Integrated spatial models foster complementarity between monitoring programmes in producing large-scale bottlenose dolphin indicators

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Abstract

Over the last decades, large-scale ecological projects have emerged that require collecting ecological data over broad spatial and temporal coverage. Yet, obtaining relevant information about large-scale population dynamics from a single monitoring programme is challenging, and often several sources of data, possibly heterogeneous, need to be integrated. In this context, integrated models combine multiple data types into a single analysis to quantify the population dynamics of a targeted population. When working at large geographical scales, integrated spatial models have the potential to produce spatialized ecological estimates that would be difficult to obtain if data were analysed separately. In this study, we illustrate how spatial integrated modelling offers a relevant framework for conducting ecological inference at large scales. Focusing on the Mediterranean bottlenose dolphins (*Tursiops truncatus*), we combined 21,464 km of photo-identification boat surveys collecting spatial capture–recapture data with 24,624 km of aerial line transect following a distance sampling protocol. We analysed spatial capture–recapture data together with distance sampling data to estimate the abundance and density of bottlenose dolphins. We compared the performances of the distance sampling model and the spatial capture–recapture model fitted independently to our integrated spatial model. The outputs of our spatial integrated models inform bottlenose dolphin ecological status in the French Mediterranean Sea and provide ecological indicators that are required for regional-scale ecological assessments like the EU Marine Strategy Framework Directive. We argue that integrated spatial models are widely applicable and relevant to conservation research and biodiversity assessment at large spatial scales.

Introduction

Macro-institutions get increasingly involved in large-scale programmes for biodiversity conservation over regional and continental areas. Whether these policies aim at assisting governments (e.g. the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services), or at implementing environmental management such as the European Union directives (Habitat Directive, 92/43/EEC, 1992, or Marine Strategy Framework Directive, MSFD, 2008/56/EC, 2008), conducting large-scale ecological monitoring is required to establish conservation status of targeted species and ecosystems, and to inform decision-making.

For biodiversity management decisions, conservation sciences require assessing the ecological status of species and ecosystems, which democratized the call for ecological indicators (Buckland *et al*., 2005; Nichols & Williams, 2006). An ecological indicator can be defined as a metric reflecting one or more components of the state of ecological systems. An ecological indicator can either be measured directly or result from the simplification of several field-estimated values (Niemi & McDonald, 2004). The Marine Strategy Framework Directive referred to the abundance/density of targeted species (e.g. seabirds and cetaceans) as ecological indicators to fulfil for national reporting. At large spatial scales, logistical and financial constraints often prevent a detailed coverage of the targeted population using a single collection effort, and different monitoring programmes coexist (Lindenmayer & Likens, 2010; Zipkin & Saunders, 2018; Isaac *et al*., 2019). The multiplication of monitoring programmes over the same conservation context has fostered the development of statistical models that can estimate

ecological quantities while accommodating several, possibly heterogeneous, datasets (Besbeas *et al.*, 2002; Isaac *et al.*, 2019; Miller *et al.*, 2019; Zipkin, Inouye, & Beissinger, 2019; Farr *et al.*, 2021). Integrating data from several monitoring protocols can give complementary insights into population structure and dynamics (Schaub & Abadi, 2011), increase space and time coverage of the population (Schaub & Abadi, 2011; Zipkin *et al.*, 2019) and produce more precise ecological estimates (Isaac *et al.*, 2019; Farr *et al.*, 2021; Lauret *et al.*, 2021).

A recurrent objective of ecological monitoring programmes is to estimate population abundance and density (Williams, Nichols, & Conroy, 2002), for which distance sampling (DS, Buckland *et al.*, 2005) and capture–recapture (CR, Williams *et al.*, 2002) methods are widely used. Abundance reflects the estimated number of animals in a specified area while density is a spatialized estimate that reflects the number of animals per unit area. DS and CR methods have strengths and weaknesses in relation to logistical and practical issues (Hammond *et al.*, 2021). DS methods can cover large areas at a reasonable cost (e.g. line transect monitoring), while CR monitoring programmes can be costly to develop at large spatial scales because more sampling effort is required over a longer time period to recapture individuals (Hammond *et al.*, 2021). Even when estimating abundance over the same study area, DS and CR do not estimate exactly the same quantity (Calambokidis & Barlow, 2004; Crum, Neyman, & Gowan, 2021). DS methods estimate abundance within a study area at the time of the survey. CR methods are based on individual sampling and estimate the number of animals that were present in the study area during the time of the monitoring (Calambokidis & Barlow, 2004). CR methods encapsulate a longer temporal extent because multiple sampling occasions are needed to build CR histories (Williams *et al.*, 2002). However, when data are collected over the same monitoring period and if animals do not move in and out of the study area during that period, CR and DS provide consistent estimates. Recent modelling tools have emerged to integrate both DS and CR methods into integrated population models (Kéry & Royle, 2020). DS and spatial CR methods (SCR) allow accounting for spatial variation in abundance and density (Miller *et al.*, 2013; Royle *et al.*, 2014; Camp *et al.*, 2020), possibly at large scales (Bischof *et al.*, 2020). The extension to integrated spatial models has been proposed to account for spatial variation in abundance and demographic parameters while analysing jointly DS data and SCR data (Chandler *et al.*, 2018). Integrated modelling holds promise for species occurring over large areas that are likely to be the target of multiple monitoring protocols. Besides, working at large geographical scales requires encapsulating spatial dimensions in the estimation of ecological quantities. Integrated spatial models allow to assess spatialized ecological inference, for example, density of individuals. To date, integrated spatial models have been developed and used on open populations to estimate temporal variation in population dynamics and vital rates such as survival and recruitment (Chandler & Clark, 2014; Chandler *et al.*, 2018; Sun, Royle, & Fuller, 2019).

These applications rely on long-term datasets that are not always compatible with conservation objectives. In many cases, ecological information is needed quickly, and data to investigate temporal variation are unavailable (Nichols & Williams, 2006; Lindenmayer & Likens, 2010). Consequently, the ecological inference is often restricted to closed population indicators (e.g. abundance or population size, density or spatial repartition of the population, distribution or spatial extent of a population). When the temporal resolution of monitoring programmes does not allow quantifying population dynamics, we argue that an application of integrated spatial models to closed populations can be useful in numerous ecological contexts to deal jointly with existing monitoring programmes and assess abundance and density.

In this study, we build an integrated spatial model and demonstrate the relevance of combining DS and SCR to build large-scale ecological indicators. We consider the monitoring of common bottlenose dolphins (*Tursiops truncatus*) that are considered as 'vulnerable' by the IUCN Red List in the North-Western Mediterranean Sea (IUCN, 2009). The protected status of bottlenose dolphins within the French seas (listed in *Annex II* of the European *Habitats Directive)* led to the development of specific programmes to monitor Mediterranean bottlenose dolphins within the implementation of the European Marine Strategy Framework Directive, which requires assessing the conservation status of this species every 6 years over the large extent of the French Mediterranean Sea (Authier *et al.*, 2017). Increasing efforts are dedicated to develop monitoring programmes in the Marine Protected Areas (MPA) network that mainly implements photo-identification protocols locally, while governmental agencies perform large-scale line transect programmes to monitor marine megafauna and fisheries. Hence, multiple data sources coexist about bottlenose dolphins in the French Mediterranean Sea. In this study, we analysed DS data collected by aerial line transect surveys over a large area covering coastal and pelagic seas (Laran *et al.*, 2017), which we combined with SCR data collected by a photo-identification monitoring programme restricted to coastal waters (Labach *et al.*, 2021). We compared the abundance and density of bottlenose dolphins estimated from DS model, SCR model, and integrated spatial models to highlight the benefits of the integrated approach in an applied ecological situation. We discussed the promising opportunities of using integrated spatial models in the context of marine monitoring planning in the French Mediterranean. Eventually, we underlined the conservation implications of using such a model to a wider extent to make the best use of available datasets.

Materials and methods

Monitoring bottlenose dolphins in the French Mediterranean Sea

Common bottlenose dolphins (*Tursiops truncatus*) occur over large areas throughout the Mediterranean Sea. Because monitoring elusive species in the marine realm is complex, multiple monitoring initiatives have emerged to collect data about

bottlenose dolphins in the French Mediterranean Sea. In the context of the Marine Strategy Framework Directive, the French government implemented large-scale aerial transects to monitor marine megafauna (Laran *et al.*, 2017). However, the large spatial coverage of the aerial monitoring is impaired by the low resolution of such data (i.e. one campaign every 6 years). Then, to collect detailed data, the French agency for biodiversity funded a photo-identification monitoring programme to investigate the ecological status of the bottlenose dolphins in the French Mediterranean Sea. This coastal boat photo-identification monitoring has been performed between 2013 and 2015. Coastal photo-identification monitoring represents a promising opportunity to produce high-resolution information because data can be collected routinely by Marine Protected Areas at a high time frequency.

Study area and datasets

We focused on an area of 255,000 km^2 covering the North-Western Mediterranean Sea within which we considered two monitoring programmes about bottlenose dolphins. We used SCR data from at-sea boat surveys over 21,464 km of the French continental shelf. Observers performed monitoring aboard small boats to locate and photo-identify bottlenose dolphins all year long between 2013 and 2015, always at a constant speed and with three observers. Taking pictures of the dorsal fin of each individual in the group makes possible the construction of detection history and hence the analysis of the population through capture–recapture methods (Labach *et al.*, 2021). Boat surveys were restricted to the coastal waters of France and adopted a search–encounter design covering approximately all the continental shelf every 3 months. We divided the duration of the monitoring programmes into eight equal sampling occasions that length for 3 months each, following a previous analysis by Labach *et al.* (2021). We also used DS data that were collected during winter and summer aerial line transect surveys covering 24,624 km of both coastal and pelagic NW Mediterranean Sea between November 2011 to February 2012 and May to August 2012 (Laran *et al.*, 2017). Two trained observers collected cetacean data following a DS protocol (i.e. recording species identification, group size, and declination angle). Aerial surveys were conditional on a good weather forecast. We divided the study area into 4356 contiguous grid cells creating a 5' × 5' Mardsen grid (WGS 84). To model the density of individuals, we used depth as an environmental covariate, which is expected to have a positive effect on bottlenose dolphins' occurrence (Bearzi, Fortuna, & Reeves, 2009; Labach *et al.*, 2021). To estimate the sampling effort of aerial and boat surveys, we calculated the transect length (in km) prospected by each monitoring protocol within each grid cell during a time period. Typically, entire transects are split into segments as they overlap multiple grid cells (Miller *et al.*, 2013). Sampling effort was therefore cell and occasion specific in the case of the SCR model, and cell specific for the DS model. Sampling efforts ranged from 0.047 km to 308 km per grid cell and per occasion for the photo-id dataset, and from 1.33 to 54560 km per grid cell for the aerial

line transect dataset. We used subjective weather conditions recorded by plane observers during the line transect protocols as a discrete variable ranging from 1 to 8. The good weather condition was expected to be positively related to the detection probability.

Spatial integrated models for closed populations

To integrate DS and SCR data, we used the hierarchical model proposed by Chandler *et al.* (2018). However, while initially developed for open populations and due to the lack of temporal depth in our datasets, we adapted the model to estimate abundance and density without accounting for demographic parameters (Fig. 1). We performed a closed population estimation of bottlenose dolphin density over the 2011–2015 period, assuming that (1) the population was demographically closed during the study period, (2) all individuals were correctly identified at each capture occasion and marks were permanent during the sampling period and (3) no migratory events occurred during the sampling period. Although being strong assumptions, bottlenose dolphin deaths and recruitments between 2011 and 2015 were likely small considering the long life cycle of bottlenose dolphins (Bearzi *et al.*, 2009; Hammond *et al.*, 2021). Besides, the Western Mediterranean bottlenose dolphin population is clustered into coastal subunits, hence we neglected migration events and movements that can occur between "French" resident groups and other populations, for example, offshore, Spanish, Italian or Atlantic groups (Louis *et al.*, 2014; Carnabuci *et al.*, 2016). We structure our integrated spatial model around two layers with i) an ecological model that describes the density of individuals based on an inhomogeneous point process (*Spatial abundance* section below) and ii) two observation models that describe how the DS and SCR data arise from the latent ecological model (*capture–recapture data* and *distance sampling data* sections below).

Spatial abundance

For the ecological model, we use a latent spatial point process modelling the density of individuals and the overall abundance. Over the study area *S*, an intensity function returns the expected number of individuals at location *s* in *S*. Here, *s*, represents an arbitrary point in the study area *S*. To account for spatial variation, we model the latent density surface as an inhomogeneous point process. For every location *s* in the study area *S*, the expected abundance λ is written as a log-linear function of an environmental covariate, say habitat:

$$log(\lambda(s)) = \mu_0 + \mu_1 habitat(s) \tag{1}$$

where parameters to be estimated μ_0 and μ_1, respectively, are the density intercept and the regression coefficient of the environmental covariate. For simplicity, we use depth as a habitat covariate possibly influencing bottlenose dolphin density, and explore a linear relationship between density and depth. The effect of habitat covariates could be further explored (e.g. by
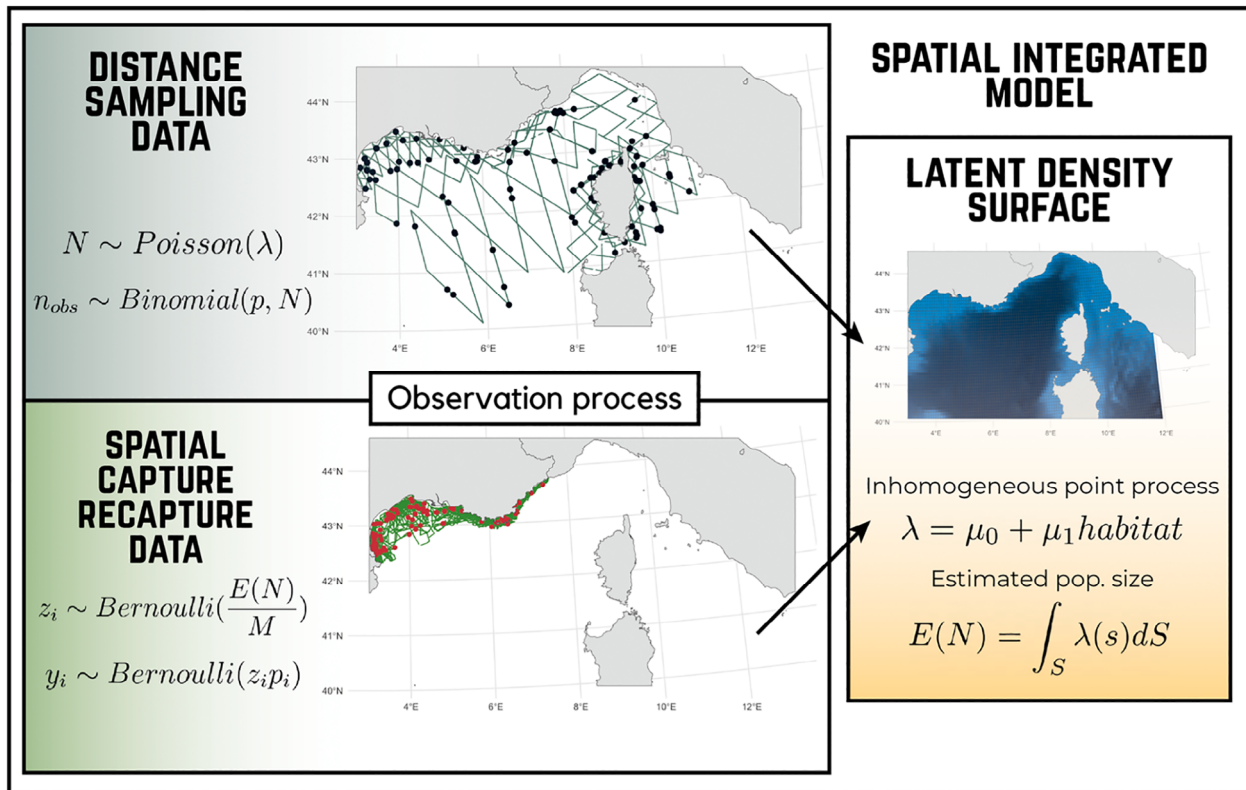
Figure 1 Graphical description of the spatial integrated model (SIM) that combines spatial capture–recapture (SCR) and distance sampling (DS). The SIM is a hierarchical model with three processes: i) latent population size E(N) and density λ informed by an inhomogeneous point process, ii) DS observation process that links the line transect dataset to the latent density surface and iii) SCR observation process that links the detection histories to the latent density. The observation process is stochastic according to detection probability. For DS model, the observed group size n_{obs} is a binomial draw in the latent abundance N at the sample grid cell. For SCR model, observing an individual *i* is a Bernoulli draw with a detection probability p_i. Through the data augmentation process with a hypothetical population size M, the probability an individual *i* belong to the study population is the result of a Bernoulli draw of probability E(N)/M.

considering other covariates such as sea surface temperature or prey availability, or by accounting for non-linear effects). Then, the estimated population size is derived by integrating the intensity function over the study area:

$$E(N) = \int_S \lambda(s) ds. \quad (2)$$

As we discretized the study area, we estimated λ_j the intensity process describing density for each grid cell *j* with $j = 1, \ldots, J = 4356$, hence $E(N) = \sum_{j=1}^J \lambda_j$. The latent ecological process defined by equation 1 is an inhomogeneous point process that is common to both the SCR and DS models. SCR and DS data are linked to density λ and informed the parameters of equation 1. To account for unseen individuals, we used the data augmentation technique and augmented the observed datasets to reach M = 20,000 individuals (Royle & Dorazio, 2012). Each individual *i* is considered being ($z_i = 1$) or not ($z_i = 0$) a member of the population according to a draw in a Bernoulli distribution of probability ψ, with $z_i \sim \text{Bernoulli}(\psi)$, where ψ is the probability for individual *i*

to be a member of the population, with $\psi = E(N)/M$ and $N = \sum_{i=1}^M z_i$.

Capture–recapture data

To link capture–recapture data with the ecological process, we built a SCR model (Royle *et al.*, 2014). Detection history of individuals was collected over T = 8 sampling occasions and capture locations were recorded. Grid cells *j* in which sampling effort was positive during an occasion were considered as active detectors for this sampling occasion, hence reflecting that animals could be observed. We stored observations in a three-dimensional array *y* with y_{ijt} indicating whether individual *i* was captured at grid-cell *j* during sampling occasion *t*. We assume that observation y_{ijt} is an outcome from a Bernoulli distribution with capture probability p_{ijt}, $y_{ijt} \sim \text{Bernoulli}\left(p_{ijt} z_i\right)$. We model capture probability with a half-normal detection function $p_{ijt} = p_0 exp\left(-\frac{d_{ij}^2}{2\sigma_{SCR}^2}\right)$, where d_{ij} is the Euclidian distance between the activity centre of individual *i* and the grid cell *j*, σ_{SCR} is the scale

parameter of the half-normal function and p_0 is the baseline encounter rate (Royle *et al.*, 2014). We accounted for spatial and temporal variation in the detection probability through the baseline detection rate p_0 that we modelled as a logit-linear function: $\text{logit}(p0_{jt}) = \delta_0 + \delta_1 E_{jt}$. When the sampling effort E_{jt} is null, we fixed $p0_{ijt}$ to 0.

The locations of the activity centre inform the density of individuals λ. For each individual i belonging to the sampled population, its activity centre is assigned as the result of a multinomial draw in the predicted density in each grid cell of the study area.

$$id_i \sim \text{Multinomial}(1, \overline{\lambda})$$

where id_i is the activity centre of individual i, and $\overline{\lambda}$ represents the vector of the predicted density in each cell of the study area. Due to the computational burden to sample the 4356 grid cells, we mimicked the multinomial distribution through the 'zeros trick' (see R codes for details). We considered that activity centres did not change between sampling occasions.

Distance sampling data

To accommodate distance data, we built a hierarchical DS model (Kéry & Royle, 2016). We model the DS data conditional on the underlying density surface defined by equations (1) and (2). We considered two sampling occasions t_{ds} as some transects were replicated. We assume that the probability of detecting dolphins is a decreasing function of the perpendicular distance between the transect and dolphin group. Because distance may not be estimated with perfection by observers, we discretized the distance of observation in B distance bins. Then, $r_{jbt} = r_{0(j,t)}\exp\left(-\frac{d_b^2}{2\eta^2}\right)$, where η is the scale parameter of the half-normal function, $r_{0(j)}$ is the probability of detection in the grid cell j and d_b is the observation distance between the flight transect and bin b where the detection occurred. The distance class d_j of the observed data at grid-cell j is modelled as a multinomial/categorical draw

$$d_{j,t} \mid n_{j,t} \sim \text{Multinomial}(1, \pi_{j,t})$$

with $\pi_{j,t}$ the vector of length B storing the detection probabilities in each bin b at grid-cell j. The bth index being $\pi_{j,b,t} = r_{j,b,t} / \left(\sum_b r_{j,b,t}\right)$.

We account for spatial variation in the baseline detection rate of the detection function modelling $r_{0(j,t)}$ as a log-linear function of weather condition $W_{j,t}$ in grid-cell j during sampling occasion t:

$$\text{logit}(r_{0(j,t)}) = \alpha_0 + \alpha_1 W_{j,t}.$$

Besides, aerial surveys only sampled a fraction of the total area of each grid cell (Appendix S1). We calculated $S_{j,t}$, the proportion of the grid cell effectively sampled by aerial

surveys considering a 1200-m-wide annulus around the transect. We assumed that density within each grid cell was uniform and remained constant across the sampling period. Then, $N_{j,t}$, the number of individuals sampled by aerial surveys in each grid-cell j during sampling occasion t is Poisson distributed, with λ_j being the density of individuals predicted by the point process in grid cell j restricted to the proportion of grid cell sampled, $S_{j,t}$.

$$N_{j,t} \sim \text{Poisson}(\lambda_j S_{j,t})$$

Then, $n_{j,t}$, the observed group size detected at grid cell j during sampling occasion t, is given by a binomial draw in the expected number of sampled individuals, and $N_{j,t}$ with probability the sum of $r_{j,b,t}$, the detection probabilities within each bin b of grid cell j during sampling occasion t.

$$n_{j,t} \mid N_{j,t} \sim \text{Binomial}\left(N_{j,t}, \sum_b r_{j,b,t}\right)$$

Bayesian implementation

To highlight the benefit of integrating data for the estimation of bottlenose dolphin density, we compared (i) the output of the spatial DS model, (ii) the SCR model and (iii) the integrated spatial model.

We ran all models with three Markov Chain Monte Carlo chains with 100,000 iterations each in the NIMBLE R package (de Valpine *et al.*, 2017). We checked for convergence calculating the *R-hat* parameter (Gelman *et al.*, 2013) and reported posterior mean and 80% credible intervals (CI) for each parameter. We considered as important the effect of a regression parameter whenever the 80% CI of its posterior distribution did not include 0. We also calculated the predicted density of bottlenose dolphins (i.e. λ). Data and codes are available on GitHub (https://github.com/valentinlauret/SpatialIntegratedModelTursiops).

Results

We detected 536 dolphins through aerial surveys clustered in 129 groups. We identified 927 dolphins over 1707 detections in photo-identification surveys, of which 638 dolphins were captured only once (68%), 144 were captured twice (15.5%), 149 were captured three times and up to eight times for one individual. The maximum distance between two sightings of the same individual was 302 km, with one individual detected twice during the same sampling occasion at 115 km distance.

We estimated 2451 dolphins (2337; 2566) with the integrated spatial model over the study area (Table 1), 11531 dolphins (10132; 12997) with the DS model and 1834 dolphins (1745; 1926) with the SCR model (Table 1). Density intercepts of the integrated spatial model ($\mu_0 = -0.85$ (−0.90; −0.79)) and SCR model ($\mu_0 = -1.18$ (−1.81; −1.07)) were lower than the intercept of the DS model ($\mu_0 = 0.95$ (0.82; 1.08)).

Table 1 Parameter estimates for the spatial integrated model (SIM), spatial capture–recapture (SCR) model and distance sampling (DS) model. For each parameter, we display the posterior mean and its 80% credible interval (CI)

Parameter	SIM		SCR model		DS model	
	Mean	80% CI	Mean	80% CI	Mean	80% CI
Estimated population size N	2451	2337, 2566	1834	1745, 1926	11531	10132, 12997
Intercept of density μ_0	−0.85	−0.90, −0.79	−1.18	−1.81, −1.07	0.95	0.82, 1.34
Effect of depth on density μ_1	0.32	0.26, 0.38	0.28	−0.47, 1.22	0.18	0.12, 0.25
SCR scale parameter: σ_{SCR}	531	156, 903	2458	500, 5920		
SCR p_0 parameter: Intercept δ_0	−12.54	−12.93, −12.16	−12.77	−13.53, −12.11		
SCR p_0 parameter: Effect of at-sea sampling effort δ_0	0.58	0.54, 0.63	0.58	0.53, 0.62		
DS scale parameter: σ_{DS}	3.21	1.09, 8.51			4.16	7.14, 9.44
DS r_0 parameter: Intercept α_0	3.32	2.80, 3.87			1.15	0.79, 1.51
DS r_0 parameter: Effect of weather condition α_1	1.64	1.15 2.1			1.86	1.52, 2.21

DS model estimated a positive effect of shallow waters ($\mu_1 = 0.18$ (0.12; 0.25), Table 1) similar to the effect estimated by the integrated spatial model ($\mu_1 = 0.32$ (0.26; 0.38), Table 1). However, the SCR model did not detect an effect of depth on density ($\mu_1 = 0.28$ (−0.47; 1.22), Table 1). Then, both integrated and DS models predicted higher densities of bottlenose dolphins in the coastal seas than in the pelagic seas, whereas the SCR model predicted no effect of depth on dolphin density.

Boat sampling effort exhibited a positive effect on detection probability for both the SCR model ($\beta_1 = 0.58$ (0.53; 0.62)) and the integrated spatial model ($\beta_1 = 0.58$ (0.54; 0.62), Table 1). For the integrated spatial model and the DS model, the detection probability increased when the weather condition improved (integrated spatial model: $\alpha_1 = 1.64$ (1.15; 2.10), DS: $\alpha_1 = 1.86$ (1.52; 2.21), Table 1).

Discussion

Integrated spatial model benefits from both distance sampling and capture–recapture data

With our integrated spatial model, we estimated bottlenose dolphin abundance within the range of what was found in previous studies in nearby areas (Gnone *et al.*, 2011; Lauriano *et al.*, 2014), and found that densities were more likely to be higher in coastal areas (Bearzi *et al.*, 2009). A striking result was the higher abundance estimated by DS compared to abundance estimated by the integrated and SCR models, which estimates were also found in previous studies analysing the same datasets in isolation. Using CR data only, Labach *et al.* (2021) estimated 2350 dolphins (95% confidence interval: 1827; 3135) inhabiting the French continental coast where our integrated model predicted 2451 dolphins (95% confidence interval: 2306; 2602). Analysing DS data, Laran et al. (2017) estimated 2946 individuals (95% confidence interval: 796; 11,462) during summer, and 10,233 (95% confidence interval: 4217; 24,861) during winter where our DS model estimated 11,531 (95% confidence interval: 9784; 13,478) all year long. Recent aerial campaigns performed in 2018–2019 on the same study area and following the same distance sampling protocol do not suggest a seasonal difference in bottlenose dolphins' abundance (Laran *et al.*, 2021).

We see several reasons that might explain the discrepancy in estimates obtained from SCR and DS models. First, although the Mediterranean bottlenose dolphins' population is clustered in coastal sub-units (Carnabuci *et al.*, 2016), groups can be encountered offshore (Bearzi *et al.*, 2009). In the DS dataset, large dolphin groups were detected in the pelagic seas at the extreme south of the sampling design (Appendix S1). These groups could either be i) occasional pelagic individuals belonging to coastal populations and that are mainly resident outside our study area (e.g. Balearic and South-Western Sardinia), or ii) resident pelagic populations that are not sampled by coastal photo-id surveys (Louis *et al.*, 2014). Second, SCR data were restricted to the French continental coast and did not sample dolphin populations that exist elsewhere in the study area, for example, in Corsica, Liguria and Tuscany (Carnabuci *et al.*, 2016). Despite this geographic sampling bias in the capture–recapture data, SCR models should predict the existence of Corsican and Italian populations if the relationship between density and habitat in equation (1) was correct and consistent throughout the study area. Predicting abundance outside the range of the data used could lead to biased estimates if the habitat–density relation is not correctly specified (Hammond *et al.*, 2021; Lee-Yaw *et al.*, 2021). As the photo-id surveys did not sample greater depths, our SCR model is likely to underestimate abundance because the relation linking dolphin density to depth was not correctly specified. Thus, we emphasised the relevance of aerial surveys that collected data in the pelagic seas, which helps to quantify the habitat–density relationship. To perform a detailed analysis of the NW Mediterranean bottlenose dolphin populations, one should consider additional environmental covariates to better capture spatial variation in density (e.g. sea surface temperature, distance to coast or 200 m contour, Lambert *et al.*, 2017). Besides, because Sardinian and Balearic populations and offshore groups can be sampled in the aerial surveys, the DS model drives upwards abundance compared to the SCR model which is unlikely to account for animals that are members of the Southern neither the Eastern nor offshore populations.

Overall, both DS and SCR data affected the estimates of the integrated spatial model. Using SCR data brought more information about population size (e.g. more detections, more individuals) than the DS data to inform the intercept of density (μ_0), making the integrated spatial model abundance estimate closer to the SCR model estimate (Table 1; Fig. 2). However, the DS data that were collected throughout the range of the habitat predictor informed the slope of the inhomogeneous point process (μ_1), that is, the effect of depth on dolphin density. Then, in the integrated spatial model, the SCR data informed the estimated population size and the DS data informed spatial repartition of individuals by correcting for the geographic sampling bias in the SCR data. The integrating approach helped to reduce the sampling limitations of each dataset and can improve the ecological inference as illustrated here about bottlenose dolphins.
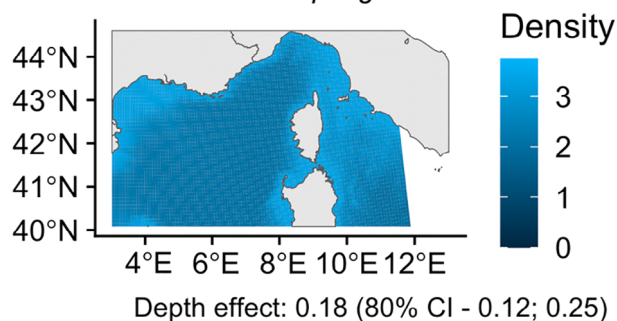
Conservation implications for monitoring bottlenose dolphins in the French Mediterranean Sea and beyond

When the conservation goal is to assess abundance in an area at a specific time, line transect surveys may be a cost-effective choice. However, if one's goal is to estimate the number of animals in an area over a longer period, CR methods could be more appropriate but have cost implications that may exceed those of conducting a line transect survey (Crum *et al.*, 2021; Hammond *et al.*, 2021). Despite differences in ecological inference, DS and CR are complementary methods depending on the conservation motivations and funding. To date, assessing the bottlenose dolphin population of the French Mediterranean Sea for the EU reporting only focuses on the DS data (Laran *et al.*, 2017). Aerial surveys provide crucial information on marine megafauna taxa, and on human pressures to fill several criteria of the Marine Strategy Framework Directive (Laran *et al.*, 2017; Pettex *et al.*, 2017; Lambert *et al.*, 2020). However, funding constraints make aerial monitoring hardly applicable at a high frequency, and it is planned to be implemented every 6 years. In parallel, the French office for biodiversity develops and supports local monitoring programmes in the French MPA network to perform photo-id data continuously; such detailed datasets represent an important asset to inform the abundance of marine mammals populations (Evans & Hammond, 2004). Ecological indicators required by the Marine Strategy Framework Directive for bottlenose dolphins would benefit from integrating aerial line transect with more data when available (Lauret *et al.*, 2021). In addition, the French Research Institute for Exploitation of the Sea (i.e. IFREMER) collected yearly bottlenose dolphins' data during line transects surveys for pelagic fisheries (Baudrier *et al.*, 2018). Ultimately, several monitoring programmes will be available for bottlenose dolphins in the Mediterranean context and integrated spatial models makes it possible to include existing datasets that have been discarded so far to inform public policies (Cheney *et al.*, 2013; Isaac *et al.*, 2019).
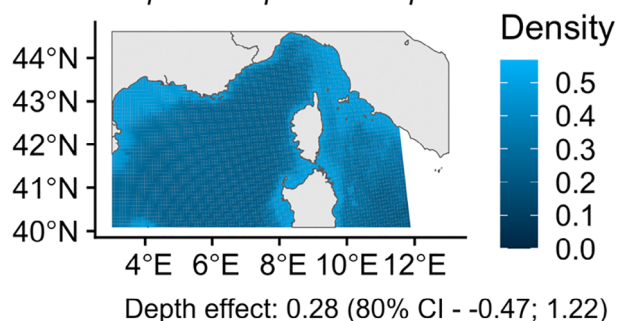
We acknowledge that our model has limitations due to several ecological features lacking, for example, spatial
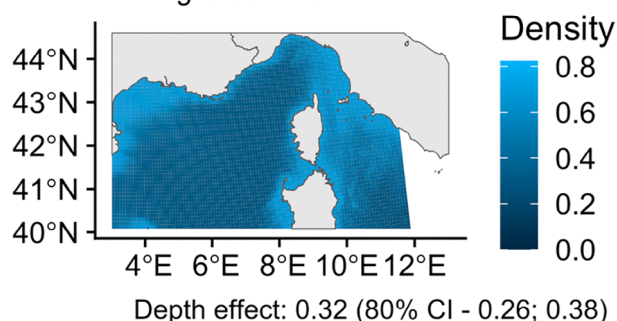

Figure 2 Estimated density surface of bottlenose dolphins (*Tursiops truncatus*) for the three models. Lighter colour indicates more individuals per area unit. Both spatial integrated model (SIM) and distance sampling model (DS) predicted higher density in coastal seas, while spatial capture–recapture model (SCR) predicted homogeneous density across the study area. Note that density scales are different between maps, indicating a higher overall population size for the DS model than for SIM and SCR model.

autocorrelation, effect of other environmental covariates, accounting for non-linear covariate effect and group behaviour of bottlenose dolphins that may generate non-

independent individual detection probabilities. One might also consider extending the activity centre process to include a movement model of individuals (Gowan, Crum, & Roberts, 2021). Moreover, ecological closure assumptions we assumed are likely to be violated but we assumed the bias introduced bias would be minimal. However, we emphasize that integrated spatial models are highly relevant considering the future monitoring planning by the French biodiversity agency that will perpetuate the coexistence of photo-identification with aerial line transect. Analysing the collected data in an integrated framework will lead to a more comprehensive understanding of how the monitoring programmes can work together and what exactly it is that they achieve in unison. It is our hope that the ability of integrating different datasets contributes to the ongoing monitoring efforts developed in the Mediterranean context and fits within the scope of what managers expect from statistical developments to inform environmental policies (Lauret, 2021).

Line transect and capture–recapture surveys are widely used monitoring methods to assess the population dynamics of marine mammals (Hammond *et al.*, 2021). Our work provides a promising modelling baseline to deal with the bottlenose dolphin evaluation but also opens perspectives for other conservation challenges about marine species that are subject to similar monitoring situations in the French Mediterranean context (e.g. fin whale and seabirds) and elsewhere.

Last, adding complementary long-term datasets to the aerial surveys would make it possible to access the demographic parameters (e.g. recruitments and survival (Chandler *et al.*, 2018)), which would represent a major opportunity for the knowledge about French Mediterranean bottlenose dolphin populations and to produce reliable conservation status. The use of integrated spatial models for the French Mediterranean bottlenose dolphin population also enables to extend the modelling approach exploring seasonality in density and to measure immigration and dispersal between bottlenose dolphin populations (Zipkin & Saunders, 2018). Finally, precising the assessment of bottlenose dolphin conservation status could ultimately lead to mitigation programmes in the context of the Marine Strategy Framework Directive, for example, marine protected areas implementation such as the Bottlenose dolphins Natura 2000 area in the French Gulf of Lion.

Spatial integrated models as a promising tool for conservation

When establishing species conservation status for large-scale environmental policies, discarding some datasets from the analysis can reduce the reliability of the ecological estimation (Bischof, Brøseth, & Gimenez, 2016). Using multiple datasets into integrated spatial models helps to overcome some limitations present when using separated information sources (e.g. limited spatial or temporal survey coverage, Zipkin & Saunders, 2018; Isaac *et al.*, 2019). However, caution should be taken as integrating data require additional modelling assumptions, for example, assuming population closure over a longer time period in our case (Dupont *et al.*,

2019; Fletcher *et al.*, 2019; Farr *et al.*, 2021; Simmonds *et al.*, 2020). Integrated spatial models are flexible tools that can include more than two datasets (Zipkin & Saunders, 2018), and various types of data that enlarge the scope of usable information (presence–absence (Santika *et al.*, 2017), count data (Chandler *et al.*, 2018) and citizen science data (Sun *et al.*, 2019)). Recent and current developments of SCR models widen perspectives to extend integrated spatial models to account for unidentified individuals, or to better describe animal movement (Milleret *et al.*, 2019; Jiménez *et al.*, 2020; Turek *et al.*, 2021). Over the last decades, the spatial scope of conservation efforts has greatly increased, and the analytical methods have had to adapt accordingly (Zipkin & Saunders, 2018). Integrated spatial models are a promising tool that can be used in multiple situations where several data sources coexist, especially for large-scale conservation policies.

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Supporting information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Appendix S1. Data and models results.