

Materials

Material needed for DNA origami (see preparation)

Procedures

1. Loading of helper strands
 - a. Prepare the M13, S-x, F-cap-x and S-PAM-cap-x mix in a microcentrifuge tube on ice by adding the components indicated below. The molar ratio of M13 (5nM) and helper strands (50 nM) is 1:10.

DNA origami (DO) (with S-PAM-cap-x and F-cap-x)

Reagents	Volume
TAE/Mg ²⁺ buffer	47µl
S-x	0.25ul × 150 = 37.5µl
F-cap-x	0.25ul × 36 = 9µl
S-PAM-cap-x	0.25ul × 6 = 1.5µl
M13	5µl

S-13	S-62	S-117	S-166	F-Cap76
S-14	S-63	S-118	S-167	F-Cap77
S-15	S-64	S-119	S-168	F-Cap78
S-16	S-65	S-120	S-169	F-Cap79
S-17	S-66	S-121	S-170	F-Cap80
S-18	S-67	S-122	S-180	F-Cap81
S-19	S-68	S-132	S-181	F-Cap82
S-20	S-69	S-133	S-182	F-Cap83
S-21	S-70	S-134	S-183	F-Cap123
S-22	S-71	S-135	S-184	F-Cap124
S-23	S-72	S-136	S-185	F-Cap129
S-24	S-73	S-137	S-186	F-Cap130
S-25	S-74	S-138	S-187	F-Cap131
S-26	S-84	S-139	S-188	F-Cap171
S-36	S-85	S-140	S-189	F-Cap172
S-37	S-86	S-141	S-190	F-Cap173
S-38	S-87	S-142	S-191	F-Cap174
S-39	S-88	S-143	S-192	F-Cap175
S-40	S-89	S-144	S-193	F-Cap176
S-41	S-90	S-145	S-194	F-Cap177
S-42	S-91	S-146	S-195	F-Cap178
S-43	S-92	S-147	S-196	F-Cap179
S-44	S-93	S-148	S-197	F-Cap125
S-45	S-94	S-149	S-198	F-Cap126
S-46	S-95	S-150	S-199	F-Cap127

S-47	S-96	S-151	S-200	F-Cap128
S-48	S-97	S-152	S-201	S-PAM-Cap102
S-49	S-98	S-153	S-202	S-PAM-Cap103
S-50	S-99	S-154	S-203	S-PAM-Cap104
S-51	S-100	S-155	S-204	S-PAM-Cap113
S-52	S-101	S-156	F-Cap27	S-PAM-Cap114
S-53	S-105	S-157	F-Cap28	S-PAM-Cap115
S-54	S-106	S-158	F-Cap29	
S-55	S-107	S-159	F-Cap30	
S-56	S-108	S-160	F-Cap31	
S-57	S-109	S-161	F-Cap32	
S-58	S-110	S-162	F-Cap33	
S-59	S-111	S-163	F-Cap34	
S-60	S-112	S-164	F-Cap35	
S-61	S-116	S-165	F-Cap75	

- b. Mix the above reagents and briefly centrifuge them. The mix is first heated at 95 °C for 10 minutes. Subsequently, annealing is performed by slowly cooling the mixture from 95 °C to 20 °C at a rate of 1 °C/min.
 - c. The Amicon® Ultra-0.5 Centrifugal Filter Devices (100kDa) are used to remove redundant helper strands three times. The concentrated DO solution is obtained.
2. Loading functional strands
- a. Add 1.5 µL of the PAM-rich stock solution and 9µl F-H stock solution to the concentrated solution. Then, restore the volume to 100µl with TAE/Mg²⁺ buffer.
 - b. Mix the above reagents and briefly centrifuge them. The mix is annealed from 45 °C to 25 °C at a rate of 5 min/°C for six cycles.
 - c. The Amicon® Ultra-0.5 Centrifugal Filter Devices (100kDa) are used to remove redundant PAM-rich and F-H three times. The concentrated Cy5-labeled DO_{PAM} solution is obtained.
3. Loading FITC-sgRNA_L/Cas9
- a. Prepare the FITC-sgRNA_L and Cas9 mix in a single microcentrifuge tube on ice by adding the components indicated below. The molar ratio of FITC-sgRNA_L and Cas9 is 1:1.
- | Reagents | Volume |
|--------------------------------|--------|
| PBS (PH=7.4, free of RNase H) | 94 µl |
| FITC-sgRNA _L (5 µM) | 3µl |
| Cas9 (5 µM) | 3µl |

- b. Mix the above reagents and briefly centrifuge them. Incubate at 37 °C

- for 10 minutes to get FITC-sgRNA_L/Cas9 complex.
- c. Add 10 μ l of FITC-sgRNA_L/Cas9 mixture to the concentrated Cy5-labeled DO_{PAM} solution, and make up to 100 μ l with TAE/Mg²⁺ Buffer. The molar ratio of FITC-sgRNA_L/Cas9 complex and Cy5-labeled DO_{PAM} is 3:1.
 - d. Mix the above reagents and briefly centrifuge them. The mix is annealed from 37 °C to 4 °C at a rate of 1°C/min.
 - e. The Amicon® Ultra-0.5 Centrifugal Filter Devices (100 kDa) are used to remove the redundant FITC-sgRNA_L/Cas9. The concentrated Cy5-labeled DO_{PAM}R^{FITC}C solution is obtained.
4. Verifying the loading of FITC-sgRNA_L/Cas9 complex
- a. Measure the absorbance of Cy5-labeled DO_{PAM}R^{FITC}C solution at 495nm and 650nm by using Thermo Fisher Scientific NanoDrop One 2.12.0 and python 3.13.
 - b. Plot the absorption spectrum of the Cy5-labeled DO_{PAM}R^{FITC}C solution.