



Dr Manoj Kumar Valluru

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EDUCATION AND QUALIFICATIONS:

- **01/2014-03/2018, PhD Molecular Biology and Biotechnology**, Project: “The role of microRNAs in development”, Dr K Sorefan. Department of Molecular biology and Biotechnology, University of Sheffield, UK.
- **2007-2008, MSc Stem Cell and Regenerative Medicine (Merit)**, Project: “Sodium butyrate and compressive loading regulated osteogenesis through ERK pathway in human mesenchymal stem cells cultured in 3D scaffolds”, Kroto Research Institute, Prof G Reilly. Department of Biomedical Science, University of Sheffield, UK.
- **2003-2007, B.Eng. Biotechnology (First)**, Project: IL-2 in multiple myeloma, Prof R. A. Nazeer. School of Bioengineering, SRM University, Chennai, INDIA.
- **2001-2003, Zoology, Botany, Physics, Chemistry & English (First)**, Hindu Higher Secondary School, Chennai, INDIA.

RESEARCH EXPERIENCE:

- **08/2019-7/2025, KRUK and PKDF Postdoctoral Researcher**, Project: “Functionally characterize the transcriptomic and proteomic signatures altered in cellular and experimental models of ADPKD”, Prof Albert Ong. IICD, University of Sheffield, UK.
- **04/2018-07/2019, Bioinformatician**, Project 1: “VEGFA isoform switch in different cancer subtypes”, Dr W R English. Project 2: “Cell-Type Signatures in Uveal Melanoma”, Dr K Sisley. Project 3: “miRNAs in invasive breast cancer bone metastasis”, Prof P Clézardin. Department of Oncology and Metabolism, University of Sheffield, UK.
- **04/2013-12/2013, Research Assistant**, Project: “Genetic engineering of ovarian cancer cells to investigate resistance mechanisms to antivascular agents”, Dr WR English and Prof GM. Tozer. Department of Oncology, University of Sheffield, UK.
- **11/2011-12/2012, Research Engineer**, Project: “Neurosphere and Human embryonic stem cell bioprocessing”, Dr V Chotteau. Biotechnology Department, KTH Royal Institute of Technology, Stockholm, SWEDEN.
- **04/2009-12/2010, Research Technician**, Project: “Regulation of Angiogenesis Initiation and Regression in Wound Healing”, Dr CA Staton, Prof NJ Brown and Prof MWR Reed. Department of Oncology, University of Sheffield, UK.

PUBLICATIONS, PREPRINTS, BOOK CHAPTERS and ABSTRACTS: ORCID iD [0000-0001-9156-866X](https://orcid.org/0000-0001-9156-866X) and RID AAP-9004-2020

Publications

(1) Chen EWC, Chong J, Valluru MK, et al., (2024) Combining genotype with height-adjusted kidney length predicts rapid progression of ADPKD. *Nephrol Dial Transplant* (20%). (2) Miranda Durkie, Christopher M. Watson, Peter Winship, Anne-Cecile Hogg, Rodney Nyanhete, Sharon Cooley, Manoj K. Valluru et al., (2023) The Common PKD1 p.(Ile3167Phe) Variant Is Hypomorphic and Associated with Very Early Onset, Biallelic Polycystic Kidney Disease, Human Mutation (5%). (3) Valluru MK et al., (2023) Genomics England Research Consortium. A founder UMOD variant is a common cause of hereditary nephropathy in the British population. *J Med Genet* (60%). (4) Puppo M, Valluru MK et al., (2023) MiR-662 is associated with metastatic relapse in early-stage breast cancer and promotes metastasis by stimulating cancer cell stemness. *Br J Cancer* (35%). (5) Durkie M, Chong J, Valluru MK et al., (2021) Biallelic inheritance of hypomorphic PKD1 variants is highly prevalent in very early onset polycystic kidney disease. *Genet Med* (20%). (6) Magayr TA, Song X, Streets AJ, Vergoz L, Chang L, Valluru MK et al., (2020) Global microRNA profiling in human urinary exosomes reveals novel disease biomarkers and cellular pathways for autosomal dominant polycystic kidney disease. *Kidney Int* (3%). (7) Lannoy M, Valluru MK et al., (2020) The positive effect of selective prostaglandin E2 receptor EP2 and EP4 blockade on cystogenesis *in vitro* is counteracted by increased kidney inflammation *in vivo*. *Kidney Int* (40%). (8) Doherty RE, Bryant HE, Valluru MK et al., (2019) Increased Non-Homologous End Joining Makes DNA-PK a Promising Target for Therapeutic Intervention in Uveal Melanoma. *Cancers (Basel)* (10%). (9) Simonini S, Deb J, Moubayidin L, Stephenson P, Valluru M et al., (2016) A non-canonical auxin-sensing mechanism is required for organ morphogenesis in *Arabidopsis*. *Genes & Dev* (10%). (10) Staton CA, Shaw LA, Valluru M et al., (2011) Expression of class 3 semaphorins and their receptors in human breast neoplasia. *Histopathology* (10%). (11) Valluru M et al., (2011) Blood vessel characterisation in human dermal wound healing. *Br J Dermatology* (80%). (12) Staton CA, Valluru M et al., (2010) Angiopoietin-1, angiopoietin-2 and Tie-2 receptor expression in human dermal wound repair and scarring. *Br J Dermatology* (30%).

Review Articles (1) Z Mao, MK Valluru *et al.*, (2021) **Drug repurposing in autosomal dominant polycystic kidney disease: back to the future with pioglitazone.** *Clinical Kidney Journal* (20%). **(2)** M Puppo, MK Valluru *et al.*, (2021) **MicroRNAs and Their Roles in Breast Cancer Bone Metastasis.** *Current osteoporosis reports* (15%). **(3)** Dardente H, English WR, Valluru MK *et al.*, (2020) **Debunking the myth of the endogenous *Vegfxxx* transcripts.** *Trends in endocrinology & metabolism* (20%). **(4)** Valluru M *et al.*, (2011) **Transforming growth factor- β and Endoglin signaling orchestrate wound healing.** *Front. Physio* (90%).

Preprints and manuscripts in progress (1) MK Valluru & K Sorefan. (2019) **Control of stem cell niche and fruit development in *Arabidopsis thaliana* by AGO10/ZWL requires the bHLH transcription factor INDEHISCENT.** *Development biology. BioRxiv* (80%). **(2)** Valluru MK, Ritchie J *et al.*, **Deconvoluting Callender Classification Cell-Type Signatures in Uveal Melanoma.** *Cancers (Basel)* (90%). **(3)** M Puppo, M Croset, D Ceresa, MK Valluru *et al.*, (2024) **Protective effects of miR-24-2-5p in breast-to-bone metastasis: a dual inhibitory action on cancer cells and bone-resorbing osteoclasts.** *Breast Cancer* (15%). **(4)** J Fingret, K Sisley, MK Valluru *et al.*, (2024) **Surgical Interventions Improve Survival in Patients with Metastatic Uveal Melanoma.** *British Journal of Ophthalmology* (25%).

Book Chapters (1) M Puppo, M K Valluru, P Clézardin. Chapter 33, **MicroRNAs and bone metastasis: how small RNAs regulate secondary tumour formation and progression in the skeleton.** Dominique Heymann (ed.) Bone Cancer 3E. Elsevier 2022 (40%).

Conference abstracts (1) Kai Xin Chung N, Valluru M *et al.*, (2022) **MO030: Familial clustering of a rare UMOD variant in undiagnosed hereditary nephropathy suggests the presence of a common ancestral founder mutation.** *Nephrology Dialysis Transplantation*. **(2)** M Puppo, MK Valluru *et al.*, (2021) **Functional studies on miRNAs with a potential role in breast cancer bone metastasis.** *RNA* 2021, P3-36. **(3)** MK Valluru *et al.*, (2019) **VEGFA isoform switching in soft tissue sarcoma is associated with decreased survival.** *International journal of experimental pathology*. 100 (4), A41-A41. **(4)** BL Aguero, M Valluru *et al.*, (2019) **Prognostic significance of soluble and ECM associated VEGFA isoforms in high-grade serous ovarian cancer.** *International journal of experimental pathology*. 100(4), A8-A9. **(5)** Carolina Å, Linn W, Erika H, Manoj V. *et al.*, “Longitudinal Study of Neurosphere Metabolism in Bioreactor Culture for Scalable Production of Human Embryonic Stem Cell-Derived Neurons”. 4th International Conference on Stem Cell Engineering (2014). American Institute of Chemical Engineers. **(6)** Valluru M *et al.*, (2010) **Class 3 semaphorins and their plexin receptors in human dermal wound healing & Class 3 semaphorins inhibit angiogenic activity in vitro.** *Microcirculation* 17:485 PC50&51.

TEACHING EXPERIENCE & AWARDS:

(1) [STA](#) at The University of Sheffield & [Bioprocess design](#) at KTH. **Teaching at The University of Sheffield (2022-present):** MED6003 Data analysis methods in genetic epidemiology and MED6071 RNA-seq File handling. Co-Supervisor: PhD student Linda Vidova (Rare Tumours Research Group). **(2)** MBB PhD Studentship Award (Sheffield, 2014) and Robert hill institute poster prize (Sheffield, 2015). **(3)** SciLifeLab (Stockholm, 2012), IRCC (Torino, 2011) travel and training award, EMBO travel grant (2022), FASEB full travel and conference grant (2022).

CONTINUING PROFESSIONAL DEVELOPMENT:

Training (1) [Cancer research training](#), Cancer Cell Biology, IRCC - Institute for Cancer Research and Treatment, Turin, Italy. (3rd July – 23rd July 2011). **(2)** [Computational Methods for Massively Parallel Sequencing](#), SciLifeLab – Uppsala University, Uppsala, Sweden (28th January – 3rd February 2012). **(3)** [Clinical bioinformatics: unlocking genomics in healthcare](#), University of Manchester, UK (Further Learn, Dr A Davies, 09/18-10/18). **(4)** [The UNIX Workbench and Genomic Data Science](#), Johns Hopkins Bloomberg School of Public Health (Coursera, Dr Jeffrey Leek, 10/18-12/18).

Workshops (1) [Introduction to Identifying & Characterising Variants from NGS Data](#), University of Sheffield (Dr M Parker, 4th & 5th June 2018). **(2)** [Python Programming Workshop](#), University of Sheffield (Dr Mark Quinn, July 2018). **(3)** [Analysis of RNA-seq data in R](#), University of Sheffield (Dr M Dunning, 9th & 10th July 2018). **(4)** [Workshop on proximity labeling](#) (Ting Lab, November 2020). **(5)** [MaxQuant - Computational Mass Spectrometry-Based proteomics](#) (Max-Planck-Institute of Biochemistry, June 2021). **(6)** [HPC for Healthcare](#), N8 CIR (Dr M Dunning and W Furnass, 12th July 2021). **(7)** [Deep Learning for microscopy image analysis](#), Human Technopole (Dr Florian Jug, 22-26th Nov 2021). **(8)** [Spatial transcriptomics data analysis in Python](#), Single Cell Omics Germany (May 23-24, 2022).

Peer review: Nature scientific rep, Frontiers Oncology and European Journal of Inflammation.

SKILLS:

Wet lab (1) Plant and mammalian cell culture (Stem cells, primary cells and cell lines) on scaffolds and using bioreactors. ***In vitro* assays:** BioProfile FLEX (culture analysis), MTS and BrdU assay (Cell proliferation), scratch assay (Cell migration), capillary tubule formation assay on Matrigel and Aortic ring assay (angiogenesis methods). ***In vivo:*** Home office certificate (2010, Mouse: Modules 1 to 4). CD-1 mouse: Incisional Wound Healing. **(2)** Molecular cloning, DNA and RNA prep, QRT-PCR, PCR, Gene knockin using ZFNs, Protein expression (*E.coli*), Protein biotinylation, Western blotting, ELISA, Pull-down assay, ChIP, RNC and EMSA. **(3)** PLA, STELLARIS smFISH, TFM, Live cell imaging, Immunocytochemistry, Immunohistochemistry on resin and paraffin-embedded sections. SEM, super-resolution confocal imaging and Fluorescence microscopy (Zeiss LSM 980 with Airyscan 2, Slide scanner and Celldiscoverer7, Olympus FV1000 and Leica THUNDER).

Data science (1) Programming languages: Bash, Python and R (intermediate), able to run different data analysis pipelines, e.g., sRNA- seq, RNA/RNC-seq, fRIP/CLIP-seq, RNA structural motif, RNA splicing, APA, Proteomic data analysis, Protein structure modelling etc. (conda environment, bash script). **(2)** Genomic data wrangling and analysis, Databases: [UKBB](#), [100K](#), [TCGA](#), [GTEx](#) and [CCLE](#). **(3)** High-Performance Computing: STANAGE (The Sheffield University HPC cluster). **(4)** Image analysis using Image J/Napari and other tools from GitHub. **(5)** Other statistics: R, Sigmaplot, Graphpad, and SPSS, Operating systems: Linux/UNIX (Ubuntu), Mac OS X, Windows.

Management (1) Medical School Athena swan data group – 2019 to 2020. **(2)** MDHRSa committee (Junior chair - Medical School Annual Research Meeting 2021). **(3)** Identifying new resources, working successfully with others, handle large amounts of data and presenting at weekly and monthly project meetings. **(4)** Contributing to grant applications.

MEMBERSHIPS:

(1) [RNA Society](#): Bethesda, Maryland, US (2020-01-01 to present | Post Doc Membership)