

## PCR Genotyping Protocol for *Ptf1a<sup>tdTomato</sup>* mouse line

### PCR reagents:

- oligo primers at 20  $\mu$ M  
F-p48            5'-CCT TCT GAC TTC TCC AAG AAG GCA-3' (top)  
R-5'p48        5'-CCC TTT ATG CCT GGC ATT TCA CTG-3' (bottom)

F-p48+ R-5'p48 amplification yields **670bp TM** (targeted mutant allele) band and a **636bp WT** (wild type allele) band.

- Perkin Elmer PCR buffer with  $MgCl_2$
- dNTP premix ((dNTP premix is made by using 100 mM NEB dNTP's. The premix contains 250  $\mu$ l of each dNTP - A, C, G, & T - and 19 ml sterile water and is stable at  $-20^{\circ}C$ . I freeze this in 1 ml aliquots and thaw and refreeze as needed.)
- Perkin Elmer Amplitaq Gold
- genomic DNA samples diluted to 50 ng/ $\mu$ l with sterile water

### PCR reaction mixture:

15.8  $\mu$ l sterile water  
2.5  $\mu$ l 10X PCR buffer  
4  $\mu$ l dNTP premix  
0.75  $\mu$ l F-p48 primer  
0.75  $\mu$ l R-5'p48 primer  
1  $\mu$ l dil. DNA template  
0.2  $\mu$ l Amplitaq Gold  
25  $\mu$ l total volume

### Cycling conditions:

1 cycle -  $94^{\circ}C$  x 6 min.  
40 cycles -  $94^{\circ}C$  x 1 min.,  $60^{\circ}C$  x 30 sec.,  $72^{\circ}C$  x 1 min.  
1 cycle -  $72^{\circ}C$  x 7 min.  
hold at  $4^{\circ}C$ .

### Analysis of PCR products:

10  $\mu$ l aliquots of reactions + 2  $\mu$ l of 6X gel loading buffer in a 1% mini-agarose gel

