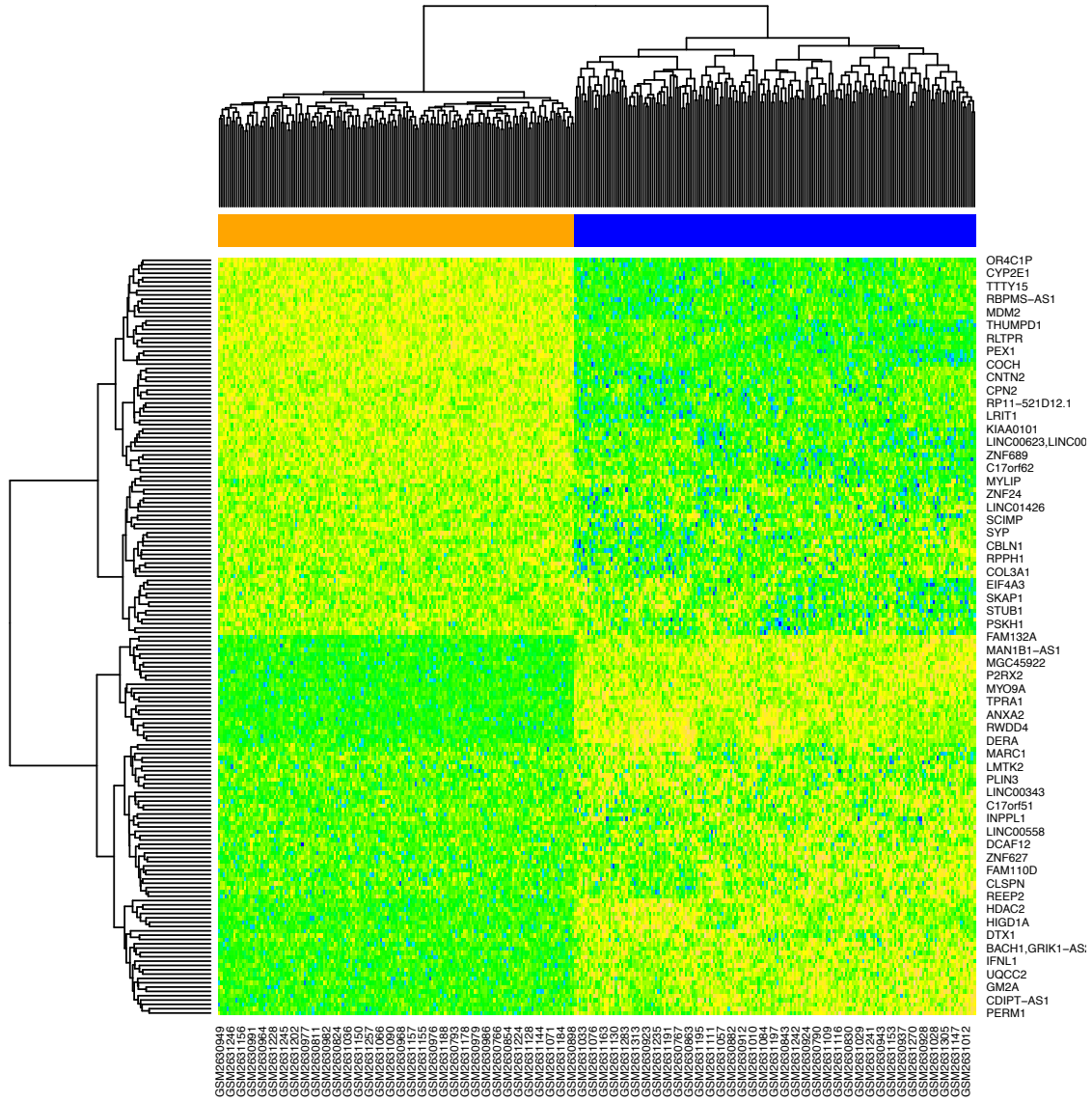
```
selected <- row.names(results)[p.adjust(results$P.Value, method="fdr")<0.001] # gets the characters that
direction <- sign(results$logFC) ## generates vector of numbers based on if the logFC has positive or negative
esetSel <- eset[selected, ] #storing the occurrences of <.001 into esetSel
nrow(esetSel) # how many occurrences of <.001
```

```
## [1] 175
```

Okay! So we're now looking at just 175 probes!

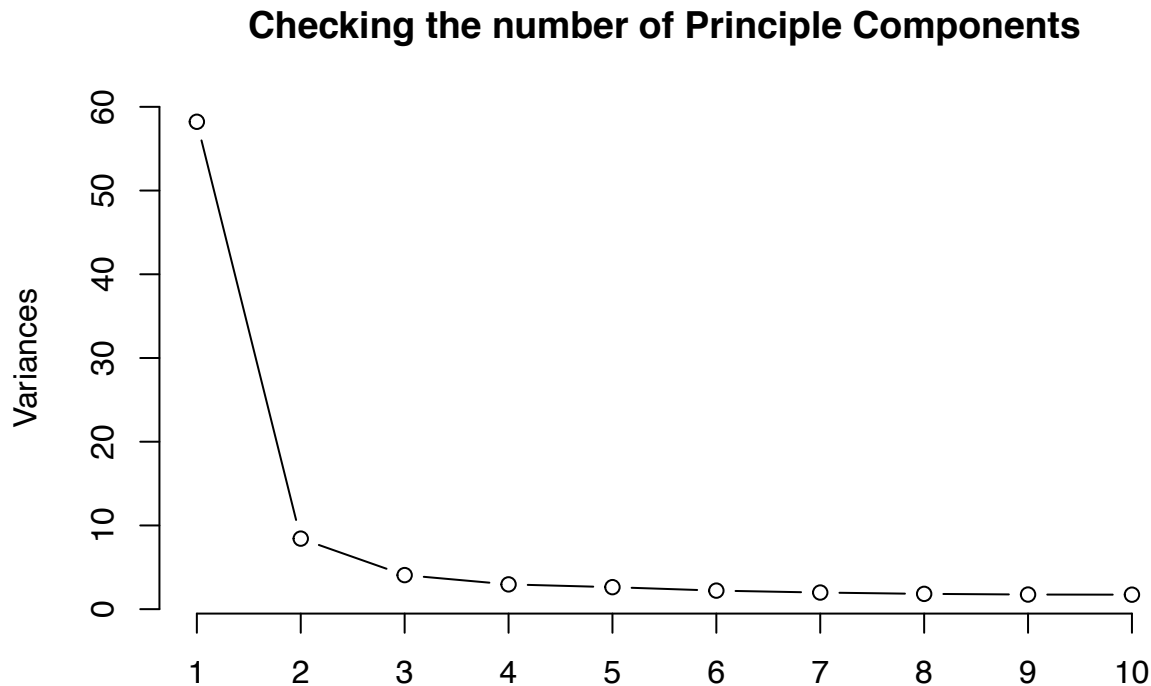
We are going to make a heat map here. I've provided the code, but try changing colours, labels, etc. to make it your own. You can try typing '?heatmap' into the console to see the help page and provide more ideas for what you'd like to change!

```
patientcolors <-ifelse(pheno$case[k]==1,"orange","blue")
heatmap(as.matrix(esetSel), col=topo.colors(100), ColSideColors=patientcolors, distfun = function(x) dist(x))
```



Notice the annotation bar along the top. This indicates PD vs not PD samples. This heat map is an example of a ‘non-supervised method’ - where we didn’t feed the labelled data to the algorithm. Instead, it is just clustering similar samples together. Because all of our PD samples cluster away from the non-PD samples, we are relatively certain we’ve picked good biomarkers! We should also check a PCA plot.

```
pc<-prcomp(t(esetSel),center=T,scale=T)
plot(pc,type="l",main="Checking the number of Principle Components")
```



Again, I’ve provided code for you here. Change it to something you like better!

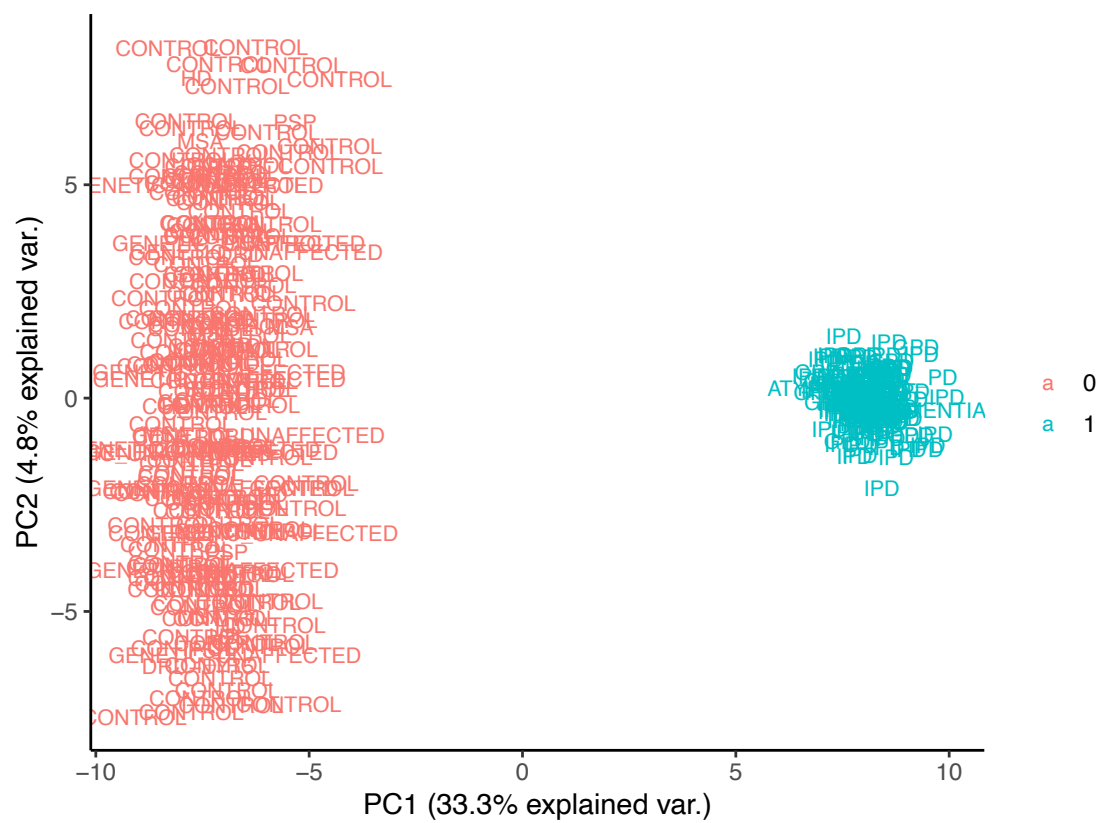
```
#install.packages("devtools")
library(devtools)

#install.packages("ggpubr")
library(ggpubr)

#install_github("vqv/ggbiplot")
#library(ggbiplot)
source("ggbiplot.R")
g <- ggbiplot(pc, obs.scale = 1, var.scale = 1,
              groups = as.factor(pheno$case[k]), ellipse = F,
              circle = F, labels=pheno$disease_label[k], var.axes = F)
g <- g + scale_color_discrete(name = '')
g <- g + theme(legend.direction = 'horizontal',
              legend.position = 'top')
```



```
g <- g + theme_classic2()
print(g)
```



We have separation! Notice the obvious differences between cases and controls.

Make a variable which only contains the differential gene names and call it diffGenes AND print out all of these gene names using one line of code. The parentheses around the full line of code do this!

```
(diffGenes<-selected)
```

```
## [1] "EXOC3L4"
## [2] "FAM132A"
## [3] "MDM2"
## [4] "CCR3"
## [5] "MYO9A"
## [6] "GADD45GIP1"
## [7] "ANXA2"
## [8] "CCNJ"
## [9] "EMC6"
## [10] "GEMIN4"
## [11] "PPM1K"
## [12] "TTL"
## [13] "DEFB103A,DEFB103B"
## [14] "STX3"
## [15] "DERA"
## [16] "HIST1H2B0"
## [17] "RGS4"
## [18] "MAN1B1-AS1"
## [19] "PSG5"
## [20] "OR4C1P"
## [21] "ABHD12B,MIR4454"
## [22] "COCH"
## [23] "RWDD4"
## [24] "FAR1"
## [25] "PEX1"
## [26] "THUMPD1"
## [27] "CTB-31020.9"
## [28] "GCLC"
## [29] "SEC16B"
## [30] "CYP2E1"
## [31] "EGLN1"
## [32] "PRKAG2"
## [33] "NPC2"
## [34] "TPRA1"
## [35] "SLC26A4"
## [36] "XAB2"
## [37] "C10orf88"
## [38] "MGC45922"
## [39] "P2RX2"
## [40] "AARS"
## [41] "RNF157"
## [42] "PSMC2"
## [43] "RBPMS-AS1"
## [44] "PCDHGB8P"
## [45] "CD207"
## [46] "RLTPR"
## [47] "TTY15"
```

[48] "TRAF5"
 ## [49] "RP11-245J9.5"
 ## [50] "KCNA10"
 ## [51] "UQCC2"
 ## [52] "RP11-324J3.1"
 ## [53] "GABRA1"
 ## [54] "CPN2"
 ## [55] "MAP6"
 ## [56] "C17orf62"
 ## [57] "TRAFFD1"
 ## [58] "HIPK4"
 ## [59] "GM2A"
 ## [60] "N6AMT1"
 ## [61] "RABGAP1L"
 ## [62] "ANP32A"
 ## [63] "ROBO2"
 ## [64] "TOPORS-AS1"
 ## [65] "STEAP2"
 ## [66] "RNF167"
 ## [67] "HDAC2"
 ## [68] "ETFB"
 ## [69] "RP11-521D12.1"
 ## [70] "PHKA1"
 ## [71] "TNS4"
 ## [72] "EIF4A2"
 ## [73] "ZNF689"
 ## [74] "BACH1,GRIK1-AS2"
 ## [75] "LRIT1"
 ## [76] "KBTBD8"
 ## [77] "B3GAT2"
 ## [78] "DNM2"
 ## [79] "DDIAS"
 ## [80] "C2CD3"
 ## [81] "CNTN2"
 ## [82] "AP1S3"
 ## [83] "CDIPT-AS1"
 ## [84] "HIGD1A"
 ## [85] "KIAA0101"
 ## [86] "PERM1"
 ## [87] "IFNL1"
 ## [88] "CYP4Z1"
 ## [89] "R3HDM4"
 ## [90] "HMGCLL1"
 ## [91] "RBM41"
 ## [92] "RP11-108P20.4"
 ## [93] "ARL6"
 ## [94] "LINC00623,LINC00869,LINC01138,LOC103091866"
 ## [95] "LINC00865"
 ## [96] "ASMTL-AS1"
 ## [97] "CASP14"
 ## [98] "OR5J2"
 ## [99] "DDX60L"
 ## [100] "ZDHHC24"
 ## [101] "MUC20"

```

## [102] "SYNP0"
## [103] "LAIR2"
## [104] "UCP3"
## [105] "REEP2"
## [106] "HDAC10"
## [107] "CBLN1"
## [108] "AP2M1"
## [109] "FOXN3-AS2"
## [110] "SYP"
## [111] "PPP6R2"
## [112] "CDH26"
## [113] "RPPH1"
## [114] "NT5DC3"
## [115] "ZNF627"
## [116] "STUB1"
## [117] "DTX1"
## [118] "CCDC136"
## [119] "FAM169A"
## [120] "LINC00558"
## [121] "CLCA2"
## [122] "GINM1"
## [123] "GHRHR"
## [124] "PKD2L2"
## [125] "RP11-742B18.1"
## [126] "LPGAT1"
## [127] "EIF4A3"
## [128] "CTD-2033C11.1"
## [129] "LEF1"
## [130] "LMTK2"
## [131] "A1BG"
## [132] "LINC00343"
## [133] "FAM110D"
## [134] "ADORA3"
## [135] "DKC1,MIR664B,SNORA56"
## [136] "BOP1,MIR7112"
## [137] "SCIMP"
## [138] "MAB21L1,MIR548F5"
## [139] "ZNF883"
## [140] "ZC3H14"
## [141] "PADI4"
## [142] "CLSPN"
## [143] "ZNF24"
## [144] "PLIN3"
## [145] "AURKC"
## [146] "RP11-320N7.2"
## [147] "FAM99B"
## [148] "LPCAT4"
## [149] "MPV17L"
## [150] "CD22"
## [151] "NEK11"
## [152] "MARC1"
## [153] "NR3C1"
## [154] "USO1"
## [155] "GJD4"

```

```
## [156] "RP11-21L23.2"
## [157] "LINC01426"
## [158] "STAT1"
## [159] "IGLC1, IGLV3-10, IGLV3-10"
## [160] "MRPL15"
## [161] "INPPL1"
## [162] "C17orf51"
## [163] "DCAF12"
## [164] "LINC00337"
## [165] "CYFIP2"
## [166] "LINC00927"
## [167] "ALK"
## [168] "SSX2, SSX2B, SSX3"
## [169] "ROCK2"
## [170] "MAGEC3"
## [171] "PSKH1"
## [172] "SKAP1"
## [173] "COL3A1"
## [174] "MYLIP"
## [175] "RP11-613M5.1"
```

To use these genes as a classifier, we will need to define a score function. Our score will be the sum of the average expression for the upregulated (positive) genes and the average for the down regulated (negative) genes. Here, I've written you a function which will do this. Please enter it and make comments to show you understand what its doing.

```
PDscore<-function(x,g,v,s){
  #x expression values for a sample
  #g all the genes
  #v the diffGenes
  #s is the sign of the logFC

  i<-which(g%in%v) #Subset for diffGenes within the entire list of all the genes
  x<-x[i] # stores the value at the ith index into x (for expression values for a sample)
  s<-s[i] # stores the value at the ith index into s (for sign of LogFC)
  #Create vectors for genes with positive and negative momentum
  p<-c()
  n<-c()
  for(i in 1:length(x)){ # loop through the entire expression values
    if(s[i]>0){ #if the LogFC sign is positive than append it to the list p for positive
      p<-append(p,(x[i]))
    }
    else if(s[i]<0){ ## if the LogFC sign is negative than append it to the list n for negative
      n<-append(n,(x[i]))
    }
  }

  #If neither positive nor negative set to 0
  if(is.null(p)){p[1]=0}
  if(is.null(n)){n[1]=0}

  # the "score" is the differential of the mean of positive and negative
  score<-mean(p)-mean(n)
  return(score)
```

```
}
```

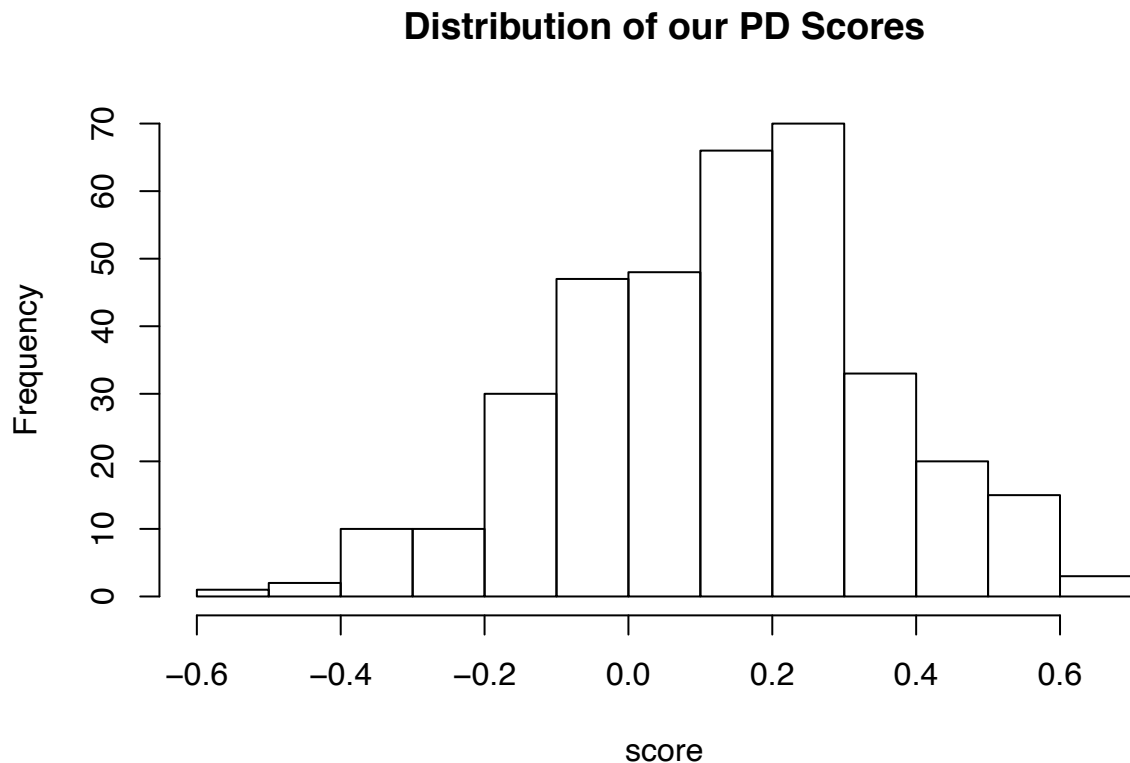
Now we can apply our function to our expression set to define a score for each patient. Comment what this is doing and why each step is necessary!

```
#Create vector
score<-c()

# create a vector of all genes in expr. Necessary for future steps to use gene stats to predict case ou
allGenes<-as.character(expr[as.character(expr$GeneName)%in%rownames(results),1])

#Apply our function to calculate scores
for(i in 1:ncol(eset)){
  score[i]<-PDscore(eset[,i],allGenes,diffGenes,direction)
}

#Generates histogram
hist(score,main="Distribution of our PD Scores")
```



Now we'll use ggplot to make and interpret a violin plot of our score. I've provided some code to do this, but try to change labels, colours, etc. to make it your own.

```
df<-data.frame(cbind(pheno$case[k],score))
```

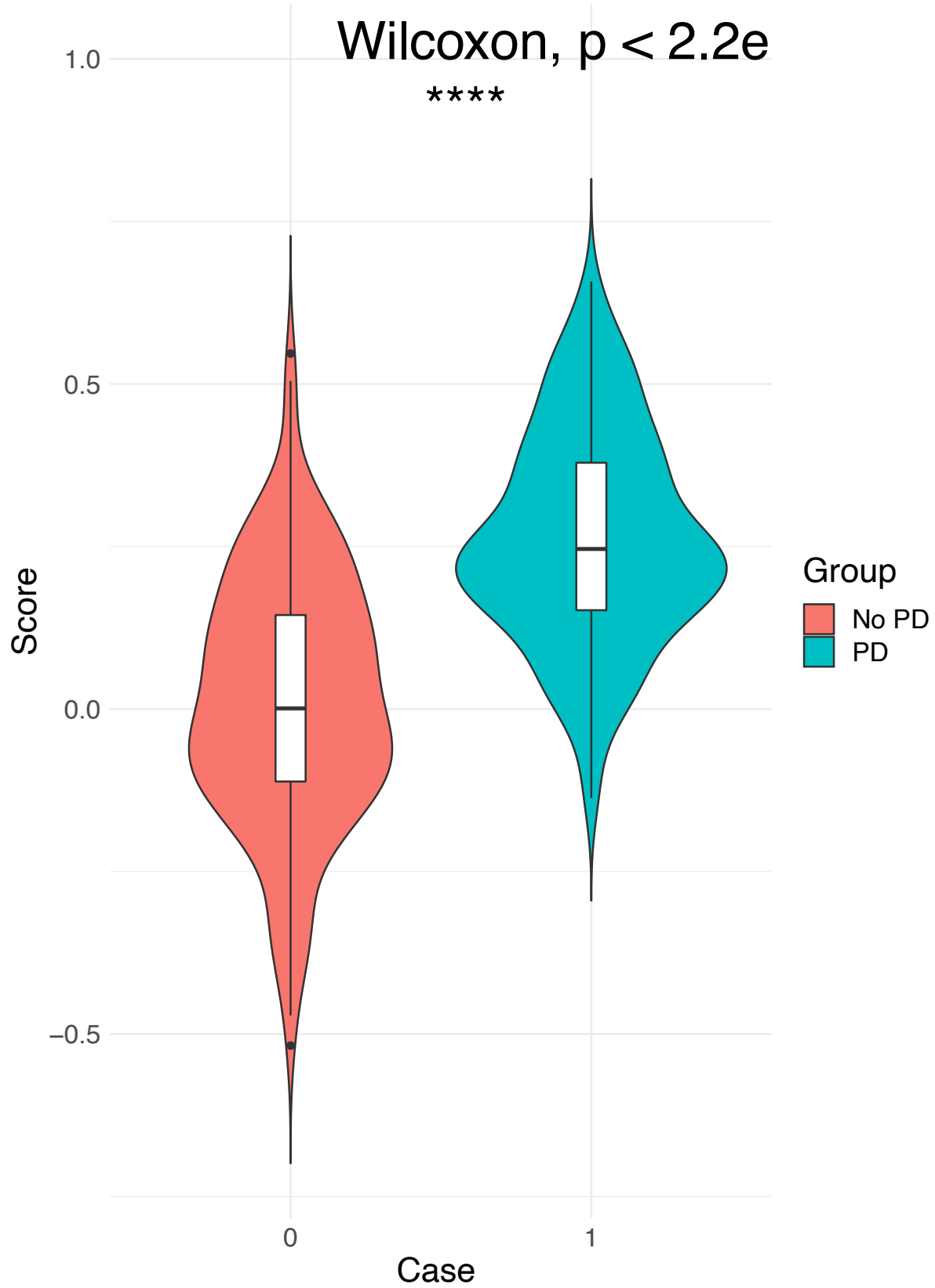
```

dp <- ggplot(df, aes(x=as.factor(pheno$case[k]), y=score, fill=as.factor(pheno$case[k]))) +
  geom_violin(trim=FALSE)+
  geom_boxplot(width=0.1, fill="white")+
  labs(title="Plot of case by score",x="Case ", y = "Score")+
  stat_compare_means(label.x = 1.5, label.y = 1, size=10)+
  stat_compare_means(aes(label = ..p.signif..),
                     label.x = 1.5, label.y = 0.9, size =10) + theme_minimal() +
  scale_fill_discrete(name = "Group", labels = c("No PD", "PD")) +
  theme(text = element_text(size = 18))

```

dp

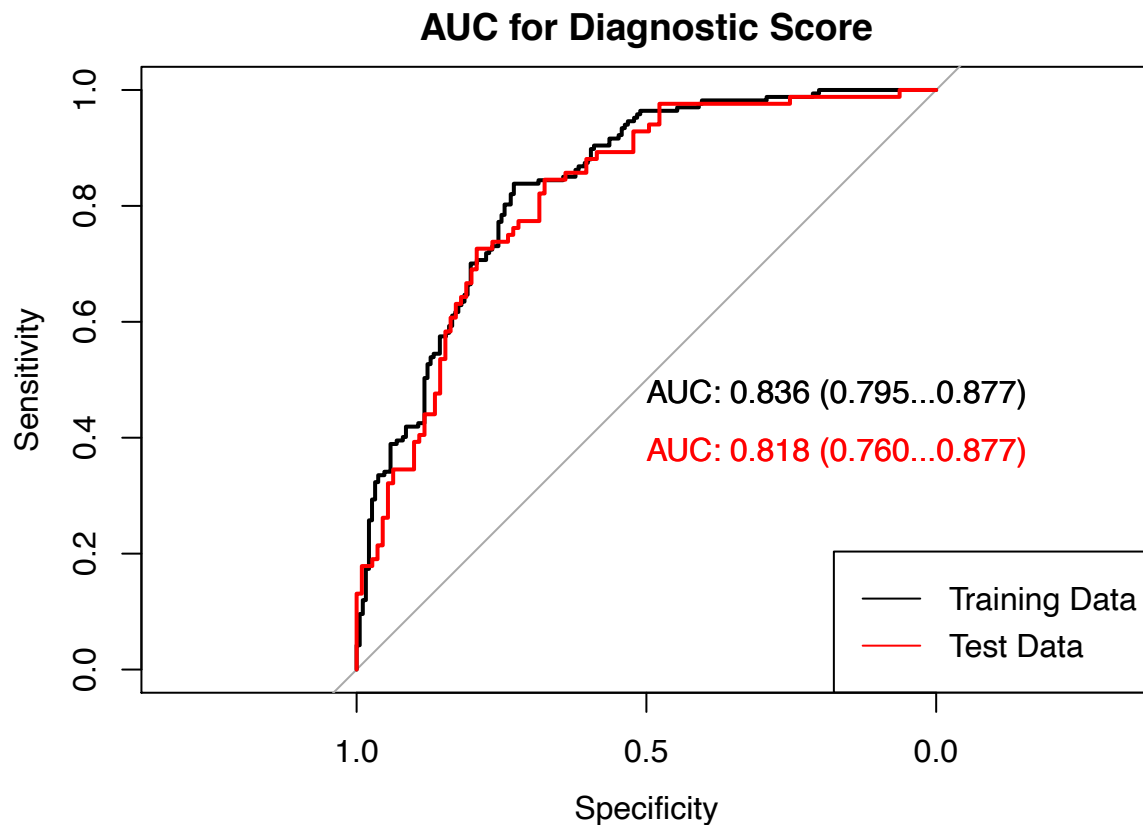
Plot of case by score



This shows not only the boxplot of our data, but also the distribution of our data points around the boxplot! As before, we can see that we don't have significant separation for our score, although we can see that the cases are trending to have a higher score. With more time and data cleaning we may be able to find something here!

Let's make an ROC plot, first with our training data, and then with our test data. As before, play with the plot options to make something you like! Note, there are MANY packages to build ROC plots, this one is just simple. Feel free to play with other packages to make publication ready plots if you'd like!

```
#install.packages("verification")
#install.packages("pROC")
library("pROC")
testEset<-expr[,2:ncol(expr)]
testEset<-testEset[,~k]
newScore<-apply(testEset,2,FUN=PDscore,allGenes,diffGenes,direction)
plot.roc(pheno$case[k]~score, data=df,legacy.axes=F,print.auc=T, ci=T, main="AUC for Diagnostic Score")
plot.roc(pheno$case[~k]~newScore,data=data.frame(cbind(pheno$case[~k],newScore)),add=T,print.auc=T, ci=T,
legend("bottomright",c("Training Data","Test Data"),lty=c(1,1),col=c("black","red"))
```



Notice that our score does better with our training data - this is expected! This is why we need to split our data, to avoid problems with over-fitting. These scores are better than random (the grey line), but we'd like to see an AUC as close to 1 as possible. Let's See if we can do better!

Statistics!

We can run a t-test to see if our score is significantly different between cases and controls. Try using the `t.test` function in R.

```
allScore<-c(score,newScore)
mergeCase<-c(pheno$case[k],pheno$case[-k]) ## to preserve order

## Do the t.test here

t.test(allScore ~ mergeCase)

##
## Welch Two Sample t-test
##
## data: allScore by mergeCase
## t = -15.842, df = 548, p-value < 2.2e-16
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -0.2743704 -0.2138353
## sample estimates:
## mean in group 0 mean in group 1
## 0.005519575 0.249622456
```

The mean scores for our cases and controls are close, but they are significantly different with an extremely small p-value of 2.787e-13. This highlights a classical statistical fallacy - while small p-values are great, they are often meaningless without a large enough effect size. Here, we have achieved significance due to the large sample size of our study, hence our study is adequately powered.

We could also run a simple regression to examine the impact of the score on the log odds of being a case.

```
smallModel<-glm(pheno$case[k]~score, family=binomial)
summary(smallModel)

##
## Call:
## glm(formula = pheno$case[k] ~ score, family = binomial)
##
## Deviance Residuals:
##      Min       1Q   Median       3Q      Max
## -2.5109  -0.7565  -0.2132   0.8773   2.1995
##
## Coefficients:
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)  -1.2373     0.1862  -6.646 3.02e-11 ***
## score         7.9472     0.8874   8.955 < 2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for binomial family taken to be 1)
##
##      Null deviance: 490.89  on 354  degrees of freedom
## Residual deviance: 350.94  on 353  degrees of freedom
```

```
## AIC: 354.94
##
## Number of Fisher Scoring iterations: 5
```

Summarize this output!

Again, we conclude that the score is a statistically significant indicator of the odds of having PD. Let's build a larger model which examines other phenotype variables.

First, build a data frame which includes all the model data we're interested in. Start with the age variables in your pheno set, and then use the `cbind()` function to add on our scores and the binary case vector. Print a summary of the model data.

Enter your own code here

```
modelData <- data.frame(cbind(age_at_exam, age_at_symptoms, allScore, mergeCase ))
modelData
```

##	age_at_exam	age_at_symptoms	allScore	mergeCase
## X	NA	53	-0.1347098260	1
## X.1	NA	64	0.1521432297	1
## X.2	NA	NA	0.1034828276	0
## X.3	NA	60	-0.0596159956	0
## X.4	NA	66	0.2008197232	0
## X.5	NA	41	0.1159874423	0
## X.6	NA	43	-0.1716217278	0
## X.7	40	NA	0.3145752497	0
## X.8	42	NA	-0.0783260787	0
## X.9	43	NA	0.3160259096	0
## X.10	49	NA	0.2077043555	0
## X.11	40	NA	0.1054308479	0
## X.12	NA	NA	0.0388705862	0
## X.13	NA	NA	-0.1137639583	0
## X.14	NA	NA	-0.3133639163	0
## X.15	NA	NA	-0.3253562144	0
## X.16	NA	NA	0.2621020054	0
## X.17	NA	NA	-0.0677631764	0
## X.18	NA	NA	-0.1049747583	0
## X.19	NA	NA	0.3126505620	0
## X.20	NA	NA	0.1711543562	0
## X.21	NA	NA	0.1299140878	0
## X.22	NA	NA	-0.1625402453	0
## X.23	NA	NA	-0.0001492302	0
## X.24	NA	NA	0.1440582062	0
## X.25	NA	NA	0.2269270956	0
## X.26	NA	NA	0.2834992804	0
## X.27	NA	NA	-0.0734041713	0
## X.28	NA	NA	0.2290789771	0
## X.29	NA	NA	-0.2087020865	0
## X.30	NA	NA	-0.3854779012	0
## X.31	NA	NA	-0.0848262372	0
## X.32	NA	NA	-0.0960736661	0
## X.33	NA	NA	0.2615004252	0
## X.34	NA	NA	0.1353291115	0
## X.35	NA	NA	-0.2326583465	0

## X.36	NA	NA -0.2099056367	0
## X.37	NA	NA 0.2205223646	0
## X.38	NA	NA -0.0605756069	0
## X.39	NA	NA -0.0207499227	0
## X.40	NA	NA -0.0142608639	0
## X.41	NA	NA -0.0021409175	0
## X.42	NA	NA 0.2607879538	0
## X.43	NA	NA 0.0834096624	0
## X.44	NA	NA 0.0686908765	0
## X.45	NA	NA 0.1759604057	0
## X.46	NA	NA -0.4334295525	0
## X.47	NA	NA -0.0825491905	0
## X.48	NA	NA -0.0973115985	0
## X.49	NA	NA 0.2189929066	0
## X.50	NA	NA 0.5046861931	0
## X.51	NA	NA -0.2715498546	0
## X.52	NA	NA -0.1108084334	0
## X.53	NA	NA -0.0175931857	0
## X.54	NA	NA 0.1213403250	0
## X.55	NA	NA 0.1760353223	0
## X.56	NA	NA -0.1352559821	0
## X.57	NA	NA -0.0863252329	0
## X.58	NA	NA 0.2269926177	0
## X.59	NA	NA -0.0904286923	0
## X.60	NA	NA 0.0073166342	0
## X.61	NA	NA 0.1191124525	0
## X.62	NA	NA 0.1460380005	0
## X.63	NA	NA -0.2058909418	0
## X.64	NA	NA 0.2144559649	0
## X.65	NA	NA 0.1374674165	0
## X.66	NA	NA -0.1305490226	0
## X.67	NA	NA -0.0502469710	0
## X.68	NA	NA 0.4213617050	0
## X.69	NA	NA -0.0160066746	0
## X.70	NA	NA 0.0731164951	0
## X.71	NA	NA -0.1353828390	0
## X.72	NA	NA -0.1952449189	0
## X.73	NA	NA 0.0020716362	0
## X.74	NA	NA 0.1601396132	0
## X.75	NA	NA -0.1962461422	0
## X.76	NA	NA 0.1259466914	0
## X.77	NA	NA -0.0337047700	0
## X.78	NA	NA 0.0475631840	0
## X.79	68	NA -0.0691503670	0
## X.80	82	NA 0.1639790493	0
## X.81	63	NA -0.3291049779	0
## X.82	70	NA 0.0235955686	0
## X.83	49	NA -0.0664110238	0
## X.84	58	NA -0.3697818244	0
## X.85	65	NA -0.3608025295	0
## X.86	78	NA -0.3191115091	0
## X.87	42	NA -0.3108080449	0
## X.88	67	NA 0.0961788529	0
## X.89	69	NA -0.0021726334	0

## GSM2630927	49	46 -0.0522823732	0
## GSM2631227	NA	46 -0.1467890637	0
## GSM2631238	NA	46 0.3225404641	0
## GSM2630771	50	46 -0.3682175723	0
## GSM2630857	49	47 -0.1639329943	0
## GSM2630868	52	47 0.4116132618	0
## GSM2630818	50	47 -0.0608316878	0
## GSM2630929	54	47 -0.1272356228	0
## GSM2630932	51	47 -0.0845866511	0
## GSM2631230	53	49 0.0032341971	0
## GSM2631232	57	49 0.3899492115	0
## GSM2631234	69	50 0.1273560475	0
## GSM2630906	53	50 0.3872403761	0
## GSM2630917	58	50 0.3127298280	0
## GSM2631298	59	50 0.0257254438	0
## GSM2631158	52	51 -0.0597313257	0
## GSM2631167	60	51 -0.0120072012	0
## GSM2631175	63	51 -0.2229541259	0
## GSM2631176	NA	51 0.0556446776	0
## GSM2631177	58	51 0.0281937628	0
## GSM2630777	NA	52 -0.3812652966	0
## GSM2630788	58	52 -0.0931504744	0
## GSM2631173	59	53 -0.0207580150	0
## GSM2630787	54	53 -0.3740003573	0
## GSM2631100	61	53 -0.1098869820	0
## GSM2631118	NA	54 0.0182209788	0
## GSM2631039	60	54 0.0342952062	0
## GSM2631024	64	54 -0.1815996949	0
## GSM2631275	NA	55 0.3135896765	0
## GSM2631061	NA	55 0.2616320367	0
## GSM2631027	NA	55 -0.0554127774	0
## GSM2630980	NA	55 0.3108372458	0
## GSM2631105	56	55 -0.2570759631	0
## GSM2631032	NA	55 0.1758353429	0
## GSM2631082	58	55 0.0491639967	0
## GSM2631131	NA	55 -0.0824422320	0
## GSM2631047	60	55 -0.0422465024	0
## GSM2631180	65	55 -0.1377294518	0
## GSM2631038	NA	56 0.0999457968	0
## GSM2631059	59	56 -0.1383420739	0
## GSM2631085	59	56 0.3021180006	0
## GSM2630761	59	56 -0.0748953939	0
## GSM2631011	64	56 -0.0903771134	0
## GSM2631137	67	57 0.0538060543	0
## GSM2631277	73	57 0.0602106297	0
## GSM2631072	61	57 -0.0939756419	0
## GSM2631026	62	57 0.0998868259	0
## GSM2631284	62	57 0.0630271069	0
## GSM2631274	71	58 0.1136231806	0
## GSM2630774	71	58 -0.1640405477	0
## GSM2630993	63	58 -0.2129823439	0
## GSM2631112	67	58 0.0362726719	0
## GSM2631142	60	58 0.0647821034	0
## GSM2630984	71	58 0.1349552299	0

## GSM2631083	67	58	0.1358762580	0
## GSM2631068	NA	59	-0.0590313000	0
## GSM2631272	64	59	0.2241232311	0
## GSM2631123	65	59	-0.2695422786	0
## GSM2630855	71	60	-0.2321818066	0
## GSM2630758	64	60	-0.1726227631	0
## GSM2630866	80	60	-0.2094973202	0
## GSM2630840	63	60	0.0192708477	0
## GSM2630778	62	60	-0.1123960834	0
## GSM2630876	63	60	0.1347894708	0
## GSM2630965	NA	60	-0.3182871966	0
## GSM2630958	61	60	0.0202881196	0
## GSM2631044	77	60	-0.2164530381	0
## GSM2631080	66	60	-0.3462555919	0
## GSM2631179	63	61	0.2444754976	0
## GSM2631181	74	61	-0.0535447428	0
## GSM2631132	65	61	-0.0323790327	0
## GSM2631122	66	62	-0.0444032947	0
## GSM2631058	70	62	0.1472488193	0
## GSM2631081	64	62	0.3407157285	0
## GSM2631140	NA	62	0.0864484831	0
## GSM2631045	66	62	0.0073262075	0
## GSM2630812	67	63	0.1645513140	0
## GSM2630816	66	63	-0.1324290947	0
## GSM2630873	63	63	-0.0638843470	0
## GSM2630874	78	63	0.1110271307	0
## GSM2630803	NA	64	0.2043994103	0
## GSM2630772	NA	64	0.1289805806	0
## GSM2630875	NA	64	-0.1921792326	0
## GSM2631303	70	64	0.2524339302	0
## GSM2631222	69	64	0.1626319108	0
## GSM2630935	77	64	-0.4060597178	0
## GSM2631250	NA	64	0.0967163831	0
## GSM2630936	NA	65	0.1594038858	0
## GSM2630926	NA	65	-0.3816749088	0
## GSM2630833	67	65	0.0294311589	0
## GSM2630920	74	65	0.0255902903	0
## GSM2630844	70	65	-0.6118184477	0
## GSM2631217	67	65	-0.1690363581	0
## GSM2631226	69	65	0.4014600534	1
## GSM2631193	67	65	0.1813764093	1
## GSM2631282	NA	66	-0.0246015592	1
## GSM2631063	69	66	0.3264019816	1
## GSM2631141	71	66	0.0420676727	1
## GSM2630946	NA	67	0.0386019162	1
## GSM2630809	71	67	0.3809984358	1
## GSM2631292	72	67	0.2501502113	1
## GSM2630887	71	67	0.0775886379	1
## GSM2631251	70	67	0.2012195917	1
## GSM2630782	71	67	0.1620374371	1
## GSM2630858	69	67	0.4540467256	1
## GSM2630859	70	68	0.4829596283	1
## GSM2630852	NA	68	0.5219211804	1
## GSM2631139	NA	68	-0.1669816927	0

## GSM2630952	69	68 -0.1193681318	0
## GSM2631086	78	68 0.3593611172	0
## GSM2630962	69	68 0.0757851007	0
## GSM2631034	NA	69 -0.0362659932	0
## GSM2631092	72	69 -0.1461183888	0
## GSM2631286	74	69 0.2439021957	0
## GSM2631113	75	70 0.1200285112	0
## GSM2631093	71	70 -0.0703832811	0
## GSM2630989	70	70 0.4284944864	1
## GSM2631127	73	70 0.2018891315	1
## GSM2631258	76	72 0.3873551052	1
## GSM2630802	74	72 0.0704288303	1
## GSM2630983	NA	72 0.3162031378	1
## GSM2630914	75	73 0.2071276910	1
## GSM2630997	NA	74 0.2971352645	1
## GSM2630921	NA	75 0.1123782133	1
## GSM2630992	NA	76 0.3446099937	1
## GSM2630823	NA	76 0.2637098838	1
## GSM2631050	78	78 0.4122471406	1
## GSM2630994	NA	78 0.1968760655	1
## GSM2631021	NA	NA 0.1856891642	1
## GSM2630825	NA	NA -0.1368355207	1
## GSM2630821	NA	NA 0.3325432561	1
## GSM2631297	NA	NA 0.2087770486	1
## GSM2631114	NA	NA 0.4229274243	1
## GSM2631117	NA	NA 0.1731362552	1
## GSM2631260	NA	NA 0.1675814527	1
## GSM2631096	NA	NA 0.2129642554	1
## GSM2631003	NA	NA 0.2228625058	1
## GSM2631095	NA	NA 0.0146353863	1
## GSM2630940	NA	NA 0.2491197581	1
## GSM2630805	NA	NA 0.2363934829	1
## GSM2631009	NA	NA 0.0128259099	1
## GSM2630944	NA	NA 0.0724033832	1
## GSM2630947	NA	NA -0.3523216244	1
## GSM2630892	NA	NA 0.4874447599	1
## GSM2630973	NA	NA 0.5383556380	1
## GSM2630828	NA	NA 0.3688085064	1
## GSM2631192	NA	NA 0.6262424973	1
## GSM2630822	NA	NA 0.0592759754	1
## GSM2630969	NA	NA 0.5034897321	1
## GSM2631296	NA	NA 0.1295880360	1
## GSM2630956	NA	NA 0.0785098765	1
## GSM2630959	NA	NA 0.0802919119	1
## GSM2631291	NA	NA 0.0116655885	1
## GSM2630981	NA	NA 0.0802672895	1
## GSM2631067	NA	NA 0.1490238129	1
## GSM2631107	NA	NA 0.2600287801	1
## GSM2631186	NA	NA 0.3400259568	1
## GSM2630942	NA	NA 0.2002346464	1
## GSM2630867	NA	NA 0.1083356794	1
## GSM2631266	NA	NA 0.3190021207	1
## GSM2630763	NA	NA 0.3409741983	1
## GSM2631267	NA	NA 0.4056896209	1

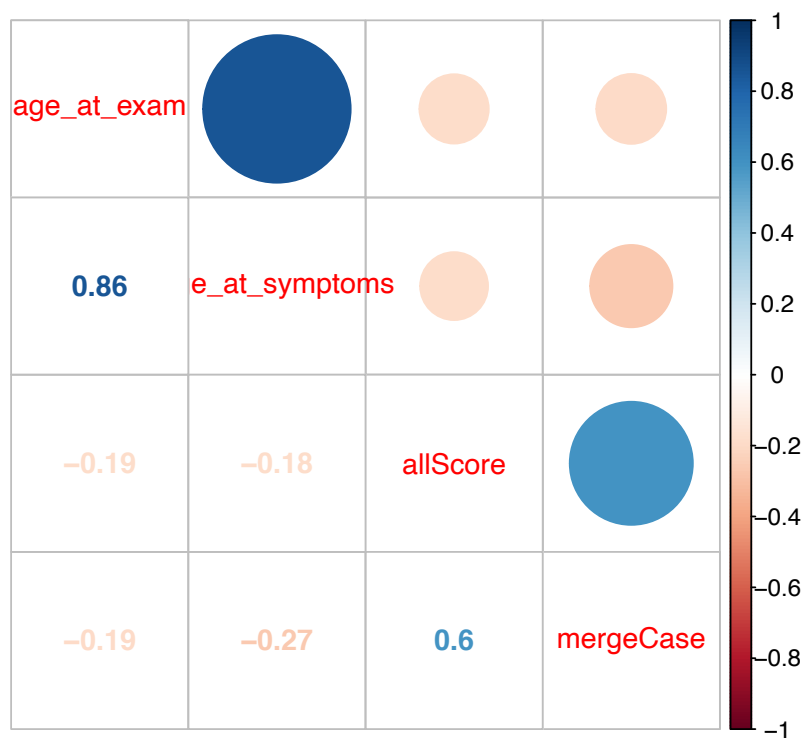

```
## GSM2631004      NA      NA 0.3515907890      1
## GSM2631104      NA      NA 0.4513433797      1
## GSM2630966      NA      NA 0.0324209020      1
## GSM2631269      NA      NA 0.1468274640      1
## GSM2631013      NA      NA 0.1451585074      1
## GSM2630995      NA      NA 0.1642121476      1
## GSM2631106      60      NA 0.2759626014      1
## GSM2631187      NA      NA 0.4110372889      1
## GSM2631261      NA      NA -0.0208964163      1
## GSM2631078      NA      51 0.3397995067      1
## GSM2631060      66      60 0.2339563384      1
## GSM2630762      NA      64 0.3560714020      1
## GSM2630838      NA      68 0.3017586700      1
## GSM2630900      75      71 -0.0246343182      1
## GSM2631244      76      72 0.2478615815      1
## GSM2630870      81      74 0.1617287673      1
## GSM2630862      NA      NA 0.2109527968      1
## GSM2631169      NA      69 0.1448082916      1
## GSM2631168      NA      NA 0.2337505989      1
## GSM2631170      63      52 -0.0018886159      1
## GSM2631019      NA      53 0.1181109146      1
## GSM2630765      76      66 0.1547808579      1
## GSM2630814      NA      72 -0.3945095738      0
## GSM2631199      NA      72 -0.0681534488      0
## GSM2631233      NA      NA 0.3345370758      1
## GSM2631216      NA      NA -0.1650202314      0
## GSM2630889      NA      NA -0.0844095755      0
## GSM2630792      NA      NA 0.3939263236      0
```

```
summary(modelData)
```

```
##   age_at_exam   age_at_symptoms   allScore   mergeCase
##   Min.   :30.00   Min.   :10.00   Min.   : -0.6118   Min.   :0.0000
##   1st Qu.:54.75   1st Qu.:45.00   1st Qu.: -0.0334   1st Qu.:0.0000
##   Median :61.00   Median :55.00   Median : 0.1298   Median :0.0000
##   Mean   :60.56   Mean   :53.61   Mean   : 0.1169   Mean   :0.4564
##   3rd Qu.:68.25   3rd Qu.:64.00   3rd Qu.: 0.2598   3rd Qu.:1.0000
##   Max.   :82.00   Max.   :78.00   Max.   : 0.6577   Max.   :1.0000
##   NA's   :266     NA's   :325
```

We should examine the correlations in our data set. You can do this quickly by building a correlation plot matrix.

```
install.packages("corrplot")
library(corrplot)
M<-cor(na.omit(modelData))
corrplot.mixed(M)
```



How would you interpret this output? Write a few sentences below!

- age_at_exam and age_at_symptoms is highly correlated to each other
- mergeCase and allScore are also very correlated

Let's build our first model. Here, we consider the case as our dependent variable, and the others as our explanatory variables.

```
model1<-glm(mergeCase~.,family=binomial,data=modelData)
```

This error is important - it represents that our model is drastically overfit. We can easily fix this using the BayesGLM model from the arm package

```
#install.packages("arm")
library(arm)
model1<-bayesglm(mergeCase~.,family=binomial,data=modelData)
summary(model1)
```

```
##
## Call:
## bayesglm(formula = mergeCase ~ ., family = binomial, data = modelData)
##
## Deviance Residuals:
##      Min       1Q   Median       3Q      Max
## -2.1132  -0.6876  -0.2873   0.7161   2.1547
##
## Coefficients:
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)   -0.16206    1.23382  -0.131   0.8955
## age_at_exam    0.04603    0.03531   1.304   0.1923
## age_at_symptoms -0.07472    0.03125  -2.391   0.0168 *
## allScore       8.25921    1.33364   6.193  5.9e-10 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for binomial family taken to be 1)
##
##      Null deviance: 215.96  on 157  degrees of freedom
## Residual deviance: 138.16  on 154  degrees of freedom
## (392 observations deleted due to missingness)
## AIC: 146.16
##
## Number of Fisher Scoring iterations: 9
```

We cannot use the step function for bayes glm. We will iteratively remove variables with the highest p-values, and then rerun the model.

Try this on your own, removing one by one and checking the output to find the next largest p-value. OR if you feel up to the challenge, write your own function to automate this process for you! There are bonus points available ;)

```
## Enter your own code here
model<-bayesglm(mergeCase~.,family=binomial,data=modelData)
model2 <- step(model)
```

```
## Start:  AIC=146.16
## mergeCase ~ age_at_exam + age_at_symptoms + allScore
##
##              Df Deviance    AIC
## <none>              138.16 146.16
## - age_at_exam      1   140.56 146.56
## - age_at_symptoms  1   145.36 151.36
## - allScore          1   203.64 209.64
```

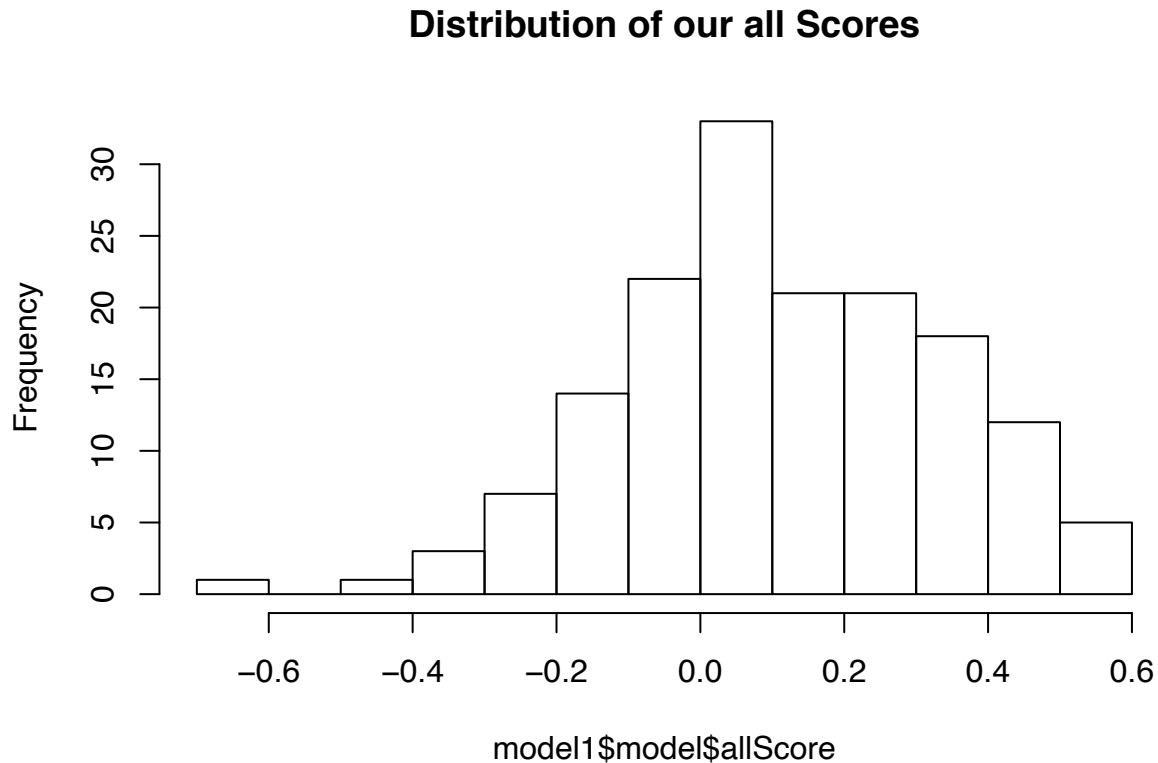
```
summary(model2)
```

```
##
## Call:
## bayesglm(formula = mergeCase ~ age_at_exam + age_at_symptoms +
##   allScore, family = binomial, data = modelData)
##
## Deviance Residuals:
##      Min       1Q   Median       3Q      Max
## -2.1132  -0.6876  -0.2873   0.7161   2.1547
##
## Coefficients:
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)   -0.16206    1.23382  -0.131   0.8955
## age_at_exam    0.04603    0.03531   1.304   0.1923
## age_at_symptoms -0.07472    0.03125  -2.391   0.0168 *
## allScore       8.25921    1.33364   6.193  5.9e-10 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for binomial family taken to be 1)
##
##      Null deviance: 215.96  on 157  degrees of freedom
## Residual deviance: 138.16  on 154  degrees of freedom
## (392 observations deleted due to missingness)
## AIC: 146.16
##
## Number of Fisher Scoring iterations: 9
```

This is our final model! Notice that our largest effect size is controlled by our genetic score. At a first glance, we might assume this means that the score has the largest effect on the model. However, if we recall how to interpret our coefficients, the estimated effect size is the change in log odds of being a case for a 1 unit increase in our score. Think about the score distribution: the range of our scores is fairly small. In contrast, the range of the updrs scores varies from 0 to 36. Keep in mind the scale of our data when interpreting these models!

Let's predict the probability of having a case given our model. Make a histogram of the score from this model.

```
## Enter your own code here
hist(model1$model$allScore,main="Distribution of our all Scores")
```



Like before, we'll build a violin plot to compare the output of our regression model. See if you can adapt the violin plot code from before to do this now.

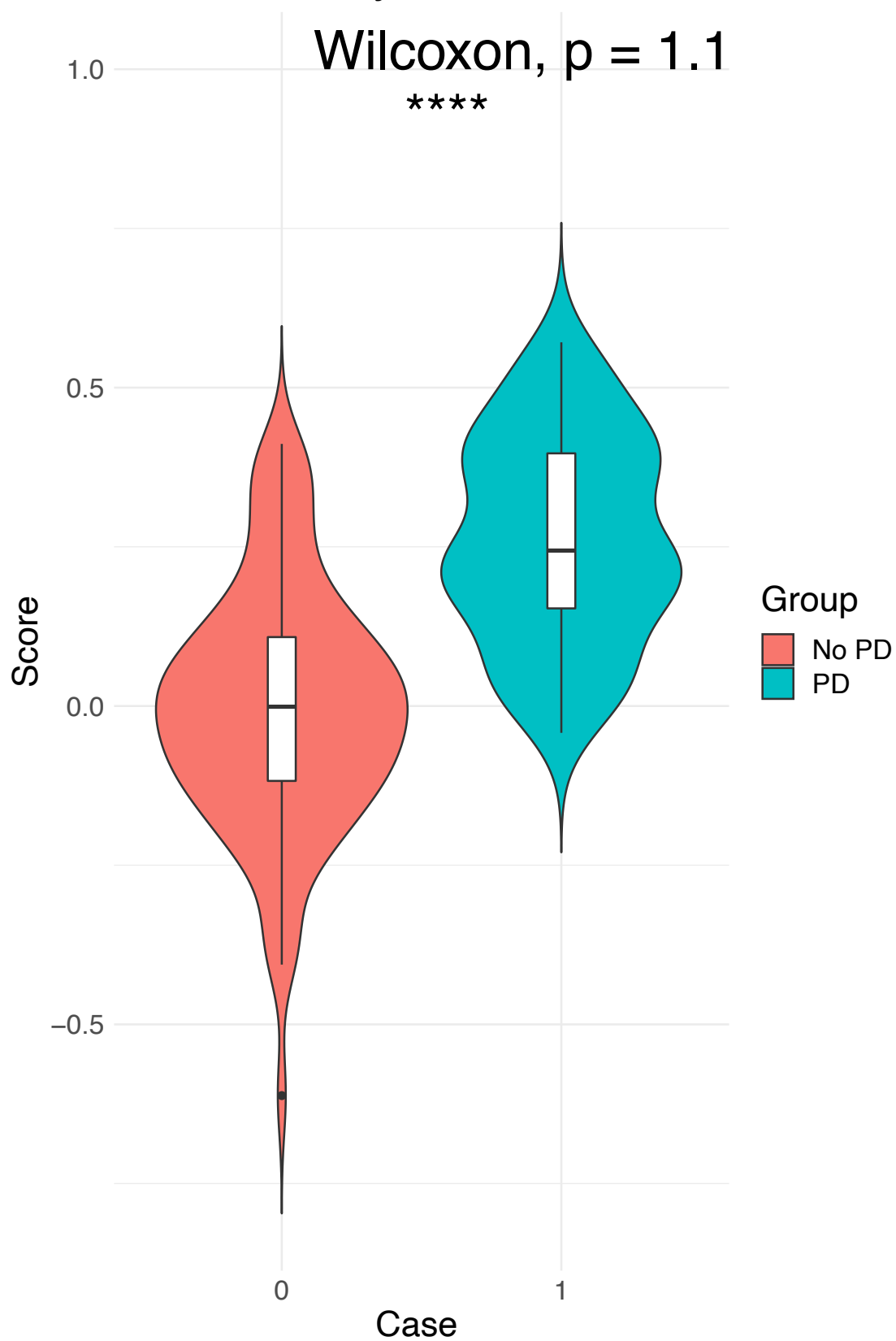
```
## Enter your own code here

df<-data.frame(cbind(model1$model$mergeCase,model1$model$allScore))

dp <- ggplot(df, aes(x=as.factor(model1$model$mergeCase), y=model1$model$allScore, fill=as.factor(model1$model$mergeCase))) +
  geom_violin(trim=FALSE)+
  geom_boxplot(width=0.1, fill="white")+
  labs(title="Plot of case by score",x="Case ", y = "Score")+
  stat_compare_means(label.x = 1.5, label.y = 1, size=10)+
  stat_compare_means(aes(label = ..p.signif..),
                     label.x = 1.5, label.y = 0.9, size =10) + theme_minimal() +
  scale_fill_discrete(name = "Group", labels = c("No PD", "PD")) +
  theme(text = element_text(size = 18))

dp
```

Plot of case by score



Now we're starting to see a clearer separation of scores! It's clear that by including the established tests to pre-screen patients for PD and other neurological diseases we have improved overall performance. While this may be an obvious conclusion, it is worth noting that the context with which our diagnostic signature would be used would be on patients already exhibiting potential PD symptoms. Clearly this needs a little more work, but for a first pass at assessing raw data, it's not bad!

Again, we can examine ROC curves. I've done some of the set up to get the data in the right format. Use the ROC code above to then build your own plot!

```
library("pROC")
nd<-cbind(pheno[-k,4:ncol(pheno)],newScore)
colnames(nd)<-c(colnames(modelData[1:ncol(modelData)-1]),"score")
newMScore<-predict(model2,newdata=nd)

## Enter your own code here
plot.roc(model1$model$mergeCase ~ model1$model$allScore, data=df, legacyaxes=F, print.auc=T, ci=T, main=

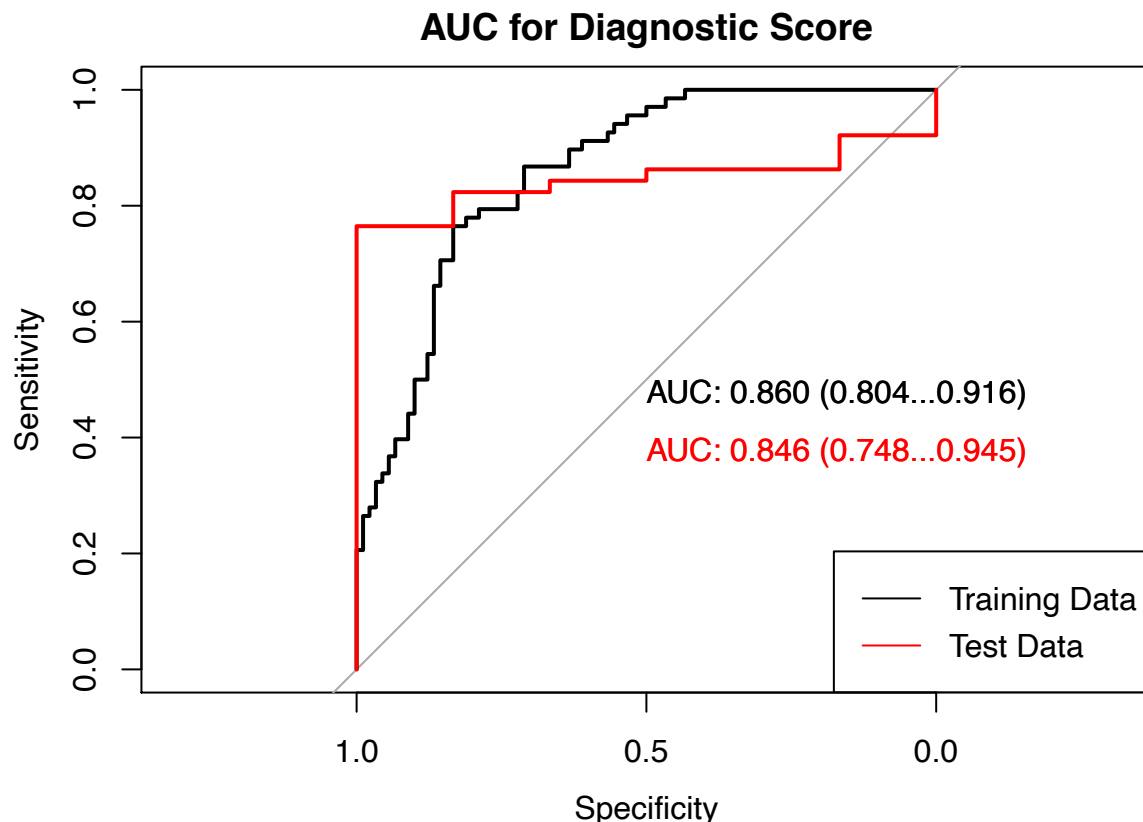
## Setting levels: control = 0, case = 1

## Setting direction: controls < cases

plot.roc(pheno$case[-k] ~ newMScore, data=data.frame(cbind(pheno$case[-k],newMScore)), add=T, print.auc=

## Setting levels: control = 0, case = 1
## Setting direction: controls < cases

legend("bottomright",c("Training Data","Test Data"),lty=c(1,1),col=c("black","red"))
```



Here, we have a notable increase in AUC, particularly for our training data. Our test data shows an overall improvement as well, although with a large confidence interval. There are clearly some data points in here which are abnormal - and perhaps worth investigating.

##Congratulations, you have finished the R Bootcamp Assignment!